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Efficient monitoring of phenology in chestnuts

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Color artwork: online only

1 Abstract

2 Chestnuts (*Castanea spp.*) are ecologically and economically important forest and fruit trees. They
3 are cultivated for their nutritious nuts. To select varieties to be cultivated in chestnut orchards, their
4 phenology needs to be considered. In this work, we adapt the international BBCH system to
5 chestnuts by building on an existing phenological scale used in some European countries for these
6 species. The proposed BBCH scale uses eight of the ten principal growth stages for fruit trees and
7 secondary growth stages that are specific to chestnut trees. We tested it by monitoring chestnut
8 trees phenology during three growing seasons, illustrating its suitability for high-throughput
9 phenotyping studies. Overall, the approach used, despite its inherent limitations, is straightforward,
10 accessible and flexible, allowing particularly precise description of the complex flowering phenology
11 of these trees.

12 Keywords

13 BBCH, Flowering phenology, Pollination, Monoecious, Male-sterile, Fagaceae, Castanea

14

15 Introduction

16 Plant phenology is the timing of plant life seasonal events, such as bud burst, flowering, fruiting, and
17 leaf abscission. It plays a fundamental role in the functioning of both natural ecosystems and
18 agrosystems (Stucky et al., 2018). Because plant phenology is influenced by climatic variables and
19 affects plant growth and reproduction, it is key to studies of the consequences of climate change. In
20 agriculture, plant phenology is also particularly useful for the planning of cultivation operations such
21 as planting, fertilizing, irrigating or harvesting (Chmielewski, 2003) and for breeding programs (Meier
22 et al., 2009). Therefore, phenological data are currently collected around the world at an accelerated
23 pace. However, phenological descriptions are often not standardized, making it difficult to make

24 sense of newly collected data in large-scale multispecies comparisons (Stucky et al., 2018). Hence, it
25 is crucial to develop universal phenological scales for all major cultivated species.

26 The *Biologische Bundesanstalt, Bundessortenamt und Chemische Industrie* (BBCH) scale developed
27 for monitoring phenological growth stages is the result of a teamwork conducted by the German
28 Federal Biological Research Centre for Agriculture and Forestry, the German Federal Office of Plant
29 Varieties, the German Agrochemical Association and the Institute for Vegetables and Ornamentals
30 (Bleiholder et al., 1989). This work generated much interest, and Hack et al. (1992) subsequently
31 published the principles of the extended BBCH, a universal scale that works with mono- and
32 dicotyledons. Since 1992, new BBCH extended scales for specific crops have regularly been
33 published, including some for fruit trees: pome fruits and stone fruits (Meier et al., 1994), *Citrus*
34 (Agusti et al., 1995), apricots (Pérez Pastor et al., 2004), mango (Hernández Delgado et al., 2011), or
35 sweet cherry (Fadón et al., 2015). Despite some limitations (Stucky et al., 2018), this method has
36 many advantages for practical applications and has become a standard in agronomy.

37 There are seven consistently recognized chestnut tree species growing in subtropical, Mediterranean
38 and temperate forests from the Northern hemisphere (Pereira-Lorenzo et al., 2012). Some are of
39 special economic importance. In particular, the Chinese chestnut (*C. mollissima*), Japanese chestnut
40 (*C. crenata*) and European chestnut (*C. sativa*) are widely cultivated for their fruits (Pereira-Lorenzo
41 et al., 2012). Chestnut production is mostly concentrated in Asia and Europe (Food and Agriculture of
42 the United Nations, 2020). China is by far the main producer of chestnuts worldwide, with over 1.9
43 million tons of fruits harvested annually. Fruit production in China has tripled in the last two decades.
44 In Europe, chestnut production, which had steadily decreased since the 19th century, has slowly
45 started to recover since the 2010s, currently reaching about 0.15 million tons per year, thanks to a
46 renewed interest of consumers for chestnut products.

