



HAL
open science

The sugar transporter SWEET10 acts downstream of FLOWERING LOCUS T during floral transition of *Arabidopsis thaliana*

Fernando Andrés, Atsuko Kinoshita, Naveen Kalluri, Virginia Fernández, Vítor S Falavigna, Tiago M D Cruz, Seonghoe Jang, Yasutaka Chiba, Mitsunori Seo, Tabea Mettler-Altmann, et al.

► To cite this version:

Fernando Andrés, Atsuko Kinoshita, Naveen Kalluri, Virginia Fernández, Vítor S Falavigna, et al.. The sugar transporter SWEET10 acts downstream of FLOWERING LOCUS T during floral transition of *Arabidopsis thaliana*. *BMC Plant Biology*, 2020, 20 (1), 10.1186/s12870-020-2266-0. hal-03129127

HAL Id: hal-03129127

<https://hal.inrae.fr/hal-03129127>

Submitted on 2 Feb 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.






Distributed under a Creative Commons Attribution 4.0 International License

RESEARCH ARTICLE

Open Access



The sugar transporter SWEET10 acts downstream of *FLOWERING LOCUS T* during floral transition of *Arabidopsis thaliana*

Fernando Andrés^{1,2*†}, Atsuko Kinoshita^{1†}, Naveen Kalluri¹, Virginia Fernández^{1,3}, Vítor S. Falavigna¹, Tiago M. D. Cruz¹, Seonghoe Jang^{1,4} , Yasutaka Chiba⁵, Mitsunori Seo⁵, Tabea Mettler-Altmann⁶, Bruno Huettel¹  and George Coupland^{1*} 

Abstract

Background: Floral transition initiates reproductive development of plants and occurs in response to environmental and endogenous signals. In *Arabidopsis thaliana*, this process is accelerated by several environmental cues, including exposure to long days. The photoperiod-dependent promotion of flowering involves the transcriptional induction of *FLOWERING LOCUS T* (*FT*) in the phloem of the leaf. *FT* encodes a mobile protein that is transported from the leaves to the shoot apical meristem, where it forms part of a regulatory complex that induces flowering. Whether *FT* also has biological functions in leaves of wild-type plants remains unclear.

Results: In order to address this issue, we first studied the leaf transcriptomic changes associated with *FT* overexpression in the companion cells of the phloem. We found that *FT* induces the transcription of *SWEET10*, which encodes a bidirectional sucrose transporter, specifically in the leaf veins. Moreover, *SWEET10* is transcriptionally activated by long photoperiods, and this activation depends on *FT* and one of its earliest target genes *SUPPRESSOR OF CONSTANS OVEREXPRESSION 1* (*SOC1*). The ectopic expression of *SWEET10* causes early flowering and leads to higher levels of transcription of flowering-time related genes in the shoot apex.

Conclusions: Collectively, our results suggest that the *FT*-signaling pathway activates the transcription of a sucrose uptake/efflux carrier during floral transition, indicating that it alters the metabolism of flowering plants as well as reprogramming the transcription of floral regulators in the shoot meristem.

Keywords: Flowering time, *FLOWERING LOCUS T*, Photoperiod, Sugar transporter

Background

In plants, the transition from vegetative growth to flowering is regulated by several environmental and endogenous stimuli. This complexity is conferred by a network of genetic pathways that has been characterized in most detail in the model species *A. thaliana*. This network includes the vernalisation, gibberellin, thermosensory, age, sugar and photoperiod-dependent pathways [1–3]. *FLOWERING LOCUS T* (*FT*) is a positive regulator of flowering whose expression leads to rapid

transcriptional reprogramming of the meristem associated with inflorescence and flower development, and is often described as a floral integrator because its transcription is activated by several genetic pathways that promote flowering [1, 4, 5]. Sugars such as sucrose and trehalose-6-phosphate also promote flowering, and there is evidence that these act both upstream and downstream of *FT* in the flowering process [6–9]. However, our understanding of the relationship between *FT* and sugar metabolism is fragmentary. Here, we demonstrate that *FT* is required for the transcriptional activation of a gene encoding a sugar uptake/efflux carrier in the vasculature of the leaf and at the shoot apex during floral transition, providing a specific link between *FT* function and sugar transport.

* Correspondence: fernando.andres-lalaguna@inrae.fr; coupland@mpipz.mpg.de

†Fernando Andrés and Atsuko Kinoshita contributed equally to this work.

¹Max Planck Institute for Plant Breeding Research, Carl-von-Linne-Weg 10, 50829 Köln, Germany

Full list of author information is available at the end of the article



FT was first placed within the photoperiodic flowering pathway of *A. thaliana* based on physiological and genetic analyses [10]. Furthermore, simultaneous loss-of-function of *FT* and its closest relative *TWIN SISTER OF FT* (*TSF*) leads to late flowering plants under long days (LDs) that are almost insensitive to photoperiod [11, 12]. Transcription of *FT* is induced by exposure to LDs downstream of the *GIGANTEA* (*GI*) and *CONSTANS* (*CO*) genes in specialized companion cells of the phloem [13–16]. *FT* encodes a small globular protein that shares high homology with mammalian phosphatidylethanolamine-binding proteins (PEBP) [4, 5], and is a major component of a systemic signal that induces flowering in response to photoperiod (a “florigen”) [17–20]. *FT* protein moves through the phloem to the shoot apical meristem (SAM) by an active mechanism [21, 22] and binds lipids in vitro [23]. An endoplasmic reticulum-membrane protein, *FT-INTERACTING PROTEIN 1* (*FTIP1*), interacts with *FT* in companion cells of the phloem and mediates its export into sieve elements [21]. In the sieve elements, *FT* interacts with a heavy metal-associated isoprenylated plant protein called *SODIUM POTASSIUM ROOT DEFECTIVE 1* (*NaKR1*), which regulates the long-distance transport of *FT* to the SAM [22]. In the SAM, *FT* is proposed to interact with two bZIP transcription factors (*FD* and *FD PARALOG* [*FDP*]) [24–26]. The transcriptional complex that is formed between *FT*, these bZIPs and 14–3-3 proteins is proposed to trigger transcriptional activation of genes that promote flowering, such as *SUPPRESSOR OF OVEREXPRESSION OF CONSTANS 1* (*SOC1*), *FRUIT-FULL* (*FUL*) and *APETALA1* (*API*), which encode MADS-box transcription factors, and several members of the *SQUAMOSA PROMOTER BINDING LIKE* (*SPL*) gene family [24, 25, 27–30]. Transcriptomic and in situ hybridization studies identified *SOC1* mRNA as the earliest activated transcript detected in the SAM during *FT*-mediated photoperiodic induction of flowering [28, 29, 31–33], while genome-wide experiments showed that *SOC1* binds to the promoters of numerous genes involved in the floral transition and floral meristem identity [34, 35]. Therefore, *SOC1* acts as an intermediate component in the *FT*-signaling pathway during the activation of flowering of *A. thaliana*. Consistently, the flowering response to *FT* overexpression is attenuated in the *soc1* single mutant [36], and this effect is even more pronounced in the *soc1 ful* double mutant [29, 36].

Ectopic expression of *FT* from heterologous promoters leads to early flowering [4, 5]. For example, overexpression of *FT* from constitutive promoters such as the *Cauliflower mosaic virus CaMV 35S* promoter [p35S] [4, 5] or phloem-specific promoters such as those of the *GALACTINOL SYNTHASE1* [*GAS1*] and *SUCROSE TRANSPORTER2* [*SUC2*] genes [17, 19, 20, 37] induces early flowering of *A. thaliana*. This effect is highly conserved

among Angiosperms, so that overexpression of *FT* or its homologues causes early flowering in a wide range of species [18, 38, 39]. Overexpression of *FT* also induces the transcription of *FUL* and *SEPALLATA3* (*SEP3*) in leaves of *A. thaliana*, conferring changes in leaf morphology that are suppressed by *ful* and *sep3* mutations [40]. Thus, at least when overexpressed, *FT* can influence the development of leaves by affecting the expression of regulatory genes, and upon transport from the leaves it promotes the floral transition at the SAM.

Here, we have further studied the regulatory role of *FT*. We analyzed global transcriptomic changes in leaves associated with the specific expression of *FT* in the phloem companion cells. Our results indicate that *FT* promotes the expression of *SWEET10*, a gene encoding a sucrose bidirectional transporter, in the leaf veins and at the shoot apex. This effect is also mediated by photoperiod and by *SOC1*. Moreover, the overexpression of *SWEET10* slightly accelerates flowering, leading us to discuss possible roles for this gene during floral transition mediated by the *FT*-signaling pathway in *A. thaliana*.

Results

FT induces the expression of *SWEET10*

The global effects on gene expression of *FT* overexpression in the phloem companion cells of the leaves were examined. To this end, transgenic *A. thaliana* plants that overexpress *FT* from the *pGAS1* promoter in a *ft-10 tsf-1* double mutant background were employed (*pGAS1:FT ft-10 tsf-1*). In these transgenic plants, the use of the *pGAS1* promoter ensures that the *FT* transgene is expressed in phloem companion cells of the minor veins, recreating the spatial pattern of expression described for the native gene [15]. Indeed, the overexpression of *FT* from the *pGAS1* promoter complements the late-flowering phenotype of *ft-10 tsf-1* double mutants [11, 17]. The transcriptome of leaves of *pGAS1:FT ft-10 tsf-1* transgenic plants was compared to that of *ft-10 tsf-1* and Col-0 plants using Tiling Arrays. Bioinformatic analysis showed that 699 genes (*p*-Value ≤ 0.01) were differentially expressed between *pGAS1:FT ft-10 tsf-1* and *ft-10 tsf-1* (Additional file 1). A final list of 14 genes (Table 1) was selected by applying more restrictive statistical criteria (adj. *P* Value ≤ 0.05). The majority of these genes were well-known flowering-related regulators that act downstream of *FT* [28, 29], such as *SOC1*, *FUL*, *SEPALLATA1* (*SEPI*) and *SEP3*, which were up-regulated in *pGAS1:FT ft-10 tsf-1*. In addition, *SWEET10* and *SWEET13*, two members of Clade III of the *SWEET* family that encode sucrose transporters [41], were in the list of genes induced by *FT* (Table 1). In particular, the mRNA levels of *SWEET10* were strongly up-regulated in the *pGAS1:FT ft-10 tsf-1* and other genetic backgrounds

