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1 **A new application of NIR spectroscopy to describe and predict purees quality**  
2 **from the non-destructive apple measurements**

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24

25 **Highlights**

26 Texture and taste of cooked purees could be predicted from the spectra of raw apples.

27 Apples and purees were well classified by NIRS according to the studied factors.

28 NIRS could predict acceptably quality traits of both apples and purees.

29

30 **Abstract**

31 The potential of NIRS was investigated on both apples and purees to (i) examine  
32 factors involving quality variability (variety, agricultural practice, cold storage, puree  
33 mechanical refining level) and (ii) establish the link between quality traits before and  
34 after processing in order to predict the quality characteristics of purees from spectral  
35 information of raw apples. Apples and purees were well-classified at over 82% and 88%  
36 according to varieties and storage times respectively. The PLS models showed a good  
37 ability to estimate puree characteristics from spectra acquired on corresponding apples  
38 such as viscosity ( $R^2 > 0.82$ ), cell wall content ( $R^2 > 0.81$ ) and also dry matter ( $R^2 >$   
39  $0.83$ ), soluble solids content ( $R^2 > 0.80$ ) and titratable acidity ( $R^2 > 0.80$ ). NIR  
40 technique should be a useful tool for industry insofar as it can give a reliable  
41 assessment of texture and taste of the final products based on the non-destructive  
42 fresh materials evaluation.

43

44 Key words: *Malus domestica* Borkh, Near infrared spectroscopy, PLS models,  
45 discriminant analyses, apples and purees

46

47 **1. Introduction**

48 Apple is one of the most widely cultivated fruits around the world (totally 68.6  
49 million tons in 2018 (USDA, 2018)), consumed both as fresh fruits and processed  
50 products. The fruit could be processed into various products to meet consumers' basic  
51 nutritious demand. Among them, apple puree has recently been reported to be a good  
52 source of polysaccharides (Le Bourvellec, Bouzerzour, Ginies, Regis, Plé, & Renard,  
53 2011) and antioxidant compounds (Loncaric, Dugalic, Mihaljevic, Jakobek, & Pilizota,  
54 2014; Oszmiański, Wolniak, Wojdyło, & Wawer, 2008). Additionally, apple purees  
55 can be used in the food industry as the basic ingredient of many fruit-based products  
56 such as jams, preserves or compotes, yogurts and pie fillings (Defernez, Kemsley, &  
57 Wilson, 1995).

58 However, the modifications of the initial physical structure, color and composition,  
59 which occur during processing, often make difficult for fruit processors to know and  
60 predict the quality characteristics of purees according to the raw apples. Qualities of  
61 apple purees depend on complex interactions between process conditions and raw  
62 material characteristics. These in turn are determined by the genetic diversity  
63 (varieties), pedoclimatic conditions, agricultural practices, maturity stages, and  
64 storage periods (Espinosa-Muñoz, Symoëaux, Reñard, Biau, & Cuvelier, 2012;  
65 Espinosa-Muñoz, To, Symoëaux, Reñard, Biau, & Cuvelier, 2011; Keenan, Brunton,  
66 Butler, Wouters, & Gormley, 2011; Picouet, Landl, Abadias, Castellari, & Viñas,  
67 2009). Apple puree manufacturers therefore encounter difficulties to maintain the  
68 expected and constant quality level of the final apple products. Until now, the research

69 studies regarding the quality assessment of apple purees have been mainly focused on  
70 the changes in polyphenol contents and total antioxidant activity (Loncaric, Dugalic,  
71 Mihaljevic, Jakobek, & Pilizota, 2014; Sukhonthara, Kaewka, & Theerakulkait, 2016),  
72 color (Oszmiański, Wolniak, Wojdyło, & Wawer, 2008), ascorbic acid (Picouet, Landl,  
73 Abadias, Castellari, & Viñas, 2009), organic acids (Bengoechea *et al.*, 1997), sugars  
74 (Keenan, Brunton, Butler, Wouters, & Gormley, 2011), polysaccharides (Le  
75 Bourvellec, Bouzerzour, Ginies, Regis, Plé, & Renard, 2011), rheological properties  
76 (Espinosa-Muñoz, Renard, Symoneaux, Biau, & Cuvelier, 2013; Espinosa-Muñoz,  
77 Symoneaux, Renard, Biau, & Cuvelier, 2012) and sensory appreciation  
78 (Espinosa-Muñoz, To, Symoneaux, Renard, Biau, & Cuvelier, 2011). However,  
79 almost all of these quality parameters have been measured through specific laboratory  
80 analyses, such as chromatography, which are time-consuming, expensive and not  
81 suitable for fast and numerous characterizations. Consequently, the development of  
82 rapid, accurate and reliable methods is required to control the quality of the raw  
83 apples and processed purees, and meet the ever-increasing demands for consistent and  
84 high quality fruit products.

85 Near infrared spectroscopy (NIRS) has been increasingly used for the safety  
86 inspection and quality assessment of agricultural products (Nicolai *et al.*, 2007). It has  
87 several advantages such as rapid spectrum acquisition, limited preparation  
88 requirements and no chemical waste, but it requires an initial calibration step, which  
89 is time consuming. Indeed, on a set of samples, representative of the expected  
90 variability, both NIRS spectra and their corresponding reference data are needed to

91 establish predictive models using multivariate statistical and mathematical data  
92 analyses. Several parameters can thus be evaluated from a single spectrum, with  
93 varying precision. Intensive investigations using NIRS have been reported regarding  
94 the measurement of apple internal attributes in the past decades (Nicolai *et al.*, 2007).  
95 Satisfactory evaluation results are reported for soluble solids contents (Peirs, Tirry,  
96 Verlinden, Darius, & Nicolai, 2003), dry matter (McGlone, Jordan, Seelye, & Clark,  
97 2003), titratable acidity (Liu & Ying, 2005), starch index (Menesatti *et al.*, 2009),  
98 chlorophyll content (Zude, Truppel, & Herold, 2002), firmness (Zude *et al.*, 2006),  
99 individual sugars (Liu, Ying, Yu, & Fu, 2006) and antioxidant capacity (Schmutzler &  
100 Huck, 2016). Further, NIRS spectra are shown to classify apples according to varieties  
101 (Luo *et al.*, 2011), geographical origins (Bobelyn *et al.*, 2010) and postharvest storage  
102 periods (Camps, Guillermin, Mauget, & Bertrand, 2007; Giovaelli, Sielli, Beghi,  
103 Guidetti, & Casiraghi, 2014).

104 Thus, NIRS can assess a diversity of quality traits in raw apples and processed  
105 products, while some of the puree characteristics are directly linked to those of the  
106 raw fruit. We therefore can suppose that NIRS spectra of the raw fruits could be used  
107 to predict the properties of the processed products, here purees, at least given a  
108 constant process operation. However, as far as we know, there is no literature related  
109 to the feasibility of using NIRS to evaluate the changes of apple puree properties and  
110 to trace back to their corresponding raw apple quality. The challenge here was to  
111 assess the possibility of predicting the properties of processed fruit products based on  
112 the raw fruit material spectral information, and so, to provide practical and suitable

113 strategies to estimate the quality potential of fruits, to monitor their processing, and to  
114 control the quality of fruit products.

115 The specific objectives of our current work were to assess the potential of NIRS to 1)  
116 detect different factors such as variety, fruit thinning, storage period and mechanical  
117 puree refining on apples and/or their corresponding purees, 2) evaluate the quality  
118 traits of interest in both apples and the corresponding purees such as  
119 textural/rheological properties, soluble solids content, titratable acidity, dry matter  
120 content, insoluble solids content 3) and then establish the links between fruit materials  
121 before and after processing.

## 122 **2. Materials and methods**

### 123 **2.1 Fruit materials**

#### 124 **2.1.1 Apples**

125 The experiment was conducted on three apple varieties: ‘Golden Smoothee’, ‘Golden  
126 Delicious’, and ‘Granny Smith’ during two subsequent harvesting seasons, 2016 and  
127 2017, that are summarized in a supplementary figure (**Suppl. Figure 1**). In 2016, 240  
128 ‘Golden Smoothee’ apples were harvested from the experimental orchard of INRA  
129 (Drôme, France). In 2017, 480 ‘Golden Delicious’ and 240 ‘Granny Smith’ apples  
130 were obtained from the experimental orchard at La Pugère (Bouches du Rhône,  
131 France).

