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1 **Central American and Caribbean population history of the *Pseudocercospora fijiensis***  
2 **fungus responsible for the latest worldwide pandemics on banana**

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12

13

14 **Abstract** Among the emerging fungal diseases threatening food security, the  
15 *Pseudocercospora fijiensis* fungus causing black leaf streak disease of banana is one of the  
16 most marked examples of a recent worldwide pandemic on a major crop. We assessed how  
17 this pathogen spread throughout the latest invaded region, i.e. Central America and the  
18 Caribbean. We retraced its population history combining detailed monitoring information on  
19 disease outbreaks and population genetic analyses based on large-scale sampling of *P.*  
20 *fijiensis* isolates from 121 locations throughout the region. The results first suggested that  
21 sexual reproduction was not lost during the *P. fijiensis* expansion, even in the insular  
22 Caribbean context, and a high level of genotypic diversity was maintained in all the  
23 populations studied. The population genetic structure of *P. fijiensis* and historical data  
24 showed that two disease waves swept northward and southward in all banana-producing  
25 countries in the study area from an initial entry point in Honduras, probably mainly through  
26 gradual stepwise spore dispersal. Serial founder events accompanying the northern and  
27 southern waves led to the establishment of two different genetic groups. A different  
28 population structure was detected on the latest invaded islands (Martinique, Dominica and  
29 Guadeloupe), revealing multiple introductions and admixture events that may have been  
30 partly due to human activities. The results of this study highlight the need to step up  
31 surveillance to limit the spread of other known emerging diseases of banana spread mainly by  
32 humans, but also to curb gene flow between established pathogen populations which could  
33 increase their evolutionary potential.

34

35 **Keywords:** Emerging disease, fungal plant pathogen, population history, dispersal, *Musa*,  
36 *Pseudocercospora fijiensis*.

37

## 38 **1. Introduction**

39

40 Of all plant pathogens, fungi are a highly serious threat to crops (Almeida et al., 2019). Global  
41 trade and large-scale standardized farming are conducive to fungal disease dispersal and  
42 establishment worldwide (McDonald and Stukenbrock, 2016). Thus many so-called  
43 ‘emerging plant diseases’ are caused by pathogens that have increased their geographical  
44 range and become invasive in new and wide areas (Anderson et al., 2004). Studying diseases  
45 that have recently emerged may help improve surveillance (Parnell et al., 2017) and they may  
46 also be useful models for understanding evolutionary and ecological processes involved in  
47 pathogen populations, such as dispersal, reproduction mode, population bottlenecks,  
48 admixture and changes in phenotypic traits (Stukenbrock and McDonald, 2008). It is first  
49 essential to reconstruct the history of pathogen populations during range expansions when  
50 documenting these processes in emerging diseases (Gladieux et al., 2015).

51

52 The detection dates of a given emerging disease across geographical areas could first be  
53 documented to retrace the history of pathogen populations. This spatiotemporal or historical  
54 data can be used to estimate the speed of past emergences, which is a key aspect for  
55 understanding and reacting to future invasive species (Evans, 2016). However, historical data  
56 are not informative enough to retrace the population history and identify the epidemiological  
57 and demographic processes that have accompanied the spread of emerging fungal diseases.  
58 Population genetics are often a valuable complementary approach for gaining clearer insight  
59 into the population biology of a fungal invasion (review in Gladieux et al., 2015; McDonald  
60 and Stukenbrock, 2016). Such studies have notably shown that, although fungi can be  
61 dispersed naturally through spore production, humans also play a role in the spread of  
62 diseases via the transport of agricultural products or infected plant materials. This role seems  
63 most evident between continents because it is harder for pathogens to travel several thousand  
64 kilometres by natural means alone. Yet it is not easy to disentangle human-mediated  
65 introductions from natural dispersal on smaller geographical scales. Several demographic or

66 evolutionary processes often accompany fungal invasions, which may promote adaptations  
67 such as population bottlenecks, or admixture between populations following multiple  
68 introductions. Lastly, changes in life history traits during fungal invasions have been  
69 described and the loss of sexual reproduction is one of the most frequent.

70

71 Banana—a major high-value crop produced in tropical regions—is threatened by several  
72 pathogens (Guzman et al., 2019), including the fungus *Pseudocercopora fijiensis* which  
73 causes black leaf streak disease (BLS). This emerging disease has spread from Asia  
74 throughout the entire intertropical zone in the last 50 years and is considered to be one of the  
75 most serious threats to food security (Pennisi, 2010). In some areas, over 50 fungicides are  
76 applied annually to control the disease, which obviously has a major impact on human health  
77 and the environment (de Lapeyre de Bellaire et al., 2010).

78 *P. fijiensis* is a heterothallic ascomycete fungus that produces two types of spores. Conidia  
79 (from asexual reproduction) disperse over scales of only a few metres, whereas ascospores  
80 (from sexual reproduction) are easily wind-dispersed over an average distance of 200 m, but  
81 with a fat-tailed dispersal kernel that may extend several kilometres (Rieux et al., 2014).

82 Sexual and asexual reproduction concomitantly prevail in banana plantations (Guzman et al.,  
83 2019). Human-mediated introductions may also accelerate regionwide disease spread through  
84 the movement of infected leaf material (planting or packing materials, (Churchill, 2011). In a  
85 continental context, sexual reproduction is not lost during invasion and—although genetic  
86 discontinuity indicative of some stochastic long-distance dispersal events has been  
87 reported—the genetic structure of *P. fijiensis* populations on a country scale suggest that the  
88 disease is spread mainly by gradual stepwise spore dispersal (Halkett et al., 2010). Supporting  
89 this observation, the rate of *P. fijiensis* spread estimated from the spatiotemporal data  
90 published in (Halkett et al., 2010) was close to that predicted under certain conditions by a  
91 modelling approach accounting for both asexual and sexual reproduction (Hamelin et al.,  
92 2016).

