

Digestive n-6 Lipid Oxidation, a Key Trigger of Vascular Dysfunction and Atherosclerosis in the Western Diet: Protective Effects of Apple Polyphenols

Gaëtan Bolea, Clothilde Philouze, Mathilde Dubois, Sydney Risdon, Anaïs Humberclaude, Christian Ginies, Anne-laure Charles, Bernard Geny, Cyril Reboul, Claire Arnaud, et al.

▶ To cite this version:

Gaëtan Bolea, Clothilde Philouze, Mathilde Dubois, Sydney Risdon, Anaïs Humberclaude, et al.. Digestive n-6 Lipid Oxidation, a Key Trigger of Vascular Dysfunction and Atherosclerosis in the Western Diet: Protective Effects of Apple Polyphenols. Molecular Nutrition and Food Research, 2021, pp.2000487. 10.1002/mnfr.202000487. hal-03152465

HAL Id: hal-03152465 https://hal.inrae.fr/hal-03152465v1

Submitted on 25 Jul 2023

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 2 3	Digestive n-6 lipid oxidation, a key trigger of vascular dysfunction and atherosclerosis in the Western diet: protective effects of apple polyphenols
4 5 6	Gaëtan BOLEA ^{1,2} , Clotilde PHILOUZE ¹ , Mathilde DUBOIS ¹ , Sydney RISDON ¹ , Anaïs HUMBERCLAUDE ¹ , Christian GINIES ² , Anne-Laure CHARLES ⁴ , Bernard GENY ⁴ , Cyril REBOUL ¹ , Claire ARNAUD ³ , Claire DUFOUR* ² , Grégory MEYER * ¹
7 8	¹ EA4278 LaPEC "Laboratory of Cardiovascular Pharm-ecology ", Avignon University, F-84000 Avignon, France
9 10	² INRAE, Avignon University, UMR408 SQPOV "Safety and Quality of Plant Products", F-84000 Avignon, France.
11 12	³ INSERM, Grenoble University, U1042 HP2 "Cardiovascular & Respiratory Pathophysiology and Hypoxia", F-38000 Grenoble, France
13 14 15	⁴ UR3072 University of Strasbourg, Translational Medicine Federation of Strasbourg (FMTS), Faculty of Medicine, Team 3072 "Mitochondria, Oxidative Stress and Muscle Protection", F-67000 Strasbourg, France
 16 17 18 19 20 21 22 23 24 25 26 27 28 29 	Corresponding authors Dr. G. MEYER Laboratoire de Pharm-Ecologie Cardiovasculaire (EA4278), Faculty of Sciences, Avignon University, 33 rue Louis Pasteur, 84000 Avignon, France Phone: +33 490162944 ; Fax: +33 490162901 ; E-mail: gregory.meyer@univ-avignon.fr Dr. C. DUFOUR INRAE, UMR SQPOV, 228 route de l'aérodrome, 84914 Avignon Cedex 9, France Phone : +33 432722515 ; Fax : +33 432722492 ; E-mail : claire.dufour@inrae.fr
30 31 32 33 34	Abbreviations: 4-HNE: 4-hydroxy-2-nonenal; Ach: Acetylcholine; ApoE: ApolipoproteinE; EPR: Electron Paramagnetic Resonance; HF: High Fat; NO: Nitric Oxide; oxLDL: oxidized LDL; PE: Phenolic Extract; PC: Proanthocyanidins; PUFA: polyunsaturated fatty acids; ROS: Reactive Oxygen Species; RM: Red Meat; SNP: Sodium Nitroprusside; SO: Sunflower Oil; WD: Western Diet.

35 Keywords: 4-Hydroxy-2-nonenal, Atherosclerosis, Endothelial function, Polyphenols,
36 Western diet

37 Abstract

38 Scope

A main risk factor of atherosclerosis is a Western diet (WD) rich in n-6 PUFA sensitive to oxidation. Their oxidation can be initiated by heme iron of red meat leading to the formation of 4-hydroxy-2-nonenal (4-HNE), a cytotoxic aldehyde. An increased 4-HNE production is implicated in endothelial dysfunction and atherosclerosis. By contrast, a diet rich in proanthocyanidins reduces oxidative stress and arterial diseases. This study evaluates the effects of a WD on vascular integrity in ApoE^{-/-} mice and the protective capacity of apple extract and puree rich in antioxidant proanthocyanidins.

46 Methods and results

ApoE^{-/-} mice were fed during 12 weeks with a WD with or without n-6 PUFA. Moreover, two WD + n-6 PUFA groups were supplemented with apple puree or phenolic extract. We report an increase in digestive 4-HNE production associated with a rise in plasmatic 4-HNE and oxLDL concentrations. Oxidizable n-6 PUFA consumption was associated with a worsened endothelial dysfunction and atherosclerosis. Interestingly, supplementations with apple polyphenol extract or puree prevented these impairments while reducing oxidative stress.

53 Conclusion

n-6 lipid oxidation during digestion may be a key factor of vascular impairments.
Nevertheless, an antioxidant strategy could limit 4-HNE formation during digestion and thus
durably protect vascular function.

- 57
- 58
- 59
- 60 61
- 62

- 63 INTRODUCTION
- 64

Atherosclerosis, a leading cause of mortality, is a multifactorial and degenerative disease 65 characterized by the progressive accumulation of lipids and inflammatory cells on the wall of 66 large and medium arteries. Such changes result in blood flow impairments, chronic 67 inflammation and oxidative stress in the vascular wall. ^[1,2] Lipid accumulation within the 68 arterial wall is preceded by an altered vascular endothelial function, which is explained by a 69 70 complex interplay between reduced nitric oxide (NO) bioavailability, oxidative stress and inflammation.^[2,3] One of the major risk factors of atherosclerosis development is the Western 71 diet rich in n-6 polyunsaturated fatty acids (n-6 PUFA) which are highly sensitive to oxidation. 72 73 In the digestive tract, food is exposed to postprandial oxidative stress and therefore the gastric tract has been proposed as a major site for lipid oxidation. ^[4-6] Indeed, in the Western diet, 74 heme iron in the form of myoglobin from red meat can initiate lipid oxidation leading to the 75 76 formation of 4-hydroxy-2-nonenal (4-HNE). 4-HNE is a specific genotoxic and cytotoxic α , β unsaturated hydroxyalkenal generated from n-6 PUFA and found in some oxidative stress-77 related and inflammatory diseases.^[7] Keller et al. (2015)^[8] have shown that marked 4-HNE 78 administered to rats by intragastric gavage was largely absorbed by the intestinal barrier since 79 it was found in urine, liver, or kidney.^[8] This highly reactive aldehyde binds covalently (via 80 Michael addition or Schiff base formation) with free amino or thiol groups of cysteine, lysine 81 and histidine.^[9] This adduction results in an impairment of the protein conformation and 82 83 function inducing subsequent cellular dysfunctions and tissular damages. As a matter of fact, the covalent binding of 4-HNE to ApoB100, which yields oxidized LDL (oxLDL), is 84 recognized as a key step in the development of an endothelial dysfunction and the subsequent 85 atheromatous plaque formation.^[10,11] 86

87 In the last decades, epidemiological, clinical and experimental studies have demonstrated that a healthy diet plays a central role in the prevention of atherosclerosis. As a matter of fact, 88 89 inverse associations are repeatedly outlined between coronary artery disease and stroke and the consumption of fruit and vegetables. ^[12,13] In recent controlled trials with cocoa and other 90 91 flavonoid-rich fruit and vegetables, the vascular function was improved along with the plasma NO status and inflammation-related markers. ^[14,15] As a matter of fact, proanthocyanidins 92 93 (PCs) appear as potential mediators in reducing blood pressure and improving endotheliummediated dilation whereas their plasma metabolites were found to reduce monocyte adhesion 94 to endothelial cells through modulation of gene expression and TNF- α . ^[16,17,18,19] PCs are the 95 second polyphenol class in the French diet with apple being the major contributor.^[20] 96

97 Monomeric and oligomeric PCs represent 80% of the apple phenolic pool followed by hydroxycinnamic acids (15%), flavonols and dihydrochalcones. After ingestion, native forms 98 of polyphenols are bioaccessible in the stomach after their release from the plant matrix.^[21] 99 Antioxidant apple polyphenols were found to inhibit lipid oxidation in *in vitro* gastric 100 101 digestion.^[22] Through their catechol core, they can reduce the pro-oxidant hypervalent iron form of myoglobin (MbFe^{IV}=O) to metmyoglobin (MbFe^{III}) as well as chelate free iron 102 preventing lipid oxidation initiation.^[23] Antioxidant effects of apple polyphenols in the 103 104 gastrointestinal tract could limit 4-HNE formation and its subsequent absorption. Limiting 4-105 HNE absorption can reduce LDL oxidation and thus protect the vascular function.