132 Two fruit thinning levels were also compared during the ripening of ‘Golden  
133 Delicious’ apples in 2017. For this, trees were thinned 40 days after flowering (on  
134 June 2<sup>nd</sup>), and the treatment named Th+ corresponded to 50 to 100 fruits/tree (a



135 standard commercial fruit load) and Th- to 150-200 fruits/tree (highly loaded trees).  
136 The 'Golden Smoothee' in 2016 and 'Granny Smith' in 2017 grew under regular fruit  
137 thinning (Th-). After harvesting (Golden Smoothee on September 14<sup>th</sup>, 2016, 'Golden  
138 Delicious' on September 11<sup>th</sup>, 2017 and 'Granny Smith' on September 25<sup>th</sup>, 2017),  
139 apples were kept in a cold storage chamber at 4°C and at around 90% of humidity  
140 during one, three and six months (respectively T1, T3 and T6), except the first group  
141 (T0) for which apples were analyzed and processed the day after harvest. These  
142 storage durations were chosen in order to increase the fruit variability linked to  
143 firmness and biochemical changes, such as demethylation and depolymerization of  
144 pectins (Billy, Mehinagic, Royer, Renard, Arvisenet, Prost, et al., 2008). Each set (T0,  
145 T1, T3 and T6) was divided into two sub-groups. The first one was dedicated for fresh  
146 apple characterization and the second one for processing. For characterization, three  
147 replicates of 10 apples, representative of the total apple set, were analyzed after one  
148 night temperature equilibration at 23°C.

149 First, nondestructive measurements (NIR, color, ethylene releasing rate, fruit weight),  
150 and then texture tests (puncture mean load and puncture linear distance) were  
151 performed on each apple. After that, each apple was cored and divided as described by  
152 Bureau *et al.* (Bureau, Scibisz, Le Bourvellec, & Renard, 2012) in order to create, for  
153 each replicate of ten apples, three batches of 40 pieces representative of each apple.  
154 The pieces were immediately put in liquid nitrogen to avoid any oxidization. Finally,  
155 one batch was stored at -20 °C and then was subsequently freeze-dried to evaluate the  
156 alcohol insoluble solids (AIS) contents. The other two batches were stored at -80°C

157 for biochemical characterization measurement.

### 158 **2.1.2 Purees**

159 For each raw apple condition, three replicates of apple puree were produced with 4 kg  
160 of apples each. After sorting and washing, apples were cored and cut in 8 portions,  
161 then processed in a multi-functional processing system (Roboqbo, Qb8-3, Bentivoglio,  
162 Italy) following a Hot Break recipe: cooked at 95°C for 5 min at a 1500 rpm grinding  
163 speed, then cooled down to 65°C while maintaining the grinding speed. Half of the  
164 batch was refined at 0.5 mm using a Robot Coupe C80 automatic refiner (Robot  
165 Coupe SNC, Vincennes, France) in order to study two levels of granularity:  
166 non-refined (NR) and 0.5 mm refined (Ra). Finally, processed purees were  
167 conditioned in hermetically sealed cans, then placed and cooled at 23 °C before  
168 measurements, which took place the next day.

## 169 **2.2 Determination of quality traits**

### 170 **2.2.1 Color**

171 The skin color (un-blushed and blushed sides) was determined using a CR-400  
172 chromameter (Minolta, Osaka, Japan) and expressed in the CIE 1976 L\*a\*b\* color  
173 space (illuminant D65, 0° view angle, illumination area diameter 8 mm). The puree  
174 color was measured three times through a dedicated glass cuvette using the same  
175 method and equipment.

### 176 **2.2.2 Ethylene production**

177 Each group of ten apples was put in a hermetic jar at 23°C for 1 hour, and ethylene  
178 production was analyzed by taking 500 µL of the headspace and injecting it in gas

179 chromatography (Agilent, California, United States) equipped with a porapak Q  
180 column and a FID detector and expressed in  $\text{nmol kg}^{-1} \text{h}^{-1}$ .

### 181 **2.2.3 Fruit texture and puree rheology**

182 Fresh apple texture was evaluated by a puncture test using a multipurpose texture  
183 analyzer (TAPlus, Lloyd Instruments, Farenham, UK). The puncture tests were  
184 operated with a punch probe (diameter 1.2 mm), which could penetrate up to a depth  
185 of 17 mm into each peeled section of apple. Firmness was then evaluated as the mean  
186 load value calculated by the division of penetration energy by the height of testing.  
187 Crunchiness of apple flesh (Gregson & Lee, 2002) was estimated as the linear  
188 distance values from the area under the force-distance curve in the range of 10 mm at  
189 the load plateau, consisting in summing the lengths between consecutive points.

190 The puree rheological measurements were carried out using a Physica MCR-301  
191 controlled stress rheometer (Anton Paar, Graz, Austria) at 22.5 °C. The flow curves  
192 were performed after a pre-shearing period of 1 minute at  $50 \text{ s}^{-1}$  followed by 5  
193 minutes at rest. The viscosity was measured at a rate of 1 point every 15 seconds, at a  
194 controlled shear rate range of  $[10; 250] \text{ s}^{-1}$  on a logarithmic ramp. The value of the  
195 viscosity at  $100 \text{ s}^{-1}$   $\eta_{100}$  was kept as an indicator of the puree viscosity. As often used  
196 to model fruit purees (Colin-Henrion, Cuvelier, & Renard, 2007), the complete flow  
197 curves were fitted with a power-law viscosity model as described by Eq. 1.

$$198 \quad \eta = K \dot{\gamma}^{n-1} \quad (Eq1)$$

199 where  $\eta$  is the apparent viscosity (Pa.s),  $\dot{\gamma}$  the shear rate ( $\text{s}^{-1}$ ),  $K$  the consistency  
200 parameter, and  $n-1$  the flow parameter.

#### 201 **2.2.4 Biochemical characterization of apples and purees**

202 Soluble solids content (SSC) was determined with a digital refractometer (PR-101  
203 ATAGO, Norfolk, VA, USA) and expressed in °Brix at 20°C. Titratable acidity (TA)  
204 was determined by titration up to pH 8.1 with 0.1 mol/L NaOH and expressed in  
205 mmol H<sup>+</sup>/kg of fresh weight (FW) using an autotitrator (Methrom, Herisau,  
206 Switzerland). Contents of sugars (glucose, fructose, and sucrose) and malic acid were  
207 quantified using an enzymatic method with kits for food analysis (Sigma-Aldrich,  
208 Deisenhofen, Germany) and expressed in g kg<sup>-1</sup> FW. These measurements were  
209 performed with a SAFAS flx-Xenius XM spectrofluorimeter (SAFAS, Monaco). The  
210 dry matter content (DMC) was estimated with the difference between the weight  
211 values of fresh samples and of freeze-dried samples upon reaching constant weight  
212 (freeze-drier, 5 days). Content of alcohol insoluble solid (AIS) was evaluated using  
213 the method proposed by Renard (Renard, 2005) and expressed as the ratio on both  
214 fresh weight (FW) and dry matter weight (DW). Three biological replicates were  
215 obtained for each biochemical trait and each sample.

#### 216 **2.3 FT-NIR spectrum acquisition**

217 The spectral data of apples and purees were both acquired with a multi-purpose  
218 analyzer spectrometer (Bruker Optics®, Wissembourg, France), which provides  
219 diffuse reflectance measurements with a spectral resolution of 2 nm from 800 to 2500  
220 nm. For each spectrum, 32 scans were recorded and averaged. The spectral acquisition  
221 and instrument adjustments were controlled by OPUS software Version 5.0 (Bruker  
222 Optics®). The apples were placed on an automated 30-positions sample wheel, each

223 position corresponding to a measured area of 18 mm diameter. For each intact apple,  
224 two spectra were collected (on the blushed and un-blushed sides) though the 18 mm  
225 diameter areas at 23°C. Puree were transferred into 10 mL glass vials (5 cm height x  
226 18 mm diameter) which were placed on the automated sample wheel of the  
227 spectrophotometer. Each puree sample was measured three times on different aliquots.  
228 A reference background measurement was automatically activated before each data  
229 set acquisition using an internal Spectralon reference.