93 In the Americas, BLSD was first detected in Honduras in 1972 and was most probably  
94 introduced by humans (Robert et al., 2012; Robert et al., 2015), spreading northwards and  
95 southwards and then reaching the Caribbean in 1990 (Guzman et al., 2019). Despite  
96 surveillance networks set up in the French West Indies (Teycheney et al., 2009), the last  
97 disease-free islands of Martinique, Dominica and Guadeloupe were reached and infested by  
98 BLSD from 2010 onwards (Ioos et al., 2011). This situation offered an opportunity to monitor  
99 a recent fungal invasion in an insular context, which could provide new information on the  
100 population biology of such organisms.

101  
102 This study set out to retrace the population history of *P. fijiensis* throughout Central America  
103 and the Caribbean to gain insight into the epidemiological and demographic processes that  
104 have accompanied its spread in this latest invaded region. To that end, we gathered detailed  
105 monitoring information on disease outbreaks within the region. At the same time, we  
106 conducted a population genetic analysis using large-scale sampling throughout all banana-  
107 producing countries of mainland Central and South America and the Caribbean Basin. We  
108 used these data to address the following questions: What were the BLSD invasion routes  
109 throughout Central America and the Caribbean Islands? Was the invasion speed slower in the  
110 Caribbean insular context in comparison to mainland areas? Did successive bottlenecks  
111 accompany this invasion? Did the expansion have an impact on sexual reproduction when the  
112 fungus passed from island to island in the Caribbean? Could this analysis generate insight into  
113 the potential invasion drivers, e.g. natural fungal dispersal by the wind, or human activities?

114

## 115 **2. Materials and Methods**

### 116 *2.1. Historical data analysis*

117 Historical *P. fijiensis* invasion data were collected in 12 Caribbean banana-producing  
118 countries. We noted the date and geographical coordinates (decimal units for latitude and  
119 longitude) for the locations corresponding to the first, middle and last detection of the disease

120 in each country (Supplementary Dataset 1). The historical dataset was divided into two  
121 subsets corresponding to a northern front and a southern front. The northern front was  
122 represented by five countries (Cuba, Jamaica, Haiti, Dominican Republic, Puerto Rico)  
123 mainly distributed along a longitudinal east-west gradient and the southern front by seven  
124 countries (Trinidad, Grenada, St Vincent, St Lucia, Martinique, Dominica, Guadeloupe)  
125 mainly distributed along a latitudinal north-south gradient. The time of occurrence in each  
126 location was calculated as the time between the date of detection in the location and the date  
127 of the first appearance in the northern or southern Caribbean, i.e. the Esmeralda location in  
128 Cuba (21.850°N, -78.100°W) and Coromandel location in Trinidad (10.117°N, -61.800°W),  
129 respectively. These locations were considered as the origin of disease spread in the two  
130 Caribbean areas. A linear model between the occurrence time and geographical distance from  
131 the origin of each front was tested using the `lm` function of R 3.6.0 ([http://www.R-](http://www.R-project.org)  
132 [project.org](http://www.R-project.org)). The Euclidian distance was chosen because the study locations for each of the  
133 fronts were more or less distributed along the same line (from west to east for the northern  
134 front and from south to north for the southern front). The speed of the invasion waves was  
135 calculated as the inverse of the regression slope.

136

## 137 *2.2. Fungal material sampling and DNA extraction*

138 Leaf samples were collected from most banana-growing countries in mainland tropical  
139 of Central, South America and the Caribbean Basin (Table1, Supplementary Dataset 1).  
140 Overall, 2,188 fungal individuals from 121 different geographical locations and 19 countries  
141 were analysed. The host plants were susceptible clones of Cavendish banana and plantain  
142 cultivars. The samples from Central and South America (Mexico, Honduras, Costa Rica,  
143 Panama, Columbia and Venezuela) had been previously collected between 1991 and 2009 for  
144 the purposes of the study of Robert et al. (2012). All of the other Caribbean samples were  
145 collected between 2009 and 2014 for this study. Overall, the samples first included 55  
146 populations from 19 countries. The sampling effort was mainly focused on the Caribbean  
147 islands (46 populations), including three populations collected from three different locations

148 per island, and even more were collected from two recently contaminated islands in the  
149 French West Indies (14 populations in Martinique and 6 in Guadeloupe). The populations  
150 consisted of around 18 to 60 fungal isolates collected simultaneously from different plants in  
151 the same banana cropfield. More intensive sampling was carried out in Martinique and  
152 Guadeloupe for the purpose of a landscape genetic analysis. About 397 isolates were collected  
153 from 27 locations in Guadeloupe and around 680 isolates were collected from 58 locations in  
154 Martinique (3 to 17 isolates per location) based on a two-dimensional sampling scheme  
155 designed to cover the two islands entirely (Supplementary Dataset 1).

