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1	Woody Plant declines. What's wrong with the microbiome?
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33 Abstract

34 Woody plant (WP) declines have multi-factorial determinants as well as a biological and 35 economic reality. The vascular system of WPs involved in the transport of carbon, nitrogen 36 and water from sources to sinks has a seasonal activity, which places it at a central position 37 for mediating plant-environment interactions from nutrient cycling to community assembly 38 and for regulating a variety of processes. To limit effects and to fight against declines, we 39 propose (i) to consider the WP and its associated microbiota as an holobiont and as a set of 40 functions, (ii) to consider simultaneously, without looking at what comes first, the 41 physiological or pathogenic disorders and (iii) to define pragmatic strategies including 42 preventive and curative agronomical practices based on microbiota engineering.

43

44 Woody plants in ecosystem functioning

Plants can be simply divided into annual, biennial (life cycle from seed to seed lasts one or two years) and perennial plants (can survive more than 2 years). Woody plants (WP) are perennial plants characterized by the presence during the non-growing season of persistent aboveground dormant parts, as well as ligneous material (mainly in vascular tissues).

50 WPs play a major role in the functioning of the ecosystem. For example, they actively 51 participate in water and CO₂ cycles, especially within complex ecosystems such as forests [1]. 52 WPs render many ecosystem services, either directly when providing economically 53 important goods and services (*e.g.* wood, fiber and fruits) or indirectly as reservoir of 54 biodiversity [1]. In a context of global warming, their carbon (C) sequestration capacity is 55 also of particular interest [2].

In their tissues or organs, WPs host a set of symbiotic and free-living microorganisms (*i.e.* archae, bacteria, fungi, oomycetes, protists and viruses) commonly referred to as the **microbiota**. Although biomolecular techniques for detecting the diversity of microorganisms are improving, host-microbiota interaction mechanisms are still poorly understood [3–5]. The emergence of the "**holobiont**" concept, *briefly defined as* a "functional entity formed by a macrobe and its long- and short-term associations with microbes and viruses" [6], opened the possibility to view an entire system.

63 In the present review, we first discuss the origin of WP declines with a focus on the 64 link between flow and storage of carbon (C), nitrogen (N) and water in WPs and their

65 transfer to/from associated microbes, in healthy or declining conditions. Then, we introduce 66 definitions and concepts about the interactions between WPs and their microbiota, often 67 inspired by knowledge on animal-associated microbiota. Finally, we discuss prospects of 68 microbiota engineering to enhance tolerance of WPs to declines.

69

70 Woody plant declines: pathological and physiological disorders

71 Decline versus dieback

72 In the last decades, WP declines are affecting more and more species [7]. They can be 73 defined as a long-term degeneration of plant tissues following a succession of multiple 74 negative events (e.g. abiotic stresses, climate deregulation, emergence of new pathogens, 75 biological invasions and agricultural strategies). WP declines may also be seen as a decrease, 76 over multiple years, of wood or fruit productivity, leading or not to a sudden death, the 77 latter being occasionally referred to as dieback (Box 1). Several authors use 'decline' strictly 78 for the progressive reduction of the vigor of the WP, whatever its causes and symptoms, 79 while they would associate 'dieback' to the drying of the plant that starts at the tips [8]. 80 However, the distinction between 'decline' and 'dieback' is not always made or accepted as 81 such by forest pathologists [8,9]. For better readability, we will retain only the term 'decline' 82 later in this review.

83

84 Origin of the declines and impact on productivity and ecosystem services

Individual tree mortality, estimated by the ICP forest network (www.icp-forests.net ; [10]), has doubled in Europe's temperate forests over the last three decades [11]. Per year, about 0.5 to 1.5% of trees died owing to different factors such as fungi, fire, grazing, climate change. Today, the impact of WP declines on ecosystem services has reached an apparent peak and brought to our attention the necessity to identify determinants and mechanisms leading to their occurrence [12]. However, the main mortality factor remains undetermined (**Box 2**).

Longevity and late sexual maturity of WPs make their physiology, development and ecology different from annual plants. WPs are especially vulnerable to pathogens since exposure lasts for a longer period of time compared to annual plants [13]. WPs are also increasingly exposed to pathogens due to silvicultural and fruit farming techniques, and

96 especially globalization, favoring fast dispersal of microbe species along commercial routes97 [14].

98 Compared to less complex or non-multifactorial diseases, it is difficult to define a 99 state of decline based on the observed symptoms, as these vary widely according to the 100 species or individuals concerned. Declines may affect different compartments and the 101 symptoms may be diverse (*i.e.* twigs drying, leaves yellowing, wood rotting). Since declines 102 result from the combination of various factors, the association of given symptoms to their 103 origin is challenging. In most declines, the whole plant is damaged including woody parts and 104 leaves. A given decline can first appear in a specific compartment, then spread to others, and 105 finally affect the balance of the whole holobiont. After a while, it leads to visible symptoms 106 such as a significant loss of biomass as shown in poplar [15] and ash [16,17], or a decrease in 107 the quantity and quality of fruits (e.g. grapevine or olive tree) [18,19]. In addition to the 108 economic impact (e.g. loss of quality, quantity, yield), forest diseases can affect some 109 ecosystem services either directly (e.g. C sequestration, water purification, soil stability) or 110 indirectly (e.g. associated biodiversity, recreational and cultural aspects) [20,21].

111

112 Carbon, nitrogen and water in WP declines: fueling and fluxes problems?

113 WP species exhibit a wide range of gas exchange responses and hydraulic strategies, 114 depending on whole-tree conductance and rooting depth. Photosynthesis essentially takes 115 place in leaves during the light period. The Rubisco-fixed C is partly accumulated (i) as starch 116 in the chloroplast to sustain metabolism during the dark period and (ii) as hexoses and 117 sucrose in the cytosol for both daily metabolism and plant long distance transport. The 118 excess of C is exported for long-term storage in the trunk and roots. Within the plant, 119 carbohydrate are transported from photosynthetic and storage tissue (sources) to areas of 120 active growth and metabolism (sinks). This phenomenon is at least partially regulated by a 121 large number of monosaccharide- and sucrose-specific transporters, identified in WPs as 122 poplar and grapevine [22,23]. Besides C, nitrogen (N) is required in significant quantities as it 123 constitutes 1–5% of the WP dry weight [24]. N compounds are taken-up directly from the 124 soil either under organic or inorganic forms directly through the plant roots- or the 125 mycorrhizal-pathway. They are then transferred to the upper parts in the form of amino 126 acids via an interconnection of the xylem and the phloem (Box 4) [23,25]. Because N-

127 containing compounds are necessary for C assimilation, N mobilization and C assimilation
128 are probably tightly co-regulated to allow the optimal development of WPs [26,27].

129 C storage organs, varying depending on the season [28], play a key role in annual and 130 seasonal metabolism and growth variations in WPs [26], as it represents resources that build 131 up in the plant and can be mobilized in the future. This is especially true in deciduous species 132 where in spring remobilization of reserves is necessary for budburst and growth of new 133 leaves, allowing functional photosynthesis [29]. N is stored before leaf fall. Two thirds of the 134 foliar N, mainly as Rubisco, which is primarily a storage protein, is recycled during the leaf 135 senescence process, then transported and stored as amino acids and proteins in the 136 perennial parts of the tree [30]. C and N stored in autumn in the trunk and roots are used for 137 wood formation, especially for the phloem [31], before N root uptake is efficient.

138 As C, N and water transport depends on source and sink activity, any disturbance 139 could influence WP functions, from growth and allocation to defense and reproduction. 140 Leaves, trunks and roots perform unique functions, necessary for a healthy development. 141 Interruptions caused by short, long term or successive stresses, in any of the functions 142 performed by leaves (decrease of photosynthesis leading to less food and fewer energy 143 storage), trunks (damage to vascular tissues reduces or inhibits C, N and water transport) 144 and roots (improper soil conditions and mechanical damages lead to lesser root extension) 145 may lead to plant starvation. These interruptions, combined with defenses against other 146 diseases (physiological or pathogenic), could finally lead to decline. In addition, several 147 groups have recently proposed to link forest declines with microbiome modification or 148 imbalance [32].