Table 1 Top 14 differentially expressed genes between p*GAS1:FT ft-10 tsf-1* and *ft-10 tsf-1*

ID	logFC	P. Value	adj.P.Val	Gene Name
AT1G65480	6,03	4E-10	7,8E-06	<i>FT</i> (FLOWERING LOCUS T)
AT5G60910	2,83	2E-09	2,5E-05	<i>FUL</i> (FRUITFULL)
AT1G24260	2,66	8E-09	6E-05	<i>SEP3</i> (SEPALLATA3)
AT5G50790	2,28	2E-07	0,00144	<i>SWEET10</i>
AT2G45660	2,25	5E-10	7,8E-06	<i>SOC1</i> (SUPPRESSOR OF OVEREXPRESSION OF CO 1)
AT5G15800	1,92	4E-06	0,01516	<i>SEP1</i> (SEPALLATA1)
AT1G80130	1,24	2E-05	0,03677	Tetratricopeptide repeat (TPR)-like superfamily protein
AT5G50800	1,09	8E-06	0,02541	<i>SWEET13</i>
AT3G58200	0,82	1E-05	0,02772	<i>TRAF</i> -like family protein
AT3G56080	0,78	2E-05	0,04354	S-adenosyl-L-methionine-dependent methyltransferases superfamily protein
AT1G62290	-1,18	8E-06	0,02541	<i>PASPA2</i> (PUTATIVE ASPARTIC PROTEINASE A2)
AT5G44400	-1,31	1E-05	0,02772	<i>ATBBE26</i> (BERBERINE BRIDGE ENZYME-LIKE 26)
AT5G23020	-1,52	3E-06	0,01516	<i>IMS2/MAM-L/MAM3</i> (METHYLTHIOALKYMALATE SYNTHASE-LIKE)
AT2G42540	-3,21	9E-06	0,02541	<i>COR15A</i> (COLD-REGULATED 15A)

overexpressing *FT* (Fig. 1a). The expression of *SWEET10* mRNA was clearly induced in plants overexpressing *FT* from companion cell specific promoters p*GAS1* and p*SUC2* [42, 43] (Fig. 1a). This experiment demonstrated that in all cases tested, *FT* overexpression increased transcription of *SWEET10* mRNA. Additionally, in silico gene co-expression analyses indicated that *FT* is highly co-regulated with *SWEET10* during Arabidopsis development (Additional file 2: Figure S1A). These analyses also showed that *SWEET10* is co-expressed with many other genes that are regulated by *FT* (Additional file 2: Figure S1B), such as *SEP3* and *APETALA1* (*API*) [40].

***FT* promotes the expression of *SWEET10* mRNA in the leaf veins**

Next, *FT*-mediated spatial and temporal regulation of *SWEET10* mRNA expression was characterized. The levels of *SWEET10* mRNA in leaves and hand-dissected shoot apices (containing SAM, a segment of the apical stem and young leaves) were quantified by RT-qPCR. As shown in Fig. 1b, *SWEET10* expression levels were higher in shoot apices compared to mature leaves and this difference was more pronounced in p*GAS1:FT ft-10 tsf-1* plants. To better characterize the spatial pattern of expression of *SWEET10*, we fused the 3 Kb region 5' of the translational start codon to a *GUS* reporter gene to create p*SWEET10:GUS*. In transgenic plants harboring this reporter, *GUS* signal was restricted to the leaf veins and notably enhanced in young leaves of p*GAS1:FT* plants (Fig. 1c). Changes in p*SWEET10:GUS* expression were also studied during the floral transition under LDs.

To this end, we monitored *GUS* signal in p*SWEET10:GUS* transgenic plants grown under LDs for 7, 9 and 14 days, the time window during which the floral transition occurs in our growth conditions. As observed in the previous experiment, the *GUS* signal was restricted to the leaf vasculature and it was stronger in young leaves (Fig. 1d). Furthermore, the *GUS* signal was more evident in plants undergoing the transition to reproductive phase (i.e. 9 to 14 days) compared to those at vegetative stage (i.e. 7 days) (Fig. 1d). This result was confirmed by a RT-qPCR experiment performed during the same time-course, in which an increase of *SWEET10* mRNA expression was observed in shoot apices containing young leaves of Col-0 plants from day 9 (Fig. 1e). The increase of *SWEET10* expression was largely suppressed in the *ft-10 tsf-1* mutant (Fig. 1 e). This indicates that the up-regulation of *SWEET10* during the floral transition of *A. thaliana* partially depends on the presence of a functional *FT* allele.

The photoperiodic flowering pathway of *A. thaliana* regulates *SWEET10* expression

FT is a major component of the photoperiodic flowering pathway that promotes floral induction of *A. thaliana* in response to LDs. As the above experiments suggest that *FT* regulates *SWEET10* mRNA expression levels during floral transition, we tested whether the photoperiodic pathway activates *SWEET10* transcription. The expression of *GUS* in p*SWEET10:GUS* was monitored in plants grown under SDs and then shifted to LDs for 3, 5 and 7 days. An increase in intensity of the *GUS* signal was

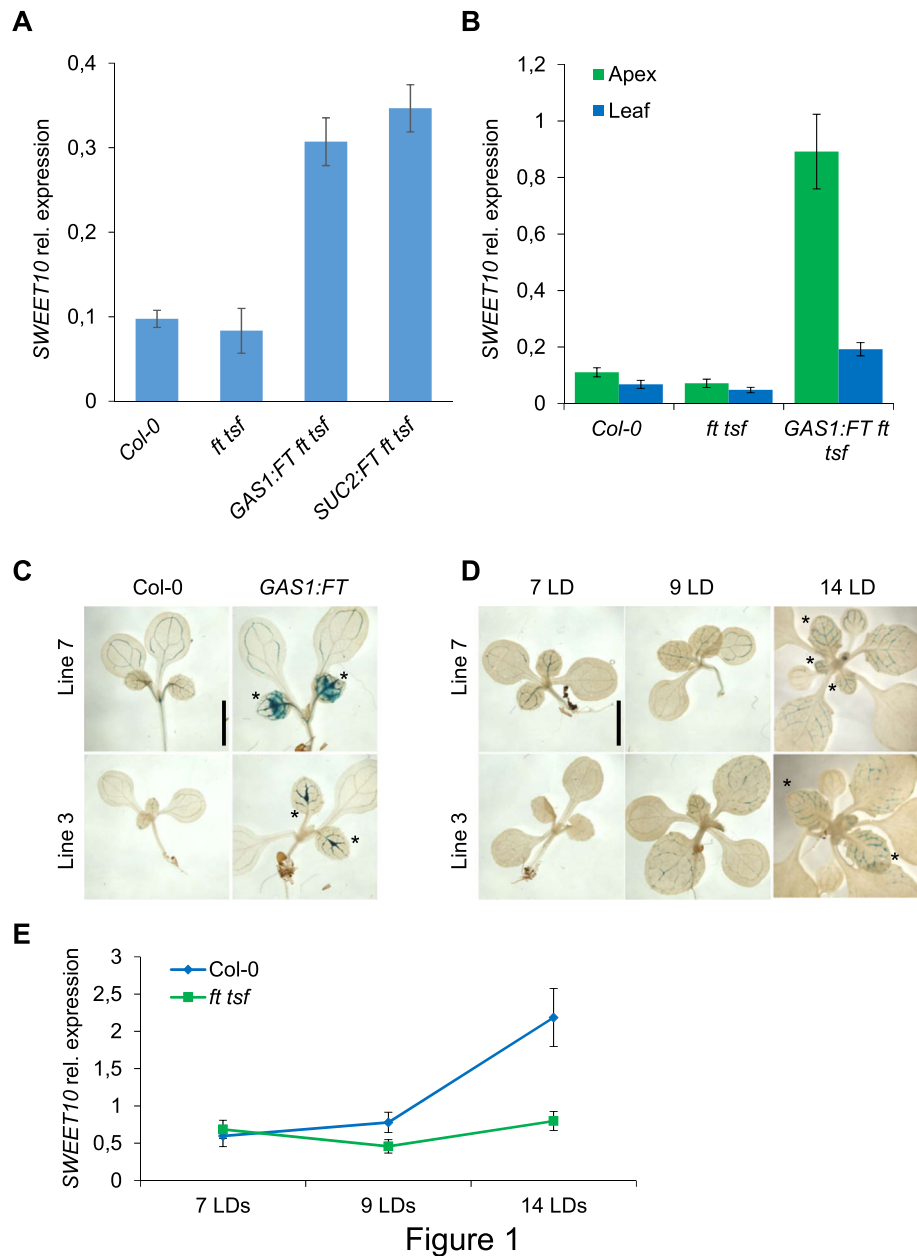


Fig. 1 *FT* activates the transcription of *SWEET10* mRNA. **a** Quantification of *SWEET10* mRNA levels in leaves of different *FT*-overexpressing plants (9 LDs at ZT16). **b** Comparison of *SWEET10* mRNA levels in leaves and shoot apices of different *FT*-overexpressing plants (9 LDs at ZT16). **c** *GUS* expression levels in T3 *pSWEET10:GUS* (Col-0) independent transgenic lines #3 and #7 (left) and in T1 plants from the cross between *pGAS1:FT* and *pSWEET10:GUS* lines #3 and #7 (right) at 7 LDs. **d** *GUS* staining of *pSWEET10:GUS* transgenic lines #3 and #7 in Col-0 background (T3 generation) during a time-course in LDs. Pictures of **(c)** and **(d)** were taken at ZT8. Scale bar = 5 mm. Asterisks indicate young leaves. **e** Quantification of *SWEET10* expression levels in shoot apices of Col-0 and *ft tsf* double mutants in a time-course under LDs. Shoot apices were sampled at ZT8. Errors bars in A, B and E indicate Standard Deviation

observed in the vascular tissue of leaves shifted to LDs compared to those grown under SDs (Fig. 2a), indicating that the *SWEET10* promoter responds to LDs. Furthermore, in cross-sections of the shoot apex of *pSWEET10:GUS* plants, *GUS* expression increased in the mature vascular tissue at the apex of plants shifted to LDs (Fig. 2b). In agreement with these observations, RT-qPCR

analysis demonstrated that the levels of *SWEET10* mRNA were higher in plants grown under LDs compared to SDs (Fig. 2c) or after the shift of SD-grown plants to LDs (Fig. 2d). Interestingly, the *GUS* expression disappeared from leaves of *pSWEET10:GUS* plants at the end of the flowering phase. Instead, *GUS* expression was observed in reproductive organs, including anthers and

