



**HAL**  
open science

## Rumen function in goats, an example of adaptive capacity

Sylvie Giger-Reverdin, Céline Domange, Laurent Broudiscou, Daniel Sauvant,  
Valérie Berthelot

► **To cite this version:**

Sylvie Giger-Reverdin, Céline Domange, Laurent Broudiscou, Daniel Sauvant, Valérie Berthelot. Rumen function in goats, an example of adaptive capacity. *Journal of Dairy Research*, 2020, 87 (1), pp.45-51. 10.1017/s0022029920000060 . hal-03040684

**HAL Id: hal-03040684**

**<https://hal.inrae.fr/hal-03040684>**

Submitted on 23 Nov 2021

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial - NoDerivatives 4.0  
International License

1 **Rumen function in goats, an example of adaptive capacity**

2

3 Sylvie Giger-Reverdin<sup>1</sup>, Céline Domange<sup>1</sup>, Laurent P. Broudiscou<sup>1</sup>, Daniel Sauvant<sup>1</sup> and  
4 Valérie Berthelot<sup>1</sup>

5

6

7 <sup>1</sup>UMR Modélisation Systémique Appliquée aux Ruminants, INRA, AgroParisTech,  
8 Université Paris-Saclay, 75005, Paris, France

9

10 Short title: **Rumen function in goats**

11

12

13

14 Author for correspondence: Sylvie Giger-Reverdin

15

16 Email: [sylvie.giger-reverdin@agroparistech.fr](mailto:sylvie.giger-reverdin@agroparistech.fr)

17

18

19

20

21 This Open Access text is the final draft version of this published article:

22

23 Giger-Reverdin S, Domange C, Broudiscou LP, Sauvant D and Berthelot V (2020) Rumen  
24 function in goats, an example of adaptive capacity *Journal of Dairy Research* **87** 45-51

25

26

27

28

29

30 **Abstract:**

31 The aim of this Research Reflection is to describe the basic rumen function of goats and its  
32 modification in response to environmental factors, as well as to discuss similarities and  
33 differences when compared to other ruminants. In so doing we shall reveal the adaptive  
34 capacity of goats to harsh environments. The basic rumen function in goats is similar to other  
35 species of ruminants, as stressed by the opportunity to apply the updates of feeding systems  
36 for ruminants to goats. The rumen epithelium acts as a protective barrier between the rumen  
37 and the host, but it can be damaged by toxic compounds or acidosis. The rumen also plays an  
38 important role in water balance, both for dehydration and rehydration. Recent studies show  
39 that the microbiota exhibits a high fractional stability due to functional redundancy and  
40 resilience, but this needs more investigation. The microbial community structure differs  
41 between goats and cows, which explains the difference in sensitivity to milk fat depression  
42 following intake of high lipid diets. Goats also differ from other ruminants by their enhanced  
43 ability to feed-sort, but as with cows they can suffer from acidosis. Nevertheless, goats can be  
44 considered to be very resistant to environmental factors such as water stress, salt stress or heat  
45 stress, and this is especially so in some endogenous breeds. They also are able to detoxify  
46 tannins, polyphenols and other secondary metabolites. Some new trials involving feeding  
47 behaviour, microbiota and omics or approaches by meta-analyses or modelling will improve  
48 our knowledge of rumen function in goats.

49 **Key-words:** digestive tract, ruminant, environmental conditions, microbiota, omics

50 **Introduction**

51 Ruminants have developed a specific multiple-stomach system to use the biomass they select  
52 with browsing or grazing. Among them, goats are known to better survive harsh conditions  
53 than other ruminants (Silanikove, 1994), but also to take profit of highly nutritive diets. These  
54 specificities might be linked to the rumen, a complex organ where microbial fermentation has  
55 a major impact on the efficiency of feed utilization. The aim of this short review is to describe  
56 the basic rumen function of goats and the modifications due to environmental factors in order  
57 to discuss if it differs from other ruminants or if goats are a good model for all species of  
58 ruminants in different environments.

## 59 **Basic rumen function**

### 60 *Degradation of dietary constituents and ruminal metabolism*

61 Goats are similar to other ruminants for the basic rumen function: biomass consumed by the  
62 animals is partly fermented in the rumen by the microbes and converted to microbial matter,  
63 volatile fatty acids (VFA), fermentation gases (methane and carbon dioxide) and ammonia, all  
64 together with the production of heat. The transit fractional rate responses to feeding level and  
65 proportion of concentrate are generally similar for cattle and small ruminants (Sauvant *et al.*,  
66 2006). On the other hand, there are no publications where the efficiency of microbial growth  
67 in the rumen of goat has been compared, for the same diet, to that of other ruminants.  
68 Nevertheless, the global similarity between goat and cows has been used in the Feeding System  
69 for ruminants, like the recently updated INRA feeding system for ruminants (INRA, 2018),  
70 even if goats present some specific digestive features. The new concept of rumen protein  
71 balance (RPB) that represents the difference between crude protein (CP) intake and non-  
72 ammonia CP flowing out at the duodenum (i.e. undegraded feed CP + microbial CP +  
73 endogenous CP) can be applied to goats as to other ruminants (Giger-Reverdin & Sauvant,  
74 2014). RPB is highly correlated with the ammonia level in the rumen, and then to the urinary  
75 N losses with no difference between goats and cattle (Sauvant *et al.*, 2018b). However, in  
76 contrast to cattle, there is no negative digestive interaction due to concentrate supply in goats  
77 (Sauvant *et al.*, 2018a). This is consistent with the fact that, for similar mixed diets, the rumen  
78 pH of goat is higher by about 0.4 point compared to cattle (Sauvant *et al.*, 2018c). For poor  
79 diets, despite the old results of Devendra (1978), the debate is still running regarding  
80 differences in digestive efficiency between goats and other ruminants, but the digestibility  
81 seems to be similar with good diets (Sales *et al.*, 2012).

82

### 83 *Role of the ruminal epithelium*

84 The ruminal epithelium acts as a barrier between the rumen and the host. It has two main  
85 functions: absorption of nutrients and protection against toxic products, as has been  
86 extensively reviewed recently (Baldwin & Connor, 2017). The rumen barrier function can be  
87 impaired when the animals suffer from an important drop of rumen pH (acidosis) as has been  
88 observed over many years in goats used as a model of ruminants (Das & Misra, 1992). Rumen  
89 epithelial tight junctions might be damaged with disruption of ruminal epithelial associated  
90 with a local inflammatory response (Liu *et al.*, 2013), electrophysiological properties are also  
91 modified with changes in net ion transfer and the ruminal epithelial permeability increases  
92 (Klevenhusen *et al.*, 2013).

