

# Prenatal maternal vitamin D deficiency sex-dependently programs adipose tissue metabolism and energy homeostasis in offspring

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1	Prenatal maternal vitamin D deficiency sex-dependently programs adipose tissue
2	metabolism and energy homeostasis in offspring
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- 25 List of nonstandard abbreviations :
- 26
- 27 25(OH)D: 25-hydroxyvitamin D
- 28 AGRP: Agouti related-peptide
- 29 AUC: Area under the curve
- 30 CTRL: Control
- 31 EE: Energy expenditure
- 32 GOX: Carbohydrate oxidation
- 33 HF: High fat
- 34 HOMA-IR: Homeostasic model assessment of insulin resistance
- 35 IPA: Ingenuity pathway analysis
- 36 ITT: Insulin tolerance test
- 37 LF: Low fat
- 38 LOX: Lipid oxidation
- 39 MC4R: Melanocortin receptor type 4
- 40 NPY: Neuropeptide Y
- 41 RQ: Respiratory quotient
- 42 VDD: Vitamin D deficiency
- 43

#### 44 Abstract

In utero environment is crucial to ensure normal development of the foetus and to program 45 metabolic health throughout the life. Beside macronutrients, the role of micronutrients, 46 including vitamin D, begins to be explore. The aim of this study was to decipher the impact of 47 48 maternal vitamin D deficiency (VDD), in normal and high fat (HF) diet context, on adipose tissue metabolism and energy homeostasis in offspring, considering sex-specific responses. 49 50 Body weight, energy expenditure and spontaneous activity was differential impacted in juvenile male and female offspring born from VDD mice. In adulthood, a HF diet combined 51 52 with maternal VDD disrupted glucose homeostasis and adiposity in male offspring but not in females. Such phenotypes were associated to different transcriptomic profiles in adipose 53 54 tissue, that could be related to differential modulation of plasma 17β-estradiol concentrations. Thus, maternal VDD sex-dependently modulated metabolic fate of the offspring, especially 55 56 when associated with HF diet in adulthood.

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58 Keywords: Vitamin D, Maternal vitamin D deficiency, offspring, adipose tissue, metabolism.59

#### 60 Introduction

61

Vitamin D is an essential micronutrient that is suspected to display an important role on 62 normal foetal development, and throughout the life of the child and future adult. Its role in 63 calcium and phosphate metabolism is well-established and it also appears to be involved in 64 many other physiological processes<sup>1</sup>, including control of adipose tissue biology<sup>2-9</sup>. 65 Consequently, vitamin D insufficiency is considered as a risk factor in several pathologies, 66 including auto-immune diseases, musculoskeletal defects<sup>10</sup>, cardiovascular and metabolic 67 diseases. Such insufficiency is characterised by plasma concentration in 25-hydroxyvitamin D 68 (25(OH)D) below the cut-off value of 50nmol/L<sup>11</sup>. In fact, vitamin D insufficiency has 69 become a large public health issue<sup>12,13</sup> and is common for women in child bearing age, 70 pregnant and breastfeeding women. Currently, 54% of pregnant women and 75% of new-71 borns present a 25(OH)D status < 50nmol/L, also 18% of pregnant women and 29% of new-72 borns present a severe vitamin D deficiency  $(25(OH)D < 25nmol/L)^{14,15}$ . It is noteworthy that 73 maternal vitamin D deficiency (VDD) was associated with pregnancy, foetal and neonatal 74 outcomes<sup>15</sup> such as increased risk factor for preeclampsia<sup>16</sup>, gestational diabetes mellitus<sup>17</sup>, 75 higher risk of small-for-gestational-age, reduced term birth weight, and lower head 76 circumference<sup>18</sup>, even if these results are sometimes controverted<sup>19</sup>. 77

78 Interestingly, recent studies linked maternal vitamin D insufficiency to overweight and increased fat mass in offspring. Indeed, in the INMA cohort, including 2358 pregnant women, 79 maternal VDD was associated with increased risk of overweight in foetal and early postnatal 80 (1-year-old) offspring<sup>20</sup>. Similarly, in the prospective cohort study Southampton Women's 81 survey (977 pregnant women), maternal VDD was associated with lower fat mass at birth and 82 greater fat mass at 4 and 6 years<sup>21</sup>. These results were confirmed in the prospective cohort 83 Rhea (532 pregnant women), where the maternal 25(OH)D concentrations < 37.7 nmol/L 84 were associated with higher body mass index and central adiposity in the 4 and 6-years-old 85 offspring<sup>22</sup>. Nevertheless, in a long term follow up (20 years), no association between 86 maternal vitamin D status and cardio-metabolic risk factors was highlighted<sup>23</sup>. Thus, those 87 associations remain controversial. 88

To gain further insight on the relationship, several preclinical studies have been implemented. Overall, it has been established that maternal VDD as potential programming long-term effect on metabolic health<sup>24,25</sup>, nevertheless several important points remain pending. Based on the Developmental origins of Health and disease (DOHaD concept), we hypothesized that the response to prenatal maternal VDD may be sex-specific and influenced by the nutritional environment during adulthood. Indeed, the maternal environment during the pre- and periconceptional period, especially nutrition, can modify epigenetic marks, leading to long term
phenotypic consequences that may vary according to the sex and the environment of the
offspring<sup>26</sup>.

98 Thus, the aim of the present study is to determine the impact of maternal VDD combined with 99 the impact of obesogenic environment, induced by a high-fat diet, on the sex-specific

100 response during adulthood on energy homeostasis and adipose tissue metabolism.

101

- **103** Material and Methods
- 104

#### 105 Animal Experiments

106 The protocol received the agreement of Aix-Marseille University Ethics Committee and the 107 French Ministry of Research (APAFIS#1300-2015072112279135). Eight-week-old female 108 and male C57BL/6JRJ mice were obtained from Janvier Labs (Le Genest-Saint-Isle, France), fed ad libitum with control food (chow diet A04 from Safe-diets, Augy France) during the 1-109 week acclimation period and with full access to drinking water. The animals were maintained 110 111 at 22°C under a 12-hour light, 12-hour dark cycle and a 20% humidity level. Female mice (15 112 per group) were randomly assigned into one of the two experimental groups depending on the 113 diet i.e. control (AIN-93G with vitamin D3, 1.0 IU/g) or vitamin D-depleted (AIN-93G without vitamin D3, 0.0 IU/g) for eight-weeks (Supplemental Figure 1), and were mated with 114 115 males. Weight gain was measured once a week and dietary at 3-weeks of pre-mate diet, at 5 days and 15 days of gestational stage (Supplemental Figure 2). After delivery, all females 116 117 were fed with control diet (AIN-93G) until weaning of the offspring. The litter size was adjusted to 6 pups per females. The body weight of the offspring was evaluated weekly from 118 119 the weaning until the study end, and not prior weaning to avoid maternal cannibalization and perinatal stress. At six-weeks of age both males and females of the offspring were randomly 120 assigned to receive Low Fat diet (AIN-93M Maintenance Purified Diet) or High Fat diet (DIO 121 Rodent Purified Diet w/45% Energy from Fat) for eight weeks. At the end of the protocol, 122 mice were subjected to food restriction overnight and blood was collected by cardiac puncture 123 anesthesia, serum was isolated by centrifugation at 3000 rpm for 15 min at 4°C and was 124 stored at -80°C. Animals were euthanized by cervical dislocation and various tissue (liver, 125 spleen, hypothalamus and various white adipose tissue deposits) were collected, weighted and 126 stored at -80°C. Eight groups of offspring mice (males and females) were designed to study 127 the impact of maternal diet (CTRL vs VDD) and adult diet (LF vs HF). 128