106

107 The aims of this study were to evaluate 1) the effects of a chronic Western diet and the 108 subsequent lipid oxidation occurring in the digestive tract on both the vascular function and 109 atherosclerosis development in $ApoE^{-/-}$ mice, and 2) the consequences of a primary prevention 110 strategy by a supplementation in polyphenols rich in PCs.

111

112 MATERIALS AND METHODS

113 Animal model and study design

All investigations are conformed to the Guide for the Care and Use of Laboratory Animals 114 115 published by the US National Institutes of Health (NIH, National Academies Press US, 8th edition, 2011) as well as European Parliament Directive 2010/63/EU (experimentation n°: 116 117 84.004. Eight weeks old male C57Bl6J mice (n = 100) knockout for apolipoproteinE (ApoE^{-/-}) 118 (Charles River, France / Breeding in HP2 laboratory, Grenoble) were used for this study. Mice 119 were housed five by cage under controlled conditions of temperature (21 ± 1 °C), hygrometry 120 $(60 \pm 10\%)$ and lightening (12:12 hours light-dark cycle), with access to water and food ad 121 *libitum*. After two weeks of acclimatization, male mice were randomly assigned to five groups 122 (n = 20 per group): a first group with a normocholesterolemic standard diet (SD group) (SAFE 123 A04), a Western diet group with High Fat with red meat (WD group), a second Western diet 124 group complemented with sunflower oil (WD+SO group) and two other groups with a 125 WD+SO diet supplemented with apple polyphenols in the form of apple puree 126 (WD+SO+PUREE group) or the corresponding phenolic extract (WD+SO+PE group). SD group was only conducted in order to confirm the effects of our Western Diet in a model of
 ApoE -/- mice. Consequently, differences observed between SD and WD groups are reported in
 supplemental data.

130 Diet protocols were sustained during 12 weeks and food intake was quantified as a mean per 131 cage (n = 3 cages with 6/7 mice per cage) each three days during the whole protocol. During 132 this period, weight gain was measured every week and dietary intake was evaluated three times 133 per week. Faeces were harvested at the start, middle and end of the protocol. During the last 134 week, blood pressure was evaluated by tail cuff method in 12 animals per group and 135 atheromatous plaque size by echography on the 8 other animals of the group. At the end of the 136 12 weeks-diet period, an overnight fasting was performed and mice were euthanized by sodium 137 pentobarbital injection (120 mg/kg I.P.). Blood was collected for biochemical analyses. Hearts 138 were excised with aortic arch and fixed in O.C.T then frozen in liquid nitrogen and stored at -139 80 °C for histological analysis. Finally, thoracic aorta was removed in order to perform ex-vivo 140 analysis of vascular function (n = 8/grp), quantification of atheromatous plaque development 141 (n = 8 for a ortic staining with Oil-Red O) or biochemical assays (n = 4 per group)

142

143 **Dietary protocol**

144 Regarding the diet protocol, the first group was fed with a High Fat and red meat diet (HF 145 230, Safe, France) (Western diet, WD). Composition in % kcal: proteins 16.1, carbohydrates 146 24.2, lipids 59.7 for a total of 12.1 kcal per day. Composition in % of diet weight: High Fat 147 79.5, red meat 19.9, cellulose 0.64, n-6 PUFA 10.5, cholesterol 0.076, Vitamin E 0.014 and 148 heme iron 0.0013. The second group was fed with a WD diet complemented with sunflower oil 149 (SO, Rustica from Leclerc, lot A21815) as a source of oxidizable n-6 PUFA (WD+SO group). 150 Composition in % kcal: proteins 12.3, carbohydrates 18.5, lipids 69.2 for a total of 13.0 kcal 151 per day. Composition in % of diet weight: High Fat 68.6, sunflower oil 13.7, red meat 17.1, 152 cellulose 0.55, n-6 PUFA 28.3, cholesterol 0.066, Vitamin E 0.021 and heme iron 0.0011. The 153 2 last groups were fed with a WD+SO diet supplemented with apple (Reinette de Flandre, 154 Ambricourt, France) polyphenols in the form of apple puree (whole apple matrix except seeds) 155 for the WD+SO+PUREE group or the corresponding phenolic extract for the WD+SO+PE 156 group. Composition for WD+SO+PUREE group in % kcal: proteins 12.1, carbohydrates 19.4, 157 lipids 68.5 for a total of 11.7 kcal per day; in % of diet weight: High Fat 60.6, sunflower oil

12.1, red meat 15.1, Puree 12.1 (polyphenols 0.060), n-6 PUFA 28.3, cholesterol 0.058,
Vitamin E 0.019 and heme iron 0.0010. Composition for WD+SO+PE group in % kcal:
proteins 12.1, carbohydrates 19.6, lipids 68.3 for a total of 12.4 kcal per day; in % of diet
weight: High Fat 67.0, sunflower oil 13.4, red meat 16.7, extract 2.3 (polyphenols 0.061),
cellulose 0.54, n-6 PUFA 28.3, cholesterol 0.064, Vitamin E 0.021 and heme iron 0.0011.

163

164 **Blood pressure measurements**

165 Before the first week and during the last week of the dietary protocol, systolic, diastolic and 166 mean blood pressures were assessed in conscious mice by tail-cuff method using CODA tail 167 cuff system (Kent Scientific, Torrington, CT, USA). To reduce stress influence, mice 168 underwent 3 periods of habituation to blood pressure measurements during the week preceding 169 final recording. Mice were warmed for 30 min before measurements so that their skin 170 temperature reached 33-34 °C during measurements. All measurements were made by the 171 same experimenter between 9 AM and 12 AM. Obtained results were the mean of at least 10 172 valid measurements out of the 20 performed in a recording session.

173

174 Ultrasound imaging (echography) of atheromatous plaque

175 Echographic assessment was performed using a Vevo 3100[™] imaging system (Fujifilm 176 VisualSonics, Toronto, Canada) equipped with the MX550D[™] probe (22-55 MHz). Images 177 were analyzed post-processing using the VevoLab[™] software (Fujifilm VisualSonics, Toronto, 178 Canada). Echographic assessment as well as image analysis were performed in a blind manner. 179 Mice anesthesia was induced with 4% isoflurane (IsoFlo[™], Zoetis, Parsippany, USA) in 180 medical air mix (22% O₂, 78% N₂) (Air Liquide, Paris, France), ventilated at 1 L/min, and then 181 maintained with 1.5% isoflurane after effective sleep. Animals were placed in a supine position 182 on a heating pad and their body temperature was monitored throughout the exam, as well as 183 their breathing rate and electrocardiogram. Cross-sectional images of ascending aorta were 184 obtained from a modified right parasternal view. Aortic plaque surface areas were determined 185 from 3 measurements at the same spot. All measurements were performed between two 186 inspiratory peaks.

