

Rewetting of soil: Revisiting the origin of soil CO2 emissions

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20 Abstract

21

Rewetting dry soils is associated with a burst of microbial activity and mineralization, 22 23 which manifests itself as a pulse in soil CO₂ emissions, long-known as the Birch effect. In 24 arid and semi-arid systems, soil CO₂ losses upon rewetting at the end of extended dry 25 periods can contribute a significant fraction to the overall carbon (C) budget. Microbial 26 biomass is one of the sources of mineralized C, as was demonstrated over 30 years ago 27 (Kieft et al., 1987). The present paper offers a perspective on how the field has 28 progressed since the 1987 paper was published in *Soil Biology & Biochemistry*, what it 29 means in terms of current concerns about global climate change, and the needs and 30 potential emphases of future research. Many studies since 1987 have addressed the 31 origin of this CO₂ pulse, finding multiple possible C sources involving both biotic and 32 abiotic processes. We propose that the magnitude of the rewetting event ($\Delta \psi$) 33 determines the relative contribution from the array of substrates that contribute to the 34 soil CO₂ pulse upon rewetting. The magnitude of the CO₂ pulse is likely related to soil 35 physical characteristics and to the size of the available C pool, which is partly controlled 36 by plants. Further, the relative contributions of the mechanisms generating soil CO₂ 37 pulses upon rewetting are likely to be modified by climate change. To understand and 38 predict the magnitude of soil CO₂ pulses upon rewetting, we advocate continued cross-39 disciplinary research involving soil microbial ecology, soil physics, soil chemistry, as 40 well as increased integration and recognition of the importance of plant-soil interactions 41 in controlling the soil C pools contributing to soil CO₂ pulses.

42 Introduction

43

The ecology of soil rewetting is especially relevant to drylands, which include arid, 44 45 semi-arid and dry sub-humid subtypes. Drylands represent 45% of terrestrial surface (i.e. 46 66.7x10⁶ km²) and carry 39% of the world population (Prăvălie, 2016; Huang et al., 2017; 47 Plaza et al., 2018). Soil wet-up is particularly important for terrestrial ecosystems in Mediterranean and savanna climates that experience sustained periods of very low or no 48 49 rainfall, followed by significant rainfall. Mediterranean climates (dry summers and wet 50 winters) occur between 30° and 45° latitude and are found in Africa, Europe, western 51 South America, Southwest and West Australia, and western North America, for a total area 52 of ~3x10⁶ km². Savanna climates occur between and 25° and 30° latitude, and include 53 almost half the surface of Africa, large areas of Central & South America, eastern India, 54 southeast Asia, eastern and northern Australia, totaling $\sim 33 \times 10^6 \, \text{km}^2$.

55 Rewetting of dry soil can have global consequences. At the ecosystem scale it 56 results in a large soil CO₂ efflux pulse, also known as the Birch effect, named after H.F. 57 Birch who reported mineralization peaks upon rewetting soils in East Africa (Birch, 58 1958). However, the phenomenon had already been documented 35 years earlier by soil 59 microbiologists (Waksman and Starkey, 1923; Winogradsky, 1924). Its putative 60 mechanisms at the soil microbial scale started gaining attention in the 1980s (Bottner, 61 1985). The sudden increase in microbial activity upon rewetting triggers a burst in carbon 62 (C) and nitrogen (N) mineralization (Miller et al., 2005; Xiang et al., 2008; Borken and 63 Matzner, 2009; Dijkstra et al., 2012). The amount of C emitted due to rain pulses is comparable in magnitude to the net annual CO₂ exchange of many terrestrial ecosystems 64 (Xu et al., 2004). In ecosystems that are characterized by distinct dry-wet seasons, the CO₂ 65 66 pulses associated with rewetting can represent a large part of the annual C budget (Jarvis et al., 2007; Ma et al., 2012; Rey et al., 2017). For example, following intense rainfall, up
to 20 % of the annual CO₂ flux from soil to atmosphere occurred in an African savanna
ecosystem (Fan et al., 2015). While a rapid change in soil water potential results in an
overall microbial reactivation, it exposes soil microorganisms to a radical change in
environmental conditions. The soil dry-down and subsequent rewetting provide such an
extreme change in microbial environment, that bacterial and fungal death commonly
results (Blazewicz et al., 2014; Blazewicz et al., 2020).

