



**HAL**  
open science

## A double-tracer radioisotope approach to assess simultaneous bioaccumulation of caesium in the olive flounder *Paralichthys olivaceus*

Roberta Hansman, Marc Metian, Simon Pouil, François Oberhänsli, Jean-Louis Teyssié, Peter Swarzenski

### ► To cite this version:

Roberta Hansman, Marc Metian, Simon Pouil, François Oberhänsli, Jean-Louis Teyssié, et al.. A double-tracer radioisotope approach to assess simultaneous bioaccumulation of caesium in the olive flounder *Paralichthys olivaceus*. *Journal of Environmental Radioactivity*, 2018, 190-191 (12), pp.141-148. 10.1016/j.jenvrad.2018.05.014 . hal-03163115

**HAL Id: hal-03163115**

**<https://hal.inrae.fr/hal-03163115>**

Submitted on 5 Sep 2023

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 **A double-tracer radioisotope approach to assess simultaneous bioaccumulation of caesium in the**  
2 **olive flounder *Paralichthys olivaceus***

3 Roberta L. Hansman, Marc Metian, François Oberhänsli, Jean-Louis Teyssié, and Peter W. Swarzenski  
4 *International Atomic Energy Agency – Environment Laboratories, Radioecology Laboratory; 4a Quai*  
5 *Antoine 1er, MC-98000 Principality of Monaco*

6

7 **Abstract**

8 To better understand bioaccumulation of radiocaesium in the commercially important Japanese  
9 flatfish, *Paralichthys olivaceus*, the uptake and depuration kinetics of caesium via both seawater and  
10 food were assessed simultaneously using controlled aquaria. The pre-conditioned fish were exposed  
11 to radionuclides via the two different pathways (aqueous versus dietary) concurrently using two  
12 isotopes of caesium,  $^{137}\text{Cs}$  and  $^{134}\text{Cs}$ , respectively. Dissolved caesium uptake was linear and did not  
13 reach a steady state over the course of the 8-day exposure period. Consumption of  $^{134}\text{Cs}$ -labelled  
14 food led to higher bioaccumulation rates of radioactive Cs than via seawater exposure of  $^{137}\text{Cs}$   
15 during uptake and following depuration, though the model-derived long-lived biological half-lives of  
16 both pathways was approximately 66 d. Further development of this method for assessing multiple  
17 radiocaesium bioaccumulation pathways simultaneously could lead to a promising new approach for  
18 studying Cs contamination in marine organisms.

19 **1. Introduction**

20 As a consequence of the accident at the Tokyo Electric Power Company (TEPCO) Fukushima Dai-ichi  
21 Nuclear Power Plant (FDNPP; IAEA, 2015), large amounts of radioactive caesium [estimates for  $^{137}\text{Cs}$   
22 vary from 3.5 PBq according to Tsumune et al. (2012) to 27 PBq reported by Bailly du Bois et al.  
23 (2012)] had been released into the ocean. This radioactive release was predominantly transported  
24 southward (Aoyama et al., 2012; Tsumune et al., 2012), and relatively high concentrations of  
25 radioactive caesium [both  $^{134}\text{Cs}$  (half-life of 2.065 y) and  $^{137}\text{Cs}$  (30.167 y)] were detected in a variety  
26 of marine organisms around the southern coast of Fukushima Prefecture after the accident  
27 (Arakawa et al., 2015; Shigenobu et al., 2014). Approximately 6 years have passed since the accident  
28 occurred, and the radioactive caesium concentrations in seawater off the coast of Fukushima  
29 Prefecture have now dropped so that they are close to pre-accident levels (0.001–0.002 Bq L<sup>-1</sup>)  
30 (Kusakabe et al., 2013; Oikawa et al., 2013). Concentration reductions have also been observed in  
31 seaweed, cephalopods, shellfish, and crustaceans; however, the rates of reduction have varied  
32 among taxonomic groups. Radiocaesium concentrations have also declined in fish species that were  
33 significantly contaminated [e.g., Japanese rockfish (*Sebastes cheni*), fat greenling (*Hexagrammos*

34 *otakii*), and marbled sole (*Pleuronectes yokohamae*] (Iwata et al., 2013; Sohtome et al., 2014; Wada  
35 et al., 2013).

36 The Japanese government banned landings of many marine species in the vicinity of Fukushima,  
37 including *Paralichthys olivaceus*, after the accident due to the presence of high levels of radioactive  
38 Cs (Wada et al., 2013). The olive flounder *P. olivaceus* is a demersal fish native to the subtropical and  
39 temperate western Pacific Ocean and widely distributed in the coastal waters around Japan. An  
40 economically important aquaculture species in East Asia since the 1990s (Kikuchi and Takeda, 2001),  
41 the olive flounder was a target species of a stock enhancement program that released around one  
42 million hatchery-raised juveniles annually in Fukushima Prefecture (Tomiyama et al., 2008). Several  
43 studies have monitored the radiocaesium contamination in *P. olivaceus* following the accident,  
44 including modelling the uptake and depuration biokinetics of this fish or assessing the depuration  
45 biokinetics using naturally exposed fish (Kurita et al., 2015; Tateda et al., 2015, 2016, 2017).

46 Studies have focused on the differences in the bioaccumulation of radionuclides in marine organisms  
47 depending on the particular contaminant pathway, be it through aqueous, dietary, sedimentary, or  
48 maternal exposure routes. The uptake and depuration of radionuclides by marine organisms is  
49 variable depending on species, element, and environmental conditions. Some studies have been  
50 able to demonstrate that radiocaesium concentrations increase with increasing trophic levels  
51 (Kasamatsu and Ishikawa, 1997; Mathews and Fisher, 2008), providing evidence for bioaccumulation  
52 and suggesting biomagnification (Mathews and Fisher, 2008; Pan and Wang, 2016; Zhao et al.,  
53 2001).

54 While these pathways have previously been evaluated separately in the laboratory for many species  
55 of marine organisms exposed to a suite of radioisotopes and metals including Cs (e.g., Bustamante et  
56 al., 2006; Metian et al., 2011, 2016; Warnau et al., 1996a, 1996b), to our knowledge no such  
57 experiments have yet been performed to quantify the simultaneous uptake and depuration of  
58 caesium radionuclides via both seawater exposure and diet. The advantages of analysing these  
59 exposure pathways concurrently are both practical and scientific. From a practical standpoint,  
60 experimental resources including time may be much reduced. Scientifically, the compounding effects  
61 of two exposure pathways can be evaluated, as contamination in the marine environment will  
62 always involve multiple concurrent sources of exposure. We were able to measure the effects of  
63 these two exposure pathways simultaneously through the use of two different radioisotopes of Cs,  
64 <sup>134</sup>Cs and <sup>137</sup>Cs.

65 Here we demonstrate the concurrent bioaccumulation and depuration of radioactive Cs in the  
66 Japanese flatfish *Paralichthys olivaceus*, commonly known as olive flounder, via both food and  
67 seawater exposure pathways. We also evaluate the utility of this double-tracer radioisotope  
68 approach in assessing these processes simultaneously in the laboratory and explore possible future  
69 applications of this methodology.

## 70 **2. Material and methods**

### 71 *2.1 Experimental organisms*

72 Japanese aquaculture juvenile fish *Paralichthys olivaceus* were obtained from a fish wholesaler  
73 (Tropic Nguyen, France). They were acclimated to laboratory conditions for 4 weeks in an open  
74 circuit 500-L aquarium; flux: 50 L h<sup>-1</sup> of 1- $\mu$ m filtered seawater; salinity: 38 g L<sup>-1</sup>; temperature:  
75 20.5  $\pm$  0.5  $^{\circ}$ C; pH: 8.0  $\pm$  0.1; light/dark cycle: 12 h/12 h. During this period fish were fed daily with  
76 frozen *Artemia salina* and *Euphasia pacifica*.