188 Isolated aortic rings and vasoreactivity

189 Under anesthesia (120 mg/kg I.P.), thoracic aortas were quickly removed and placed in cold 190 Krebs-Henseleit bicarbonate buffer (composition in mM: NaCl 118, NaHCO₃ 25, KCl 4.8, 191 KH₂PO₄ 1.2, CaCl₂ 2.5 and glucose 11). After removal of adherent perivascular adipose 192 tissues, aortas were cut into 2 mm long rings. Then, aortic rings were set onto stainless steel 193 connected to an isometric force transducer, linked to an amplifier and a computerized 194 acquisition system, to record changes in isometric force (EMKA technologies, EMKA Paris, 195 France). They were suspended in organ bathes containing Krebs-Henseleit buffer at 37 °C continuously bubbled with gas mixture (95% $O_2 - 5\% CO_2$). Resting tension was adjusted to 196 197 0.8 g and aortic rings were allowed to stabilize during 60 min. During this period, Krebs-198 Henseleit buffer was changed two times and rings were re-stretched to maintain a tension of 199 0.8 g. First, smooth muscle cells and endothelial integrity were assessed by a single dose of phenylephrine (PE, 10⁻⁶ M) followed by a single vasorelaxing dose of acetylcholine (ACh, 10⁻⁵ 200 M). Then, each vessel ring was pre-constricted with phenylephrine (10^{-6} M) . After pre-201 constriction reached a plateau, endothelium-dependent and independent relaxations were 202 examined by challenging aortic rings with cumulative concentrations of ACh $(10^{-9} \text{ to } 10^{-5} \text{ M})$ 203 or sodium nitroprusside (SNP, 10^{-9} M to 10^{-5} M) respectively. Vasodilatation was expressed as 204 a percentage of maximum contractile response to PE. Maximal relaxation (Emax) and 205 206 sensitivity (EC50) to each drug were used to characterize vasorelaxation.

207

208 Blood analysis

Blood analyses were performed on plasma obtained by centrifuging blood at 300 *g* for 10 min at 4 °C and plasma was stored at -80 °C for biochemical analyses. Total cholesterol (TC), high-density lipoprotein cholesterol (HDL) and low-density lipoprotein cholesterol (LDL) concentrations were determined by the CHOD-PAP method (Biolabo SAS, France). Triglycerides (TG) were assessed by the GPO method (Biolabo SAS, France). Finally, malondialdehyde-modified oxLDL were assessed by an ELISA kit (Cloud-Clone Corp., USA).

215

216 **Biochemical assays**

217

Measurement of reactive oxygen species (ROS) by electron paramagnetic resonance (EPR)

ROS production was measured by EPR in aortic homogenates as previously described.^[24] 218 219 Briefly, 50 µL of homogenates were incubated with 5 µL of Krebs-Hepes buffer (pH 7.4) 220 containing 1 mM of CMH (1-hydroxy-3-methoxycarbonyl-2,2,5,5-tetramethylpyrrolidine) and 221 25 µM deferroxamine methane sulfonate. This solution was placed inside the e-scan 222 spectrometer (Bruker, Ettlingen, Germany) for data acquisition at 37 °C. The acquisition EPR parameters were: microwave frequency = 9.652 GHz; modulation frequency = 86 kHz, 223 modulation amplitude = 0.01 G; center field = 3495.9 G, sweep width = 300 G; microwave 224 power = 1.0 mW; number of scans = 10 and receiver gain = 2.10^{-1} . Adducts generated by the 225 226 probe reaction with free radicals were acquired and the spectra sequentially recorded for about 227 5 min in order to calculate the ROS production rate. The ROS production was normalized to the protein content of each sample and then expressed in μ mol min⁻¹mg⁻¹. 228

229 Nitrites in aorta

Nitrites from descending aorta, used as an index of total NO production, were quantified by using the high-sensitivity nitrite assay kit (Measure-iTTM High-Sensitivity Nitrite Assay Kit; InvitrogenTM) and was performed according to the manufacturer's instructions. Measurements were performed at the end of 12 weeks of protocol diet. NO production was expressed relative to protein content of each sample.

235

236 *Quantification of faeces and plasma 4-HNE*

237 Lipid oxidation during digestion was evaluated by the quantification of 4-HNE in faeces and 238 plasma. Faeces were collected 24 hours after diet change at the start, middle and end of the 12 239 weeks protocol. Faeces (200 mg) were crushed with mortar and pestle in liquid nitrogen. The 240 resulting powder (ca. 30 mg) was vortexed once with 500 µL of acetonitrile containing 50 ng 241 of 4-HNE-D3 (internal standard) during 5 min. After centrifugation (16 000 g, 5 min, room 242 temperature), the supernatant was reacted with 100 µL of a solution of 2,4-243 dinitrophenylhydrazine (75 mg of 2,4-DNPH added with 0.6 mL of formic acid and 10 mL of 244 acetonitrile) during 1 hour at room temperature. For plasma, 70 µL were vortexed with 200 µL 245 of acetonitrile and 20 µL of 4-HNE-D3 (10 ng/mL in ACN) during 5 min. After centrifugation 246 as above, the supernatant is dried under nitrogen and reacted with 50 μ L of a solution of 2,4-247 DNPH (25 mg of 2,4-DNPH added with 0.2 mL of formic acid and 10 mL of acetonitrile) 248 during 1 hour at room temperature. Separation and quantification of derivatized 4-HNE were 249 performed by using LC/MS/APCI (EVOQ Elite, Bruker Daltonics, Bremen, Germany) with an 250 ACOUITY HSS T3 column (50 mm x 2.1 mm; 1.7 µm, Waters, Saint-Quentin-en-Yvelines, 251 France) at 40 °C. For chromatographic separation, the mobile phase was constituted by a 252 binary solvent system with water/formic acid (99.95/0.05, v/v, solvent A) and acetonitrile 253 (solvent B) at the flow rate of 0.4 mL/min. The volume injected was 4 µL. The elution gradient 254 was as follows: 0-1 min isocratic 50% B; 1-9 min, linear 50-98% B; 9-10 min, isocratic 98% 255 B; 10-10.1 min, linear 98-50% B and 10.1-11 min, isocratic 50% B. Mass spectrometry 256 conditions were as follows: ionization by APCI in negative mode, spray current 20 µA; cone 257 temperature 300 °C; heated probe temperature 300 °C; cone gas flow 20; probe gas flow 40; 258 nebulizer gas flow 50. Spectra were recorded in the MRM mode. Derivatized 4-HNE-D3 was 259 quantified by following the transition from ion at m/z 338 to ion at m/z 167 and derivatized 4-260 HNE with transition from m/z 335 to m/z 167. The retention time of both compounds was 3.5 261 min. For quantification, 5 point-calibrations were injected at constant concentration for 4-262 HNE-D3 (0.28 µM) and between 1.02 µM and 0.064 µM for 4-HNE. The adducts with DNPH 263 were stable over 6 h with a variation lower than 5%.

264

265 *OIL red coloration*

266 Atheromatous plaque size was assessed by measuring lipid aggregation in thoraco-267 abdominal aortas that were fixed in paraformaldehyde (4%) during 24 hours at 4 °C. Then, 268 aortas were rinsed three times with Krebs-Henseleit buffer and incubated overnight in an Oil-269 Red-O solution in isopropanol/water (60/40, v/v, 0.33% of Oil-Red-O, Sigma-Aldrich, Saint-270 Louis, USA) at room temperature. Atheromatous plaque size was assessed by capturing images 271 directly from a color camera (Motic Moticam 2300, Motic China group Co.) attached to a binocular loupe. Images were displayed on a RGB monitor by using Motic Image 2.0 (Motic 272 China group Co.). Analysis was carried out using Image J[®] software (Image J, NIH, USA). Oil 273 274 red staining was expressed as the percentage of aortic area.