74 The 1987 study by Kieft et al. was designed to investigate the fate of soil microbial 75 biomass C after the rapid rewetting of a dry soil. Using two Californian soils from a 76 Mediterranean-type climate, Kieft and his colleagues found a net loss of microbial biomass 77 C upon rewetting, that could be due to the death of microbial cells or the rapid 78 decomposition of the microbial solutes; the authors favoured the latter explanation. 79 Mechanisms of C loss, the nature of the lost C, and changes in microbial community 80 composition or relation to biogeochemical functioning were not investigated. However, 81 by opening the microbial 'black box' in the general context of dry-wet cycles, this study 82 brought attention to the large influence that rapid increases in soil water content can 83 exert on microbial and terrestrial ecosystem functioning. Exploration of the processes 84 comprising the Birch effect has been a particularly active field of research lately, as global 85 change impacts precipitation cycles worldwide. In many regions, especially under 86 Mediterranean climates, more extreme climate conditions increase the contrast between 87 dry and wet conditions to which ecosystems are exposed. It has been more than three 88 decades since the Kieft et al. study was published; here we provide a perspective on the 89 current status of the field, its importance in the context of global change, and areas in need 90 of more research.

92 What is wet-up and where does it occur?

93 We use the term wet-up to describe a real or simulated precipitation input of liquid 94 water to dry soil causing a large and rapid change in soil water potential ($\Delta \psi$). The 95 magnitude of the change in water potential is a determinant of the size of the resulting 96 soil CO₂ efflux pulse (Lado-Montserrat et al., 2014). Here we suggest that the addition of 97 liquid water to a very dry soil causes CO₂ pulses that differ not only in rate/magnitude but 98 also in available carbon sources and impact on microbial physiology and survival 99 depending on the magnitude of the water potential increase. Since soil wet-up includes a 100 prominent biological component, we focus on the biologically active zone of the soil, i.e. 101 between 0 and \sim 50 cm depth; this zone can experience rapid rewetting from very dry 102 conditions, depending on presence of macropores, surface cracking, and soil texture. While we will not concentrate on other inputs of moisture in this article, it is important to 103 104 note that in arid and semi-arid ecosystems, several other processes can introduce 105 moisture to dry surface soil. For example, surface litter decomposition occurs in response 106 to fog and dew events and even as a result of high relative humidity, particularly in 107 combination with UV-mediated degradation of exposed surface litter (Dirks et al., 2010; 108 Jacobson et al., 2015). Adsorption of water from atmospheric humidity has been 109 demonstrated to extend into soil (perhaps to a depth of 5 cm) over a range of atmospheric 110 relative humidity (20-60%) and to stimulate CO₂ production (McHugh et al., 2015). Thus, 111 transient episodes of decomposition at or near the soil surface occur in arid and semi-arid 112 ecosystems in response to non-precipitation events (fog, dew, sorption from atmospheric 113 humidity) and can drive CO₂ fluxes that are significant in ecosystem C budgets (Dirks et 114 al., 2010). Recognition and understanding of the importance of these alternative means of 115 increasing soil moisture (non-rainfall) has developed over the past 30 years and are 116 important advances in the subject.

We present a perspective that primarily addresses rewetting dynamics of soil near the soil surface and in the rooting zone of shallow-rooted plants. Under laboratory conditions, drying-rewetting cycles resulted in the release of "old" C from soil layers below 20 cm (Schimel et al., 2011). While macropore flow of incident rain water might result in the wet-up of soil deeper than 20 cm, rapid rewetting of soil by rainfall will be most important near the surface of soil and generally play a less important role in the mobilization of deep old soil carbon.