149

150 **The WP holobiont**

The recent explosion of the number of studies on human gut microbiota highlighted links between altered microbiota (dysbiosis) and development of immune-mediated and metabolic pathologies such as obesity, type-2 diabetes, or cancer [33]. In this context, one could imagine a transposition to the study of links between taxonomic composition and functioning of the WP microbiota and decline outcomes.

156

157 *Key concepts on the plant holobiont*

158 Plants do not live alone, but are colonized and surrounded by microbes. Microbial 159 colonization, whether **epiphytic** or **endophytic**, vary between plant tissues. A wide diversity 160 of microbial taxa is involved in these associations going from pathogenic to beneficial 161 through neutral [34,35] (Figure 1) as revealed by more and more sophisticated methods (see 162 for instance Mercado-Blanco, 2018 [3] and Outstanding question 1). The microbiota 163 corresponds to all microbes associated with a plant and the pool of microbial genes 164 corresponds to the **microbiome** [4,36]. In addition, it appears that (i) from one individual to 165 another the microbiota and the microbiome are different, (ii) within the same individual, 166 several microbiomes or microbiota co-exist among different tissues [37,38] and (iii) microbes 167 might support various plant functions. Therefore, to investigate a potential link between the microbiota or microbiome and declines, it is necessary to (i) analyze the diversity of 168 169 associated microbes, (ii) determine how microbes interact amongst themselves and with the 170 plant and (iii) highlight their respective potential functions and their impact on WP 171 metabolism and development.

172 The short and long-term interactions between host-microbe and microbe-microbe 173 mediate both host and microbial properties and are essential for the health of the host 174 [3,34,39]. Plant microbiota is carried out either by vertical transmission through the seeds or 175 by horizontal transmission through environmental sources [40]. Within plant microbiota, 176 beneficial microbes (i) provide or extend major metabolic functions of plants, especially in 177 terms of nutrient acquisition [3,41,42], (ii) promote plant resistance against biotic and 178 abiotic stresses [43,44], (iii) mediate metabolites production and (iv) support seed 179 germination [45]. Considering the importance of the gained functions, some authors account 180 microbiome for a 'second plant genome'. Combination of plant host and microbiota is thus 181 referred to as 'holobiont' (Figure 1), which can be defined as 'the genomic reflection of the 182 complex network of symbiotic interactions that link an individual of a given taxon with its 183 associated microbiome' [42,46,47]. The holobiont concept applied to plants allows a new 184 interpretation of the facilitation concept in plant ecology, the microbiota being considered 185 as a 'facilitator' element providing additional functions to the holobiont and adjusting to 186 environmental conditions that are optimal for survival and to keep or reinforce plant 187 homeostasis [48].

The **hologenome** theory takes the holobiont as a unit of selection in evolution [49], where the microbiome is associated with the plant genome both evolving in parallel. Pioneer descriptions of plant microbiota have been published for model plants such as *Arabidopsis thaliana, Medicago truncatula* and *Lotus japonicus* [50–52]. Microbiota of WPs has been described on poplar [53–55], oak [56,57] and grapevine [5], either on a plant compartment, including root system [34], trunk [58], **phyllosphere** [55], fruits [59], or as a comprehensive analysis [54,60].

196

197 Concept of 'core microbiota': any bacterial or fungal species shared in healthy and diseased 198 plants

199 Plant diseases, either physiological or ecological, are often associated with changes 200 or compositional deviations from a 'core microbiota' of the holobiont [50] (Figure 2). The 201 core microbiota concept emerged with the discovery that plant microbiota differs from 202 rhizospheric microbiota. Briefly, healthy plants favor colonization by commensal microbes 203 and avoid its colonization by pathogens [3,50]. The core microbiota varies with plant age and 204 compartment as well as soil properties [61,62]. The idea of a 'functional core microbiota' 205 playing a central role in plant physiology and health is now admitted [63], and is often 206 described as the sum of 'the good' (plant beneficial microbes), the 'bad' (plant pathogenic 207 microbes) and 'the ugly' (human pathogenic microbes) [35]. From one plant to another, core 208 microbiota does not consist of identical microbial taxa, but rather different taxa that ensure 209 the same set of essential functions for the holobiont fitness [63]. Key or hub species of this 210 core microbiota can be affected during the onset of declines, making the holobiont 211 ineffective against the spread of symptoms.

212

213 *The holobiont functioning: exchange of good practices between partners?*

A chemical and molecular dialogue within holobiont. Within the holobiont, plant-microbe and microbe-microbe communicate mainly through chemical and molecular signaling [64]. A large part of microbiome studies focused on the rhizosphere were dedicated to quorum sensing (strictly microbe-microbe communication) and signaling molecules [65] such as volatile organic compounds, phytohormones or antimicrobial peptides [66]. Different communication tools are used to recruit beneficial microbes, to activate desirable microbe traits, induce systemic defense, coordinate microbial population behavior and activity, shape rhizomicrobiome or establish mutualistic symbiosis with fungi or bacteria [66]. For instance, mycorrhizal fungi form mutualistic associations called **mycorrhizas** [57,67–70], able to transfer signaling molecules and nutrients between plants through **common mycorrhizal networks** [71] (**Box 4**). Disruption of this communication tools could be linked to the evolution of declines in WP (see **Outstanding question 2**).

226

227 Around the roots: a top exchange zone. The rhizosphere and the mycorrhizosphere refer to 228 the zone of influence created by roots alone or with the mycelium of mycorrhizal fungi, 229 influencing the composition and function of its microbiota through their exudates, and vice 230 versa [71,72]. The soil is subjected to numerous abiotic and biotic stresses, affecting the 231 diversity, abundance and activity of living organisms in the soil and consequently the 232 functioning of the ecosystem [73]. Soil microbial community can affect plant survival, growth 233 and tolerance to biotic and abiotic stresses and induce systemic resistance against pests and 234 pathogens both in leaves and roots [34,74,75]. The microbial lifestyle could switch from 235 rhizospheric to mycorrhizospheric or from epiphytic to endophytic depending on 236 environmental factors [76].

237 C and N are the main fuels for plant development and the basis of **rhizodeposition** by 238 root exudation of primary metabolites. Rhizodeposition describes the C-root loss from 239 plants, ranging from 10 to 40 % of assimilated C [77]. The flux of root-derived C (sugars, 240 sugar alcohols, amino acids and phenolics) constitutes a substantial nutrient release in soils, 241 with sugars the main C source for microbes as mycorrhizal fungi and bacteria [78,79]. It was 242 suggested that the more C translocated from leaves to roots, the more C ought to be exuded 243 from roots (e.g. [80]) and that C exudation by roots is controlled by factors drawing C into 244 the rhizosphere, such as root colonization by fungi (e.g. [81]). In addition to sugars, released 245 amino acids are used by microbial communities as C-N source, but are also recognized by 246 microbial chemoreceptors involved in the early root colonization [82]. Several authors have 247 proposed that differences in sugars and amino acids concentration between root cells and 248 root vicinity mediate root exudation [83], but mechanisms and transport systems remain 249 largely unknown.

250

251 C-N pools, WP declines and associated microbes: a vicious circle?

252 WP storage is highly sensitive to variations in environmental factors. Pathogens 253 involved in declines accentuate some of these physiological dysfunctions in C-N production 254 and flow. In several WP declines, physiological functions needed to assure proper storage 255 are disrupted, although such disruptions might be associated with other type of diseases. For 256 example, in the olive tree CoDiRO disease (Box 1), leaves are subjected to chlorosis or 257 wilting, leading to the reduction of the leaf surface area and a lower photosynthesis rate. In 258 Eutypa dieback or Esca disease in grapevine [84], anatomical analyses have shown a 259 degradation of the photosynthetic systems (e.g. chloroplast alteration, thylakoid and 260 endomembrane deformation). In addition, leaf degradation leads to a reduction of stomatal 261 conductance, gas exchanges between atmosphere and the plant, and hydraulic lift from the 262 soil to the leaves [84].

