

Effects of the earthworm Pontoscolex corethrurus on rice P nutrition and plant-available soil P in a tropical Ferralsol

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Jean Trap, Eric Blanchart, Onja Ratsiatosika, Malalatiana Razafindrakoto, Thierry Becquer, et al.. Effects of the earthworm Pontoscolex corethrurus on rice P nutrition and plant-available soil P in a tropical Ferralsol. Applied Soil Ecology, 2021, 160, pp.103867. 10.1016/j.apsoil.2020.103867. hal-03152409

HAL Id: hal-03152409 https://hal.inrae.fr/hal-03152409

Submitted on 2 Jan 2023

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1	Effects of the earthworm <i>Pontoscolex corethrurus</i> on rice P nutrition and plant-available soil
2	P in a tropical Ferralsol
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Abstract

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Madagascar

20 We conducted a greenhouse experiment in mesocosms for 28 d to assess the effects of the endogeic earthworm Pontoscolex corethrurus on plant-available soil phosphorus (P) and rice 22 (Oryza sativa) P nutrition in a Malagasy Ferralsol. To assess plant-available soil P, we 23 determined the *L-value* by measuring the specific activity of P taken up from the soil by rice 24 and by applying the isotopic dilution principle. Despite earthworm mortality, *P. corethrurus* significantly increased rice shoot biomass (+26%) and P nutrition (+65%), confirming that the 25 26 soil used for the experiment was P-deficient. The L-value also markedly increased from $6.8 \pm$ 0.9 to 14.2 ± 1.3 mg P mesocosm⁻¹ in the presence of P. corethrurus. We estimated that the 27 28 orthophosphate ions released due to earthworm mortality contributed to 30% of the *L-value* 29 increase. We attributed the remaining 70% increase to the solubilization of native soil P 30 during its transit through the digestive tract. Thereafter, we discussed the sources of 31 uncertainty associated with the *L-value* calculation and their utilization to assess earthworm 32 effects on P availability in further studies. 33 Keywords: endogeic earthworms, ³²P-labeling, isotopic dilution, *L-value*, upland rice, 34

1. Introduction

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38 Earthworms are involved in key soil functions (Lavelle et al., 2007; Liu et al., 2019). They 39 enhance soil nutrient availability for plants (Chapuis-Lardy et al., 2011; van Groenigen et al., 40 2019). These terrestrial invertebrates can ingest considerable amounts of soil, for example, a community of earthworms in Lamto savannas was reported to have consumed 1250 t ha⁻¹ yr⁻¹ 41 42 (Lavelle et al., 1992). Endogeic earthworms can daily ingest soil quantity several times their own weight, in the range of 1-10 g of dry soil g⁻¹ fresh weight day⁻¹, and specifically for 43 Pontoscolex corethrurus, a value between 1.5 and 3.15 g g⁻¹ d⁻¹ has been reported (Barois et 44 al., 1999). This can result in high nitrogen (N) mineralization, approximately 30-50 kg N ha⁻¹ 45 yr⁻¹ (Lavelle et al., 2004), and considerably contribute to annual plant N uptake (Blouin et al., 46 47 2013). Despite increasing evidence and understanding, little is known about the distinct processes 48 49 involved in the effects of earthworms on phosphorus (P) flows (Le Bayon and Milleret, 2009; 50 Ros et al., 2017). Earthworms may contribute to diffusible inorganic P production by 51 enhancing microbial activity and organic P mineralization (e.g., phosphatase activity), and/or 52 by changing the dynamics of orthophosphate ions via desorption in stable bio-structures 53 produced by earthworms due to shifts in soil pH during the gut transit (Vos et al., 2019), or possibly due to competitive adsorption of elevated dissolved organic matter (Ros et al., 2017). 54 55 The effects of earthworms on P availability have, however, rarely been explored in the rice-56 Ferralsol system. There is a dearth of research on this aspect and this is a serious concern, 57 considering that P is the main nutrient limiting plant production in acidic soils in tropical 58 regions (Hou et al., 2020; Raminoarison et al., 2020). 59 The *L-value* technique was proposed by Larsen (1952) to assess the fraction of soil P that was 60 plant-available. Its principle relies on the homogeneous labeling of orthophosphate ions in the 61 solid phase of the soil, that can later dilute orthophosphate ions in solution. The tracking is