47 Chestnut trees are self-incompatible (Stout, 1926; Xiong et al., 2019). Hence, orchards must include
48 enough varieties for successful cross-fertilization. When female flowers of a variety are receptive,

49 male flowers of nearby trees from different compatible varieties must release pollen at the same
50 time for successful pollination, fertilization and fruit production (Solignat and Chapa, 1975). Because
51 there is a wide variation in flowering phenology across varieties, a knowledge of that variation is
52 essential for the design of productive orchards. Chestnut trees are monoecious and have a complex
53 flowering phenology with two separate peaks of pollen emission (Stout, 1928; Hasegawa et al.,
54 2017). To monitor chestnut phenology, it is therefore critical to develop an effective method taking
55 into account these biological features.

56 Solignat and Chapa (1975) have proposed the first phenological scale for chestnuts growth stages.
57 This system has been widely used by chestnut breeders and germplasm curators to screen and
58 characterize chestnut cultivars phenology and for establishing plant variety rights in Europe using
59 harmonized descriptions of new varieties fulfilling criteria of distinctness, uniformity and stability, as
60 defined in Kiewiet (2005). Badeau et al. (2017) then proposed a very simplified BBCH scale for
61 chestnut to be used in a citizen science program, in which they monitored only male flowers. Here
62 we propose a complete phenology scoring system combining Solignat and Chapa (1975) stages and
63 the international BBCH system to facilitate comparisons across studies thanks to a uniform coding
64 system of phenologically equivalent growth stages in plants (Hack et al., 1992).

65

66 [Materials and methods](#)

67 [Development of a BBCH scale](#)

68 The BBCH scale is a decimal code (from 00 to 99) divided into principal and secondary growth stages.
69 An arithmetically greater code always indicates a plant at a later growth stage (Meier et al., 2009).
70 The first digit corresponds to the main growth stage common to all plants. It allows comparisons
71 between different crops, including mono- and dicotyledons. These stages begin from stage 0:
72 “Germination / sprouting / bud development” and end with stage 9: “Senescence, beginning of

73 dormancy” (Hack et al., 1992). The second digit corresponds to secondary growth stages, i.e. short
74 developmental stages characteristics of the studied plant species. These secondary growth stages,
75 also coded from 0 to 9, can represent percentages or average developmental stages: for example,
76 stage 5 could represent a plant with 50% of flowers open or a plant with a relatively high proportion
77 of unfolded leaves (Meier, 2001). If two or more growth scales are used to describe separate
78 phenological events proceeding in parallel, such as the phenology of male and female flowers in
79 monoecious species, they are separated by a slash.

80 To develop a BBCH phenological scale for chestnut trees, we relied on the principal growth stages
81 used for pome fruits and stone fruits by Meier et al. (1994). To further describe chestnut flower
82 development, we identified secondary growth stages and assigned them specific scores. For this
83 purpose, we selected as much as possible phenological scores matching with those proposed by
84 Solignat and Chapa (1975).

85 Chestnut flowering

86 Chestnut trees (genus *Castanea*) have a remarkably complex reproductive system. At the flower
87 level, they are monoecious. Their small female and male flowers are borne on inflorescences called
88 catkins. At the inflorescence level, however, chestnut trees are andromonoecious: they have two
89 types of catkins, unisexual male catkins and bisexual catkins. Bisexual catkins are composed of a few
90 female flowers generally grouped by three at the basis of the catkin. After pollination, they develop
91 into an infructescence composed of a spiny burr enclosing up to three fruits, one per flower. The distal
92 part of bisexual catkins harbors numerous male flowers grouped into small glomerules spirally
93 organized on the catkin. Finally, at the tree level, chestnut trees are either bisexual or unisexual
94 female. In female trees, also called male-sterile trees, male catkins are still present, but their flowers
95 have aborted anthers that produce little or no pollen. Interestingly, these male flowers still produce
96 nectar and attract insects (Pereira-Lorenzo et al., 2017 ; Larue et al., in press).