### 77 *2.2 Radiotracers and counting*

78 The uptake and depuration of radiocaesium in *P. olivaceus* were determined using radiotracers  
79 purchased from Polatom (<sup>134</sup>CsCl in aqueous solution) and Areva Cerva Lee (<sup>137</sup>CsCl in 0.1 N HCl).  
80 <sup>134</sup>Cs and <sup>137</sup>Cs were counted using a high-resolution  $\gamma$ -spectrometer system composed of four  
81 high-purity germanium (HPGe) detectors (efficiency = 50%) connected to a multi-channel analyzer  
82 and a computer equipped with spectra analysis software Interwinner 6. Precise activities of <sup>134</sup>Cs  
83 (605, 796 keV) and <sup>137</sup>Cs (662 keV) were determined using standards (i.e., phantoms, as described in  
84 Cresswell et al., 2017) of known activity and appropriate geometries, and measurements were  
85 subsequently corrected for counting efficiencies and radioactive decay (Cresswell et al., 2017).  
86 Counting times ranged from 20 to 73 min with an average of 50 min. The counting times were  
87 adjusted to obtain propagated counting errors generally less than 5%, although a few samples with  
88 very low activities had counting errors up to 15%.

### 89 *2.3 Experimental procedure*

90 A single experiment was conducted to investigate Cs bioaccumulation in the Japanese flatfish  
91 simultaneously through seawater and dietary exposure pathways over a long period (87 d total  
92 consisting of 8 d of uptake followed by 79 d of depuration). The experiment was conducted using  
93 eleven *P. olivaceus* fish (mean initial weight 5.19  $\pm$  1.85 g) in 70-L closed-circuit aquaria constantly  
94 aerated with an aquarium water pump under the following conditions: salinity = 38 g L<sup>-1</sup>,  
95 temperature = 20.5  $\pm$  0.5  $^{\circ}$ C, pH = 8.0  $\pm$  0.1, light/dark cycle = 12 h/12 h. All 11 organisms were  
96 exposed for 8 d to seawater spiked with <sup>137</sup>Cs dissolved in 1  $\mu$ m-filtered seawater (1 Bq mL<sup>-1</sup>), and 10

97 of these were fed food labelled with  $^{134}\text{Cs}$  to allow for one single-exposed ( $^{137}\text{Cs}$  via seawater)  
98 control.

99 Radiolabelled food was prepared by growing *Artemia salina* in seawater containing 220 kBq  $^{134}\text{Cs}$ ,  
100 with *Isochrysis galbana* to keep the prey fed and healthy over 8 d, leading to labelled *A. salina*. Fish  
101 were fed this  $^{134}\text{Cs}$ -labelled *A. salina* (mean daily weight  $2.7 \pm 0.2$  g; mean daily activity =  $232 \pm 13$   
102 Bq) for six morning feedings (days 0, 1, 2, 3, 4, and 7) and supplemented with unlabelled krill every  
103 afternoon. With regards to the multiple feeding approach used here,  $^{134}\text{Cs}$  activity also reflects prior  
104 feedings, as well as any depuration that occurred during the following day. During depuration, the  
105 same daily feeding schedule was kept using both unlabelled *A. salina* and krill. For seawater  
106 exposure, a daily spike of  $^{137}\text{Cs}$  accompanied six daily water changes (days 0, 1, 2, 3, 4, and 7) for an  
107 average seawater  $^{137}\text{Cs}$  activity of  $1.066 \pm 0.063$  Bq  $\text{g}^{-1}$  over the exposure period ( $^{137}\text{Cs}$  radioactivity  
108 in the water was measured before and after each seawater renewal; i.e., time-integrated activity).  
109 This concentration is a fraction of the maximum  $^{137}\text{Cs}$  concentrations in the discharge following the  
110 accident and comparable in magnitude to values observed in surface seawater near Fukushima  
111 (Buesseler et al., 2011).

112 During the 79-day depuration period, 7 fish were placed under uncontaminated conditions  
113 (constantly aerated, open-circuit aquarium; flow =  $50 \text{ L h}^{-1}$ ; salinity =  $38 \text{ g L}^{-1}$ ,  
114 temperature =  $20.5 \pm 0.5$  °C, pH =  $8.0 \pm 0.1$ , light/dark cycle = 12 h/12 h), collected at different time  
115 intervals, and whole-body radioanalyzed alive.

#### 116 1.4 Data analyses

117 The uptake kinetics of dissolved  $^{137}\text{Cs}$  was expressed in terms of change in concentration factor (CF,  
118 ratio of whole-body fish  $^{137}\text{Cs}$  activity in Bq  $\text{g}^{-1}$  wet weight as a function of the time-integrated  
119 seawater  $^{137}\text{Cs}$  activity in Bq  $\text{g}^{-1}$ ) over time for the seawater exposure. Kinetics were best described  
120 using a linear model (Eq. (1))

$$121 \quad (1) \quad CF_t = k_u t$$

122 where  $CF_t$  is the concentration factor at time  $t$  (d) and  $k_u$  are the biological uptake rate constants  
123 ( $\text{d}^{-1}$ ; e.g. Whicker and Schultz, 1982).

124 Depuration kinetics for  $^{134}\text{Cs}$  and  $^{137}\text{Cs}$  were fit to a simple, two-component exponential loss model  
125 (Eq. 2):

$$(2) A_t = A_{0s}e^{-k_{es}t} + A_{0l}e^{-k_{el}t}$$

126 where  $k_e$  is the depuration rate constant ( $d^{-1}$ ), and  $A_t$  and  $A_0$  are the total activities (Bq) at time  $t$  (d)  
127 and 0, respectively; 's' and 'l' subscripts denote the short- and long-lived exponential components.  
128 Biological half-lives ( $T_{b_{1/2s}}$  and  $T_{b_{1/2l}}$ ) were calculated from the corresponding depuration rate constant  
129 ( $k_{es}$  and  $k_{el}$ , respectively) according to the relation  $T_{b_{1/2}} = \ln 2/k_e$  as in Whicker and Schultz (1982).  
130 Model constants and statistics were estimated by iterative adjustment of the model using the non-  
131 linear curve fitting routines in the Statistica software package (StatSoft, Inc., 2004) and statistical  
132 methods as in Warnau et al. (1996a, 1996b) and Metian et al. (2011). Additional statistical analyses  
133 were performed using R (R Core Team, 2016).

134 The percentage of  $^{134}\text{Cs}$  food activity assimilated was calculated by dividing the total  $^{134}\text{Cs}$  activity  
135 measured in the fish each day during the uptake phase by the total cumulative  $^{134}\text{Cs}$  activity in the  
136 food given (as  $^{134}\text{Cs}$ -labelled *A. salina*). The relative contribution of  $^{134}\text{Cs}$  (food) and  $^{137}\text{Cs}$  (seawater)  
137 to total activity was calculated as the proportion of the mean activity of each radioisotope ( $^{134}\text{Cs}$  or  
138  $^{137}\text{Cs}$  in Bq) to the mean total activity (i.e.,  $^{134}\text{Cs} + ^{137}\text{Cs}$  in Bq) in the fish each day measurements  
139 were taken during the experiment.

### 140 3. Results

#### 141 3.1 Uptake

142 The simultaneous uptake of  $^{134,137}\text{Cs}$  by *P. olivaceus* through both aqueous and dietary exposure  
143 pathways is shown in Fig. 1 as total activity (Bq) over time (d). Multiple feedings of  $^{134}\text{Cs}$ -labelled  
144 food resulted in higher total activities in the fish than through seawater exposure. Over the initial  
145 four days, the rate of accumulation was more than double for dietary uptake of Cs than seawater  
146 exposure ( $3.441 \text{ Bq d}^{-1}$  vs.  $1.216 \text{ Bq d}^{-1}$ ;  $R^2 = 0.992$  and  $0.971$  for linear regression, respectively).  
147 Although some depuration occurred during the two-day pause in  $^{134}\text{Cs}$ -labelled feedings, the total  
148 activity increased over the entire exposed period and  $11.5 \pm 1.0\%$  of the total food  $^{134}\text{Cs}$  activity  
149 given to the fish was assimilated (Fig. 2). As seawater  $^{137}\text{Cs}$  exposure continued during the two-day  
150 pause in feeding and counting, total activity in fish for  $^{137}\text{Cs}$  increased linearly over the entire  
151 exposure period ( $1.225 \text{ Bq d}^{-1}$ ,  $R^2 = 0.996$ ). While the multiple feeding strategy utilized in this  
152 experiment does not allow for the calculation of assimilation efficiency (AE) as in single feeding  
153 studies, the calculated concentration factor (CF) for seawater exposure reached a value of  
154  $1.61 \pm 0.47$  at the end of the exposure (day 8) with an uptake rate constant ( $k_u$ ) of  $0.205 \text{ d}^{-1}$ .