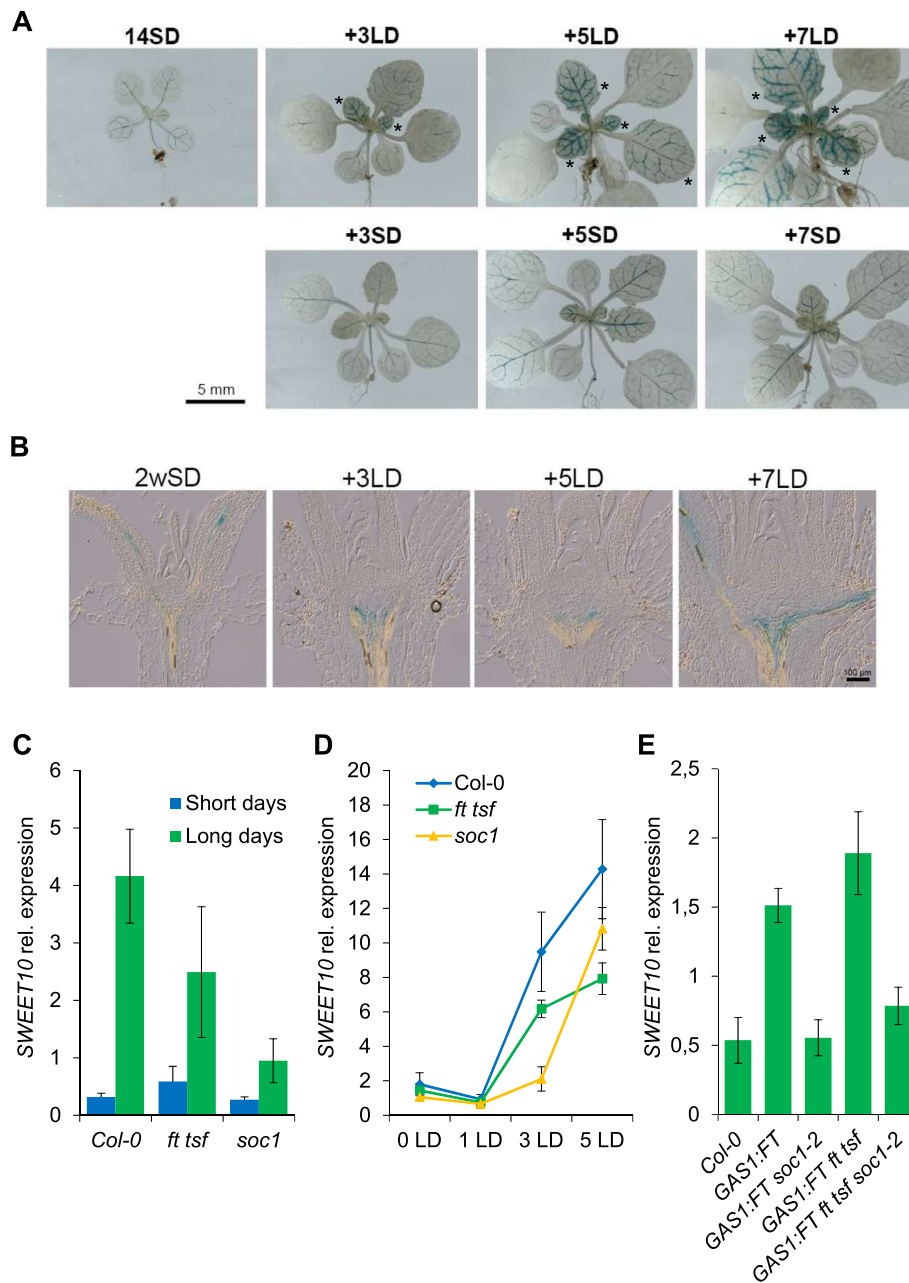


Fig. 2 Photoperiod signaling pathway contributes to *SWEET10* mRNA induction. **a** and **b** GUS expression in plants expressing p*SWEET10*:GUS (line #7, T4 generation) grown under SDs for 2 weeks, shifted to LDs and collected for GUS staining at ZT8 after exposure to different numbers of long days. **a** Whole seedlings. Scale bar = 5 mm. Asterisks indicate young leaves. **b** Shoot apices were dissected and visualized under an optical microscope (× 20). Bar scale: 100 μm. **c**, **d** and **e** shows the expression levels of *SWEET10* mRNA in shoot apices of different genotypes at ZT8. In **(c)**, plants were grown under LDs and SDs. In **(d)**, plants were grown under SDs for 2 weeks, shifted to LDs shoot apices were harvested at ZT8 in different days. In **(e)**, plants were grown under LDs and shoot apices sampled at ZT8. Errors bars in **c**, **d** and **e** indicate Standard Deviation

siliques (Additional file 2: Figure S2). The photoperiod-dependent up-regulation of *SWEET10* mRNA also involves FT, because it is reduced in *ft-10 tsf-1* mutant plants (Fig. 2c and d). *SOC1* is a key component of the photoperiod signaling pathway that acts immediately downstream of FT [31, 44]. Therefore, whether *SWEET10* transcriptional regulation requires *SOC1*

downstream of FT was investigated. Remarkably, *SWEET10* mRNA levels were strongly reduced in a null mutant allele of *SOC1* (*soc1-2*) (Fig. 2d and e). Moreover, the introduction of the *soc1-2* mutation into transgenic plants overexpressing *FT* from the *GAS1* promoter was sufficient to largely suppress the enhanced transcriptional induction of *SWEET10* mediated by higher

levels of FT (Fig. 2e). Collectively, these results indicate that *SWEET10* is transcriptionally regulated by the photoperiodic flowering pathway and this regulation involves the activities of *FT* and *SOC1*.

Overexpression of *SWEET10* causes early flowering and affects the expression levels of genes that promote floral induction

The results presented so far suggest that *SWEET10* transcription is induced by FT-signaling pathway via *SOC1*. In order to explore this possibility, we overexpressed *SWEET10* in *A. thaliana* plants and evaluated its effect on flowering time. We obtained several T1 transgenic lines that ectopically expressed *SWEET10* from the 35S promoter (p35S:*SWEET10*). A higher level of *SWEET10* mRNA expression was observed for several of these lines compared to the control Col-0 lines (Additional file 2: Figure S3). We scored the flowering-time of homozygous single copy T3 transgenic lines. Seven out of 8 tested independent transgenic lines displayed a significant acceleration of flowering compared to the control plants under LDs (Fig. 3a and b). We also overexpressed *SWEET10* in the companion cells of the phloem from the *SUC2* promoter. However, most of the p*SUC2*:

SWEET10 transgenic plants did not flower earlier than the controls (Additional file 2: Figure S4). To address whether the overexpression of *SWEET10* could accelerate flowering independently of the photoperiodic pathway, we scored the flowering time of p35S:*SWEET10* plants under SD conditions. Under these conditions, p35S:*SWEET10* transformants flowered at similar times to the controls (Fig. 3c). This result suggests that the acceleration of flowering mediated by increased *SWEET10* mRNA levels requires LDs. Therefore, the flowering function of *SWEET10* could also depend on FT function. To further characterize the function of *SWEET10*, a T-DNA insertion line and transgenic plants expressing an artificial microRNA (amiR) that targets *SWEET10* mRNA were employed. None of these genetic backgrounds displayed a significant change in flowering time compared to the wild-type plants (Fig. 3d and e; and Additional file 2: Figure S5).

In order to clarify the nature of the effect of *SWEET10* overexpression on flowering time, the expression levels of key regulators of flowering in *A. thaliana* were quantified in plants overexpressing *SWEET10* (Fig. 4). In this analysis, the mRNA levels of *FD* and some *SPL* genes (*SPL4* and 9) were higher in shoot apices of p35S:*SWEET10* during

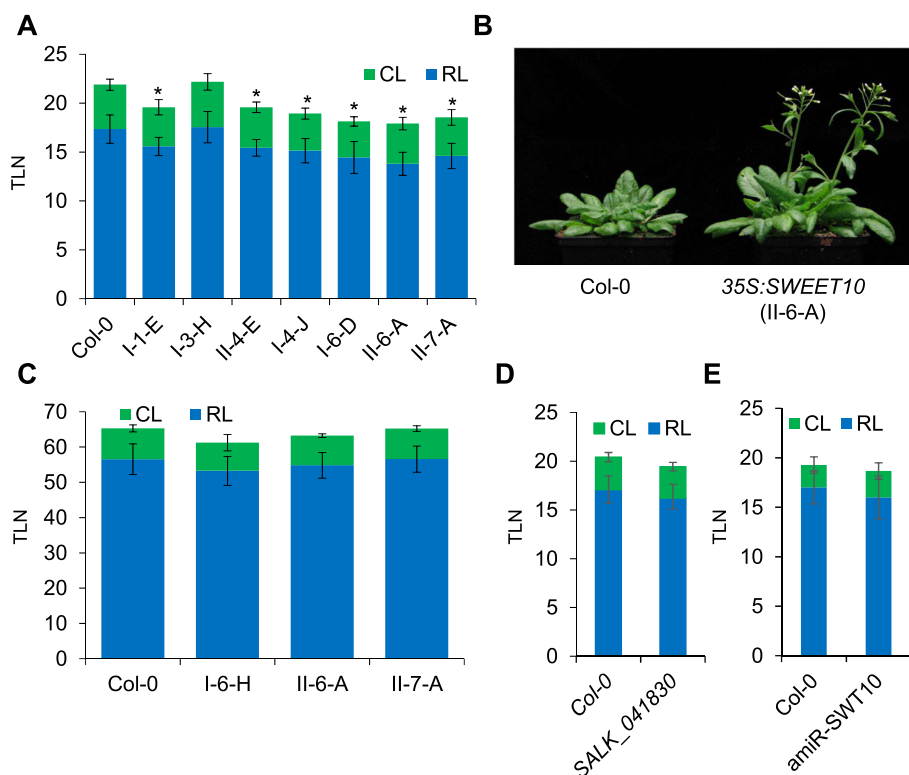


Fig. 3 Overexpression of *SWEET10* promotes flowering under LDs. Flowering time of transgenic plants overexpressing *SWEET10* from the 35S promoter under LDs (a) and (b), and under SDs (c). d Flowering time of the SALK_041830 T-DNA line compared to Col-0 under LDs. e Flowering time of a T3 transgenic line (#17-3) silencing *SWEET10* gene expression compared to Col-0 under LDs. At least 10 plants were used for each experiment. Asterisk indicates a significant difference compared to Col-0 (T-test, p -Value ≤ 0.05). TLN: Total Leaf Number; RL: Rosette Leaf number; CL: Cauline Leaf number. Errors bars in A, C, D and E indicate Standard Deviation