97 The phenology of chestnuts is particularly complex. Male flowers of unisexual catkins bloom much
98 earlier than male flowers of bisexual catkins, whereas female flowers have a long receptivity period.
99 This rare flowering system (Renner, 2014), first described by Meehan (1879), is called duodichogamy
100 (Stout, 1928). The two peaks of pollen emission do not overlap (Hasegawa et al., 2017) and are very
101 unequal, the first one being an order of magnitude greater than the second one in terms of number
102 of flowers produced and amount of pollen released (Larue et al., in press).

103 [Study site and monitoring](#)

104 The studied trees belong to the INRAE chestnut genetic resources collection (Figure S1). They grow in
105 two nearby orchards located near Bordeaux in southwestern France (44.788319 N, -0.577062 E). The
106 collection includes 117 *C. sativa*, 22 *C. crenata*, 20 *C. mollissima* and 81 interspecific hybrids,
107 including 56 *C. sativa* x *C. crenata* hybrids, some of which belong to popular varieties widely
108 cultivated in the region. All the 240 trees are grafted on two well-known rootstocks: 'Marsol' (CA07)
109 or 'Maraval' (CA74). The first orchard was planted in 1970 and comprises 29 widely spaced trees on
110 2.3 ha. The second orchard was planted in 1990 and includes 211 trees on 3.5 ha.

111 In late spring of 2018, we monitored flowering phenology of all trees twice a week. In 2019 and 2020,
112 we monitored a subset of these trees. We photographed phenological growth stages in the field with
113 APS-C camera (Fujifilm X-T3 and Nikon D500) equipped with a macro lens objective (Fujinon XF 80
114 mm f/2.8 R LM OIS W Macro and AF-S VR Micro-Nikkor 105mm f/2.8G).

115

116 [Results](#)

117 The proposed BBCH phenological scale for chestnut trees includes eight principal growth stages
118 (Table 1). We used three different scores to describe accurately the whole complexity of flowering
119 phenology in this andro-monoecious species. The first score is for male flowers from unisexual

120 catkins. The second score is for female flowers from bisexual catkins. The third score is for male
121 flowers from bisexual catkins. For example, at chestnut tree scored 65/65/59 (Figure 1) corresponds
122 to unisexual catkins with at least 50% of opened male flowers and to bisexual catkins with at least
123 50% of receptive female flowers and with all male flowers still unopened. Note that the scores used
124 for the two types of male flowers are the same. We illustrate these flower developmental stages in
125 Figure 2.



126

127 **Figure 1:** [Color: online only] This flowering shoot is composed of eight unisexual male catkins (black
128 symbols) and two bisexual catkins (white symbols) at the tip. The phenological score that we
129 attributed to this tree on that particular day using the BBCH scale was 65/65/59. On that branch,
130 seven unisexual catkins are at full bloom, whereas the most distal unisexual catkin is just starting to
131 flower (overall BBCH score for unisexual catkins: 65). Each bisexual catkin has a single female
132 inflorescence formed by three female flowers at full receptivity (BBCH = 65) and a short male catkin.
133 Flowers of the male part of the bisexual catkins are still not open, but catkins are already well
134 elongated (BBCH = 59).



135
 136 **Figure 2:** [Color: online only] Illustration of flowering stages. Left pictures: unisexual male catkins. a)
 137 59: Male catkins just before flowering. Catkins have almost their final length and have turned yellow
 138 but their flowers grouped in well-differentiated glomerules are still closed. b) 65: Male catkins from
 139 male-sterile trees at full flowering. Flowers are open but stamens are not visible. c) 65: Male catkins
 140 from male-fertile trees at full flowering. Stamens are visible. d) 67: Male catkins are fading and
 141 turning brown. Central pictures: bisexual catkins. e) 59: Male part of bisexual catkins just before
 142 flowering. f) 65: Male part of bisexual catkins from male-sterile trees at full bloom. Flowers are open,
 143 aborted stamens have short filaments and do not protrude from the flowers. g) 65: Male part of
 144 bisexual catkins from male-fertile trees at full bloom. Flowers are open and stamens have long
 145 filaments. h) 67: Male part of a bisexual catkin that has turned brown. Right pictures: female
 146 inflorescence. i) 61: Only the stigmas of the central flower are visible. j) 63: Stigmas of central flower
 147 are well developed and stigmas of lateral flowers are visible. k) 65: full receptivity. Stigmas of the
 148 three flowers are well developed. l) 67: tips of stigmas from female flowers have turned brown.