93

94 *Water storage and resistance to dehydration*

95 The rumen plays an important role in water balance both at times of dehydration and  
96 rehydration, because it acts as a water reservoir containing a large volume of water (Jaber *et*  
97 *al.*, 2013). Some breeds that are well-adapted to harsh conditions such as the Black Bedouin  
98 goat might face a four day water deprivation with a loss of 40 % body-weight. Since a large  
99 portion (50-70%) of the water lost during dehydration comes from the rumen, the animal is  
100 able to maintain a normal water balance in blood and body tissues to ensure a body water  
101 level compatible with life (Silanikove, 1994). During rapid rehydration, the rumen may store  
102 water for some hours to prevent haemolysis and osmotic shock to tissues. For example, Black  
103 Bedouin goats are able to drink water equivalent to 20-40 % of their body mass in one episode  
104 every four days in the Sinai desert (Middle East), which is an extremely valuable trait in arid  
105 regions with few available feeds (Silanikove, 1994). There is a large difference in the capacity  
106 to cope with both dehydration and rehydration between animal species or breeds within  
107 species, such that European breeds like the Saanen goat are more sensitive than breeds  
108 indigenous to arid lands like Bedouin goats (Silanikove, 1994).

109

110 *Microbiota*

111 As in all ruminants, Bacteroidetes and Firmicutes are the dominant phyla in goats with low  
112 abundance of Fibrobacteres. The microbiota is usually dominated by Prevotella followed by  
113 Butyrivibrio and Ruminococcus, as well as unclassified *Lachnospiraceae*, *Ruminococcaceae*,  
114 *Bacteroidales*, and *Clostridiales*. Diversity within the archaea is much lower than for bacteria,  
115 with only a few methanogenic groups dominating the rumen microbiota (Methanosphaerae,  
116 Methanobacteriaceae and/or Methanobrevibacter). The genera *Entodinium* and *Epidinium* are  
117 dominant for the protozoa. Even though the main micro-organisms are widespread in  
118 ruminants, the communities of the microbiota can be different according to the host species.  
119 For example, whatever the diets, goats have a higher relative abundance of unclassified  
120 *Veillonellaceae* and a lower relative abundance of *Fibrobacter* (Henderson *et al.*, 2015).  
121 Usually, diets fed to the ruminant are the major determinant of the bacteria structure  
122 (Henderson *et al.*, 2015). Even though diets affect the rumen microbiota structure, the  
123 microbiota usually exhibits a high functional stability due to functional redundancy and  
124 resilience. Nevertheless, diets rich in concentrate or supplemented with lipids can affect both  
125 the structure and function of the microbiota. In line with cows, high grain diets reshape the  
126 rumen microbial community by reducing its richness and diversity and changing the microbial

127 composition in goats. Zhang *et al.* (2019) showed that 30 taxa were affected by the diet, there  
128 being 5 enriched taxa (*Selenomonas 1*, *Ruminococcus* and unclassified *Veillonellaceae*) in the  
129 high grain diet group and 25 enriched taxa in the hay diet group (*Butyrivibrio*,  
130 *Pseudobutyrvibrio*, *Fibrobacter* and several unclassified taxa such as unclassified  
131 *Christensenellaceae*, *Ruminococcaceae* and *Ruminococcaceae*) at the genera level. These  
132 changes in the composition of the microbiota were associated with modifications in the rumen  
133 metabolome with enhanced capacity to influence amino acid and nucleotide metabolisms. The  
134 linkages between rumen bacteria and metabolites are extremely complex (Zhang *et al.*, 2019).  
135 The composition of the rumen microbiota is also altered by the dietary crude protein (CP)  
136 content. Min *et al.* (2019) observed that the proportions of proteolytic bacterial species tended  
137 to be higher in goats grazing sunn hemp (*Crotalaria juncea*, 17 % CP) compared to  
138 bermudagrass (*Cynodondactylon*, 10 % CP). Similarly, the *Prevotella* and *Selenomonas*  
139 genera proportions were increased in cows fed alfalfa rather than a cornstalk-based diet  
140 (Zhang *et al.*, 2014).

141 Each adult animal harbours its own microbiota even when animals are bred and fed  
142 identically, suggesting that the host also has a significant impact on the composition of its  
143 microbiota. In dairy cows, Weimer *et al.* (2017) showed that the ruminal bacteria  
144 communities moved toward re-establishment of the pre-exchange communities within days to  
145 weeks at a similar diet, suggesting a high specificity and resilience of the rumen microbiota  
146 within its host. The animal might exert some influence over its rumen microbiota through its  
147 intake behaviour or its digesta passage rates such as the fractional turn-over rate of the solid  
148 particles. Because of the specificity of goats regarding their intake behaviour (intermediate  
149 feeder vs grazer for cow) and their potential higher rumen turn-over rates compared to cows  
150 (Clauss *et al.*, 2006), specific studies in goats are needed. The influence of the host on ruminal  
151 functions is poorly documented except on the methanogenesis function. A better  
152 understanding of the microbial composition, the functional role of microbes in fermentation  
153 and how the host controls its own microbiota is essential to be able to manipulate the rumen  
154 microbiota.

155

#### 156 *Lipid metabolism and biohydrogenation*

157 Dietary fatty acids (FA) in forage, cereals and oilseed are mainly C18-carbon polyunsaturated  
158 fatty acid (PUFA) especially linoleic (C18:2 9c,12c) and alpha linoleic (C18:3 9c,12c,15c)  
159 acids. Dietary unsaturated lipids undergo bacterial lipolysis and extensive biohydrogenation  
160 of released FA in the rumen resulting in the formation of saturated FA, and of a variety of

