



**HAL**  
open science

## Gene banks for wild and cultivated sunflower genetic resources

Sreten Terzić, Marie-Claude Boniface, Laura L. Marek, Daniel Alvarez, Karin Baumann, Vera Gavrilova, Maria Joita-Pacureanu, Mulpuri Sujatha, Daniela Valkova, Leonardo Velasco, et al.

### ► To cite this version:

Sreten Terzić, Marie-Claude Boniface, Laura L. Marek, Daniel Alvarez, Karin Baumann, et al.. Gene banks for wild and cultivated sunflower genetic resources. OCL Oilseeds and fats crops and lipids, EDP, 2020, 27 (9), pp.1-14. 10.1051/ocl/2020004 . hal-03053994

**HAL Id: hal-03053994**

**<https://hal.inrae.fr/hal-03053994>**

Submitted on 11 Dec 2020

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution| 4.0 International License

## Gene banks for wild and cultivated sunflower genetic resources<sup>☆</sup>

Sreten Terzić<sup>1,a</sup>, Marie-Claude Boniface<sup>2,a</sup>, Laura Marek<sup>3,a</sup>, Daniel Alvarez<sup>4</sup>, Karin Baumann<sup>5</sup>, Vera Gavrilova<sup>6</sup>, Maria Joita-Pacureanu<sup>7</sup>, Mulpuri Sujatha<sup>8</sup>, Daniela Valkova<sup>9</sup>, Leonardo Velasco<sup>10</sup>, Brent S. Hulke<sup>11</sup>, Siniša Jocić<sup>1</sup>, Nicolas Langlade<sup>2</sup>, Stéphane Muñoz<sup>2</sup>, Loren Rieseberg<sup>12</sup>, Gerald Seiler<sup>11</sup> and Felicity Vear<sup>2,\*</sup>

<sup>1</sup> IFVCNS, 21000 Novi Sad, Serbia

<sup>2</sup> UMR LIPM, INRAE, CNRS, Université de Toulouse, 31326 Castanet-Tolosan, France

<sup>3</sup> ISU & USDA-ARS, NCRPIS, 50014 Ames, IA, USA

<sup>4</sup> INTA, Manfredi, 5988 Pcia de Córdoba, Argentina

<sup>5</sup> IPK, Gatersleben, Germany

<sup>6</sup> N.I. Vavilov Institute of Plant Genetic Resources, 190000 St-Petersburg, Russia

<sup>7</sup> NARDI, Fundulea, 915200 Calarasi, Romania

<sup>8</sup> ICAR, 500030 Hyderabad, India

<sup>9</sup> DAI, 9520 General Toshevo, Bulgaria

<sup>10</sup> IAS-CSIC, 14004 Cordoba, Spain

<sup>11</sup> USDA-ARS, Edward T. Schafer Agricultural Research Center, ND58102-2765 Fargo, USA

<sup>12</sup> UBC, Botany Dept., V6T 1Z4 Vancouver B.C., Canada

Received 19 December 2019 – Accepted 29 January 2020

**Abstract** – Modern breeding of sunflower (*Helianthus annuus* L.), which started 100 years ago, increased the number and the diversity of cultivated forms. In addition, for more than 50 years, wild sunflower and other *Helianthus* species have been collected in North America where they all originated. Collections of both cultivated and wild forms are maintained in gene banks in many countries where sunflower is an important crop, with some specificity according to the availability of germplasm and to local research and breeding programmes. Cultivated material includes land races, open pollinated varieties, synthetics and inbred lines. The majority of wild accessions are ecotypes of wild *Helianthus annuus*, but also 52 other species of *Helianthus* and a few related genera. The activities of three gene banks, in USA, France and Serbia, are described in detail, supplemented by data from seven other countries. Past and future uses of the genetic resources for environmental adaptation and breeding are discussed in relation to genomic and improved phenotypic knowledge of the cultivated and wild accessions available in the gene banks.

**Keywords:** inbred lines / open pollinated varieties / wild / annual / perennial

**Résumé** – Les centres de ressources génétiques mondiaux du tournesol sauvage et cultivé : un réservoir de gènes et d'allèles. L'amélioration moderne du tournesol (*Helianthus annuus* L.) a débuté il y a un siècle, diversifiant et augmentant le nombre des formes cultivées du tournesol. De plus, des collectes de tournesols sauvages et d'espèces du genre *Helianthus* ont lieu depuis 50 ans en Amérique du Nord d'où ils sont tous originaires. Ainsi, des collections de tournesols cultivés et sauvages sont conservées par des centres de ressources génétiques dans de nombreux pays où le tournesol est une culture importante. Chacun d'eux présente des spécificités par rapport aux ressources génétiques maintenues, en fonction des programmes de recherche ou de sélection variétale locales. Le matériel génétique cultivé comprend des écotypes, des populations et des lignées tandis que les accessions sauvages correspondent eux écotypes d'*Helianthus annuus* sauvages et des 52 autres espèces apparentées du genre *Helianthus*. Les activités de

<sup>☆</sup> Contribution to the Topical Issue "Sunflower / Tournesol".

\*Correspondence: [felicity.vear@wanadoo.fr](mailto:felicity.vear@wanadoo.fr)

<sup>a</sup> These authors contributed equally to this work.

trois centres de ressources génétiques des États-Unis, de la France et de la Serbie sont décrites en détail, complétées par des données provenant des centres de sept autres pays. L'historique de l'utilisation des ressources génétiques et les perspectives futures pour l'adaptation des variétés à l'environnement sont discutés ainsi que leur caractérisation au niveau génomique et phénotypique.

**Mots clés** : tournesol / *Helianthus* / lignées / écotypes / populations à pollinisation libre / variétés

## 1 Introduction

Preservation of important crop species, cultivars, landraces, and crop wild relatives (CWR) provides the basis for a sustainable agricultural system and ensures the security of our global food supply (Campbell *et al.*, 2010). Collection of germplasm is the first step in conserving genetic resources. The mission of gene banks is to serve as the central repository for maintaining the accessions and related information, to distribute seeds for conducting germplasm-related research and to encourage the use of CWR for crop improvement and product development. Germplasm resources can be categorized as *in situ* resources (maintained as wild populations and landraces in natural habitats) or *ex situ* resources (accessions preserved in gene banks). *In situ*, resources are conserved in natural conditions where they continue to evolve in response to their environment. This is the ideal way to maintain populations, but such natural populations are often at risk of encroachment by human activities compared to populations maintained in gene banks. The most important advantage of *ex situ* conservation is that it prioritizes both availability and preservation of the resource.

Crop genetic resources consist of the total genetic variability in the crop or within sexually compatible species (Holden *et al.*, 1993). The crop genetic variability can be further divided in three pools; primary, secondary, and tertiary. The primary gene pool is the crop itself, landraces, cultivars and populations and the closely related wild species that lack crossing barriers, the secondary gene pool consists of many of the more distantly related crop wild species relatives that produce partially fertile hybrids with the crop, but require little or no manipulation for crossing, and the tertiary gene pool is the most distantly related CWR that require special techniques such as bridge species or embryo rescue to use for breeding. Although many secondary and tertiary gene pools may appear to be unfit for use in breeding programs, they may contain useful genetic variation that will protect crops against new emerging pests in the future. Since we cannot predict with an acceptable level of confidence, the occurrence, severity, or even the nature of future stresses, germplasm with the widest range of genetic diversity as possible should be preserved for future breeding purposes. Hopefully, the present gene bank collections have captured the broadest genetic diversity possible, preserving it for future generations, since many of the accessions have no immediate value.

For over 100 years, CWR have been undeniably beneficial for modern agriculture, by providing breeders with a diverse pool of potentially useful genetic resources (Hajjar and Hodgkin, 2007). Anderson *et al.* (2004) predicted that emerging plant diseases and agricultural pests will become more common. Furthermore, some authors (Palmgren *et al.*, 2015) have

suggested that modern crops have lost properties through domestication and breeding that their CWR possessed to tolerate emerging pests and ever-changing climate.

The total area of the sunflower (*Helianthus annuus* L.) crop worldwide in 2019 was estimated at 25.9 million ha in 72 different countries (USDA, 2019). Globally, oilseed sunflower accounts for 90% of the value of the sunflower crop, with the remaining 10% coming from confectionary sunflower (Hladni, 2016). Oilseed sunflower accounts for up to 12% of the worldwide production of vegetable oils, ranking fourth after palm, soybean and canola oil (Rauf *et al.*, 2017).

Compared to the other main temperate crops, cultivated sunflower is a recent crop. It experienced a domestication bottleneck that narrowed its genetic base but the large number of sunflower CWR makes it possible to mine a vast genetic pool for crop improvement. The primary centre of origin of the genus *Helianthus* in North America makes collection and preservation possible with minimal difficulties. This genus contains 53 species of which 14 are annuals and 39 perennials (Schilling, 2006; Stebbins *et al.*, 2013). They are found from the Atlantic coast to the Pacific and from Canada to Mexico. Since the sunflower CWR and their pathogens and insect pests have been subjected to co-evolution over a long period, the wild species constitute an important source of resistance genes. In addition, knowledge of particular habitats and adaptations of wild species can help to identify potential sources of tolerance genes in ecotypes which survive in areas where abiotic challenges exist (Seiler *et al.*, 2017). Through adaptation, genetic diversity goes hand in hand with habitat diversity, which can often be the key to identify potential sources of genes.

While *H. annuus* was first domesticated in North America, the first active scientific breeding was in Russia at the end of the 19th century and first half of the 20th century. V.S. Pustovoit developed open pollinated varieties from local populations present in different parts of the ex-Soviet Union, especially the Mariupol region in Ukraine (for details, see [Supplementary data and references](#)). A large part of present-day cultivated genetic resources come from these Russian open pollinated varieties, but not all. Wild *H. annuus* and cultivated sunflower can be crossed with no loss of fertility and the first important genetic resource that formed the base of the restorer pool came from an unintended cross of Canadian material (developed from that of Russian emigrants) with wild *H. annuus* in a winter nursery in Texas (Putt, 1964). This difference in origins between the CMS (PET1, Leclercq, 1969) maintainer “female” pool from open pollinated Russian varieties and restorers with some wild *H. annuus* genome may have led to the heterosis observed, one of the reasons for the rapid success of sunflower hybrids since 1970.