#### 230 **2.4 Statistical analyses and chemometrics**

231 After ensuring the normal distribution of dataset, the results were presented as mean  
232 values and the data dispersion within our experimental dataset expressed as standard  
233 deviation values (SD). Analysis of variance (ANOVA) was carried out to determine  
234 the significant differences due to the tested factors on both apples and purees using  
235 XLSTAT (version 2018.5.52037, Addionsoft SARL, Paris, France) data analysis  
236 toolbox. The pairwise comparison between means was performed using Tukey's test  
237 at the 95% level of certainty ( $p < 0.05$ (\*), 0.01 (\*\*), and 0.001 (\*\*\*)). For apples, a  
238 one-way ANOVA was applied to access the effect of storage period on 'Golden  
239 Smoothee' in 2016 and 'Granny Smith' in 2017; a two-way ANOVA concerned the  
240 effects of storage period and fruit thinning on 'Golden Delicious' in 2017. For purees,  
241 a two-way ANOVA accessed the effects of storage periods and refining treatments on  
242 'Golden Smoothee' and Granny Smith, and a three-way ANOVA for 'Golden  
243 Delicious' in terms of fruit thinning, storage periods and puree refining levels.  
244 Pearson's determination coefficients ( $R^2$ ) were calculated in order to study the

245 significance of the relationship between apples and purees and then output as an heat  
246 map using R software (version 3.5.2) (R Core Team, 2018) and the additional package  
247 named ‘ComplexHeatmap’ (Gu, Eils, & Schlesner, 2016).

248 Spectral pre-processing and multivariate data analysis were performed with Matlab  
249 7.5 (Mathworks Inc. Natick, MA) software using the SAISIR package (Bertrand &  
250 Cordella, 2008). All the NIR data were pre-processed with standard normal variate  
251 (SNV) and a derivative transform calculation (Savitzky–Golay method, gap size = 11,  
252 21, 31, 41) of first or second order. Each of the preprocessing methods was tested in  
253 the discrimination models. As SNV pre-processing had the best performances to  
254 correct multiplicative interferences and variations in baseline shift, the results shown  
255 are those obtained with the SNV pretreatment. Principal Component Analysis (PCA)  
256 and Factor Discriminant Analysis (FDA) were carried out on spectral data to evaluate  
257 the possibility to discriminate samples according to the tested factors (cultivars,  
258 thinning and storage). The specificity and sensitivity values of FDA discriminations,  
259 which help for a better evaluation of the rate of sample differentiation, were  
260 calculated by the already reported method of Nargis (Nargis *et al.*, 2019). The Partial  
261 least-square (PLS) regression method was used to develop predictive models of the  
262 quality traits of interest in apples and purees. The whole spectral dataset included 840  
263 spectra of apples and 240 spectra of purees. The dataset was randomly split, two third  
264 of dataset (560 spectra of apples and 160 spectra of purees) were used for calibration  
265 and one third of dataset (280 spectra of apples and 80 spectra of purees) for validation.  
266 The procedure was repeated 10 times in order to obtain the suitable dimensions of the

267 PLS models. The latter performance was described by the root mean square error of  
268 calibration (RMSEC), the root mean square error of validation (RMSEV), the number  
269 of latent variables (LVs), the determination coefficient ( $R^2$ ) between the predicted and  
270 measured parameters and the RPD (Residual Predictive Deviation) value as described  
271 by Nicolai (Nicolai *et al.*, 2007).

## 272 **3. Results and discussion**

### 273 **3.1 Apple and puree characteristics measured by classical methods**

#### 274 **3.1.1 Fresh apples**

275 In this experiment, three cultivars, two agricultural conditions and a cold storage for 6  
276 months provided an interesting apple fruit variability (**Figure 1a and 1b**). Clear  
277 discriminations were shown between the different storage periods along the first  
278 principal component, and apple varieties and thinning levels along the second  
279 principal component (**Figure 1a**). ‘Granny Smith’ was clearly differentiated from the  
280 two ‘Golden’ cultivars. Remarkably, the two non-thinned ‘Golden’ samples were  
281 close to each other (blue in 2016 and red in 2017), in spite of different growing  
282 seasons and locations, while the thinned samples were clearly differentiated. The most  
283 discriminant quality traits were: mean load, linear distance, AIS content (FW and  
284 DW), TA, malic acid content, ethylene production rate and color changes ( $L^*$ ,  $a^*$  and  
285  $b^*$ ) on the first principal component, and SSC, DMC and sucrose content on the  
286 second principal component (**Figure 1b**). The totality of the acquired data is presented  
287 in a supplementary table (**Suppl. Table 1**).

288 During cold storage, mean load, linear distance and AIS content (FW and DW)

289 decreased remarkably ( $p < 0.001$ ) in all apples, indicating an intensive reduction of  
290 apple firmness, crunchiness and cell wall material contents (Johnston, Hewett, &  
291 Hertog, 2002). Good correlations were observed between AIS and mean load ( $R^2$   
292 =0.78 in ‘Golden Smoothie’,  $R^2=0.75$  in ‘Granny Smith’,  $R^2 =0.82$  in ‘Golden  
293 Delicious’). The acidity in all apples decreased significantly with storage ( $p < 0.001$ )  
294 at a large range from 103.4 to 26.5 meq/kg FW for TA and 6.5 to 2.3 g/kg FW for  
295 malic acid. The ethylene production rate increased and then decreased during storage  
296 with significant changes ( $p < 0.001$ ). All color parameters ( $L^*$ ,  $a^*$  and  $b^*$ ) increased  
297 clearly for all apples, linked to a degreening and a yellowing during the long-term  
298 storage. The changes of all individual sugar contents were significant in ‘Golden  
299 Smoothie’ ( $p < 0.05$ ) and ‘Golden Delicious’ ( $p < 0.001$ ), but not in ‘Granny Smith’  
300 ( $p > 0.05$ ). For ‘Golden Delicious’, SSC and DMC from thinned trees (Th +) appeared  
301 to be significantly ( $p < 0.001$ ) higher than from non-thinned trees (Th-).

### 302 **3.1.2 Apple purees**

303 The fresh apple variability described above affected the characteristics of the  
304 corresponding non-refined (NR) purees cooked using the same recipe (**Figure 1c and**  
305 **1d**). The NR purees were discriminated according to the apple variety, fruit thinning  
306 and storage periods (**Figure 1c**). The first principal component was positively  
307 correlated to TA, content of malic acid, AIS (DW and FW) and rheological parameters  
308 ( $\eta_{100}$ ,  $K$ ), and negatively linked with colors ( $L^*$ ,  $b^*$ ) and fructose content. The storage  
309 periods could be well-classified with this component. The second principal  
310 component was highly related to DMC, sucrose content, SSC and AIS content (FW)



311 allowing the separation of varieties and fruit thinning conditions.

312 In all NR purees, clear decreases ( $p < 0.001$ ) of TA, malic acid and AIS (in DW) were  
313 observed during storage, which were highly consistent with their changes in raw  
314 apples. At the same time, the rheological properties ( $\eta_{100}$  and  $K$ ) decreased, with  
315 statistically significant differences ( $p < 0.01$ ) in all NR purees, but not in Ra purees  
316 (**Suppl. Table 2**). A good correlation was found between AIS expressed in fresh  
317 weight (FW) and the values of  $\eta_{100}$  in ‘Golden Smoothee’ ( $R^2=0.77$ ), ‘Granny Smith’  
318 ( $R^2=0.73$ ) and non-fruit thinned (Th-) ‘Golden Delicious’ ( $R^2=0.84$ ), meaning there  
319 was a good relationship between cell wall content and viscosity in NR purees.

320 Visually perceptible differences of color with an increase of  $L^*$  and  $b^*$  (with  $\Delta E > 2$ )  
321 were detectable only after 6 months of storage for ‘Golden Delicious’ and ‘Golden  
322 Smoothee’ (Hunter, & Harold, 1987). Fructose content, as the major individual sugar,  
323 increased significantly ( $p < 0.001$ ) in ‘Golden Smoothee’ and ‘Golden Delicious’  
324 purees during storage, but not for ‘Granny Smith’, again in good agreement with the  
325 behavior observed in the raw fruits.

326 The changes of SSC, sucrose content and DMC in purees were still the major  
327 discriminative contributors for apple varieties and fruit thinning conditions during  
328 puree processing. Obvious differentiations were observed for SSC and sucrose  
329 content between ‘Granny Smith’ purees and the other purees, in accordance with their  
330 changes in raw apples. In ‘Golden Delicious’, tree thinning (Th+) led to a significant  
331 increase ( $p < 0.001$ ) of DMC both in apples and their corresponding processed purees  
332 at each storage period (supplementary information, **Suppl. Table 1 and 2**), in

333 accordance with the fact that the thinned apples, in addition to being larger, also  
334 accumulates more cell materials per volume unit (Palmer, Harker, Tustin, & Johnston,  
335 2010). Additionally, the apples of the tree thinning (Th+) gave purees more viscous  
336 with significant higher values of  $\eta_{100}$  and  $K$  ( $p < 0.001$ ) than the non-thinning  
337 condition (Th-). The tree thinning treatments, by affecting individual apple growth  
338 potential, affected physical properties of raw apples and processed purees, including  
339 their viscosity. Small fruits from non-thinned trees (Th-) resulted in less viscous  
340 purees than large fruits from thinned trees.

