An efficient protocol for functional studies of apple transcription factors using a glucocorticoid receptor fusion system

Joan Estevan, Sara Gómez-jiménez, Vítor da Silveira Falavigna, Alicia Camuel, Lisa Planel, Evelyne Costes, Fernando Andrés

To cite this version:
Joan Estevan, Sara Gómez-jiménez, Vítor da Silveira Falavigna, Alicia Camuel, Lisa Planel, et al.. An efficient protocol for functional studies of apple transcription factors using a glucocorticoid receptor fusion system. Applications in Plant Sciences, Wiley, 2020, 8 (10), 10.1002/aps3.11396. hal-03025007

HAL Id: hal-03025007
https://hal.inrae.fr/hal-03025007
Submitted on 26 Nov 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L’archive ouverte pluridisciplinaire HAL, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d’enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Distributed under a Creative Commons Attribution 4.0 International License
An efficient protocol for functional studies of apple transcription factors using a glucocorticoid receptor fusion system

Joan Estevan1, Sara Gómez-Jiménez1, Vítor da Silveira Falavigna1,2, Alicia Camuel1, Lisa Planel1, Evelyne Costes1,3, and Fernando Andrés1,3

PREMISE: We report a protocol for studying the function of apple (Malus × domestica) transcription factors based on the glucocorticoid receptor (GR) system, which allows the dexamethasone (DEX)-mediated activation of plant transcription factors to monitor the expression levels of their potential target genes.

METHODS AND RESULTS: Apple leaves are transformed with a vector that allows the expression of the studied transcription factor (i.e., FLOWERING LOCUS C [MdFLC]) fused to GR. Calli derived from the transformed leaves are treated with DEX and cycloheximide, a protein synthesis inhibitor. Compared with other methods, combining the GR system with cycloheximide treatments enables the differentiation between direct and indirect transcription factor target genes. Finally, the expression levels of putative MdFLC target genes are quantified using quantitative reverse transcription PCR.

CONCLUSIONS: We demonstrate the efficiency of our GR system to study the function of apple transcription factors. This method is accessible to any laboratory familiar with basic molecular cloning procedures and the apple leaf–mediated agro-transformation technique.

KEYWORDS apple; GR system; MdFLC; target gene identification; transcription factor.
of TFs on the transcriptional regulation of their target genes. As a proof of concept, we investigated the effect of the TF MdFLC on the expression of the putative target genes identified in our DAP-seq experiment. To this end, we transformed apple calli with a vector expressing MdFLC TF fused to the GR protein and used quantitative reverse transcription PCR (RT-qPCR) to monitor the effect of the target gene induction by a DEX treatment. These experiments were performed in the presence of the protein synthesis inhibitor cycloheximide, favoring the detection of direct target genes. Our results indicate that the TF-GR protocol presented here is an efficient and reliable method to evaluate the MdFLC TF target genes, and is therefore a valuable tool in gene functional studies.