In spite of the fact that the Russian Institute VNIIMK was the only important breeding centre in the world until the 1960s,

spread of the sunflower crop in the last 100 years led to a wide variety of oil, confectionary and ornamental types with large differences in physiological and harvesting maturity, height, and disease resistances. Thus, in addition to the old open pollinated varieties, genetic resources with these characteristics are largely represented by inbred lines, which are easier to maintain. The present-day collections of cultivated resources in different countries probably cover most of the available variation, with some specialization according to the interests and locations of research institutes. These resources are important for quite rapid modification of modern cultivated material since they provide sources of disease resistance, oil content, different maturities, and other agronomic characters which can be easily introduced into elite parental lines or hybrids without loss of the most important agronomic traits.

Since sunflower is truly a global crop, grown in a large number of countries, gene banks are maintained in different parts of the world, with varying numbers and diversity of sunflower accessions. Some information about the different sunflower gene banks can be obtained from the GENESYS portal, which is a collaboration between Bioversity International on behalf of System-wide Genetic Resources Program of the CGIAR (Consultative Group on International Agricultural Research), the Global Crop Diversity Trust and the Secretariat of the International Treaty on the Plant Genetic Resources for Food and Agriculture (link: <https://www.genesys-pgr.org/>). However, some of the information in this database is outdated and incomplete. The aim of this paper is to provide an update of the main sunflower gene banks around the world, the numbers of accessions they contain and their specificities, and to provide in-depth details of accessions held in the USDA (USA), INRAE (France) and IFVC (Serbia) gene banks. The discussion will consider how genomics and modern phenotyping methods can improve our knowledge of the resources, mining of the traits of interest, and their importance to provide adequate material for research and breeding in the future.

## 2 Details of the main gene banks

The US, French and Serbian gene banks are described in detail as examples. Other gene banks (Argentina, Bulgaria, Germany, India, Romania, Russia, Spain) are described more succinctly and details are given in supplementary documents. Table 1 provides the basic information provided by curators for all these collections. There are also collections in China and Turkey.

### 3 USA – USDA Gene bank

The USDA sunflower gene bank is part of the U.S. National Plant Germplasm System (NPGS) and is currently housed in Ames, Iowa, at the North Central Regional Plant Introduction Station (NCRPIS), a partnership between the USDA and Iowa State University. The NCRPIS is one of the original US Plant Introduction Stations and the first one to become operational. The cultivated sunflower collection has been in Ames since the station opened in 1948. The wild sunflower collection was started in Texas in the late 1970s at the USDA Bushland location and was transferred to Ames in 1985 along with a more modest wild collection from the

USDA location in Davis, CA. An overarching NPGS goal is for the gene banks to maintain and distribute their collections to researchers and educators worldwide. The standard distribution amount for sunflowers is 100 seeds per accession. Because sunflower is native to North America, it is relatively uncomplicated to add germplasm from wild populations in the US and Canada; therefore, the USDA sunflower gene bank has the goal to have as comprehensive a collection of the wild *Helianthus* species as possible, sampling populations from across the geographic range of each species. To attain that goal, the gene bank continues to add accessions from unrepresented regions or unusual habits within the standard ranges of each species. No sunflower breeding takes place at the Ames, IA gene bank location.

#### 3.1 Cultivated collection

The first donations to the USDA cultivated sunflower collection were all open pollinated populations from international programmes. The collection also contains landrace material, accessions were collected in the south-western US and received in the 1970s, and a large donation of Spanish landrace material received in 1990. The first inbred lines came to the collection in the late 1960s from the breeding program at Texas A&M University, USA. By 1980, the USDA sunflower-breeding program established in Fargo, ND was developing and releasing inbred lines which then became part of the collection. Sunflower breeding has a long history of introgression of wild traits to improve the crop and much of the material developed by the USDA group in Fargo contains introgressed wild genetic segments. The majority of these lines are highly inbred although a group of 56 accessions received in 2018 are partial inbreds or populations. The collection also contains 360 pre-bred lines developed at the University of British Columbia from crossing a cultivated line with 11 different wild annual species resulting in partially inbred, breeding material. For details, see: <https://www.ag.ndsu.edu/fss/ndsu-varieties/usda-sunflower-inbred-lines>.

#### 3.2 Wild *Helianthus* collection

The majority of the wild accessions in the gene bank are wild *H. annuus* (1057). The collection also includes 636 accessions of other wild annual species as well as 904 accessions of wild perennial taxa. The only extant wild *Helianthus* taxa not represented in the collection is *H. niveus* ssp. *niveus*, which is endemic to Baja California, Mexico. The majority of the accessions were collected within the United States except for 10 *H. annuus* accessions collected in Mexico, 24 in Canada, and 56 samples of naturalized *H. annuus* populations from Australia. Of the accessions of the other wild annual species, eight were collected in Mexico and Canada, and 16 were collected as naturalized populations in Australia, Argentina, Moldova, Mozambique and the former Soviet Union. Thirty-six perennial accessions were collected in Canada and five originated from the former Soviet Union. The USDA collection also includes samples from four Asteraceae genera related to *Helianthus*: *Phoebanthus*, *Tithonia*, *Verbesina* and *Viguiera*.

Table 1. Basic information provided by curators for all these collections.

Country	Organisation	Curator	E-mail address	Website	Date first entries	Numbers of accessions			Specificities			
						Cultivated lines	Wild annual populations	Other annual <i>Helianthus</i> spp.				
						CMS	<i>H. annuus</i>	Perennial <i>Helianthus</i> spp.	Other genera			
Argentina	INTA, Cordoba	Daniel Alvarez	alvarez.daniel@inta.gob.ar		1948	1	63	38	Prebreeding			
Bulgaria	DAI General Toshevo	Daniela Valkova	valkova_d@abv.bg		1970–1980	30	125	50	Hybridization with annual and perennial species, hybridization methodology			
France	INRA, Toulouse	Marie-Claude Boniface	crb.tourneso@inra.fr	<a href="https://urgi.versailles.inra.fr/siregal/siregal/grc.do">https://urgi.versailles.inra.fr/siregal/siregal/grc.do</a>	1962	10	369	435	Variability in cultivated, sources of biotic, abiotic stress resistance in wild			
Germany	IPK, Gatersleben	Karin Baumann	baumann@ipk-gatersleben.de	<a href="https://www.ipk-gatersleben.de/">https://www.ipk-gatersleben.de/</a>	1946	2	17	52	35+90 cultivated <i>H. tuberosus</i>			
India	1. ICAR-IJIR	1. Mangesh Y. Dudhe and M. Sujatha	1. director.ijir@icar.gov.in; mangesh.dudhe@icar.gov.in; mulpuri.sujatha@icar.gov.in	1. <a href="http://www.icar-ijir.org.in">www.icar-ijir.org.in</a> 2. <a href="http://www.nbpggomet.in/">http://www.nbpggomet.in/</a>	1969	1	447	50	Prebreeding			
Romania	NARDI Fundulea	Maria Joita-Pacureanu	2. director.nbpgg@icar.gov.in; jalli.radhamani@icar.gov.in; mariapacureanu@yahoo.com	<a href="http://www.winnda-fundulea.ro">www.winnda-fundulea.ro</a>	1965	1	10	20	Genetics of characteristics; CMS, restorer, morphological and physiological, interspecific hybridization			
Russia	VIR, Saint-Petersberg	Vera Gavrilova	v.gavrilova@vir.nw.ru	<a href="http://www.vir.nw.ru">http://www.vir.nw.ru</a>	1922	9	340	13	Genetics of characters (morphological traits, CMS, restoration of pollen fertility, seed storage proteins), interspecific hybridization			
Serbia	IFVC, Novi-Sad	Sreten Terzić	sreten.terzic@ifvms.ns.ac.rs	<a href="https://www.ifvms.rs/kolekcija-divljeg-suncokreta/wild.html">https://www.ifvms.rs/kolekcija-divljeg-suncokreta/wild.html</a>	1980	75	186	304+28 cultivated <i>H. tuberosus</i>	Prebreeding			
Spain	INIA, Madrid	Isaura Martin	martin@inia.es	<a href="http://webx.inia.es/web_coleccionescrf/">http://webx.inia.es/web_coleccionescrf/</a>	1986	196			Confectonery sunflower landraces collected in Spain			
USA	USDA	Laura Marek	lmarek@astate.edu	PassporteCRFeng.asp <a href="https://npgsweb.ars-grin.gov/gringlobal/search.aspx">https://npgsweb.ars-grin.gov/gringlobal/search.aspx</a>	1948	4	1057	636	904	4	Striving for as complete as possible geographic representation of <i>Helianthus</i> crop wild relatives	
												"Cultivated" includes all <i>H. annuus</i> which are not wild

*Helianthus tuberosus* accessions (90) are maintained as seeds, not as clones, due to costs of maintenance for tubers. Maintaining the collection in the field in Ames is problematic due to rodent activity in addition to issues due to the invasive nature of the plants. The majority of the *H. tuberosus* accessions represent samples from wild populations.

## 4 France – INRAE gene bank

The sunflower gene bank (Centre de Ressources Biologiques [CRB]) at INRAE, Toulouse (<https://www6.toulouse.inra.fr/lipm/Recherche/Genetique-et-Genomique-du-Tourne-sol/CRB-Tournesol>), was started after the transfer of INRA research activities at Clermont-Ferrand and Montpellier to Toulouse in 2006–2015. The entries from Clermont-Ferrand resulted from collection and breeding of cultivated populations and inbred lines especially for resistance to diseases, particularly downy mildew and *Sclerotinia* head rot. The material from Montpellier included many accessions of wild *H. annuus* and other *Helianthus* species and a quite large proportion of cultivated lines obtained from crosses of these wild ecotypes with cultivated sunflower followed by selection for a wide range of characters. The aim of the INRAE sunflower gene bank is to maintain, develop and distribute genetic resources of *Helianthus*.

### 4.1 Two thousand and three hundred inbred cultivated sunflower lines

These lines were bred by INRA between 1963 and 2012, or received (in variably homozygous states) during exchange programmes with many different research institutes. There are 1500 lines of French origin, the others come mainly from the USA (340 lines), Bulgaria (88 lines), Russia (87 lines) and Romania (82 lines), but also from 16 other countries. There are 1690 maintainers of CMS PET1 (B lines), their CMS form being available for 560 of them, and 640 lines are male fertility restorers. This material is stored at 4 °C and multiplied under bags about every 10 years. About half, chosen to represent the complete collection, make up the “sunflower network collection”, which is multiplied by INRAE with the help of private sunflower breeders (Caussade Semences, MAS Seeds, Corteva, RAGT, Soltis, Syngenta). The INRA breeding programme for parental lines that could be used commercially terminated in 2012 and this collection is now the basis for genetic and genomic studies (e.g., association mapping, markers, gene identification) for most of the economically important characters in sunflower. Therefore, much of this material is available for research or breeding programmes under an MTA. The list is available at <https://urgi.versailles.inra.fr/siregal/siregal/grc.do>.