performed by adding and by thorough mixing of a solution of carrier-free ³²P-ions (or ³³P-62 63 ions) to the soil. The plant is used to sample the specific activity, i.e., ratio of radioactive P to unlabeled P, of the P taken up by an individual plant after a defined period of growth. 64 65 Assuming no isotopic discrimination between labeled and unlabeled orthophosphate ions, the L-value is calculated by applying the isotopic dilution principle. The L-value represents the 66 67 amount of plant-available exchangeable P. Thus, it is possible to quantify the effect of a particular treatment on the fate of P from a particular soil source to the plant. The *L-value* 68 69 technique has been extensively used to assess and compare the amount of plant-available P in 70 soils (Bühler et al., 2003; Frossard et al., 1994; Pypers et al., 2006; Russell et al., 1957) and 71 the ability of plants to access different pools of soil P (Hedley et al., 1982; Hinsinger and 72 Gilkes, 1997). However, it has not been used thus far to assess the effects of earthworm 73 presence on plant-available soil P. 74 We conducted a greenhouse experiment to assess the effects of the endogeic earthworm P. 75 corethrurus (Rhinodrilidae) inoculation on rice (Oryza sativa) nutrition and the L-value in a 76 native Ferralsol of the Malagasy Highlands. We hypothesized that the earthworm-induced 77 solubilization of soil P during transit through the digestive tract enhanced rice P nutrition and 78 the *L-value*. 79 80 2. Materials and Methods 81 2.1. Soil sampling and preparation 82 We sampled the first 0-10-cm layer of a Ferralsol under natural grassland from the experimental station of the "Laboratoire des Radio-Isotopes" (18°46'55" S, 47°32'46" E, 83

1274 m asl, Madagascar) in October 2016. The soil had a sandy-clay texture, with pH 5.5,

total organic C content of 20.8 g dry kg⁻¹, C:N ratio of 16, total P content of 380 mg dry kg⁻¹,

and Olsen P content of 4.7 mg dry kg⁻¹. Using the procedure described by Randriamanantsoa

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et al. (2013) for P-deficient soil, the concentration of orthophosphate ions in solution was determined as 0.004 mg P L⁻¹.

The soil was air-dried to a constant weight (7 d), sieved to obtain 2 mm particle size, rehumidified with sterile water to 50% of its water-holding capacity, and finally autoclaved for 10 min at 100°C to kill earthworms (adults, juveniles, and cocoons). The autoclaved soil was air-dried to a constant weight (7 d) and sieved again at 2 mm. A microbial suspension was prepared from fresh soil using a two-step protocol (see Ranoarisoa et al. (2018) for details on soil preparation). Briefly, we provided 28.2 mg-N and 3.4 mg-P per kg dry soil as bacterial biomass N and P, and 28.2 mg-N as NH₄Cl and 3.4 mg-P as KH₂PO₄ per kg dry soil as a basal mineral fertilization.

2.2. Biological materials

We focused on the endogeic earthworm *P. corethrurus* (Ponto), which is highly abundant in Madagascar, and the main species in cropped soils in the highlands of Madagascar (Razafindrakoto et al., 2010). *P. corethrurus* belongs to the family Rhinodrilidae. It is a peregrine species found throughout the tropics. It is 5-8 cm in length, weighing 0.7 g on average at the adult stage. It is an endogeic geophagous species living in an exceptionally substantial range of soil and climate environments and easily adapts to man-made agrosystems. Individuals of *P. corethrurus* were collected using a spade from the experimental station and inoculated in the same fresh soil (not autoclaved) for 5 d. Before inoculation, earthworms were rinsed three times with distilled water. Twenty individuals were weighed after 48 h at 65°C. Dried tissues were finely and manually ground and total P in the *P. corethrurus* tissues was analyzed (3 replicates) after calcination at 550°C for 5 h and digestion with nitric acid (65%). The P concentration was determined in the digest solutions using the molybdenum blue method (Murphy and Riley, 1962).

2.3. Soil ³²P labeling

The soil was labeled with a carrier-free orthophosphate ³²P ion radionuclide source delivered in HCl 1M (NEX054010MC (323 TBq mmol⁻¹), PerkinElmer®, Belgium), and was diluted in water with an activity rate of approximately 0.1 mCi kg⁻¹ dry soil. The soil was mixed by hand for 15 min in a large plastic container, and 500 g of ³²P-labelled dry soil was introduced in 1-L plastic mesocosms. A basal nutrient solution containing N, Ca, K, Mg, S, Fe, Mn, Zn, Cu, Co, B, and Mo was added to achieve 70%-80% of soil water holding capacity. Eight seeds of rice (cultivar B22) were planted in each mesocosm and three rinsed adults of *P. corethrurus* were inoculated per +Ponto mesocosm (treatment with *P. corethrurus*, see below). Thereafter, 100 g of acid-washed sand was provided at the top of the soil to optimize germination conditions and to protect workers from radiation.