129

#### 130 Biochemical analysis

To confirm the maternal and offspring vitamin D status, 25(OH)D serum concentration were
measured using an in vitro diagnostic enzyme immunoassay kit 25-OH Vitamin D (direct)
ELISA kit (PromoKine). Insulin, leptin, testosterone and 17β-estradiol were measured in
plasma using an enzyme-linked immuno-sorbent assay ELISA (Insulin ALPCO Diagnostics,
New Hamshire, United States; DuoSet mouse leptin, R&D systems, Minneapolis, United
States, Testosterone Demeditec Diagnostics GmbH, Germany; ab108667, Abcam, Cambridge,

England, respectively). The manufacturer's protocols were followed. For glucose concentration, mice were subjected to food restriction for 5h and glycemia was measured from tail blood (Accu-Check glucometer, Roche). The HOMA-IR index was calculated according to the following formula: fasting insulin (microU/L) x fasting glucose (nmol/L)/22.5.

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#### 143 Histological analysis

144 Visceralal adipose tissue samples were fixed in 10% buffered formalin, embedded in paraffin 145 and sliced to prepare 5  $\mu$ m tissue sections whose were stained with hematoxylin and eosin 146 (H&E). The images were captured by a light microscope (EZAD, Leica, Germany; 10X 147 magnification). The adipocyte area ( $\mu$ m<sup>2</sup>) were determined using (Image J) software.

148

#### 149 Indirect calorimetry

At 5 and 12 weeks old the offspring was acclimated and kept for 24h in an indirect 150 calorimetric cage (Physiocage, Bioseb, Vitrolles, France), as previously described<sup>6</sup>. The 151 calorimetric appliance was composed of a gas analyzer (to measure O<sub>2</sub> consumption and CO<sub>2</sub> 152 153 production as VO<sub>2</sub> and VCO<sub>2</sub>) and an activity recorder (locomotion and rearing). The temperature of the calorimetric room was set to 22°C. Four were connected to each gas 154 analyzer, but each cage had specific inlets and outlets. A constant inlet flow (5 cm<sup>3</sup>/min) was 155 maintained throughout the experiment. Gases were continuously analyzed with the following 156 sequence: 3 min from cage 1, 3 min from cage 2, 3 minutes from cage 3, 3 minutes from cage 157 4 and then 3 min from room air and thus the volume (mL/min) of O<sub>2</sub> consumed (VO<sub>2</sub>) and 158  $CO_2$  produced (VCO<sub>2</sub>) were measured for each mouse. Energy expenditure (EE) was 159 calculated as following (EE=  $(16.3 \times \text{VO2} + 4.57 \times \text{VCO2})/60$  (watt)). Lipid (LOX) and 160 carbohydrate (GOX) oxidation were calculated according to the following equations: LOX = 161  $(1.69 \times VO_2 - 1.69 \times VCO_2) \times (9.46 \times 4.186/60)$  (watt) and GOX =  $(4.57 \times VCO_2 - 1.69 \times VCO_2) \times (9.46 \times 4.186/60)$  (watt) and GOX =  $(4.57 \times VCO_2 - 1.69 \times VCO_2) \times (9.46 \times 4.186/60)$  (watt) and GOX =  $(4.57 \times VCO_2) \times (9.46 \times 4.186/60)$  (watt) and GOX =  $(4.57 \times VCO_2) \times (9.46 \times 4.186/60)$  (watt) and GOX =  $(4.57 \times VCO_2) \times (9.46 \times 4.186/60)$  (watt) and GOX =  $(4.57 \times VCO_2) \times (9.46 \times 4.186/60)$  (watt) and GOX =  $(4.57 \times VCO_2) \times (9.46 \times 4.186/60)$  (watt) and GOX =  $(4.57 \times VCO_2) \times (9.46 \times 4.186/60)$  (watt) and GOX =  $(4.57 \times VCO_2) \times (9.46 \times 4.186/60)$  (watt) and GOX =  $(4.57 \times VCO_2) \times (9.46 \times 4.186/60)$  (watt) (wa 162  $3.23 \times VO_2$  × (3.74×4.186/60) (watt)<sup>27</sup>. Total activity was evaluated by summing 163 spontaneous activity and rearing activity, measured in indirect calorimetric cages 164 (Physiocage, Bioseb, Vitrolles, France), and was normalized to the control value. 165

166

#### 167 Insulin Tolerance Test (ITT)

168 One week before the end of the protocol, mice were subjected to ITT. Mice were subjected to 169 food restriction for 5h and stuffed with an insulin solution (0.05 UI/mL) prepared in saline (0.5 UI/ kg body weight). Glycemia was measured from tail blood at 0, 15, 30, 60, 90, 120
minutes after injection (Accu-Check glucometer, Roche Diagnostic, Meylan, France).

172

#### 173 RNA extraction real time PCR and RNA sequencing

Total RNA was extracted from retroperitoneal adipose tissue or hypothalamus using TRIzol 174 reagent (Thermo Fischer Scientific, Les Ulis, France). For the real time PCR, one µg of total 175 RNA from hypothalamus was used to synthetize cDNAs using random primers and Moloney 176 murine leukemia virus reverse transcriptase (Thermo Fischer Scientific, Les Ulis, France). 177 Real-time quantitative PCR analyses were performed using the Mx3005P Real-Time PCR 178 System (Stratagene, La Jolla, USA) as previously described<sup>28</sup>. For each condition, expression 179 was quantified in duplicate, and 18S rRNA was used as the endogenous control in the 180 comparative cycle threshold (CT) method<sup>29</sup>. 181

182 The sequences of the primers used in this study are reported in supplemental data183 (Supplemental Table 1).

For the RNA sequencing, total RNA was isolated from 3 mice per group and was used for the
RNA-seq library preparation, using the kit TruSeq Stranded mRNA by Illumina.

Libraries were paired-end sequenced on the Illumina NextSeq 500 sequencer. Reads with a phred score lower than 20 and shorter than 25 bp were removed using Sickle (v1,33). Quality of trim reads were checked using multiQC (v1.0). Trim reads were aligned using STAR aligner (v2.7.0d) with arguments "outFilterMismatchNoverLmax" and "outFilterMultimapNmax" set to 0.08 and 1, respectively.