First studies of the available variability and identification of representative core collections started in 2006. Coque *et al.* (2008) found three groups, one of B lines and two groups of restorers, one of which included some B lines. Mandel *et al.* (2011, 2013) used the first INRA core collection together with USDA lines and also distinguished restorer and maintainer lines. Using the INRA lines, Cadic *et al.* (2012) defined three groups but with some lines that were not well classified. The whole collection has now been genotyped with SNP arrays

and/or re-sequenced, which should make it possible to eliminate duplicates and to develop improved core collections.

### 4.2 Four hundred and three cultivated sunflower populations

These are samples of open pollinated varieties (OPV) and land races together with pools or synthetics developed as a base for breeding programmes. Collection of this type of material started in 1962 with samples of Russian OPV imported directly from Russia or via Eastern Europe for trials to determine if they would be suitable for cropping in France. The accessions come from 34 countries, with 72 from Russia, 54 from France, 34 from Romania, 27 from the USA, 26 from Spain and 26 from Morocco. As in the case of inbred lines, these populations are maintained at 4 °C, but samples have also been frozen at –18 °C. They are multiplied about every 10 years, mostly under 50–100 m<sup>2</sup> insect proof cages by the *Helianthus* network mentioned above. They are also listed on the same site as the inbred lines (<https://urgi.versailles.inra.fr/siregal/siregal/grc.do>) and are also available with an MTA.

Mangin *et al.* (2017) studied 102 of these populations and showed that variability is structured firstly by oil content (oil vs. confectionary) and then by the presence or absence of restorer genes and by flowering date.

### 4.3 Six hundred and sixty-five wild *Helianthus* accessions

The majority of these accessions (369) are wild *H. annuus*, but there are 66 accessions of other annual *Helianthus* species and 230 accessions of perennial species. There are also five representatives of genera close to *Helianthus*: *Verbesina*, *Tithonia*, *Viguiera* and *Simsia*. Most of the accessions came from USDA but some were obtained during international research collaboration. For example, 216 accessions, including 79 *H. annuus*, were received from IFVC, Novi-Sad, although 64 of them originated from the USDA collection. Thirty-four entries, mostly perennials, were received from Eastern Europe, especially Russia and Bulgaria. A total of 262 accessions came from European or Australian research centres, but again 173 originally came from USDA. A small number of accessions were collected outside North America, seven from South America and 20 from other continents. The first few accessions were sent by C. Heiser to P. Leclercq at INRA, Clermont-Ferrand in 1964, one sample of *H. petiolaris* (numbered 6312) being the origin of the CMS PET1. However, most of the accessions were introduced, maintained and studied at INRA Montpellier by H. Serieys between 1975 and 2008. They were transferred to Toulouse in 2010–2012. In order for this collection to be useable in research programmes, the Toulouse gene bank has undertaken regeneration of seed stocks. Maintenance and multiplication are carried out under insect-proof netting cages, with cross-pollination between several plants of each accession. Accessions are regenerated every ten years and may be distributed using an MTA when seed stocks are sufficient.

Many interspecific crosses were made at Montpellier between 1975 and 2000, mostly with annual species, providing up to 20 sources of CMS (Serieys and Christov, 2005) and one

source of resistance to broomrape (from *H. debilis*). Some successful crosses were made with hexaploids (*H. resinosus*, *H. pauciflorus* (*rigidus*)) in a search for good resistance to *Sclerotinia*) and *H. mollis*, a perennial diploid, to obtain sunflower with sessile leaves (Faure *et al.*, 2002). Wild *H. annuus* accessions provided several sources of CMS that are difficult to restore and also original genes providing resistance to downy mildew (Pecrix *et al.*, 2018). More recently, at Toulouse, other *H. annuus* accessions have been studied in several programmes in collaboration with breeders or other research institutes and field and growth chamber evaluations have shown new sources of resistance to broomrape. Recently, new interspecific crosses have been made between cultivated sunflower and *H. petiolaris*, *H. debilis*, *H. anomalus*, *H. bolanderi*, *H. deserticola*, *H. exilis*, *H. argophyllus*, *H. neglectus* and *H. niveus*.

#### 4.4 One hundred and eighty clones of cultivated forms of Jerusalem artichoke (*Helianthus tuberosus*)

The INRAE collection of cultivated forms of Jerusalem artichoke, which dates from collection and breeding programmes at Clermont-Ferrand and Rennes in the period 1960–1970, and at present maintained at INRAE Montpellier, will be transferred to Toulouse in 2020. Since tubers need to be planted each year, costs are high, and the transfer required a specific support programme and collaboration with the experimentation unit at Toulouse to undertake the maintenance of this material. The clones were characterised for their morphology and phenology at Clermont Ferrand and Rennes, and more recently, at Montpellier, for a certain number of SSR markers. The INRAE collection was also included as part of a comprehensive phenotyping and SNP genotyping study (Bock *et al.*, 2018), which showed that wild and cultivated Jerusalem artichoke are distinct genetically and phenotypically, and that invasive Jerusalem artichoke genotypes have arisen multiple times, with independent ancestry from wild and cultivated material.

## 5 Serbia – IFVC Gene bank

The main goal of this collection was to use wild species in cultivated sunflower breeding. They are mostly used at IFVCNS for disease resistance breeding, but also to work on oil quality, CMS, herbicide tolerance and new phenotype traits. Interspecific crosses and early generation prebreeding line development are a part of the gene bank activities. Besides evaluation for breeding, the work on wild sunflowers expanded with further collecting efforts, maintenance improvements and diversity estimates (Škorić, 2008). The collection was established in 1980 and started with 59 accessions obtained from the collection at INRA, Montpellier (Atlagić and Terzić, 2015). During the period from 1980 to 1991, five collecting trips were conducted in cooperation with the USDA-ARS stations in Bushland, TX and Fargo, ND. Varying numbers of species (1–37) and populations (52–384) were collected during each trip covering 6 to 21 US states per expedition. The first joint collection mission outside the USA was conducted in 1994 in several provinces of Canada. Seeds of the collected species were divided for the collections in USA and Serbia.

Additional seed samples were obtained from collection trips to Montenegro in 1991 and USA in 2001, as well as by exchange from other gene banks (Atlagić *et al.*, 2006).

During establishment of the collection, the method of sowing had to be adjusted to obtain plants from collected seeds. Poor overwintering in Serbian climatic conditions caused the loss of 11 perennial species, while the inappropriate chernozem soil type contributed to the loss of four annual species. The most common problem was low self-fertility due also to the limited number of plants for each regeneration and to manual pollination (Atlagić and Terzić, 2015). The collection in Novi Sad now contains 21 perennial and eight annual species, represented by 332 and 185 accessions respectively. Each year, 60 to 80 accessions of annual species are grown for seed regeneration. Seed reserves are kept in cold storage at +4 °C and vary from a few seeds to several thousand per accession (Tab. S1). Most of the perennial species are maintained as a living collection in quarantine fields; while some also have seed available (Tab. S2). Detailed information about the collected accessions including passport data can be accessed online on the IFVCNS site: <https://www.ifvcns.rs/kolekcija-divljeg-suncokreta/wild.html>. For both annuals and perennials, accessions are available with an MTA when seed stocks are sufficient. Details of wild species are given in Tables S1 and S2, additional references in Supplementary data: Supplementary references 1.

F<sub>1</sub> interspecific hybrids derived from hybridization between perennial species and cultivated sunflower are also maintained at IFVCNS. Plants usually have very low pollen viability, but they can be reproduced vegetatively, and have been grown in the field using clonal maintenance for more than 30 years (Atlagić *et al.*, 2012).

Research was first concentrated on evaluation of most economically important traits. Based on the findings, intensive interspecific crosses were made between wild species and cultivated sunflower (Atlagić *et al.*, 2006). Oil quantity was determined for all accessions (Čuk, 1982), while resistance to *Phomopsis* stem canker and *Sclerotinia* was determined for selected species (Škorić and Rajčan, 1992). The prebreeding programme produced interspecific lines for which combining ability was evaluated (Mihaljčević, 1988) while restorer genes and CMS sources were also determined (Škorić *et al.*, 1988). Cytogenetic analyses were used to monitor and optimize the prebreeding process by studying the number and structure of chromosomes, microsporogenesis and pollen viability (Atlagić, 2004). More recently, efforts were focused more on collection maintenance. Smaller numbers of "targeted" hybrid combinations were made to transfer disease resistance from wild to cultivated sunflower with the help of biotechnology (Terzić *et al.*, 2006). The species in the collection have been characterised using both phenotypic and molecular markers. Large variability was found not only between the species but also among accessions within a single species (Saftić-Panković *et al.*, 2005). Wild species were screened for resistance to broomrape, *Macrophomina*, powdery mildew etc. and the collection was also monitored for natural infection and resistance in field conditions (Terzić *et al.*, 2010; Tančić *et al.*, 2012). The effect of seed ageing on germination was studied on all annual species in the collection (Terzić, 2018). The sunflower association mapping population (UGA-SAM1) described by Mandel *et al.* (2011) was used to evaluate the current crop ontology

and usefulness of qualitative traits for the discrimination of genotypes (Terzić *et al.*, 2019). A detailed morphological evaluation of both the wild and cultivated germplasm is in progress (Lazarević *et al.*, 2016).

All the characterization efforts significantly increased the value of the collection making it more attractive to breeders. Nonetheless, diversity of the species also resulted in frequent problems for their usage, related to cross incompatibility, embryo abortion, sterility and reduced fertility of interspecific hybrids. Variable cross compatibility was found when hybridizing different accessions of the same species with cultivated sunflower. Despite the difficulties, seven annual and 14 perennial species have been crossed with cultivated sunflower using conventional hybridization methods (Atlagić, 2004). The interspecific lines obtained have then been selected for desired traits and used for creation of new elite germplasm (Hladni *et al.*, 2018). The whole prebreeding program resulted in direct use of the wild germplasm, justifying the long-term commitment and investments in collection maintenance and evaluations. The wild species thus remained a constant source of specific genes and variability for cultivated sunflower breeding.