263 C and N storage can also be reduced after vascular system degradation or plug-264 blocking. Cankers in the trunk disturb water flux by degrading xylem vessels. The bacterium 265 *Xylella fastidiosa*, involved in Pierce disease and CoDiRO, forms plugs and blocks xylem sap 266 circulation [85]. In the case of horse chestnut bleeding canker, presence of Pseudomonas 267 syringae is related to the stopping of water flow through the trunk [13]. In grapevine, in vitro 268 tests showed disappearance of all starch reserves after 18 months of Eutypa lata activity 269 [86]. Following the damage, three allocations for the non-structural sugars are possible: (i) 270 storage for re-growth when the threat has passed (i.e. poplar; [87]), (ii) growth of sink 271 tissues such as fine roots or young leaves, and (iii) synthesis of defensive compounds 272 (poplar: [88]; *Quercus ilex*: [89]). Similar reactions following the damage were recorded for 273 N allocation on common milkweed and red oak [90,91]. N deprivation in the previous 274 autumn (N blocked in roots and woods) leads to budburst disturbance, C resource depletion 275 and tree decline in spring. Finally, root architecture could be modified leading to the 276 reduction of water and N uptake [92].

Finally, the question of which came first, the physiological or ecological disease, is still open (Figure 3). Is it possible that a monofactorial disease, readily identifiable, leads to a weakening of the WP? This would make the affected WP more susceptible to other stresses leading to an overall decline. Conversely, it can be assumed that multiple stresses applied to a WP weaken it, lowering its innate immune response when a specific pathogen arrives, acting like an opportunist (**Box 2**).

283 This question is still open depending on the type of pathogen involved and the type of stress 284 applied. Indeed, a strict biotrophic agent can be disadvantaged during the infection of a 285 plant already stressed and deficient, which cannot provide the necessary nutrients for its 286 metabolism. On the contrary, saprophytes (e.g. the GTD responsible fungi) will be favored 287 within already weakened plants. These saprophytic pathogens are often latent members of 288 the plant holobiont, but their virulence would only be expressed during a modification of the 289 microbial community due to the previous weakening of the plant (with for example an 290 exclusion of beneficial endophytes when the carbon status of the plant no longer allows the 291 survival of these).

292

293 Nevertheless, we know that imbalances of microbiota are observed in many declines 294 (Boxes 2 & 3). In Acute Oak Decline, comparison between diseased and healthy trees 295 revealed microbiota modifications, with a predominance of Brenneria goodwinii, Rahnella 296 victoriana and Gibbsiella quercinecans in diseased tissues ([57,67–70]. In silver fir dieback, 297 the link between modifications of the microbiota and the presence of disease is not clearly 298 established, even if tree decline has an impact on soil microbiota composition [93]. Decline 299 of photosynthesis rate leads to modify roots and mycorrhizal exudation, changing bacterial 300 growth and community composition as well as nitrification process in and near 301 the rhizosphere and mycorrhizosphere [94–96].

302

303 Engineering the microbiota, a realistic perspective to cure WPs from declines?

Plant microbiota protect the plant against pathogens *via* direct (production of bacterial toxins) or indirect effect (stimulation of the host immune response and competition for ecological niches) [13,36,39]. Cultural practices are known to impact the composition of WP associated microbiota [97–99]. Therefore, adjusting these practices might be a straightforward option to reduce, prevent or even cure declines [3].

The development of agroecological practices, based on the valorization of ecosystem services and of natural biological regulations, responds to a strong societal demand [100]. In this context, beneficial microbes promoting the growth, nutrition and health of plants constitute the basis for a model of functional community useful in the development of agroecological practices [101]. The majority of the results on these approaches come from crops, mostly annual plants like rice, tobacco, tomato, maize or potato (for review, see

Compant et al., 2019 [102]). Transferring this knowledge for WP management, requires
specific adaptation to perennial plants:

317 Development of new rootstock-graft assemblies and ungrafted plants: having a better 318 tolerance to diseases using beneficial synthetic communities within the selection 319 process. It is tempting to hypothesize that better ecosystem services might be explained 320 by roots and rootstock capacity to associate with different microbiota [103]. The 321 physiological aspects of the rootstock-graft interactions may affect the physiology of the 322 plant and it is possible to optimize the assembly between the two vegetative parts [104– 323 106]. In addition, WP habitat can be also dependent on the local microbiome and 324 especially on symbiotic organisms [107].

Association of tree species with beneficial microbiota: sharing the benefits of the microbiota from one individual to another can be considered directly by pooling the microbiota in the soil (especially thanks to the mycorrhizae, see Box 4) or indirectly by chemical communication between microorganisms. After identification of the microbiota associated with a tree species and the functional benefits associated with it (*i.e.* root growth promotion), it will be possible to select species and associate them with others for a "microbial complementarity".

332

333 Conserving microbial functions, an important issue for microbiota engineering

334 Plant-associated microbiota is a key factor to buffer the effects of biotic and abiotic 335 constraints [65]. However, the description of the taxonomic composition of plant microbiota 336 remains incomplete with the usual and current molecular tools and the processes regulating 337 their assembly need to be further studied [108]. The understanding of the assembly of the 338 microbiota is a research theme of fundamental importance to understand the phenomena of 339 facilitation and rapid adaptation of plants to local environmental constraints. In this line of 340 research, Lapsansky and collaborators [109] proposed the concept of "soil memory" : from 341 one plant generation to another, a given soil would hold its associated rhizospheric and 342 edaphic microbiota. The fitness of perennial plants of the n+1 generation (or of a new woody 343 plant set up in a plot replanting context) would thus be improved, taking advantage of the 344 pre-existing beneficial microbes for their development [109]. Time scales are often 345 considered on an annual or biannual basis for agronomic plants with a short life cycle, but

this concept can also be applied to the time scales necessary for the renewal of a forest, a vineyard or at least an orchard. The concept of soil memory could be exploited artificially for example by sampling a given well-balanced forest soil microbiota, multiply it in controlled conditions, before introducing it into nursery greenhouses in plantlets soil.

350

351 Acting directly on the microbiota composition by microbial inoculation

352 Some bacteria, endophytic and mycorrhizal fungi can be used as microbial inoculants, 353 owing to their functional properties of antibiosis, lysis, competition or nutrient acquisition, 354 induced resistance or hormonal stimulation. Their functions depend on both abiotic and 355 biotic context [110]. With an annual growth rate of about 10 %, the economic market of 356 microbial inoculants increases rapidly [111]. Although field results on annual crops 357 compared to market gardening are often disappointing compared to promising advances 358 under controlled conditions, we can already try to modify root microbiota of woody species 359 such as oak for Acute Oak Decline [56]. For example, the beneficial role of *Fraxinus ornus* 360 fungal endophytic microbiota against ash decline was suggested, although results need to be 361 further supported [112].

362 Microbial inoculation (see **Outstanding question 3**) could be achieved:

in nursery by inoculating a single or a cocktail of microbial strains by selecting of native
 or locally adapted inoculants (considering both biotic and abiotic context) or strains
 with a marketing authorization [113]

- on established or mature WP by direct addition of bacterial strains or cocktails of
 strains by spraying (rather for endophytes) or by reintroduction of mycelial strains as
 spores or mycelium directly in the soil (*i.e.* mycorrhizal fungi, [114]). Such additions
 could be repeated until the microbial imbalance ends.
- by indirect regular supplementation by complantation (*i.e* replacing dead plants, a
 cultural practice largely used in vineyards) or by using plant cover as a reservoir of
 diversity [115].

A key issue is that the introduction of beneficial microbes into an established holobiont is not easy to sustain over time. The microbial populations introduced by humans can decrease over time and thus their beneficial effects disappear. They can also eliminate native beneficial microbial species, changing the functional balance of the ecosystem.

377 Nursery manipulations (early stages of plant development) would reduce the odds of 378 disturbing an established microbial equilibrium [109]. Additionally, it can be difficult to 379 switch from an 'unbalanced microbiota state' to a 'healthy microbiota state', without coming 380 back to the original microbial communities because of the resilience of the system, which 381 would make it difficult to keep the new equilibrium over time [116]. Additionally, when 382 microbial strains are characterized as beneficial for plants, the question of the specificity of 383 certain strains for targeted cultivars will prevail. Results from lab experiments may be 384 difficult to implement in the field [39]. Finally, it becomes clearly necessary to include 385 microbiota in plant breeding strategies and in the analysis of varieties and rootstocks 386 performance to limit the loss of microbial genetic diversity, inherent in current agricultural 387 systems and practices.