275

276 *Histological sections and colorations*

The upper part of the heart (with aortic root) was cut using a cryostat (Leica CM1950; Leica systems, France) at -20 °C. The distal portion of the aortic sinus was recognized by the appearance of the three leaflets of the valves. Sections of 8 µm thickness were performed. For 280 each aortic root, we quantified atheromatous plaque size and necrotic core size (acellular area) 281 with Oil-Red-O and Hematoxylin and Eosin staining (Sigma-Aldrich, France) and fibrosis 282 (Red Sirius, Labomoderne, Paris, France) from five sections separated from each other by 80 283 µm. Photography was performed using a digital camera (Baumer VCXU31C, Baumer SAS 284 France), mounted with a macro lens VS Technology 0513 (VS Technology Corporation, 285 Japan) and Baumer GAPI acquisition software (Baumer SAS France). Quantitative analyses were performed blindly using Image J[®] software to assess the stained area in mm² and express 286 it relative to the diameter of aorta in mm (atheromatous plaque size and necrotic core size) or 287 288 in percentage of atheroma (fibrosis).

289

290 Statistical analysis

Data were expressed as mean \pm SEM. Normality was evaluated using D'Agostino and Pearson test when n≥8. Otherwise, Shapiro-Wilk test was performed. When applicable, comparison of multiple experimental conditions were performed using analysis of variance (ANOVA) or repeated measures ANOVA followed by a Tukey post hoc test. A value of p<0.05 was considered statistically significant. Statistics were performed using GraphPad Prism 8 (GraphPad Software, La Jolla, USA).

297

298

299 **RESULTS**

Diet Characterization

ApoE^{-/-} mice, classically used to evaluate atherosclerosis development, were distributed to 301 302 four groups: a group with a high fat and red meat diet (WD), a second group with a WD diet 303 complemented with sunflower oil as a source of oxidizable n-6 PUFA (WD+SO group) and 304 two other groups with a WD+SO diet supplemented with apple polyphenols in the presence of 305 the apple matrix (Puree) or as a phenolic extract, respectively WD+SO+PUREE and WD+SO+PE groups. In parallel, a negative control group of ApoE^{-/-} mice fed with standard 306 307 diet was performed (SD, n=20, SAFE A04) to evaluate the deleterious effects of WD 308 (Supplementary data Figure 2).

As previously reported, the composition of the sunflower oil used was: 9.8% of saturated fatty acids (6.3% C16; 3.3% C18; 0.2% C20), 27.8% of monounsaturated fatty acids (0.1% C16:1; 27.0% C18:1n-9; 0.6% C18:1n-7; 0.1% C20:1) and 62.4% of polyunsaturated fatty
acids (62.4% C18:2; 0.01% C18:3). ^[25] The α-tocopherol content was 665 ppm.

313 Identification and quantification of apple polyphenols were performed by UPLC/DAD/ESI-MS and thioacidolysis as previously reported. ^[25] Forty-six compounds distributed in four 314 315 phenolic classes were identified: proanthocyanidins, hydroxycinnamic acids, dihydrochalcones 316 and flavonols. The selected variety (Reinette de Flandre) is markedly rich in polyphenols (4.3 g/kg) compared to classically consumed dessert apples (1 g/ kg). [26] Polyphenols were even 317 318 more concentrated in puree after thermal treatment of Reinette de Flandre reaching 5 g/kg. 319 Monomeric and oligomeric PCs represented 5-7% and 67-68%, respectively, of the phenolic 320 pool followed by the hydroxycinnamic acid, 5-caffeoylquinic acid (21-22%). Quercetin derivatives and dihydrochalcones were only minor contributors.^[21] In our model of apple 321 322 supplementation, no major difference was observed for polyphenol contents and composition 323 between Puree and PE. Apples used to prepare the diet also contained, 3.15 g of glucose, 5.85 324 g of fructose, 1.09 g of saccharose, 0.04 g of citric acid, 1.61 g of malic acid, 17.7 mg of 325 dehydroascorbic acid and 5.9 mg of ascorbic acid per 100 g of fresh fruit.

326

327 Lipid oxidation during digestion and blood oxidative state

328 In our model of Western Diets (high fat associated with red meat), body weight increased 329 all along the protocol to the same extent in the 4 groups (Figures 1A and 1B). In line with 330 these results, we observed no difference regarding adiposity index defined as (epididymal fat 331 + inguinal fat + perinephric fat)/mouse weight (Figure 1C), blood HDL (Figure 1D), LDL 332 (Figure 1E), HDL-C/LDL-C (Figure 1F), total cholesterol (Figure 1G) and triglycerides 333 (Figure 1H) between groups. These results suggest that there is no difference between groups 334 regarding lipid absorption. In addition, no difference between groups in term of fasting 335 glycemia as well as glucose and insulin tolerance (supplemental data) was observed. To 336 evaluate the impact of a Western diet enriched in n-6 PUFA on lipid oxidation during 337 digestion and the ability of Puree and PE to modulate this phenomenon, we next measured the 338 level of 4-HNE in faeces. Before the beginning of the diet (Week 0), only low contents of 4-339 HNE were reported at the detection limit of LC/MS/APCI (Figure 2A). After 6 (Figure 2B) 340 and 12 (Figure 2C) weeks of diet a stronger increase in 4-HNE was observed in the group 341 supplemented with SO (WD+SO) compared to the WD group. It is worth noting that 4-HNE 342 was neither detected in freshly reconstituted diets nor after 48 hours at room temperature (data 343 not shown). This suggests that lipid oxidation can occur during in vivo gastrointestinal 344 digestion. The higher levels of 4-HNE in WD+SO compared to WD can be linked to the diet 345 contents in n-6 PUFA, which are respectively of 28.3% and 10.5% for the WD+SO and WD 346 diets. Interestingly, the increase observed by the addition of n-6 PUFA was largely prevented 347 by both Puree and PE supplementations (Figure 2BC). Next, to evaluate whether lipid 348 oxidation products were also observed in systemic circulation, we measured, after 12 weeks 349 of diet, plasmatic concentrations of 4-HNE also known to be reliable biomarkers of oxidative stress.^[9] As reported in Figures 2D and E, plasmatic level of 4-HNE was higher in the 350 351 WD+SO group compared to WD by 87%. In line with the results obtained on 4-HNE in 352 faeces, this phenomenon was blunted by both apple Puree and PE supplementations by 32% 353 and 51%, respectively (Figure 2D). Finally, we measured oxLDL, another biomarker of oxidative stress, which is also known as a strong biomarker of cardiovascular risk. ^[10] The 354 355 same pattern was observed since oxLDL level was significantly increased by 14% in the 356 WD+SO group compared to WD (Figure 2E) while the supplementation by both Puree and 357 PE markedly attenuated this phenomenon (-22% and -29%, respectively) (Figure 2E).

358

359

360 **4-HNE and cardiovascular risk factors**

High levels of 4-HNE and oxLDL are associated in the literature with an increased 361 cardiovascular risk.^[10] Then, we evaluated how our Western diet model enriched in n-6 362 PUFA, supplemented or not with apple polyphenols, affected some key elements of 363 364 cardiovascular risk such as arterial blood pressure, endothelial function and finally 365 atheromatous plaque development. We first evaluated arterial blood pressure by the tail-cuff method. When the Western diet was enriched with n-6 PUFA (WD+SO), both systolic and 366 367 mean arterial pressures were increased compared to WD (Figure 3AC). However, no change in arterial diastolic pressure was observed between WD and WD+SO groups (Figure 3B). 368 369 When the WD+SO diet was supplemented with either Puree or PE, systolic, diastolic and 370 mean arterial blood pressures were significantly lower than in WD+SO group (Figure 3ABC).