## 6 Other important gene banks

The Vavilov Research Institute for Plant Genetic Resources (Russia) is the oldest gene bank for sunflower, with the first entries in 1922. The sunflower collection at VIR totals 2730 accessions with 2288 accessions of cultivated (*Helianthus annuus* L.) and 442 wild sunflowers belonging to 24 species (of which five are annual and 19 are perennial species). Viable accessions are maintained in the active (+4 °C) and long-term (−10 °C) collections. Studies of sunflower genetic diversity in field and laboratory conditions conducted for many years resulted in the definition of collections with variation for particular traits and for their genetic control. The collection of cultivated sunflower contains local varieties and landraces and cultivars from national and international breeding programmes. It also includes the first CMS-based hybrids which are conserved as hybrid populations and lines (supplementary data 2, supplementary references, Tab. S3).

DAI-General Toshevo (Bulgaria) maintains a collection of wild annual and perennial *Helianthus* species. The first wild sunflower accessions were obtained from the USDA-ARS, Bushland, Texas and planted in the 1970s. A second group of wild sunflower accessions was obtained in 1983. However, a major part of the collection of wild sunflower species was obtained in the course of collections made in collaboration with USDA in the USA in 1999 and 2004. The DAI collection includes accessions maintained in the field and in cold chambers at +4 °C and at −14 °C (long term preservation). Perennial species are permanently grown in the field as a living collection, and annuals are planted each growing season. The DAI collection includes 215 accessions from 30 perennial *Helianthus* species (Tab. S1), 175 accessions from seven annual *Helianthus* species (Tab. S2), 30 different CMS sources and some species from different genera of the Asteraceae family. These accessions are important initial material for research and breeding for resistance to abiotic and biotic stress

factors and variation in fatty acid content (Supplementary data 1, Supplementary references 2).

The INTA (Argentina) sunflower gene bank at Manfredi (Cordoba) is part of the National Network of Genetic Resources. The first accessions grown at the station were OPV incorporated in 1948. The wild sunflower collection is made up of introductions from the USA and naturalized wild collections in Argentina. The first inbred lines were incorporated in 1980 from the INTA breeding programme and other sources. The main objective of this gene bank is to maintain, characterize and promote the use of genetic resources for research and breeding (the INTA programme is also at Manfredi).

ICAR (India) ICAR-IIOR collects, maintains, evaluates and supplies the germplasm to all the centres under the sunflower network in India while ICAR-NBPGR facilitates import of germplasm and maintenance of the material multiplied (Tab. S4).

The INIA (Spain) gene bank contains an important collection of confectionary type landraces collected from many parts of Spain (listed in Tab. S5). The germplasm has been evaluated for seed quality traits and molecular diversity (Supplementary references 4).

The other gene banks with details in Table 1 are: IPK, Germany and NARDI, Romania.

## 7 Discussion

### 7.1 Use of CWR up to the present

During the domestication process, sunflower certainly lost many CWR traits and, perhaps in part due to the domesticated ideotype with a single large head and to production practices, is now vulnerable to many biotic and abiotic stresses. Sunflower CWR possess genetic diversity useful for breeding a productive, nutritious, and resilient crop. In a survey of the introduction of genes from CWR in 13 crops of major importance to global food security from the mid-1980s to 2005, Hajjar and Hodgkin (2007) reported that sunflower CWR had contributed seven traits for the crop, including disease and insect resistance, abiotic factors, male fertility restoration and cytoplasmic male sterility, fifth among crops surveyed. This survey did not include an eighth trait of tolerance to the herbicides imidazolinone and sulfonylurea, which is now widely deployed and also gives partial control of the parasitic weed broomrape (Alonso *et al.*, 1998; Škorić, 2012). Tolerance to imidazolinone and sulfonylurea herbicides were discovered in a wild *H. annuus* population from Kansas in the 1990s (Al-Khatib *et al.*, 1998).

Sunflower CWR have been very important in the global development of sunflower as the fourth largest oilseed crop. Annual losses to diseases, weeds, and insects in the mid-1990s were estimated at \$1.36 billion (Hesley, 1999). At that time, diseases accounted for losses of \$642 million, weeds \$489 million, and insects \$229 million. Prescott-Allen and Prescott-Allen (1986) estimated that sunflower CWR contributed 25.9% of the annual value of the crop, in monetary terms about \$185 million (Tyack and Dempewolf, 2015) or as much as \$267–384 million (Hunter and Heywood, 2011).

The PET1 CMS used as the female parent for the commercial production of sunflower hybrids is the most



valuable trait obtained from the CWR, followed by disease resistance. Sunflower CWR have been a valuable resource of resistance genes since the beginning of scientific breeding programmes and have become particularly useful since the development of hybrids (reviewed by Seiler, 2010, 2012). Resistance genes for rust, downy mildew, Verticillium wilt, Alternaria leaf spot, powdery mildew, Phomopsis stem canker, and Sclerotinia wilt/rot resistance, and broomrape have been introduced into the crop from CWR (Seiler *et al.*, 2017). Dempewolf *et al.* (2017) reviewed more than 350 publications using sunflower CWR, with 210 concerning resistance to biotic stresses, 60 male fertility restoration, 40 resistances to abiotic stresses, 25 quality characters, 10 phenological traits and 5 agronomic traits. For biotic stresses, the majority were for disease resistance, with 12 of the 14 annual species cited tending to provide single dominant resistance genes, most notably for rust, downy mildew, and Verticillium wilt, with annual *H. argophyllus* the most frequently cited species. Only 10 of the 39 perennial species were cited, with *H. tuberosus* in first place, mostly contributing polygenetic resistance to Phomopsis and Sclerotinia (Seiler, 2010). Genomic studies confirm these reports, showing that:

- about 10% of the cultivated genome derives from the secondary gene pool, with 4.7% from *H. argophyllus*;
- introgressions from wild species are enriched for disease resistance genes;
- there are only traces of genetic material from perennials (Hübner *et al.*, 2019).

Sunflower CWR contains considerable variability for tolerance to abiotic stresses such as drought, salinity, heat, flooding, low nutrient, and heavy metal tolerance (Ortiz, 2015). *Helianthus paradoxus* has salinity tolerance which has been successfully transferred to cultivated sunflower (Miller, 1995; Miller and Seiler, 2003). Hajjar and Hodgkin (2007) suggested that hybrids developed using this trait could give a yield increase of 25% in saline soils. CWR also provide the opportunity to study physiological processes involved in the survival mechanisms of desert-inhabiting species, for example Bowsher *et al.* (2016) studied the desert-adapted *Helianthus niveus* ssp. *tephrodes* endemic to the Algodones Dunes in California. *Helianthus argophyllus* has been extensively used for drought tolerance breeding. Baldini and Vannozzi, (1998); Baldini *et al.* (1993) and Martin *et al.* (1992) reported that interspecific lines obtained by divergent selection for physiological traits from *H. argophyllus* had higher water use efficiency, better drought susceptibility index, and higher harvest index under drought conditions than conventional sunflower lines.

## 7.2 From one cultivated sunflower reference genome to reference genome for all the CWR

By the end of 2019, more than 1100 plant genomes (from more than 500 species) have been sequenced (<https://www.ncbi.nlm.nih.gov/genome/browse#!/overview/>) and projects are underway to sequence more plant genomes in the near future, such as the 10,000 Plant Genome Sequencing Project (Twyford, 2018). Sequencing the sunflower genome (Badouin *et al.*, 2017) was a challenging project and a research network

between Canada, France and USA started a decade ago, with this objective (Kane *et al.*, 2011). The size of the genome (approx. 3.6 Gb, 17 chromosome pairs) and the numerous repetitive sequences made the assembly very difficult. Sequencing technologies are evolving rapidly by producing more, cheaper, longer and higher-quality sequences. However, for a long time, sequences were too short to differentiate and compare the long and highly similar repeats. In 2015, the RSII PacBio sequencer produced sequences longer than the known repeats from the sunflower genome and made possible sequencing of the 17 chromosomes of the inbred line XRQ. The raw data did not themselves solve the complexity of the sunflower genome. A long and complex process of bioinformatic computation compared and assembled the millions of sequence fragments as chromosomes. Improvement of bioinformatic software also plays an important role in genomics. As an example, at the same time as using long-read sequences for the sunflower genome, NRGene developed a software permitting genome sequences to be compiled using short-read fragments (Lu *et al.*, 2015). This approach made it possible to obtain a complete sequence of the genome of the line HA412 of similar quality to that obtained for the line XRQ.

To date, three high quality genomes are available (XRQ, HA412 and the restorer line PSC8). By comparing these three genomes, we found several large inversions. In addition, a pan genome for cultivated sunflower based on re-sequencing of 287 cultivated sunflower lines revealed extensive gene presence/absence variation (Hübner *et al.*, 2019; Owens *et al.*, 2019). These structural variations had been very difficult to identify previously without such genomic resources. Single nucleotide polymorphisms are the molecular markers most used either to map traits or to characterize the diversity of sunflower germplasm. Many genotyping methods are available but the most used for high-throughput genotyping are genotyping by sequencing (Elshire *et al.*, 2011; Narum *et al.*, 2013; Baute *et al.*, 2016, Celik *et al.*, 2016; Badouin *et al.*, 2017; Todesco *et al.*, 2019) or genotyping arrays such as the two AXIOM<sup>®</sup> arrays developed and used in sunflower for either mapping traits (Louam *et al.*, 2016; Duriez *et al.*, 2019) or to describe the diversity of germplasm (Mangin *et al.*, 2017) or the Illumina<sup>®</sup> Infinium iSelect used for diversity analysis and genomic prediction (Livaja *et al.*, 2016).

There are many examples of structural variations (Saxena *et al.*, 2014), some of them being involved in plant traits. More recently, it has been shown in sunflower that structural variations are involved in adaptive traits to the environment (Todesco *et al.*, 2019). However, there are no efficient tools for the high-throughput of these structural variations even if a recent report used the AXIOM<sup>®</sup> array in maize (Mabire *et al.*, 2019).

The International Consortium on Sunflower Genomics, grouping four public research institutes and eight private partners, aims to sequence reference genomes for wild sunflowers and wild relatives. The genomes of the CWR are more complex than those of cultivated sunflower. They are highly heterozygous, and it is more difficult to obtain the two haplotypes of the chromosomes for the diploid species. Some wild relatives have more complex ploidy and can be larger, up to 11.21 Gb in *H. agrestis* (Qiu *et al.*, 2018).

Recent advances in sequencing technologies have been successfully used to sequence the genome of two new

cultivated sunflower lines in only few weeks (SUNRISE project) and that should make it possible:

- to obtain one reference genome for all *Helianthus* species in the coming years;
- to describe the diversity within each species more precisely;
- to compare the gene contents and allelic diversity of the germplasms;
- to use all information for mapping new traits or adaptive genes to improve understanding of evolution or to breed new varieties adapted to their changing abiotic and biotic environment.