161 positional or geometric (cis, trans) isomers of unsaturated FA (Lourenço *et al.*, 2010).  
162 *Butyrivibrio*-related bacteria isolated in the rumen were thought to be the main active  
163 population carrying out the biohydrogenation process. However, with the development of  
164 culture-independent high-throughput next-generation sequencing techniques, it was shown  
165 that uncultivated microbial species including *Prevotella*, *Lachnospiraceae incertae sedis*, and  
166 unclassified *Bacteroidales*, *Clostridiales* and *Ruminococcaceae* might also be involved (Huws  
167 *et al.*, 2011). Knowledge is still limited on the microbial ecology of FA metabolism,  
168 especially in goats. The apparent biohydrogenation values of linoleic and linolenic acids  
169 ranged between 85 and 95% depending upon rumen conditions such as pH and microbial  
170 populations. Low ruminal pH observed with increasing amounts of concentrates can result in  
171 incomplete biohydrogenation leading to increased production of trans FA (Lourenço *et al.*,  
172 2010). A shift in the biohydrogenation pathways, from the 11t to the 10t pathways can also be  
173 observed with production of rumen biohydrogenation intermediates (C18:2 10t-12c; C18:1  
174 10t...) with supposed antilipogenic effects on the mammary gland, inducing milk fat  
175 depression (MFD). However, interspecies differences in the rumen biohydrogenation process  
176 were poorly investigated except through indirect comparison of milk FA profiles. In line with  
177 interspecies differences in microbial population and composition (Henderson *et al.*, 2015) and  
178 in rumen enzymes activities and DM degradation (Moon *et al.*, 2010), it could not be ruled  
179 out that there might be differences in the biohydrogenation process between cows and goats.  
180 In a direct comparison of the ruminal lipid metabolism in dairy cows and goats, Toral *et al.*  
181 (2016) suggest that *Ruminococcaceae* may be linked to the saturation of C18:1 in the rumen  
182 of cows and *Pseudobutyrvibrio* in goats. Moreover, microorganisms are able to synthesize  
183 their own FA from carbohydrates or amino acids contributing up to 60 % of the total FA  
184 outflows from the rumen leading to FA duodenal flows higher than FA intake in cows fed low  
185 fat diets (Schmidely *et al.*, 2008). They also synthesize specific FA such as odd FA and  
186 methyl branched-chain FA (BCFA) of the iso and anteiso forms. As variation in the odd FA  
187 and BCFA profile leaving the rumen was expected to reflect changes in the relative  
188 abundance of specific bacterial populations in the rumen, these FA were thought to be useful  
189 as markers of rumen function and microbial synthesis (Fievez *et al.*, 2012). But as dietary FA  
190 contents and treatments might affect the contribution of microbial FA to total FA outflows  
191 and also affect the odd-FA and BCFA bacterial content and outflows differently, these  
192 outflows as potential markers of changes in the relative abundance of rumen bacteria strains  
193 should be used with care (Berthelot *et al.*, 2019).

194

195 *Role of epigenetics and development of the rumen with age*

196 Development of the digestive compartments begins at around the same prenatal stage in sheep  
197 and goats, but later than in cattle (Garcia *et al.*, 2012). Microbial colonization pattern and  
198 fermentation differs between young goats reared during the first month of life under different  
199 (natural vs artificial) milk feeding systems (Abecia *et al.*, 2014). However, the rumen  
200 epithelial immune development was not modified by distinct microbial colonization patterns  
201 (Abecia *et al.*, 2014). It must be stressed that some supplementation in early life could  
202 temporarily be of interest, as for example medium chain FA to decrease methane production,  
203 but might also have a negative effect on daily gain of kids and modify some rumen papillae  
204 characteristics (Debruyne *et al.*, 2018).

205

206 **Responses to environmental factors**

207 *Feeding behaviour and high concentrate diets*

208 The rumen can be considered as a fermenter and the intake of feedstuffs as the supply of  
209 substrate for the fermenter. Thus, the pattern and the quality of intake play an important role  
210 on the fermentation occurring in the rumen (Desnoyers *et al.*, 2011) Sheep and goats have  
211 quite similar feeding behaviour and graze selectively on heterogeneous resources in order to  
212 eat a diet of higher quality than offered (Baumont *et al.*, 2000), however, goats eat more  
213 slowly than sheep because they tend to select their feeds more carefully (Morand-Fehr *et al.*,  
214 1991). The supply of concentrate might be up to 50% or more of the dry matter intake in some  
215 high producing herds, which can have the effect of inducing rumen acidosis. Eating and  
216 ruminating behaviours are key parameters to be considered in the occurrence of this disease in  
217 goats (Giger-Reverdin, 2018) as in cattle (Maekawa *et al.*, 2002). When facing an acidogenic  
218 diet, goats develop different individual strategies. They can decrease their intake rate and  
219 duration and hence the dry matter eaten during the first eating bout (Serment & Giger-  
220 Reverdin, 2012). They can also sort against concentrates and search for fibre (Giger-Reverdin,  
221 2018). Without concentrate, mean daily chewing time ( $962 \pm 35$  min/d) is close to the mean  
222 maximum of 1000 min/d generally observed in ruminants, but each supply of 100g/day of  
223 concentrate decreases daily chewing duration by  $23.3 \pm 2.8$  min/d as obtained from the  
224 bibliography data base “Caprinut” (Sauvant & Giger Reverdin, 2018). This decrease in  
225 mastication causes a proportional reduction in salivary input to the rumen and buffer  
226 recycling, and thus increases the risk of rumen acidosis. With a total mixed ration (TMR),  
227 chewing duration decreases  $57.6 \pm 6.6$  min/d for an increase of 10 % concentrate. When  
228 compared to cattle the chewing time per g of dry matter intake (DMI) in goats is about 10



229 time higher (Sauvant *et al.*, 2008). This difference could impact the flow of bicarbonate  
230 recycling/g of DMI and explain the higher value of rumen pH for similar diets, mentioned  
231 above.