Transcripts discovery was performed using Cufflinks (v2.2.1) with the "library-type" 191 argument set to fr-firstrand, and a GTF file obtained from GENCODE ("Comprehensive gene 192 annotation", vM1) provided as the genomic annotation. The GTF files produced for each 193 sample by Cufflinks were combined using Cuffmerge. The "class code" assigned to each 194 195 transcript by Cuffmerge was used to defined unknown transcripts (class code"u"). Only de novo transcripts with counts greater than 0 in at least one RNA-seq sample were kept for 196 197 subsequent analyses. These de novo transcripts were combined with the GENCODE GTF file to produce the final genomic annotation that was provided to FeatureCounts (v1.6.1) for 198 quantification. 199

- 200 Differential gene expression was performed using DESEQ2 between conditions. To create
- bigwig files, reads from Watson and Crick strands were selected using SAMtools (v1.9) and
- provided to the bam2wig.py script from the RseQC program suite (v2.6.4). RNA-seq profiles
- 203 were visualized using the IGV genome browser.

204

#### 205 Ingenuity Pathway Analysis (IPA)

Differential gene expression (with p-value adjusted < 0.05), obtained from the RNA seq analysis between our conditions, were used in the IPA software to identify the canonical pathways differentially impacted by the maternal diet.

209

#### 210 Statistical analysis

211 Data are expressed as mean  $\pm$  SEM. Significant differences were determined by unpaired 212 Student's *t* test or by ANOVA followed by the Fisher's LSD post hoc test using GraphPad 213 Prism. *p* < 0.05 was considered to be statistically significant.

214

- 216 **Results**
- 217

#### 218 Maternal vitamin D deficiency affects post-weaning energy metabolism of offspring.

Maternal VDD diet consumption prior and during gestation led to significant smaller post-219 220 weaning (5 weeks old) body weight in male mice offspring (Fig.1A), that was non-significant in female offspring (Fig.1H). The food consumption remained the same between CTRL and 221 VDD, either for males or females (Fig. 1B, 1I). In VDD males, spontaneous activity, 222 respiratory quotient (RQ) and energy expenditure (EE) were increased (Fig. 1C, D, E) 223 compared with the control group (p < 0.05). No difference of lipid oxidation (LOX) was 224 noticed in male (Fig. 1F), but carbohydrate oxidation (GOX) was increased in VDD males 225 (Fig. 1G). For female, despite the increased activity and RQ (Fig. 1J, K) of the VDD group, 226 there was no statistical difference for the EE (Fig. 1L). This was associated with decreased 227 228 LOX and increased GOX (Fig. 1M, N).

229

# Maternal vitamin D deficiency combined with HF diet affects energy metabolism in adult offspring.

232 To explore the combined effect of maternal VDD and post-weaning nutrition, mice were 233 submitted to low fat (LF) or high fat (HF) diet, leading thus to 8 groups (CTRL-LF, CTRL-HF, VDD-LF and VDD-HF, for both male and female). Firstly, plasma 25(OH)D 234 concentration was measured in 6 weeks old offspring. Only VDD females presented 235 decreased 25(OH)D values (108.6nmol/L  $\pm$  2.852) compared with the CTRL (128.9nmol/L  $\pm$ 236 6.186; p < 0.01) (Fig. 2A, B). During the diet period (from week 6 to 12), no statistical 237 difference between CTRL-LF and VDD-LF males body weight was observed (except at 9 238 weeks old, p < 0.05; Fig. 2C). Under HF diet, the body weight of CTRL males was gradually 239 higher than VDD males (significant from week 9 to 12; Fig. 2C). Concerning females, there 240 was non-significant differences between CTRL-LF and VDD-LF (from week 6 to 12). Under 241 HF diet, the CTRL females showed increased body weight compared with the VDD 242 243 (significant from week 9 to 12; Fig. 2D).

The energy metabolism was explored by indirect calorimetry (performed at 12-weeks of age). No modification of the energy metabolism of VDD offspring (*i.e.* body weight, activity, RQ, EE, LOX and GOX) was observed compared with the CTRL offspring when exposed to LF diet in adulthood, either for male and female (Supplemental Figure 3). Under HF diet, energy metabolism of VDD offspring was disturbed when compared with CTRL (Fig. 2). Males exposed to VDD associated-HF diet displayed a smaller body weight than CTRL-HF (p <

- 250 0.05) and increased food consumption, EE and GOX (p < 0.05) (Fig. 2E, F, I, J). No 251 modification of activity, RQ and LOX was observed in males. Female exposed to VDD-HF 252 diets also exhibited smaller body weight than CTRL-HF (Fig. 2L). The food intake tended to 253 be increased (non-significant) in the VDD-HF compared with the CTRL-HF, whereas the 254 activity decreased and EE and GOX increased (p < 0.05) (Fig. 2M, N, P, R). No modification 255 of RQ and LOX was observed in females.
- In order to explain the disruption of feeding behaviour in VDD group, we investigated the 256 expression of the genes of hypothalamic melanocortin pathway known to be involved in the 257 regulation of food intake<sup>30</sup>. For males, exposed to LF diet in adulthood, agouti related-peptide 258 (AGRP) and melanocortin receptor type 4 (MC4R) were under-expressed in the VDD 259 260 compare to the CTRL (Supplemental Figure 4A). No statistical differences were observed in 261 HF condition (Supplemental Figure 4B). For females in VDD and CTRL-LF there was not 262 statistical difference (Supplemental Figure 4C). Under HF diet, Neuropeptide Y (NPY) and AGRP were overexpressed (p < 0.05) and MC4R under-expressed in VDD-HF compare to 263 264 CTRL HF (Supplemental Figure 4D). There is no difference in POMC expression between the different groups (Supplemental Figure 4A-D). 265
- In order to explain energy metabolism sex discrepancies, the  $17\beta$ -estradiol and testosterone plasma concentrations were evaluated in all groups (Table 1). No significant modification was observed in male groups (CTRL-LF, CTRL-HF, VDD-LF, VDD-HF) for  $17\beta$ -estradiol and testosterone. In females, no difference was observed in CTRL condition, whereas in VDD condition,  $17\beta$ -estradiol was strongly increased under HF diet (Table 1). No modification of testosterone was observed.
- 272

#### 273 Maternal VDD and HF diet-associated affect glucose homeostasis sex-specifically

Biochemical analysis of male offspring highlighted that the glycemia tended to be higher in both CTRL-HF and VDD-HF groups when compared with LF groups (performed at 13 weeks; Fig. 3A). Similar pattern was observed for plasma insulin, with an increased concentration for the VDD-HF compared with the CTRL-HF (p < 0.05) (Fig. 3B). Thus, the VDD-HF males presented the higher values of plasma insulin, glycemia and HOMA-IR when compared with CTRL-HF, and to CTRL-LF and VDD-LF males (Fig. 3C, D).