Further new bioinformatic tools will make it possible:

- to compare genomes and to identify polymorphisms (structural variations included);
- to visualize many genomes at the same time in Genomes Browsers.

### 7.3 High-throughput phenotyping of CWR resources for research and breeding

Usually, CWR genetic resources are phenotypically characterized by a series of manually acquired traits comprising phenology, architecture, ligule colour, leaf colour and shape and seed traits including colour, number, weight, oil content and composition. These data are time-consuming and expensive to acquire and therefore are usually only complete for a subset of a collection. In addition, these traits are largely subjected to environmental effects such as temperature, water deprivation or spatial variation within field nurseries. These factors considerably impair selection of the most promising accessions to start pre-breeding programmes and to integrate genetic innovation into cultivated or elite material.

To break this bottleneck and improve utilization of gene banks, in the last decade, high-throughput phenotyping platforms have been developed to measure plant traits efficiently while controlling or precisely characterizing growing conditions. Glasshouse-based platforms allow a very fine control of the environment (light, water, nutrients; Cobb *et al.*, 2013; Granier *et al.*, 2006), and have been successfully used to conduct genetic association studies (Cabrera-Bosquet *et al.*, 2012) and to derive plant trait ranges that can be entered in crop models (Lenz-Wiedemann *et al.*, 2010; Steduto *et al.*, 2009).

In field conditions, phenotyping has been mainly developed using unmanned aerial vehicles (UAV) as they are portable, adaptable with different captors, low-cost and suitable for all crops and stages (Zhao *et al.*, 2019). When equipped with digital cameras, UAVs can be used to estimate canopy surface and biomass (Liebisch *et al.*, 2015) with multispectral cameras or hyperspectral sensors to characterize physiological processes such as chlorophyll fluorescence or nitrogen levels (Camino *et al.*, 2018) or plant water status using thermal imaging (Gómez-Candón *et al.*, 2016; Gonzalez-Dugo *et al.*, 2015). For short crops, such as wheat, ground based phenotyping robot often termed “Phenobile” have also been developed (Maded *et al.*, 2017; Qiu *et al.*, 2019). In addition to cameras similar to those on UAVs, they can carry heavy and energy-demanding sensors such as LiDAR and additional lights to be independent of natural sunlight varying

in quality and quantity. This facilitates and improves the quality of subsequent image analysis although a limitation of these ground-based vehicles is their suitability for tall crops such as sunflower.

Different high-throughput phenotyping technologies have been developed and tested for sunflower but very few have been published so far. In controlled environments, different tests have been performed on the Phenotoul-TPMP platform (Maviane-Macia *et al.*, 2019) to assess biotic and abiotic stresses on sunflower. This platform is based on a conveyor system in a greenhouse where plants are automatically imaged in visible, fluorescence and near-infrared spectra. Using the RGB images and the IPSO Phen software INRAE teams successfully characterized the responses of sunflower genotypic panels for drought stress at early stages (SUNRISE project) and Orobanche infection (Plant2Pro Phenor project). Exploitation of these methodological developments will certainly follow shortly.

In semi-controlled conditions, still in the frame of the SUNRISE project, the Phenotoul-Heliaphen phenotyping platform was developed by Gosseau *et al.* (2019) to study the effects of different drought scenarios at vegetative and post-flowering stages of sunflower. This platform allowed the authors to impose precisely different drought intensities and characterize yield sensitivity to post-flowering stress of sunflower varieties, but seed weight measurement still required time-consuming manual measurements. At earlier stages, they characterized leaf expansion and transpiration thresholds of different genotypes that were subsequently used to simulate yields with the SUNFLO crop model (Casadebaig *et al.*, 2011). Using Heliaphen-grown plants, Gélard *et al.* (2016, 2017, 2018) applied structure from motion techniques and multi-view stereo imaging to reconstruct and analyse 3D point clouds of adult sunflower plants. They successfully extracted individual leaves, petioles and stem dimensions and orientations on time-series and characterized the differences in genotypic responses to drought (Gélard *et al.*, 2018). Future developments of the high-throughput phenotyping Heliaphen platform will require the automatic acquisition of adequate images for this 3D reconstruction software and its automatic implementation to allow researchers to exploit the facility in full.

Field based phenotyping has also been tested on sunflower by different public and private teams, however few results are publicly available. One example of application for both nursery and phenotyping is plant counting using UAV as illustrated in the Delair and MAS Seeds project [www.delair.aero](http://www.delair.aero). This can be used to assess plant emergence as a phenotypic response to early stresses or to assess density as a proxy of plot quality for further phenotyping. A similar approach was conducted in the SUNRISE project using UAV visible images for plant counting and flowering time. In addition, multi-spectral images could be used to estimate NDVI, that will make it possible to study the evolution of radiation interception during the entire crop cycle. In collaboration with the Phenome EMPHASIS project, ground vehicles could also be tested on sunflower. The Phenobile V2 was first used on sunflower trials on the Phenotoul-Agrophen platform. Equipped with multispectral cameras, LiDARs and RGB cameras, methodological developments are currently underway to estimate leaf area index and above-ground biomass more accurately and dynamically.

The development of high-throughput phenotyping platforms allowed the acquisition of large amounts of images and 3D point clouds but, as in many cases, the main bottleneck is the development of sunflower-specific image analysis pipelines taking into account its leaf architecture and the position of the head. This will require new expertise in image analysis and also the collaboration of experts for the different traits and diseases. Soft- and hardware tools to manipulate these data must also be constructed. In particular, a sunflower crop ontology needs to be developed with international collaboration, and an open phenotype database developed. HTPP-produced data could then follow the FAIR principle and be accessible as on the PhenoDB INRAE database (<https://sunrise.toulouse.inra.fr/phenoDB/>) for further long-term usage and characterization of CWR.

In addition to the development of high-throughput genotyping tools that are widely used to characterize the CWR genomic diversity, the emerging field of phenomics is starting to develop more robust and precise phenotyping methods for sunflower that will allow the characterization of entire collections and therefore both help their conservation and accelerate their exploitation for breeding and help to understand the genetic control of their tremendous phenotypic diversity.

#### 7.4 Use of cultivated sunflower and CWR resources for environmental adaptation

While sunflower breeding for abiotic tolerance has mainly focused on drought and salt tolerance to date, heat and flooding tolerance are likely to be of increasing importance in the future due to higher temperatures, as well more frequent extreme climate events (Coumou and Rahmstorf, 2012). A recent study of the impacts of heat stress on reproductive traits in cultivated and wild *H. annuus* showed that the latter is much more tolerant of heat stress (Hernández *et al.*, 2018). Surprisingly, the most tolerant populations were from wetter rather than hotter environments, with invasive populations of wild *H. annuus* showing the greatest tolerance. Populations of wild *H. annuus* from wet habitats also exhibit tolerance to flooding (Torres and Diederhufen, 1981). However, considerable variation for flooding tolerance can be found in the cultivated sunflower gene pool, so it is unlikely to be necessary to tap wild resistance alleles for this trait (Gao *et al.*, 2019). Likewise, tolerance to heavy metal soils has been reported in wild species, most notably *H. exilis* (Sambatti and Rice, 2007), but substantial variation already exists among sunflower cultivars (Rizwan *et al.*, 2016). On the other hand, various wild sunflowers from sand dune habitats such as *H. anomalus*, *H. neglectus*, *H. niveus* ssp. *tephrodes*, and dune ecotypes of *H. petiolaris*, are tolerant of low nutrient soils (Donovan *et al.*, 2010), a trait that appears to have been lost in cultivated sunflowers. Introduction of such a trait to cultivated sunflowers could reduce fertilizer usage when sunflower is grown in nutrient poor soils.

#### 7.5 Use of cultivated sunflower and CWR resources in breeding

In the breeding and germplasm enhancement program of the USDA, use of CWR has recently focused on the primary

and secondary gene pools, in particular, the annual wild species and open pollinated varieties. Many of USDA's activities are continuation of previous uses of CWR, mostly centred on diversification of sources of resistance to disease. Introgressions from *H. praecox*, *H. argophyllus*, and *H. petiolaris* have been associated with *Sclerotinia* basal stalk rot resistance (Qi *et al.*, 2016; Talukder *et al.*, 2019) and were combined with downy mildew (caused by *Plasmopara halstedii*) resistance previously recovered from the primary CWR (Hulke *et al.*, 2010). Similar work in diversification of downy mildew resistance sources has come from the secondary CWR pool, as well, with the independent genes *PI18*, *PI20*, and *PI35* recently introgressed from *H. argophyllus* (Qi *et al.*, 2019). New work of similar methodology is ongoing with *Sclerotinia* head rot resistance. Recently, however, USDA has diversified the number of breeding projects that have involved CWR. This year, USDA released two germplasm lines (HA 488 and HA 489) that provided, for the first time, resistance to the red sunflower seed weevil (*Smicronyx fulvus* LeConte) and the banded sunflower moth (*Cochylis hospes* Walsingham), respectively. In seeming contradiction to the idea that such resistance would be derived from germplasm that originates within the range of the pest (Seiler *et al.*, 2017), the resistance was found in open pollinated varieties that were developed in South Eastern Europe (DeGreef *et al.*, 2020; Wronski *et al.*, 2020). However, it is unknown if the European varieties had a recent ancestor from North America, where these pests are common. Occasionally, these resources contain genetic surprises. In the case of the banded sunflower moth resistance work, a source of very low saturated fat composition in the seed oil was also discovered, which was released separately as HOLS4 (Hulke *et al.*, 2017). USDA has also used CWR in recent years to expand the maturity range of the breeding program. CWR resources from the primary gene pool were used to develop early maturing oilseed types for use in Canada and "double crop" systems in the central and southern U.S. (Hulke and May, 2017). Double cropping is the growing of a winter grain, which is promptly harvested, followed by a summer crop, which allows harvesting of two crops in a year. This work is ongoing and was recently expanded to include development of full-season sunflower lines to be used in the central and southern U.S., which are regions with much longer growing seasons than Fargo, ND.