#### 155 3.2 Depuration

156 Depuration of  $^{134}\text{Cs}$  and  $^{137}\text{Cs}$  over the 79-day experiment is shown in Fig. 3A and B, with total activity  
157 plotted in (A) and the percentage of remaining activity in (B). Depuration kinetics were best  
158 described by a simple two-component exponential model (Fig. 3A; Table 1). The initial depuration  
159 rate was higher for  $^{137}\text{Cs}$  than  $^{134}\text{Cs}$  ( $k_{\text{es}} = 0.41$  and  $0.16 \text{ d}^{-1}$ , respectively), though both appeared to  
160 reach a steady plateau by the end of the experiment. Dietary exposure to Cs through multiple  
161 feedings led to a higher total activity of  $^{134}\text{Cs}$  at this plateau compared to  $^{137}\text{Cs}$  seawater exposure;  
162 however, the amount of remaining activity compared to the maximum values reached were similar  
163 for both exposure pathways. The remarkable similarities in the derived long-lived biological half-lives  
164 ( $T_{\text{b}_{1/2\text{l}}} = 65.63 \pm 27.74 \text{ d}$  and  $65.65 \pm 17.79 \text{ d}$  for  $^{134}\text{Cs}$  and  $^{137}\text{Cs}$ , respectively) from the fitted two-  
165 component exponential loss models for both food and seawater exposure clearly highlight this  
166 observation.

### 167 3.3 Global bioaccumulation

168 The relative contribution of  $^{134}\text{Cs}$  vs.  $^{137}\text{Cs}$  to total activity over the course of the entire experiment is  
169 shown in Fig. 4. The average contribution of Cs activity from seawater exposure over all 87 d was  
170  $34.6 \pm 2.5\%$  ( $\pm$ one standard deviation), and though slightly more variable during the uptake period,  
171 there was no significant difference in relative contribution when compared to the loss phase  
172 ( $33.9 \pm 4.6\%$  and  $34.7 \pm 1.2\%$ , respectively;  $p > 0.05$ ). Approximately two-thirds of the total Cs  
173 radioactivity in *P. olivaceus* during both uptake and depuration is due to consumption of Cs-  
174 contaminated food.

## 175 4. Discussion

176 The olive flounder is a commercially important fishery that was essentially closed in the waters  
177 around Fukushima following the accident due to observed increased levels of radiocaesium  
178 contamination above the Japanese standard limit for food safety of  $100 \text{ Bq kg}^{-1}$  wet weight enforced  
179 in April 2012 (Wada et al., 2013). Concentrations of  $^{134}\text{Cs}$  +  $^{137}\text{Cs}$  in the surrounding seawater  
180 immediately after the accident were initially very high but decreased rapidly (Aoyama et al., 2016),  
181 yet concentrations in *P. olivaceus* tissues remained high and could be found in excess of the limit up  
182 to 3 years later (up to  $230 \text{ Bq kg}^{-1}$ ; Kurita et al., 2015). By these standards, both dietary and aqueous  
183 exposure to radiocaesium at the concentrations used in the present experiment led to  
184 contamination levels in *P. olivaceus* within one day. During depuration, concentrations of  
185 radiocaesium did not fall below the food safety limit by the end of the experiment 79 d after  
186 exposure as final average concentrations were  $569 \pm 211$  and  $288 \pm 76 \text{ Bq kg}^{-1}$  for  $^{134}\text{Cs}$  and  $^{137}\text{Cs}$ ,  
187 respectively. This accounted for 19.2% and 16.9% of the maximum  $^{134}\text{Cs}$  and  $^{137}\text{Cs}$  concentrations  
188 at the beginning of the depuration period, respectively. This direct comparison from our laboratory

189 experiment and the field should be taken in context however, as the juvenile fish used here can have  
190 different uptake and depuration biokinetics than commercial-sized adult flounder (e.g., Suzuki et al.,  
191 1992). Nonetheless, it is still useful to make intermediate connections between laboratory and field  
192 measurements with the goal of further understanding contamination pathways in the marine  
193 environment.

194 Delineating Cs bioaccumulation pathways in aquatic organisms contributes to our understanding of  
195 Cs measurements reported from the field in biota after a contamination event. In ecotoxicological  
196 studies, the contribution of different contamination pathways (water, food, and sediment) is usually  
197 estimated using bioenergetic models developed by Thomann (1981) implemented with kinetic data  
198 measured in controlled conditions (Reinfelder et al., 1998; Thomann et al., 1995; Wang et al., 1996).  
199 One of the main disadvantages of this methodology is that it requires the implementation of difficult  
200 and complex experimental protocols (e.g., Hédouin et al., 2010; Metian et al., 2009, 2016). In the  
201 present study, we carried out a simple experiment using a double-tracer radioisotope approach to  
202 more easily provide the first information regarding the contribution of dietary and aqueous sources  
203 of Cs in its global accumulation by *P. olivaceus*. This approach has some limitations (Table 2), and the  
204 relative contribution of dietary versus aqueous exposure pathways to radiocaesium bioaccumulation  
205 was over-simplified in this study due to the multiple and partially sporadic feedings as compared to  
206 implementing a biodynamic model. Nevertheless, the simultaneous exposure using two  
207 radioisotopes of caesium suggests the predominant role of food in the bioaccumulation of Cs in *P.*  
208 *olivaceus* (approximately two-thirds of the Cs whole-body activity derived from food; Fig. 4). This  
209 finding is in agreement with previous studies with other fish species (Mathews et al., 2008; Zhao et  
210 al., 2001).

211 Sediment exposure, which was not tested in this experiment, is expected to be an additional  
212 pathway for Cs contamination in *P. olivaceus* due to their benthic niche. Nevertheless, it could be  
213 considered in the feeding pathway (particulate pathway). As seawater Cs concentrations are  
214 typically much lower than those of sediment, one might expect bioaccumulation from sediment to  
215 be higher than via seawater exposure in the marine environment for demersal species. Limited  
216 studies comparing seawater and sediment radiocaesium exposure pathways have shown sediment-  
217 bound Cs to be bioavailable (Wang et al., 2016), though its contribution to Cs bioaccumulation  
218 compared to seawater exposure is variable (<1–31% and 6–24% for seawater and sediment <sup>134</sup>Cs  
219 uptake pathways, respectively; Metian et al., 2016). Further investigations are needed to  
220 characterize the importance of this Cs bioaccumulation pathway in *P. olivaceus* and properly confirm  
221 our results using a bioenergetic model over a long-term experiment.



222 In fish, the trophic transfer of radionuclides can be best assessed experimentally by two main  
223 methods: (1) the “single-feeding” approach where fish are fed radiolabelled food for a unique pulse-  
224 chase feeding [as described by Wang and Fisher (1999)], and (2) the “multi-feeding” approach where  
225 fish are regularly exposed to radiolabelled food (e.g., Pouil et al., 2017). The latter has the advantage  
226 of tracking more similarly to marine organisms consuming contaminated food over a period of time  
227 as would be expected in natural systems with prolonged sources of Cs contamination. However, the  
228 “multiple feeding” approach utilized in this experiment does not allow for the calculation of  
229 assimilation efficiency (AE; see the review of Pouil et al., 2018). Nevertheless, an analogous  
230 parameter to AE is the percentage of remaining  $^{134}\text{Cs}$  activity when the data plateau after  
231 approximately 60 d of depuration, which was  $36.0 \pm 17.8\%$  for *P. olivaceus* in this experiment (Fig.  
232 3B). Comparing this to calculated AEs from other single-feeding studies, juvenile cuttlefish displayed  
233 an AE of  $29.2 \pm 3.6\%$  and a similar long-lived biological half-life  $T_{b1/2}$  of 66 d after a single feeding of  
234  $^{134}\text{Cs}$ -contaminated *A. salina*, though depuration biological half-lives following seawater exposure  
235 and dietary exposure in adults were much different ( $T_{b1/2} = 6.1$  and 16 d, respectively) than for *P.*  
236 *olivaceus* (Bustamante et al., 2006). From the same flatfish order as *P. olivaceus* (Pleuronectiformes),  
237 the turbot *Psetta maxima* had a higher AE of  $63 \pm 2\%$  and  $T_{b1/2}$  of 36.5 d following consumption of  
238  $^{134}\text{Cs}$ -contaminated prey (Mathews et al., 2008). An even greater AE of  $79.6 \pm 8.6\%$  with a  $T_{b1/2}$  of  
239 13.9 d was determined in the killifish *Fundulus heteroclitus* after consuming  $^{137}\text{Cs}$ -contaminated  
240 blackworms (Wang et al., 2016).