Another key element of increased cardiovascular risk factor, which can contribute to the regulation of blood arterial pressure is endothelial function. Thus, we next evaluated the impact of our experimental conditions on vascular endothelial function on isolated aortic rings. In the aorta of animals fed for 12 weeks with a WD enriched in n-6 PUFA, maximal

vascular relaxation in response to ACh was reduced by 16% compared to WD (Figure 3D left 375 376 and central panel). Considering that no difference was observed between these 2 groups 377 regarding the response to SNP (Figure 3D right panel), these results strongly support that a 378 Western diet, enriched in n-6 PUFA, is associated with a vascular endothelium impairment 379 without smooth muscle cell alteration. In line with our previous results, endothelial 380 dysfunction was prevented when the animals were supplemented with both apple Puree and 381 polyphenol extract (Figure 3D left and central panel). Indeed, the maximal response to ACh 382 was increased by 41% in WD+SO+PUREE and 40% in WD+SO+PE groups compared to 383 WD+SO. An interesting point is that contrary to the WD+SO group which presented an 384 impairment of vasodilation to ACh in 75% of the tested arteries (with an impairment threshold 385 of < 70% of precontraction), no impaired aorta was reported in both supplemented groups. 386 No impact on the dose-response to SNP was observed.

Since, endothelial dysfunction is a key trigger in the pathogenesis of atherosclerosis ^[10] we 387 388 next assessed the consequences of endothelial dysfunction on the development of atherosclerosis. In our study, all experiments have been performed on ApoE^{-/-} mice, which are 389 390 prompt to develop subsequent atherosclerotic lesions at aortic bifurcations in the aortic arch, in the descending aorta and in other large arteries.^[27]. The enrichment of WD with n-6 PUFA 391 392 increased atheromatous plaque size in aortic arch section as measured by echography (Figure 393 4A). However, no difference in atherosclerosis was observed by Oil-Red-O coloration in 394 descending thoraco-abdominal aorta or by histological staining of aortic root in this WD+SO 395 group when compared to the WD group (Figure 4BC). To evaluate plaque stability, 396 percentage of fibrosis and necrotic core area size in atheromatous plaque were assessed. No 397 difference between any groups was observed regarding fibrosis (Figure 4D). However, we 398 observed that the necrotic core area increased in mice by 52% compared to the WD group 399 when the Western diet was enriched in n-6 PUFA (Figure 4E). In line with the protective 400 effect of both apple Puree and PE on endothelial dysfunction, reduced atheromatous plaque 401 developments were reported by echography in aortic cross (respectively by -20% and -20%) 402 and by Oil-Red O histological staining in a rtic root (respectively by -73% and -37%) as well 403 as by Oil-Red O staining in thoraco-abdominal aorta (respectively -51% and -45% when 404 compared to WD+SO) (Figure 4ABC). Another interesting result was that both apple Puree 405 and PE were able to prevent the increase of the necrotic core area size by 55% and 42%. 406 respectively (Figure 4E). This last result indicates that plaque stability, which is affected 407 negatively in mice fed with WD+SO, was preserved with polyphenol supplementation.

408

409 **ROS production and NO pathway in endothelial function**

410 In the vascular wall, the sensitive equilibrium between NO bioavailability and ROS 411 production contributes mainly to the regulation of the endothelial function. Moreover, 412 increased ROS production and alteration in NO bioactivity are known to be the key triggers of endothelial dysfunction and atherosclerosis.^[28] Thus, we measured the impact of our 413 414 experimental conditions on NO and ROS production in the aortic wall as well as eNOS expression and phosphorylation on its main activation site serine¹¹⁷⁷ (eNOS^{Ser1177}). No 415 416 difference was observed between WD and WD+SO groups regarding total eNOS expression and phosphorylation on Ser¹¹⁷⁷ (Suppl Figure 2). However, with both supplementations, 417 418 eNOS level tended to be lower when compared to the WD+SO group (p=0.06 and 0.08) and 419 they both seem to increase eNOS activation state through its phosphorylation on serine¹¹⁷⁷ 420 (Suppl Figure 2). To evaluate the impact of our experimental conditions on NO 421 bioavailability, we next measured the level of nitrites in aortic tissues. We report an increased 422 nitrite level in WD+SO group compared to WD mice (Figure 5A). Neither Puree 423 (WD+SO+PUREE) nor polyphenols supplementations (WD+SO+PE) were able to impact 424 nitrite levels compared to WD+SO group. Finally, since in pro-oxidative conditions, ROS can 425 react with NO to generate nitro-oxidative stress, which can also contribute to reduce NO 426 bioavailability, we also measured ROS production by EPR. We report that WD+SO led to 427 increased ROS production compared to WD (Figure 5B). By contrast, apple Puree 428 significantly reduced ROS production by 30% whereas polyphenol extract only tended to 429 reduce it (-26 %, p=0.08) when compared to the WD+SO diet. No statistical difference 430 between PUREE and PE groups could be further evidenced. Altogether, these results point out 431 that despite the beneficial effect of apple polyphenol supplementations on endothelial 432 function, their impact on the NO pathway appears limited. Nonetheless, the modulation of 433 aortic ROS production may contribute to explain some changes in NO bioavailability in our 434 groups.

435

437 **DISCUSSION**

The aim of this study was to give an insight into the mechanism involved in vascular 438 dysfunction induced by the consumption of a Western diet enriched with n-6 PUFA. ApoE^{-/-} 439 mouse is the most widely used animal model to evaluate the effects of a diet on 440 atherosclerosis development. ApoE^{-/-} mice display an increased peroxidation of lipoproteins, 441 442 an elevated level of endothelial cell adhesion molecules, an increased proliferation and migration of VSMCs as well as a decreased anti-atherogenic function.^[29] Moreover, the anti-443 444 inflammatory properties of APOE, combined with its impact on lipoprotein metabolism, 445 explain why ApoE knockout mice display a very strong development of atherosclerosis compared to other mouse models. ^[30-32] However, this model has imperfections and other 446 models such as LDLr - have been considered. Nonetheless, our diets have a low cholesterol 447 content (<0.01% of diet weight) and it has been reported that long diets with higher 448 cholesterol contents are needed to induce significant lesion development in LDLr -/- mice. 449 ^[33,34] Consequently, the ApoE^{-/-} model appeared as the most appropriate to study the link 450 between lipid oxidation, endothelial function and atherosclerosis development with our diets. 451

The main results of our work with ApoE ^{-/-} mice showed that oxidative stress generated in the gastrointestinal tract (4-HNE) is correlated with an increased cardiovascular risk (endothelial dysfunction and atherosclerosis) and that preventing lipid oxidation could avoid these impairments.

Lipid oxidation products can be generated *in vivo* and the gastric tract has been proposed as the major site for diet-induced lipid oxidation. ^[35] This was confirmed in our study since we observed that enrichment of a Western diet with n-6 PUFA strongly increases the formation of 4-HNE during digestion, as shown by the higher recovery of 4-HNE in faeces from WD+SO group. This result is in line with previous studies reporting that a Western meal based on sunflower oil and beef led to the formation of TBARS, lipid-derived conjugated dienes and 4-HNE during gastric digestion in mini-pig. ^[36,37]

In order to prevent the deleterious effects of 4-HNE (oxidative stress, cell proliferation, or cell death), ^[38] we considered using an antioxidant strategy. Since, we previously reported that an apple phenolic extract and an apple matrix (Puree) were able to reduce lipid oxidation and 4-HNE formation in an *in vitro* model of digestion, ^[25] we tried to use both of them *in vivo*. By adding either the whole apple matrix (WD+SO+PUREE) or the corresponding phenolic extract (PE) to the diet, we managed to prevent lipid oxidation during digestion. We can 469 hypothesize that in our model, a reduction of the pro-oxidant hypervalent heme iron form (MbFe^{IV}=O) into metmyoglobin (MbFe^{III}) by chlorogenic acid as well as by monomeric and 470 471 oligomeric PCs may have occurred. This reduction inhibits the initiation phase of the lipid 472 oxidation process and has been previously reported as a major antioxidant effect of PCs and hydroxycinnamic acids in *in vitro* digestion. ^[5] A remarkable result is that both 473 474 WD+SO+PUREE and WD+SO+PE managed to prevent digestive lipid oxidation with the 475 same extent. This result could be explained by an increased polyphenol bioaccessibility in 476 WD+SO+PUREE group when compared to raw fruit that we have reported in a previous work.^[25] The relative similarity in polyphenol bioaccessibilities for both apple matrix and 477 478 phenolic extract may explain the very similar results obtained throughout this study for both 479 supplementations.