388

389 Acting indirectly on the microbiota composition: impact of agricultural practices.

Some agricultural methods are specific of WPs agro-management and might act indirectly on microbiota composition. For example, yearly dead parts of woody plants are often removed because they are thought to contain detrimental microorganisms. However, they represent a valuable carbon source and their removal would contribute to the reduction of organic compounds within the soil which in turn affect soil microbiome [117,118]. If the risks of contamination are too high for some declines and prevent application of grinding dead wood, another option might be to spread dead parts after complete or partial combustion.

397

398 Grafting is a technique widely used within WPs, such as grapevine, olive and citrus 399 trees [73]. Some cultivars or rootstocks as well as clones are more tolerant to pathogens or 400 are physiologically different: the nature of the host influences the susceptibility to a disease 401 (e.g. GTDs in grapevine [119]), an apoplectic disorder or a physiological disturbance (e.g. the 402 decline of the grapevine associated with the use of the rootstock 161-49C, see Box 1) 403 [120,121]. As for GTDs, some olive tree cultivars are more susceptible to CoDiRO than 404 others. For example, cultivar 'Leccino' seems to be more resistant [122]. For the grapevine 405 Syrah decline, it has been shown that even if all rootstocks are concerned, 110R and 99R are 406 more sensitive than others, so their use is not recommended. A very strong clone effect was 407 also observed [123]. These differences in susceptibility could be related to differences in 408 associated microbiota. Rhizodeposition, which is related to the recruitment of rhizospheric

409 microbes, has been shown to depend on the genotype of the rootstock in apple trees [124]. 410 Within other plant compartments, an influence of the rootstock-scion combination genotype 411 and of WP genotype on microbial diversity is also suggested (*e.g.* the endophytic microbiota 412 in apple trees [125], the bacterial endophytes in olive trees [126], the rhizospheric 413 microbiota in grapevine [127] or the arbuscular mycorrhizal community in citrus [128].

414

It has been shown that the diversity of organisms (plants, animals and microbes) is positively affected in organically managed or low-farming systems, compared to systems under integrated management [102]. Soil diversity has been studied, particularly in vineyards, where a link was found between soils under green manure and highest soil microbiota diversity, compared to other organic or biodynamic agro-management practices [129].

Finally, since microbiota associated with plants can vary from one species to another, and even from one individual to another within the same species, strategies of association of different species within the same agricultural system can be considered [130]. Thus, a plant could take advantage of the beneficial effects of the microbiota of the neighboring plant, in particular the rhizospheric microbiota, and *vice versa* (see **Outstanding questions 4 and 5**).

425

426 **Concluding remarks and future perspectives**

427 Improving longevity of WPs is a major challenge considering their declines. A major feature 428 would be to understand, not only the physiology and the ecology of WPs, but also of their 429 holobionts. Microbiota and by extension the microbiome could offer new engineering 430 solutions to prevent and even cure WP declines only if (i) we take into account the specific 431 physiology of WPs, especially fluxes of C, N and water between sources and sinks that are 432 changing over seasons and years and (ii) the chemical and molecular dialogues between 433 microbes and between the WP and microbes. Keeping the homeostasis of WP holobiont 434 would necessitate to carefully balance its resources between growth, storage, reproduction 435 and defense. From nursery to fruit and berry plantations, considering the use of microbiota portfolio adapted to specific conditions (i.e. climatic, chemical and biological stresses) will 436 437 constitute a major step towards a better understanding and management of the agro-438 ecological outcome of WPs holobionts (see **Outstanding questions**).

439

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448

449 **References**

- 450 1 Costanza, R. *et al.* (1997) The value of the world's ecosystem services and natural capital.
 451 *Nature* 387, 8
 452 2 Jackson B. P. *et al.* (2002) Ecosystem carbon loss with weadly plant invasion of smaller decaystem.
- 452 2 Jackson, R.B. *et al.* (2002) Ecosystem carbon loss with woody plant invasion of grasslands.
 453 *Nature* 418, 623–626
- 454 3 Mercado-Blanco, J. *et al.* (2018) Belowground Microbiota and the Health of Tree Crops.
 455 *Frontiers in Microbiology* 9,
- 456 4 Schlaeppi, K. and Bulgarelli, D. (2015) The plant microbiome at work. *Molecular Plant-Microbe* 457 *Interactions* 28, 212–217
- 458 5 Pinto, C. *et al.* (2014) Unravelling the diversity of grapevine microbiome. *PLoS ONE* 9, e85622
- Gordon, J. *et al.* (2013) Superorganisms and Holobionts: Looking for a term for the functional
 entity formed by a macrobe and its associated symbiotic microbes and viruses? The term is
 "holobiont." *Microbe Magazine* 8, 152–153
- 4627Anderson, P.K. *et al.* (2004) Emerging infectious diseases of plants: pathogen pollution, climate463change and agrotechnology drivers. *Trends in Ecology & Evolution* 19, 535–544
- 464 8 Fraedrich, B. and Experts, B.T. Dieback and Decline.
- 465 9 Ciesla, W.M. *et al.* (1994) *Decline and Dieback of Trees and Forests: A Global Overview*, Food &
 466 Agriculture Org.
- 467 10 Neumann, M. *et al.* (2017) Climate variability drives recent tree mortality in Europe. *Global*468 *Change Biology* 23, 4788–4797
- 469 11 Senf, C. *et al.* (2018) Canopy mortality has doubled in Europe's temperate forests over the last
 470 three decades. *Nature Communications* 9, 4978
- 471 12 Boyd, I.L. *et al.* (2013) The Consequence of Tree Pests and Diseases for Ecosystem Services.
 472 *Science* 342, 1235773–1235773
- 473 13 Koskella, B. *et al.* (2017) A signature of tree health? Shifts in the microbiome and the ecological
 474 drivers of horse chestnut bleeding canker disease. *New Phytologist* 215, 737–746
- 475 14 Ennos, R.A. (2015) Resilience of forests to pathogens: an evolutionary ecology perspective.
 476 *Forestry* 88, 41–52
- 477 15 Steenackers, J. *et al.* (1996) Poplar diseases, consequences on growth and wood quality.
 478 *Biomass and Bioenergy* 10, 267–274
- 47916Coker, T.L.R. *et al.* (2019) Estimating mortality rates of European ash (*Fraxinus excelsior*) under480the ash dieback (*Hymenoscyphus fraxineus*) epidemic. *PLANTS, PEOPLE, PLANET* 1, 48–58
- 481 17 Matisone, I. *et al.* (2018) Statistics of ash dieback in Latvia. *Silva Fennica* 52,
- 482 18 Gramaje, D. *et al.* (2018) Managing Grapevine Trunk Diseases With Respect to Etiology and
 483 Epidemiology: Current Strategies and Future Prospects. *Plant Disease* 102, 12–39
- 484 19 Andrea Luvisi *et al.* (2017) Sustainable Management of Plant Quarantine Pests: The Case of
 485 Olive Quick Decline Syndrome. *Sustainability* 9, 659