METHODS AND RESULTS

Vector construction and transformation in Agrobacterium tumefaciens

The complete coding sequence of MdFLC (MD09G1009100) cloned into the pENTR/D-TOPO vector was provided by Dr. Luís F. Revers (Embrapa Uva e Vinho, Bento Gonçalves, Brazil). Gene-specific primers (Table 1) were used to amplify the coding sequence of MdFLC without the stop codon using KOD Hot Start DNA Polymerase (Merck, Darmstadt, Germany). The PCR protocol consisted of one step at 95°C for 2 min, followed by 40 cycles at 95°C for 20 s, 60°C for 10 s, and 70°C for 10 s. The PCR product was visualized in a 1% agarose gel and purified using a Zymoclean Gel DNA Recovery Kit (Zymo Research, Irvine, California, USA). The purified PCR product was cloned into the pDONR207 vector (Karimi et al., 2007) using Gateway BP Clonas II Enzyme Mix (Thermo Fisher Scientific, Waltham, Massachusetts, USA), according to the manufacturer's instructions. This construct was recombined into the pDONR207 vector (Karimi et al., 2007) using Gateway BP Clonas II Enzyme Mix (Thermo Fisher Scientific, Waltham, Massachusetts, USA), according to the manufacturer's instructions. This construct was recombined into the pDONR207 vector (Karimi et al., 2013), provided by Dr. Gloria Coruzzi (New York University, New York, New York, USA), using Gateway LR Clonas II Enzyme Mix (Thermo Fisher Scientific). The MdFLC-GR sequence was amplified and purified as described above using the primers FLC-Fw-BP and GR-Rv2-BP (Table 1). The purified MdFLC-GR PCR product was cloned into the pDONR207 vector as described above. The resulting vector was recombined into the binary vector pCamway35S (Leclercq et al., 2015), which carries the in planta selection marker 35S::GFP (GREEN FLUORESCENT PROTEIN) and an additional constitutive-expression promoter CaMV 35S to drive the expression of MdFLC-GR. The structure of the final vector (pCamway35-MdFLC-GR) is shown in Fig. 1A. The vectors resulting from each different cloning step were confirmed by sequencing. The final expression vector harboring the 35S::MdFLC-GR cassette was used in the transformation of the A. tumefaciens strain EHA105 (Hood et al., 1993).

Production of transformed calli from leaf explants

The apple cultivar 'Gala' was used in this study. In vitro cuttings of apple were subcultured at one-month intervals on a micropropagation medium, a modified Lepoivre medium (Leblay et al., 1991) supplemented with 0.5 mg/L 6-benzylaminopurine and 0.1 mg/L indole-3-butyric acid, at 22°C under a 16-h light (80 µmol m−2 s−1)/8-h dark photoperiod. Previously described apple leaf transformation protocols (James et al., 1993) were modified and used to produce transformed calli that could be easily stored and multiplied for TF functional studies. Transformed A. tumefaciens was grown for 48 h on a Luria–Bertani solid medium containing 50 µg/mL kanamycin. The bacteria inoculum at OD600 = 1 was resuspended in an induction medium containing 150 µM acetosyringone (James et al., 1993) and incubated at room temperature with gentle agitation for 3–4 h. The day before the transformation, the youngest leaves of four-week-old micropropagated shoots were harvested (from auxillary shoot cultures) and incubated in the dark on a regeneration medium, which was a modified Lepoivre medium (see above) supplemented with 5 mg/L thidiazuron and 0.2 mg/L a-naphtalene acetic acid. The leaves were wounded perpendicularly to the midrib using a scalpel (two or three internal sections) and vacuum-infiltrated for 1 min with the