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125 C released upon rewetting: origin and mechanisms

126 The CO₂ pulse upon rewetting typically exhibits a pattern of very high rates during 127 the first minutes and hours after rewetting and this response declines over time (Kim et 128 al., 2012). The origin of the CO₂ pulse upon rewetting soil can be both abiotic and biotic. 129 Abiotic processes include solubilization of carbonates (depending on carbonate 130 concentration in soils), CO₂ displacement from the soil pores to the atmosphere by water, 131 as well as the quantitatively low degassing of CO₂ dissolved in rain and soil degassing due 132 to the decrease in barometric pressure over time (Lee et al., 2004; Barnard et al., 2015). 133 However, in non-carbonate-rich soils, abiotic processes generate less CO₂ than do the 134 biotic processes, which are the main contributors of the soil CO₂ pulse upon rewetting. 135 The nature and sources of the C compounds that are mineralized upon rewetting a dry 136 soil has been extensively explored and somewhat fervently argued. Here we highlight the 137 most prevalent sources and mechanisms that fuel the C mineralization pulse: compatible 138 solutes; microbial necromass; changes in water film connectivity; sustained extracellular 139 enzyme activity during dry-down; C desorption from mineral surfaces; and disruption of 140 aggregates. Each of these is discussed below.

141 Compatible solutes, accumulated by microbial cells in response to drying, were 142 suggested by Kieft et al. (1987) as candidate compounds fueling the soil CO₂ efflux pulse; 143 this substrate pool was also addressed by later studies (Halverson et al., 2000; Fierer and 144 Schimel, 2003; Schimel et al., 2007). Soil microorganism accumulation of compatible 145 solutes within their membrane-bound cells reduces cell dehydration, but cells are 146 required to rapidly dispose of them upon rewetting, or face membrane rupture 147 (plasmoptysis). These processes will release solutes, such as proline, glutamine, glycine 148 betaine, and trehalose into the soil (Kempf and Bremer, 1998; Halverson et al., 2000; 149 Welsh, 2000; Warren, 2019) and these can be assimilated and mineralized rapidly by 150 other microorganisms. Some of the earlier studies focusing on the intracellular 151 accumulation of compatible solutes as soil gets drier, found no evidence for osmolytes as 152 a significant C-source (Boot et al., 2013; Kakumanu et al., 2013). More recent evidence 153 (Warren, 2014; Warren, 2016) for this mechanism however provides some support. 154 While some studies detected no increased amount of compatible solutes in the soil upon 155 rewetting (Williams and Xia, 2009; Warren, 2014), recent studies have reported an 156 accumulation of osmolytes in microbial communities during dry periods (Chowdhury et 157 al., 2019; Malik et al., 2019; Slessarev et al., 2020). The elusive nature of osmolytes may 158 be related to: (i) their low residence time in the soil, as they are produced and consumed 159 at measurable rates even in moist soils (Warren, 2019), albeit at lower rates than in dry 160 soils (Warren et al, unpubl. data); and (ii) their probable specificity to a microbial 161 community. Some microbes may not accumulate compatible solutes and instead undergo 162 passive water loss, especially under conditions of rapid drying (Potts, 1994). Possible 163 fates of these plasmolyzed cells are death from water loss, death upon rewetting, or survival of both desiccation and re-wetting. 164

166 Microbial death due to the rapid water potential increase (from very negative 167 values to zero or nearly zero) upon rewetting was also invoked by Kieft et al. (1987) as a 168 possible source of C substrates. Dead microbial cells can provide mineralizable substrates 169 upon rewetting a dry soil, arising from cells that died during the previous dry-down 170 period or as a direct result of the rewetting. Evidence for this mechanism has been 171 presented by measuring gross rates of bacterial death and growth upon rewetting 172 (Blazewicz et al., 2014; Blazewicz et al., 2020). High bacterial mortality was documented 173 to have occurred within the first three hours after a rewetting event. The authors noted 174 that some of the mortality is likely to have occurred during the dry-down process but the 175 microbial necromass remained largely intact until the wet-up event.