241 Although the assimilation of Cs is very variable among fish species (from 50 to 95%; Pouil et al.,  
242 2018), the remaining activity values in the present study are still considered low compared to AEs of  
243 other high predatory species such as the seabass *Dicentrarchus labrax* (Mathews and Fisher, 2008)  
244 and the false kelpfish *Sebastes marmoratus* (Pan and Wang, 2016). It is generally assumed that  
245 AEs of Cs are higher in predator fish compared to planktivorous and herbivorous species (Pan and  
246 Wang, 2016; Rowan and Rasmussen, 1994). In the present study, the amount of remaining activity  
247 suggests that this statement is not always true. In fish, the mechanisms underlying species-  
248 dependent AE of radionuclides are unclear though Chan et al. (2003) attributed the differences of  
249 radionuclide AEs between the mudskipper *Periophthalmus modestus* and the rabbitfish *Siganus*  
250 *canaliculatus* to the gut passage time (GPT), with a longer GPT corresponding to a higher AE.

251 In many studies considering the trophic transfer of Cs in fish, emphasis is on the potential for  
252 biomagnification of this radionuclide in marine food chains (e.g., Pan and Wang, 2016; Zhao et al.,  
253 2001). To determine this potential, the most common approach consists of calculating the trophic  
254 transfer factor (TTF; Reinfelder et al., 1998) from the kinetic parameters (AE and  $k_e$ ) and the

255 ingestion rate (IR). When TTF >1, it indicates a potential Cs biomagnification; when TTF <1,  
256 biomagnification is unlikely (Mathews et al., 2008; Reinfelder et al., 1998). Several studies have  
257 concluded that biomagnification of Cs can occur in the marine environment. In our study we cannot  
258 calculate the TTF since we have not adopted an approach allowing for the proper measurement of  
259 the required kinetic parameters; however, the “multi-feeding” approach carried out here can be  
260 used to characterize when biomagnification is effective (i.e., when Cs concentrations are higher in  
261 fish than in food). As such, based on the first 4 days of feeding with radiolabelled brine shrimp where  
262 concentrations of Cs in fish were multiplied by approximately 4.5 (Fig. 2) and assuming a linear  
263 increase in Cs concentrations in *P. olivaceus* (Fig. 1), biomagnification could occur in less than one  
264 month. These preliminary results raise the interest of using the multiple feeding approach to confirm  
265 experimentally previous results obtained by modelling.

266 Our results indicated a limited bioaccumulation of Cs in *P. olivaceus* from seawater exposure. The  
267 concentration factor (CF) calculated for *P. olivaceus* in this experiment of  $1.61 \pm 0.47$  is generally low  
268 compared to other fish species (Jeffree et al., 2010; Zhao et al., 2001), and much lower than  
269 invertebrates such as cephalopods and decapods (e.g., Bustamante et al., 2006; Metian et al., 2016).  
270 It is nevertheless important to note that contrary to what has occurred in past studies, the  
271 radiocaesium uptake kinetics did not reach a plateau during the exposure period; thus, it seems we  
272 can expect a high CF value in steady-state conditions for this fish species. However, such results  
273 suggest low Cs bioaccumulation capacities from aqueous exposure in *P. olivaceus* (very low uptake  
274 rate constant), and we can assume based on this experiment that bioaccumulation of Cs is mainly  
275 derived by dietary intake in this species.

276 A similar double-tracer method has been used previously to assess dietary versus aqueous exposure  
277 pathways in the bioaccumulation of radioactive polonium in decapods and fish (Carvalho and  
278 Fowler, 1994). The time and resources saved through use of this technique are significant, yet the  
279 technical challenges to source, administer, and analyse multiple radioisotopes of a specific element  
280 of interest can be great (Table 2). Furthermore, in such experiments full control of single-tracer  
281 exposure is not possible and potential cross-contamination could occur such as seawater adsorbed  
282 to food or leaching of radiolabelled food into the seawater. Further improvements and future  
283 directions for this methodology include utilizing a single pulse-chase feeding rather than the multiple  
284 feedings as in this experiment (Pouil et al., 2017), extending exposure time to reach a steady-state  
285 concentration factor (Fig. 1), and incorporating Cs bioaccumulation via exposure to contaminated  
286 sediments.

## 287 **5. Conclusions**

288 To maximize resources, the double-radioisotope approach used in this study allows for a novel  
289 assessment of the simultaneous determination of caesium bioaccumulation via both dietary and  
290 aqueous exposure pathways. Using this method, the results of this work indicate that food was the  
291 predominant uptake pathway for radiocaesium in the olive flounder *P. olivaceus*, relative to  
292 seawater exposure. Implications for this work would extend to seafood safety programmes that  
293 must examine all vectors for contamination.

## 294 **Acknowledgements**

295 This work was supported by the IAEA Environment Programme. The IAEA is grateful for the support  
296 provided to its Environment Laboratories by the Government of the Principality of Monaco. The  
297 authors thank Dr. M. Warnau for his fruitful advice in the design of the experiment.

## 298 **References**

- 299 Aoyama, M., Hamajima, Y., Hult, M., Uematsu, M., Oka, E., Tsumune, D., Kumamoto, Y., 2016. 134Cs  
300 and 137Cs in the North Pacific Ocean derived from the March 2011 TEPCO Fukushima Dai-  
301 ichi Nuclear Power Plant accident, Japan. Part one: surface pathway and vertical  
302 distributions. *J. Oceanogr.* 72, 53–65. <https://doi.org/10.1007/s10872-015-0335-z>
- 303 Aoyama, M., Tsumune, D., Uematsu, M., Kondo, F., & Hamajima, Y. (2012). Temporal variation of  
304 134Cs and 137Cs activities in surface water at stations along the coastline near the  
305 Fukushima Dai-ichi Nuclear Power Plant accident site, Japan. *Geochemical Journal*, 46(4),  
306 321-325.
- 307 Arakawa, H., Tokai, T., Miyamoto, Y., Akiyama, S., Uchida, K., Matsumoto, A., ... & Hirakawa, N.  
308 (2015). Distribution of radioactive material in marine ecosystems off the Fukushima coast:  
309 radioactive cesium levels in Fukushima marine organisms. In *Marine Productivity:  
310 Perturbations and Resilience of Socio-ecosystems: Proceedings of the 15th French-Japanese  
311 Oceanography Symposium* (pp. 71-78). Springer International Publishing.
- 312 Bailly du Bois, P., Laguionie, P., Boust, D., Korsakissok, I., Didier, D., & Fiévet, B. (2012). Estimation of  
313 marine source-term following Fukushima Dai-ichi accident. *Journal of Environmental  
314 Radioactivity*, 114, 2-9.
- 315 Buesseler, K., Aoyama, M., Fukasawa, M., 2011. Impacts of the Fukushima nuclear power plants on  
316 marine radioactivity. *Environ. Sci. Technol.* 45, 9931–9935.  
317 <https://doi.org/10.1021/es202816c>
- 318 Bustamante, P., Teyssié, J.-L., Fowler, S.W., Warnau, M., 2006. Assessment of the exposure pathway  
319 in the uptake and distribution of americium and cesium in cuttlefish (*Sepia officinalis*) at  
320 different stages of its life cycle. *J. Exp. Mar. Biol. Ecol.* 331, 198–207.  
321 <https://doi.org/10.1016/j.jembe.2005.10.018>