2.4. Experimental design

We conducted a mesocosm experiment in a greenhouse with two treatments, namely a control without earthworms (-Ponto) and a treatment containing *P. corethrurus* (+Ponto). To limit the handling of radioactive materials, ten mesocosms (five without earthworms and five with earthworms) without the addition of ³²P were used to perform analyses on fresh soil and plant samples. Furthermore, ten supplementary mesocosms (five without earthworms and five with earthworms) with the addition of ³²P were used to perform *L-value* analyses. All mesocosms (labeled or not labeled) were treated in the same way. The mesocosms were randomly placed in the greenhouse. During the period of rice culture, the soil moisture content was controlled twice a day to maintain constant conditions. The climatic conditions (mean T (°C) and mean PAR) within the greenhouse were monitored with a climatic station (WatchDog Series Weather Stations, USA) (Supporting Information 1).

2.5. Measurements from unlabeled mesocosms

After 28 d, whole rice plants were removed, and the associated roots, seed residues, and shoots of rice were separated, washed, dried at 65°C for 48 h, and weighed. Total P content in shoots, roots, and seeds was determined after crushing, calcination at 550°C for 5 h, and digestion of plant tissues with nitric acid (65%). P content in plant tissues was determined in the digest solutions using the molybdenum blue colorimetric method (Murphy and Riley, 1962). We analyzed the total Mg, K, and Ca contents in the digest solutions using atomic absorption spectroscopy (Thermo Scientific iCE 3000 SERIES, China). Total C and N contents in plants (shoots and roots) were determined using a CHNS/O elemental microanalyzer (Flash 2000 Series, CHNS/O 122 Analyzers Thermo Scientific, IRCOF, France). We collected the earthworms (individuals and cocoons) by manually sieving the fresh soil. We determined the survival rate of *P. corethrurus* by comparing the final earthworm abundance to the initial number. Soil aggregation was determined by sieving airdried soil at 2 mm, which corresponded to a mass proportion of soil greater than 2 mm (Blanchart, 1992). We used soil aggregation as a proxy for *P. corethrurus* activity.

154 2.6. Measurements from labeled mesocosms

After 28 d, only plant shoots were removed from the labeled mesocosms, dried at 65°C for 48 h, and weighed. The P content in shoots was determined as described above. The shoot digests were analyzed for ³²P radioactivity using a liquid scintillation analyzer (Tri-Carb 2800 TR, PerkinElmer) with a liquid scintillation cocktail (Ultima Gold XR). The ³²P activities, R (introduced radioactivity), and r (remaining radioactivity in harvested shoots), were corrected for radioactive decay. The specific activity of shoot P was calculated from the ratio between r/R and shoot P uptake. The *L-value* was calculated by applying the isotopic dilution

principle, that is, the specific activity of the *L-value* equalled the specific activity of P taken

up by a plant, as demonstrated in the follow equation:

- 164 $R/L = r / (P_{shoot} P_{seed}) \Rightarrow L = (P_{shoot} P_{seed}) / (r/R),$
- with r representing the ³²P-radioactivity in rice shoot at harvest, R representing the ³²P-

radioactivity of the initially applied to soils to each mesocosm, and P_{seed} representing the

seed-borne P in rice shoots. To correct the *L-value* for P_{seed}, we followed the procedure

proposed by Achat et al. (2014). Briefly, the fraction of P in rice shoots derived from seed P

mobilization (P_{seed}) was calculated by subtracting the initial amount of P in seed at seedling

and the remaining amount of P in seed residues found in the not-labeled soils (see above) after

28 d of cultivation. This amount was proportionally distributed between the shoot and root of

rice on a mass-basis. We did not handle fresh soil from the labeled mesocosms due to

173 radioactivity.

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- 2.7. Statistical analysis
- Mean and standard deviation (SD) values were computed per treatments for all variables. We
- performed Student's t-tests with earthworm treatments as a factor for all variables using the R
- software (R Core Team, 2016). Statistical significance was set at P < 0.05.

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3. Results and discussion

- 181 3.1. Earthworm survival rates and effect on soil aggregation
- Neither Ponto individuals nor cocoons, nor any decaying tissues were observed in unlabeled
- mesocosms at the end of the experiment. The last observation of fresh casts was conducted on
- the 19th day after the commencement of the experiment. We found casts in +Ponto soils and
- the soil aggregation rate increased from 3.5% in -Ponto to 15.5% in +Ponto treatments (Fig. 1.
- A). In this soil and for similar experimental conditions, Chapuis-Lardy et al. (2009) measured

1.5 g casts produced per day and per earthworm. Therefore, we determined the production of casts during the 19-day period of Ponto activities as 85.5 g corresponding to 17.7% of the soil weight. This result was in agreement with the percentage of aggregates measured.