176 Bacteriophage predation may also contribute to cell death after rewetting. Phages 177 infect a large proportion of soil bacteria (Williamson et al., 2007) and can become lytic when soil conditions become favorable for the growth of their hosts (Williamson et al., 178 179 2017). Indeed, viral abundance in the soil has been shown to be correlated with soil water 180 content (Williamson et al., 2005), possibly due to the effects of soil water content on 181 bacterial numbers as well as on the adsorption of viruses in the soil (Zhao et al., 2008). 182 Viral diffusion/advection is also enhanced by soil wetting, increasing the chances of virus-183 host encounter and attachment. At this time, however, there is no direct evidence for wet-184 up-associated predation as a significant mechanism of cell death and labile C input. 185 Similarly, eukaryotic predation may contribute to microbial mortality upon soil wetting, 186 but, to our knowledge, this has not been demonstrated.

Upon rewetting a dry soil, microorganisms have to access and then metabolize organic substrates. Connectivity of soil water films is gradually lost as soils dry and this restricts diffusional access to substrate (Manzoni et al., 2012) but enhances bacterial coexistence and diversity (Carson et al., 2010) by limiting competitive exclusion (Wang

and Or, 2013; Šťovíček et al., 2017b). Conversely, rewetting a soil restores water film
connectivity (Smith et al., 2017), providing conditions for highly competitive and mobile
microorganisms to access substrates and to thrive, thereby generating a soil CO₂ efflux
pulse (see modelling study by Evans et al., 2016).

Another possible source of substrates during rewetting requires sustained extracellular enzyme activity during dry-down (Miller et al., 2005; Schimel et al., 2017). The reduction in microbial activity as soils dry can lead to a net increase in substrate that may become available to soil microorganisms upon rewetting (Lawrence et al., 2009; Zhang et al., 2014; Fraser et al., 2016). However, a recent study found no evidence for sustained exoenzyme activity in drying soils (Homyak et al., 2018).

Soluble organic compounds in the soil can associate with minerals, particularly during dry periods (Qualls, 2000). Upon rewetting, the desorption of organic compounds from mineral surfaces can become a source of C (Blankinship and Schimel, 2018) as changes in pH and ionic strength can directly affect organic matter-mineral binding (Clarke et al., 2011; Newcomb et al., 2017).

In dryland ecosystems, photodegradation of surface litter increases the breakdown of carbon compounds to substrates that can be metabolized by microorganisms upon rewetting (Austin and Vivanco, 2006; Baker and Allison, 2015; Day et al., 2018). Although photodegradation alone cannot explain differences in decomposition rates across an aridity gradient (Brandt et al., 2010), it enhances soil CO₂ emissions upon soil rewetting (Ma et al., 2012).

Disruption of organic matter occlusion in soil aggregates is another physical consequence of rewetting that can enhance substrate availability. Rewetting a dry soil can break up aggregates, exposing organic substrates that were previously occluded (Denef

et al., 2001a; Cosentino et al., 2006; Borken and Matzner, 2009; Navarro-García et al.,
2012).

217 Modeling has provided solid insight into the mechanisms that drive the soil CO₂ 218 pulse associated with rewetting. Most modeling studies have focused on the sources of 219 soil organic matter at a large scale (Lawrence et al., 2009) and were not designed to take 220 into account the biophysical mechanisms at the pore-scale. The gap between pore physics, 221 soil organic matter cycling and microbiology has been bridged, using empirical models 222 and simplified model systems (Moyano et al., 2013; Manzoni et al., 2016; Tecon and Or, 223 2017), yet challenges remain (Baveye et al., 2018). Recent modeling efforts support the 224 coexistence of physical as well as microbiological processes as drivers of the rewetting 225 soil CO₂ pulse (Zhang et al., 2014; Evans et al., 2016; Šťovíček et al., 2017b; Yan et al., 226 2018).

The question of which C sources fuels the Birch effect has been under discussion for many decades. It is an important topic in the context of global change, as rainfall patterns and amounts change in areas around the globe, potentially impacting the C budget of ecosystems (Kim et al., 2012; Vargas et al., 2012).