When challenged with an HF diet, the glycemia of the VDD female tended to be decreased (nonsignificant) compared with the CTRL-HF group to reach the same values as LF groups (Fig. 3G), and plasma insulin of VDD female (VDD-HF) decreased compared with the control group (CTRL-HF) (p < 0.05) to reach the same values as LF groups (both CTRL-LF

- and VDD-HF) (Fig. 3H). The CTRL-HF females displayed higher values than the other three
  groups regarding of combined plasma insulin, glycemia and HOMA-IR (Fig. 3I, J).
- Insulin tolerance tests were undertaken in CTRL and VDD offspring (performed at 13 weeks).
- 287 No statistically differences were observed between LF groups, for both male and female (Fig.
- 3E, 3K). In obesogenic condition, the area under the curve (AUC) of VDD-HF was increased
- compared with the CTRL HF group of males (p < 0.01) (Fig. 3F) and the AUC of VDD-HF
- 290 was decreased compared with the CTRL of females (p < 0.05) (Fig. 3L).
- 291

### 292 Maternal VDD and HF diet-associated modify morphological parameters and adipose 293 tissue cellularity in sex-specific manner.

- After euthanasia, CTRL-LF and VDD-LF males showed similar body weight and weight gain; 294 similarly, to CTRL-HF and VDD-HF (Table 1). Significant differences were observed 295 296 between LF (both CRTL and VDD) and HF groups (both CTRL and VDD). Compare to the three other groups, VDD-HF males presented the highest fat pad weight (*i.e.* perigonadal, 297 298 retroperitoneal, inguinal fat pad; Table 1), leading to increased adiposity index and leptin plasma level (Fig. 4A, Table 1)). HF diet in CTRL females increased body weight, weight 299 300 gain (between weaning and the end of the protocol), fat pad weight, adiposity index and leptin plasma level, whereas VDD-HF displayed similar parameters than LF groups (both CTRL-LF 301 and VDD-LF). HF diet (both CTRL and VDD) increased liver weight in male compare to the 302 LF (both CTRL and VDD). For females, only CTRL-HF increased liver weight compared 303 with other groups. No statistical difference of spleen's weight of males, but the females 304 exposed to HF diet showed increased spleen weight compare to the LF (Table 1). 305
- The mean adipocyte area of CTRL and VDD males under LF diet were reduced compared with the adipocytes area of males under HF diet (both CTRL and VDD; Fig. 4A, C and E). The mean adipocyte area increased in CTRL-HF females, but was not different in VDD-HF females, compared with LF groups (both CTRL-LF and VDD-LF; Fig. 4B, D and F).
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## 311 Maternal and adult offspring diet modulate the expression of mRNA and associated 312 canonical pathways in adipose tissue.

To identify the impacts of maternal (CTRL, VDD) and adult diets (LF, HF) on the offspring transcriptome, we performed RNA sequencing on visceral adipose tissue. Two set of data were used to characterize the impact of the maternal diet and the adult diet. The first one to study the impact of the HF diet on the same condition of maternal diet (*i.e.* CTRL-LF vs CTRL-HF and VDD-LF vs VDD-HF). The second one to study maternal VDD on the same condition of adult diet (*i.e.* CTRL-LF vs VDD-LF and CTRL-HF vs VDD-HF). We
highlighted the differential expression of transcripts between our conditions as established in
the Supplemental Table 2.

321 Using Ingenuity pathway analysis, we put forward canonical pathways with determinant z-322 score and non-similar between conditions. We highlighted 16 differentially expressed canonical pathways between CTRL-LF and CTRL-HF males. Interestingly, the triacylglycerol 323 biosynthesis, mitochondrial L-carnitine Shuttle Pathway, and fatty acid beta oxidation were 324 differentially decreased in CTRL-HF compared to CTRL-LF. 19 pathways were differentially 325 regulated between the VDD-LF and the VDD-HF males; including the oxidative 326 phosphorylation which was down-regulated in VDD-HF compared to VDD-LF. Also, 6 327 pathways seemed to be common between the offspring exposed to maternal CRTL diet or 328 VDD diet (Fig. 5). 329

When comparing CTRL-LF and VDD-LF males, 1 canonical pathway was expressed, while 3
pathways were expressed between CTRL-HF and VDD-HF males.

332 For the female born from CTRL mice, 31 canonical pathways were differentially expressed between LF and HF, including the fatty acid beta oxidation, the oxidative phosphorylation and 333 334 triacylglycerol biosynthesis which decreased in CTRL-HF compared to CTRL-LF (Fig. 6). No pathway was regulated when comparing VDD-LF to VDD-HF. Thus, when studying the 335 impact of LF diet between CTRL and VDD females, 3 canonical pathways were differentially 336 expressed, while 27 were differentially expressed for females on HF diet, including oxidative 337 phosphorylation, fatty acid beta oxidation, mitochondrial L-carnitine Shuttle pathway that 338 were induced in VDD-HF compared to CTRL-HF. Interestingly, estrogen biosynthesis 339 pathway appeared as induced in VDD-HF compared to CTRL-HF. One pathway was common 340 between LF and HF diets. 341

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#### 345 **Discussion**

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The maternal diet is now well-established as a key player in the foetal development and longterm effects programming in the offspring. In this study, we highlighted the impacts of maternal VDD on offspring metabolism in normal nutritional condition and under HF challenge. We also explored the sexual dimorphism of the metabolic response.

- In agreement with previously published data, we reported here that maternal VDD diet 351 differentially affected the energy homeostasis and the body weight of the juvenile offspring (5 352 weeks old). Indeed, the maternal VDD was associated with reduction of body weight of the 353 male mice offspring in our experiment, similarly to previous reports in 15 days old mice<sup>24</sup> or 354 at weaning<sup>31</sup>. Other studies reported that VDD male offspring displayed higher body weight<sup>32</sup> 355 or not differences  $^{33-35}$ , but it is noteworthy that body weight evaluation have been evaluated in 356 adult rats<sup>32,33</sup> or mice<sup>34,35</sup>, not in juvenile. Interestingly, in our experiment, no discrepancy in 357 body weight was observed in female, demonstrating thus a strong metabolic sex-specific 358 359 response which has never been highlighted so far.
- Such body weight modifications are strongly related to energy metabolism as highlight by 360 361 indirect calorimetry. Indeed, the increased spontaneous activity and energy expenditure 362 (through the carbohydrate oxidation) of the VDD males, together with the absence food intake difference, could explain the smaller body weight of juvenile VDD males. The lack of body 363 weight impact in female may result from the absence of food intake modification in VDD 364 female. The observed increase in VDD female spontaneous activity could explain the increase 365 in carbohydrate oxidation, which was associated to a decrease in lipid oxidation, resulting in 366 overall no modification of energy expenditure. Interestingly, we observed that both male and 367 female VDD of the offspring were characterized by an increase in spontaneous activity. 368 Similar observations have already been reported in males VDD mice<sup>36</sup>, and it has been 369 370 reported that in human, VDD during pregnancy is strongly correlated with the risk of attention deficit and hyperactivity disorder in children<sup>37</sup>. Nevertheless, in juvenile offspring of VDD 371 rats<sup>38</sup>, no modification of locomotor activity was observed. The origin of this phenotype is 372 presently not well understood. Neurobehavioral development could explain this phenotype<sup>39</sup>, 373 but this assumption will require further investigations. 374
- 375