486	20	Freer-Smith, P.H. and Webber, J.F. (2017) Tree pests and diseases: the threat to biodiversity
487		and the delivery of ecosystem services. <i>Biodiversity and Conservation</i> 26, 3167–3181
488	21	Singh, J. et al. (2019) Potential role of weather, soil and plant microbial communities in rapid
489		decline of apple trees. PLOS ONE 14, e0213293
490	22	Lecourieux, F. et al. (2014) An update on sugar transport and signalling in grapevine. Journal of
491		Experimental Botany 65, 821–832
492	23	Liesche, J. (2017) Sucrose transporters and plasmodesmal regulation in passive phloem loading:
493		Mechanism and regulation of passive phloem loading. <i>Journal of Integrative Plant Biology</i> 59,
494		311–321
495	24	Sauter, J.J. and van Cleve, B. (1989) Immunochemical localization of a willow storage protein
496	25	with a poplar storage protein antibody. <i>Protoplasma</i> 149, 175–177
497	25	Prautsch, S. et al. (2015) Philoem as Capacitor: Radial Transfer of Water into Xylem of Tree
498	26	Stems Occurs via Symplastic Transport in Ray Parenchyma. Plant Physiology 167, 963–971
499 500	20	Noronna, H. <i>et ul.</i> (2018) A molecular perspective on starch metabolism in woody tissues.
501	27	Pluillu 246, 555–506 Zhang H. et al. (2018) Posponsos of Woody Plant Eurostianal Traits to Nitrogon Addition: A
502	27	Meta-Analysis of Leaf Economics, Gas Exchange, and Hydraulic Traits, Econtiers in Plant Science
502		0 683
504	28	5,005 Furze M E <i>et al.</i> (2019) Whole-tree nonstructural carbohydrate storage and seasonal dynamics
505	20	in five temperate species. New Phytologist 221, 1466–1477
506	29	Merchant A (2016) The importance of storage and redistribution in vascular plants. <i>Tree</i>
507	25	Physiology 36, 533–535
508	30	Dickson, R.E. (1989) Carbon and nitrogen allocation in trees. Annales des Sciences Forestières
509		46. 631s-647s
510	31	Hansen, J. <i>et al.</i> (1996) Assimilation, allocation and utilization of carbon by 3-year-old Scots
511		pine (. <i>Trees</i> 11, 83
512	32	Terhonen, E. <i>et al.</i> (2019) Forest Tree Microbiomes and Associated Fungal Endophytes:
513		Functional Roles and Impact on Forest Health. Forests 10, 42
514	33	Forslund, K. et al. (2015) Disentangling type 2 diabetes and metformin treatment signatures in
515		the human gut microbiota. <i>Nature</i> 528, 262–266
516	34	Berendsen, R.L. et al. (2012) The rhizosphere microbiome and plant health. Trends in Plant
517		Science 17, 478–486
518	35	Mendes, R. et al. (2013) The rhizosphere microbiome: significance of plant beneficial, plant
519		pathogenic, and human pathogenic microorganisms. FEMS Microbiology Reviews 37, 634–663
520	36	Compant, S. et al. (2016) Editorial special issue: soil, plants and endophytes. Plant Soil 405, 1–
521		11
522	37	Hardoim, P.R. et al. (2015) The Hidden World within Plants: Ecological and Evolutionary
523		Considerations for Defining Functioning of Microbial Endophytes. Microbiology and Molecular
524		Biology Reviews 79, 293–320
525	38	Philippot, L. <i>et al.</i> (2013) Going back to the roots: the microbial ecology of the rhizosphere.
526		Nature Reviews Microbiology 11, 789–799
527	39	Berg, G. <i>et al.</i> (2016) The plant microbiome explored: implications for experimental botany.
528		Journal of Experimental Botany 67, 995–1002
529	40	Baltrus, D.A. (2017) Adaptation, specialization, and coevolution within phytobiomes. <i>Current</i>
530		Opinion in Plant Biology 38, 109–116
531	41	Hacquard, S. et dl. (2015) Microbiota and Host Nutrition across Plant and Animal Kingdoms. Cell
532 522	40	Host & Microbe 17, $603-616$
525 521	42	Valuenkoommuyse, P. et al. (2015) The importance of the microbiome of the plant holobiont.
534 535	12	New Fily cologist 200, 1130-1200 Podolich O at al. (2015) Reviving of the enderbytic bactorial community as a putative
536	+J	mechanism of plant resistance. <i>Plant and Soil</i> 388, 367–377
550		$\frac{1}{3}$

Rolli, E. et al. (2015) Improved plant resistance to drought is promoted by the root-associated microbiome as a water stress-dependent trait: Root bacteria protect plants from drought. Environmental Microbiology 17, 316–331 Truyens, S. et al. (2015) Bacterial seed endophytes: genera, vertical transmission and interaction with plants: Bacterial seed endophytes. Environmental Microbiology Reports 7, 40-Berg, G. et al. (2014) Unraveling the plant microbiome: looking back and future perspectives. Frontiers in Microbiology 5, Guerrero, R. et al. (2013) Symbiogenesis: the holobiont as a unit of evolution. International Microbiology DOI: 10.2436/20.1501.01.188 Sánchez-Cañizares, C. et al. (2017) Understanding the holobiont: the interdependence of plants and their microbiome. Current Opinion in Microbiology 38, 188–196 Carrier, T.J. and Reitzel, A.M. (2017) The Hologenome Across Environments and the Implications of a Host-Associated Microbial Repertoire. Frontiers in Microbiology 8, 802 Lundberg, D.S. et al. (2012) Defining the core Arabidopsis thaliana root microbiome. Nature 488, 86-90 Tkacz, A. et al. (2015) Stability and succession of the rhizosphere microbiota depends upon plant type and soil composition. The ISME Journal 9, 2349-2359 Zgadzaj, R. et al. (2016) Root nodule symbiosis in Lotus japonicus drives the establishment of distinctive rhizosphere, root, and nodule bacterial communities. Proceedings of the National Academy of Sciences 113, E7996–E8005 Blair, P.M. et al. (2018) Exploration of the Biosynthetic Potential of the Populus Microbiome. 3, Cregger, M.A. et al. (2018) The Populus holobiont: dissecting the effects of plant niches and genotype on the microbiome. Microbiome 6, Hacquard, S. and Schadt, C.W. (2015) Towards a holistic understanding of the beneficial interactions across the Populus microbiome. New Phytologist 205, 1424–1430 Maghnia, F.Z. et al. (2019) The rhizosphere microbiome: A key component of sustainable cork oak forests in trouble. Forest Ecology and Management 434, 29-39 Sapp, M. et al. (2016) Metabarcoding of Bacteria Associated with the Acute Oak Decline Syndrome in England. Forests 7, 95 Bruez, E. et al. (2016) Various fungal communities colonise the functional wood tissues of old grapevines externally free from grapevine trunk disease symptoms: Fungal microflora of GTD-free old vines. Australian Journal of Grape and Wine Research 22, 288-295 Droby, S. and Wisniewski, M. (2018) The fruit microbiome: A new frontier for postharvest biocontrol and postharvest biology. Postharvest Biology and Technology 140, 107–112 Kovalchuk, A. et al. (2018) Mycobiome analysis of asymptomatic and symptomatic Norway spruce trees naturally infected by the conifer pathogens *Heterobasidion* spp.: Mycobiome of Norway spruce. Environmental Microbiology Reports 10, 532–541 Hamonts, K. et al. (2018) Field study reveals core plant microbiota and relative importance of their drivers: Core microbiota of field-grown sugarcane and their drivers. Environmental Microbiology 20, 124–140 Zarraonaindia, I. et al. (2015) The soil microbiome influences grapevine-associated microbiota. mBio 6, Lemanceau, P. et al. (2017) Let the Core Microbiota Be Functional. Trends in Plant Science 22, 583-595 Rosier, A. et al. (2016) A perspective on inter-kingdom signaling in plant-beneficial microbe interactions. Plant Molecular Biology 90, 537–548 Berg, G. et al. (2017) Plant microbial diversity is suggested as the key to future biocontrol and health trends. FEMS Microbiology Ecology 93, Venturi, V. and Keel, C. (2016) Signaling in the Rhizosphere. Trends in Plant Science 21, 187–198