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232 Soil microbial response

Ascribing the CO₂ released upon rewetting to a particular source of C can be akin to the Indian parable of six blind men examining an elephant, each of them failing to consider the multiple facets of the objective truth while experiencing it from their own limited perspective. Many C sources contribute to the soil CO₂ pulse upon rewetting (Unger et al., 2010; Evans et al., 2016). Apart from carbonate solubilization, CO₂ emissions from the soil upon rewetting are driven by coupled biophysical or biochemical

mechanisms, in which soil microorganisms act as catalysts, conditional upon whether ornot they have access to substrate.

241 In arid and semi-arid systems, adaptation to cyclic dry-down and rewetting 242 enables soil functions to be carried out under alternating contrasting environmental 243 conditions. Soil microorganisms seem to have evolved life strategies related to prolonged 244 dry periods followed by rapid rewetting (Placella et al., 2012; Barnard et al., 2013; Evans 245 and Wallenstein, 2014). At the seasonal scale, microbial strategies during dry-down differ 246 from strategies during rewetting (Evans et al., 2014). Soil microorganisms that are able 247 to sustain activity longer as soils dry down may access easily degradable organic matter 248 under less intense competitive conditions, while the other microorganisms that are not 249 able to maintain activity under dry conditions have already entered a state of reduced 250 activity, or even dormancy (including formation of spores). This mechanism could 251 support the existence of taxa (e.g. bacteria belonging to the Sphingomonadaceae, 252 Comamonadaceae and Oxalobacteraceae; Aanderud et al., 2015) that are present in low 253 relative abundance in the soil (Lennon and Jones, 2011; Meisner et al., 2018). The 254 duration of the dry period can affect the response to subsequent rewetting (Barnard et 255 al., 2015), especially in microbial communities that have not evolved in contrasting dry-256 wet conditions (Meisner et al., 2015; Meisner et al., 2017). For example, in soils where 257 microbes are not adapted to drought or cyclic dry periods, a lag period before exponential 258 microbial growth has been detected, the duration of which depended on the intensity of 259 drying (Meisner et al., 2017). Indeed, altered precipitation patterns before rewetting 260 impacted not only the community structure of the microorganisms that were active upon 261 rewetting, but also of those that were not, as they were recruited into the active bacterial 262 pool (Engelhardt et al., 2019).

263 The importance of microbial death as a source of C for the CO₂ pulse has been 264 measured using quantitative ¹⁸O stable isotope probing. Upon rewetting a soil from a 265 Mediterranean climate, initial bacterial growth was detected in select taxa dominated by 266 Proteobacteria (Azotobacter, Pseudomonas, Vibrio) and Firmicutes (Streptococcus, 267 *Clostridium, Mycoplasma*) while mortality was taxonomically widespread (Blazewicz et 268 al., 2020). In addition, the density-dependence of growth upon rewetting contrasted with 269 density independence of mortality, supporting the hypothesis that population-level 270 biological interactions, such as competition and predation, likely controlled the growth 271 rates of taxa but not their death (Blazewicz et al., 2020).

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273 A number of studies have shown differential sensitivity of soil bacterial taxa to dry-274 down and rewetting; this differential sensitivity is conserved at phylum and class 275 phylogenetic groupings (Lennon et al., 2012; Placella et al., 2012; Barnard et al., 2013, 276 2015; Engelhardt et al., 2018). This phylogenetic coherence reflects the evolutionary 277 roots of microbial tolerance to drying and rewetting in microorganisms that are 278 indigenous to and more common in semi-arid soil systems. The functional response of 279 arid and semi-arid systems to dry-wet cycles can thus be expected to be deterministic and 280 predictable (Averill et al., 2016). The two flagship phyla are Actinobacteria, which are 281 commonly abundant in arid soils and include many taxa that are resistant to low moisture 282 conditions (Goodfellow and Williams, 1983; Zvyagintsev et al., 2007) and Acidobacteria, 283 a versatile group that can be adapted to changing environments (Eichorst et al., 2018); 284 taxa from the Acidobacteria group have been found to dynamically track soil water 285 conditions, flourishing rapidly upon rewetting (Placella et al., 2012; Barnard et al., 2013). Dry-down typically increases the relative abundance of the present or potentially active 286 287 thick-walled, Gram-positive, high G+C content, spore-forming, and EPS-producing