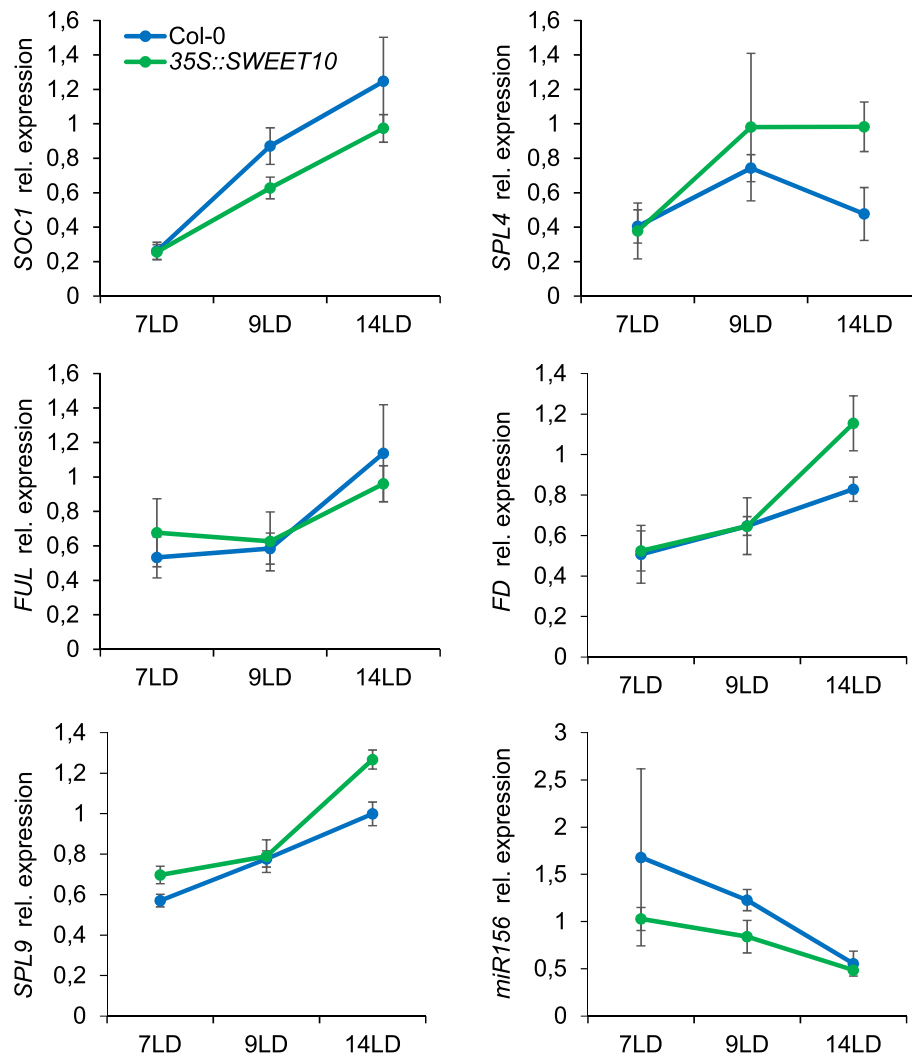


Fig. 4 Expression levels of flowering-time related genes in *SWEET10* overexpressing plants. The expression levels of flowering-time related genes was quantified in Col-0 and p35S:*SWEET10* transgenic plants (Line II-6-A, T4 generation) under LDs. Shoot apices were collected at ZT8. Errors bars indicate Standard Deviation

reproductive development (14 LDs). This pattern of expression correlates with a possible role of *SWEET10* in promoting flowering. However, *SOC1* expression was slightly lower in p35S:*SWEET10* transgenics compared wild-type plants, whereas *FUL* mRNA levels were not differentially expressed at this developmental stage. Notably, the expression level of one precursor of miR156 (*MIRNA156C*), which targets several mRNAs encoding SPLs, was reduced after 7 and 9 LDs in the p35S:*SWEET10* compared to wild type plants.

SWEET10* might be the only member of the *SWEET* family involved in the FT-flowering pathway in *A. thaliana

SWEET10 belongs to a large family of genes composed by 17 members in *A. thaliana* [45]. At least two of them, *SWEET10* and 13, were deregulated in the microarray

experiment that we performed (Table 1). We extracted the expression data of all 17 members of the family from the microarray analysis (Additional file 1). As shown in Fig. 5a, only the mRNA levels of *SWEET10* and 13 were significantly affected in p*GAS1:FT ft-10 tsf-1* plants compared to *ft-10 tsf-1* double mutants. Furthermore, we made use of promoter:*GUS* fusions to monitor the spatial and temporal expression of some other Clade III *SWEET* genes (*SWEET11*, 12, 13 and 14). In all the tested transgenic plants the *GUS* signal was detected in the vasculature under SDs, but did not increase after exposure to LDs, as was observed for p*SWEET10:GUS* (Fig. 2a and Additional file 2: Figure S6). Moreover, plants overexpressing *SWEET13* and *SWEET14* did not show acceleration of flowering under LDs. Instead, some of the tested lines displayed late flowering compared to the wild-type plants (Fig. 5b).

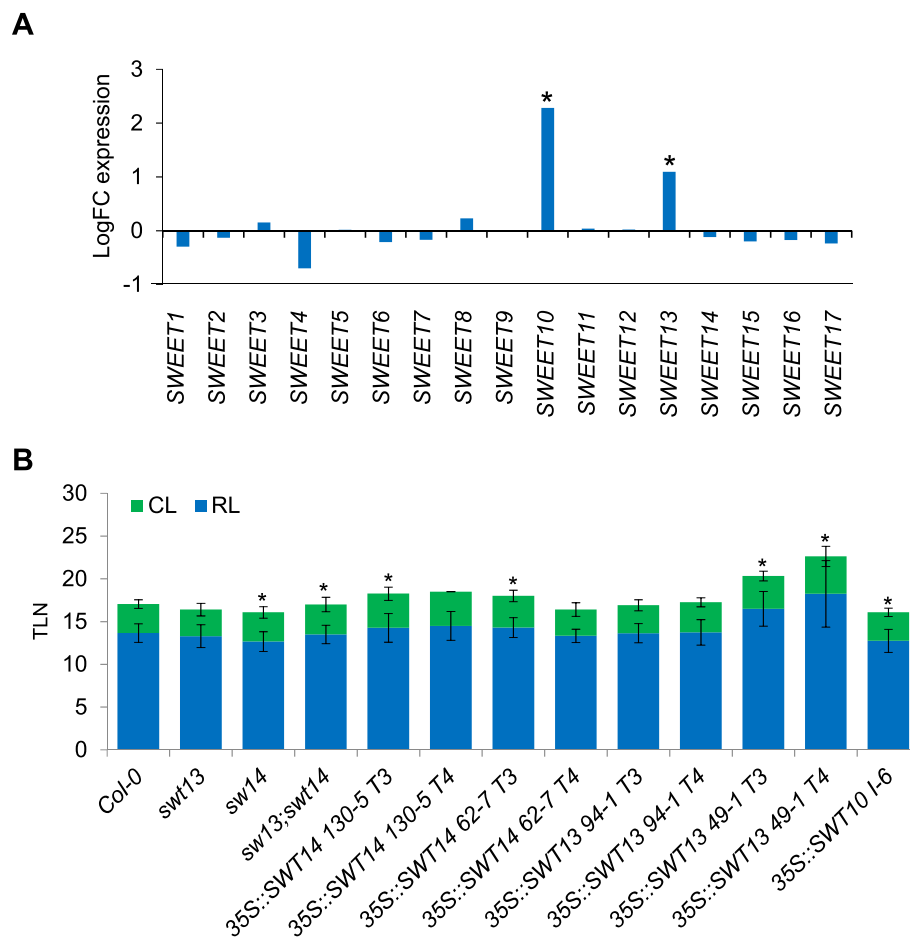


Fig. 5 Involvement of *SWEET* family members in the FT-flowering pathway. **a** Expression of *SWEET* family members in *GAS1:FT ft tsf* plants. The logFC between *GAS1:FT ft tsf* and Col-0 is represented. Significant differences are indicated with an asterisk (adj. *P* Value ≤ 0.05). **b** Effect of the overexpression of *SWEET13* and *14* in flowering-time under LDs. At least 10 plants were used for each experiment. Asterisk indicates a significant difference compared to Col-0 (T-test, *p*-Value ≤ 0.05). Scale bar = 5 mm. Errors bars in B indicate Standard Deviation

Measurement of concentrations of fructose, glucose and sucrose at shoot apices during floral transition

The increased expression of *SWEET10* at the shoot apex during floral transition (Fig. 2), suggested that sugar levels might increase in this tissue during the transition. Therefore, concentrations of sucrose, fructose and glucose were tested in shoot apices harvested from plants grown for 2 weeks under short days and then transferred to LDs for 7 days to induce the transition. Although *SWEET10* expression increases under these conditions (Fig. 2d), no significant change in concentration of any of the sugars was detected across the time course (Additional file 2: Figure S7). Also, there was no significant difference in levels of these sugars between Col-0 and *ft tsf* genotypes (Additional file 2: Figure S7). Thus, based on this analytical method, no changes in sugar levels that correlated with the floral transition could be detected in apical samples.

Discussion

Here, we demonstrate that *FT* and *SOC1* activate the transcription of the *SWEET10* gene. The pattern of transcription of *SWEET10* and the effect of its overexpression suggests that the sugar transporter it encodes contribute to the floral transition in *A. thaliana* under LDs. *SWEET10* represents a previously undescribed link between sugar transport and the photoperiod-dependent control of flowering time.