3.2. Effects of Pontoscolex corethrurus on rice biomass and nutrition

Despite mortality, *P. corethrurus* significantly increased rice shoot biomass (+26%), total biomass (+15%), and rice P nutrition (+65%) (Table 1). Owing to the application of the basal nutrient solution, rice N, K, Ca, and Mg did not differ significantly across the -Ponto and +Ponto treatments (Table 1). However, a general trend was observed with a slight increase in N, K, Ca, and Mg uptake in the +Ponto treatment. These results were in agreement with those of the previously reported studies (Hauser, 1993). These effects are likely due to the release and decomposition of Ponto tissues at death, considering the content of N (1.8%), K (0.5%), Ca (0.2%), and Mg (0.2%) in earthworm tissues is important for such studies (Lourdumary and Uma, 2013).

3.3. Effects of Pontoscolex corethrurus on L-values

In our study, the P content in seed residuals at the end of the experiment was 7.9 (± 0.49) μ g P mesocosm⁻¹ in –Ponto and 13.1 (± 0.41) μ g P mesocosm⁻¹ in +Ponto (Table 1), which represented an average of 2.6% of the initial amount of P in seeds at seedling. After subtracting it from the P accumulation in rice shoots for 28 d of culture, *L-value*s were calculated considering only the P derived from plant-available soil P. The *L-value*s were 6.8 mg P mesocosm⁻¹ soil in –Ponto and 14.2 mg P mesocosm⁻¹ in +Ponto, which represented an increase of +109% when comparing the treatment to the control.

The decomposition of Ponto tissues after their death releases nutrients, especially orthophosphate ions. This process is usually recognized as negligible even if earthworm

212 survival in pot experiments can be below 100% (Bertrand et al., 2015; Chassé et al., 2019; 213 Paul et al., 2012; Vos et al., 2019), and on a few occasions, it can be below 60% (Derouard et 214 al., 1997; Fonte et al., 2019; Huang et al., 2015; Ratsiatosika et al., 2019). In our study, we considered the initial amount of P in +Ponto (2.21 mg P mesocosm⁻¹ soil), which was released 215 216 by the Ponto decomposition. This contributed to an increase in the *L-value* by $\sim 30\%$ 217 $(2.21\times100/(14.2-6.8))$ with a maximal value of 38.1% and the lowest value of 22.1% (Fig. 1. 218 B). The remaining 70% of the *L-value* increase could be attributed to the Ponto activities, 219 which partially solubilized soil P through the transit in their digestive tract. 220 The following two questions arise from these observations: 1) What are the mechanisms that 221 might explain the increase in the *L-value*?; and 2) What are the possible sources of 222 uncertainty that could lead to overestimation or underestimation of the *L-value*? 223 We could not clearly identify the main mechanisms that could lead to solubilization of native 224 soil P during its transit in the digestive tract. Little is known regarding the distinct processes 225 involved in their effects on P flows (Le Bayon and Milleret, 2009; Ros et al., 2017). P in 226 acidic soils, including soils containing orthophosphate ions, forms complexes with Al- and 227 Fe-oxyhydroxides; additionally, P precipitates as amorphous and crystalline Al and Fe 228 minerals and organic P compounds (Sample et al., 1980; Turner, 2006). According to this 229 variable speciation, several biological and physicochemical processes might transform native 230 P soil and modify the equilibrium between added orthophosphate ³²P ions that dilute native 231 ³¹P soil. Since about 31% of the total P in Malagasy soils is available in the organic form 232 (Turner, 2006), P. corethrurus may contribute to the release of orthophosphate ions in 233 solution that diffuse at the solid-to-solution interface by enhancing mineralization of soil 234 organic matter (Chapuis-Lardy et al., 2011; Ghosh et al., 1999; Le Bayon and Milleret, 2009). 235 For instance, Coulis et al. (2014) observed lower amounts of soil organic P extracted with 236 NaOH, but higher water-soluble inorganic P content when earthworms (Allobophora