156 Most of the populations were composed of fungal isolates grown in the laboratory. In  
157 this case, each individual corresponded to a strain derived from a single spore (sexual or  
158 asexual spores, depending on the case) isolated from an infected banana leaf. Growing  
159 mycelium resulting from spore germination was placed on solid medium (300 mL V8, 3 g  
160 CaCO<sub>3</sub>, 20 g/L agar, pH 6) for 2 weeks at 25°C, before being dried for 2 days at 55°C and  
161 ground. DNA was extracted from growing mycelium as described in (Halkett et al., 2010).  
162 For some populations, young lesions (“leaf streak”) were cut from banana leaves, dried for a  
163 day at 55°C and stored at –20°C. Each lesion was considered to have been caused by a single  
164 fungal strain. DNA was directly extracted from the sampled lesions using the protocol  
165 described in Robert et al. (2010).

166

### 167 *2.3. Microsatellite marker genotyping*

168 All individuals were genotyped using 16 microsatellite markers. These markers had  
169 already been used in other studies (Robert et al., 2010; Zapater et al., 2008). They were  
170 combined in three multiplex panels of four markers for the first panel, and six markers for the  
171 other two panels (Table S1, Supplementary file 1). PCR amplification and genotyping were  
172 carried out as described in (Robert et al., 2010). The genotype was obtained for at least 10  
173 markers for all of the studied isolates, which was largely enough for genetic assignment  
174 (Arthofer et al., 2018).

175



176 2.4. Linkage disequilibrium and genetic diversity analysis

177 Multi-locus gametic linkage equilibrium was estimated using the standardized index of  
178 association  $\bar{r}_d$  (Agapow and Burt, 2001) and tested with the *poppr* R-package (Kamvar et al.,  
179 2014). Gametic linkage disequilibrium between all pairs of loci was also tested in all  
180 populations using Fisher's exact tests implemented in the *genepop* R-package with default  
181 parameters (Rousset, 2008), and the false discovery rate (FDR) procedure implemented in the  
182 *qvalue* R-package version 2.12.0 (<http://github.com/jdstorey/qvalue>).

183 Genetic variation within the 55 populations with more than 18 isolates was estimated  
184 according to the number of polymorphic markers (out of 16), the number of multi-locus  
185 genotypes (MLG), and by 3 descriptive indices averaged across markers: gene diversity index  
186 (equivalent to the mean expected heterozygosity  $H_e$  (Nei, 1987), Simpson's index  $\lambda$  (Hill,  
187 1973) computed using the *poppr* R-package (Kamvar et al., 2014) and the mean allelic  
188 richness ( $A_r$ ), calculated using the rarefaction method implemented in the *hierfstat* R-package  
189 (Goudet, 2005). The cumulative geographic distance from the Honduras population HND3  
190 (close to the site where the epidemic was assumed to have started in the Americas) was  
191 calculated for each population following a point-by-point pathway northward and southward.  
192 The correlation between that geographic distance and  $A_r$  or  $H_e$  was tested using the *lm*  
193 function of R 3.6.0 (<http://www.R-project.org>).

194 The BOTTLENECK program (Piry et al., 1999) was used to detect a potential signal of  
195 recent population bottlenecks based on the detection of excess heterozygosity (in the sense of  
196 Nei's (1987) gene diversity) compared to heterozygosity expected under the mutation-drift  
197 equilibrium assumption, from the observed number of alleles. The expected heterozygosity  
198 distribution was estimated based on 1000 replications assuming a two-phase mutation model  
199 (TPM: 70% stepwise mutation model (SMM), 30% IAM)), as recommended for  
200 microsatellite markers (Dirienzo et al., 1994). We then used the Wilcoxon significance test to  
201 determine whether loci displayed significant excess heterozygosity.

202

203 2.5. Population structure analyses

204 An admixture analysis was conducted by estimating the individual ancestral coefficients  
205 with the sNMF algorithm implemented in the LEA R-package (Frichot and François, 2015).  
206 This approach is similar to Bayesian clustering programs, such as STRUCTURE (Pritchard et  
207 al., 2000). The K value representing the number of ancestral populations ranged from 1 to 10,  
208 and 10 independent runs were performed for each K value. The number of ancestral  
209 populations that best explained the genotypic data was chosen based on the cross-entropy  
210 criterion. This criterion evaluates the quality of fit of the statistical model to the data using a  
211 cross-validation technique (Frichot and François, 2015). The maximum local contribution to  
212 ancestry was spatially interpolated at each geographical map point. Barplots were drawn up  
213 by taking the ancestry coefficient estimated for individuals or averaged for individuals  
214 belonging to the 55 populations presented in Table 2. Principal components analysis (PCA) of  
215 these populations was performed with the dudi.pca function using the R software ade4  
216 package (Dray and Dufour, 2007).

217

218 To further evaluate the population structure and the degree of differentiation on different  
219 scales, we used an analysis of molecular variance (AMOVA) framework, as implemented in  
220 the HierFstat R-package (Goudet, 2005). The significance of the F-statistics estimated on  
221 different geographical levels was tested using 1,000 permutations between units. The 95%  
222 confidence intervals of these F-statistics were computed by bootstrapping over loci 1,000  
223 times. Pairwise *F<sub>st</sub>* (Weir and Cockerham, 1984) were also estimated and tested (Fisher exact  
224 test) using the *Genepop* R-package. Genetic distances between populations were estimated  
225 using the Cavalli-Sforza chord distance and represented using a neighbour-joining tree, as  
226 implemented in POPULATIONS1.2.31 software ([www.bioinformatics.org/~tryphon/  
227 populations/](http://www.bioinformatics.org/~tryphon/populations/)).