In adulthood, morphological parameters (body weight, fat pad weight, liver weight, adipocyte
area) and energy balance were not modified in male and female offspring of VDD or CTRL
mice, when exposed to a control diet (VDD-LF and CTRL-LF), in agreement with previous

reports<sup>35</sup>. All altered parameters in juvenile mice were normalized in adulthood, including 379 reduced body weight and energy expenditure, and increased spontaneous activity, for both 380 CTRL and VDD offspring. In offspring of VDD rats, similar results were reported in 381 adulthood, *i.e.* no effect on food intake, spontaneous activity in 14 weeks old male rats<sup>32</sup>. 382 Nevertheless, this study also reported differences on body weight and others biological 383 parameters (total cholesterol, triglycerides, HDL, blood glucose, that were higher in VDD 384 compared with CTRL offspring)<sup>32</sup>. The origin of these contradictory observations is not clear 385 but could be due to different compositions of control diets used, that could be sufficient to 386 387 unveil the metabolic phenotype associated to maternal VDD in offspring. In agreement with this assumption, we highlighted that challenged offspring with an HF diets (45% of energy 388 from lipids from 9 to 12 weeks of age) led to a smaller body weight of VDD offspring 389 compared with the CTRL offspring for both males and females. Such observation is not fully 390 consistent with previous reports<sup>31</sup>, but could be related to the genetic background of the mice 391 which differ between studies. Nevertheless, our results in males are consistent with the fact 392 393 that the increased energy expenditure (mainly due to the carbohydrate oxidation), could exceed the increased food intake, leading to smaller body weight of the VDD-HF males 394 395 compared with CTRL-HF males.

Female from VDD-HF diets also presented smaller body weight under HF, compared with LF diet, with a tend to food intake increase. This limitation of body weight might be related to the increase of energy expenditure (through the carbohydrate oxidation). An important issue that remain presently unsolved is the drastic decrease of spontaneous activity in VDD-HF female compared with CTRL-HF.

An interesting observation was that in VDD group the level of expression of the hypothalamic melanocortin pathway genes, *i.e.* NPY, AgRP and MC4R, is different of the CTRL mice level, and could explain at least in part food intake modulations. This suggest that VD is important for the accurate development of the melanocortin pathway. In accordance with this hypothesis, VD is known to be important for the brain development<sup>40</sup>. However, further investigations are needed to understand the impact of VD on the development of neural pathways involved in feeding behaviour.

Beside body weight and energy metabolism, the glucose homeostasis was also investigated in
offspring adulthood. In LF condition, both in males and females, no modification of glucose
homeostasis was observed, especially glycemia, consistently with other studies<sup>24,32-34</sup>.
Nevertheless, several studies reported that VDD in male rats led to an increase of fasting
insulin, HOMA-IR levels and insulin tolerance at 16 weeks<sup>33</sup>, and in VDD male mice to an

increase of insulinemia<sup>34</sup>. Such differences could be related to the model used (mice vs rats<sup>33</sup>) 413 or duration of the protocol (14 weeks vs 6 months<sup>34</sup>). Interestingly, in males submitted to HF 414 diet, we observed higher insulin plasma level, glycemia, HOMA IR and insulin resistance, as 415 previously reported in control conditions<sup>24,32-34</sup>. Surprisingly, in females, no perturbation of 416 the glucose homeostasis was observed. At the opposite, VDD-HF females displayed similar 417 values as CTRL-LF or VDD-LF for several parameters, including insulinemia, glycemia and 418 HOMA-IR. Such observation has never been reported yet, since most of the studies included 419 420 only males.

Since insulin resistance is strongly linked to adiposity<sup>41</sup>, we investigated adiposity index of 421 animals, leptin plasma level and adipose tissue cellularity. Interestingly, adiposity index and 422 leptin plasma concentration were not impacted in males by the VDD in LF condition but was 423 strongly induced by the combination of VDD and HF, similarly to Belenchia et al.<sup>31</sup> and 424 425 appeared to be strongly correlated with insulin resistance. No additive effect was reported for adipose cellularity. In females, under LF diet, no modification was observed for adiposity 426 427 index, leptin plasma level and adipocytes area. As expected, the adiposity index, plasma leptin and cellularity were induced by HF diet in offspring of CTRL mice (CTRL-HF), but 428 429 surprisingly, VDD-HF group displayed similar adiposity index, plasma leptin and adipocyte area compared with LF groups (both CTRL-LF and VDD-LF). Such observation is fully 430 consistent with the improvement of insulin sensitivity observed in the VDD-HF group. 431 Nevertheless, such effect has never been reported yet and could correspond to an adaptative 432 mechanism based on an adequate diet supply of macronutrients and micronutrients, that 433 434 appears to be highly sex-specific.

To investigate the origin of such adiposity discrepancies, RNA-seq experiments were 435 undertaken on retroperitoneal adipose tissue. We hypothesized that differential transcriptomes 436 could be explain the variation of adiposity between group. In agreement, when comparing 437 male offspring from control mice exposed to LF or HF diet in adulthood (i.e. CTRL-LF vs 438 CTRL-HF), we observed that the "mitochondrial L-carnitine shuttle pathway" and the "fatty 439 acid beta oxidation" were down-regulated in the CTRL-HF, which was associated to increase 440 441 of adiposity. In males from VDD mice submitted to LF or HF (i.e. VDD-LF vs VDD-HF), those canonical pathways related to lipid catabolism were not regulated, but we observed a 442 strong repression of oxidative phosphorylation which correlated with fat pad accretion, weight 443 gain and a higher adiposity index. Thus, the discrepancy between offspring of CTRL or VDD 444 mice could be related to the ability to induce lipid oxidation/oxidative phosphorylation in 445 adipose tissue. In addition, the transcriptomic response to maternal diet combined to 446

adulthood diet (i.e. CTRL-LF vs VDD-LF and CTRL-HF vs VDD-HF) did not results in 447 major differences in term of lipid metabolism pathways, nor major difference in adiposity, 448 which reinforces the putative role of adipose tissue lipid metabolism modulations to drive 449 450 adiposity phenotype. It is noteworthy that the contribution of white adipose tissue to whole body energy expenditure is considered relatively small. Nevertheless, there are examples of 451 nutritional and pharmacological interventions in animals resulting in obesity resistance 452 associated with increased oxidative capacity in WAT<sup>42-44</sup>. In agreement with the modest 453 contribution of white adipose tissue on the whole energy expenditure , we did not lipid 454 455 oxidation induction by indirect calorimetry, suggesting that such local induction of lipid oxidation may have beneficial effect in terms of adiposity, but not impact the overall lipid 456 457 oxidation which involve many other organs.