232

### 233 *Lipid supplementation*

234 In most ruminant diets, fat represents less than 5% of total dry matter. However, fat can be  
235 added to the diet to improve its energetic value in dairy production. It is also often used to  
236 modify the FA profile of ruminant products (milk, meat) to improve their nutritional,  
237 organoleptic or technological properties. However, fat supplementation often decreases  
238 microbial growth, especially fibrolytic bacteria and protozoa, and rumen fiber digestibility. It  
239 also decreases the DMI of cows and goats except in goats in early lactation (Faverdin *et al.*,  
240 2018). As in cows, diets rich in lipid increase the level of trans FA in goats (Cremonesi *et al.*,  
241 2018). The biohydrogenation intermediates may vary according to the type of lipids. Those  
242 rich in C18:3 9c,12c,15c (linseed) favor biohydrogenation intermediates characteristic of  
243 C18:3 biohydrogenation (C18:3 9c,11t, 15c, C18:2 11t,15c, C18:1 15c, C18:1 15t) and those  
244 rich in C18:2 9c,12c produce intermediates more characteristic of C18:2 biohydrogenation  
245 (C18:1 6t-9t, C18:1 10t, C18:1 11t) (Cremonesi *et al.*, 2018). In this study, despite different  
246 biohydrogenation pathways, *Butyrivibrio* and *Pseudobutyrvibrio* were not affected by the  
247 lipid supplementation. *Fibrobacteriaceae* and *Prevotellaceae* were the bacterial families  
248 showing the highest and significant correlation with FA involved in the biohydrogenation  
249 pathway of C18:3 and C18:2. When ruminal lipid metabolism was compared in dairy cows  
250 and goats with diets supplemented with starch and plant oil or fish oil, an interaction between  
251 diets and species was observed indicating that the responses of cows and goats to dietary  
252 treatments were different. With the plant or fish oil diets, goats exhibited greater increases in  
253 C18:1 trans FA in the rumen fluid compared to cows but the shift from C18:1 11t to 10t and  
254 the increase in C18:2 10t, 12c was greater in cows fed the starch and C18:2 oil-enriched diet.  
255 This suggests that the biohydrogenation pathways are more stable and robust in response to  
256 high starch diet with plant oils in goats (Toral *et al.*, 2016). This is consistent with the higher  
257 sensitivity of cows to MFD. In line with these interactions, the bacterial populations affected  
258 by lipid supplemented diets differ between cows and goats, in agreement with species specific  
259 microbial community structures. *Ruminococcaceae*, *Lachnospiraceae* and  
260 *Succinivibrionaceae* were affected in cows whereas *Prevotella*, *Clostridium cluster IV* and  
261 *Veillonellaceae* were modified in goats (Toral *et al.*, 2016).

262

264 A peculiarity of ruminants is the ability to avoid potentially toxic plant species in their diet  
265 and/or to be more resistant to secondary metabolites which represent potential toxic  
266 compounds, (for example, alkaloids, terpenes and terpenoids, organic acids like oxalic acid and  
267 phytic acid, glucosinolates, cyanides, saponins and phenolic compounds like tannins and  
268 flavonoids). Due to its geographical distribution, a large proportion of the goat population is  
269 exposed to these situations, particularly in countries where climatic and soil conditions favour  
270 the development of plants which produce all the more secondary metabolites to defend  
271 themselves against heat or water stress. In these areas, small ruminants, including goats,  
272 appear particularly resistant to ingestion of large amounts of anti-nutritional compounds and  
273 even of toxic metabolites (Silanikove *et al.*, 1996). They are also less sensitive to mycotoxins  
274 than monogastrics because of the rumen microbiota and the interactions inside the rumen with  
275 feed particles enabling the degradation, deactivation and hence detoxification of these  
276 metabolites (Gallo *et al.*, 2015). Moreover, in the ruminants, there is a difference in  
277 detoxification capacity, one such example being the degradability of mycotoxins like  
278 aflatoxin B1 which is higher in goats than in steers (Upadhaya *et al.*, 2009). Even so, this  
279 degradation of aflatoxin B1 in the rumen of the goat leads to the formation of aflatoxin M1  
280 excreted in the milk like in other ruminant species (Battacone *et al.*, 2009). This ability can be  
281 linked to a behavioural adaptation towards some secondary metabolites. It may lead to  
282 modifications of the dietary selection pattern (Duncan *et al.*, 2000, Mkhize *et al.*, 2018), but  
283 also to specific detoxification enzymatic batteries of secondary metabolites which can be  
284 realized at different places in ruminants but mainly in the epithelium of the rumen. In the case  
285 of rhodanese, a ubiquitous enzyme playing a central role in cyanide detoxification, the activity  
286 was highest in the epithelium of the rumen of goats (Nazifi *et al.*, 2003).

287 Currently there is considerable research interest in the tannins and the benefits of agro-  
288 industrial by-products containing tannins (for example, chestnut husk, grape skin, winery  
289 residue) introduced into the diet of ruminants (Kondo *et al.*, 2016). Tannins are part of the  
290 group of phenolic compounds and because of their multiple phenolic hydroxyl groups, one of  
291 their main properties is the ability to form complexes with proteins. Moreover, because of  
292 their varied natures (hydrolysable or condensed tannins), these metabolites can lead to  
293 beneficial or detrimental effects on the ruminant health and feed efficiency according to their  
294 concentration (Makkar, 2003). One of the interest aspects of dietary tannins is protection of  
295 proteins against ruminal degradation. This could become a handicap when the only sources of  
296 protein are provided by legumes rich in condensed tannins, reducing nitrogen availability to

297 rumen microorganisms and inhibiting growth of the main ruminal bacteria. Nevertheless,  
298 McSweeney *et al.* (1999) could show that in sheep and goats fed a tannin-containing shrub  
299 legume *Calliandracalothyrsus*, some rumen bacteria isolated from goats had an ability to  
300 digest protein in the presence of condensed tannins, attesting to the specific digestion and  
301 resistance characteristics of the caprine species to secondary metabolites. The architecture of  
302 terpens (the presence of oxygen-containing ring structures) which are also important  
303 secondary metabolites has a strong influence on their rumen degradability (Malecky *et al.*,  
304 2009). These observations may prove useful to rationalize the use of essential oils and plant  
305 dry extracts which are increasingly incorporated as additives to the diet of other ruminant  
306 species to optimize ruminal fermentations (Calsamiglia *et al.*, 2007).

307

#### 308 *Adaptation to salt or salt stress*

309 Animals may intake large amounts of salt with either feedstuffs or drinking water. Quite  
310 often, both sources of salt are combined because water available for drinking is the same as  
311 that used by the plants to grow on salty soils. This can be of critical importance when the  
312 animals are grazing halophytes and when the saline water from underground wells is the only  
313 available drinking water (Ashour *et al.*, 2016). According to the recent review of Attia-Ismail  
314 (2016), intake of salt might modify the rumen fermentative profile with an impact on the acid  
315 base equilibrium, especially on Na<sup>+</sup>, K<sup>+</sup> and Cl<sup>-</sup>, and thus on the osmotic pressure within the  
316 rumen. The animal drinks more water to balance this effect, which can decrease the adhesion  
317 of bacteria to feed particles in the rumen and increase the turnover rate of solid and liquid  
318 phases in the rumen. The consequence is a lower digestion in the rumen. Large differences in  
319 salinity tolerance between animal species or between breeds within species are observed, and  
320 it seems that sheep and goats are more tolerant to salt stress than cattle when adapted, and that  
321 goats have a slight tolerance advantage over sheep (Dunson, 1974, McGregor, 2004, Squires,  
322 2016). Goats are able to cope without any detrimental effect on digestibility up to levels of  
323 8326 mg TDS (total dissolved solutes) in water (Paiva *et al.*, 2017) but, as in heifers, rumen  
324 function and cell wall digestibility decreases with an increase in TDS (Alves *et al.*, 2017).