The transcriptional activation of *SWEET10* might be part of a genetic network controlled by the FT-pathway in leaves

FT protein is expressed in the vascular tissue of leaves and is transported to the shoot apex as a component of the florigenic signal that activates flowering [17, 19, 20]. However, whether *FT* has additional roles in the vascular tissue or in leaves remains unclear. Furthermore, overexpression of *FT* from the constitutive *35S* promoter

caused the transcriptional induction in leaves of *FULL* and *SEP3*, which in wild-type plants are activated by FT signaling at the shoot apex [40]. Thus, we reasoned that transcriptomic analysis of plants overexpressing *FT* from a promoter specific to companion cells of the phloem, the cell type in which *FT* is expressed [15, 37], could identify additional genes that respond to FT function in wild-type plants. The majority of genes identified by this approach as significantly regulated by FT were already known to act downstream of FT during the floral transition and flower development, such as *SOC1*, *FULL*, *SEPI* and *SEP3* (Table 1). In addition, *SWEET10* was one of the most significantly differentially expressed genes in leaves of *GASI:FT* plants and was co-regulated with these flowering genes (Additional file 2: Figure S1), suggesting that it could be activated in leaves by FT along with other known floral regulators.

FT is proposed to activate gene expression directly by interacting with the bZIP transcription factor FD that is expressed in the shoot meristem [24, 25]. However, it could also activate expression of genes such as *SWEET10* indirectly through the action of downstream transcription factors. We have shown that *SOC1* regulates the transcription of *SWEET10* (Fig. 2d and e). This regulation would probably occur in the leaves, as *SOC1* is also expressed in these organs [46]. Thus, *SWEET10* is placed downstream of FT and *SOC1*, within a genetic network that operates in the leaves.

Spatial-temporal distribution of *SWEET10* mRNA

Several *SWEET* genes belonging to Clade III (e.g. *SWEET11*, *12*, *13* and *14*) have been shown to be expressed in the vascular tissue of *A. thaliana*, most likely in the phloem [41, 47] (Chen et al., 2011; Kanno et al., 2016). In particular, *SWEET11* and *12* are expressed in specialized cells that form files along the veins and probably correspond to phloem parenchyma cells [41]. *SWEET10* was also expressed in the phloem (Fig. 1 c, d; Fig 3a and b), most likely in phloem parenchyma cells as was suggested for other members of the Clade III [41]. After floral transition, the GUS expression driven by p*SWEET10:GUS* was dramatically reduced in the leaves and highly detected in the reproductive organs (Additional file 2: Figure S2). This pattern of expression suggests diverse *SWEET10* functions throughout plant development as proposed for some *SWEET* genes [48]. In the presence of high levels of FT in the phloem such as in *GASI:FT* transgenic plants, the expression of *SWEET10* was remarkably increased and restricted to the vasculature (Fig. 1c). This poses the question of how FT activates the transcription of *SWEET10* in the phloem parenchyma cells. One possibility is that *FT* is expressed in this cell type, as suggested for the rice *FT*-like gene *Hd3a* mRNA [49]. However, it was recently

shown that in *A. thaliana* *FT* mRNA is synthesized in specific companion cells of the phloem [15] from where its protein is exported to the sieve elements. In this context, it would be more plausible that the movement of FT and/or *SOC1* to the phloem parenchyma cells is responsible for the transcriptional activation of *SWEET10*. Detailed imaging studies of the spatial distribution of FT, *SOC1* and *SWEET10* using fluorescent markers would contribute to address specifically in which phloem cells they are present. The possibility that *SOC1* acts as a mediator of FT-signaling to activate the transcription of *SWEET10* is particularly interesting. Recently, the direct targets of *SOC1* were identified using genome-wide approaches [34, 35]. In these studies, the binding of *SOC1* to *SWEET10* was not found, suggesting the existence of a third *SWEET10* activating-factor downstream of *SOC1* and FT. This factor could be encoded by one of the genes that are highly co-expressed with *SWEET10* such as *SEP3* and *FTM5* (Additional file 2: Figure S1). The identification of transcription factors that bind to *SWEET10* regulatory regions would help understand how the FT-signaling pathway induces the expression of this gene in the vasculature.

Potential functions of *SWEET10* in regulating flowering

Sugars are believed to promote flowering in several species [50]. In many of these species, floral induction correlates with a rapid increase in the concentration of sucrose in the phloem, especially near the shoot apex. This phenomenon was shown for instance in *Sinapis alba* (white mustard) [51] and *Xanthium strumarium* (rough cocklebur) [52]. In *A. thaliana*, the inductive LD treatment triggers a transient increase of sucrose in the leaf exudate [53]. Moreover, transgenic plants of different species, such as tomato, potato and *A. thaliana*, which over-accumulated sucrose in leaves flowered earlier than the control wild-type plants [53–55]. These results suggest that sucrose acts as a signal during the photoperiod flowering induction. Whether the levels of sucrose or other sugars change during floral transition in the SAM remains unclear, as its quantification in this tissue is technically challenging. Indeed, we did not detect significant changes in the concentrations of sucrose, fructose or glucose in shoot apices of *A. thaliana* plants shifted from SDs to LDs (Additional file 2: Figure S7). However, these apical samples include young leaves, a segment of the apical stem and meristems, so we cannot exclude that local changes in sugar concentration occur. Furthermore, in a previous report, sucrose was found to increase in concentration during the floral transition in shoot apices of plants grown under continuous LDs [6]. Therefore, sugars, and sucrose in particular, could act in the SAM to induce or facilitate the floral transition in response to LDs. In this context, sugar transporters such

as SWEET proteins might play an important role in this process. In agreement with this, the overexpression of *SWEET10* in *A. thaliana* triggered a significant acceleration of flowering (Fig. 3a and b). Interestingly, other sugar transporters have also been related to flowering-time control. For example, *A. thaliana* mutants deficient in *SUCROSE TRANSPORTER 9 (SUC9)* were early flowering under SDs, probably by an increase in the phloem-loading of sucrose [56]. Therefore, the transport of sugars from leaves to the SAM mediated by specialized transporters could contribute to the floral transition in *A. thaliana* and other species. However, the precise role of *SWEET10* in controlling flowering time is still unknown. One possible scenario is that *SWEET10* is transcriptionally induced downstream of FT (and SOC1) in order to supply sugars to the SAM at the time that floral transition occurs. This would contribute to satisfying increased energy requirements of the shoot meristem in order to undergo the increased growth and cell division associated with the floral transition and the initiation of floral organogenesis. Remarkably, the transcription factor CO, which is part of the photoperiodic flowering pathway of Arabidopsis, is responsible for the mobilization of sugars from amylose during the floral transition [57]. Therefore, the photoperiod pathway could affect sugar transport at least at two distinct levels: through CO to mobilize sugars [57] and then through FT to facilitate sugar transport to the SAM. In an alternative scenario, sugars transported by *SWEET10* would contribute to the movement of FT towards the SAM. However, so far there is no evidence that sugars are involved in FT transport, although it is proposed to move through the phloem in the photosynthate stream. In both situations, the effect of *SWEET10* overexpression on flowering time would depend on FT activity. In agreement with this, the early-flowering phenotype of *35S:SWEET10* transgenics was suppressed under SD conditions (Fig. 3c). Moreover, the overexpression of *SWEET10* resulted in the induction of genes in the shoot apex related to FT function (Fig. 4). Among them, *SPL4* and *SPL9* that are also known to be upregulated by gibberellin signaling under inductive LD conditions [58]. Interestingly, SWEET proteins were proposed to transport gibberellins as well as sucrose [47], suggesting that FT could regulate both sucrose and gibberellin levels at the apex during flowering by upregulating *SWEET10*. Also, in potato FT was proposed to regulate SWEET function at the post-translational level to prevent leakage of sugar into the apoplast [59].

Overexpression of *SWEET10* caused early flowering, but loss of function mutants were not affected in flowering time. Overexpression from the *35S* promoter is widely used to address the function of genes, but loss-of-function genetics would provides more definitive

evidence on the role of *SWEET* genes in flowering time-control. *SWEET10* single mutants examined here did not show any striking phenotype related to flowering-time (Fig. 3d and e), which could be explained by functional redundancy between members of the *SWEET* family. *SWEET13* is an obvious candidate to play a redundant function, as its expression was also upregulated in *pGAS1:FT ft-10 tsf-1* plants compared to *ft-10 tsf-1* double mutants (Table 1). However, *SWEET13* overexpression did not result in early flowering (Fig. 5b), and higher order mutants might also show pleiotropic phenotypes. Thus, a systematic study of higher order loss-of-function mutants could be necessary to obtain a more complete picture of the *SWEET* genes function in flowering-time. Furthermore, induction of SWEET proteins during flowering might contribute to the altered metabolic state of the vasculature during floral transition without visibly altering leaf number or flowering time. Nevertheless, the reduced expression of *SWEET10* in *ft tsf* double mutants and its increased expression after transfer to LDs, support a relationship between *SWEET10* transcription and flowering.

Conclusions

The data shown here indicate that transcriptional activation of *SWEET10* by FT and SOC1 occurs during the promotion of flowering mediated by inductive photoperiod and that overexpression of *SWEET10* causes early flowering consistent with a functional role in this process. This emphasizes the likely significance of changing patterns in sugar transport during the floral transition. Moreover, it supports the idea that FT not only plays a role as a long-distance signaling molecule but that it can also function in leaves to bring about transcriptional changes that eventually contribute to flowering-time regulation in the SAM.