480 Next, we hypothesized that lipid oxidation products generated during digestion may have 481 an effect on vascular health (endothelial dysfunction, atherosclerosis) after their absorption. 482 Indeed, we report here that higher lipid oxidation in the digestive tract (4-HNE) is associated 483 with a higher plasmatic concentration of 4-HNE. These data are in accordance with results 484 from Keller et al. who observed 4-HNE in urine and various organs (liver, kidney, brain, 485 heart) after its ingestion.^[8] It further indicates that even if the majority of 4-HNE could be 486 excreted in faeces, a part of it is absorbed and can be implicated in vascular impairments. 487 Moreover, it has been shown that eating a fatty diet can alter the intestinal barrier physiological state by increasing its permeability ^[39] which could lead to an increased 488 489 absorption of 4-HNE.

490 In our work, fasting plasma concentration of 4-HNE after chronic WD+SO diet was 2.7 491 µM (0.43 mg/L). Such concentration of 4-HNE is known to trigger a variety of biological responses such as inflammation and cell proliferation.^[9] Moreover, this plasmatic level of 4-492 493 HNE is also known to induce oxidative stress in the vascular system. It is important to note 494 that 4-HNE values observed in our work were assessed after a prolonged fasting state and that 495 we can expect higher values in the postprandial state. Nevertheless, the increased oxidative 496 stress was confirmed here since the group with the higher 4-HNE concentration displays an 497 increase in ROS production and plasmatic malondialdehyde-modified oxLDL. Yet, an 498 increase in oxLDL production has been largely reported in the literature as a key step in the 499 development of vascular impairments such as endothelial dysfunction or atherosclerosis, which are generally described as the consequence of oxidative stress. ^[40,41] Moreover, it is 500 considered that high NO levels can account for nitro-oxidative stress in obese mice. ^[42] The 501

high levels of nitrites observed in the WD+SO group may be the results of the reaction between NO and superoxide to form peroxynitrite, a potent cytotoxic reactive species. Such a nitro-oxidative stress is largely reported as a key factor of vascular endothelial impairments and atherosclerosis. ^[43] Consequently, it was not surprising to observe the worst endothelial function and the largest atherosclerosis development in the group with the highest lipid oxidation.

508 We previously pointed out the deleterious effects of digestive lipid oxidation on vascular 509 integrity and the fact that both WD+SO+PUREE and WD+SO+PE diets were able to prevent 510 lipid oxidation. We therefore hypothesized that inhibition of lipid oxidation by polyphenol 511 consumption may prevent vascular impairments. This hypothesis was confirmed regarding the 512 decrease of ROS production and vascular impairments (reduced endothelial impairments and 513 atherosclerosis development). Surprisingly, vascular function was not only restored but even 514 positively enhanced in both supplemented groups. Few assumptions can be performed to 515 explain the beneficial effect of our diets on vascular function. In a first place, our protocol 516 does not allow us to exclude a cell-mediated effect of polyphenols included in the extract and 517 puree since these supplementation were not tested on a group with a standard diet. Indeed, a 518 direct effect of phase-II and colonic metabolites from proanthocyanidins on vascular function could not be excluded according to our results and the literature. ^[19,44,45]. Nonetheless, it has 519 520 been previously reported that 4-HNE was responsible for an impairment of Akt 521 phosphorylation, which is a main regulator of vascular function especially through eNOS phosphorylation on Ser1177.^[46] Despite eNOS phosphorylation on its activation site seems to 522 523 be increased in both WD+SO+PUREE and WD+SO+PE groups, the extent of this increase 524 was not satisfactory and certainly did not appear to be a key element to explain our results on 525 endothelial function. This idea is reinforced by the lack of effect of both PUREE and PE on 526 nitrites level when compared to the WD+SO group. Nevertheless, we have to consider that 527 even if nitrite levels are unchanged, a decrease of oxidative stress can result in a reduction of 528 NO reaction with O_2^- to form ONOO⁻. Thus, considering that both PUREE and PE treatment 529 reduced or tended to reduce ROS production in the vascular wall, we can hypothesize that the 530 bioavailability of NO may be increased. This hypothesis is supported by the impact of our 531 experimental conditions on endothelial vasodilatation to ACh. Indeed, on aortic tissue 532 endothelium-dependent vasodilatation is almost exclusively dependent of the eNOS-NO 533 pathway, since the pre-incubation of L-NAME almost completely abolishes it in aortas of 534 ApoE^{-/-} mice. ^[47–49] Further studies, evaluating eNOS coupling, NO bioavailability and the 535 level of nitro-oxidative stress will be needed to confirm this point.

These results are in accordance with numerous studies reporting the interest of a chronic uptake of apple polyphenols to reduce risk factors (arterial pressure, blood lipid profile, inflammation, oxidative stress) and cardiovascular diseases (anti-angiogenic, anti-atherogenic and anti-hypertensive) in rodents or humans. ^[45,50–53] However, there is no study as this time that are able to link these effects with a potential antioxidant effect during digestion.

541

542 CONCLUSION

543 In conclusion, a Western diet rich in n-6 PUFA appears to exacerbate lipid oxidation 544 during digestion leading to the formation of 4-HNE among other lipid oxidation products. The 545 absorption of these products is associated with plasmatic oxidative stress, endothelial dysfunction and development of atherosclerosis in the ApoE^{-/-} mice model. The involvement 546 of digestive lipid oxidation in these phenomena was confirmed since the inhibition of lipid 547 548 oxidation and, consequently, 4-HNE synthesis in the gastrointestinal tract, by a 549 supplementation in apple polyphenols under two different matrix forms, remarkably prevents 550 those impairments. Consequently, the consumption of proanthocyanidin-rich fruits such as 551 apple or pear should be encouraged to attenuate the deleterious effect of the Western diet.

552

553

554 **Financial support**

555 The study was funded by the Foundation of Avignon University and SFR Tersys. This work 556 was supported by the Platform 3A, funded by the European Regional Development Fund, the 557 French Ministry of Research Higher Education and Innovation, the Provence-Alpes-Côte-558 d'Azur Region, the Departmental Council of Vaucluse and the Urban Community of Avignon 559

560 Acknowledgements

561 Gaëtan Boléa acknowledges the foundation of Avignon University for PhD grant. We 562 gratefully thank Mrs Christine Boutin (Bio verger, Ambricourt, France) for the kind gift of 563 apple *Reinette de Flandre*. 564

565 **Disclosures**

566 Authors declare that they have no conflict of interest. Claire Dufour and Gregory Meyer 567 designed the study protocols and secured funding.

568

569 Author Contributions

570 G.B.; C.P.; A.H.; M.D.; C.G.; S.R.; A-L.C. B.G. G.M. performed experiments. C.A. 571 developed ApoE^{-/-} model and histological protocols. G.B., G.M., C.D. designed the scope, 572 analyzed the data and wrote the manuscript. C.R., C.A. and B.G. revised the manuscript. All 573 authors read and approved the final manuscript.