325

#### 326 *Heat stress*

327 Heat stress is often associated with water deprivation or infrequent drinking in animals living  
328 in arid areas (Silanikove, 1992). Feed intake decreases during heat stress for several reasons.  
329 Thermoregulation operates to decrease heat production arising from rumen fermentation, and  
330 there is limited availability of water and of feeds, the majority of which have a poor nutritive

331 value (Morand-Fehr & Doreau, 2000). Feeding pattern is also modified with an increase in  
332 night grazing. In these conditions, reduction of passage rate through the digestive tract might  
333 increase digestibility, but this benefit is overridden by the negative effects of heat stress and  
334 water deprivation (Silanikove, 1992). Rumen fermentation is modified by heat stress: rumen  
335 pH decreased at equivalent dry matter intake (Castro-Costa *et al.*, 2015) and the rumen  
336 bacterial community changes in goats (Zhong *et al.*, 2019). Indigenous goats adapted to harsh  
337 conditions are more capable of coping with heat stress than non-desert breeds (Silanikove,  
338 1992). Moreover, goats have a poor insulation capacity in contrast to sheep, but have the  
339 advantage of dissipating heat by sweating (Silanikove, 1992).

#### 340 **Perspectives**

341 This review exposes some areas in which knowledge is lacking and there is need for further  
342 research and new approaches.

343

#### 344 *Feeding behaviour*

345 Goats exhibit an important sorting behaviour compared to other ruminants, which impacts  
346 rumen function and the efficiency of microbial growth. More studies are needed to better  
347 separate the influence of feed sorting from the intrinsic species effect, and to find an  
348 explanation to the lack of digestive interaction due to the proportion of concentrate, or to the  
349 higher rumen pH compared to cattle for a similar diet.

350

#### 351 *Microbiota*

352 Despite the wealth of information provided by modern omics techniques, little progress has  
353 been made in the understanding of the relationship between the structure and functions of  
354 rumen microbiota. The methodological effort needed to quantify the microbiota structural and  
355 metabolic characteristics is tedious enough to hinder the implementation of dedicated  
356 experiments. Moreover, the strong redundancy among the main functions in the ruminal  
357 ecosystem limits the potential number of unequivocal and specific relations between  
358 microbial species and functional abilities. However, two areas are worth exploring in this  
359 relationship; firstly, the consideration of the host phenotype for some important functions of  
360 the microbiota such as methanogenesis, and secondly, the consideration of smaller scales,  
361 close to the size of the plant tissues, that are potential ecological niches capable of harbouring  
362 specialized microbial communities.

363

364 *Omics*

365 As previously pointed out, the many interactions occurring between the different animal  
366 tissues and cells but also, at different levels, between the cell (genome) and exogenous events  
367 (environment) are hindrances to understanding the underlying mechanisms and the role of the  
368 host compared to that of the rumen microbiota. One of the ways to access all of the systemic  
369 and/or tissue-specific signatures is the approach *via* “omics”. Indeed, these approaches are  
370 complementary in the search for interrelationships between genotypes and phenotypes  
371 (Shahzad & Loor, 2012). Metabolomics, in which advanced analytical chemistry techniques  
372 and multidimensional statistical analyses are applied to measure large numbers of small  
373 molecule metabolites in cells, tissues and biofluids (end products of these complex  
374 interactions), after being first exploited in biomedical research, is progressively used also in  
375 research and monitoring of livestock (Goldansaz *et al.*, 2017). Most of the time, it is the  
376 association between different complementary approaches which provides most information.  
377 For example, by combining metabolomics and proteomic studies, it is possible to get a better  
378 knowledge of the role of the rumen epithelium in goats adapted to grain-rich feeding  
379 compared to hay feeding (Guo *et al.*, 2019). The joint and simultaneous use of metabolomics  
380 and pyrosequencing studies in goats informs about the metabolic pathways preferentially  
381 involved in the response to high-grain diets (Zhang *et al.*, 2019), whilst the links between the  
382 ruminal bacterial community and metabolites represent a powerful tool in terms of prediction  
383 or monitoring of certain nutritional diseases such as acidosis (Mao *et al.*, 2016, Hua *et al.*,  
384 2017). These approaches also make it possible to investigate more finely and specifically *via*  
385 co-culture the key role of microorganisms such as fungi and methanogens, but also the nature  
386 of the metabolites produced (Cheng *et al.*, 2013). Interrelationships between the different  
387 bacteria of the ruminal community in goat kids after birth and before weaning (Abecia *et al.*,  
388 2018) can also be studied. These first studies using the “omics” approaches in livestock  
389 (including small ruminants and goats), based on non-invasive sampling methodologies and  
390 analysing a high quantity of small molecules in different biological fluids and matrices to  
391 identify putative biomarkers, are probably only just the start of much more extensive research  
392 exploiting the opportunities offered by multi-omics studies (Goldansaz *et al.*, 2017).

### 393 **Conclusion**

394 This review points out that goats have globally similar rumen function when compared to  
395 other ruminants, even if there is a lack of detailed comparison between species in similar  
396 conditions. Knowledge needs to be improved in some areas, such as microbial efficiency and

397 ecology or feeding behaviour. Moreover, some breeds of goats have developed specific  
398 characteristics to sustain them in harsh conditions, because they are able to cope with anti-  
399 nutritional or toxic compounds derived from secondary plant metabolites, and are quite  
400 tolerant to environmental stressors, which is a key point in the context of climate change.