Concerning females, CTRL-HF compared with CTRL-LF repressed several canonical 458 459 pathways linked to lipid metabolism (*i.e.* fatty acid, beta oxidation, oxidative phosphorylation, triacyl glycerol biosynthesis, mitochondrial l-carnitine shuttle pathway). Similarly to males, 460 461 such pattern corresponded to a signature of body weight gain and adiposity. Importantly, when comparing VDD-HF to VDD-LF, we did not observe any modification of metabolic 462 463 canonical pathways, nor modification of adiposity. In addition, the transcriptomic response to maternal diet combined to adulthood diet (i.e. CTRL-LF vs VDD-LF and CTRL-HF vs VDD-464 HF) results in the induction of canonical pathways implicated in lipid metabolism (i.e. beta 465 oxidation, oxidative phosphorylation, triacyl glycerol biosynthesis, mitochondrial l-carnitine 466 shuttle pathway) in VDD-HF compared to CTRL-HF. This was in agreement with the 467 decreased of adiposity index, and body weight observed in VDD-HF compared to CTRL-HF. 468 Altogether these observations suggested that adiposity is strongly associated to lipid 469 metabolism pathway that may influence adipose tissue accretion. The origin of such gene 470 expression profiles modifications is presently not established but we speculate that epigenetic 471 mechanisms could link maternal VDD to long-term transcriptional modifications in 472 offspring's adipose tissue. Such assumption will require further investigations. 473

It is obvious that the adiposity and gene expression profile are strongly sex-specific, and notably the effect of VDD maternal under HF diet which leads to highly divergent adiposity in males and females. Sex-specific metabolic discrepancies and notably adiposity are strongly related to estradiol status<sup>45</sup>. Indeed, it is well-established that estrogens promotes subcutaneous fat accumulation<sup>46</sup> and improve glucose homeostasis<sup>47</sup>. Consequently, the decrease of estrogens associated to menopause is linked to an increase in visceral fat and greater risk for the metabolic syndrome in postmenopausal compared with premenopausal

women<sup>45</sup>. Thus, we focussed on  $17\beta$ -estradiol plasma level to explain observed phenotypes. 481 Interestingly, we noticed that in males, no significant differences of plasma concentrations 482 were observed, suggesting that estradiol by itself is not a driver element. It is important to 483 keep in mind that estradiol plasma level is not the sole element that impact adipose tissue 484 biology, and estrogen receptors are also important<sup>45</sup> and would deserve attention in further 485 experiments. Nevertheless, in females, when comparing CTRL-LF to CTRL-HF, 17β-486 estradiol plasma level tended to increase but did not reach statistical significance, whereas in 487 VDD mice (VDD-LF vs VDD-HF), the plasma level of 17β-estradiol was strongly and 488 significantly induced. This is an important point since 17β-estradiol is well-known to induce 489 metabolic catabolism, leading to weight gain limitation and glucose homeostasis 490 improvement<sup>45,47</sup>, and such increase of  $17\beta$ -estradiol could explain by itself the metabolic 491 improvement observed in VDD-HF female mice. The origin of the induction of 17β-estradiol 492 is presently unknown, even if epigenetic mechanism are suspected to be involved. Further 493 investigations will be mandatory to explain this observation. 494

To conclude, our study brings new informations on the impact of VDD in metabolic disruption in offspring and notably its predisposition to long term metabolic health complications. Importantly it sheds light on the sex-specific adipose tissue / adiposity response, that need to be taking into account in terms of public health.

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512	L Mounien, B Loriod, M Puceat analysed data; E Seipelt, F Tourniaire, C Couturier, J Astier,
513	H Vachon performed research; JF Landrier, E Seipelt, L Mounien wrote the paper.

- 515 Figures legends
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Fig 1. Indirect calorimetry on juvenile offspring during 24hours to compare the energetic metabolism of the VDD and the CTRL. Parameters measured during 24 h for males and females were Body weight (A, H), Food intake (B, I), spontaneous activity (C, J), Respiratory quotient (RQ, D, K), Energy expenditure (EE, E, L), Lipid oxidation (LOX, F, M) and carbohydrate oxidation (GOX, G, N). Values are presented as mean  $\pm$  SEM. Bars not sharing the same letter were significantly different in Fisher's LSD post hoc test. *p* < 0.05

- Fig 2. Morpho-metabolic follow-up of juvenile and 12 weeks-old offspring. (A, B) 524 25OH(D) concentration of 5-weeks-old offspring male (A) and female (B) before the adult 525 diet induced (Low fat LF or HF, High fat diet). Growing curves of males (C) and females (D) 526 527 from the beginning of hf diet to indirect calorimetry at 12-weeks-old. Indirect calorimetry of adults (E-R) offspring during 24hours to compare the energetic metabolism of the VDD and 528 529 the CTRL on HF diet. Parameters measured during 24 h for males and females were Body weight (E, L), Food intake (F, M), activity (G, N), Respiratory quotient (RQ, H, O), Energy 530 531 expenditure (EE, I,P), Lipid oxidation (LOX, J, Q) and carbohydrate oxidation (GOX, K, R). 532 Values are presented as mean  $\pm$  SEM. Bars not sharing the same letter were significantly different in Fisher's LSD post hoc test. p < 0.05533
- 534

Fig 3. Glucose homeostasis of 13-weeks old offspring. Measured of glucose from tail blood of males (A) and females (G). Insulinemia of male (B) and females (H), Ratio glucose/insulin for males (C) and females (F), HOMA-IR for males (D) and females (J). Insulin tolerance test for males on LF diet (E) and HF diet (F), also for females on LF diet (K), and HF diet (L). Values are presented as mean  $\pm$  SEM. Bars not sharing the same letter were significantly different in Fisher's LSD post hoc test. p < 0.05

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**Fig 4. Adiposity index and adipose tissue histology of offspring.** Adiposity index of the offspring has been established for males (A) and females (B). Representative histological images of visceral adipose tissue of males (E) and females (F) adult offspring after eosinhematoxylin coloration (10X magnification). Mean adipocyte area determined using Image J software for males (C) and females (D).. Values are presented as mean  $\pm$  SEM. Bars not sharing the same letter were significantly different in Fisher's LSD post hoc test. *p* < 0.05