588 67 Meaden, S. et al. (2016) The effects of host age and spatial location on bacterial community 589 composition in the English Oak tree (Quercus robur): Age-related decline in bacterial diversity of 590 Oak trees. Environmental Microbiology Reports 8, 649–658 591 68 Broberg, M. et al. (2018) Integrated multi-omic analysis of host-microbiota interactions in acute 592 oak decline. Microbiome 6, 593 69 Denman, S. et al. (2018) Microbiome and infectivity studies reveal complex polyspecies tree 594 disease in Acute Oak Decline. The ISME Journal 12, 386-399 595 70 Doonan, J. et al. (2019) Genomic analysis of bacteria in the Acute Oak Decline pathobiome. 596 Microbial Genomics 597 Wipf, D. et al. (2019) Trading on the arbuscular mycorrhiza market: from arbuscules to common 71 598 mycorrhizal networks. New Phytologist DOI: 10.1111/nph.15775 599 72 Shi, S. et al. (2016) The interconnected rhizosphere: High network complexity dominates 600 rhizosphere assemblages. Ecol Lett 19, 926–936 601 73 Warschefsky, E.J. et al. (2016) Rootstocks: Diversity, Domestication, and Impacts on Shoot 602 Phenotypes. Trends in Plant Science 21, 418–437 603 74 Syed Ab Rahman, S.F. et al. (2018) Emerging microbial biocontrol strategies for plant 604 pathogens. Plant Science 267, 102–111 605 75 Pineda, A. et al. (2017) Steering Soil Microbiomes to Suppress Aboveground Insect Pests. 606 Trends in Plant Science 22, 770–778 607 Uroz, S. et al. (2019) Plant Symbionts Are Engineers of the Plant-Associated Microbiome. Trends 76 608 in Plant Science 24, 905–916 609 77 Hennion, N. et al. (2019) Sugars en route to the roots. Transport, metabolism and storage 610 within plant roots and towards microorganisms of the rhizosphere. Physiologia Plantarum 165, 611 44-57 612 Finzi, A.C. et al. (2015) Rhizosphere processes are quantitatively important components of 78 613 terrestrial carbon and nutrient cycles. Global Change Biology 21, 2082–2094 614 79 Kawasaki, A. et al. (2016) Microbiome and Exudates of the Root and Rhizosphere of 615 Brachypodium distachyon, a Model for Wheat. PLOS ONE 11, e0164533 616 Hill, P. et al. (2007) Response of root respiration and root exudation to alterations in root C 80 617 supply and demand in wheat. Plant and Soil 291, 131–141 618 81 Meier, I.C. et al. (2013) Fungal communities influence root exudation rates in pine seedlings. 619 FEMS Microbiology Ecology 83, 585–595 620 Sasse, J. et al. (2018) Feed Your Friends: Do Plant Exudates Shape the Root Microbiome? Trends 82 621 in Plant Science 23, 25–41 622 83 Durand, M. et al. (2018) Carbon source-sink relationship in Arabidopsis thaliana: the role of 623 sucrose transporters. Planta 247, 587–611 624 84 Fontaine, F. et al. (2016) The effects of grapevine trunk diseases (GTDs) on vine physiology. 625 European Journal of Plant Pathology 144, 707–721 626 85 Bucci, E.M. (2018) *Xylella fastidiosa*, a new plant pathogen that threatens global farming: 627 Ecology, molecular biology, search for remedies. Biochemical and Biophysical Research 628 Communications 502, 173–182 629 Bertsch, C. et al. (2013) Grapevine trunk diseases: complex and still poorly understood. Plant 86 630 Pathology 62, 243-265 631 87 Babst, B.A. et al. (2008) Lymantria dispar herbivory induces rapid changes in carbon transport 632 and partitioning in Populus nigra. Entomologia Experimentalis et Applicata 128, 117–125 633 88 Massad, T.J. et al. (2014) An optimal defense strategy for phenolic glycoside production in 634 Populus trichocarpa - isotope labeling demonstrates secondary metabolite production in 635 growing leaves. New Phytologist 203, 607-619 636 89 Sardans, J. et al. (2014) Metabolic responses of Quercus ilex seedlings to wounding analysed 637 with nuclear magnetic resonance profiling. Plant Biology 16, 395–403

638 90 Tao, L. and Hunter, M.D. (2013) Allocation of resources away from sites of herbivory under 639 simultaneous attack by aboveground and belowground herbivores in the common milkweed, 640 Asclepias syriaca. Arthropod-Plant Interactions 7, 217-224 641 91 Ullmann-Zeunert, L. et al. (2013) Quantification of growth-defense trade-offs in a common 642 currency: nitrogen required for phenolamide biosynthesis is not derived from ribulose-1,5-643 bisphosphate carboxylase/oxygenase turnover. The Plant Journal 75, 417–429 644 92 Jin, K. et al. (2017) Shaping an Optimal Soil by Root–Soil Interaction. Trends in Plant Science 22, 645 823-829 646 93 Gazol, A. et al. (2018) Beneath the canopy: Linking drought-induced forest die off and changes 647 in soil properties. Forest Ecology and Management 422, 294–302 648 94 Bardgett, R.D. et al. (2014) Going underground: root traits as drivers of ecosystem processes. 649 Trends in Ecology & Evolution 29, 692–699 650 Fransson, P. et al. (2016) Ectomycorrhizal exudates and pre-exposure to elevated CO 2 affects 95 651 soil bacterial growth and community structure. Fungal Ecology 20, 211–224 652 van Dam, N.M. and Bouwmeester, H.J. (2016) Metabolomics in the Rhizosphere: Tapping into 96 653 Belowground Chemical Communication. Trends in Plant Science 21, 256-265 654 97 Hartman, K. et al. (2018) Cropping practices manipulate abundance patterns of root and soil 655 microbiome members paving the way to smart farming. Microbiome 6, 656 98 Pascazio, S. et al. (2015) Phyllosphere and carposphere bacterial communities in olive plants 657 subjected to different cultural practices. International Journal of Plant Biology 6, 658 99 Pinto, C. and Gomes, A.C. (2016) Vitis vinifera microbiome: from basic research to technological 659 development. BioControl 61, 243-256 660 100 Altieri, M.A. et al. (2015) Agroecology and the design of climate change-resilient farming 661 systems. Agronomy for Sustainable Development 35, 869–890 662 101 Busby, P.E. et al. (2017) Research priorities for harnessing plant microbiomes in sustainable 663 agriculture. PLOS Biology 15, e2001793 664 102 Compant, S. et al. (2019) A review on the plant microbiome: Ecology, functions, and emerging 665 trends in microbial application. Journal of Advanced Research 19, 29–37 666 103 Vivas, M. et al. (2015) Maternal effects on tree phenotypes: considering the microbiome. 667 Trends in Plant Science 20, 541–544 668 104 De Roy, K. et al. (2014) Synthetic microbial ecosystems: an exciting tool to understand and 669 apply microbial communities: Synthetic microbial ecosystems. Environ Microbiol 16, 1472–1481 670 105 Großkopf, T. and Soyer, O.S. (2014) Synthetic microbial communities. Current Opinion in 671 Microbiology 18, 72–77 672 106 Johns, N.I. et al. (2016) Principles for designing synthetic microbial communities. Current 673 *Opinion in Microbiology* 31, 146–153 674 107 Rodriguez, R.J. et al. (2008) Stress tolerance in plants via habitat-adapted symbiosis. ISME J 2, 675 404-416 676 108 Morrison-Whittle, P. and Goddard, M.R. (2015) Quantifying the relative roles of selective and 677 neutral processes in defining eukaryotic microbial communities. The ISME Journal 9, 2003–2011 678 Lapsansky, E.R. et al. (2016) Soil memory as a potential mechanism for encouraging sustainable 109 679 plant health and productivity. Current Opinion in Biotechnology 38, 137–142 680 110 Hoeksema, J.D. et al. (2010) A meta-analysis of context-dependency in plant response to 681 inoculation with mycorrhizal fungi. Ecology Letters 13, 394–407 682 111 Berg, G. (2009) Plant–microbe interactions promoting plant growth and health: perspectives for 683 controlled use of microorganisms in agriculture. Applied Microbiology and Biotechnology 84, 684 11 - 18685 112 Schlegel, M. et al. (2016) Effects of endophytic fungi on the ash dieback pathogen. FEMS 686 *Microbiology Ecology* 92, fiw142 687 113 Calvente, R. et al. (2004) Analysing natural diversity of arbuscular mycorrhizal fungi in olive tree 688 (Olea europaea L.) plantations and assessment of the effectiveness of native fungal isolates as 689 inoculants for commercial cultivars of olive plantlets. Applied Soil Ecology 26, 11–19