chlorotica) were present in a durum wheat-chickpea intercropping glasshouse experiment. Additionally, dissolution of phosphate minerals in the native soil could occur in the digestive tract. However, this effect is likely minor because the internal pH is close to 7.0 (Barois and Lavelle, 1986). Transitions in the digestive tract may also affect soil-to-solution dynamics of orthophosphate ions by changing soil properties in stable bio-structures produced by earthworms due to shifts in soil pH (Vos et al., 2019) or due to competitive adsorption of elevated dissolved organic matter (Ros et al., 2017). The substantial increase (+109%) of the *L-value* associated with Ponto inoculation should be considered with considerable caution even after correction for P_{seed} because several sources of uncertainties can disturb its calculation and interpretation. In this study, especially, one source of bias could be the result of non-homogeneous soil-casts-(P. corethrurus after decomposition) mixture, since the mass of casts represented 85.5 g and the remaining soil in mesocosms amounted to 414.5 g. Both P. corethrurus casts and soil fertilized by the addition of P. corethrurus decaying tissues create small volumes favorable to growth of rice roots and absorption of orthophosphate ions. As a result, the specific activity of P taken up by rice may be preferentially absorbed from these small volumes having a lower specific activity than that in other soil volumes. These local effects do not validate the basic hypothesis to calculate the L-value, in which a homogeneous mixture is necessary. Consequently, it is likely that this leads to a significant overestimation of the *L-value*. The same drawback was encountered in studies on the capacity of different crop species to access poorly available soil P as assessed by the *L-value* technique (Hocking, 2001). The *L-value* was considerably larger with white lupin compared to *L-values* obtained with other crop species. This increase was explained by the ability of white lupin which developed specialized proteoid roots in P-deficient soil, which exuded, locally in their rhizosphere, considerable amounts of citrate or citric acid to mobilize soil P. This indicated that P uptake was substantial in proteoid root clusters than that in apical

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segments of non-proteoid roots (Neumann et al., 1999). Thus, although the isotopic tracing technique has shown to be an extremely powerful and precise method in many soil-plant-fertilizer studies, it should be used with caution in ecological studies investigating the role of earthworms on P availability.

In summary, earthworms considerably increased plant growth and P nutrition. Considering all interpretation precautions cited above for the *L-value*, we can conclude that most of the earthworm-driven P availability is attributed to *P. corethrurus* activities that partially solubilize soil P through soil transit in its digestive tract. However, our experiment showed that the contribution of dead *P. corethrurus* decomposition to plant P nutrition was significant (approximately 30%). The effects of earthworms on P availability are dependent on species and ecological categories (Suárez et al., 2004). Thus, the relative contribution of the different mechanisms quantified in this study should not be considered generic. However, according to the worm density inoculated and the duration of the pot experiments, the survival of earthworms should be considered while evaluating plant-available P.

Acknowledgements

This work was supported by the French National Programme EC2CO-

Biohefect/Ecodyn/Dril/MicrobiEn/BAC-RIP. We acknowledge the Laboratoire des

RadioIsotopes of the University of Antananarivo, Madagascar, for providing access to the

experimental station in Lazaina and the greenhouse. We thank Modeste Rakotondramanana,

Marie-Paule Razafimanantsoa, Andrianantenaina Hilaire Damase Razafimahafaly, and Eric

N. Randrianasolo for laboratory assistance. We also thank the two anonymous reviewers who

allowed us to improve the quality of the manuscript.

- 287 **Reference**
- Achat, D.L., Sperandio, M., Daumer, M.-L., Santellani, A.-C., Prud'Homme, L., Akhtar, M.,
- Morel, C., 2014. Plant-availability of phosphorus recycled from pig manures and dairy
- effluents as assessed by isotopic labeling techniques. Geoderma 232, 24-33.
- Barois, I., Lavelle, P., 1986. Changes in respiration rate and some physicochemical properties
- of a tropical soil during transit through *Pontoscolex corethrurus* (Glossoscolecidae,
- Oligochaeta). Soil Biol Biochem 18, 539-541.
- Barois, I., Lavelle, P., Brossard, M., Tondoh, J., Martinez, A., Rossi, J., Senapati, B., Angeles,
- A., Fragoso, C., Jimenez, J., 1999. Ecology of earthworm species with large environmental
- tolerance and/or extended distributions. In Lavelle P. (ed), Brussaard L. (ed), Hendrix P. (ed).
- 297 Earthworm management in tropical agroecosystems. Wallingford: CABI, p. 57-84. ISBN 0-
- 298 85199-270-6.
- Bertrand, M., Blouin, M., Barot, S., Charlier, A., Marchand, D., Roger-Estrade, J., 2015.
- 300 Biocontrol of eyespot disease on two winter wheat cultivars by an anecic earthworm
- 301 (Lumbricus terrestris). Appl Soil Ecol 96, 33-41.
- 302 Blanchart, E. 1992. Restoration by earthworms (Megascolecidae) of the macroaggregate
- 303 structure of a destructured savannah soil under field conditions. Soil Biol Biochem 24,
- 304 1587:1594.
- Blouin, M., Hodson, M.E., Delgado, E.A., Baker, G., Brussaard, L., Butt, K.R., Dai, J.,
- Dendooven, L., Pérès, G., Tondoh, J., 2013. A review of earthworm impact on soil function
- and ecosystem services. Eur J Soil Sci 64, 161-182.
- 308 Bühler, S., Oberson, A., Sinaj, S., Friesen, D.K., Frossard, E., 2003. Isotope methods for
- assessing plant available phosphorus in acid tropical soils. Eur J Soil Sci 54, 605-616.
- Chapuis-Lardy, L., Le Bayon, R.-C., Brossard, M., López-Hernández, D., Blanchart, E., 2011.
- Role of soil macrofauna in phosphorus cycling, Phosphorus in Action. Springer, pp. 199-213.