574 **REFERENCES**

- 575
- 576 [1] J.-J. Chiu, S. Chien, *Physiol. Rev.* **2011**, *91*, 327–387.
- 577 [2] R. Ross, Am. Heart J. 1999, 138, S419–S420.
- 578 [3] G. K. Hansson, A.-K. L. Robertson, C. Söderberg-Nauclér, *Annu. Rev. Pathol. Mech.*579 *Dis.* 2006, *1*, 297–329.
- 580 [4] J. Kanner, Mol. Nutr. Food Res. 2007, 51, 1094–1101.
- 581 [5] B. Lorrain, O. Dangles, M. Loonis, M. Armand, C. Dufour, J. Agric. Food Chem. 2012,
 582 60, 9074–9081.
- 583 [6] E. Vulcain, P. Goupy, C. Caris-Veyrat, O. Dangles, *Free Radic. Res.* 2005, *39*, 547–584
 563.
- 585 [7] S. J. Chapple, X. Cheng, G. E. Mann, *Redox Biol.* **2013**, *1*, 319–331.
- 586 [8] J. Keller, M. Baradat, I. Jouanin, L. Debrauwer, F. Guéraud, *Redox Biol.* 2015, *4*, 136–
 587 148.
- 588 [9] A. Nègre-Salvayre, S. Garoby-Salom, A. Swiader, M. Rouahi, M. Pucelle, R. Salvayre,
 589 *Free Radic. Biol. Med.* 2017, *111*, 127–139.
- 590 [10] S. Gargiulo, G. Testa, P. Gamba, E. Staurenghi, G. Poli, G. Leonarduzzi, *Free Radic*.
 591 *Biol. Med.* 2017, *111*, 140–150.
- 592 [11] I. Staprans, X.-M. Pan, J. H. Rapp, K. R. Feingold, *Mol. Nutr. Food Res.* 2005, 49, 1075–1082.
- 594 [12] D. Aune, E. Giovannucci, P. Boffetta, L. T. Fadnes, N. Keum, T. Norat, D. C.
 595 Greenwood, E. Riboli, L. J. Vatten, S. Tonstad, *Int. J. Epidemiol.* 2017, 46, 1029–1056.

- 596 [13] J. Zhan, Y.-J. Liu, L.-B. Cai, F.-R. Xu, T. Xie, Q.-Q. He, *Crit. Rev. Food Sci. Nutr.*597 2017, 57, 1650–1663.
- 598 [14] L. Hooper, C. Kay, A. Abdelhamid, P. A. Kroon, J. S. Cohn, E. B. Rimm, A. Cassidy,
 599 Am. J. Clin. Nutr. 2012, 95, 740–751.
- 600 [15] A. L. Macready, T. W. George, M. F. Chong, D. S. Alimbetov, Y. Jin, A. Vidal, J. P. E. 601 Spencer, O. B. Kennedy, K. M. Tuohy, A.-M. Minihane, M. H. Gordon, J. A. Lovegrove,
- 602 FLAVURS Study Group, Am. J. Clin. Nutr. **2014**, 99, 479–489.
- 603 [16] C. C. Lee, J. H. Kim, J. S. Kim, Y. S. Oh, S. M. Han, J. H. Y. Park, K. W. Lee, C. Y.
 604 Lee, *Int. J. Mol. Sci.* 2017, *18*, 1363.
- 605 [17] S. Claude, C. Boby, A. Rodriguez-Mateos, J. P. E. Spencer, N. Gérard, C. Morand, D.
 606 Milenkovic, *Mol. Nutr. Food Res.* 2014, 58, 1016–1027.
- 607 [18] C. D. Kay, L. Hooper, P. A. Kroon, E. B. Rimm, A. Cassidy, *Mol. Nutr. Food Res.*608 2012, 56, 1605–1616.
- 609 [19] R. Sansone, A. Rodriguez-Mateos, J. Heuel, D. Falk, D. Schuler, R. Wagstaff, G. G. C.
- 610 Kuhnle, J. P. E. Spencer, H. Schroeter, M. W. Merx, M. Kelm, C. Heiss, for the Flaviola
- 611 Consortium, European Union 7th Framework Program, Br. J. Nutr. 2015, 114, 1246–1255.
- 612 [20] J. Pérez-Jiménez, L. Fezeu, M. Touvier, N. Arnault, C. Manach, S. Hercberg, P. Galan,
- 613 A. Scalbert, Am. J. Clin. Nutr. 2011, 93, 1220–1228.
- 614 [21] C. Dufour, M. Loonis, M. Delosière, C. Buffière, N. Hafnaoui, V. Santé-Lhoutellier, D.
 615 Rémond, *Food Chem.* 2018, 240, 314–322.
- 616 [22] G. Boléa, C. Ginies, M.-J. Vallier, C. Dufour, Food Funct. 2019, 10, 3942–3954.
- 617 [23] B. Lorrain, O. Dangles, C. Genot, C. Dufour, J. Agric. Food Chem. 2010, 58, 676–683.
- 618 [24] S. Battault, F. Singh, S. Gayrard, J. Zoll, C. Reboul, G. Meyer, *Hypertens. Res.* 2016,
 619 39, 70–78.
- 620 [25] G. Boléa, C. Ginies, M.-J. Vallier, C. Dufour, Food Funct. 2019, 10, 3942–3954.
- [26] Guyot, S., Le Bourvellec, C., Marnet, N., and Drilleau, J.F., *Procyanidins Are the Most Abundant Polyphenols in Dessert Apples at Maturity.*, 2002.
- 623 [27] D. E. Venegas-Pino, N. Banko, M. I. Khan, Y. Shi, G. H. Werstuck, J. Vis. Exp. 2013,
 624 50933.
- 625 [28] V. Victor, Curr. Pharm. Des. 2009, 15, 2986–2987.
- 626 [29] S. Oppi, T. F. Lüscher, S. Stein, Front. Cardiovasc. Med. 2019, 6, 46.
- 627 [30] A. S. Plump, J. D. Smith, T. Hayek, K. Aalto-Setälä, A. Walsh, J. G. Verstuyft, E. M.
 628 Rubin, J. L. Breslow, *Cell* 1992, *71*, 343–353.
- 629 [31] Y. Nakashima, A. S. Plump, E. W. Raines, J. L. Breslow, R. Ross, *Arterioscler*.
 630 *Thromb. J. Vasc. Biol.* **1994**, *14*, 133–140.
- 631 [32] J. D. Smith, J. L. Breslow, J. Intern. Med. 1997, 242, 99–109.
- 632 [33] Y. T. Lee, H. Y. Lin, Y. W. F. Chan, K. H. C. Li, O. T. L. To, B. P. Yan, T. Liu, G. Li,
- 633 W. T. Wong, W. Keung, G. Tse, *Lipids Health Dis.* 2017, 16, 12.
- 634 [34] G. S. Getz, C. A. Reardon, Arterioscler. Thromb. Vasc. Biol. 2012, 32, 1104–1115.