288 bacterial groups (e.g. Actinobacteria, Firmicutes) but decreases that of thin-walled, Gram-289 negative or fast-growing groups such as Acidobacteria, Verrucomicrobia or Chloroflexi 290 (Placella et al., 2012; Angel and Conrad, 2013; Barnard et al., 2013; Maestre et al., 2015; 291 Zhou et al., 2016; Šťovíček et al., 2017a; Ochoa-Hueso et al., 2018). Upon rewetting, 292 different bacterial groups respond on a variable timeframe that likely reflects their ability 293 to have formed spores during the dry period, thus being potentially ready for a quick 294 response and rapid growth (Keijser et al., 2007; Placella et al., 2012; Barnard et al., 2013; 295 Sinai et al., 2015). Taxon-specific growth documented by quantitative stable isotope 296 probing has shown that members of the Bacillales order of Firmicutes grew within 3h of 297 rewetting a dry Mediterranean soil, followed in the first 24h by β -proteobacteria of the 298 Burkholderia order (Blazewicz et al., 2020). Fungi generally display an overall greater 299 resistance to desiccation-hydration cycles than bacteria (Bapiri et al., 2010; Curiel Yuste 300 et al., 2011; Barnard et al., 2015), likely due to their hyphal network structure and 301 frequent mutualistic strategies (de Boer et al., 2005), which complement other drought-302 resistance traits that can also be found in bacteria, such as production of extracellular 303 polymeric substances (EPS) and heat-shock proteins (Crowther et al., 2014). As a 304 consequence, the fungal community displays a higher stability than the overall bacterial 305 community during dry-down and subsequent rewetting (de Vries et al., 2018).

In temperate climates, contrasting dry and wet conditions arise during a period of drought, i.e. water limitations outside the normal climatic envelope. Since the temperate soil biosphere may not be well-adapted to these extreme conditions, its response to drying and rewetting is expected to be more stochastic and less predictable than that of highly-adapted arid and semi-arid systems, although a trait-based phylogenetic framework could help predict microbial responses (Lennon et al., 2012; Amend et al., 2016). Drought legacy experiments have shown that in some systems having been

exposed to past drought conditions can shape the response of soil microbial communities
to subsequent drought or rewetting (Bouskill et al., 2013; Göransson et al., 2013; Hawkes
et al., 2017; Meisner et al., 2018), with consequences that can extend to soil processes
(Evans and Wallenstein, 2012; Martiny et al., 2017) as well as plant communities (Lau and
Lennon, 2012; Meisner et al., 2013; Kaisermann et al., 2017).

318 The intensity and duration of dry conditions determine the response of soils to 319 rewetting (Zhang et al., 2019). After a severe dry period, soil rewetting triggers a larger 320 CO₂ pulse (Xu et al., 2004; Cable et al., 2008; Göransson et al., 2013; Barnard et al., 2015), 321 which is consistent with a greater loss of C in carbon-rich ecosystems that experience 322 more pronounced dry-wet events (Canarini et al., 2017). Under extreme conditions (i.e. 323 dry soil for a long period of time or large $\Delta \psi$), maintaining metabolic functions upon 324 rewetting may become more challenging than under more favorable conditions and there 325 may be a greater frequency of microbial death and subsequent larger relative 326 contribution of compatible solutes to the soil CO₂ pulse. However, the speed of dry-down 327 should also be considered. When dry-down is slow, more compatible solutes accumulate 328 in the soil relative to depolymerization products generated by exoenzymes (Warren, 329 2016). In addition, microbial production of extracellular polymeric substances (EPS), 330 such as exopolysaccharides, can both retain water during soil drying and slow down 331 rewetting of the habitats of microbial cells. (Roberson and Firestone, 1992).

Maximum compatible solute concentration in soil microbes occurred at relatively high ψ (-5 MPa) under slow dry-down conditions (Warren, 2016), compared to the estimated -14 MPa at which microbial activity ceases in the soil (Manzoni et al., 2012). Thus, compatible solute accumulation can only protect indigenous soil organisms from dry down events of limited magnitude.