- 312 Chapuis-Lardy, L., Ramiandrisoa, R., Randriamanantsoa, L., Morel, C., Rabeharisoa, L.,
- 313 Blanchart, E., 2009. Modification of P availability by endogeic earthworms
- 314 (Glossoscolecidae) in Ferralsols of the Malagasy Highlands. Biol Fert Soils 45, 415-422.
- Chassé, P., Pelosi, C., Lata, J.-C., Barot, S., 2019. Impact of crop genetic diversity on a litter
- 316 consumer. Basic and Appl Ecol 36, 1-11.
- Coulis, M., Bernard, L., Gerard, F., Hinsinger, P., Plassard, C., Villeneuve, M., Blanchart, E.,
- 318 2014. Endogeic earthworms modify soil phosphorus, plant growth and interactions in a
- 319 legume–cereal intercrop. Plant Soil 379, 149-160.
- 320 Derouard, L., Tondoh, J., Vilcosqui, L., Lavelle, P., 1997. Effects of earthworm introduction
- on soil processes and plant growth. Soil Biol Biochem 29, 541-545.
- Fonte, S.J., Botero, C., Quintero, D.C., Lavelle, P., van Kessel, C., 2019. Earthworms regulate
- 323 plant productivity and the efficacy of soil fertility amendments in acid soils of the Colombian
- 324 Llanos. Soil Biol Biochem 129, 136-143.
- Frossard, E., Morel, J., Fardeau, J., Brossard, M., 1994. Soil isotopically exchangeable
- phosphorus: a comparison between E and L values. Soil Sci. Soc. Am. J. 58, 846-851.
- 327 Ghosh, M., Chattopadhyay, G., Baral, K., 1999. Transformation of phosphorus during
- 328 vermicomposting. Bioresource Technol 69, 149-154.
- Hauser, S., 1993. Distribution and activity of earthworms and contribution to nutrient
- recycling in alley cropping. Biol Fert Soils 15, 16-20.
- Hedley, M., White, R., Nye, P., 1982. Plant-induced changes in the rhizosphere of rape
- 332 (Brassica napus var. Emerald) seedlings: III. Changes in L value, soil phosphate fractions and
- 333 phosphatase activity. New Phytol 91, 45-56.
- Hinsinger, P., Gilkes, R., 1997. Dissolution of phosphate rock in the rhizosphere of five plant
- species grown in an acid, P-fixing mineral substrate. Geoderma 75, 231-249.

- Hocking, P.J., 2001. Organic acids exuded from roots in phosphorus uptake and aluminum
- tolerance of plants in acid soils. Adv Agron 74, 63-67.
- 338 Hou, E., Luo, Y., Kuang, Y., Chen, C., Lu, X., Jiang, L., Luo, X., Wen, D., 2020. Global
- meta-analysis shows pervasive phosphorus limitation of aboveground plant production in
- natural terrestrial ecosystems. Nat Commun 11, 1-9.
- Huang, J., Zhang, W., Liu, M., Briones, M.J., Eisenhauer, N., Shao, Y., Fu, S., Xia, H., 2015.
- 342 Different impacts of native and exotic earthworms on rhizodeposit carbon sequestration in a
- 343 subtropical soil. Soil Biol Biochem 90, 152-160.
- Larsen, S., 1952. The use of P 32 in studies on the uptake of phosphorus by plants. Plant Soil
- 345 4, 1-10.
- Lavelle, P., Barot, S., Blouin, M., Decaëns, T., Jimenez, J.J., Jouquet, P., 2007. Earthworms
- as key actors in self-organized soil systems. Ecosystem Engineers: Plants to Protists 405.
- Lavelle, P., Blanchart, E., Martin, A., Spain, A., Martin, S., 1992. Impact of soil fauna on the
- properties of soils in the humid tropics. SSSA Spec Publ 29, 157-185.
- Lavelle, P., Charpentier, F., Villenave, C., Rossi, J.-P., Derouard, L., Pashanasi, B., André, J.,
- Ponge, J.-F., Bernier, N., 2004. Effects of earthworms on soil organic matter and nutrient
- dynamics at a landscape scale over decades. Earthworm Ecology 2, 145-160.
- 353 Le Bayon, R.-C., Milleret, R., 2009. Effects of earthworms on phosphorus dynamics–a
- review. Dynamic Soil, Dynamic Plant 3, 21-27.
- Liu, T., Chen, X., Gong, X., Lubbers, I.M., Jiang, Y., Feng, W., Li, X., Whalen, J.K.,
- Bonkowski, M., Griffiths, B.S., 2019. Earthworms coordinate soil biota to improve multiple
- ecosystem functions. Curr Biol 29, 3420-3429.
- Lourdumary, A.B., Uma, K., 2013. Nutritional evaluation of earthworm powder (Lampito
- 359 mauritii). J Appl Pharm Sci 3, 82.