341 Concerning the refined (Ra) purees, an expected clear reduction was obtained for both  
342 AIS content (in DW) and viscosity ( $\eta_{100}$ ) after refining (Suppl. Table 2). That could  
343 be due to the loss of insoluble fibers in the removed particle fraction (Colin-Henrion,  
344 Mehinagic, Renard, Richomme, & Jourjon, 2009) leading to a loss of puree viscosity  
345 (Espinosa-Muñoz, To, Symoneaux, Renard, Biau, & Cuvelier, 2011; Leverrier,  
346 Almeida, & Cuvelier, 2016).

### 347 **3.1.3 Relationship between the fresh apples and puree characteristics**

348 In order to study the link between physical and chemical parameters of raw apples and  
349 their processed purees, the coefficients of determination ( $R^2$ ) were calculated with the  
350 dataset including all three varieties under two thinning conditions (Th+ and Th-) and  
351 two refining levels and are displayed as heat maps for  $R^2$  values from red ( $R^2 > 0.8$ ) to  
352 blue ( $R^2 < 0.2$ ) (Figure 2). A clear similarity was observed between the two maps  
353 (Figure 2a and 2b) with the same blue and red areas. Between all apples and their  
354 processed purees, high  $R^2$  values (0.92 in NR and 0.91 in Ra) were obtained for TA.

355 Acceptable correlations were also found for SSC (0.79 in NR and 0.81 in Ra), DMC  
356 (0.72 in NR and 0.73 in Ra) and malic acid content (0.65 in NR and 0.61 in Ra). For  
357 the AIS (DW) contents, good correlations ( $R^2$ ) were obtained for each variety between  
358 raw apples and NR purees (not for Ra purees): 0.76 in ‘Golden Smoothie’, 0.83 in  
359 ‘Granny Smith’ and 0.77 in ‘Golden Delicious’, but lower when using all NR purees  
360 ( $R^2 = 0.65$ ). Moreover, acceptable correlations ( $R^2 > 0.71$ ) were obtained between  
361 texture characteristics (mean load and linear distances) of all apples and rheological  
362 parameters ( $\eta_{100}$  and  $K$ ) of their corresponding purees, whether non-refined (NR)  
363 (**Figure 2a**) or refined (Ra) (**Figure 2b**). The rheological variations in processed  
364 purees under the effects of different genotypes, storage periods and refining  
365 treatments were consistent with the textural changes in apples. However, no  
366 significant correlations ( $R^2 < 0.44$ ) were found for individual sugars (glucose, sucrose  
367 and fructose) between all apples and their purees (NR and Ra), probably because: i)  
368 these concentrations changed less during long-term storage than TA and malic acid  
369 content, and ii) water content varied during thermal processing and refining  
370 treatments.

371 The good correlations of acidity (TA and malic acid content), SSC, DMC and physical  
372 properties (textural and rheological parameters) between apples and their purees were  
373 in line with PCA results (**Figure 1b and 1d**). The chemical and physical variations in  
374 purees were shown to be potentially linked to the raw apple properties.

### 375 **3.2 Apple and puree characteristics measured by NIRS**

376 ANOVA was performed on the SNV pre-treated NIR spectra of apples and processed  
377 purees (**Figure 3**), in order to point out the wavelengths that varied during processing.  
378 According to the F-values, the variability was clearly higher for the spectra of apples  
379 (**Figure 3a and 3b**) than those of purees (**Figure 3c and 3d**), as could be expected  
380 given that each puree was prepared from 4 kg of fruit. For raw apples, the effect of  
381 variety (F-values of 800) was higher than the effect of storage (F-value of 120)  
382 (**Figures 3a and 3b**). However, after processing into purees, the opposite conclusion  
383 was obtained: the effect of storage (F-values of 110) was almost three times higher  
384 than the effect of variety (F-value of 40) (**Figure 3c and 3d**). Combine with their  
385 averaged spectral results (not shown), when apples were processed into puree, the  
386 peaks at 1930 nm in apples (**Figure 3a**) were not variable in purees (**Figure 3c**),  
387 demonstrating that the water contents had very limited variations in the purees.

### 388 **3.2.1 Discrimination of fresh apples**

389 The wavelength range with the most variability (between 1700 and 2350 nm),  
390 identified by the ANOVA (**Figure 3a**), was chosen to discriminate the effects of  
391 cultivar and storage. This range was used to perform PCA and FDA (**Figure 4a and**  
392 **4b**).

393 The first PCA displayed the discrimination of apples according to the variety (**Figure**  
394 **4a**). The first PC-score (PC1) discriminated ‘Granny Smith’ (GS) on the left and  
395 ‘Golden Delicious’ (GD) and ‘Golden Smoothie’ (GO) on the right, and accounted for  
396 83.5% of the total variability. As observed for the reference data, ‘Golden Smoothie’  
397 spectra were overlapped with those of ‘Golden delicious’. The wavelengths at around

398 1880 nm, 1930 nm and 2100-2300 nm were the main contributors of the PC1 (not  
399 shown).The two bands at 1880 nm and 1930 nm are explained by the O-H  
400 combinations, which have been reported to characterize the water content in apples  
401 (Camps, Guillermin, Mauget, & Bertrand, 2007). The broad band at 2100-2300 nm  
402 corresponds to the first combination band of C-H bond of sugars or organic acids,  
403 already used to determine the concentration of individual sugars in apple juices (León,  
404 Kelly, & Dowd, 2005; Liu, Yi, Yu, & Fu, 2006). These fingerprint wavelengths  
405 are consistent with the discrimination of apple cultivars harvested in France (Camps,  
406 Guillermin, Mauget, & Bertrand, 2007).

407 In a second step, the different storage periods of all apples could be separated (100%  
408 of discrimination sensitivity and specificity between T0 and T1 apples, and 98.5% for  
409 sensitivity and 99.5% for specificity between T1 and T3 apples) by FDA (**Figure 4b**).

410 It was observed that T3 and T6 apples were overlapped (**Figure 4b**), in line with their  
411 changes of mean load and linear distance (PCA could not well-classified storage  
412 periods). However, this result was inconsistent with previous report regarding well  
413 classification of storage periods of ‘Golden Delicious’ at 2°C by FDA (Giovanelli,  
414 Sinelli, Beghi, Guidetti, & Casiraghi, 2014). In our experiment, the strong variability  
415 and heterogeneity from different apple cultivars and fruit-thinning treatments could  
416 provide more variations of water contents and carbohydrates, and thus introduced  
417 difficulties to well classify the storage stages after 3 months (T3). The use of different  
418 storage temperatures might also be involved, with a faster evolution at 4°C than at  
419 2°C. The relevant wavelengths were mainly located in the ranges from 1700-1900 nm

420 and 2250 nm (not shown).

421 The wavelengths around 1880 nm, 1930 nm, and 2100-2300 nm could be applied to  
422 the discrimination of apple varieties, while those at 1700-1900 nm and 2250 nm could  
423 be used for the classification of apple storage periods.

### 424 **3.2.2 Discrimination of apple purees**

425 For purees, the ANOVA indicated major variations in the following wavelength  
426 ranges: 800-1050 nm, 1550-1730 nm, 1870 nm and 2100-2200 nm (**Figures 4c and**  
427 **4d**). Thus, the whole wavelength range from 800 to 2500 nm was used for FDA on the  
428 spectral dataset of all purees (not well-classified with PCA). The first two factors of  
429 the FDA allowed the discrimination of the three cultivars, ‘Golden Smoothie’ (GO),  
430 ‘Granny Smith’ (GS) and ‘Golden Delicious’ (GD), with the discrimination specificity  
431 and sensitivity values of 86.8% and 84.6% in GD and GO apples; 84.0% and 82.2%  
432 in GD and GS apples; 88.5% and 91.9% in GD and GS apples (**Figure 4c**). The F1  
433 and F2 coefficients were both highly correlated with the area between 800 and 1000  
434 nm (not shown), which is known as the absorption of apple carbohydrates and water  
435 (Giovanelli, Sinelli, Beghi, Guidetti, & Casiraghi, 2014; Zude, Herold, Roger,  
436 Bellon-Maurel, & Landahl, 2006), and already used for apple cultivar classification  
437 (Bobelyn *et al.*, 2010). Purees could be classified according to the storage periods  
438 with a distinct group for T0 and T1 (91.7% of sensitivity and 95.0% of specificity),  
439 but a mixed group for T3 and T6 (**Figure 4d**). Besides the aforementioned absorbance  
440 region between 800 and 1000 nm, the wavelengths around 1400 nm and between  
441 2100 and 2300 nm were also major contributors for discrimination of storage

442 durations. These regions have been shown to be related to water loss and SSC  
443 variations, and could be regarded as the fingerprint wavelengths of apple storage  
444 periods (Camps, Guillermina, Mauget, & Bertrán, 2007; Giovanelli, Sicelli, Beghi,  
445 Guidetti, & Casiraghi, 2014; León Kelly, & Downey, 2005).