---

**TABLE 1.** List of primers used in this study.

<table>
<thead>
<tr>
<th>Purpose</th>
<th>Primer name</th>
<th>Accession code</th>
<th>Primer sequence (5′–3′)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Molecular cloning</td>
<td>FLC-Fw-BP</td>
<td>MD09G1009100</td>
<td>GGGGACAAGTTTGTACAAAAAAGCAGGCTCTAT</td>
</tr>
<tr>
<td></td>
<td>FLC-Rv-BP-nostop2</td>
<td>MD09G1009100</td>
<td>GGGGCAGGGAAGGTCCAG</td>
</tr>
<tr>
<td></td>
<td>MdFLC-like-F</td>
<td>MD09G1009100</td>
<td>GGGGACCACTTTGACTCAAGAAAGCTGGGTTAACAC</td>
</tr>
<tr>
<td></td>
<td>MdFLC-like-R</td>
<td>MD09G1009100</td>
<td>AACCTGATATGGTGGCCG</td>
</tr>
<tr>
<td></td>
<td>MDH-F</td>
<td>MD16G1219000</td>
<td>GGGGACCACTTTGACTCAAGAAAGCTGGGTTAG</td>
</tr>
<tr>
<td></td>
<td>MDH-R</td>
<td>MD16G1219000</td>
<td>ATGCGCGGTTTTTGTAG</td>
</tr>
<tr>
<td></td>
<td>MDH-R</td>
<td>MD16G1219000</td>
<td>TGCCAATTACCTTCTTTCTG</td>
</tr>
<tr>
<td></td>
<td>HAI3-F</td>
<td>MD01G1220800</td>
<td>TGGCAGTAGTCAGGGAATGA</td>
</tr>
<tr>
<td></td>
<td>HAI3-R</td>
<td>MD01G1220800</td>
<td>TGGCAGTAGTCAGGGAATGA</td>
</tr>
<tr>
<td></td>
<td>FUL-F</td>
<td>MD09G1074000</td>
<td>ACCCTGACCTTTGACTCAAGAAAGCTGGGTTAG</td>
</tr>
<tr>
<td></td>
<td>FUL-R</td>
<td>MD09G1074000</td>
<td>GGAGGAGTGGATTGCTCAAA</td>
</tr>
<tr>
<td></td>
<td>ABA2-F</td>
<td>MD07G1033200</td>
<td>CCGGTTGCCGTTCCAAA</td>
</tr>
<tr>
<td></td>
<td>ABA2-R</td>
<td>MD07G1033200</td>
<td>GGGGAGGATGGATTGCTCAAA</td>
</tr>
<tr>
<td></td>
<td>GASA4-F</td>
<td>MD17G1041500</td>
<td>CATGGGACAGGGAAGGGCAT</td>
</tr>
<tr>
<td></td>
<td>GASA4-R</td>
<td>MD17G1041500</td>
<td>GGCACACAGGGAAGGGCAT</td>
</tr>
</tbody>
</table>

a Nomenclature of accession codes from the apple genome GDDH13 version 1.1 (Daccord et al., 2017).
inoculum at −0.09 mPa. The inoculated leaves were placed abaxial surface up on a co-cultivation medium (regeneration medium with the addition of 100 μM acetosyringone), and the plates were incubated in the dark at 22°C for 48 h. After co-cultivation, the leaves were transferred, adaxial surface up, onto a selection/callusogenesis medium, a modified Lepoivre medium (see above) supplemented with 0.5 mg/L of kinetin, 0.5 mg/L 2,4-dichlorophenoxyacetic acid, 50 mg/L kanamycin, 300 mg/L cefotaxime, and 150 mg/L ticarcillin disodium, before being placed in darkness at 22°C. One month after transformation, the putatively transformed calli that proliferate in the presence of antibiotics were observed under a fluorescence stereo zoom microscope (Leica MZFLIII; Leica Camera AG, Wetzlar, Germany) (Fig. 1B). The GFP-expressing calli were further selected and subcultured into the selection medium with three-week intervals in the dark at 22°C. Six months after the leaf transformation, about 30 transformed calli (approximatively 5 mm in diameter) were obtained.

**Chemical induction of expression**

Successfully transformed calli were subjected to different treatments, while non-transformed calli of a similar age were used as wild-type controls. Each sample comprised a pool of 10 calli. One wild-type and one transformed sample (T0) of calli were frozen in liquid nitrogen at the beginning of the assay (no treatment) and stored at −80°C. The other calli samples were treated with DEX to induce the nuclear accumulation of MdFLC TF, as described elsewhere (Schena and Yamamoto, 1988; Bargmann et al., 2013). A list of required materials and reagents is provided in Appendix 1. Transformed calli were pretreated with 40 μM cycloheximide for 30 min and then rinsed with distilled water. Subsequently, 10 μM DEX was added to the treated samples (+DEX) and incubated at room temperature for 1 h before being rinsed with distilled water and stored at room temperature for 3 (T4) or 7 (T8) h. Negative controls (−DEX) were treated with solvent alone (i.e., the 95% ethanol used to dissolve DEX). Calli treated with DEX or the solvent were collected and frozen at −80°C.