549	Fig 5. Overview of canonical pathway differentially expressed on visceral adipose tissue of
550	the male offspring.
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552	Fig 6. Overview of canonical pathway differentially expressed on visceral adipose tissue of
553	the female offspring.
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555	Table 1. General morphological parameters and $17\beta$ -estradiol plasma concentrations obtain
556	from the offspring at the protocol end. Values are presented as mean $\pm$ SEM. Bars not sharing
557	the same letter were significantly different in Fisher's LSD post hoc test. $p < 0.05$
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563	Refer	References		
564				
565	1.	Bendik I, Friedel A, Roos FF, Weber P, Eggersdorfer M. Vitamin D: A critical and		
566		essential micronutrient for human health. Front Physiol. 2014;5 JUL(July):1-14.		
567		doi:10.3389/fphys.2014.00248		
568	2.	Bonnet L, Karkeni E, Couturier C, et al. Gene Expression Pattern in Response to		
569		Cholecalciferol Supplementation Highlights Cubilin as a Major Protein of 25(OH)D		
570		Uptake in Adipocytes and Male Mice White Adipose Tissue. Endocrinology.		
571		2018;159(2):957-966. doi:10.1210/en.2017-00650		
572	3.	Karkeni E, Bonnet L, Marcotorchino J, et al. Vitamin D limits inflammation-linked		
573		microRNA expression in adipocytes in vitro and in vivo: A new mechanism for the		
574		regulation of inflammation by vitamin D. Epigenetics. 2018;13(2):156-162.		
575		doi:10.1080/15592294.2016.1276681		
576	4.	Karkeni E, Marcotorchino J, Tourniaire F, et al. Vitamin D Limits Chemokine		
577		Expression in Adipocytes and Macrophage Migration In Vitro and in Male Mice.		
578		Endocrinology. 2015;156(5):1782-1793. doi:10.1210/en.2014-1647		
579	5.	Marcotorchino J, Gouranton E, Romier B, et al. Vitamin D reduces the inflammatory		
580		response and restores glucose uptake in adipocytes. Mol Nutr Food Res.		
581		2012;56(12):1771-1782. doi:10.1002/mnfr.201200383		
582	6.	Marcotorchino J, Tourniaire F, Astier J, et al. Vitamin D protects against diet-induced		
583		obesity by enhancing fatty acid oxidation. J Nutr Biochem. 2014;25(10):1077-1083.		
584		doi:10.1016/j.jnutbio.2014.05.010		
585	7.	Landrier J-F, Karkeni E, Marcotorchino J, Bonnet L, Tourniaire F. Vitamin D		
586		modulates adipose tissue biology: possible consequences for obesity? Proc Nutr Soc.		
587		2015;25(March):1-9. doi:10.1017/S0029665115004164		
588	8.	Bonnet L, Hachemi MA, Karkeni E, et al. Diet induced obesity modifies vitamin D		
589		metabolism and adipose tissue storage in mice. J Steroid Biochem Mol Biol.		
590		2019;185(May 2018):39-46. doi:10.1016/j.jsbmb.2018.07.006		
591	9.	Landrier J, Mounien L, Tourniaire F. Obesity and Vitamin D Metabolism		
592		Modifications. J Bone Miner Res. 2019;34(7):1383-1383. doi:10.1002/jbmr.3739		
593	10.	Borg SA, Buckley H, Owen R, et al. Early life Vitamin D depletion alters the postnatal		
594		response to skeletal loading in growing and mature bone. <i>PLoS One</i> . 2018;13(1):1-17.		
595		doi:10.1371/journal.pone.0190675		
596	11.	Turck D, Bresson J, Burlingame B, et al. Dietary reference values for vitamin D. EFSA		

- 597 *J*. 2016;14(10):e04547. doi:10.2903/j.efsa.2016.4547
- Palacios C, Gonzalez L. Is vitamin D deficiency a major global public health problem? *J Steroid Biochem Mol Biol*. 2014;144 Pt A(787):138-145.

600 doi:10.1016/j.jsbmb.2013.11.003

- Cashman KD, Dowling KG, Gonzalez-Gross M, et al. Vitamin D deficiency in Europe:
  pandemic? *Am J Clin Nutr*. 2016;(C):1-12. doi:10.3945/ajcn.115.120873.
- 603 14. Saraf R, Morton SMB, Camargo CA, Grant CC. Global summary of maternal and
  604 newborn vitamin D status a systematic review. *Matern Child Nutr.* 2016;12(4):647605 668. doi:10.1111/mcn.12210
- Miliku K, Blanken LME, Gaillard R, et al. Maternal Vitamin D concentrations during
  pregnancy, fetal growth patterns, and risks of adverse birth outcomes. *Am J Clin Nutr*.
  2016;103(6):1514-1522. doi:10.3945/ajcn.115.123752.1514
- Achkar M. Vitamin D status in early pregnancy and risk of preeclampsia. *Am J Obstet Gynecol.* 2016;212(4):1-14. doi:10.1016/j.ajog.2014.11.009.Vitamin
- 611 17. Amraei M, Mohamadpour S, Sayehmiri K, Mousavi SF, Shirzadpour E, Moayeri A.
- Effects of vitamin D deficiency on incidence risk of gestational diabetes mellitus: A
  systematic review and meta-analysis. *Front Endocrinol (Lausanne)*. 2018;9(FEB):1-11.
  doi:10.3389/fendo.2018.00007
- 615 18. Gernand AD, Simhan HN, Klebanoff MA, Bodnar LM. Maternal serum 25-
- hydroxyvitamin D and measures of newborn and placental weight in a U.S. multicenter
  cohort study. *J Clin Endocrinol Metab.* 2013;98(1):398-404. doi:10.1210/jc.2012-3275
- Wang H, Xiao Y, Zhang L, Gao Q. Journal of Steroid Biochemistry and Molecular
  Biology Maternal early pregnancy vitamin D status in relation to low birth weight and
  small-for-gestational-age o ff spring. *J Steroid Biochem Mol Biol*. 2017;(157):0-1.

621 doi:10.1016/j.jsbmb.2017.09.010

- Morales E, Rodriguez A, Valvi D, et al. Deficit of vitamin D in pregnancy and growth
  and overweight in the offspring. *Int J Obes*. 2015;39(1):61-68.
- 624 doi:10.1038/ijo.2014.165
- Crozier SR, Harvey NC, Inskip HM, Godfrey KM. Maternal vitamin D status in
   pregnancy is associated with adiposity in the offspring : prospective observational
- 627 study. *Am J Clin Nutr*. 2012;96(1):57-63. doi:10.3945/ajcn.112.037473.Maternal
- 628 22. Daraki V, Roumeliotaki T, Chalkiadaki G, et al. Low maternal vitamin D status in
- pregnancy increases the risk of childhood obesity. *Pediatr Obes*. 2018;13(8):467-475.
- 630 doi:10.1111/ijpo.12267