- Murphy, J., Riley, J.P., 1962. A modified single solution method for the determination of
- 361 phosphate in natural waters. Anal Chim Acta 27, 31-36.
- Neumann, G., Massonneau, A., Martinoia, E., Römheld, V., 1999. Physiological adaptations
- 363 to phosphorus deficiency during proteoid root development in white lupin. Planta 208, 373-
- 364 382.
- Paul, B.K., Lubbers, I.M., van Groenigen, J.W., 2012. Residue incorporation depth is a
- 366 controlling factor of earthworm-induced nitrous oxide emissions. Glob Change Biol 18, 1141-
- 367 1151.
- Pypers, P., van Loon, L., Diels, J., Abaidoo, R., Smolders, E., Merckx, R., 2006. Plant-
- available P for maize and cowpea in P-deficient soils from the Nigerian Northern Guinea
- 370 Savanna–comparison of E-and *L-values*. Plant Soil 283, 251-264.
- Raminoarison, M., Razafimbelo, T., Rakotoson, T., Becquer, T., Blanchart, E., Trap, J., 2020.
- 372 Multiple-nutrient limitation of upland rainfed rice in ferralsols: a greenhouse nutrient-
- omission trial. J Plant Nutr 43, 270-284.
- Randriamanantsoa, L., Morel, C., Rabeharisoa, L., Douzet, J.M., Jansa, J., Frossard, E., 2013.
- 375 Can the isotopic exchange kinetic method be used in soils with a very low water extractable
- 376 phosphate content and a high sorbing capacity for phosphate ions? Geoderma 200, 120-129.
- Ranoarisoa, M.P., Morel, C., Andriamananjaraa, A., Jourdan, C., bernard, L., Becquer, T.,
- Rabeharisoa, L., Rahajaharilazaa, K., Plassard, C., Blanchard, E., Trap, J., 2018. Effects of a
- bacterivorous nematode on rice ³²P uptake and root architecture in a high P-sorbing ferrallitic
- 380 soil. Soil Biol Biochem 122, 39-49.
- Ratsiatosika, O., Bernard, L., Rabary, B., Rainihanjarimanana, I., Randriamanantsoa, R.,
- Razafimbelo, T., Razafindrakoto, M., Trap, J., Blanchart, E., 2019. Earthworm functional
- groups, residue quality and management impact on upland rice growth and yield–an
- experimental study in the Madagascar highlands. J Exp Agric Int 30, 1-14.