228

## 229 2.6. Approximate Bayesian Computation (ABC) model choice analyses

230 A model choice analysis was carried out using the ABC random forest (ABC-RF)  
231 methodology developed by Pudlo et al. (2016), to test scenarios of *P. fijiensis* introduction

232 into Martinique. An ABC model choice analysis involves a comparison of observed data with  
233 data simulated under the evolutionary scenarios to be compared. Like Fraimout et al. (2017),  
234 we carried out two analyses with two different sets of populations as representatives of their  
235 genetic cluster (e.g. the Honduras ancestral population, the northern group, the southern group  
236 and the two populations from Martinique; Table S2, Supplementary File 1).

237 For each of the three scenarios tested (Fig. S6, Supplementary File 1; see results for a  
238 description), we simulated 60,000 datasets (20,000 per scenario) using DIYABC v2.1.0  
239 software (Cornuet et al., 2014). Parameters were drawn up from the prior distributions  
240 described in Table S3 (Supplementary File 1). Time was expressed as numbers of  
241 generations, assuming 10 successive generations per year (8). Historical data were used to  
242 define the bounds of the introduction events. A generalized stepwise mutation model as  
243 implemented in the software was used to simulate mutations at microsatellite markers.  
244 Observed and simulated datasets were summarized using all the one-sample and two-sample  
245 statistics for microsatellite markers, as well as the linear discriminant analysis (LDA) axes,  
246 calculated in DIYABC v2.1.0 (Cornuet et al., 2014). The quality of the simulated models  
247 (scenarios and priors) was checked using PCA on the space of the summary statistics, as  
248 implemented in DIYABC v2.1.0 (Cornuet et al., 2014) (Fig. S7, Supplementary File 1).  
249 After simulating the datasets, we carried out an ABC-RF model choice analysis using the  
250 `abcrf` R-package (Pudlo et al., 2016). An ABC-RF analysis provides a classification vote for  
251 competing scenarios where the number of times a scenario is selected represents the model  
252 best fitting the dataset from a forest of 900 trees, as well as the posterior probability of the  
253 best selected model. The consistency of the ABC-RF model choice analysis was ensured by  
254 replicating the simulations and ABC-RF analysis 10 times and calculating the prior error rate  
255 (Table S4, Supplementary File 1). The number of votes and the prior error rate for each  
256 scenario, as well as the posterior probability of the best selected model, were averaged over  
257 the 10 replicates. Preliminary analyses using different numbers of datasets per scenario were  
258 carried out to ensure that we used a sufficient number of datasets and of trees for the random  
259 forest analyses.

260

### 261 3. Results

262 Overall, 2,188 fungal individuals from 121 different geographical locations were  
263 isolated (Table 1, Fig. 1A). The isolates were genotyped using 16 microsatellites. Historical  
264 records were concurrently obtained from various locations (Table 1, Fig. 1A and  
265 Supplementary Dataset 1). These data were used to conduct statistical and population genetic  
266 analyses.

267

#### 268 3.1. Random mating reproduction mode in all invasive populations

269 A within-population genetic analysis of 55 populations with more than 18 isolates  
270 suggested the existence of sexual reproduction and random mating in all populations studied  
271 (Table 2). Overall, 1,354 multi-locus genotypes (MLGs) were detected for 1,617 isolates.  
272 Around 12.5% (170/1,354) of the total number of MLGs were repeated in the whole sample.  
273 Of these repeated MLGs, 68.2% (116/170) were found only twice in the whole sample.  
274 Around 37.6% (64/170) were repeated within and 62.4% (106/170) between populations.  
275 Thus, most of the repeated MLGs were found only twice overall and were distributed in two  
276 different populations. Around 38.2% (65/170) of the repeated MLGs included at least one  
277 isolate which was cloned from ascospores derived from sexual reproduction. Taken together,  
278 these observations did not enable us to conclude that isolates having the same MGLs were  
279 clones. Due to the limited number of markers used, in most cases they may have  
280 corresponded to MLGs that had been sampled twice by chance even in a sexual reproduction  
281 setting. Regardless, the presence of repeated MLGs had little impact on the population  
282 structure observed and the number of MLGs detected in each population was very close to the  
283 total number of isolates sampled (Simpson's index  $> 0.93$ ). Furthermore, in all the  
284 populations, no pairs of loci were found to be in significant linkage disequilibrium using  
285 Fisher's exact test. The multilocus association index  $r_d$  was significantly not null in 42%  
286 (23/55) of the populations and 16% (9/55) of the populations, at 5% and 1% FDR,  
287 respectively. However,  $r_d$  was significantly and negatively correlated with the gene ( $r=-0.46$ ,

288  $p=2.9 \cdot 10^{-4}$ ) and genotypic diversity index ( $r=-0.53$ ,  $p=2.5 \cdot 10^{-5}$ ; Fig. S1, Supplementary File  
289 1), suggesting that some linkage disequilibrium may have resulted from bottleneck effects, as  
290 described in theoretical studies (Rogers, 2014).

291

### 292 *3.2. Two epidemic waves from Honduras with serial founder effects shaping the population* 293 *structure*

294 The chronology of the first disease occurrence dates in the different countries and  
295 locations (Table 1, Fig. 1A) suggested that *P. fijiensis* spread via two separate epidemic  
296 waves throughout the Central America-Caribbean Basin region. Starting from Honduras in  
297 1972, a northern wave followed a route from Guatemala to Porto Rico (first detection in  
298 2004), while a southern wave followed a route from Nicaragua to Guadeloupe (first detection  
299 in 2012). In the Caribbean, significant linear regressions were obtained between the  
300 occurrence times and the distances from the point considered as being the origin for each  
301 wave ( $R^2=0.77$  and  $0.85$  and  $p = 1.6e-05$  and  $1.9e-09$ , respectively, for the northern and  
302 southern waves, Fig. 2). The speed of invasion was calculated as the inverse of the slope of  
303 each linear regression and was very close for the two waves, with  $6.8$  km/month ( $81$  km/year)  
304 for the northern wave and  $6.7$  km/month ( $80$  km/year) for the southern wave.