The use of CWR in the sunflower breeding programme at IFVCNS, Serbia, has a long tradition dating back to 1980, when the collection was first established. In the past two decades, a significant number of inbred lines have been created by interspecific crosses. These lines, together with the wild populations, represent a valuable resource of useful alleles that are abundantly used in the IFVCNS breeding program for increasing genetic variability, introducing resistance genes to economically significant pathogens and parasitic plants, increasing tolerance to herbicides and altering the architecture of the plant. Current research on CWR is focused on the discovery of new resistance genes to the more virulent populations of *Orobanche cumana* and *Macrophomina phaseolina* (Tassi) Goid. In order to discover new resistance genes to *Orobanche*, both CWR per se (Terzić *et al.*, 2010; Jocković *et al.*, 2018) and inbred lines of cultivated sunflower originating from interspecific crosses have been tested. As a result, resistance genes to new races of *O. cumana* were recently detected in inbred lines originating from crosses

with *H. deserticola* (Hladni *et al.*, 2012), *H. tuberosus* and *H. divaricatus* (Cvejić *et al.*, 2014). The resistance genes from these sources have been subsequently mapped, and the developed molecular markers will expedite their transfer into commercial lines (Imerovski *et al.*, 2013, 2016, 2019). Interestingly, *Orobanche cumana* is not found in the centre of sunflower origin, but CWR have proven to be a useful source of resistance genes to this parasite which hampers sunflower production across Europe. A similar workflow was used to find resistance to *Macrophomina phaseolina*. First, CWRs were tested per se to narrow down the sources of potential resistance genes that could be used in the breeding program (Tančić *et al.*, 2012). Later, inbred lines originating from interspecific crosses with potential donors were screened to determine if they could be used as a source of tolerance to this pathogen (Tančić Živanov *et al.*, 2020).

Apart from being a reservoir of diverse resistance genes, CWR served as donors of some valuable agronomic traits as well. For example, wild *H. annuus* was a donor of early flowering time locus that enabled research at IFVCNS to develop ultra-early hybrids with vegetation periods of less than 90 days. These types of hybrids are particularly well suited for areas where sunflower is grown as a second crop (*i.e.*, after winter crops). Additionally, the ultra-early hybrids could be critical for sunflower production in northern Europe and Russia, where in the last decade sunflower production has been expanding due to climate change.

## 7.6 Cultivated and wild *H. annuus* vs. CWR

While much of the focus of this paper has been on the benefits of CWRs, which are undeniable, it is important to keep in mind that wild species are genetically and chromosomally divergent from the domesticated sunflower. As a consequence, in addition to beneficial alleles targeted by breeders, introgressions may introduce maladaptive alleles or deleterious structural variants into the cultivated gene pool, a phenomenon known as linkage drag. For example, *H. petiolaris*, which is the wild donor several components of the hybrid production system, differs from the cultivated sunflower by 6–8 translocations and 50–60 inversions (Ostevik *et al.*, 2019). These rearrangements hamper introgression from much of the genome and, if successfully introgressed into cultivars, can introduce genetic load and reduce recombination rates. Gene presence/absence variation, which affects 27% of the genes in the cultivated sunflower pan genome (Hübner *et al.*, 2019), is also often associated with introgressions from wild species (Owens *et al.*, 2019). Such linkage drag could be reduced by focusing pre-breeding efforts on wild *H. annuus*, which is geographically widespread, highly genetically and ecologically variable (Kane and Rieseberg, 2007), and fully interfertile with cultivated sunflower. Kantar *et al.* (2015) showed that wild *H. annuus* encompasses about 60% of the ecological niche space of all species. Thus, when traits required to improve the sunflower crop are not found in cultivated material, the first step should be to search in wild *H. annuus*. Studies of the other *Helianthus* species will be most important for research concerning characters for which they are the only source and to understand evolution and adaptation of the *Helianthus* genome.

## Supplementary material

Supplementary data, references and Tables S1–S5

The Supplementary Material is available at <https://www.ocl-journal.org/10.1051/oc/2020004/olm>.

*Acknowledgements.* We would like to thank Patrick Vincourt for critical reading of the manuscript.

## References

- Al-Khatib K, Baumgartner JR, Peterson DE, Currie RS. 1998. Imazethapyr resistance in common sunflower (*Helianthus annuus*). *Weed Sci* 46: 403–407.
- Alonso LC, Rodriguez-Ojeda MI, Fernandez-Escobar J, Lopez-Calero G. 1998. Chemical control of broomrape (*Orobanche cernua* Loefl.) in sunflower (*Helianthus annuus* L.) resistant to imazethapyr herbicide. *Helia* 21: 45–54.
- Anderson PK, Cunningham AA, Patel NG, Morales FJ, Epstein PR, Daszak P. 2004. Emerging infectious diseases of plants: pathogen pollution, climate change and agrotechnology drivers. *Trends Ecol Evol* 19(10): 535–544.
- Atlagić J. 2004. Roles of interspecific hybridization and cytogenetic studies in sunflower breeding. *Helia* 27(41): 1–24.
- Atlagić J, Terzić S. 2015. The challenges of maintaining a collection of wild sunflower (*Helianthus*) species. *Genet Resour Crop Evol* 63: 1219–1236.
- Atlagić J, Terzić S, Škorić D, Marinković R, Vasiljević Lj, Panković D. 2006. The wild sunflowers collection in Novi Sad. *Helia* 29 (44): 55–64.
- Atlagić J, Terzić S, Marinković R, Jocić S, Miklič V. 2012. Feasibility of keeping F1 interspecific sunflower hybrids. Proc. 18th International sunflower conference, Mar Del Plata & Balcarce, Argentina. February 27–March 1, 2012, pp. 588–593.
- Badouin H, Gouzy J, Grassa CJ, *et al.* 2017. The sunflower genome provides insights into oil metabolism, flowering and Asterid evolution. *Nature* 546(7656): 148–152. Available from <https://doi.org/10.1038/nature22380>. Epub 2017 May 22.
- Baldini M, Vannozzi GP. 1998. Agronomic and physiological assessment of genotypic variation for drought tolerance in sunflower genotypes obtained from a cross between *H. annuus* and *H. argophyllus*. *Agric Med* 128: 232–240.
- Baldini M, Ceconi F, Vannozzi GP. 1993. Influence of water deficit on gas exchange and dry matter accumulation in sunflower cultivars and wild species (*Helianthus argophyllus* T. & G.). *Helia* 16: 1–10.
- Baute GJ, Owens GL, Bock DG, Rieseberg LH. 2016. Genome-wide genotyping-by-sequencing data provide a high-resolution view of wild *Helianthus* diversity, genetic structure, and interspecies gene flow. *Am J Bot* 103: 2170–2177. Available from <https://doi.org/10.3732/ajb.1600295>.
- Bock DG, Kantar M, Caseys C, Matthey-Doret R, Rieseberg LH. 2018. Evolution of invasiveness by genetic accommodation. *Nat Ecol Evol* 2: 991–999.
- Bowsher A, Milton E, Donovan L. 2016. Comparison of desert-adapted *Helianthus niveus* (Benth.) Brandegees ssp. *tephrodes* (A. Gray) Heiser to cultivated *H. annuus* L. for putative drought avoidance traits at two ontogenetic stages. *Helia* 39(64): 1–19.
- Cabrera-Bosquet L, Crossa J, von Zitzewitz J, Serret MD, Luis Araus J. 2012. High-throughput phenotyping and genomic selection: the