- Razafindrakoto, M., Csuzdi, C., Rakotofiringa, S., Blanchart, E., 2010. New records of
- as earthworms (Oligochaeta) from Madagascar. Opuscula Zoologica (Budapest) 41, 231-236.
- Ros, M.B., Hiemstra, T., van Groenigen, J.W., Chareesri, A., Koopmans, G.F., 2017.
- Exploring the pathways of earthworm-induced phosphorus availability. Geoderma 303, 99-
- 389 109.
- Russell, R.S., Russell, E., Marais, P., 1957. Factors affecting the ability of plants to absorb
- 391 phosphate from soils: I. The relationship between labile phosphate and absorption. J Soil Sci
- 392 8, 248-267.
- 393 Sample, E., Soper, R., Racz, G., 1980. Reactions of phosphate fertilizers in soils. Am. Soc.
- 394 Agron., Madison, WI.
- Suárez, E.R., Pelletier, D.M., Fahey, T.J., Groffman, P.M., Bohlen, P.J., Fisk, M.C., 2004.
- 396 Effects of exotic earthworms on soil phosphorus cycling in two broadleaf temperate forests.
- 397 Ecosystems 7, 28-44.
- R Core Team, 2016. R Studio: Integrated Development for R [Internet].[cited 2015 Nov 20].
- Turner, B.L., 2006. Organic phosphorus in Madagascan rice soils. Geoderma 136, 279-288.
- 400 van Groenigen, J.W., van Groenigen, K.J., Koopmans, G.F., Stokkermans, L., Vos, H.M.,
- 401 Lubbers, I.M., 2019. How fertile are earthworm casts? A meta-analysis. Geoderma 338, 525-
- 402 535.
- Vos, H.M., Koopmans, G.F., Beezemer, L., de Goede, R.G., Hiemstra, T., van Groenigen,
- J.W., 2019. Large variations in readily-available phosphorus in casts of eight earthworm
- species are linked to cast properties. Soil Biol Biochem 138, 107583.

Fig. 1. (A) Soil aggregation rate according to earthworm presence expressed in mass percentage. Whiskers correspond to standard deviation. (B) Contribution of the decomposition pathway on plant P nutrition following earthworm inoculation expressed as percentage of the *L-value*.

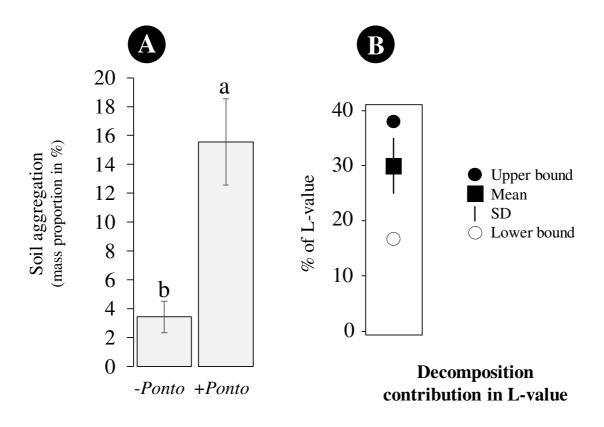


Table 1. Plant variables measured after 28 d of growth in the presence (+*Ponto*) or absence (-*Ponto*) of the earthworm *Pontoscolex corethrurus* in a Ferralsol from Madagascar.

Variables	Unit	Treatments					
		-Ponto			+Ponto)	
Shoot biomass	mg mesocosm ⁻¹	537	(65)	b	677	(36)	a
Root biomass	mg mesocosm ⁻¹	441	(71)	a	445	(27)	a
Shoot: root ratio	ratio	1.2	(0.2)	a	1.5	(0.1)	a
Total biomass	mg mesocosm ⁻¹	978	(118)	b	1122	(29)	a
Shoot P amount	mg mesocosm ⁻¹	0.37	(0.05)	b	0.61	(0.09)	a
Root P amount	mg mesocosm ⁻¹	0.24	(0.05)	b	0.32	(0.02)	a
Total P amount	mg mesocosm ⁻¹	0.61	(0.09)	b	0.95	(0.11)	a
Shoot L-value	mg-P mesocosm ⁻¹	6.8	(0.9)	b	14.2	(1.3)	a
Shoot N amount	mg mesocosm ⁻¹	15.1	(2.3)	a	16.1	(0.7)	a
Shoot K amount	mg mesocosm ⁻¹	19.1	(2.6)	a	20.2	(1.1)	a
Shoot Ca amount	mg mesocosm ⁻¹	1.8	(0.5)	a	1.9	(0.1)	a
Shoot Mg amount	mg mesocosm ⁻¹	1.2	(0.2)	a	1.2	(0.1)	a
Shoot N:P ratio	ratio	38.8	(6.7)	a	22.4	(2.7)	b
Shoot Ca:Mg ratio	ratio	1.4	(0.1)	a	1.5	(0.1)	a
Shoot K:(Ca+Mg) ratio	ratio	6.5	(1.16)	a	6.3	(0.6)	a
Final P in seed residues	μg-P mesocosm ⁻¹	7.9	(0.49)	b	13.1	(0.41)	a
Earthworm P content Initial P in seed at seedling	mg-P dry g ⁻¹	5.3	(2.24)				
stage	mg-P dry kg ⁻¹	1343	(73)				

Mean (SD). * P < 0.05; ** P < 0.01; *** P < 0.001. Letters (a and b) indicate significant differences between '-Ponto' and '+Ponto' treatments according to the Student t test at P < 0.05.