305

306 The population genetic structure of *P. fijiensis* in America was found to be in accordance with  
307 historical data and showed that serial founder events accompanying the northern and southern  
308 waves had led to the establishment of different genetic groups. The highest levels of allelic  
309 richness ( $A_r$ ) and Nei's gene diversity ( $H_e$ ; (Nei, 1978) were detected in Honduras, where the  
310 disease was first introduced in the area (Table 2). Significant signatures of bottlenecks on the  
311 distribution of allele frequencies were observed in  $34\%$  ( $19/55$ ) and  $16\%$  ( $9/55$ ) of the  
312 populations, at  $5\%$  and  $1\%$  FDR, respectively (Table 2). The number of microsatellite  
313 markers and sample sizes used may have been too low to have sufficient statistical power for  
314 tests on some samples (Peery et al., 2012). To further highlight the bottleneck effects, gene

315 diversity in each location, as estimated by the  $A_r$  or  $H_e$  parameter, was plotted against the  
316 cumulative step-by-step geographical distance from Honduras following the migration waves  
317 (Fig. 3). A significant negative correlation was detected when jointly considering the northern  
318 and southern waves and genetic parameters. It was found that the population gene diversity  
319 decreased with the distance from Honduras.

320 Individual ancestry coefficients were first estimated using the sNMF algorithm (Frichot and  
321 François, 2015). The cross-entropy curve suggested a  $K$  value of 3 to 4, which could be  
322 considered as representative of major ancestral populations (Fig. S2, Supplementary File 1).  
323 The study populations were then considered to have derived from this ancestral gene pool and  
324 the ancestry coefficients gave the proportions of individual genomes originating from the  
325 different components of the pool. For a clear display of the results, barplots for the 55  
326 populations with more than 18 isolates were drawn up from the mean values of the ancestry  
327 coefficients with  $K$  ranging from 3 to 9 (Fig. S3, Supplementary File 1). The major population  
328 structure in the study area was actually observed from a value of  $K=4$  and a barplot of  
329 individual ancestry coefficients for this value is also presented in Fig. S4 (Supplementary File  
330 1). To highlight the population genetic structure, the maximum value of the ancestry  
331 coefficients for each isolate was spatially interpolated (Fig. 1B), and a barplot of the mean  
332 values of the ancestry coefficients for  $K=4$  and for 55 populations with more than 18 isolates  
333 is presented below (Fig. 1 C), as well as a neighbour-joining tree constructed from the genetic  
334 distance (Fig. 1 D). These analyses highlighted the existence of three major genetic groups  
335 corresponding to three geographical areas: a central group contained populations located in  
336 Central America, Colombia and Jamaica; a second northern group contained populations  
337 located in the northern Caribbean, from Cuba to Puerto Rico (there was some sub-structuring  
338 in this group); and a third, fairly uniform, southern group contained populations from  
339 Venezuela to the southern Caribbean islands to as far as Saint Lucia. PCA analysis revealed a  
340 very close structure (Fig. S5, Supplementary File 1).

341  $F$ -statistics were estimated at different hierarchical levels using AMOVA, while excluding  
342 populations from Martinique, Dominica and Guadeloupe (Table 3). Three levels were

343 considered: populations, countries and groups corresponding to the one defined from the  
344 clustering analysis. Significant values were obtained at all the levels tested. However, a larger  
345 proportion of the genetic diversity was distributed between groups ( $F=0.23$ ) rather than  
346 between countries within groups ( $F=0.05$ ). Genetic differentiation between populations within  
347 groups and within countries was similar ( $F=0.16$  and  $0.12$ , respectively). These estimations  
348 seemed to indicate that the groups were genetically quite homogeneous and different from  
349 each other.

350

### 351 *3.3. Long-distance introduction and admixture events*

352 We found a different population structure on islands consecutively located north of  
353 Saint Lucia (Martinique, Dominica and Guadeloupe), revealing multiple introductions and  
354 admixture events (Fig. 1). Based on some preliminary results, dense two-dimensional  
355 sampling was carried out in Martinique and Guadeloupe (Table 1, Fig. 1A), which enabled us  
356 to map the introduced genetic groups. Some individuals and populations from both ends of  
357 Martinique were related to the southern genetic group, while other samples located in the  
358 centre of the island were related to the northern genetic group, thus suggesting that there had  
359 been at least two independent introductions on that island (Fig. 1B and 1C). Three populations  
360 from Martinique (MTQ2, MTQ3, MTQ13) were grouped on the middle branches of the  
361 tree presented in Fig. 1 D. Geographically, these samples were located at junctions between  
362 differentiated populations belonging to the northern and southern genetic groups and we  
363 suspected some admixtures between them.  $F$ -statistics between populations in Martinique  
364 belonging to different genetic groups were sometimes very high, with a maximum at  $F = 0.51$   
365 (Table 3), and the highest values were obtained when comparing populations from the ends of  
366 Martinique (MTQ1 and MTQ14) with all the others (Supplementary Dataset S1). The PCA  
367 projection of the populations from Martinique, Dominica and Guadeloupe were either close to  
368 one of the three major groups (southern, northern and central) or between these groups, thus  
369 supporting the idea that there have been multiple introductions and admixture events on these  
370 islands (Fig. S5, Supplementary File 1). Based on this descriptive analysis, three different