337 The physical effects of rewetting on soil aggregates are expected to become 338 relevant once the point at which large soil aggregates become disrupted is reached (Lado-339 Montserrat et al., 2014). Rewetting aggregates with a large $\Delta \psi$ would enable access to C 340 pools that were previously inaccessible, i.e. occluded organic matter. How big an upshock, 341 i.e. the size of $\Delta \psi$, is required to access occluded organic matter? As water travels down 342 the soil profile, the vapor phase front likely influences microbial cell metabolism, but 343 liquid water is required to disrupt soil aggregates. Does the disruption threshold depend 344 on the size of $\Delta \psi$ or is it an absolute threshold of ψ (i.e. soil water potential falling below 345 the threshold during dry-down and rising above it upon rewetting)? While the dryness 346 reached by the soil can play a role, the size of $\Delta \psi$ seems to be the most relevant parameter 347 (Xu et al., 2004; Chowdhury et al., 2011; Lado-Montserrat et al., 2014). Estimating this 348 parameter could provide a metric of sensitivity of ecosystems to intense dry periods. Soil 349 CO₂ emission rates before rewetting are significantly negatively correlated with the post-350 rewetting rates (Kim et al., 2012) and can be predicted from rewetting-related changes in 351 soil moisture (Rey et al., 2017). Thus, the soil water content within the preceding years 352 could possibly be used as a proxy to estimate the relative contribution of rewetting events 353 to the overall C budget in systems that experience seasonally dry climatic conditions. 354 Moreover, the speed of soil rewetting frames the impact of the event; this would depend 355 on soil physico-chemical characteristics, among which texture (including preferential 356 flow paths) and soil salinity are important (Cable et al., 2008; Chowdhury et al., 2011) as 357 well as EPS content which can influence soil repellency as well as speed of rewetting 358 (Seaton et al., 2019).

We propose that: (i) all the previously discussed substrates potentially contribute
to soil CO₂ pulse upon rewetting, but that their relative contributions depend on climate,

361 soil type and magnitude of water potential upshock $\Delta \psi$; and (ii) exceeding a threshold in 362 dry-wet $\Delta \psi$ is required to mobilize occluded organic matter (Fig. 1).

363

364 Importance of plants

365 Plants are the primary contributors to the quantity and quality of C entering soil 366 and the size and composition of the available C pool controls the magnitude of the CO₂ 367 pulse. Many of the studies investigating the nature of the C fueling the CO₂ pulse upon 368 rewetting were undertaken in systems that were devoid of live plants at the time of 369 rewetting (Borken and Matzner, 2009). This reflects the fact that wet-up studies, and the 370 $\Delta \psi$ associated with substantial CO₂ pulses, require extremely dry soils, in which annual 371 plants are generally no longer alive. However, the plants that were present during a 372 growing period immediately preceding the dry-down may affect the microbial response 373 to rewetting as: (i) labile C pools support CO₂ pulses ; (ii) above- and below-ground litter 374 from semi-arid ecosystems can generate a large and labile pool of soil organic C that is 375 present during the dry season; and (iii) the presence of plants can impact the number and 376 size of soil aggregates (Blankinship et al., 2016). Thus, plants may have been absent in 377 many studies due either to the soil-centric focus of the researchers and the question, or 378 due to the absence of living plants at the time of rewetting. For example, in Mediterranean 379 systems the annual plant cover is often dead by the end of the dry season, when the first 380 rains constitute the largest soil water potential upshocks.