#### 401 **References**

- 402 Abecia L, Ramos-Morales E, Martinez-Fernandez G, Arco A, Martin-Garcia AI, Newbold CJ  
403 & Yanez-Ruiz DR 2014. Feeding management in early life influences microbial  
404 colonisation and fermentation in the rumen of newborn goat kids. *Animal Production  
405 Science* **54**, 1449-1454
- 406 Abecia L, Martinez-Fernandez G, Waddams K, Martin-Garcia AI, Pinloche E, Creevey CJ,  
407 Denman SE, Newbold CJ & Yanez-Ruiz DR 2018. Analysis of the rumen microbiome  
408 and metabolome to study the effect of an antimethanogenic treatment applied in early  
409 life of kid goats. *Frontiers in Microbiology* **9**, article 2227
- 410 Alves JN, Araujo GGL, Neto SG, Voltolini TV, Santos RD, Rosa PR, Guan L, McAllister T  
411 & Neves ALA 2017. Effect of increasing concentrations of total dissolved salts in  
412 drinking water on digestion, performance and water balance in heifers. *Journal of  
413 Agricultural Science* **155**, 847-856
- 414 Ashour G, Badawy MT, El-Bassiony MF, El-Hawy AS & El Shaer HM 2016. Chapter 14.  
415 Impact of halophytes and salt tolerant plants on physiological performance of  
416 livestock. In *Halophytic and salt-tolerant feedstuffs. Impacts on nutrition, physiology  
417 and reproduction of livestock* Eds. HM El Shaer & VR Squires), pp. 261-286. Crc  
418 Press-Taylor & Francis Group, Boca Raton, FL, USA
- 419 Attia-Ismail SA 2016. Chapter 19. Rumen physiology under high salt stress. Crc Press-Taylor  
420 & Francis Group, Boca Raton, FL, USA
- 421 Baldwin RL & Connor EE 2017. Rumen function and development. *Veterinary Clinics of  
422 North America, Food Animal Practice* **33**, 427-439
- 423 Battacone G, Nudda A, Palomba M, Mazzette A & Pulina G 2009. The transfer of aflatoxin  
424 M1 in milk of ewes fed diet naturally contaminated by aflatoxins and effect of  
425 inclusion of dried yeast culture in the diet. *Journal of Dairy Science* **92**, 4997-5004
- 426 Baumont R, Prache S, Meuret M & Morand-Fehr P 2000. How forage characteristics  
427 influence behaviour and intake in small ruminants: a review. *Livestock Production  
428 Science* **64**, 15-28

429 Berthelot V, Albarello H & Broudiscou LP 2019. Effect of extruded linseed supplementation,  
430 grain source and pH on dietary and microbial fatty acid outflows in continuous  
431 cultures of rumen microorganisms. *Animal Feed Science and Technology* **249**, 76-87

432 Calsamiglia S, Busquet M, Cardozo PW, Castillejos L & Ferret A 2007. Essential oils as  
433 modifiers of rumen microbial fermentation. *Journal of Dairy Science* **90**, 2580-2595

434 Castro-Costa A, Salama AAK, Moll X, Aguilo J & Caja G 2015. Using wireless rumen  
435 sensors for evaluating the effects of diet and ambient temperature in nonlactating dairy  
436 goats. *Journal of Dairy Science* **98**, 4646-4658

437 Cheng YF, Jin W, Mao SY & Zhu WY 2013. Production of citrate by anaerobic fungi in the  
438 presence of co-culture methanogens as revealed by H-1 NMR spectrometry. *Asian-*  
439 *Australasian Journal of Animal Sciences* **26**, 1416-1423

440 Clauss M, Hummel J & Streich WJ 2006. The dissociation of the fluid and particle phase in  
441 the forestomach as a physiological characteristic of large grazing ruminants: an  
442 evaluation of available, comparable ruminant passage data. *European Journal of*  
443 *Wildlife Research* **52**, 88-98

444 Cremonesi P, Conte G, Severgnini M, Turri F, Monni A, Capra E, Rapetti L, Colombini S,  
445 Chessa S, Battelli G, Alves SP, Mele M & Castiglioni B 2018. Evaluation of the  
446 effects of different diets on microbiome diversity and fatty acid composition of rumen  
447 liquor in dairy goat. *Animal* **12**, 1856-1866

448 Das SK & Misra SK 1992. Liver function in experimental rumen acidosis in goats. *Indian*  
449 *Journal of Animal Sciences* **62**, 243-244

450 Debruyne S, Ruiz-González A, Artiles-Ortega E, Ampe B, Van den Broeck W, Keyser Ed,  
451 Vandaele L, Goossens K & Fievez V 2018. Supplementing goat kids with coconut  
452 medium chain fatty acids in early life influences growth and rumen papillae  
453 development until 4 months after supplementation but effects on in vitro methane  
454 emissions and the rumen microbiota are transient. *Journal of Animal Science* **96**,  
455 1978-1995

456 Desnoyers M, Giger-Reverdin S, Sauvant D & Duvaux-Ponter C 2011. The use of a  
457 multivariate analysis to study between-goat variability in feeding behavior and  
458 associated rumen pH patterns. *Journal of Dairy Science* **94**, 842-852

459 Devendra C 1978. The digestive efficiency of goats. *World Review of Animal Production* **14**,  
460 9-22

461 Duncan AJ, Frutos P & Young SA 2000. The effect of rumen adaptation to oxalic acid on  
462 selection of oxalic-acid-rich plants by goats. *British Journal of Nutrition* **83**, 59-65

463 Dunson WA 1974. Some aspects of salt and water balance of feral goats from arid islands.  
464 *American Journal of Physiology - Renal Physiology* **226**, R662-R669

465 Faverdin P, Sauvant D, Delaby L, Lemosquet S, Daniel JB & Schmidely P 2018. 9. Dry  
466 matter intake and milk yield responses to dietary changes. In INRA Feeding System  
467 for Ruminants, pp. 149-176, Wageningen Academic Publishers, Wageningen, NLD

468 Fievez V, Colman E, Castro-Montoya JM, Stefanov I & Vlaeminck B 2012. Milk odd- and  
469 branched-chain fatty acids as biomarkers of rumen function - an update. *Animal Feed*  
470 *Science and Technology* **172**, 51-65

471 Gallo A, Giuberti G, Frisvad JC, Bertuzzi T & Nielsen KF 2015. Review on mycotoxin issues  
472 in ruminants: Occurrence in forages, effects of mycotoxin ingestion on health status  
473 and animal performance and practical strategies to counteract their negative effects.  
474 *Toxins* **7**, 3057-3111