371 scenarios for *P. fijiensis* introduction into Martinique were tested using an ABC procedure.  
372 The first scenario assumed two independent introductions into Martinique from two  
373 populations belonging to the northern and southern migration waves (Fig. S6, Supplementary  
374 File 1). Two other scenarios assumed a single introduction event from one population  
375 belonging to the northern or southern migration wave, with the second population from  
376 Martinique emerging from the first one after its establishment. In all the scenarios, Honduras  
377 was considered as the ancestral population and all the founded populations came through a  
378 bottleneck when introduced. In the first scenario, we considered that genetic differentiation  
379 between Martinican populations was the result of the combination of bottleneck effects and  
380 introductions from different origins, whereas in the other two scenarios genetic differentiation  
381 resulted only from bottleneck effects. The former scenario was found to be the most probable  
382 using two different sample sets (mean posterior probability of  $0.95 \pm 0.008$ ; Table S4,  
383 Supplementary File 1).

384 Populations from Dominica and Guadeloupe showed intermediate patterns for ancestry  
385 coefficients (Fig. 1 C), and were all located in the middle of the tree in Fig. 1D, except  
386 population GLP5, which was more related to the southern genetic group. This distinct  
387 population had the lowest genetic diversity level in Guadeloupe (Table 2) and a very high  
388 level of genetic differentiation between all the other populations from Guadeloupe ( $0.27 < F$   
389  $< 0.41$ ,  $p < 0.0001$ ; Supplementary Dataset 1). Thus, population GLP5 may have stemmed from  
390 a rare intense bottleneck and/or an independent introduction from the southern wave. It was  
391 not included in further analysis so as to be able to highlight the general trends. The range of  
392 pairwise  $F$ -statistics between populations from Guadeloupe and Dominica was lower or very  
393 close to that obtained between populations within countries in the central, northern or  
394 southern genetic groups ( $F < 0.14$ , Table 3 and Supplementary Dataset 1). Taken together, all  
395 of the above observations suggested that the populations in Guadeloupe and Dominica were  
396 founded from an admixture between the northern and southern genetic groups.



397 The correlations between genetic diversity (estimated by  $A_r$  and  $H_e$ ) and the cumulative  
398 geographical distance from Honduras for samples from Martinique, Dominica and  
399 Guadeloupe were significantly positive (Fig. 3), thus indicating that genetic diversity had  
400 increased on those islands, in contrast with samples from other countries. The estimated  
401 values of both parameters for the furthest samples from Honduras located in Guadeloupe were  
402 close to those obtained from Honduran samples. These results strongly indicated that there  
403 had been multiple introductions and/or admixtures on these islands.

404

#### 405 **4. Discussion**

406 This study set out to gain insight into epidemiological and demographic processes that have  
407 accompanied the spread of the *P. fijiensis* fungus which causes black leaf streak disease of  
408 banana in the latest invaded region, i.e. Central America and the Caribbean. To that end, the  
409 population history of *P. fijiensis* in this region was retraced based on an analysis of successive  
410 recorded outbreaks and on the population genetic structure. Taken together, the findings of  
411 these analyses revealed that two different epidemic waves of *P. fijiensis* in the northern and  
412 southern Central America-Caribbean Basin region, starting from Honduras, shaped the *P.*  
413 *fijiensis* population structure through serial founder events. This led to the formation of  
414 relatively homogenous genetic groups in the northern and southern Caribbean region, with  
415 lower genetic diversity compared to Central America and with genetic differentiation between  
416 them. The situation was unique in Martinique, Dominica and Guadeloupe. The results  
417 suggested independent introductions of *P. fijiensis* in Martinique from both the northern and  
418 southern origins and an admixture between these introductions accompanying the consecutive  
419 spread of the disease in Dominica and Guadeloupe.

420

421 In contrast with what is frequently observed in fungi (Gladieux et al., 2015), the genotypic  
422 diversity and linkage disequilibrium analysis showed that sexual reproduction was not lost in  
423 *P. fijiensis* during the invasion in the study region, above all following the serial founder  
424 events that occurred in the Caribbean region. Thus, all the populations probably kept the

425 ability to produce wind-dispersed ascospores, which have been shown to disperse over long  
426 distances (Rieux et al., 2014). Ascospores might be essential for *P. fijiensis* to colonize new  
427 areas. Furthermore, the correlation between the occurrence time and the distance from a  
428 geographical point considered as the origin in each wave, and the close values obtained for  
429 the invasion speed between the two waves (about 80 km/year), suggested a similar and  
430 regular step-by-step migration trend. This speed was also close to that estimated in another  
431 study in a continental context (about 90 km/year) and using a modelling approach accounting  
432 for both sexual and asexual reproduction (about 100 km/year but with mate limitation;  
433 (Hamelin et al., 2016)). Lastly, the existence of homogeneous genetic groups within each  
434 wave was also in favour of regular step-by-step migration. Taken together, all the data  
435 obtained in this study suggested that *P. fijiensis* spread throughout the Caribbean mainly via  
436 windborne spore dispersal. Although insular, the Caribbean geography formed by around 30  
437 islands distributed along arcs and separated by 50 to 200 km might have made this spreading  
438 pattern possible. However, we cannot rule out that there may have been some impacts due to  
439 human activities, especially with regard to island-to-island dissemination. Indeed, for human  
440 activities as well as windborne spore dispersal, the shorter the distance between islands, the  
441 more likely exchanges were and both means of dispersal might have had a similar impact on  
442 the pathogen population structure.