- 322 Carvalho, F.P., Fowler, S.W., 1994. A double-tracer technique to determine the relative importance  
323 of water and food as sources of polonium-210 to marine prawns and fish. *Mar. Ecol. Prog.*  
324 *Ser.* 103, 251–264. <https://doi.org/10.2307/24842668>
- 325 Chan, S. M., Wang, W. X., & Ni, I. H. (2003). The uptake of Cd, Cr, and Zn by the macroalga  
326 *Enteromorpha crinita* and subsequent transfer to the marine herbivorous rabbitfish, *Siganus*  
327 *canaliculatus*. *Archives of Environmental Contamination and Toxicology*, 44, 0298-0306.
- 328 Cresswell, T., Metian, M., Golding, L. A., & Wood, M. D. (2017). Aquatic live animal radiotracing  
329 studies for ecotoxicological applications: Addressing fundamental methodological  
330 deficiencies. *Journal of Environmental Radioactivity*, 178, 453-460.
- 331 Hédouin, L., Metian, M., Teyssié, J.-L., Fichez, R., Warnau, M., 2010. Delineation of heavy metal  
332 contamination pathways (seawater, food and sediment) in tropical oysters from New  
333 Caledonia using radiotracer techniques. *Mar. Pollut. Bull.* 61, 542–553.  
334 <https://doi.org/10.1016/j.marpolbul.2010.06.037>
- 335 IAEA, I. (2015). The Fukushima Daiichi Accident. In Report by the Director General. Vienna:  
336 International Atomic Energy Agency.
- 337 Iwata, K., Tagami, K., & Uchida, S. (2013). Ecological half-lives of radiocesium in 16 species in marine  
338 biota after the TEPCO's Fukushima Daiichi Nuclear Power Plant accident. *Environmental*  
339 *science & technology*, 47(14), 7696-7703.
- 340 Jeffree, R. A., Oberhansli, F., & Teyssie, J. L. (2010). Phylogenetic consistencies among  
341 chondrichthyan and teleost fishes in their bioaccumulation of multiple trace elements from  
342 seawater. *Science of the total environment*, 408(16), 3200-3210.
- 343 Kasamatsu, F., Ishikawa, Y., 1997. Natural variation of radionuclide <sup>137</sup>Cs concentration in marine  
344 organisms with special reference to the effect of food habits and trophic level. *Mar. Ecol.*  
345 *Prog. Ser.* 160, 109–120. <https://doi.org/10.2307/24858838>
- 346 Kikuchi, K., Takeda, S., 2001. Present status of research and production of Japanese flounder,  
347 *Paralichthys olivaceus*, in Japan. *J. Appl. Aquaculture* 11, 165–175.  
348 [https://doi.org/10.1300/J028v11n01\\_12](https://doi.org/10.1300/J028v11n01_12)
- 349 Kurita, Y., Shigenobu, Y., Sakuma, T., Ito, S., 2015. Radiocesium Contamination Histories of Japanese  
350 Flounder (*Paralichthys olivaceus*) After the 2011 Fukushima Nuclear Power Plant Accident,  
351 in: Nakata, K., Sugisaki, H. (Eds.), *Impacts of the Fukushima Nuclear Accident on Fish and*  
352 *Fishing Grounds*. Springer Japan, Tokyo, pp. 139–151. [https://doi.org/10.1007/978-4-431-](https://doi.org/10.1007/978-4-431-55537-7_11)  
353 [55537-7\\_11](https://doi.org/10.1007/978-4-431-55537-7_11)
- 354 Kusakabe, M., Oikawa, S., Takata, H., Misonoo, J., 2013. Spatiotemporal distributions of Fukushima-  
355 derived radionuclides in nearby marine surface sediments. *Biogeosciences* 10, 5019–5030.  
356 <https://doi.org/10.5194/bg-10-5019-2013>
- 357 Mathews, T., Fisher, N.S., 2008. Trophic transfer of seven trace metals in a four-step marine food  
358 chain. *Mar. Ecol. Prog. Ser.* 367, 23–33.

- 359 Mathews, T., Fisher, N.S., Jeffree, R.A., Teyssié, J.-L., 2008. Assimilation and retention of metals in  
360 teleost and elasmobranch fishes following dietary exposure. *Mar. Ecol. Prog. Ser.* 360, 1–12.
- 361 Metian, M., Bustamante, P., Hédouin, L., Oberhänsli, F., & Warnau, M. (2009). Delineation of heavy  
362 metal uptake pathways (seawater and food) in the variegated scallop *Chlamys varia*, using  
363 radiotracer techniques. *Marine ecology progress series*, 375, 161-171.
- 364 Metian, M., Pouil, S., Hédouin, L., Oberhänsli, F., Teyssié, J.-L., Bustamante, P., Warnau, M., 2016.  
365 Differential bioaccumulation of <sup>134</sup>Cs in tropical marine organisms and the relative  
366 importance of exposure pathways. *J. Environ. Radioact.* 152, 127–135.  
367 <https://doi.org/10.1016/j.jenvrad.2015.11.012>
- 368 Metian, M., Warnau, M., Teyssié, J.-L., Bustamante, P., 2011. Characterization of <sup>241</sup>Am and <sup>134</sup>Cs  
369 bioaccumulation in the king scallop *Pecten maximus*: investigation via three exposure  
370 pathways. *J. Environ. Radioact.* 102, 543–550.  
371 <https://doi.org/10.1016/j.jenvrad.2011.02.008>
- 372 Oikawa, S., Takata, H., Watabe, T., Misonoo, J., & Kusakabe, M. (2013). Distribution of the  
373 Fukushima-derived radionuclides in seawater in the Pacific off the coast of Miyagi,  
374 Fukushima, and Ibaraki Prefectures, Japan. *Biogeosciences*, 10(7), 5031-5047.
- 375 Pan, K., & Wang, W. X. (2016). Radiocesium uptake, trophic transfer, and exposure in three estuarine  
376 fish with contrasting feeding habits. *Chemosphere*, 163, 499-507.
- 377 Pouil, S., Bustamante, P., Warnau, M., & Metian, M. (2018). Overview of trace element trophic  
378 transfer in fish through the concept of assimilation efficiency. *Marine Ecology Progress  
379 Series*, 588, 243-254.
- 380 Pouil, S., Warnau, M., Oberhänsli, F., Teyssié, J.-L., Bustamante, P., Metian, M., 2017. Comparing  
381 single-feeding and multi-feeding approaches for experimentally assessing trophic transfer of  
382 metals in fish. *Environ. Toxicol. Chem.* 36, 1227–1234. <https://doi.org/10.1002/etc.3646>
- 383 R Core Team, 2016. R: A Language and Environment for Statistical Computing. R Foundation for  
384 Statistical Computing, Vienna, Austria.
- 385 Reinfelder, J. R., Fisher, N. S., Luoma, S. N., Nichols, J. W., & Wang, W. X. (1998). Trace element  
386 trophic transfer in aquatic organisms: a critique of the kinetic model approach. *Science of  
387 the Total Environment*, 219(2-3), 117-135.
- 388 Rowan, D. J., & Rasmussen, J. B. (1994). Bioaccumulation of radiocesium by fish: the influence of  
389 physicochemical factors and trophic structure. *Canadian Journal of Fisheries and Aquatic  
390 Sciences*, 51(11), 2388-2410.
- 391 Shigenobu, Y., Fujimoto, K., Ambe, D., Kaeriyama, H., Ono, T., Morinaga, K., ... & Watanabe, T.  
392 (2014). Radiocesium contamination of greenlings (*Hexagrammos otakii*) off the coast of  
393 Fukushima. *Scientific reports*, 4(1), 6851.
- 394 Sohtome, T., Wada, T., Mizuno, T., Nemoto, Y., Igarashi, S., Nishimune, A., ... & Ishimaru, T. (2014).  
395 Radiological impact of TEPCO's Fukushima Dai-ichi Nuclear Power Plant accident on

396 invertebrates in the coastal benthic food web. *Journal of environmental radioactivity*, 138,  
397 106-115.

398 StatSoft, Inc., 2004. *Statistica* (data analysis software system). Version 6.

399 Suzuki, Y., Nakamura, K., Nakamura, R., Nakahara, M., Ishii, T., Matsuba, M., & Nagaya, Y. (1992).  
400 Radioecological studies in the marine environment.