475 Garcia A, Masot J, Franco A, Gazquez A & Redondo E 2012. Histomorphometric and  
476 immunohistochemical study of the goat rumen during prenatal development.  
477 *Anatomical Record-Advances in Integrative Anatomy and Evolutionary Biology* **295**,  
478 776-785

479 Giger-Reverdin S & Sauvant D 2014. Relationships of both urine nitrogen output and plasma  
480 urea concentration with rumen protein balance in lactating goats. *Animal Production*  
481 *Science* **54**, 1822-1825

482 Giger-Reverdin S 2018. Recent advances in the understanding of subacute ruminal acidosis  
483 (SARA) in goats, with focus on the link to feeding behaviour. *Small Ruminant*  
484 *Research* **163**, 24-28

485 Goldansaz SA, Guo AC, Sajed T, Steele MA, Plastow GS & Wishart DS 2017. Livestock  
486 metabolomics and the livestock metabolome: A systematic review. *Plos One* **12**, 26

487 Guo CZ, Sun DM, Wang XF & Mao SY 2019. A combined metabolomic and proteomic study  
488 revealed the difference in metabolite and protein expression profiles in ruminal tissue  
489 from goats fed hay or high-grain diets. *Frontiers in Physiology* **10**, 11

490 Henderson G, Cox F, Ganesh S, Jonker A, Young W, Janssen PH & Global Rumen Census C  
491 2015. Rumen microbial community composition varies with diet and host, but a core  
492 microbiome is found across a wide geographical range. *Scientific Reports* **5**, 14567

493 Hua CF, Tian J, Tian P, Cong RH, Luo YW, Geng YL, Tao SY, Ni YD & Zhao RQ 2017.  
494 Feeding a high concentration diet induces unhealthy alterations in the composition and  
495 metabolism of ruminal microbiota and host response in a goat model. *Frontiers in*  
496 *Microbiology* **8**, 12



497 Huws SA, Kim EJ, Lee MRF, Scott MB, Tweed JKS, Pinloche E, Wallace RJ & Scollan ND  
498 2011. As yet uncultured bacteria phylogenetically classified as *Prevotella*,  
499 *Lachnospiraceae* incertae sedis and unclassified *Bacteroidales*, *Clostridiales* and  
500 *Ruminococcaceae* may play a predominant role in ruminal biohydrogenation.  
501 *Environmental Microbiology* **13**, 1500-1512

502 INRA 2018. INRA feeding system for ruminants. Wageningen Academic Publishers,  
503 Wageningen, NLD

504 Jaber L, Chedid M & Hamadeh S 2013. Water stress in small ruminants. In Responses of  
505 Organisms to Water Stress (ed. S Akinci), pp. 115-149, InTech, Rijeka

506 Klevenhusen F, Hollmann M, Podstatzky-Lichtenstein L, Krametter-Frotscher R, Aschenbach  
507 JR & Zebeli Q 2013. Feeding barley grain-rich diets altered electrophysiological  
508 properties and permeability of the ruminal wall in a goat model. *Journal of Dairy*  
509 *Science* **96**, 2293-2302

510 Kondo M, Jayanegara A, Uyeno Y & Matsui H 2016. Variation of tannin contents in selected  
511 agro-industrial by-products and their biological activity in precipitating protein.  
512 *Advances in Animal and Veterinary Sciences* **4**, 66-70

513 Liu JH, Xu TT, Liu YJ, Zhu WY & Mao SY 2013. A high-grain diet causes massive  
514 disruption of ruminal epithelial tight junctions in goats. *American Journal of*  
515 *Physiology - Regulatory, Integrative and Comparative Physiology* **305**, R232-R241

516 Lourenço M, Ramos-Morales E & Wallace RJ 2010. The role of microbes in rumen lipolysis  
517 and biohydrogenation and their manipulation. *Animal* **4**, 1008-1023

518 Maekawa M, Beauchemin KA & Christensen DA 2002. Effect of concentrate level and  
519 feeding management on chewing activities, saliva production, and ruminal pH of  
520 lactating dairy cows. *Journal of Dairy Science* **85**, 1165-1175

521 Makkar HPS 2003. Effects and fate of tannins in ruminant animals, adaptation to tannins, and  
522 strategies to overcome detrimental effects of feeding tannin-rich feeds. *Small*  
523 *Ruminant Research* **49**, 241-256

524 Malecky M, Fedele V & Broudiscou LP 2009. *In vitro* degradation by mixed rumen bacteria  
525 of 17 mono- and sesquiterpenes typical of winter and spring diets of goats on  
526 Basilicata rangelands (southern Italy). *Journal of the Science of Food and Agriculture*  
527 **89**, 531-536

528 Mao SY, Huo WJ & Zhu WY 2016. Microbiome-metabolome analysis reveals unhealthy  
529 alterations in the composition and metabolism of ruminal microbiota with increasing  
530 dietary grain in a goat model. *Environmental Microbiology* **18**, 525-541

531 McGregor BA 2004. The use and macro-mineral content of saline water for goat production.  
532 *South African Journal of Animal Science* **34**, 215-218

533 McSweeney CS, Palmer B, Bunch R & Krause DO 1999. Isolation and characterization of  
534 proteolytic ruminal bacteria from sheep and goats fed the tannin-containing shrub  
535 legume *Calliandra calothyrsus*. *Applied and Environmental Microbiology* **65**, 3075-  
536 3083

537 Min BR, Gurung N, Shange R & Solaiman S 2019. Potential role of rumen microbiota in  
538 altering average daily gain and feed efficiency in meat goats fed simple and mixed  
539 pastures using bacterial tag-encoded FLX amplicon pyrosequencing. *Journal of*  
540 *Animal Science* **97**, 3523-3534

541 Mkhize NR, Heitkonig IMA, Scogings PF, Hattas D, Dziba LE, Prins HHT & de Boer WF  
542 2018. Seasonal regulation of condensed tannin consumption by free-ranging goats in a  
543 semi-arid savanna. *Plos One* **13**, 17

544 Moon YH, Ok JU, Lee SJ, Ha JK & Lee SS 2010. A comparative study on the rumen  
545 microbial populations, hydrolytic enzyme activities and dry matter degradability  
546 between different species of ruminant. *Animal Science Journal* **81**, 642-647

547 Morand-Fehr P, Owen E & Giger-Reverdin S 1991. Feeding behaviour of goats at the trough.  
548 In *Goat Nutrition* (ed. P Morand-Fehr), pp. 3-12, Pudoc, Wageningen, The  
549 Netherlands