Methods

Plant materials

Arabidopsis thaliana Columbia-0 (Col) was used as wild-type in all experiments and for plant transformation. The transgenic plants *pGAS1:FT ft-10 tsf-1*, *pGAS1:FT* and *pGAS1:FT soc1-2* were previously described [11, 60]. The mutant alleles used were *soc1-2* [33] and *ft-10 tsf-1* [11]. The *SWEET10* CDS sequence was obtained from the Arabidopsis Biological Resource Center (<http://www.arabidopsis.org/>) (clone U15254) and cloned in the pAlligator-2 [61] and pSUC2:GW [11] vectors to generate the *p35S:SWEET10* and *pSUC2:SWEET10* lines, respectively. To generate *35S:SW13* and *35S:SW14* transgenic plants, *SWEET13* and *SWEET14* cDNAs were amplified (primer combinations in Additional file 2: Table S1) and inserted into a cloning vector. The inserted sequences were then cloned into the

binary vector pBE2113 [62] with XbaI and SmaI restriction sites. The *pSWEET10:GUS* lines were obtained by cloning a 3 Kb region upstream of the transcriptional starting site of the *SWEET10* gene (primers in Additional file 2: Table S1) into the pGreen-GW-GUS vector [63]. For *pSWEET11:GUS* and *pSWEET12:GUS* constructs, promoter regions (approximately 2 kb) of *SWEET11* and *SWEET12* were amplified (primer combinations in Additional file 2: Table S1). The amplified fragments were cloned into pENTR/D-TOPO and then into pGWB3 [64]. *pSWEET13:GUS* and *pSWEET14:GUS* transgenic plants were described previously [47]. T-DNA line SALK_041830 was obtained from the Nottingham Arabidopsis Stock Center (NASC) (<http://arabidopsis.info/>). For the production of the *SWEET10* silencing lines, a amiRNA targeting this gene was generated by using the online tool WMD3 (primers in Additional file 2: Table S1) and the artificial miRNA vector pRS300 [65]. The resulting *amiRNA-SWEET10* construct was cloned in the vector pAlligator-2. Arabidopsis plants were transformed following the floral dip method [66].

Plant growth conditions

Seeds were stratified on soil for 3 day in the dark at 4 °C. Plants were grown under controlled environmental conditions at 22 °C and white fluorescent light (150 μmol/m²/s), either in LDs (16 h light/8 h dark) or in SDs (8 h light/16 h dark). Flowering time was scored by counting total leaves number (caulines and rosettes) of at least 10 plants per genotype. Each experiment was performed at least twice. For RT-qPCR experiments leaves and shoot apices (containing a segment of the apical stem, SAM and young leaves) were dissected manually.

Microarray experiment

Col-0, *ft-10 tsf-1* and *pGAS1:FT ft-10 tsf-1* plants were grown under LD conditions during 9 days. Leaves of each genotypes were harvested at the end of the light period (ZT16). RNA from three independent biological replicates was extracted using the RNA Plant Mini kit, QIAGEN (www1.qiagen.com/). The concentration of the total RNA was determined using a NanoDrop ND1000 spectrophotometer. The probe synthesis and the hybridization were performed as previously described in [67]. One microgram of total RNA was reverse transcribed into cDNA using an oligo(dT)-T7 primer, and was then converted into cRNA and linearly amplified by T7 in vitro transcription reaction using the standard Ambion protocol (MessageAmp aRNA Kit, Ambion). cRNA was then reverse transcribed with random primers to dUTP-containing ds cDNA (WT ds cDNA Synthesis Kit, catalog no. 900813; Affymetrix). Fragmentation and labeling was performed with the GeneChip WT double-stranded DNA Terminal Labeling Kit

(catalog no. 900812, Affymetrix). After fragmentation, 7.5 μg of ds-cDNA was hybridized for 16 h at 45C on GeneChip Arabidopsis Tiling 1.0R Array. GeneChips were washed and stained with Fluidics Script FS450_0001 in the Affymetrix Fluidics Station 450. Then, the GeneChips were scanned using the GeneChip Scanner 3000 7G. Data was processed in R v2.8.1 using the probe annotation *athtiling1.0rcdf* as described in [68]. Probe-level data were pre-processed using the RMA algorithm implemented in the Bioconductor package *Affy* v1.24.2. Linear models and empirical Bayes methods from the *Limma* package v2.14 of Bioconductor were applied to derive a *P* value, false discovery rate (FDR; *P* adjusted), and mean of log₂-based ratio across replicates. The data were deposited in the Gene Expression Omnibus at the National Center for Biotechnology Information (GEO accession number GSE125054).

RT-qPCR

RNA expression analyses were performed as described in [69]. The RNA was extracted from plant tissue (leaves or shoot apices) by using the RNeasy Plant Mini Kit (Qiagen) and treated with DNA-free DNase (Ambion). One microgram of total RNA (quantified in a Nanodrop ND-1000) was used for reverse transcription by using the Superscript III (Invitrogen). Levels of gene expression were quantified by qPCR in a LightCycler 480 instrument (Roche) using the *PEX4* gene (*AT5G25760*) as a reference. Three biological replicates were performed for each qRT-PCR assay. The average of the three replicates is shown. The list of primers used for expression analyses can be found in the Additional file 2: Table S1.

Histochemical staining for GUS activity

Transgenic plants of *pSWEET10:GUS*, *pSWEET11:GUS*, *pSWEET12:GUS*, *pSWEET13:GUS* and *pSWEET14:GUS* were fixed with cold 90% (v/v) acetone for 30 min on ice, then washed with 50 mM sodium phosphate buffer twice. The samples were then immersed in the X-Gluc staining solution [50 mM NaPO₄ buffer (pH 7.0), 0.5 mM K₃Fe(CN)₆, 0.5 mM K₄Fe(CN)₆, 0.1% (v/v) Triton X-100, 0.5 mg/ml 5-bromo-4-chloro-3-indolyl-beta-D-glucuronide (X-Gluc) in H₂O] under vacuum for 15 min, and then incubated at 37 °C in the dark for 40 h. After the reaction, the samples were washed with 50 mM sodium phosphate buffer, dehydrated through an ethanol series and observed under stereo microscope (Zeiss, Stemi 508).

For histological analysis, the samples were embedded in paraffin, and sliced with the microtome (Leica, RM2125 RTS) to make serial sections of 8-μm thickness. After deparaffinization and rehydration, the sections were observed with the differential interference contrast (DIC) microscope (Zeiss, Axio Imager M2).

Sugar measurements

For each sample, 30 apices were harvested and frozen in liquid nitrogen. The samples were extracted in chloroform/methanol/water according to [70]. The aqueous phase was used for sugar measurement and the chloroform phase for protein determination. Sucrose, fructose and glucose were determined photospectrometrically using a 96-well plate reader (Synergy HT from BioTek, U.S.A.) based on the method described in [71] and adapted to the 96-well format by [72]. Protein content was measured according to [73] using the DC™ Protein Assay kit (Bio-Rad Laboratories, U.S.A.) and the values were used for normalization of the sugar data.

Supplementary information

Supplementary information accompanies this paper at <https://doi.org/10.1186/s12870-020-2266-0>.

Additional file 1. Results of the expression studies performed on the GeneChip Arabidopsis Tiling 1.0R Array.

Additional file 2: Table S1. Primers used in this study. **Figure S1.** In silico analyses of co-expressed gene networks around *SWEET10*. (A) Gene network representation and list of genes correlated to *SWEET10* during development generated by the GENEVESTIGATOR software [74]. (B) The *Arabidopsis thaliana* trans-factor and cis-element prediction database ATTED-II [75], implemented in www.arabidopsis.org, was used to predict and visualize co-expressed genes around *SWEET10*. **Figure S2.** Expression of *pSWEET10:GUS* in adult Arabidopsis plants. (A) GUS expression in a whole plant expressing *pSWEET10:GUS*. The T4 transgenic plant shown in (A) was grown under LDs until siliques were produced. Detail of an inflorescence (B) and a silique (C) showing GUS expression. **Figure S3.** Expression levels of *SWEET10* in T1 transgenic lines overexpressing *SWEET10*. The expression levels of *SWEET10* was quantified in Col-0 and 35S:*SWEET10* T1 lines under LDs. Leaves were collected at ZT8. Errors bars indicate Standard Deviation. **Figure S4.** Effect of the overexpression of *SWEET10* from *SUC2* promoter on flowering time under LDs. At least 10 plants were used for each experiment. Asterisk indicates a significant different compared to Col-0 (T-test, p -Value ≤ 0.05). Errors bars indicate Standard Deviation. **Figure S5.** Analysis of *amiR-SWEET10* transgenic plants. (A) Flowering time of 44 *amiR-SWEET10* T1 lines compared to Col-0 under LDs. TLN: Total Leaf Number. (B) *SWEET10* expression levels in a subset of T3 *amiR-SWEET10* lines. **Figure S6.** Photoperiod-dependent expression profile of *SWEET11*, 12, 13 and 14. GUS expression in plants expressing *pSWEET10:GUS*. Plants were grown under SDs for 2 weeks, shifted to LDs and collected for GUS staining at ZT8 in different days. Scale bar = 5 mm. **Figure S7.** Levels of sugar during the photoperiodic induction of flowering. Col-0 and *ft tsf* plants were grown under SDs for 2 weeks, shifted to LDs shoot apices were harvested at ZT8 in different days. Shoot apices were harvested and used to quantify the concentration of fructose (fru), glucose (glu) and sucrose (suc). Errors bars indicate Standard Deviation.

Abbreviations

FDR: False discovery rate; LDs: Long days; RT-qPCR: Quantitative reverse transcription polymerase chain reaction; SAM: Shoot apical meristem; SDs: Short days

Acknowledgements

Not Applicable.

Authors' contributions

FA performed experimental work and data analysis. FA and GC designed the study and contributed to writing and revising the manuscript. AK, MS, YC and TMA contributed to experimental work and revising the manuscript. NK,

VF, VSF, TMDC, BH and SJ contributed to experimental work. All authors read and approved the final manuscript.

Authors' information

All information is provided on the title page.

Funding

This work was financially supported by the Deutsche Forschung Gemeinschaft (DFG) through the Cluster of Excellence in Plant Science CEPLAS (EXC 1028) to T.M.A. and G.C., the European Union via a Marie Curie Intra-European Fellowship for Career Development (Project Intra-European Grant Agreement-2009-251839) (to F.A.), Alexander von Humboldt Fellowship for Postdoctoral Researchers and Japan Society for the Promotion of Science (JSPS) Overseas Research Fellowships (to A.K.), and the Max Planck Society through a core grant (to G.C.). The Funding bodies were not involved in the design of the study and collection, analysis, and interpretation of data and in writing the manuscript.

Availability of data and materials

The transcriptomic datasets generated during the current study are available in the GEO NCBI repository under the accession number GSE125054 and in the supplementary information files. *Arabidopsis thaliana* Columbia, which was used throughout this work, is available from the Arabidopsis Stock Centre, Nottingham University, UK or the Arabidopsis Biological Resource Centre, Ohio State University or from the authors. No permissions were required to use *Arabidopsis thaliana*. All other plants materials are available from the corresponding authors.