401 Tateda, Y., Tsumune, D., Misumi, K., Aono, T., Kanda, J., Ishimaru, T., 2017. Biokinetics of  
402 radiocesium depuration in marine fish inhabiting the vicinity of the Fukushima Dai-ichi  
403 Nuclear Power Plant. *J. Environ. Radioact.* 166, 67–73.  
404 <https://doi.org/10.1016/j.jenvrad.2016.02.028>

405 Tateda, Y., Tsumune, D., Tsubono, T., 2013. Simulation of radioactive cesium transfer in the southern  
406 Fukushima coastal biota using a dynamic food chain transfer model. *J. Environ. Radioact.*  
407 124, 1–12. <https://doi.org/10.1016/j.jenvrad.2013.03.007>

408 Tateda, Y., Tsumune, D., Tsubono, T., Misumi, K., Yamada, M., Kanda, J., Ishimaru, T., 2016. Status of  
409 <sup>137</sup>Cs contamination in marine biota along the Pacific coast of eastern Japan derived from a  
410 dynamic biological model two years simulation following the Fukushima accident. *J. Environ.*  
411 *Radioact.* 151, 495–501. <https://doi.org/10.1016/j.jenvrad.2015.05.013>

412 Thomann, R. V. (1981). Equilibrium model of fate of microcontaminants in diverse aquatic food  
413 chains. *Canadian Journal of Fisheries and Aquatic Sciences*, 38(3), 280-296.

414 Thomann, R. V., Mahony, J. D., & Mueller, R. (1995). Steady-state model of biota sediment  
415 accumulation factor for metals in two marine bivalves. *Environmental Toxicology and*  
416 *Chemistry: An International Journal*, 14(11), 1989-1998.

417 Tomiyama, T., Watanabe, M., & Fujita, T. (2008). Community-based stock enhancement and fisheries  
418 management of the Japanese flounder in Fukushima, Japan. *Reviews in Fisheries Science*,  
419 16(1-3), 146-153.

420 Tsumune, D., Tsubono, T., Aoyama, M., & Hirose, K. (2012). Distribution of oceanic <sup>137</sup>Cs from the  
421 Fukushima Dai-ichi Nuclear Power Plant simulated numerically by a regional ocean model.  
422 *Journal of environmental radioactivity*, 111, 100-108.

423 Wada, T., Nemoto, Y., Shimamura, S., Fujita, T., Mizuno, T., Sohtome, T., Kamiyama, K., Morita, T.,  
424 Igarashi, S., 2013. Effects of the nuclear disaster on marine products in Fukushima. *J.*  
425 *Environ. Radioact.* 124, 246–254. <https://doi.org/10.1016/j.jenvrad.2013.05.008>

426 Wang, C., Baumann, Z., Madigan, D.J., Fisher, N.S., 2016. Contaminated marine sediments as a  
427 source of cesium radioisotopes for benthic fauna near Fukushima. *Environmental Science &*  
428 *Technology* 50, 10448–10455. <https://doi.org/10.1021/acs.est.6b02984>

429 Wang, W. X., & Fisher, N. S. (1999). Assimilation efficiencies of chemical contaminants in aquatic  
430 invertebrates: a synthesis. *Environmental Toxicology and Chemistry: An International*  
431 *Journal*, 18(9), 2034-2045.

- 432 Wang, W. X., Fisher, N. S., & Luoma, S. N. (1996). Kinetic determinations of trace element  
433 bioaccumulation in the mussel *Mytilus edulis*. *Marine ecology progress series*, 140, 91-113.
- 434 Warnau, M., Fowler, S.W., Teyssié, J.-L., 1996a. Biokinetics of selected heavy metals and  
435 radionuclides in two marine macrophytes: the seagrass *Posidonia oceanica* and the alga  
436 *Caulerpa taxifolia*. *Mar. Environ. Res.* 41, 343–362. [https://doi.org/10.1016/0141-](https://doi.org/10.1016/0141-1136(95)00025-9)  
437 [1136\(95\)00025-9](https://doi.org/10.1016/0141-1136(95)00025-9)
- 438 Warnau, M., Teyssié, J.-L., Fowler, S.W., 1996b. Biokinetics of selected heavy metals and  
439 radionuclides in the common Mediterranean echinoid *Paracentrotus lividus*: sea water and  
440 food exposures. *Mar. Ecol. Prog. Ser.* 141, 83–94.
- 441 Whicker, F.W., Schultz, V., 1982. *Radioecology: Nuclear Energy and the Environment*. CRC Press,  
442 Boca Raton, Florida.
- 443 Zhao, X., Wang, W.-X., Yu, K.N., Lam, P.K.S., 2001. Biomagnification of radiocesium in a marine  
444 piscivorous fish. *Mar. Ecol. Prog. Ser.* 222, 227–237.
- 445

446 **Figure Captions**

447 **Figure 1.** Uptake of  $^{134}\text{Cs}$  via food and  $^{137}\text{Cs}$  via seawater in Japanese flatfish (*P. olivaceus*) over 8 d.  
448 Values are means  $\pm$  one standard deviation (n = 9-11).

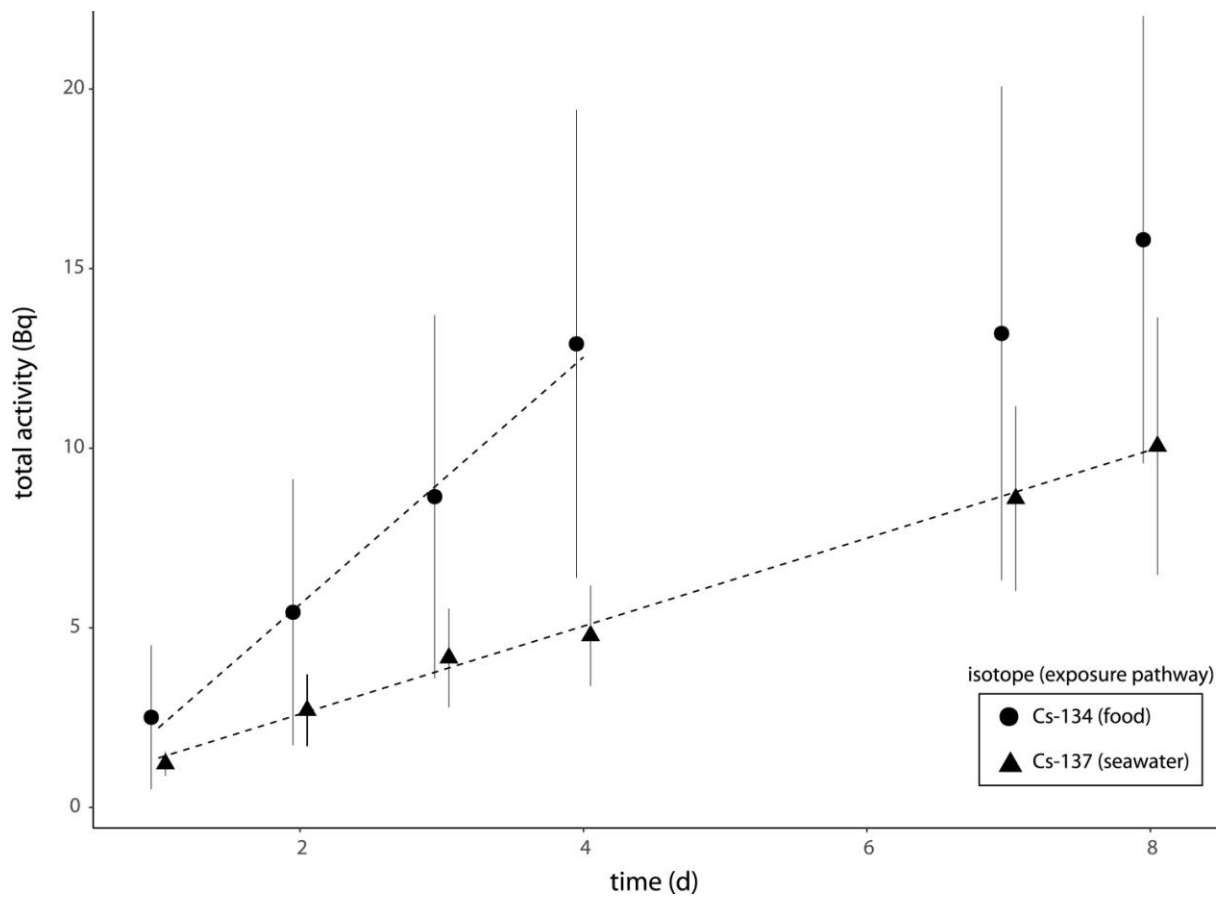
449 **Figure 2.** Daily change and total  $^{134}\text{Cs}$  activity (Bq) in Japanese flatfish (*P. olivaceus*) exposed via  
450 food over 8 d. Also plotted is the percentage of total food activity assimilated by the fish over time.