- frontiers of crop breeding converge F. *J Integrative Plant Biol* 54: 312–320.
- Cadic E, Coque M, Vear F, *et al.* 2012. Combined linkage and association mapping of flowering time in Sunflower (*Helianthus annuus* L.). *Theor Appl Genet* (2013) 126: 1337–1356. Available from <https://doi.org/10.1007/s00122-013-2056-2>.
- Camino C, González-Dugo V, Hernández P, Sillero JC, Zarco-Tejada PJ. 2018. Improved nitrogen retrievals with airborne-derived fluorescence and plant traits quantified from VNIR-SWIR hyperspectral imagery in the context of precision agriculture. *Int J Appl Earth Observ Geoinfo* 70: 105–117.
- Campbell BT, Saha S, Percy R, *et al.* 2010. Status of the global cotton germplasm resources. *Crop Sci* 50: 1161–1179.
- Casadebaig P, Guilioni L, Lecoœur J, Christophe A, Champolivier L, Debaeke P. 2011. SUNFLO, a model to simulate genotype-specific performance of the sunflower crop in contrasting environments. *Agricult Forest Meteorol* 151: 163–178.
- Celik I, Bodur S, Frary A, Doganlar S. 2016. Genome-wide SNP discovery and genetic linkage map construction in sunflower (*Helianthus annuus* L.) using a genotyping by sequencing (GBS) approach. *Mol Breed* 36: 133. Available from <https://doi.org/10.1007/s11032-016-0558-8>.
- Cobb JN, DeClerck G, Greenberg A, Clark R, McCouch S. 2013. Next-generation phenotyping: requirements and strategies for enhancing our understanding of genotype–phenotype relationships and its relevance to crop improvement. *Theor Appl Genet* 126: 867–887. DOI: [10.1007/s00122-013-2066-0](https://doi.org/10.1007/s00122-013-2066-0).
- Coque M, Mesnildrey S, Romestant M, *et al.* 2008. Sunflower line core collections for association studies and phenomics. Proc 17th Int Sunflower Conf, 8-12/6/2008 Cordoba, Spain, 2008, pp. 725–728.
- Coumou D, Rahmstorf S. 2012. A decade of weather extremes. *Nat Clim Change* 2: 491–496.
- Čuk L. 1982. Variability in oil content in seed of *Helianthus* spp. Proc. 10th International sunflower conference, Surfers Paradise, Australia, 1982, p. 211.
- Cvejić S, Jocić S, Dedić B, Radeka I, Imerovski I, Miladinović D. 2014. Determination of resistance to broomrape in newly developed sunflower inbred lines. p. 184–188. In: Proc. 3rd Int. Symp. on Broomrape (Orobanchae spp.) in Sunflower, Córdoba, Spain, 2014.
- DeGreef MG, Prasifka JR, Koehler BD, Hulke BS. 2020. Registration of oilseed sunflower maintainer germplasm HA 488, with resistance to the red sunflower seed weevil (*Smicronyx fulvus* LeConte). *J Plant Registrations* (in review).
- Dempewolf H, Baute G, Anderson J, Kilian B, Smith C, Guarino L. 2017. Past and future use of wild relatives in crop breeding. *Crop Sci* 57: 1070–1082.
- Donovan LA, Rosenthal DR, Sanchez-Velenosi M, Rieseberg LH, Ludwig F. 2010. Hybrid species are not necessarily adapted to their current habitats. *J Evol Biol* 23: 805–816.
- Duriez P, Vautrin S, Auriac MC, *et al.* 2019. A receptor-like kinase enhances sunflower resistance to Orobanchae cumana. *Nat Plants* 5: 1211–1215. Available from <https://doi.org/10.1038/s41477-019-0556-z>.
- Elshire RJ, Glaubitz JC, Sun Q, *et al.* 2011. A robust, simple genotyping-by-sequencing (GBS) approach for high diversity species. *PLoS One* 6: e19379. Available from <https://doi.org/10.1371/journal.pone.0019379>.
- Faure N, Serieys H, Cazaux E, Kaan F, Bervillé A. 2002. Partial hybridization in wide crosses between cultivated sunflower and the Perennial *Helianthus* Species *H. mollis* and *H. orgyalis*. *Ann Bot* 89 (1): 31–39. Available from <https://doi.org/10.1093/aob/mcf003>.
- Gao L, Lee JS, Hübner S, Hulke BS, Qu Y, Rieseberg LH. 2019. Genetic and phenotypic analyses indicate that resistance to flooding stress is uncoupled from performance in cultivated sunflower. *New Phytol* 223: 1657–1670.
- Gélard W, Burger P, Casadebaig P, *et al.* 2016. 3D plant phenotyping in sunflower using architecture-based organ segmentation from 3D point clouds, in: 5th International workshop on image analysis methods for the plant sciences, 2016.
- Gélard W, Herbulot A, Devy M, *et al.* 2017. Leaves segmentation in 3d point cloud, in: International conference on advanced concepts for intelligent vision systems. Springer, pp. 664–674.
- Gélard W, Herbulot A, Devy M, Casadebaig P. 2018. 3D leaf tracking for plant growth monitoring, in: 2018 25th IEEE International Conference on Image Processing (ICIP). IEEE, 2018, pp. 3663–3667.
- Gómez-Candón D, Virlet N, Labbé S, Jolivot A, Regnard JL. 2016. Field phenotyping of water stress at tree scale by UAV-sensed imagery: new insights for thermal acquisition and calibration. *Precision Agriculture* 17: 786–800.
- Gonzalez-Dugo V, Hernandez P, Solis I, Zarco-Tejada P. 2015. Using high-resolution hyperspectral and thermal airborne imagery to assess physiological condition in the context of wheat phenotyping. *Remote Sensing* 7: 13586–13605.
- Gosseau F, Blanchet N, Varès D, *et al.* 2019. Heliaphen, an outdoor high-throughput phenotyping platform for genetic studies and crop modeling. *Front Plant Sci* 9. Available from <https://doi.org/10.3389/fpls.2018.01908>.
- Granier C, Aguirrezabal L, Chenu K, *et al.* 2006. PHENOPSIS, an automated platform for reproducible phenotyping of plant responses to soil water deficit in Arabidopsis thaliana permitted the identification of an accession with low sensitivity to soil water deficit. *New Phytol* 169: 623–635.
- Hajjar R, Hodgkin T. 2007. The use of wild relatives in crop improvement: a survey of developments over the last 20 years. *Euphytica* 156: 1–13.
- Hernández FA, Poverene M, Presotto A. 2018. Heat stress effects on reproductive traits in cultivated and wild sunflower (*Helianthus annuus* L.): evidence for local adaptation within the wild germplasm. *Euphytica* 214: 1–15.
- Hesley J. 1999. The economic impact of weeds, diseases, and insects on world sunflower production. *Sunflower Mag* 25(4).
- Hladni N. 2016. Present status and future prospects of global confectionary sunflower production. Proc. 19th Int. Sunflower Conf. Edirne, Turkey, 2016, pp. 47–60.
- Hladni N, Dedić B, Jocić S, Miklič V, Dušanić N. 2012. Evaluation of resistance of new sunflower hybrids to broomrape in the breeding programs in Novi Sad. *Helia* 35(56): 89–98.
- Hladni N, Zorić M, Terzić S, *et al.* 2018. Comparison of methods for the estimation of best parent heterosis among lines developed from interspecific sunflower germplasm. *Euphytica* 214(7): 108.
- Holden J, Peacock J, Williams T. 1993. Genes, crops, and the environment. New York: Cambridge Press.
- Hübner S, Bercovich N, Todesco M, *et al.* 2019. Sunflower pan-genome analysis shows that hybridization altered gene content and disease resistance. *Nature Plants* 5: 54–62.
- Hulke BS, May WE. 2017. Registration of oilseed sunflower restorer germplasms RHA 476 and RHA 477, adapted for short season environments. *J Plant Registrations* 12: 148–151.
- Hulke BS, Miller JF, Gulya TJ, Vick BA. 2010. Registration of the oilseed sunflower genetic stocks HA 458, HA 459, and HA 460 possessing genes for resistance to downy mildew. *J Plant Registrations* 4(1): 93–97.
- Hulke BS, Gao QM, Foley ME. 2017. Registration of the sunflower oilseed maintainer genetic stocks HOLS1, HOLS2, HOLS3, and

- HOLS4 possessing genes for high oleic and low saturated fatty acids and tolerance to imidazolinone herbicides. *J Plant Registrations* 11: 200–203.
- Hunter DV, Heywood H. 2011. Crop wild relatives, a manual of in situ conservation. Earthscan, London, 411 p.
- Imerovski I, Dimitrijević A, Miladinović D, *et al.* 2013. Identification of PCR markers linked to different or genes in sunflower. *Plant Breed* 132: 115–120.
- Imerovski I, Dimitrijević A, Miladinović D, *et al.* 2016. Mapping of a new gene for resistance to broomrape races higher than F. *Euphytica* 209: 281–289.
- Imerovski I, Dedić B, Cvejić S, *et al.* 2019. BSA-seq mapping reveals major QTL for broomrape resistance in four sunflower lines. *Mol Breed* 39: 41. Available from <https://doi.org/10.1007/s11032-019-0948-9>.
- Jocković M, Jocić S, Cvejić S, *et al.* 2018. *Helianthus* species as a source for broomrape resistance. Proc. 4th Int. Symp. Broomrape in Sunflower, Bucharest, Romania, 2–4 July, 2018, pp. 178–186.
- Kane NC, Rieseberg LH. 2007. Selective sweeps reveal candidate genes for adaptation to drought and salt tolerance in common sunflower, *Helianthus annuus*. *Genetics* 175: 1803–1812.
- Kane NC, Gilla N, King MG, *et al.* 2011. Progress towards a reference genome for sunflower. *Botany* 89: 429–437.
- Kantar MB, Sosa CC, Khoury CK, *et al.* 2015. Ecogeography and utility to plant breeding of the crop wild relatives of sunflower (*Helianthus annuus* L.). *Front Plant Sci* 6, article 841.
- Lazarević J, Luković J, Terzić S, *et al.* 2016. Micro-morphological achene features of annual species of wild sunflower. *Matica Srpska J Nat Sci Novi Sad* 131: 73–80.
- Leclercq P. 1969. Une stérilité mâle cytoplasmique chez le tournesol. *Ann Amélior Pl* 19: 99–106 (available at [www.isasunflower.org/publications](http://www.isasunflower.org/publications) – older publications).
- Lenz-Wiedemann VIS, Klar CW, Schneider K. 2010. Development and test of a crop growth model for application within a Global Change decision support system. *Ecol Model* 221: 314–329.
- Liebisch F, Kirchgessner N, Schneider D, Walter A, Hund A. 2015. Remote, aerial phenotyping of maize traits with a mobile multi-sensor approach. *Plant Methods* 11: 9. Available from <https://doi.org/10.1186/s13007-015-0048-8>.
- Livaja M, Unterseer S, Erath W, *et al.* 2016. Diversity analysis and genomic prediction of *Sclerotinia* resistance in sunflower using a new 25 K SNP genotyping array. *Theor Appl Genet* 129: 317–329. Available from <https://doi.org/10.1007/s00122-015-2629-3>.
- Louarn J, Boniface MC, Pouilly N, *et al.* 2016. Sunflower resistance to broomrape (*Orobanche cumana*) is controlled by specific QTLs for different parasitism stages. *Front Plant Sci* 7: 590. Available from <https://doi.org/10.3389/fpls.2016.00590>.
- Lu F, Romay MC, Glaubitz JC, *et al.* 2015. High-resolution genetic mapping of maize pan-genome sequence anchors. *Nat Commun* 6: 6914. Available from <https://doi.org/10.1038/ncomms7914>.
- Mabire C, Duarte J, Darracq A, *et al.* 2019. High throughput genotyping of structural variations in a complex plant genome using an original Affymetrix® axiom® array. *BMC Genom* 20: 848. Available from <https://doi.org/10.1186/s12864-019-6136-9>.
- Madec S, Baret F, De Solan B, *et al.* 2017. High-throughput phenotyping of plant height: comparing unmanned aerial vehicles and ground LiDAR estimates. *Front Plant Sci* 8: 2002.
- Mandel JR, Dechaine JM, Marek LF, Burke JM. 2011. Genetic diversity and population structure in cultivated sunflower and a comparison to its wild progenitor, *Helianthus annuus* L. *Theor Appl Genet* 123: 693–704. PMID 21638000. Available from <https://doi.org/10.1007/s00122-011-1619-3>.
- Mandel JR, Nambesan S, Bowers JE, *et al.* 2013. Association mapping and the genomic consequences of selection in sunflower. *Plos Genet* 9: e1003378. PMID 23555290. Available from <https://doi.org/10.1371/journal.pgen.1003378>.
- Mangin B, Pouilly N, Boniface MC, *et al.* 2017. Molecular diversity of sunflower populations maintained as genetic resources is affected by multiplication processes and breeding for major traits. *Theor Appl Genet* 130: 1099. Available from <https://doi.org/10.1007/s00122-017-2872-x>.
- Martin M, Molfetta P, Vannozzi GP, Zerbi G. 1992. Mechanisms of drought resistance of *Helianthus annuus* and *H. argophyllus*. Proc. 13th Int. Sunflower Conf., Pisa, Italy, 1992, pp. 571–586.
- Maviane-Macia F, Ribeyre C, Buendia L, *et al.* 2019. Experimental system and image analysis software for high throughput phenotyping of mycorrhizal growth response in *Brachypodium distachyon*. *bioRxiv* 779330. Available from <https://doi.org/10.1101/779330>.
- Mihaljčević M. 1988. Combining ability and heterosis in *H. annuus* x *H. annuus* (wild) crosses. Proc. 12th International sunflower conference, Novi Sad, Yugoslavia, 2, 1988, pp. 494–495.
- Miller JF. 1995. Inheritance of salt tolerance in sunflower. *Helia* 18: 9–16.
- Miller JF, Seiler GJ. 2003. Registration of five oilseed maintainer (HA 429-HA 433) sunflower germplasm lines. *Crop Sci* 43: 2313–2314.
- Narum SR, Buerkle CA, Davey JW, Miller MR, Hohenlohe PA. 2013. Genotyping-by-sequencing in ecological and conservation genomics. *Mol Ecol* 22: 2841–2847. Available from <https://doi.org/10.1111/mec.12350>.
- Ortiz R. 2015. The importance of crop wild relatives, diversity, and genetic potential for adaptation to abiotic stress-prone environments. In: Redden R, Yadav SS, Maxted N, Dulloo ME, Guarino L, Smith P, eds. Crop wild relatives and climate change. John Wiley & Sons, pp. 80–87.
- Ostevik KL, Samuk K, Rieseberg LH. 2019. Ancestral reconstruction of sunflower karyotypes reveals dramatic chromosomal evolution. *bioRxiv* preprint. Available from <https://doi.org/10.1101/737155>.
- Owens GL, Baute GJ, Hübner S, Rieseberg LH. 2019. Genomic sequence and copy-number evolution during hybrid crop development in sunflowers. *Evol Appl* 12: 54–65.
- Palmgren MG, Edenbrandt AK, Vedel SE, *et al.* 2015. Are we ready for back-to-nature crop breeding? *Trends Plant Sci* 20(3): 155–164.
- Pecrix Y, Penouilh-Suzette C, Muñoz S, Vear F, Godiard L. 2018. Ten broad spectrum resistances to downy mildew physically mapped on the sunflower genome. *Front Plant Sci* 9: 1780. Available from <https://doi.org/10.3389/fpls.2018.01780>.
- Prescott-Allen CP, Prescott-Allen R. 1986. The first resource: wild species in the North American economy. London, UK: Yale Univ. Press, 529 p.
- Putt ED. 1964. Sunflower breeding in Canada. In: Proc. 1st Int. Sunflower Conference, College Station, Texas, USA, June 17–18, 1964, available at: [www.isasunflower.org/publications](http://www.isasunflower.org/publications).
- Qi LL, Long Y, Taluker ZI, Seiler GJ, Block CC, Gulya TJ. 2016. Genotype-by-sequencing uncovers the introgression alien segments associated with *Sclerotinia* basal stalk rot resistance from wild species – I. *Helianthus argophyllus* and *H. petiolaris*. *Front Genet* 7: 219.
- Qi LL, Ma GJ, Li XH, Seiler GJ. 2019. Diversification of the downy mildew resistance gene pool by introgression of a new gene, Pl 35, from wild *Helianthus argophyllus* into oilseed and confection sunflower (*Helianthus annuus* L.). *Theoretical Appl Genet* 132: 2553–2565.
- Qiu F, Baack EJ, Whitney KD, *et al.* 2018. Phylogenetic trends and environmental correlates of nuclear genome size variation in *Helianthus* sunflowers. *New Phytol* 221: 1609–1618. Available from <https://doi.org/10.1111/nph.15465>.