443

444 The latest islands to be invaded (Martinique, Dominica and Guadeloupe) appeared to have  
445 been as a special case and the existence of two differentiated genetic groups between the  
446 northern and southern Caribbean led us to detect multiple introductions. What historical  
447 scenario could explain the genetic pattern observed? Through surveillance networks  
448 (Teycheney et al., 2009), we found that Dominica and Guadeloupe—located north of  
449 Martinique—were disease-free when BLSA was first detected in Martinique in 2010 (Table  
450 1). Evidence of the introduction of BLSA in Martinique was the detection of an admixture  
451 between the northern and southern genetic groups in Dominica and Guadeloupe, while these  
452 groups were not found to be admixed in different locations in Martinique. Populations

453 belonging to the southern genetic group might have been introduced into Martinique from  
454 Saint Lucia by windborne spore dispersal. However, an introduction into Martinique from the  
455 northern wave via wind would imply that the disease had first jumped to Guadeloupe and  
456 Dominica and then returned. This movement pattern would be contrary to the prevailing  
457 winds (east to west in the northern Caribbean) and hurricane trajectories (Andraca-Gómez et  
458 al., 2014; Varlas et al., 2018). Although the existence of rare and stochastic long-distance  
459 disease dispersal events cannot be ruled out, another likely introduction scenario for these  
460 islands that were invaded last might be: windborne introductions into Martinique from the  
461 south (via the south to north prevailing winds) or via human activities from the north. The  
462 disease could then have been introduced into Dominica and Guadeloupe from Martinique,  
463 likely by wind dispersal, accompanied by an admixture of the two introduced genetic groups.  
464

465 The *P. fijiensis* fungus may have been at least partly introduced into Martinique by humans  
466 despite the surveillance network operating in the Caribbean (Teycheney et al., 2009). Such  
467 events accompanying plant pathogen colonization on local or regional scales are difficult to  
468 detect and could be more frequent than imagined. Furthermore, the surveillance systems are  
469 not considered to be effective enough to avoid human-mediated introduction of crop diseases  
470 (Carvajal-Yepes et al., 2019). Only an estimated 2 to 6% of all cargo entering a country can  
471 be effectively controlled (Carvajal-Yepes et al., 2019) and there is a lot of uncontrolled boat  
472 traffic in the Caribbean, especially small private boats illegally transporting vegetables  
473 between neighbouring islands. Our results supported this view and highlighted the need to  
474 reinforce surveillance to cope with other emerging crop diseases that have yet to be spread  
475 worldwide. The surveillance system in Australia has been efficient enough to eradicate BLSD  
476 eight times with the application of strict and intensive measures (Guzman et al., 2019)  
477 requiring full collaboration of many actors (from growers to the Ministry of Agriculture). On  
478 a global scale, strong international collaboration between such actors is key for preventing  
479 and excluding emerging plant diseases. Furthermore, reinforcing surveillance could have  
480 more evident impacts on the invasion patterns of pathogens mainly spread through human

481 activities. In the case of banana, this could be crucial for limiting the spread of the very  
482 destructive soilborne *Fusarium* wilt tropical race 4 (*Fusarium oxysporum* fsp cubense TR4)  
483 fungus as a first outbreak in the Americas was suspected very recently in Colombia (Stokstad,  
484 2019), and humans are considered to be the principal vector of dispersal (Dita et al., 2018;  
485 Qazi, 2015). The same reasoning could be applied to the banana bunchy top virus, which is  
486 not present in the Americas and is thought to be spread over long distances through infected  
487 plant material (Jacobsen et al., 2019).

488

489 Even after a serial founder effect during its expansion in Central and South America and the  
490 Caribbean, all of the *P. fijiensis* populations studied had maintained a relatively high level of  
491 genetic diversity and sexual reproduction. It has been suggested that *P. fijiensis* is among the  
492 plant pathogens with the highest evolutionary potential (McDonald and Linde, 2002) and this  
493 feature might remain true even in recently invaded regions. This assumption is supported by  
494 the erosion of quantitative resistance in newly disseminated hybrids on two Caribbean islands  
495 (Cuba and Dominican Republic), although local adaptation of *P. fijiensis* populations was  
496 demonstrated recently in cross-inoculation experiments (Dumartinet et al., 2019).

497 Furthermore, admixtures between different genetic backgrounds, as observed in Dominica  
498 and Guadeloupe, increase the genetic diversity upon which selection can act (Lavergne and  
499 Molofsky, 2007) and produce novel genotypes that may facilitate adaptation to novel habitats  
500 (Rius and Darling, 2014), corresponding here to novel resistant varieties. This hypothesis  
501 could be tested by comparing the pathogenicity of non-admixed or admixed *P. fijiensis*  
502 genotypes on different hosts through cross-inoculation experiments. Furthermore, it can  
503 already be stressed that an effective surveillance system could be an integral part of  
504 sustainable strategies for plant resistance deployment, which can be seen as the most  
505 promising way of controlling emerging plant diseases (Mundt, 2014). Indeed, a second  
506 outcome of such a system might be to restrain gene flows between pathogen populations,  
507 which might affect their evolutionary potential.