451 **Figure 3.** Depuration kinetics of  $^{134}\text{Cs}$  and  $^{137}\text{Cs}$  in Japanese flatfish (*P. olivaceus*) over 79 d  
452 following food and seawater exposure. Total activity (Bq) and kinetic models are displayed in (A) and  
453 the percentage of remaining activity in (B). Values are means  $\pm$  one standard deviation (n = 5–7).

454 **Figure 4.** Relative contribution (%) of the uptake pathways (seawater or food) to the total activity of  
455 Cs in Japanese flatfish (*P. olivaceus*) over the course of the experiment (87 d). The end of exposure is  
456 indicated following day 8 by \*. The dashed line marked X is the average for the entire experiment.

457

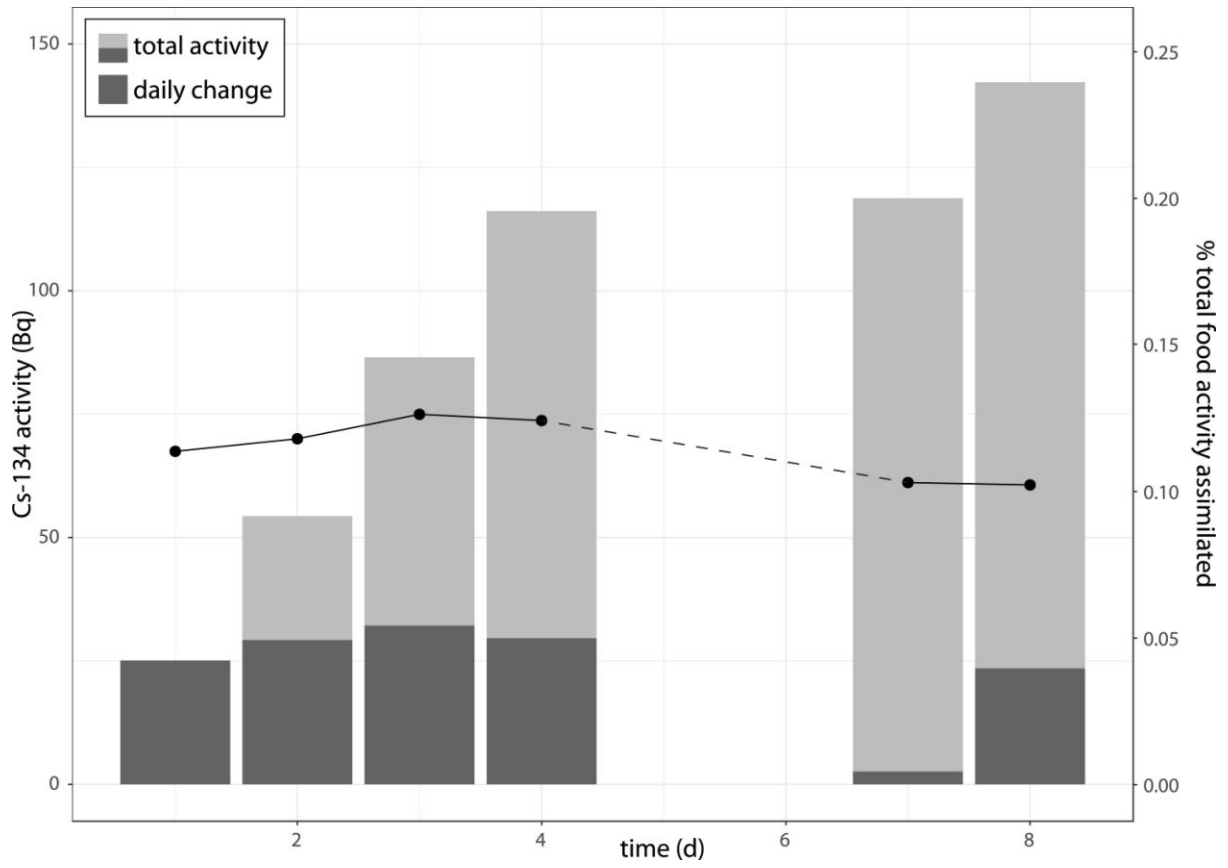




458

459 **Figure 1**

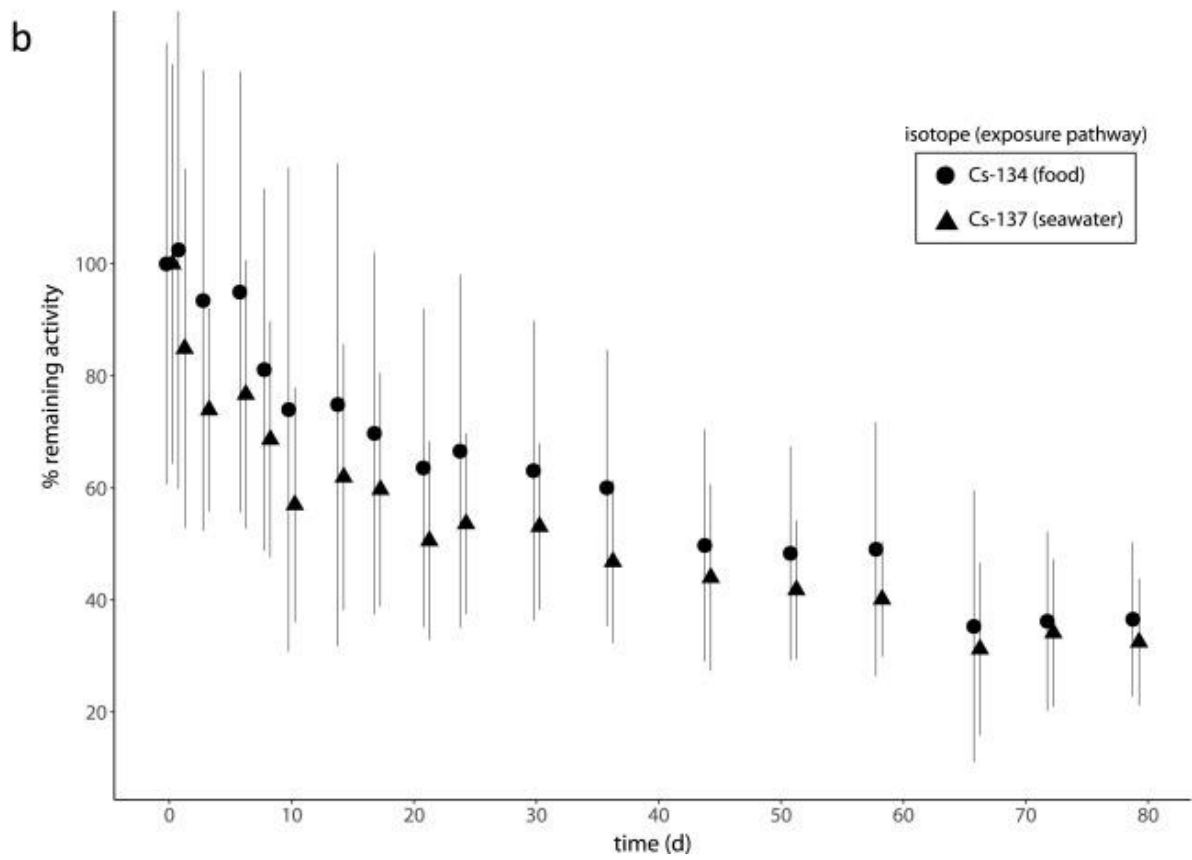
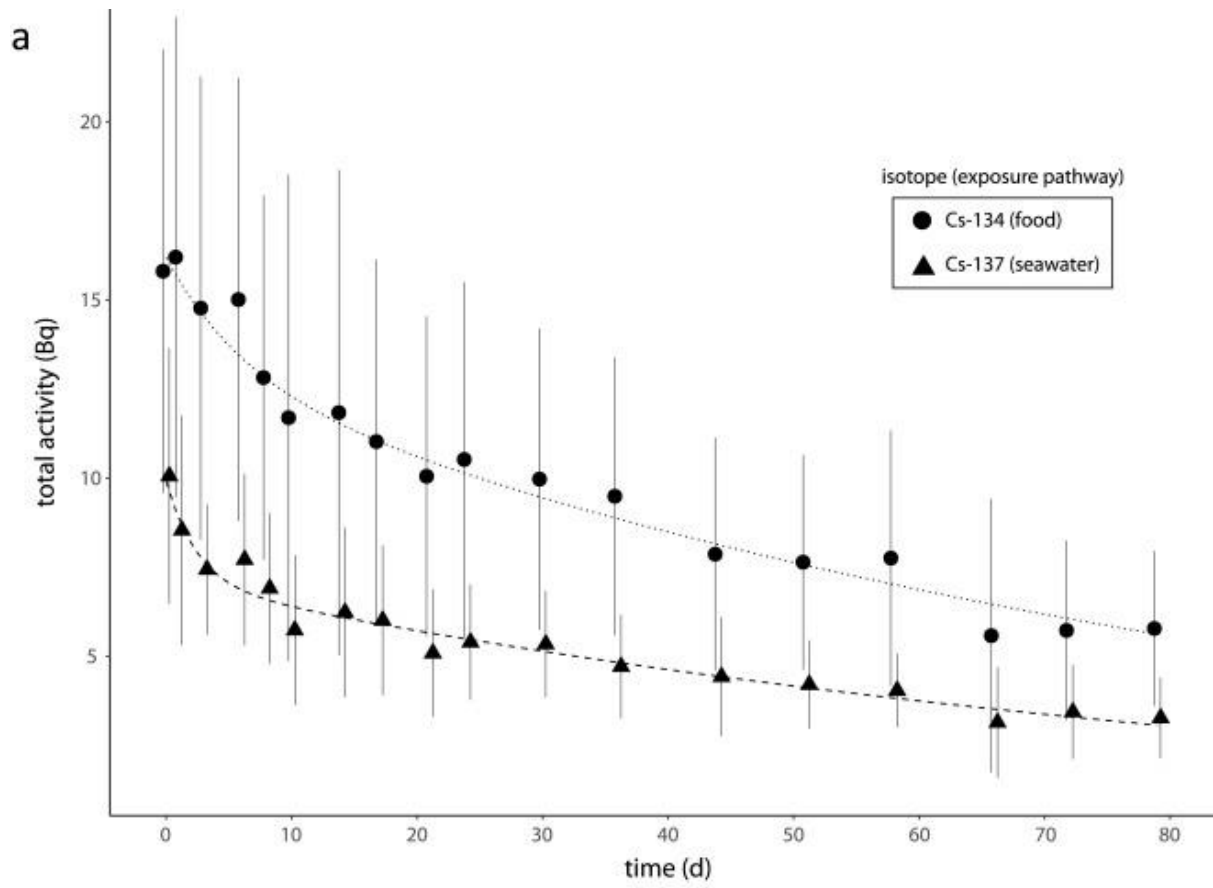
460