**Expression studies**

Total RNA was isolated using a commercial kit (Spectrum Plant Total RNA kit; Sigma-Aldrich, St. Louis, Missouri, USA) following the manufacturer’s instructions. The integrity of the total RNA was analyzed using a 4200 TapeStation instrument (Agilent Technologies, Santa Clara, California, USA), while the RNA quantification was performed with a NanoQuant instrument (Infinite 200 NanoQuant; Tecan, Männedorf, Switzerland). The RNA was subjected to a DNase treatment using a Turbo DNA-free kit (Thermo Fisher Scientific), and cDNA was synthesized from the RNA template using a Superscript III Reverse Transcriptase kit (Thermo Fisher Scientific), following the manufacturer’s instructions.

For the RT-qPCR reactions, 2 μL of the cDNA samples (diluted 1 : 10) was used as a template in a 6-μL final reaction volume containing 3 μL of 2× LightCycler 480 SYBR Green I Master mix (Roche, Basel, Switzerland) and 3 μM of each primer. The real-time PCR reactions were run on a LightCycler 480 (Roche) with an initial denaturation step of 5 min at 95°C followed by 40 cycles of 20 s at 95°C, 20 s at 60°C, and 20 s at 72°C. The PCR products were analyzed in a melting curve analysis to verify the presence of a gene-specific PCR product. The melting curve analysis was performed immediately after the PCR amplification using a single step at 95°C for 1 min, 40°C for 1 min, and an annealing procedure starting at 65°C and increasing to 95°C at increments of 0.02°C/s. Each reaction included negative and positive controls and each cDNA sample was analyzed in three technical replicates. In a pilot experiment, two housekeeping genes—WD-40 repeat family protein (WD40) and malate dehydrogenase (MDH)—were used to normalize the amount of plant RNA in each sample (Perini et al., 2014). The two housekeeping genes gave a very similar result, and thus only one of them, WD40, was used in further experiments (Appendix S1). The putative MdFLC TF target genes analyzed here were previously identified using DAP-seq (Falavigna et al., unpublished data). The transcript levels were calculated with LightCycler 480 software version 1.5.0.39 (Roche), and the efficiency of the primers was determined using LinRegPCR (Ruijter et al., 2009). The ΔΔCt method (Livak and Schmittgen, 2001) was used to analyze the data. The list of primers used to quantify the expression of the genes of interest is provided in Table 1. A Student’s t-test was performed to estimate the significance of the difference between the +DEX and −DEX treatments at each timepoint.

The level of MdFLC mRNA expression in the calli samples was quantified to confirm the efficiency of its overexpression driven by the 35S promoter. As shown in Fig. 2A, the expression levels of MdFLC were significantly higher in all transgenic calli compared
with the control (non-transformed) calli. The transcript levels of four potential target genes of MdFLC—HIGHLY ABA-INDUCED PP2C GENE 3 (HAI3), FRUITFULL (FUL), ABA DEFICIENT 2 (ABA2), and GAST1 PROTEIN HOMOLOG 4 (GASA4)—were studied using RT-qPCR over two timepoints (4 h [T4] and 8 h [T8]) in the mock (−DEX) or DEX-treated transformed calli (Fig. 2B, Appendix S1). The reference gene MDH (Perini et al., 2014) was used as a negative control (it is not bound by MdFLC TF). Notably, the expression levels of the four tested putative MdFLC TF target genes were altered by the DEX treatments in at least one of the assayed timepoints. The expression levels of HAI3 and FUL were significantly higher in the +DEX samples compared with the −DEX samples at T8. On the other hand, the levels of ABA2 expression at the two timepoints (T4 and T8) and GASA4 at T4 were significantly downregulated in the +DEX samples compared with the −DEX treatment. The expression levels of the negative control gene MDH did not show a statistical difference at any of the assayed timepoints.

CONCLUSIONS

Here, we report a protocol for functional genomics studies that facilitates the quantification of the expression of TF target genes using a GR system. This protocol has been demonstrated to be efficient and sensitive for the detection of changes in the expression of MdFLC TF target genes.