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Author details

¹Max Planck Institute for Plant Breeding Research, Carl-von-Linne-Weg 10, 50829 Köln, Germany. ²Present Address: UMR AGAP, Univ. Montpellier, INRAE, CIRAD, INSAE, Montpellier, France. ³Present Address: BPMP, Univ Montpellier, CNRS, INRAE, Montpellier SupAgro, Montpellier, France. ⁴Present Address: World Vegetable Center Korea Office (WKO), 100 Nongsaeangmyeong-ro, Iseo-myeon, Wanju-gun, Jellabuk-do 55365, Republic of Korea. ⁵RIKEN Center for Sustainable Resource Science, 1-7-22 Suehiro-cho, Tsurumi-ku, Yokohama, Kanagawa 230-0045, Japan. ⁶Cluster of Excellence on Plant Sciences and Institute of Plant Biochemistry, Heinrich-Heine University, 40225 Düsseldorf, Germany.

Received: 26 March 2019 Accepted: 27 January 2020

Published online: 03 February 2020

References

- Andrés F, Coupland G. The genetic basis of flowering responses to seasonal cues. *Nat Rev Genet.* 2012;13(9):627–39.
- Srikanth A, Schmid M. Regulation of flowering time: all roads lead to Rome. *Cell Mol Life Sci.* 2011;68(12):2013–37.
- Bouche F, Lobet G, Tocquin P, Perilleux C. FLOR-ID: an interactive database of flowering-time gene networks in Arabidopsis thaliana. *Nucleic Acids Res.* 2016;44(D1):D1167–71.
- Kardailsky I, Shukla VK, Ahn JH, Dagenais N, Christensen SK, Nguyen JT, Chory J, Harrison MJ, Weigel D. Activation tagging of the floral inducer FT. *Science.* 1999;286(5446):1962–5.
- Kobayashi Y, Kaya H, Goto K, Iwabuchi M, Araki T. A pair of related genes with antagonistic roles in mediating flowering signals. *Science.* 1999; 286(5446):1960–2.
- Wahl V, Ponnuraj J, Schlereth A, Arrivault S, Langenecker T, Franke A, Feil R, Lunn JE, Stitt M, Schmid M. Regulation of flowering by Trehalose-6-phosphate signaling in Arabidopsis thaliana. *Science.* 2013;339(6120):704–7.

7. Ohto M, Onai K, Furukawa Y, Aoki E, Araki T, Nakamura K. Effects of sugar on vegetative development and floral transition in Arabidopsis. *Plant Physiol.* 2001;127(1):252–61.
8. Cho LH, Pasriga R, Yoon J, Jeon JS, An G. Roles of sugars in controlling flowering time. *J Plant Biol.* 2018;61(3):121–30.
9. Gomez LD, Gilday A, Feil R, Lunn JE, Graham IA. AtTPS1-mediated trehalose 6-phosphate synthesis is essential for embryogenic and vegetative growth and responsiveness to ABA in germinating seeds and stomatal guard cells. *Plant J.* 2010;64(1):1–13.
10. Koornneef M, Alonso-Blanco C, Blankestijn-de Vries H, Hanhart CJ, Peeters AJ. Genetic interactions among late-flowering mutants of Arabidopsis. *Genetics.* 1998;148(2):885–92.
11. Jang S, Torti S, Coupland G. Genetic and spatial interactions between FT, TSF and SVP during the early stages of floral induction in Arabidopsis. *Plant J.* 2009;60(4):614–25.
12. Yamaguchi A, Kobayashi Y, Goto K, Abe M, Araki T. TWIN SISTER OF FT (TSF) acts as a floral pathway integrator redundantly with FT. *Plant Cell Physiol.* 2005;46(8):1175–89.
13. Takada S, Goto K. Terminal flower2, an Arabidopsis homolog of heterochromatin protein1, counteracts the activation of flowering locus T by CONSTANS in the vascular tissues of leaves to regulate flowering time. *Plant Cell.* 2003;15(12):2856–65.
14. Adrian J, Farrona S, Reimer JJ, Albani MC, Coupland G, Turck F. cis-Regulatory elements and chromatin state coordinately control temporal and spatial expression of FLOWERING LOCUS T in Arabidopsis. *Plant Cell.* 2010;22(5):1425–40.
15. Chen Q, Payyavula RS, Chen L, Zhang J, Zhang C, Turgeon R. FLOWERING LOCUS T mRNA is synthesized in specialized companion cells in Arabidopsis and Maryland mammoth tobacco leaf veins. *Proc Natl Acad Sci U S A.* 2018;115(11):2830–5.
16. Suarez-Lopez P, Wheatley K, Robson F, Onouchi H, Valverde F, Coupland G. CONSTANS mediates between the circadian clock and the control of flowering in Arabidopsis. *Nature.* 2001;410(6832):1116–20.
17. Corbesier L, Vincent C, Jang S, Fornara F, Fan Q, Searle I, Giakountis A, Farrona S, Gissot L, Turnbull C, et al. FT protein movement contributes to long-distance signaling in floral induction of Arabidopsis. *Science.* 2007;316(5827):1030–3.
18. Tamaki S, Matsuo S, Wong HL, Yokoi S, Shimamoto K. Hd3a protein is a mobile flowering signal in rice. *Science.* 2007;316(5827):1033–6.
19. Jaeger KE, Wigge PA. FT protein acts as a long-range signal in Arabidopsis. *Curr Biol.* 2007;17(12):1050–4.
20. Mathieu J, Warthmann N, Kuttner F, Schmid M. Export of FT protein from phloem companion cells is sufficient for floral induction in Arabidopsis. *Curr Biol.* 2007;17(12):1055–60.
21. Liu L, Liu C, Hou X, Xi W, Shen L, Tao Z, Wang Y, Yu H. FTIP1 is an essential regulator required for florigen transport. *PLoS Biol.* 2012;10(4):e1001313.
22. Zhu Y, Liu L, Shen L, Yu H. NaKR1 regulates long-distance movement of FLOWERING LOCUS T in Arabidopsis. *Nat Plants.* 2016;2(6):16075.
23. Nakamura Y, Andres F, Kanehara K, Liu YC, Dormann P, Coupland G. Arabidopsis florigen FT binds to diurnally oscillating phospholipids that accelerate flowering. *Nat Commun.* 2014;5:3553.
24. Abe M, Kobayashi Y, Yamamoto S, Daimon Y, Yamaguchi A, Ikeda Y, Ichinoki H, Notaguchi M, Goto K, Araki T. FD, a bZIP protein mediating signals from the floral pathway integrator FT at the shoot apex. *Science.* 2005;309(5737):1052–6.
25. Wigge PA, Kim MC, Jaeger KE, Busch W, Schmid M, Lohmann JU, Weigel D. Integration of spatial and temporal information during floral induction in Arabidopsis. *Science.* 2005;309(5737):1056–9.
26. Jaeger KE, Pullen N, Lamzin S, Morris RJ, Wigge PA. Interlocking feedback loops govern the dynamic behavior of the floral transition in Arabidopsis. *Plant Cell.* 2013;25(3):820–33.
27. Searle I, He YH, Turck F, Vincent C, Fornara F, Krober S, Amasino RA, Coupland G. The transcription factor FLC confers a flowering response to vernalization by repressing meristem competence and systemic signaling in Arabidopsis. *Genes Dev.* 2006;20(7):898–912.
28. Schmid M, Uhlenhaut NH, Godard F, Demar M, Bressan R, Weigel D, Lohmann JU. Dissection of floral induction pathways using global expression analysis. *Development.* 2003;130(24):6001–12.
29. Torti S, Fornara F, Vincent C, Andres F, Nordstrom K, Gobel U, Knoll D, Schoof H, Coupland G. Analysis of the Arabidopsis shoot meristem transcriptome during floral transition identifies distinct regulatory patterns and a leucine-rich repeat protein that promotes flowering. *Plant Cell.* 2012;24(2):444–62.
30. Taoka K, Ohki I, Tsuji H, Furuita K, Hayashi K, Yanase T, Yamaguchi M, Nakashima C, Purwestri YA, Tamaki S, et al. 14-3-3 proteins act as intracellular receptors for rice Hd3a florigen. *Nature.* 2011;476(7360):332–U397.
31. Samach A, Onouchi H, Gold SE, Ditta GS, Schwarz-Sommer Z, Yanofsky MF, Coupland G. Distinct roles of CONSTANS target genes in reproductive development of Arabidopsis. *Science.* 2000;288(5471):1613–6.
32. Borner R, Kampmann G, Chandler J, Gleissner R, Wisman E, Apel K, Melzer S. A MADS domain gene involved in the transition to flowering in Arabidopsis. *Plant J.* 2000;24(5):591–9.
33. Lee H, Suh SS, Park E, Cho E, Ahn JH, Kim SG, Lee JS, Kwon YM, Lee I. The AGAMOUS-LIKE 20 MADS domain protein integrates floral inductive pathways in Arabidopsis. *Genes Dev.* 2000;14(18):2366–76.
34. Immink RG, Pose D, Ferrario S, Ott F, Kaufmann K, Valentim FL, de Folter S, van der Wal F, van Dijk ADJ, Schmid M, et al. Characterization of SOC1's central role in flowering by the identification of its upstream and downstream regulators. *Plant Physiol.* 2012;160(1):433–49.
35. Tao Z, Shen LS, Liu C, Liu L, Yan YY, Yu H. Genome-wide identification of SOC1 and SVP targets during the floral transition in Arabidopsis. *Plant J.* 2012;70(4):549–61.
36. Melzer S, Lens F, Gennen J, Vanneste S, Rohde A, Beeckman T. Flowering-time genes modulate meristem determinacy and growth form in Arabidopsis thaliana. *Nat Genet.* 2008;40(12):1489–92.
37. An H, Rousset C, Suarez-Lopez P, Corbesier L, Vincent C, Pineiro M, Hepworth S, Mouradov A, Justin S, Turnbull C, et al. CONSTANS acts in the phloem to regulate a systemic signal that induces photoperiodic flowering of Arabidopsis. *Development.* 2004;131(15):3615–26.
38. Navarro C, Abelenda JA, Cruz-Oro E, Cuellar CA, Tamaki S, Silva J, Shimamoto K, Prat S. Control of flowering and storage organ formation in potato by FLOWERING LOCUS T. *Nature.* 2011;478(7367):119–22.
39. Zhang H, Harry DE, Ma C, Yuceer C, Hsu CY, Vikram V, Shevchenko O, Etherington E, Strauss SH. Precocious flowering in trees: the FLOWERING LOCUS T gene as a research and breeding tool in Populus. *J Exp Bot.* 2010;61(10):2549–60.
40. Teper-Bamnolker P, Samach A. The flowering integrator FT regulates SEPALLATA3 and FRUITFULL accumulation in Arabidopsis leaves. *Plant Cell.* 2005;17(10):2661–75.
41. Chen LQ, Qu XQ, Hou BH, Sosso D, Osorio S, Fernie AR, Frommer WB. Sucrose efflux mediated by SWEET proteins as a key step for phloem transport. *Science.* 2012;335(6065):207–11.
42. Stadler R, Sauer N. The Arabidopsis thaliana AtSUC2 gene is specifically expressed in companion cells. *Bot Acta.* 1996;109(4):299–306.
43. Haritatos E, Ayre BG, Turgeon R. Identification of phloem involved in assimilate loading in leaves by the activity of the galactinol synthase promoter. *Plant Physiol.* 2000;123(3):929–37.
44. Yoo SK, Chung KS, Kim J, Lee JH, Hong SM, Yoo SJ, Yoo SY, Lee JS, Ahn JH. CONSTANS activates SUPPRESSOR OF OVEREXPRESSION OF CONSTANS 1 through FLOWERING LOCUS T to promote flowering in Arabidopsis. *Plant Physiol.* 2005;139(2):770–8.
45. Chen LQ, Hou BH, Lalonde S, Takanaga H, Hartung ML, Qu XQ, Guo WJ, Kim JG, Underwood W, Chaudhuri B, et al. Sugar transporters for intercellular exchange and nutrition of pathogens. *Nature.* 2010;468(7323):527–32.
46. Hepworth SR, Valverde F, Ravenscroft D, Mouradov A, Coupland G. Antagonistic regulation of flowering-time gene SOC1 by CONSTANS and FLC via separate promoter motifs. *EMBO J.* 2002;21(16):4327–37.
47. Kanno Y, Oikawa T, Chiba Y, Ishimaru Y, Shimizu T, Sano N, Koshiba T, Kamiya Y, Ueda M, Seo M. AtSWEET13 and AtSWEET14 regulate gibberellin-mediated physiological processes. *Nat Commun.* 2016;7:13245.
48. Durand M, Mainson D, Porcheron B, Mauroussat L, Lemoine R, Pourtau N. Carbon source-sink relationship in Arabidopsis thaliana: the role of sucrose transporters. *Planta.* 2018;247(3):587–611.
49. Pasriga R, Cho LH, Yoon J, An G. Identification of the regulatory region responsible for vascular tissue-specific expression in the Rice Hd3a promoter. *Mol Cells.* 2018;41(4):342–50.
50. Bernier G, Havelange A, Houssa C, Petitjean A, Lejeune P. Physiological signals that induce flowering. *Plant Cell.* 1993;5(10):1147–55.
51. Pryke JA, Bernier G. Acid Invertase activity in apex of Sinapis-Alba during transition to flowering. *Ann Bot-London.* 1978;42(179):747–9.
52. Houssa P, Bernier G, Kinet JM. Qualitative and quantitative analysis of carbohydrates in leaf exudate of the short-day plant, xanthium-Strumarium L during floral transition. *J Plant Physiol.* 1991;138(1):24–8.