461

462 **Figure 2**

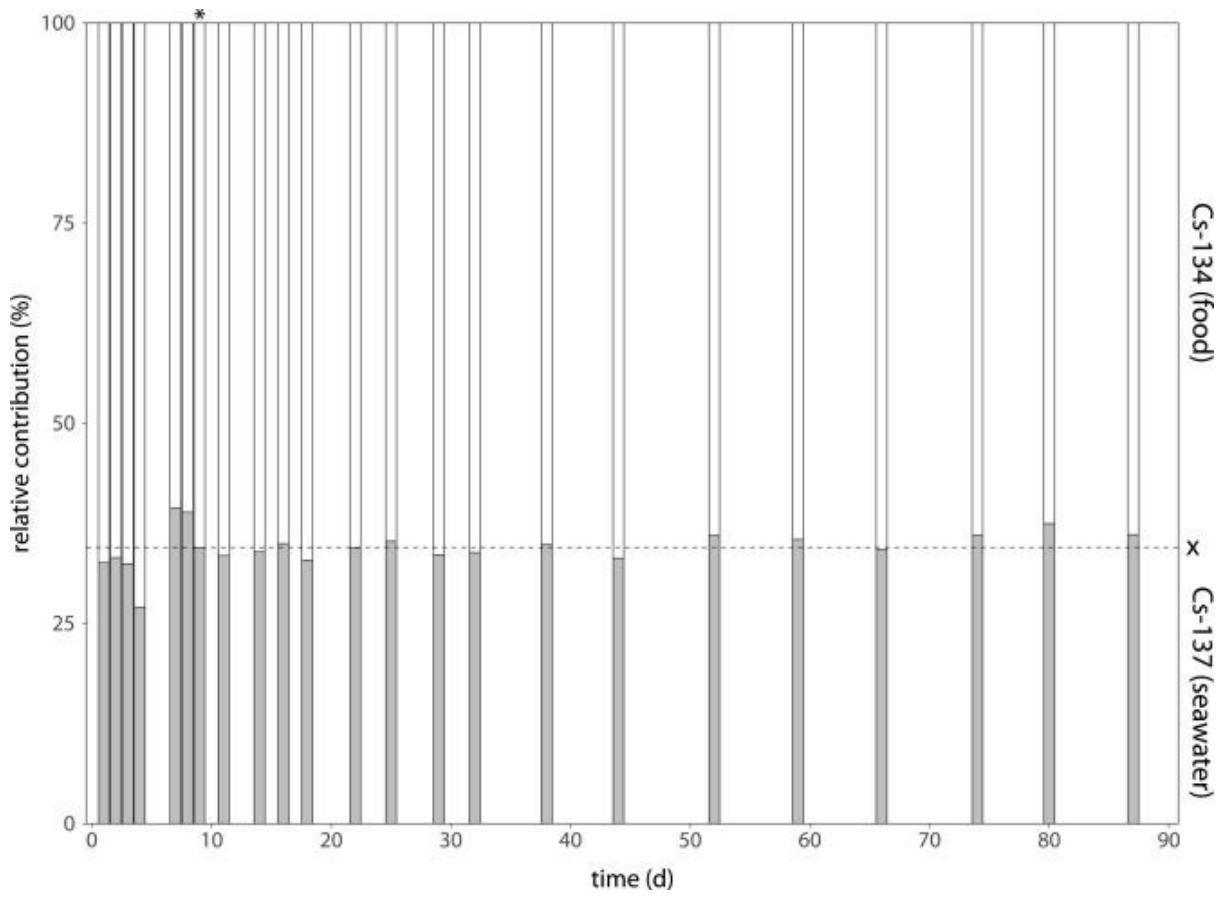
463



464

465 **Figure 3**

466



467

468 **Figure 4**

469 **Tables**

470 **Table 1.** Model parameters for the depuration kinetics of <sup>134</sup>Cs and <sup>137</sup>Cs in Japanese flatfish (*P.*  
 471 *olivaceus*) exposed via food and seawater. A<sub>0s</sub> and A<sub>0l</sub>: activity (Bq) lost according to the short- and  
 472 long-lived exponential component, respectively; T<sub>b½</sub>: biological half-life (d) [T<sub>b½</sub> = ln2/k<sub>e</sub>]; ASE:  
 473 asymptotic standard error; R<sup>2</sup>: determination coefficient of kinetics. Probability (p) of each  
 474 parameter estimation is indicated as follows: <sup>NS</sup>Not significant (p > 0.05), \* p < 0.05, \*\* p < 0.001.

Isotope	Exposure Pathway	A <sub>0s</sub> ± ASE	T <sub>b½s</sub> ± ASE	A <sub>0l</sub> ± ASE	T <sub>b½l</sub> ± ASE	R <sup>2</sup>
Cs-134	Food	3.24 ± 2.81 <sup>NS</sup>	4.31 ± 6.10 <sup>NS</sup>	12.94 ± 2.58 <sup>**</sup>	65.63 ± 27.74 <sup>*</sup>	0.32
Cs-137	Seawater	2.85 ± 0.79 <sup>**</sup>	1.68 ± 2.28 <sup>NS</sup>	7.06 ± 0.79 <sup>**</sup>	65.65 ± 17.79 <sup>**</sup>	0.49

475

476 **Table 2.** List of advantages and disadvantages of the double-tracer radioisotope approach used in  
 477 this study.

Advantages	Disadvantages
saves time by running single concurrent experiment (also labour, lab resources)	requires purchasing two different radioactive sources, which implies an increasing cost
can evaluate simultaneous and/or compounding effects on single fish exposed by both pathways	potential analytical issues resolving both isotopes  potential risk to not have full control of single tracer exposure (potential cross-contamination could occur such as seawater on food or leaching of labelled food into seawater)
	limited to two simultaneous exposure pathways studied per experiment

478