550 Morand-Fehr P & Doreau M 2000. Effect of climate uncertainty on feed intake and digestion  
551 in ruminants. In *Livestock production and climatic uncertainty in the Mediterranean*.  
552 *Proceedings of the joint ANPA-EAAP-CIHEAM-FAO symposium, Agadir, Morocco*,  
553 pp. 95-105

554 Nazifi S, Aminlari M & Alaibakhsh MA 2003. Distribution of rhodanese in tissues of goat  
555 (*Capra hircus*). *Comparative Biochemistry and Physiology B-Biochemistry &*  
556 *Molecular Biology* **134**, 515-518

557 Paiva GN, Araújo GGLd, Henriques LT, Medeiros AN, Beltrão Filho EM, Costa RG,  
558 Albuquerque ÍRRd, Gois GC, Campos FS & Freire RMB 2017. Water with different  
559 salinity levels for lactating goats. *Semina: Ciências Agrárias (Londrina)* **38**, 2065-  
560 2074

561 Sales J, Jancik F & Homolka P 2012. Quantifying differences in total tract nutrient  
562 digestibilities between goats and sheep. *Journal of Animal Physiology and Animal*  
563 *Nutrition* **96**, 668-678

564 Sauvant D, Assoumaya C, Giger-Reverdin S & Archimède H 2006. [A comparative study of  
565 the ways of expressing the feeding level in ruminants]. In 13<sup>èmes</sup> Rencontres autour des  
566 Recherches sur les Ruminants, Paris, France, p. 103

567 Sauvant D, Giger-Reverdin S, Archimède H & Baumont R 2008. [Modelling relationships  
568 between chewing activities in ruminants, dietary characteristics and digestion]. In  
569 15<sup>èmes</sup> Rencontres autour des Recherches sur les Ruminants, Paris (FRA), pp. 331-334

570 Sauvant D, Chapoutot P, Ortigues-Marty I & Nozière P 2018a. 3. Energy supply. In INRA  
571 Feeding System for Ruminants, pp. 43-59, Wageningen Academic Publishers,  
572 Wageningen, NLD

573 Sauvant D, Faverdin P, Peyraud JL & Nozière P 2018b. 13. Faecal and urinary nitrogen  
574 excretion. In INRA Feeding System for Ruminants, pp. 203-207, Wageningen  
575 Academic Publishers, Wageningen, NLD

576 Sauvant D & Giger Reverdin S 2018. 21. Dairy and growing goats. In INRA Feeding System  
577 for Ruminants, pp. 339-374, Wageningen Academic Publishers, Wageningen, NLD

578 Sauvant D, Giger Reverdin S & Peyraud J-L 2018c. 15. Digestive welfare and rumen  
579 acidosis. In INRA Feeding System for Ruminants, pp. 213-218, Wageningen  
580 Academic Publishers, Wageningen, NLD

581 Schmidely P, Glasser F, Doreau M & Sauvant D 2008. Digestion of fatty acids in ruminants:  
582 a meta-analysis of flows and variation factors. 1. Total fatty acids. *Animal* **2**, 677-690

583 Serment A & Giger-Reverdin S 2012. Effect of the percentage of concentrate on intake  
584 pattern in mid-lactation goats. *Applied Animal Behaviour Science* **141**, 130-138

585 Shahzad K & Loor JJ 2012. Application of Top-Down and Bottom-up Systems Approaches in  
586 Ruminant Physiology and Metabolism. *Current Genomics* **13**, 379-394

587 Silanikove N 1992. Effects of water scarcity and hot environment on appetite and digestion in  
588 ruminants: a review. *Livestock Production Science* **30**, 175-194

589 Silanikove N 1994. The struggle to maintain hydration and osmoregulation in animals  
590 experiencing severe dehydration and rapid rehydration: the story of ruminants.  
591 *Experimental Physiology* **79**, 281-300

592 Silanikove N, Gilboa N, Perevolotsky A & Nitsan Z 1996. Goats fed tannin-containing leaves  
593 do not exhibit toxic syndromes. *Small Ruminant Research* **21**, 195-201

594 Squires VR 2016. Chapter 15. Water requirements of livestock fed on halophytes and salt  
595 tolerant forage and fodders. In Halophytic and salt-tolerant feedstuffs: Impacts on  
596 nutrition, physiology and reproduction of livestock (eds. HM El Shaer & VR Squires),  
597 pp. 287-302, Crc Press-Taylor & Francis Group, Boca Raton, FL, USA

598 Toral PG, Bernard L, Belenguer A, Rouel J, Hervas G, Chilliard Y & Frutos P 2016.  
599 Comparison of ruminal lipid metabolism in dairy cows and goats fed diets  
600 supplemented with starch, plant oil, or fish oil. *Journal of Dairy Science* **99**, 301–316  
601 Upadhaya SD, Sung HG, Lee CH, Lee SY, Kim SW, Cho KJ & Ha JK 2009. Comparative  
602 study on the aflatoxin B1 degradation ability of rumen fluid from Holstein steers and  
603 Korean native goats. *Journal of Veterinary Science* **10**, 29-34  
604 Weimer PJ, Cox MS, Vieira de Paula T, Lin M, Hall MB & Suen G 2017. Transient changes  
605 in milk production efficiency and bacterial community composition resulting from  
606 near-total exchange of ruminal contents between high- and low-efficiency Holstein  
607 cows. *Journal of Dairy Science* **100**, 7165-7182  
608 Zhang RY, Zhu WY, Zhu W, Liu JX & Mao SY 2014. Effect of dietary forage sources on  
609 rumen microbiota, rumen fermentation and biogenic amines in dairy cows. *Journal of*  
610 *the Science of Food and Agriculture* **94**, 1886-1895  
611 Zhang RY, Liu YJ, Yin YY, Jin W, Mao SY & Liu JH 2019. Response of rumen microbiota,  
612 and metabolic profiles of rumen fluid, liver and serum of goats to high-grain diets.  
613 *Animal* **13**, 1855-1864  
614 Zhong S, Ding Y, Wang YY, Zhou GC, Guo HR, Chen YL & Yang YX 2019. Temperature  
615 and humidity index (THI)-induced rumen bacterial community changes in goats.  
616 *Applied Microbiology and Biotechnology* **103**, 3193-3203  
617