446 Other interesting results were obtained with the FDA applied on ‘Granny Smith’  
447 purees taking into account the refining levels and the storage durations (not shown).  
448 According to the F1 and F2 axes, the two refining levels (Na and Ra) were separated  
449 both at T0 and T1, but not at T3 and T6. This result is highly consistent with the  
450 rheological changes of refined and non-refined purees of ‘Granny Smith’ (**Suppl.**  
451 **Table 2**). This refining treatment led to stronger losses of viscosity and cell wall (AIS)  
452 content before the first month of apple storage (T1), compared to purees prepared  
453 after three months (T3 and T6).

### 454 **3.3 Prediction of quality traits by NIRS**

455 In this study, we tested the ability of NIR spectra and reference data coupled with PLS  
456 to predict the physical and biochemical parameters of: (1) all apples from their NIR  
457 spectra (**Suppl. Table 3**); (2) all processed purees (NR and Ra) from their NIR spectra  
458 (**Suppl. Table 4**); (3) and all purees from the spectral information of apples (**Tables 1**  
459 **and 2**).

#### 460 **3.3.1 Prediction of quality traits by NIRS on fresh apples**

461 The prediction models were developed based on the 840 NIR spectra of apples  
462 combining 3 varieties, 2 years, 2 fruit thinning practices and 4 storage periods (**Suppl.**  
463 **Table 3**). Selected results ( $R^2 > 0.8$  obtained for the validation and RPD values  $> 2$ )

464 were further discussed.

465 The prediction results of AIS content ( $R^2=0.85$  expressed in dry weight and  $R^2=0.83$   
466 in fresh weight) during cold storage stood out in **Suppl. Table 3**. The prediction of  
467 AIS content has already been studied but using the destructive mid-infrared technique  
468 on freeze-dried apple powder (Canteri, Renard, Le Bourvellec, & Bureau, 2019), and  
469 using NIRS to predict AIS content on ‘Golden Delicious’ apples during seven months  
470 storage at  $2 \pm 0.5^\circ\text{C}$  with a  $R^2$  of 0.96 (Lovász, Merész, & Salgó, 1994). The lower  $R^2$   
471 value in our study was probably in relation with the fact that three varieties and fruit  
472 thinning conditions were introduced in the same models. For the crunchiness (linear  
473 distance), the prediction result was acceptable ( $R^2=0.82$ , RPD=2.34), in accordance  
474 with previous results obtained on ‘Golden Delicious’, ‘Braeburn’ and ‘Fuji’ apples  
475 during a 7 months cold storage ( $R^2=0.84$ ), but using the crunchiness data from  
476 sensory evaluation and the averaged NIR spectra of each group (Mehinagic *et al.*,  
477 2003). Good predictions were obtained with DMC ( $R^2=0.87$ , RPD= 2.53) and SSC  
478 ( $R^2=0.81$ , RPD= 2.21), which were in accordance with previous studies (Giovanelli,  
479 Siñelli, Beghi, Guidetti, & Casiraghi, 2014; McGloñe, Jordañ, Seelye, & Clark, 2003).  
480 Moreover, acceptable correlation coefficients were obtained for TA ( $R^2=0.80$ , RPD=  
481 2.09) and for the main organic malic acid ( $R^2=0.78$ , RPD=2.03). For the other  
482 individual sugar compounds, results were acceptable for fructose content ( $R^2=0.81$ ,  
483 RPD=1.93) and sucrose content ( $R^2=0.81$ , RPD=2.14).

484 Consequently, in apples, NIR spectroscopy was a powerful tool to qualify the  
485 crunchiness (linear distance), SSC, TA, DMC, content of individual sugars (fructose



486 and sucrose) and AIS. The benefit of AIS content prediction by NIRS was evident  
487 because the classical method of extraction and analysis needs a long time and lots of  
488 chemical solvents. Our models were also robust, given the large fruit variability used,  
489 with factors such as varieties, thinning practices and cold storage periods.

### 490 **3.3.2 Prediction of quality traits of all purees**

491 For purees, good predictions were observed for global parameters such as DMC  
492 ( $R^2=0.85$ , RPD=2.42) and SSC ( $R^2=0.92$ , RPD=3.12) (**Suppl. Table 4**). The higher  $R^2$   
493 regarding SSC in purees than in apples, could possibly due a better homogeneity of  
494 the puree samples after processing. Additionally, acceptable results were also be  
495 observed for TA ( $R^2=0.80$ , RPD=2.22). For individual compounds, results were  
496 acceptable only for fructose ( $R^2=0.83$ , RPD=2.51). For the physical properties,  
497 rheological parameters ( $\eta_{100}$ ,  $K$  and  $n$ ) and color ( $L^*$ ,  $a^*$  and  $b^*$ ), only poor results  
498 were obtained in all purees ( $R^2 < 0.51$ ). However, in ‘Granny Smith’ purees (not  
499 shown), good correlations were obtained between NIRS and  $\eta_{100}$  ( $R^2=0.94$ ,  
500 RPD=6.53), and  $K$  ( $R^2=0.93$ , RPD=3.52), and  $n$  ( $R^2=0.87$ , RPD=2.94). It seemed  
501 NIRS provided the possibility to access the evolution of rheological parameters in  
502 ‘Granny Smith’ purees from different apple storage times, but this relationship was  
503 not robust if a large variability of genotypes and agricultural practices were involved.  
504 Moreover, the results were surprising for AIS which was well-predicted in the  
505 corresponding intact apples (**Suppl. Table 3**), but not in all processed purees ( $R^2 <$   
506  $0.69$ , RPD  $< 1.59$ ). The acceptable prediction of AIS content in apples probably  
507 depended on the good correlation between AIS and textural changes (firmness and

508 crunchiness).

### 509 **3.3.3 Prediction of puree quality traits from NIR spectra of fresh apples**

510 In this part, PLS models were developed by combining spectral data acquired on fresh  
511 apples and reference data acquired on purees with two approaches: a) use the 48  
512 averaged apple spectra (means of spectra of 2 faces x 10 apples by set) and the 48  
513 reference data of their corresponding NR or Ra purees (3 replicates x 4 storage  
514 periods x 4 puree groups); b) use the 480 averaged spectra of individual apples  
515 (means of faces a and b only) and their 48 reference data of corresponding NR or Ra  
516 purees. In this case, the same values of puree characteristics were linked to the 10  
517 apples of the same set. These two methods (a and b) obtained similar prediction  
518 results and only results of the method b taking into account the apple spectra  
519 variability are shown, for both the NR purees (**Table 1**) and Ra purees (**Table 2**).

520 In NR and Ra purees, good predictions were obtained for rheological parameters  
521 ( $\eta_{100}$ ,  $K$  and  $n$ ). Especially for  $\eta_{100}$ , impressive  $R^2$  and RPD values were observed  
522 for NR purees ( $R^2=0.88$ , RPD=2.31) and Ra purees ( $R^2=0.82$ , RPD=2.44). Good  
523 results were also obtained for AIS content (expressed in FW and DW) in NR purees  
524 ( $R^2=0.81$ , RPD=2.23) and Ra purees ( $R^2=0.84$ , RPD=2.48). As the AIS content is one  
525 of the main contributors of puree viscosity (Leverrier, Almeida, & Cuvelier, 2016),  
526 these concomitant results between AIS content and rheological parameters could  
527 probably be related to their good correlations in purees. In all studied purees, good  
528 correlations were obtained between their AIS and viscosity behaviors ( $\eta_{100}$ ) ( $R^2$   
529 =0.75), but not for the AIS and SSC values ( $R^2=0.32$ ). For coloration, acceptable

530 prediction results of  $b^*$  value were obtained both in NR purees ( $R^2=0.81$ ,  $RPD=2.19$ )  
531 and Ra purees ( $R^2=0.79$ ,  $RPD=2.12$ ). Moreover, considering the DMC, SSC and TA,  
532 the PLS regression models had a good ability to estimate each characteristic for all  
533 purees on the basis of acceptable  $R^2$  and RPD values ( $R^2 > 0.80$ ,  $RPD > 2.11$ ).  
534 However, the NIR technique cannot be used to estimate satisfactorily the content of  
535 individual sugars (fructose, sucrose, glucose) and of malic acid of purees depending  
536 on the spectral information of raw apples.