53. Corbesier L, Lejeune P, Bernier G. The role of carbohydrates in the induction of flowering in *Arabidopsis thaliana*: comparison between the wild type and a starchless mutant. *Planta*. 1998;206(1):131–7.
54. Micallef BJ, Haskins KA, Vanderveer PJ, Roh KS, Shewmaker CK, Sharkey TD. Altered photosynthesis, flowering, and fruiting in transgenic tomato plants that have an increased capacity for sucrose synthesis. *Planta*. 1995;196(2):327–34.
55. Muller-rober B, Sonnewald U, Willmitzer L. Inhibition of the Adp-glucose Pyrophosphorylase in transgenic potatoes leads to sugar-storing tubers and influences tuber formation and expression of tuber storage protein genes. *EMBO J*. 1992;11(4):1229–38.
56. Sivitz AB, Reinders A, Johnson ME, Krentz AD, Grof CPL, Perroux JM, Ward JM. *Arabidopsis* sucrose transporter AtSUC9. High-affinity transport activity, intragenic control of expression, and early flowering mutant phenotype. *Plant Physiol*. 2007;143(1):188–98.
57. Ortiz-Marchena MI, Albi T, Lucas-Reina E, Said FE, Romero-Campero FJ, Cano B, Ruiz MT, Romero JM, Valverde F. Photoperiodic control of carbon distribution during the floral transition in *Arabidopsis*. *Plant Cell*. 2014;26(2):565–84.
58. Porri A, Torti S, Romera-Branchat M, Coupland G. Spatially distinct regulatory roles for gibberellins in the promotion of flowering of *Arabidopsis* under long photoperiods. *Development*. 2012;139(12):2198–209.
59. Abelenda JA, Bergonzi S, Oortwijn M, Sonnewald S, Du MR, Visser RGF, Sonnewald U, Bachem CWB. Source-sink regulation is mediated by interaction of an FT Homolog with a SWEET protein in potato. *Curr Biol*. 2019;29(7):1178–+.
60. Andres F, Romera-Branchat M, Martinez-Gallegos R, Patel V, Schneeberger K, Jang S, Altmuller J, Nurnberg P, Coupland G. Floral induction in *Arabidopsis* by FLOWERING LOCUS T requires direct repression of BLADE-ON-PETIOLE genes by the Homeodomain protein PENNYWISE. *Plant Physiol*. 2015;169(3):2187–99.
61. Bensmihen S, To A, Lambert G, Kroj T, Giraudat J, Parcy F. Analysis of an activated ABI5 allele using a new selection method for transgenic *Arabidopsis* seeds. *FEBS Lett*. 2004;561(1–3):127–31.
62. Mitsuohara I, Ugaki M, Hirochika H, Ohshima M, Murakami T, Gotoh Y, Katayose Y, Nakamura S, Honkura R, Nishimiya S, et al. Efficient promoter cassettes for enhanced expression of foreign genes in dicotyledonous and monocotyledonous plants. *Plant Cell Physiol*. 1996;37(1):49–59.
63. Zhou Y, Wang YJ, Krause K, Yang TT, Dongus JA, Zhang YJ, Turck F. Telobox motifs recruit CLF/SWN-PRC2 for H3K27me3 deposition via TRB factors in *Arabidopsis*. *Nat Genet*. 2018;50(5):638–+.
64. Nakagawa T, Kurose T, Hino T, Tanaka K, Kawamukai M, Niwa Y, Toyooka K, Matsuoka K, Jinbo T, Kimura T. Development of series of gateway binary vectors, pGWBs, for realizing efficient construction of fusion genes for plant transformation. *J Biosci Bioeng*. 2007;104(1):34–41.
65. Ossowski S, Schwab R, Weigel D. Gene silencing in plants using artificial microRNAs and other small RNAs. *Plant J*. 2008;53(4):674–90.
66. Clough SJ, Bent AF. Floral dip: a simplified method for agrobacterium-mediated transformation of *Arabidopsis thaliana*. *Plant J*. 1998;16(6):735–43.
67. Gregis V, Andres F, Sessa A, Guerra RF, Simonini S, Mateos JL, Torti S, Zambelli F, Prazzoli GM, Bjerkan KN, et al. Identification of pathways directly regulated by SHORT VEGETATIVE PHASE during vegetative and reproductive development in *Arabidopsis*. *Genome Biol*. 2013;14(6):R56.
68. Naouar N, Vandepoele K, Lammens T, Casneuf T, Zeller G, van Hummelen P, Weigel D, Ratsch G, Inze D, Kuiper M, et al. Quantitative RNA expression analysis with Affymetrix tiling 1.0R arrays identifies new E2F target genes. *Plant J*. 2009;57(1):184–94.
69. Andres F, Porri A, Torti S, Mateos J, Romera-Branchat M, Garcia-Martinez JL, Fornara F, Gregis V, Kater MM, Coupland G. SHORT VEGETATIVE PHASE reduces gibberellin biosynthesis at the *Arabidopsis* shoot apex to regulate the floral transition. *Proc Natl Acad Sci U S A*. 2014;111(26):E2760–9.
70. Lunn JE, Feil R, Hendriks JHM, Gibon Y, Morcuende R, Osuna D, Scheible WR, Carillo P, Hajirezaei MR, Stitt M. Sugar-induced increases in trehalose 6-phosphate are correlated with redox activation of ADPglucose pyrophosphorylase and higher rates of starch synthesis in *Arabidopsis thaliana*. *Biochem J*. 2006;397:139–48.
71. Stitt M, Lilley RM, Gerhardt R, Heldt HW. Metabolite levels in specific cells and subcellular compartments of plant-leaves. *Methods Enzymol*. 1989;174:518–52.
72. Cross JM, von Korff M, Altmann T, Bartzetko L, Sulpice R, Gibon Y, Palacios N, Stitt M. Variation of enzyme activities and metabolite levels in 24 *Arabidopsis* accessions growing in carbon-limited conditions. *Plant Physiol*. 2006;142(4):1574–88.
73. Lowry OH, Rosebrough NJ, Farr AL, Randall RJ. Protein measurement with the Folin phenol reagent. *J Biol Chem*. 1951;193(1):265–75.
74. Hruz T, Laule O, Szabo G, Wessendorp F, Bleuler S, Oertle L, Widmayer P, Gruissem W, Zimmermann P. Genevestigator v3: a reference expression database for the meta-analysis of transcriptomes. *Adv Bioinforma*. 2008;2008:420747.
75. Obayashi T, Kinoshita K, Nakai K, Shibaoka M, Hayashi S, Saeki M, Shibata D, Saito K, Ohta H. ATTED-II: a database of co-expressed genes and cis elements for identifying co-regulated gene groups in *Arabidopsis*. *Nucleic Acids Res*. 2007;35(Database issue):D863–9.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