690	114	Appleton et al - 2003 - Mycorrhizal fungal inoculation of established street trees.pdf
691	115	van der Putten, W.H. (2017) Belowground drivers of plant diversity. <i>Science</i> 355, 134–135
692	116	Toju, H. et al. (2018) Core microbiomes for sustainable agroecosystems. Nature Plants 4, 247–
693		257
694	117	Achat, D.L. et al. (2015) Forest soil carbon is threatened by intensive biomass harvesting. Sci
695		Rep 5, 15991
696	118	Trumbore, S.E. and Czimczik, C.I. (2008) An Uncertain Future for Soil Carbon. <i>Science</i> 321,
697		1455–1456
698	119	Armijo, G. <i>et al.</i> (2016) Grapevine Pathogenic Microorganisms: Understanding Infection
699		Strategies and Host Response Scenarios, Front, Plant Sci. 7
700	120	McElrone, A L <i>et al.</i> (2010) The role of tyloses in crown hydraulic failure of mature walnut trees.
701	120	afflicted by apoplexy disorder. Tree Physiology 30, 761–772
702	121	Torregrosa and Boursiguot - 2011 - The 161-49C rootstock and the decline of young
702	171	vinevards ndf
703	122	Sabella E <i>et al.</i> (2018) Xylella fastidiosa induces differential expression of lignification related-
704	122	genes and lignin accumulation in tolerant olive trees cy. Leccino, Journal of Plant Physiology
705		
700	177	220, 00-00
707	123	2011 adf
700	174	2011pul
709	124	Leisso, R. <i>et al.</i> (2018) Targeteu Metabolic Profiling Indicates Apple Rootstock Genotype-
710		Specific Differences in Primary and Secondary Metabolite Production and Validate Quantitative
/11	405	Contribution From Vegetative Growth. Frontiers in Plant Science 9, 1336
/12	125	Liu, J. <i>et al.</i> (2018) Apple endophytic microbiota of different rootstock/scion combinations
/13	4.9.6	suggests a genotype-specific influence. <i>Microbiome</i> 6, 18
/14	126	Muller, H. <i>et al.</i> (2015) Plant genotype-specific archaeal and bacterial endophytes but similar
/15		Bacillus antagonists colonize Mediterranean olive trees. Front. Microbiol. 6,
716	127	D'Amico, F. <i>et al.</i> (2018) The Rootstock Regulates Microbiome Diversity in Root and
717		Rhizosphere Compartments of Vitis vinifera Cultivar Lambrusco. Frontiers in Microbiology 9,
718		2240
719	128	Song, F. et al. (2015) The Scion/Rootstock Genotypes and Habitats Affect Arbuscular
720		Mycorrhizal Fungal Community in Citrus. Frontiers in Microbiology 6,
721	129	Longa, C.M.O. <i>et al.</i> (2017) Soil microbiota respond to green manure in organic vineyards. <i>J</i>
722		Appl Microbiol 123, 1547–1560
723	130	Wei, Z. and Jousset, A. (2017) Plant Breeding Goes Microbial. <i>Trends in Plant Science</i> 22, 555–
724		558
725	131	Bruez, E. et al. (2013) Overview of grapevine trunk diseases in France in the 2000s.
726		Phytopathologia Mediterranea 52, 14
727	132	Martelli, G.P. et al. (2016) The olive quick decline syndrome in south-east Italy: a threatening
728		phytosanitary emergency. European Journal of Plant Pathology 144, 235–243
729	133	Songy, A. et al. (2019) Grapevine trunk diseases under thermal and water stresses. Planta DOI:
730		10.1007/s00425-019-03111-8
731	134	Gomes, T. et al. (2019) Bacterial disease induced changes in fungal communities of olive tree
732		twigs depend on host genotype. <i>Sci Rep</i> 9, 5882
733	135	Dissanayake, A. (2016) Botryosphaeriaceae: Current status of genera and species. Mycosphere
734		7, 1001–1073
735	136	Aćimović, S.G. et al. (2018) Characterization and Pathogenicity of Botryosphaeriaceae Fungi
736		Associated with Declining Urban Stands of Coast Redwood in California. Plant Disease 102,
737		1950–1957
738	137	Úrbez-Torres, J.R. et al. (2014) Grapevine Trunk Diseases in British Columbia: Incidence and
739		Characterization of the Fungal Pathogens Associated with Esca and Petri Diseases of Grapevine.
740		Plant Disease 98, 469–482

Moral, J. *et al.* (2017) Identification of Fungal Species Associated with Branch Dieback of Olive
 and Resistance of Table Cultivars to *Neofusicoccum mediterraneum* and *Botryosphaeria dothidea. Plant Disease* 101, 306–316

139 Úrbez-Torres, J.R. *et al.* (2013) Olive Twig and Branch Dieback: Etiology, Incidence, and
 745 Distribution in California. *Plant Disease* 97, 231–244

746 140 Fontaine, F. *et al.* (2016) Grapevine trunk diseases. A review. *OIV Publications* DOI:
747 10.1094/PHYTO-12-18-0488-FI

Kennelly, M.M. *et al.* (2007) *Pseudomonas syringae* Diseases of Fruit Trees: Progress Toward
 Understanding and Control. *Plant Disease* 91, 4–17

- Garcia, K. *et al.* (2016) Take a Trip Through the Plant and Fungal Transportome of Mycorrhiza.
 Trends in Plant Science 21, 937–950
- Simard, S.W. *et al.* (1997) Net transfer of carbon between ectomycorrhizal tree species in the
 field. *Nature* 388, 579–582
- 144 Deja-Sikora, E. *et al.* (2019) The Contribution of Endomycorrhiza to the Performance of Potato
 755 Virus Y-Infected Solanaceous Plants: Disease Alleviation or Exacerbation? *Frontiers in* 756 *Microbiology* 10, 516
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- 758 Text Boxes
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760 Box 1. WP declines and consequences on the productivity of fruits

761 The question of which came first, the pathological or physiological disorder, is difficult to 762 answer. Most of the time, WPs are considered as asymptomatic as no visual declines are 763 observed and some plant compartments (i.e. vessels, roots) are difficult to access. So, WP 764 weakening is already under way when the first symptoms of the disease or decline appear. 765 In addition, there is a bias on several species since we are more focused on plants producing 766 wood, fibers, berries and fruits for human nutrition. Grapevine (Vitis vinifera L.) and olive 767 trees (Olea europaea L.) are worldwide distributed crops of significant economic importance 768 and are currently facing several diseases, due to physiological imbalance of WP tissues and 769 to fungal or bacterial agents. The rootstock 161-49C in grapevine develops limited C storage 770 and displays disturbance in xylem (*i.e.* thylose) and phloem (*i.e.* smaller diameter) leading up 771 to 50% of unproductive vineyard [121]. The main declines affecting mature grapevine plants 772 are grapevine trunk diseases (GTDs) caused by several fungal species [86,131]. GTDs are 773 currently affecting vineyards throughout the world. Approximately 13 % and 14% of 774 respectively the French and Californian vineyards are unproductive. The Olive Quick Decline 775 Syndrome (or CoDiRO, for Complesso del disseccamento rapido dell'olivo), whose symptoms 776 look like those of grapevine declines, is mainly caused by the bacterium Xylella fastidiosa. 777 CoDiRO, originally detected in the olive groves of Salento in Italy in the late 2000s, is

- expected to expend to the rest of the Mediterranean basin and California [132]. Worldwide
- olive production has recently decreased by about 25% and in Italy by 50%.
- 780

781 Box 2. Link between WP declines, pathogens and holobiont imbalance

782 It seems that pathogenic agents (fungal, bacterial or viral) can trigger diseases on healthy 783 trees acting as "primary factors" when using biotic diffusion vectors or as "secondary 784 factors" when the pathogen benefits from the weakening of the tree (e.g. another disease, 785 pruning wounds). This weakening can result from significant climatic (*i.e.* drought, extreme 786 fluctuation in weather conditions), chemical (*i.e.* excessive fertilizers) and biological stresses 787 (*i.e.* insects), poor plant selection for the location, physical constraints, improper planting 788 practices, poor soil conditions and physical injury. It is likely that a given disease is not 789 always related to the presence of a particular pathogen alone, but would result from a 790 combination of adverse factors and the presence of one or several pathogens. In addition, 791 external factors are necessary to influence the transition from an endophytic to a pathogenic 792 state of some fungi in asymptomatic tissues state [133]. The intensity of the disease related 793 to pathogen leads to the modification of the holobiont and depends on either the WP (*i.e.* 794 cultivar) or the balance of microbes (*i.e.* epiphytic and endophytic microbiota) [134].

795

Box 3. Are some microbial species always present in the microbiota of woody plants exhibiting different declines related to pathogens?

798 The same pathogenic species are often involved in declines affecting various WPs and act as 799 opportunists. Similar fungal species from the *Botryosphaeriaceae* (endophytes, saprophytes 800 and some pathogenic) are associated with many declines [135] and found in GTDs, Coast 801 Redwood Trees Decline [136,137] and Branch Dieback of olive [138,139]. More than 300 802 species of plants are susceptible to *Xylella fastidiosa* in the world [85,140] and some strains 803 carried by xylem-sap feeding insects (e.g. leafhoppers and spittlebugs), cause fatal diseases 804 in grapevine (Pierce disease), olive and citrus trees. The bacterium Pseudomonas syringae 805 infect orchards (e.g. cherry, pear, plum and apricot) weakened by water stress or injury and 806 are involved in horse chestnut bleeding canker [13,141].