537 What stands out in these results was the better predictions of some quality traits of  
538 puree from fresh apple spectra (**Table 1 and 2**) than from the puree spectra directly  
539 (**Suppl. Table 4**). It was the case for rheological parameters,  $R^2=0.82$  from apples and  
540  $R^2 < 0.44$  from purees, possibly owing to the acceptable links ( $R^2 > 0.71$ ) between  
541 apple texture (mean load and linear distance) and puree rheological properties (**Figure**  
542 **2**). In addition, better PLS results to predict AIS content from fresh apple spectra  
543 ( $R^2=0.81$  in FW and 0.84 in DW) were obtained than from purees spectra ( $R^2 < 0.69$ )  
544 (**Suppl. Table 4**), probably due to good relationships between puree viscosity and AIS  
545 content mentioned above. Besides, the prediction of DMC, SSC and TA were still  
546 acceptable in all cases. Therefore, NIR technique showed a potential to directly  
547 predict the viscosity properties,  $b^*$ , AIS content, SSC, DMC and TA of processed  
548 purees using their corresponding apple spectral information directly.

549 Compared with the PLS models used to predict the characteristics of purees based on  
550 their own spectra (**Suppl. Tables 3 and 4**), more LVs and lower prediction accuracy  
551 (RPD values) have generally found when models were built using the spectra of the

552 intact apples to predict the characteristics of the processed purees. This fact has been  
553 also observed when other raw materials, e.g. meat (Meullenet, Jonville, Grezes, &  
554 Owens, 2004) or whole grain (Windham et al., 1997) were used to predict quality  
555 traits of the final cooked food. Such indirect prediction is a challenge as the spectra  
556 were not acquired on the material for which prediction was done, and because the  
557 chemical and textural traits from the material on which the spectra were acquired (the  
558 raw apples) are modified by processing. However, such predictions, albeit only  
559 semi-quantitative, are relevant for industrial use. Indeed, these developed models  
560 provide a promising solution to evaluate puree viscosity, a primary quality trait of this  
561 product, and cell wall contents (AIS), only based on spectra of fresh apples. A  
562 remarkable fact was that these predictions could not be done using the NIR spectra of  
563 the purees themselves (Suppl. Table 4).

#### 564 **4. Conclusion**

565 As far as we know, this was the first report concerning the assessment of quality  
566 variation of apple purees depending on NIR spectral information of the corresponding  
567 raw apples. Up to now, in apple industry, manufacturers use their experience and  
568 knowhow to make blend of apples in order to obtain always the same puree. From our  
569 results, NIR had the potential to predict internal quality of apples but also that of their  
570 processed products: a reliable assessment of texture and taste of the purees could be  
571 obtained based only on spectral data of fresh apples. This opens the possibility to sort  
572 or select apples according to the expected purees. By systematically scanning all  
573 apples, this could provide some objective data to predict the final product

574 characteristics and thus reduce waste of materials along the processing chain. Further  
575 work will be needed to investigate the interaction of the processing conditions  
576 (temperature, time, oxygen and so on) with raw apples under various growing and  
577 storage conditions, so as to provide guidance for adapted processing procedures to  
578 reach stable final puree qualities.

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711 **Figure captions**

712 **Figure 1.** Principal Component Analysis of physical, physiological and biochemical  
713 compositions for: apples **(a)** and processed NR (no refined) purees **(c)** in three  
714 varieties (GO: ‘Golden Smoothee’ , GD: ‘Golden Delicious’ and GS: Granny Smith)  
715 growing under two different thinning conditions (Th+ marked with red solid circle  
716 and Th- with red dotted circle) and stored at 4°C from harvest (T0), 1 (T1), 3 (T3) and  
717 6 (T6) months. Correlation plot of the first principal components for apples **(b)** and  
718 NR purees **(d)**.

719 **Figure 2.** Determination coefficient ( $R^2$ ) of all physical and biochemical parameters  
720 between the ‘Golden Smoothee’, ‘Granny Smith’ and ‘Golden Delicious’ apples  
721 (titled with “F”) and their processed purees (titled with “P”): **(a)** raw apples and  
722 non-refined (NR) purees; **(b)** raw apples and refined (Ra) purees.

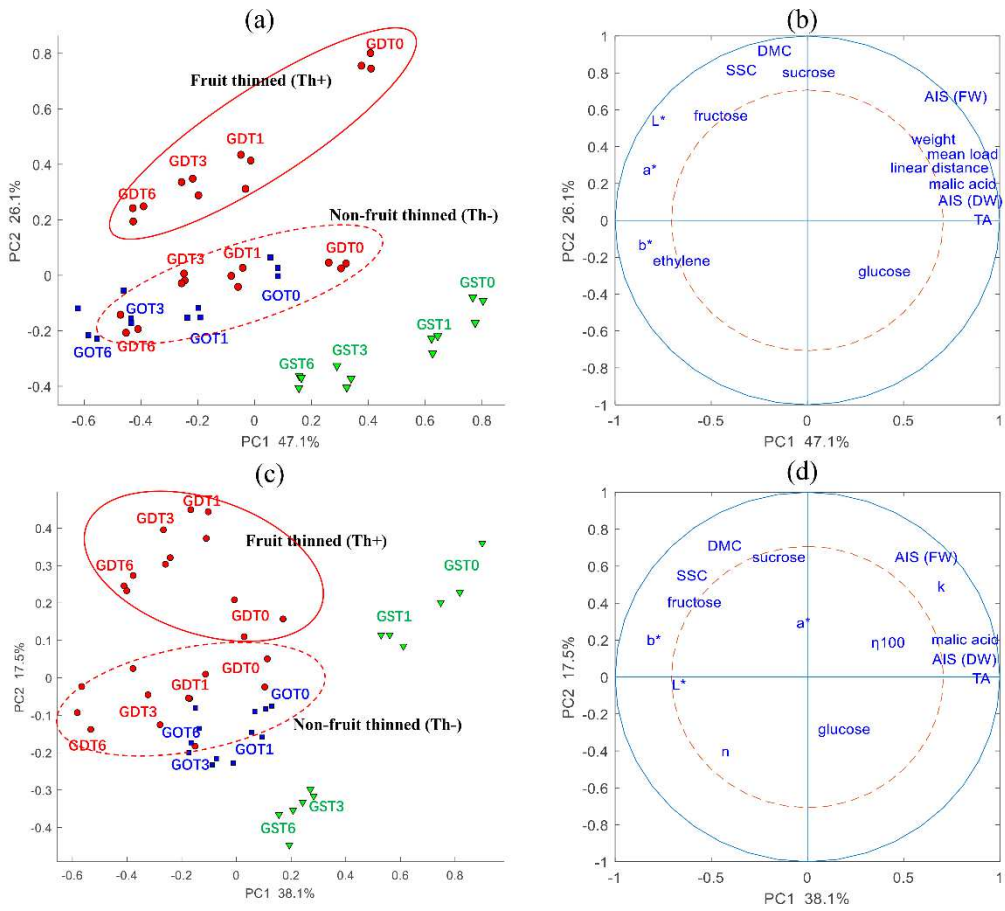
723 **Figure 3.** ANOVA results of the SNV pre-treated NIR spectra between 800 and 2500  
724 nm: **(a)** effect of variety on all apples spectra; **(b)** effect of storage period on all apples  
725 spectra; **(c)** effect of variety on all purees spectra; **(d)** effect of storage period on all  
726 purees spectra;

727 **Figure 4.** Factorial maps of the SNV pre-treated NIR spectra of all apples between  
728 1700 and 2350 nm or all purees between 800 and 2500 nm: **(a)** Principal Component  
729 Analysis showing apples cultivars; **(b)** the Factorial Discriminant Analysis of storage  
730 periods of apples; **(c)** Factorial Discriminant Analysis of cultivars of all purees; **(d)**  
731 Factorial Discriminant Analysis of storage periods of all purees.