- Rytter D, Bech BH, Halldorsson TI, et al. Maternal Vitamin D status at week 30 of
  gestation and offspring cardio-metabolic health at 20 years: A prospective cohort study
  over two decades. *PLoS One*. 2016;11(10):1-12. doi:10.1371/journal.pone.0164758
- 634 24. Reichetzeder C, Chen H, Föller M, et al. Maternal Vitamin D Deficiency and Fetal
- Programming Lessons Learned from Humans and Mice. *Kidney Blood Press Res.*2014;39(4):315-329. doi:10.1159/000355809
- 637 25. Ideraabdullah FY, Belenchia AM, Rosenfeld CS, et al. Maternal vitamin D deficiency
  638 and developmental origins of health and disease (DOHaD). *J Endocrinol*.
- 639 2019;241(2):R65-R80. doi:10.1530/joe-18-0541
- Goyal D, Limesand SW, Goyal R. Epigenetic responses and the developmental origins
  of health and disease. *J Endocrinol*. 2019;242(1):T105-T119. doi:10.1530/JOE-190009
- Even PC, Mokhtarian A, Pele A. Practical aspects of indirect calorimetry in laboratory
  animals. *Neurosci Biobehav Rev.* 1994;18(3):435-447.
- Fenni S, Hammou H, Astier J, et al. Lycopene and tomato powder supplementation
  similarly inhibit high-fat diet induced obesity, inflammatory response, and associated
  metabolic disorders. *Mol Nutr Food Res.* 2017;61(9):1601083.
- 648 doi:10.1002/mnfr.201601083
- 64929.Livak KJ, Schmittgen TD. Analysis of Relative Gene Expression Data Using Real-650Time Quantitative PCR and the 2- $\Delta\Delta$ CT Method. *Methods*. 2001;25(4):402-408.
- 651 doi:10.1006/meth.2001.1262
- 30. Derghal A, Djelloul M, Trouslard J, Mounien L. The Role of MicroRNA in the
  Modulation of the Melanocortinergic System. *Front Neurosci.* 2017;11(APR):1-8.
  doi:10.3389/fnins.2017.00181
- Belenchia AM, Johnson SA, Ellersieck MR, Rosenfeld CS, Peterson CA. In utero
  vitamin D deficiency predisposes offspring to long-term adverse adipose tissue effects. *J Endocrinol.* 2017;234(3):301-313. doi:10.1530/JOE-17-0015
- Wen J, Hong Q, Wang X, et al. The effect of maternal vitamin D deficiency during
  pregnancy on body fat and adipogenesis in rat offspring. *Sci Rep.* 2018;8(1):365.
  doi:10.1038/s41598-017-18770-4
- 33. Zhang H, Chu X, Huang Y, et al. Maternal vitamin D deficiency during pregnancy
  results in insulin resistance in rat offspring, which is associated with inflammation and
- 663 Iκbα methylation. *Diabetologia*. 2014;57(10):2165-2172. doi:10.1007/s00125-014-
- 664 3316-7

- Nascimento FAM, Ceciliano TC, Aguila MB, Mandarim-de-lacerda CA. 34. 665 Transgenerational Effects on the Liver and Pancreas Resulting from Maternal Vitamin 666 D Restriction in Mice. J Nutr Sci Vitaminol (Tokyo). 2013;59(5):367-374. 667 http://jlc.jst.go.jp/DN/JST.JSTAGE/jnsv/59.367?lang=en&from=CrossRef&type=abstr 668 669 act. 35. Belenchia AM, Jones KL, Will M, et al. Maternal vitamin D deficiency during 670 pregnancy affects expression of adipogenic-regulating genes peroxisome proliferator-671 activated receptor gamma (PPARy) and vitamin D receptor (VDR) in lean male mice 672 673 offspring. Eur J Nutr. 2018;57(2):723-730. doi:10.1007/s00394-016-1359-x Fu L, Chen Y-H, Chen X, Xu S, Yu Z, Xu D-X. Vitamin D deficiency impairs 674 36. 675 neurobehavioral development in male mice. Physiol Behav. 2017;179(July):333-339. 676 doi:10.1016/j.physbeh.2017.07.017 677 37. Morales E, Julvez J, Torrent M, et al. Vitamin D in Pregnancy and Attention Deficit Hyperactivity Disorder-like Symptoms in Childhood. Epidemiology. 2015;26(4):458-678 679 465. doi:10.1097/EDE.00000000000292 Pan P, Jin DHS, Chatterjee-Chakraborty M, et al. The effects of vitamin D3 during 680 38. 681 pregnancy and lactation on offspring physiology and behavior in Sprague-Dawley rats. 682 Dev Psychobiol. 2014;56(1):12-22. doi:10.1002/dev.21086 39. Hawes JE, Tesic D, Whitehouse AJ, Zosky GR, Smith JT, Wyrwoll CS. Maternal 683 vitamin D deficiency alters fetal brain development in the BALB/c mouse. Behav Brain 684 Res. 2015;286:192-200. doi:10.1016/j.bbr.2015.03.008 685 40. 686 Eyles D, McGrath J. Vitamin D Brain Development and Function. In: Vitamin D. Vol 1. Fourth Edi. Elsevier; 2018:563-581. doi:10.1016/B978-0-12-809965-0.00033-1 687 41. Rosen ED, Spiegelman BM. Adipocytes as regulators of energy balance and glucose 688 homeostasis. Nature. 2006;444(7121):847-853. doi:10.1038/nature05483 689 690 42. Kusminski CM, Scherer PE. Mitochondrial dysfunction in white adipose tissue. Trends Endocrinol Metab. 2012;23(9):435-443. doi:10.1016/j.tem.2012.06.004 691 692 43. Tourniaire F, Musinovic H, Gouranton E, et al. All- trans retinoic acid induces oxidative phosphorylation and mitochondria biogenesis in adipocytes. J Lipid Res. 693 2015;56(6):1100-1109. doi:10.1194/jlr.M053652 694 44. Flachs P, Rossmeisl M, Kuda O, Kopecky J. Stimulation of mitochondrial oxidative 695 capacity in white fat independent of UCP1: A key to lean phenotype. Biochim Biophys 696 Acta - Mol Cell Biol Lipids. 2013;1831(5):986-1003. doi:10.1016/j.bbalip.2013.02.003 697
- 698 45. Brown L, Clegg D. Central effects of estradiol in the regulation of food intake, body

699		weight, and adiposity. J Steroid Biochem Mol Biol. 2010;122(1-3):65-73.
700		doi:10.1016/j.jsbmb.2009.12.005
701	46.	Krotkiewski M, Bjorntorp P, Sjostrom L, Smith U. Impact of obesity on metabolism in
702		men and women. Importance of regional adipose tissue distribution. J Clin Invest.
703		1983;72(3):1150-1162.
704	47.	Riant E, Waget A, Cogo H, Arnal J-F, Burcelin R, Gourdy P. Estrogens Protect against
705		High-Fat Diet-Induced Insulin Resistance and Glucose Intolerance in Mice.
706		Endocrinology. 2009;150(5):2109-2117. doi:10.1210/en.2008-0971
707		
708		