- Qiu Q, Sun N, Wang Y, *et al.* 2019. Field-based high-throughput phenotyping for maize plant using 3D LiDAR point cloud generated with a “phenomobile.” *Front Plant Sci* 10: 554.
- Rauf S, Jamil N, Ali Tariq S, Khan M, Kausar M. 2017. Progress in modification of sunflower oil to expand its industrial value. *J Sci Food Agric* 97: 1997–2006.
- Rizwan M, Ali S, Rizvi H, *et al.* 2016. Phytomanagement of heavy metals in contaminated soils using sunflower – A review. *Crit Rev Environ Sci Technol*. Available from <https://doi.org/10.1080/10643389.2016.1248199>.
- Saftić-Panković D, Atlagić J, Miljanović T, Radovanović N. 2005. Morphological and molecular variability of *Helianthus giganteus* L. and *Helianthus maximiliani* Sch. species. *Genetika* 37: 121–130.
- Sambatti JB, Rice KJ. 2007. Functional ecology of ecotypic differentiation in the Californian serpentine sunflower (*Helianthus exilis*). *The New Phytologist* 175: 107–119.
- Saxena RK, Edwards D, Varshney RK. 2014. Structural variations in plant genomes. *Brief Funct Genom* 13: 296–307. Available from <https://doi.org/10.1093/bfgp/elu016>.
- Schilling EE. 2006. *Helianthus*. In: Flora of North America Editorial Committee, editors, Flora of North America north of Mexico. Vol. 21. New York and Oxford: Oxford Univ. Press, pp. 141–169.
- Seiler GJ. 2010. Utilization of wild *Helianthus* species in breeding for disease resistance. Proc. I.S.A. symposium “Sunflower breeding on resistance to diseases”, Krasnodar, Russia, 2010, pp. 36–50.
- Seiler GJ. 2012. Utilization of wild *Helianthus* species in sunflower breeding. In: Škorić D, ed. Sunflower genetics and breeding international monogram. Novi Sad, Serbia: Serbian Academy of Sciences and Arts, pp. 355–413.
- Seiler GJ, Qi LL, Marek LF. 2017. Utilization of sunflower crop wild relatives for cultivated sunflower improvement. *Crop Sci* 57: 1083–1101.
- Serieys H, Christov M. 2005. European Cooperative Research Network on sunflower, FAO working group: “Identification, study and utilization in breeding programs of new CMS sources”, Progress Report 1999–2004. X Consultation Meeting, Novi Sad, Serbia, 2005, 80 p.
- Škorić D. 2008. Report on plant breeding and related biotechnology capacity in Serbia. Global partnership initiative for plant breeding capacity building (GIPB), Global crop diversity trust (GCDT), pp 1–49. <http://www.fao.org/in-action/plant-breeding/our-partners/europe/serbia/en/>.
- Škorić D. 2012. Sunflower breeding. sunflower genetics and breeding international monogram. Novi Sad, Serbia: Serbian Acad. Sciences and Arts, pp. 165–354.
- Škorić D, Rajčan I. 1992. Breeding for Sclerotinia tolerance in sunflower. Proc. 13th International sunflower conference, Pisa, Italy, 2, 1992, pp. 1257–1262.
- Škorić D, Marinković R, Atlagić J. 1988. Determination of restorer genes for sources of cytoplasmic male sterility in wild sunflower species. Proc. 12th International sunflower conference, Novi Sad, Yugoslavia, 2, 1988, pp. 282–286.
- Stebbins JC, Winchell CJ, Constable JVH. 2013. *Helianthus winteri* (Asteraceae), a new perennial species from the southern Sierra Nevada foothills, California. *Aliso* 31: 19–24.
- Steduto P, Hsiao TC, Raes D, Fereres E. 2009. AquaCrop—The FAO crop model to simulate yield response to water: I. Concepts and underlying principles. *Agronomy J* 101: 426–437.
- Talukder ZI, Long YM, Seiler GJ, Underwood W, Qi LL. 2019. Introgression and monitoring of wild *Helianthus praecox* alien segments associated with Sclerotinia basal stalk rot resistance in sunflower using genotyping-by-sequencing. *Plos One* 14: e0213065.
- Tančić S, Terzić S, Dedić B, Atlagić J, Jocić S, Miklič V. 2012. Disease resistance of wild sunflower species to *Macrophomina Phaseolina*. Proc. 18th International sunflower conference, Mar Del Plata & Balcarce, Argentina, February 27–March 1, 2012, pp. 304–308.
- Tančić Živanov S, Dedić B, Zorić M, Cvejić S, Jocić S, Miklič V. 2020. Screening of sunflower genotypes for charcoal rot (*Macrophomina phaseolina* (Tassi) Goid.) Tolerance under the field conditions. *J Plant Dis Protection* (in review).
- Terzić S. 2018. The longevity of annual wild *Helianthus* seeds in short to medium-term storage conditions. Serbian Plant Physiology Society, 3rd International conference on plant biology (22nd SPPS meeting), June 9–12th, Belgrade, Serbia, 2018, p. 95.
- Terzić S, Atlagić J, Panković D. 2006. Characterization of F1 interspecific hybrids between wild *Helianthus annuus* L. populations and cultivated sunflower. *Genetika* 38(2): 159–168.
- Terzić S, Dedić B, Atlagić J, Jocić S, Tančić S. 2010. Screening wild sunflower species and F1 interspecific hybrids for resistance to broomrape. *Helia* 33(53): 25–30.
- Terzić S, Zorić M, Seiler G. 2019. Qualitative traits in sunflower breeding: UGA-SAM1 phenotyping case study. *Crop science—First Look*. Available from <https://doi.org/10.2135/cropsci2019.02.0112>.
- Todesco M, Owens GL, Bercovich N, *et al.* 2019. Massive haplotypes underlie ecotypic differentiation in sunflowers. *bioRxiv* 790279. Available from <https://doi.org/10.1101/790279>.
- Torres AM, Diedenhofen U. 1981. Sunflower alcohol-dehydrogenase genotypes – germination rates and response to flooding. *Environ Exp Botany* 21: 35–44.
- Twyford AD. 2018. The road to 10,000 plant genomes. *Nat Plants* 4: 312–313. Available from <https://doi.org/10.1038/s41477-018-0165-2>.
- Tyack N, Dempewolf H. 2015. The economics of crop wild relatives under climate change. In: Redden R, Yadav SS, Maxted N, Dulloo ME, Guarino L, Smith P, eds. Crop wild relatives and climate change. John Wiley and Sons, pp. 281–291.
- USDA, Foreign Agricultural Service. 2019. World agricultural production: Sunflower seed Area, Yield, and Production. *Circular Series WAP* 7–19: 26. Available: <https://apps.fas.usda.gov/psdonline/circulars/production.pdf>.
- Wronski AR, Prasifka JR, Grove MS, *et al.* 2020. Registration of oilseed sunflower maintainer germplasm HA 489, with resistance to the banded sunflower moth (*Cochylis hospes* Walsingham). *Journal of Plant Registrations* (in review).
- Zhao C, Zhang Y, Du J, *et al.* 2019. Crop phenomics: current status and perspectives. *Front Plant Sci* 10. Available from <https://doi.org/10.3389/fpls.2019.00714>.

**Cite this article as:** Terzić S, Boniface M-C, Marek L, Alvarez D, Baumann K, Gavrilova V, Joita-Pacureanu M, Sujatha M, Valkova D, Velasco L, Hulke BS, Jocić S, Langlade N, Muñoz S, Rieseberg L, Seiler G, Vear F. 2020. Gene banks for wild and cultivated sunflower genetic resources. *OCL* 27: 9.