The putative MdFLC TF target genes interrogated in this study were previously identified using DAP-seq (Falavigna et al., unpublished); however, in addition to the analysis of known TF target genes, this system can be applied to discover novel TF target genes in combination with RNA-seq or ChIP-seq techniques. Our system allows the rapid and timely TF activation by DEX, and thus enables the monitoring of the dynamic transcriptional changes of the target genes using time-course experiments. This feature is instrumental for selecting the optimal conditions for transcriptomic experiments; for example, we observed that the expression levels of some MdFLC target genes were only significantly affected at one particular timepoint after DEX treatment (Fig. 2B). By contrast, other methods used in the study of apple TF function using ectopic expression protocols do not allow the fine-tuning of TF activity and therefore might not be efficient in detecting genome-wide TF target genes. Additionally, by making use of the translational inhibitor cycloheximide, it is possible to distinguish between direct and indirect TF target genes, which is not likely by relying solely on constitutive expression systems.

In conclusion, we have developed a protocol for functional studies of apple that can be easily performed in any laboratory familiar with basic molecular cloning procedures and the apple leaf-mediated agro-transformation technique. Coupled with genome-wide approaches, this protocol can be instrumental for unraveling the molecular function of apple TFs and for deciphering the gene regulatory networks in which they are involved.

ACKNOWLEDGMENTS

This project was supported by the Agropolis Fondation under the reference ID 1702-023 through the Investissements d’avenir program (Labex Agro: ANR-10-LABX-0001-01), under the frame of
I-SITE MUSE (ANR-16-IDEX-0006). V.S.F. received a grant from the AgreenSkills+ EU fellowship program (FP7-609398). The authors thank Dr. Luis F. Revers (Embrapa Uva e Vinho, Bento Gonçalves, Brazil), Dr. Gloria Coruzzi (New York University, New York, New York, USA), and Dr. Julie Leclercq (AGAP, University of Montpellier, Montpellier, France) for generously providing materials, and Dr. Gabriel Krouk (Biochemistry and Plant Molecular Physiology [BPMP], Montpellier, France) and Dr. Elisabeth Chevreau (Research Institute of Horticulture and Seeds [IRHS], Angers, France) for advising on the GR system and apple genetic transformation, respectively.

**AUTHOR CONTRIBUTIONS**

J.E., S.G.-J., and A.C. performed the genetic transformation experiments. V.S.F. performed the molecular cloning and vector construction. J.E. and L.P. performed the gene expression studies and analyzed the results. J.E. and F.A. conceived the protocol and designed the experiments. J.E., V.S.F., E.C., and F.A. wrote the manuscript. All the authors approved the final version.

**DATA AVAILABILITY**

The authors confirm that all data underlying the findings are fully available without restriction. All data are included within the manuscript and Appendix S1.

**SUPPORTING INFORMATION**

Additional Supporting Information may be found online in the supporting information tab for this article.

**APPENDIX S1.** Pilot experiment and gene expression levels in transformed apple calli upon treatment with dexamethasone (DEX).

**LITERATURE CITED**


**APPENDIX 1.** Materials and reagents required to perform the chemical induction of MdFLC transcription factor nuclear accumulation.

**Chemicals/solutions**

- 95% ethanol
- Dimethylsulfoxide (DMSO)
- Liquid nitrogen
- Distilled water
- Dexamethasone (DEX) (catalog no.: D4902; Sigma-Aldrich, St. Louis, Missouri, USA): 1 mM DEX stock was dissolved in 95% ethanol and stored at −20°C
- Cycloheximide (catalog no.: C4859; Sigma-Aldrich, St. Louis, Missouri, USA): a 50 mM cycloheximide stock was dissolved in DMSO and stored at −20°C

**Equipment**

- Tweezers
- 10 µL and 100 µL micropipettes and tips
- 1.5 mL microcentrifuge tubes (catalog no.: 87003-294; VWR International, Radnor, Pennsylvania, USA)
- −80°C freezer
- Fume hood