732 ‘Golden Smoothee’ (GO), ‘Golden Delicious’ (GD), ‘Granny Smith’ (GS); storage

733 duration at 4°C from harvest (T0), 1 (T1), 3 (T3) and 6 (T6) months.

734 **Figures**



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**Figure 1**

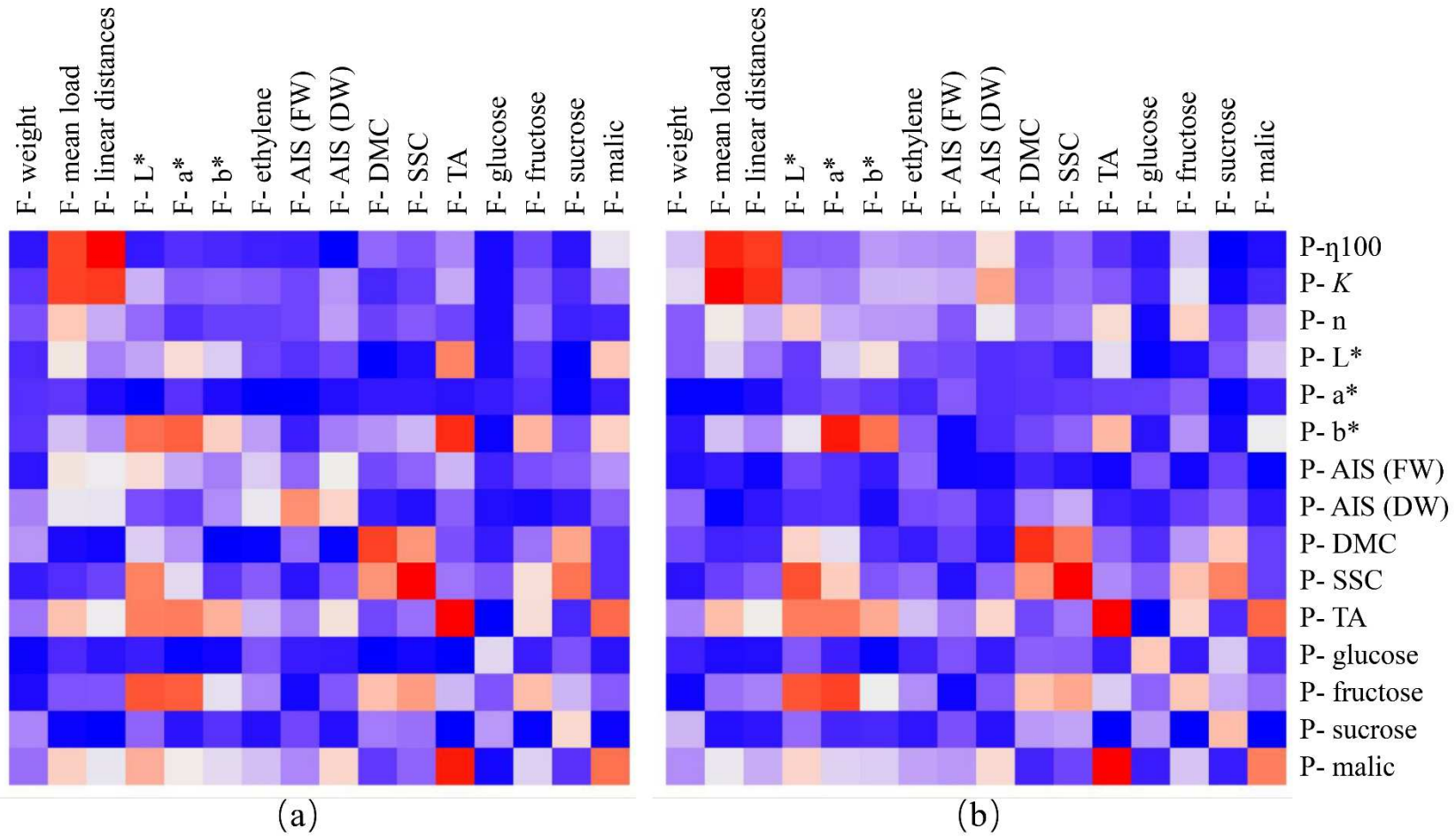
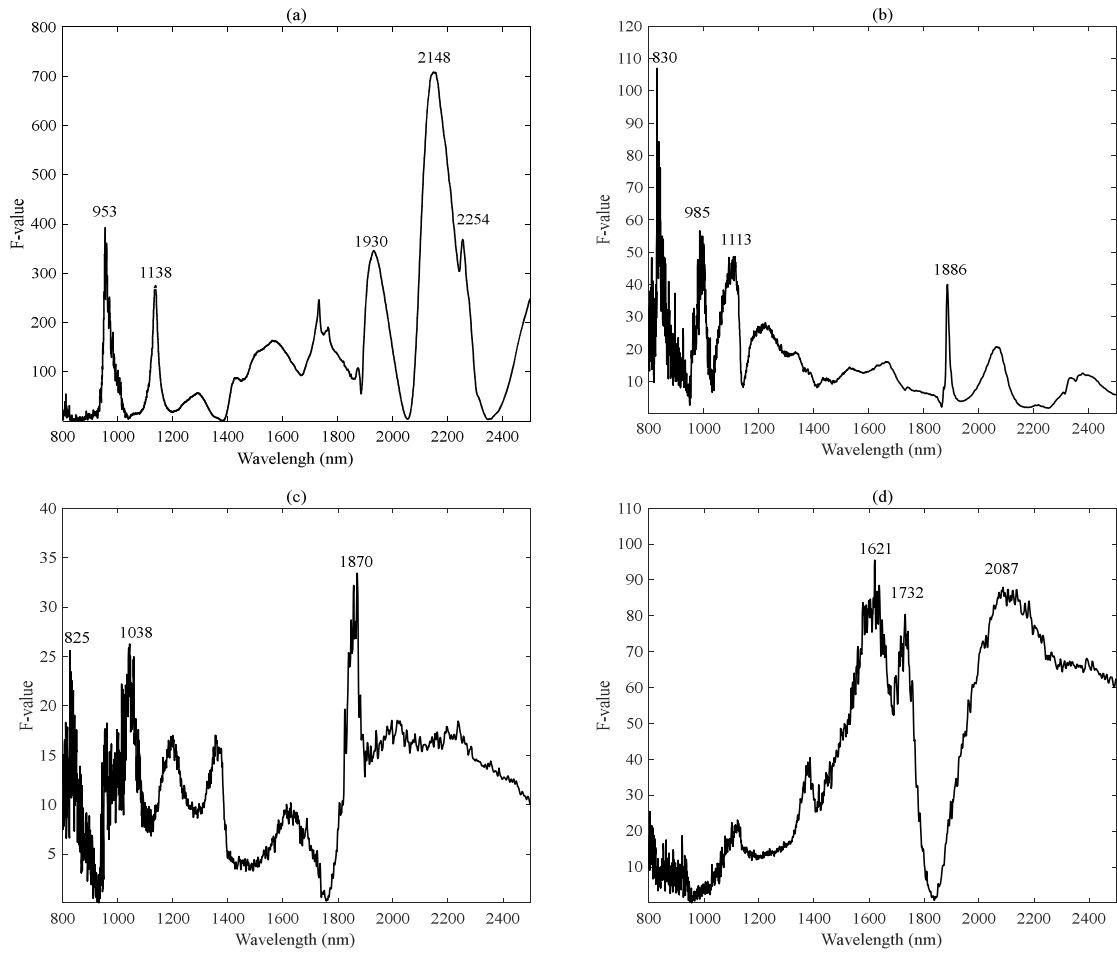
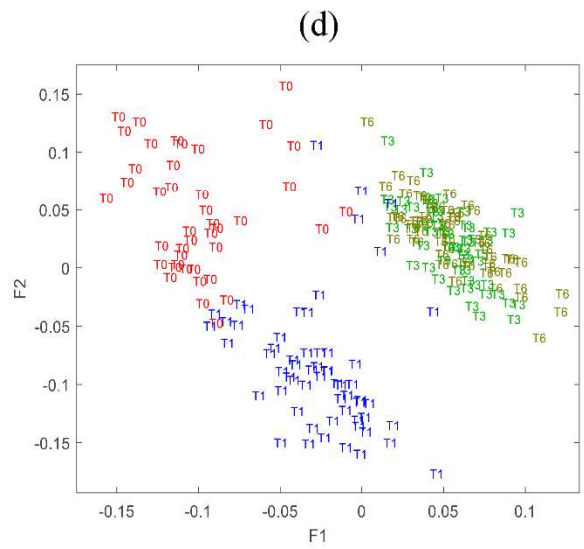
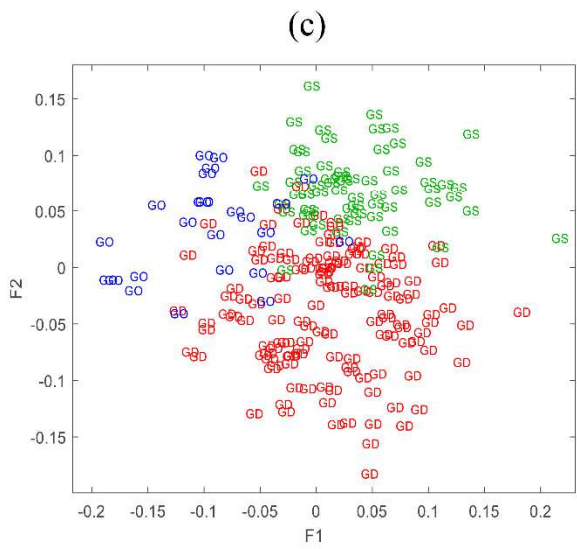
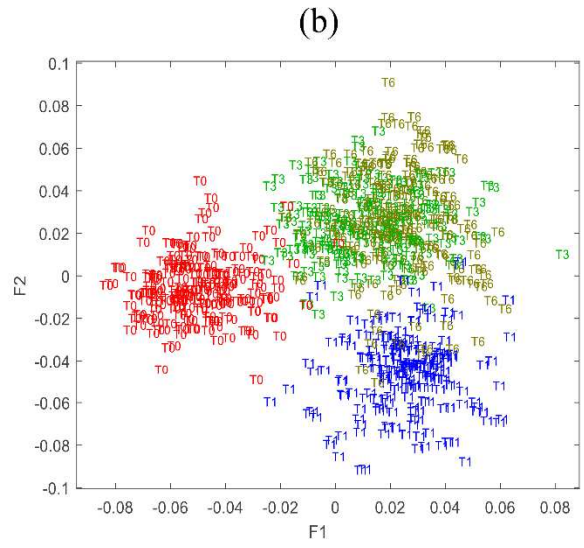
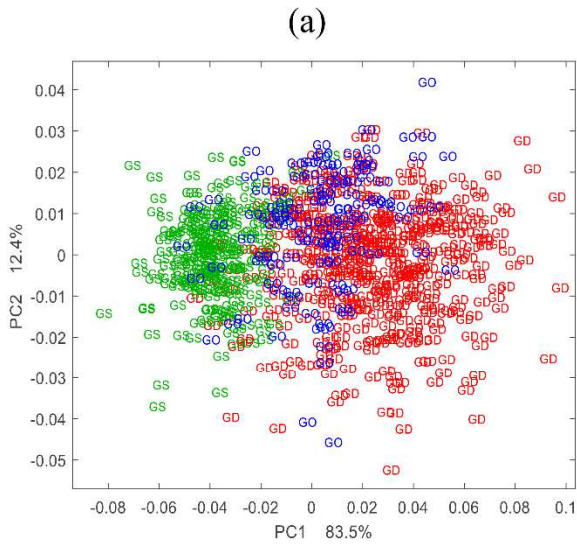