149 To assess the phenology of the male flowers from male-sterile trees, we relied on the proportion of
 150 open male flowers instead of estimating the proportion of flowers with conspicuous stamens
 151 emerging from the catkins (Table 1, Stage 5). To evaluate the phenology of female flowers, we
 152 propose two options. One option is to rely on the appearance of diagnostic stages, following Solignat
 153 and Chapa (1975) (Table 1, Stage 6, Female flowers option 1). Alternatively, a semi-quantitative scale
 154 can be used to describe tree receptivity, similar to that used for male flowering (Table 1, Stage 6;
 155 Female flowers option 2).

| BBCH Code | Description | Conversion |
|---|---|-------------------|
| Stage 0: Sprouting/Bud development | | |
| 0 | Dormant buds | A - Af |
| 07 | Beginning of bud break | B |
| 09 | Green leaf tips visible: first green leaf tips just visible | C |
| Stage 1: Leaf development | | |
| 11 | First leaves unfolded | D |
| 15 | More leaves unfolded, not yet at full size | D |
| 19 | All leaves unfolded and fully expanded | DI |
| Stage 3: Shoot development | | |
| 31 | Beginning of shoot growth | |
| 35 | Shoots about 50% of final length | |
| 39 | Shoots about 90% of final length | |
| Stage 5: Catkins growth (unisexual catkins / Female inflorescences / bisexual catkins) | | |
| <i>Male catkins (unisexual or bisexual)</i> | | |
| 50 | Appearance of male catkins | Dm-Da |
| 55 | Glomerules are visible, male catkins grow | |
| 59 | Glomerules well differentiated, male catkins about 90% of final length | Em |
| <i>Female inflorescences</i> | | |
| 50 | Appearance of buds of female inflorescences | Df |
| 55 | Buds of female inflorescences are visible, bisexual catkins grow | |
| 59 | Female inflorescences well differentiated, bisexual catkins about 90% of final length | Ef |
| Stage 6: Flowering (Male flowers of unisexual catkins / Female flowers / Male flowers of bisexual catkins) | | |

Male flowers (unisexual or bisexual catkins)

| | | |
|----|--|---------|
| 60 | First male flowers open | Fm-Fa |
| 61 | Beginning of the flowering: 10-20% of male flowers open | |
| 62 | 20-30% of male flowers open | |
| 63 | 30-40% of male flowers open | |
| 64 | 40-50% of male flowers open | |
| 65 | Full flowering: at least 50% of male flowers open | Fm2-Fa2 |
| 67 | Catkins fading: at least 50% of brown male catkins | Gm-Ga |
| 69 | End of flowering: at least 50% of male catkins have fallen | Hm |

Female flowers (Option 1): Phenotypic stages

| | | |
|----|---|-----|
| 60 | Female flowers visible | |
| 61 | Stigmas of the central flower of the inflorescence visible | Ff |
| 63 | Stigmas of the central flower elongated, stigmas of lateral flowers visible | |
| 65 | Full receptivity: stigmas of three female flowers are well developed | Ff2 |
| 67 | At least 50% of female flowers have brown stigmas | |
| 69 | End of flowering: all female flowers have brown stigmas | |

Female flowers (Option 2): Receptivity

| | | |
|----|--|--|
| 61 | Beginning of the flowering: 10-20% of female flowers are receptive | |
| 62 | 20-30% of female flowers are receptive | |
| 63 | 30-40% of female flowers are receptive | |
| 64 | 40-50% of female flowers are receptive | |
| 65 | Full flowering: at least 50% of female flowers are receptive | |
| 67 | At least 50% of female flowers have brown stigmas | |
| 69 | End of flowering: all female flowers have brown stigmas | |