Nevertheless, plants matter, whether dead or alive. Dry episodes can take place during the growing season without killing plant cover. In systems in which live plants are present, dry-wet cycling directly impacts plant-derived soil labile C inputs, both in their quantity, generally increasing exudation under moderate drought but having a more variable effect under intense drought (Preece and Peñuelas, 2016), and quality (Preece

386 and Peñuelas, 2016; Gargallo-Garriga et al., 2018), with consequences on ecosystem 387 functioning (Oikawa et al., 2014; Canarini and Dijkstra, 2015; Williams and de Vries, 388 2019). Dry-wet cycles can also affect plant N uptake, hydraulic redistribution (Fu et al., 389 2018) and trapping of windblown soil particles (Ravi et al., 2010), which all likely interact 390 with major processes involved in the soil CO₂ pulse upon rewetting (Wang et al., 2015). 391 Root exudation is of particular relevance, as it not only provides a direct labile C source 392 for soil microorganisms, but can also contribute to shaping the root microbiome (Naylor 393 and Coleman-Derr, 2017; Hartman and Tringe, 2019), a major driver of the soil CO₂ pulse 394 upon rewetting.

395 Dry soils as well as those subject to infrequent watering events can hamper plant-396 microbial coupling by impeding the transfer of recent photo-assimilates from the roots to 397 soil microorganisms (Ruehr et al., 2009; Fuchslueger et al., 2014; Engelhardt et al., 2018; 398 Karlowsky et al., 2018). This may be due to reduced rhizodeposition rates (but see Preece 399 and Peñuelas, 2016) or to challenging microbial access to rhizodeposits under diminished 400 water film connectedness (Moyano et al., 2013; Fuchslueger et al., 2014; Tecon and Or, 401 2016; von Rein et al., 2016; Karlowsky et al., 2018). The plant-microbial interaction 402 component of the mechanisms underlying the soil CO₂ efflux pulse upon rewetting may 403 also vary with depth (Engelhardt et al., 2018).

Few experiments have addressed rewetting events during the spring in Mediterranean systems, when the plants are still alive, actively growing and the high flux of rhizodeposits to the soil exerts a strong influence on soil microbial activity (Curiel Yuste et al., 2007; Ma et al., 2012). The seasonal component of microbial response to changes in water availability has been documented in several studies (Cruz-Martínez et al., 2009; Bell et al., 2014; Zhao et al., 2016). The plant-microbial interaction component of the soil CO₂

pulse after rewetting may vary seasonally and experiments exploring rewetting after dryspring periods could provide a valuable perspective.

We propose that the origin of the C released upon soil rewetting (Fig. 2) is mainly controlled by three factors: the magnitude of the rewetting upshock ($\Delta \psi$), soil physical characteristics, and ecosystem productivity (NEP). The $\Delta \psi$ controls the pool of microbialderived C, i.e. compatible solutes and dead cells. Soil physics controls pore connectivity, which in turn controls access to microbial- and plant-derived C. NEP controls the pool of plant-derived labile C which determines much of the soil enzyme pool.

418 Dry-wet cycles can impact soil C stabilization both positively and negatively 419 (Bailey et al., 2019). A few dry-wet cycles increase the turnover of aggregates (i.e. as they 420 are broken down then formed again), releasing inter-aggregate particulate organic matter 421 and decreasing stabilization of fresh organic matter within the aggregate structure (Denef 422 et al., 2001b). However, aggregates become slake-resistant with time, and more dry-wet 423 cycles result in increased microaggregate stability (Denef et al., 2001a; Rahman et al., 424 2018). Dry-wet cycles increase microbial biomass C turnover, as microbial C lost through 425 microbial death is replaced. Recent studies have proposed that enhanced microbial 426 turnover under dry-wet cycles could result in an increased contribution of previously 427 stable organic matter to microbial C (Lopez-Sangil et al., 2018; Schimel, 2018).

428

429 **Conclusion**

The large CO₂ pulse that is produced by rapid rewetting of very dry soil results
from biological, physical and chemical mechanisms. The past 30+ years of research in this
area provides an excellent example of integration across the subdisciplines of soil science.
We suggest that significant future advances on the topic will be facilitated by greater
inclusion and recognition of plant-microbe soil interactions.

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436

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851 Figure legends

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853 Figure 1. Proposed mechanisms fueling soil CO₂ efflux pulse upon rewetting.

- Figure 2. Main drivers (gray) of the mechanisms releasing CO₂ upon rewetting a dry soil,
- 856 involving interactions between soil microorganisms (blue, center), plants (green, left) and
- 857 soil physico-chemical properties (red, right).

858 Figure 1