Figure 2



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**Figure 3**



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Figure 4

746 **Table 1.** Prediction of puree quality traits from spectral data of fresh apples: PLS (Partial Least Squares) results using NIR spectra of fresh  
747 apples from three varieties ('Granny Smith', 'Golden Delicious' and 'Golden Smoothee'), two thinning conditions and two harvest seasons, at 4  
748 cold storage durations (0, 1, 3 and 6 months) for prediction of quality traits of non-refined (NR) purees.

Parameter	range	SD	Calibration n=320		Validation n=160		LVs	RPD	spectral range(nm)
			r <sup>2</sup>	RMSEC	R <sup>2</sup>	RMSEV			
CSR ( $\eta_{100}$ )	0.49-1.45	0.19	0.89	0.06	0.88	0.08	10	2.31	900- 2500
viscosity-K	16.8-59.5	11.24	0.79	5.0	0.79	5.2	12	2.15	900- 2500
viscosity-n value	0.06-0.39	0.07	0.83	0.03	0.82	0.03	12	2.06	900- 2500
L*	44.0-53.5	1.9	0.81	0.8	0.81	1.1	10	1.77	900- 2500
a*	-(5.0-3.4)	0.3	0.48	0.3	0.46	0.3	10	1.36	900- 2500
b*	9.2-23.0	3.5	0.84	1.4	0.81	1.6	12	2.19	900- 2500
AIS (mg/g DW)	114.0-171.7	32.6	0.82	16.4	0.80	16.3	11	2.00	900- 2500
AIS (mg/g FW)	19.3-48.9	5.8	0.82	2.5	0.81	2.6	13	2.23	900- 2500
DMC (g/g FW)	0.16-0.23	0.02	0.84	0.01	0.83	0.01	12	2.11	900- 2500
SSC (°Brix)	10.5-18.6	2.2	0.85	0.9	0.80	1.0	11	2.25	900- 2500
TA (meq/kg FW)	25.0-103.9	20.0	0.83	8.3	0.80	9.4	11	2.14	900- 2500
glucose (g/kg FW)	13.5-25.4	3.1	0.65	1.8	0.60	2.0	10	1.55	900- 2500
fructose (g/kg FW)	40.0-98.7	17.2	0.80	7.6	0.73	9.1	11	1.90	900- 2500
sucrose (g/kg FW)	32.2-118.5	22.0	0.80	9.6	0.76	11.1	11	1.98	900- 2500
malic (g/kg FW)	2.4-9.0	1.5	0.77	0.7	0.76	0.8	10	1.91	900- 2500

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750

751 **Table 2.** Prediction of puree quality traits from spectral data of fresh apples: PLS (Partial Least Squares) results using NIR spectra of fresh  
752 apples from three varieties ('Granny Smith', 'Golden Delicious' and 'Golden Smoothie'), two thinning conditions and two harvest seasons, at 4  
753 cold storage durations (0, 1, 3 and 6 months) for prediction of quality traits of refined (Ra) purees.

T Parameter	range	SD	Calibration n=320		Validation n=160		LVs	RPD	spectral range(nm)
			r <sup>2</sup>	RMSEC	R <sup>2</sup>	RMSEV			
CSR ( $\eta_{100}$ )	0.25-0.99	0.18	0.89	0.06	0.82	0.07	11	2.44	900- 2500
viscosity-K	2.8-39.1	8.81	0.87	3.0	0.86	3.7	10	2.40	900- 2500
viscosity-n value	0.17-0.49	0.07	0.85	0.03	0.82	0.03	13	2.11	900- 2500
L*	43.9-53.8	2.2	0.6	1.3	0.61	1.4	10	1.54	900- 2500
a*	-(5.2-3.6)	0.4	0.5	0.3	0.51	0.4	8	1.04	900- 2500
b*	7.8-22.9	3.4	0.8	1.6	0.79	1.6	12	2.12	900- 2500
AIS (mg/g DW)	90.9-189.6	18.3	0.79	8.4	0.76	8.9	12	2.05	900- 2500
AIS (mg/g FW)	14.8-33.3	4.1	0.85	1.6	0.84	1.6	12	2.48	900- 2500
DMC (g/g FW)	0.15-0.24	0.02	0.84	0.01	0.84	0.01	12	2.37	900- 2500
SSC (°Brix)	10.3-17.6	2.0	0.82	0.9	0.80	0.9	11	2.16	900- 2500
TA (meq/kg FW)	25.2-109.1	21.6	0.83	8.9	0.80	9.6	11	2.26	900- 2500
glucose (g/kg FW)	13.9-25.7	3.1	0.65	1.8	0.62	2.1	10	1.50	900- 2500
fructose (g/kg FW)	42.1-99.9	15.6	0.71	9.5	0.70	10.0	10	1.56	900- 2500
sucrose (g/kg FW)	33.4-123.1	23.5	0.78	11.0	0.76	11.0	11	2.14	900- 2500
malic (g/kg FW)	2.9-8.3	1.5	0.79	0.7	0.75	0.8	10	1.93	900- 2500

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