Stage 7: Burr development

| | | |
|----|---|---|
| 72 | Involucre is 3x larger than when the female inflorescence was receptive | I |
| 75 | Burrs about 50% of final volume | J |
| 79 | Burrs about 90% of final volume | J |

Stage 8: Fruit maturity

| | | |
|----|--|-------|
| 81 | Burrs turn brown | K |
| 83 | First burrs open | Lo |
| 85 | At least 50% of burrs open | Mo |
| 87 | At least 50% of chestnuts/burrs fallen | N - O |
| 89 | All chestnuts/burrs fallen | N - O |

Stage 9: Leaf senescence

| | | |
|----|---|----|
| 90 | Leaves begin to discolor or start to fall | Dj |
| 91 | About 10% of leaves discolored or fallen | Dz |

| | | |
|----|--|----|
| 95 | About 50% of leaves discolored of fallen | Dz |
| 97 | All leaves fallen | Dz |

156

157 **Table 1:** Phenological growth stages of chestnuts according to the BBCH scale and conversion from
 158 Chapa and Solignat (1975) scale.

159

160 We successfully applied this new BBCH scale to the trees from INRAE chestnut genetic resources
 161 collection during three years. To briefly illustrate the type of results obtained, mean date, minimum
 162 date and maximum date for onset of full flowering of the two types of male catkins are provided in
 163 Table 2 for measures performed in 2018 on all trees, distinguishing the three pure species and one
 164 class of hybrids. On average, male flowers from bisexual catkins reach full bloom about 10 days after
 165 male flowers from unisexual catkins. Date of flowering of male catkins vary slightly among species
 166 but greatly within each species. Overall, for unisexual catkins, 25 days separate the earliest and the
 167 latest chestnut trees, whereas for bisexual catkins, 20 days separate the earliest and the latest trees.

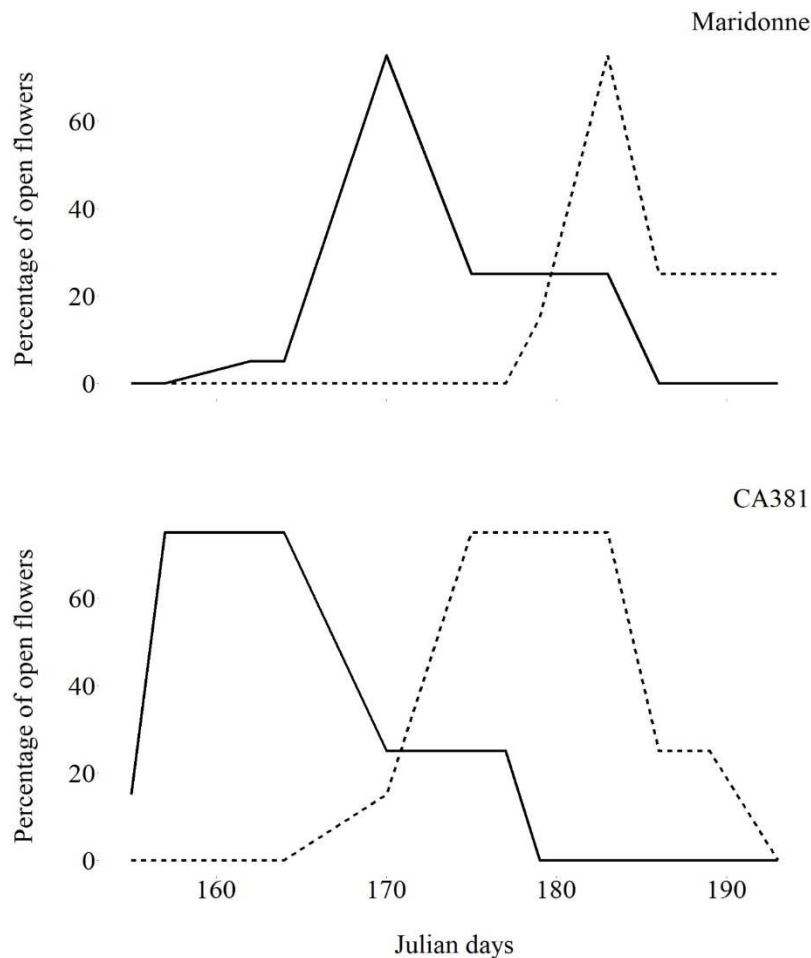
| Type | N | Unisexual male catkins (BBCH = 65) | Male flowers of bisexual catkins (BBCH = 65) |
|-------------------------------|-----|---------------------------------------|---|
| <i>C. sativa</i> | 117 | 163.8 (155.5-177) | 173.7 (165.5-182) |
| <i>C. sativa x C. crenata</i> | 56 | 161.3 (155.5-170) | 172 (162-180) |
| <i>C. crenata</i> | 22 | 160.2 (152-165.5) | 169.9 (162-178.5) |
| <i>C. mollissima</i> | 20 | 159.8 (155.5-170) | 173.2 (170-177) |