807

808 Box 4. Common mycorrhizal networks: a role of sentinel in prevention of decline?

809 Plant-plant interactions (e.g. facilitation, competition) are among the fundamental 810 processes that shape structure and functioning of plant communities. Within the fungal 811 component of the root microbiota, mycorrhizal fungi form mutualistic symbiotic associations 812 called mycorrhizas [142]. Around 90 % of terrestrial plant species including fruit and olive 813 trees, grapevine and tree species cultivated for their wood are mycorrhized. Mycorrhizal 814 fungi are able to form **common mycorrhizal networks (CMN)** linking roots from the same or 815 different species. CMNs benefit to host plant in many ways, including plant-plant facilitation 816 or nutrition [3]. Since the highlighting of intra-kingdom communication between plants and 817 microbes, especially between plants and bacteria, it has been shown that molecules other 818 than nutrients can pass between microbes and their host plant. Bidirectional nutrient 819 transfers of carbon (C) between ectomycorrhizal tree species such as birch and Douglas fir 820 [143] and nutrient exchanges between endomycorrhizal plants [71] have been shown. We 821 thus define a CMN formed by ecto- or endo-mycorrhizal fungi that link plants together. 822 Transfer of different molecules may be bidirectional between plants and CMN can improve 823 interplant nutrition and growth through plant-plant facilitation. CMNs can also induce plant 824 defense responses and plant communication through phytohormones (e.g. jasmonic acid, 825 methyl jasmonate, zeatin riboside) and therefore have a strong potential as a biocontrol 826 agent. The role of arbuscular mycorrhizal fungi as potential bridge between different plants 827 in the spread of a disease has recently been hypothesized, for example with Potato virus Y or 828 following a necrotrophic fungus attack [144]. Indeed, in addition to the competitive role of 829 arbuscular mycorrhizal fungi against pathogens by allelopathy, CMN can act as a conduit 830 carrying chemical signaling compounds emitted by the attacked plant. These links make it 831 possible for a plant to send warning signals during an insect or pathogenic attack to the 832 network of plants connected by the CMN 'informing them' of the attack. In this context, an 833 interesting perspective would be to study how this CMN would transfer the information of a 834 pathogen attack for a 'sentinel' plant to a neighboring plant, which could preventively 835 activate their defense mechanisms and enhance their immunity.

836

837 Figure legends

838 Figure 1. The woody plant holobiont.

839 It is essential to consider the woody plant (WP) as a "holobiont", *e.g.* the tree and its 840 microbial continuum from soil to leaves. The microbiota resides on (epiphytic microbiota) or

within (endophytic microbiota in the phyllosphere, the lignosphere and the rhizosphere) any
of a number of WP tissues. Interactions exist between the WP and its associated microbes,
and among microbes. These microbes can be beneficial (*e.g.* better fitness, nutrition,
immune defense), pathogenic (*e.g.* diseases) or neutral for the WP.

845

846 Figure 2. Woody plant declines: a role of the microbiota

847 The woody plant (WP)-associated microbiota is made up of bacteria, fungi, viruses, protists, 848 archae and oomycetes. The microbiota could either enrich or weaken some physiological 849 functions of the WP. We hypothesize that the taxonomic composition and functions of the 850 WP-associated microbiota can influence the WP physiological status. The plant stays healthy 851 with a given balance of microbiota. On the contrary, an imbalance of microbiota (as cause or 852 consequence of the decline), leads to visible symptoms, whose origin can be physiological, 853 cultural and pathological. Various options may be used to cure some WP declines: a direct 854 inoculation of specific microbial species (*e.g.* involved in phosphorus mobilization) in order 855 to restore the balance, or by providing specific microbial species with targeted nutrients to 856 promote their growth.

In this figure, pie charts only reflect the distribution of the microbiota and the microbiome.
We have chosen some genes, not an exhaustive list, encoding proteins involved in functions
of WP metabolism. Similarly, a balanced microbiota may correspond to different proportions
of microorganisms belonging to each taxon. Taxonomic diversity does not necessarily mirror
the functional diversity.

862

863

Figure 3. Tag cloud of physiological and ecological diseases related to woody plant declines Woody plant (WP) declines have multi-factorial determinants (*e.g.* cultural practices, parasites, abiotic stresses) and their importance is not clearly established, and may depend on the WP species. This tag cloud (built using https://nuagedemots.co/) depicts factors and keywords commonly mentioned on WP declines in references used in this review, without any causal relationships. In WP declines, it would appear that microbiota could be a reliable indicator.

- 871
- 872 Glossary

873

Allelopathy: the chemical (excretory products, metabolites) inhibition or regulation of oneorganism by another.

876

877 **Common mycorrhizal network**: A continuum network formed by both endo- and 878 ectomycorrhizal fungi connecting plants with each other *via* their mycelium. Common 879 mycorrhizal network allows exchanges of nutrients and other chemical substances between 880 connected plants.

881

882 Core microbiota: All microbial species (bacteria, fungi, viruses, protists) which predominate
 883 at surface or inside a living organism (here plants). Plant core microbiota could be defined by
 884 a set of microbial species always found in plant of different genus.

885

886 Core microbiome: Set of essential functions provided by the microbiota of a plant found887 from one plant to another.

888

Decline: Gradual reduction of growth, vigor and productivity in a woody plant due to multiple abiotic (water or thermic stresses...) and biotic stresses (pathogens, insects...) and likely to an unbalanced microbiota.

892

893 **Dieback**: Progressive death of twigs and branches which generally starts at the tips.

894

Endophyte: Microorganism (mostly bacteria or fungi) that lives inside tissues of other organisms, in a symbiotic way, with beneficial effects or at least with no negative consequences for the host fitness. Microorganisms involved can therefore be commensal or beneficial, but they can also be saprobes or latent pathogens.

899

900 **Endosphere**: Internal regions of plant tissues that can be colonized by microorganisms.

901

902 Epiphyte: Microorganism (mostly bacteria or fungi) that lives on the surface of a given903 organism.

905 Facilitation: A positive interaction that can contribute to assembling ecological communities906 and preserving global biodiversity.

907

Holobiont: Sum of the different species that forms an ecological unit. Here, we limit ourdefinition to the plant and all its symbiotic microbiota.

910

911 **Hologenome**: The host and microbiome genomes. The host genome is highly conserved with 912 slow genetic changes. The microbiome genome is dynamic (*e.g.* horizontal gene transfer, 913 mutation) and can change rapidly by modifying microbial populations in response to 914 environmental changes.

915

916 Homeostasis: The maintenance of a relatively steady state or equilibrium in a biological917 system by intrinsic regulatory mechanisms.

918

919 Key or hub species: A small number of taxa that are strongly interconnected and having a920 severe effect on communities.

921

922 Lignosphere: Botanically, it is the micro-ecosystem of the trunk and branches surface,923 considered as a possible habitat for microbes.

924

925 Microbiome: Set of genes brought by microbes forming the microbiota within a holobiont.926

927 Microbiota: Community of microbes associated with the plant (bacteria, archaea, fungi,
928 viruses, protists and other microeukaryota).

929

Mycorrhiza: From *mukes*, fungus; and *rhiza*, root. Intimate association between roots from
90% of land plants and specialized soil fungi. At least seven types of mycorrhiza exist, but
ectomycorrhiza and arbuscular mycorrhiza are the most common.

933

934 Phyllosphere: Botanically, it is the micro-ecosystem of the leaf surface, considered as a935 possible habitat for microbes.

936

- 937 **Rhizosphere**: Volume of soil surrounding living plant roots that is influenced by root activity
- 938 and hosting specific rhizospheric microbes.
- 939
- 940 Rhizodeposition: The release of materials from plant roots into the rhizosphere, including
- 941 soluble and insoluble exudates, lysates and gases.
- 942
- 943 Symbiont: Organism establishing a close and long-term interaction with its host (here the944 plant).





Woody plant declines: A role of the microbiota ?