- 635 [35] J. Kanner, T. Lapidot, Free Radic. Biol. Med. 2001, 31, 1388–1395.
- 636 [36] M. Delosière, V. Santé-Lhoutellier, C. Chantelauze, D. Durand, A. Thomas, C. Joly, E.
- Pujos-Guillot, D. Rémond, B. Comte, C. Gladine, A. Guy, T. Durand, M. Laurentie, C. Dufour, *Food Funct.* 2016, 7, 3497–3504.
- 639 [37] M. Gobert, D. Rémond, M. Loonis, C. Buffière, V. Santé-Lhoutellier, C. Dufour, *Food*640 *Funct.* 2014, *5*, 2166.
- 641 [38] S. Dalleau, M. Baradat, F. Guéraud, L. Huc, Cell Death Differ. 2013, 20, 1615–1630.
- 642 [39] S. Masumoto, A. Terao, Y. Yamamoto, T. Mukai, T. Miura, T. Shoji, *Sci. Rep.* 2016, 6,
 643 31208.
- 644 [40] D. Gradinaru, C. Borsa, C. Ionescu, G. I. Prada, *Mech. Ageing Dev.* 2015, 151, 101–
 645 113.
- [41] Y. Steffen, G. Vuillaume, K. Stolle, K. Roewer, M. Lietz, J. Schueller, S. Lebrun, T.
 Wallerath, *Nitric Oxide* 2012, 27, 176–184.
- 648 [42] A. Kleindienst, S. Battault, E. Belaidi, S. Tanguy, M. Rosselin, D. Boulghobra, G.
- 649 Meyer, S. Gayrard, G. Walther, B. Geny, G. Durand, O. Cazorla, C. Reboul, Basic Res.
- 650 *Cardiol.* **2016**, *111*, 40.
- [43] C. R. White, T. A. Brock, L. Y. Chang, J. Crapo, P. Briscoe, D. Ku, W. A. Bradley, S.
 H. Gianturco, J. Gore, B. A. Freeman, *Proc. Natl. Acad. Sci.* **1994**, *91*, 1044–1048.
- [44] L. Hooper, P. A. Kroon, E. B. Rimm, J. S. Cohn, I. Harvey, K. A. Le Cornu, J. J. Ryder,
 W. L. Hall, A. Cassidy, *Am. J. Clin. Nutr.* **2008**, 88, 38–50.
- 655 [45] M.-H. Oak, C. Auger, E. Belcastro, S.-H. Park, H.-H. Lee, V. B. Schini-Kerth, *Free* 656 *Radic. Biol. Med.* **2018**, *122*, 161–170.
- [46] I. Nakashima, W. Liu, A. A. Akhand, K. Takeda, Y. Kawamoto, M. Kato, H. Suzuki,
 Mol. Aspects Med. 2003, 24, 231–238.
- [47] M. O. R. Mian, N. Idris-Khodja, M. W. Li, A. Leibowitz, P. Paradis, Y. Rautureau, E.
 L. Schiffrin, J. Pharmacol. Exp. Ther. 2013, 347, 30–37.
- 661 [48] L. V. d'Uscio, L. A. Smith, Z. S. Katusic, *Stroke* **2001**, *32*, 2658–2664.
- 662 [49] C. M. Balarini, M. A. Leal, I. B. S. Gomes, T. M. C. Pereira, A. L. Gava, S. S.
 663 Meyrelles, E. C. Vasquez, *J. Transl. Med.* 2013, *11*, 3.
- 664 [50] C. M. Andre, J. M. Greenwood, E. G. Walker, M. Rassam, M. Sullivan, D. Evers, N. B.
 665 Perry, W. A. Laing, *J. Agric. Food Chem.* 2012, *60*, 10546–10554.
- [51] D. Aune, E. Giovannucci, P. Boffetta, L. T. Fadnes, N. Keum, T. Norat, D. C.
 Greenwood, E. Riboli, L. J. Vatten, S. Tonstad, *Int. J. Epidemiol.* 2017, 46, 1029–1056.
- 668 [52] D. A. Hyson, *Adv. Nutr.* **2011**, *2*, 408–420.
- 669 [53] A. Koutsos, K. Tuohy, J. Lovegrove, *Nutrients* **2015**, *7*, 3959–3998.

670

672 Figure Legends

Figure 1. Effects of a Western diet with or without supplementation on weight gain and plasma lipoproteins. A: Mean of daily food intake during 12 weeks per mice. B: Follow up of body mass during the 12 weeks of different diets. C: Adiposity index at the end of the protocol diets. Plasma HDL (D), LDL (E), HDL-cholesterol/LDL-cholesterol (F), total cholesterol (G) and triglyceride (H), were assessed at the end of the dietary protocols. Values are expressed as means \pm SEM (n=6/8 per group for LDL, HDL, HDL-c/LDL-c, total cholesterol and triglyceride). One way ANOVA (Tukey's multiple comparisons test).

680

Figure 2. 4-Hydroxy-2-nonenal (4-HNE) formation during gastrointestinal digestion and lipid
oxidation products in plasma. 4-HNE concentration in faeces before (A), at 6 weeks (B) and 12
weeks (C) of diet measured by LC/MS/APCI. Plasma 4-HNE (D) and oxLDL (E) at the end of
the 12 weeks. Values are expressed as means ± SEM (plasma 4-HNE n=5/7 per grp, oxLDL
n=6/7 per grp). *P<0.05 vs. WD. #P<0.05 vs. WD+SO. §P<0.05 vs. WD. \$P<0.05 vs.
WD+SO+PUREE. One way ANOVA (Tukey's multiple comparisons test).

687

688 Figure 3. Effects of Western diets and protection by apple polyphenol supplementation on 689 vascular function. Systolic (A), diastolic (B) and mean (C) blood pressure measured at the end 690 of dietary protocols by tail cuff method. D: Dose-dependent response to cumulative doses of 691 acetylcholine (ACh) and sodium nitroprusside (SNP) on aortic rings. Left: Dose-dependent 692 response to cumulative doses of ACh. Middle: Maximal relaxation of aortic rings to ACh. 693 Right: Dose-dependent response to cumulative doses of sodium nitroprusside (SNP). Data are 694 expressed in percent of relaxation relative to maximal contraction obtained with 1 µM of 695 phenylephrine. Values are expressed as means \pm SEM (blood pressure n=9/11 per group, 696 vasoreactivity n=6/8 per group). *P<0.05 vs. WD group. #P<0.05 vs. WD+SO group. §P<0.05 697 vs. WD. \$P<0.05 vs. WD+SO+PUREE. £P<0.05 vs. Week 0. Dose-response were analyzed 698 with a repeated measures ANOVA followed with a Tukey multiple comparisons test. Blood 699 pressures and maximal relaxation were analyzed using a one way ANOVA (Tukey's multiple 700 comparisons test.)

701

702 Figure 4. Development and characterization of atheromatous plaque size induced by Western 703 diets and protection by apple polyphenol supplementation. (A) Evaluation of aortic cross 704 atheromatous plaque size performed by echography. (B) Atheromatous plaque size in 705 descending thoraco-abdominal aorta assessed by Oil-Red-O coloration. (C) Atheromatous 706 plaque size in aortic root relative to aortic diameter. (D) Percent of fibrosis in atheromatous 707 plaque of aortic valve and (E) Necrotic core size in atheromatous plaque of aortic root relative 708 to aortic diameter assessed on histological sections. All measurements were performed at the 709 end of dietary protocol. Values are expressed as means \pm SEM (echography n=8 per group, 710 histological sections n=5 animals per group). One way ANOVA (Tukey's multiple 711 comparisons test.) *P<0.05 vs. WD group. #P<0.05 vs. WD+SO group. §P<0.05 vs. WD. 712 \$P<0.05 *vs*. WD+SO+PUREE.

713

Figure 5. Dietary effects on vascular NO and ROS production. (A) Quantification of nitrite in aorta. (B) Total ROS production measured by electron paramagnetic resonance in aortic tissue

- homogenate. Values are expressed as means ± SEM (nitrite and ROS production n=4/grp).
 *P<0.05 vs. WD group. #P<0.05 vs. WD+SO group. \$P<0.05 vs. WD. \$P<0.05 vs.
 WD+SO+PUREE. ROS and Nitrite productions were analyzed using a one way ANOVA (Tukey's multiple comparisons test.)

D



















FIGURE 2





FIGURE 3