168 **Table 2:** Date in Julian days of onset of full flowering for male catkins in 2018 in the INRAE chestnut
 169 collection, according to chestnut species.

170

171 To illustrate the dynamic of chestnut flowering, we provide an example of the flowering phenology of
 172 two trees from two different varieties in year 2018, illustrating the two peaks of male flowering in
 173 this duodichogamous species (Figure 3).

174



175

176 **Figure 3:** Male flowering phenology of two trees of the 'Maridonne' and CA381 varieties in year
 177 2018. The x-axis is expressed in Julian days and the y-axis in percentage of open male flowers.
 178 Phenology of male flowers from unisexual catkins is represented by a continuous line and phenology
 179 of male flowers from bisexual catkins is represented by a dotted line. CA381 flowers two weeks
 180 earlier than Maridonne and its pollen emission lasts longer. Percentage of open flowers can be
 181 estimated from the BBCH scale, with the scores 60, 61, 62, 63, 64, 65 and 67 representing
 182 respectively roughly 5%, 15%, 25%, 35%, 45%, 75% and 25% of open flowers.

183

184 Discussion

185 In principle, to model accurately the phenology of a studied species, one should rely on a complete
 186 ontology of phenological traits to be measured exhaustively at the whole plant level (Stucky et al.,
 187 2018). However, this can quickly become time consuming. Diagnosing phenological stages using pre-

188 established scores, as performed with the BBCH scale, is much faster, making it possible to monitor
189 many more trees. We chose that latter strategy to compare varieties in orchards.

190 We successfully tested this new BBCH scale on trees from three chestnut species and their hybrids.
191 Given the great homogeneity of the genus, it is likely that other chestnut species, including the
192 American chestnut (*C. dentata*), can be scored with the proposed system. Furthermore, the proposed
193 chestnut BBCH scale is suitable for both male-fertile and male-sterile trees. On male flowers from
194 male-fertile trees, the stamens are conspicuous and easily scored. For male-sterile trees, the aborted
195 stamens do not emerge out of the flower so a closer look on the opening of male flowers themselves
196 is required.

197 The scoring of the phenology of female flowers can be adapted to meet different objectives. A simple
198 approach is to monitor what happens at the scale of inflorescences, allowing comparison with
199 Solignat and Chapa (1975) phenological stages. Instead, if the objective is to study the temporal
200 compatibility between pollen emission and female flower receptivity, we recommend evaluating the
201 percentage of receptive female flowers using class intervals. To investigate in even more details the
202 mating system of the species, a more rigorous but more labor-intensive approach is to monitor and
203 track individually a sample of flowers on each tree, as performed by Hasegawa et al. (2017).

204 To the best of our knowledge, this study is the first to use phenological stages based on the BBCH
205 scale in chestnut. Despite some limitations inherent in the approach used, this scale allows a rapid
206 semi-quantitative assessment of the growth stages of the three types of flowers found in this tree,
207 making it possible to gather precious phenological knowledge on all chestnut species worldwide.

208

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218 Photo credits

219 Photography of Figure 1 and of the two insets of Figure 2 are by RJP, all others are by CL.

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