508

509 **5. Conclusions**

510 The present study of the population history of the *P. fijiensis* fungus causing the latest  
511 major epidemic on bananas in the America-Caribbean area provided fresh insight into the  
512 epidemiological and demographic processes involved in this emerging plant disease. The  
513 results suggested that the fungus may have spread mainly through spores in this new area,  
514 which encompasses a large insular zone. However, probable human involvement in the spread  
515 of the disease was suspected on some islands. The evolutionary potential of new invasive *P.*  
516 *fijiensis* populations may remain high after serial founder events and could even increase  
517 following admixture, even on some islands. This study also highlighted that surveillance and  
518 quarantine measures need to be strengthened to cope with other emerging diseases of banana  
519 and potential gene flow between established pathogen populations. Possible pathogen  
520 adaptation following the release of new resistant hybrids should also be monitored in order to  
521 adjust resistance deployment strategies and hamper pathogen adaptation.

522

523 **CRedit authorship contribution statement**

524 **Jean Carlier:** Funding acquisition, Project administration, Conceptualization, Data curation,

525 Formal analysis, Investigation, Writing - original draft. **Stéphanie Robert:**

526 Conceptualization, Data curation, Formal analysis, Investigation, Writing - original draft.

527 **Véronique Roussel :** Investigation, Data curation, Methodology, Resources. **Yolande Chilin-**

528 **Charles<sup>1,3</sup>, Nadia Lubin-Adjanoh :** Methodology, Resources. **Aude Gilabert :** Formal

529 analysis, Investigation, Writing - review & editing. **Catherine Abadie:** Funding acquisition,

530 Project administration, Formal analysis, Investigation, Writing - review & editing.

531 **Declaration of competing interest**

532 The authors declare that there is no conflict of interest.

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556

#### 557 **Data availability**

558 All the data are available in the manuscript or in the supplementary data.

#### 559 **Appendix A. Supplementary materials**

560 Supplementary Dataset 1

561 Supplementary File 1

- 563 Agapow, P. M., Burt, A., 2001. Indices of multilocus linkage disequilibrium. *Molecular Ecology Notes*.  
564 1, 101-102.
- 565 Almeida, F., et al., 2019. The Still Underestimated Problem of Fungal Diseases Worldwide. *Front*  
566 *Microbiol.* 10, 214.
- 567 Anderson, P. K., et al., 2004. Emerging infectious diseases of plants: pathogen pollution, climate  
568 change and agrotechnology drivers. *Trends in Ecology & Evolution.* 19, 535-544.
- 569 Andraca-Gómez, G., et al., 2014. A potential invasion route of *Cactoblastis cactorum* within the  
570 Caribbean region matches historical hurricane trajectories. *Biological Invasions.* 17, 1397-  
571 1406.
- 572 Arthofer, W., et al., 2018. Identifying the minimum number of microsatellite loci needed to assess  
573 population genetic structure: A case study in fly culturing. *Fly (Austin).* 12, 13-22.
- 574 Carvajal-Yepes, M., et al., 2019. A global surveillance system for crop diseases. *Science.* 364, 1237-  
575 1239.
- 576 Churchill, A. C. L., 2011. *Mycosphaerella fijiensis*, the black leaf streak pathogen of banana: progress  
577 towards understanding pathogen biology and detection, disease development, and the  
578 challenges of control. *Molecular plant pathology.* 12, 307-328.
- 579 Cornuet, J. M., et al., 2014. DIYABC v2.0: a software to make approximate Bayesian computation  
580 inferences about population history using single nucleotide polymorphism, DNA sequence  
581 and microsatellite data. *Bioinformatics.* 30, 1187-1189.
- 582 de Lapeyre de Bellaire, L., et al., 2010. Black Leaf Streak Disease is challenging the banana industry.  
583 *Fruits.* 65, 327-342.
- 584 Dirienzo, A., et al., 1994. MUTATIONAL PROCESSES OF SIMPLE-SEQUENCE REPEAT LOCI IN HUMAN-  
585 POPULATIONS. *Proceedings of the National Academy of Sciences of the United States of*  
586 *America.* 91, 3166-3170.
- 587 Dita, M., et al., 2018. Fusarium Wilt of Banana: Current Knowledge on Epidemiology and Research  
588 Needs Toward Sustainable Disease Management. *Front Plant Sci.* 9, 1468.
- 589 Dray, S., Dufour, A. B., 2007. The ade4 package: Implementing the duality diagram for ecologists.  
590 *Journal of Statistical Software.* 22, 1-20.
- 591 Dumartinet, T., et al., 2019. Pattern of local adaptation to quantitative host resistance in a major  
592 pathogen of a perennial crop. *Evolutionary Applications.*
- 593 Evans, A., 2016. The Speed of Invasion: Rates of Spread for Thirteen Exotic Forest Insects and  
594 Diseases. *Forests.* 7.
- 595 Fraimout, A., et al., 2017. Deciphering the Routes of invasion of *Drosophila suzukii* by Means of ABC  
596 Random Forest. *Mol Biol Evol.* 34, 980-996.
- 597 Frichot, E., François, O., 2015. LEA: AnRpackage for landscape and ecological association studies.  
598 *Methods in Ecology and Evolution.* 6, 925-929.
- 599 Gladieux, P., et al., 2015. The population biology of fungal invasions. *Mol Ecol.* 24, 1969-86.
- 600 Goudet, J., 2005. HIERFSTAT, a package for R to compute and test hierarchical F-statistics. *Molecular*  
601 *Ecology Notes.* 5, 184-186.
- 602 Guzman, M., et al., Black leaf streak. In: D. R. Jones, (Ed.), *Handbook of diseases of banana, abaca*  
603 *and enset.* CABI, Wallingford, 2019, pp. 41-115.
- 604 Halkett, F., et al., 2010. Genetic discontinuities and disequilibria in recently established populations  
605 of the plant pathogenic fungus *Mycosphaerella fijiensis*. *Mol Ecol.* 19, 3909-23.
- 606 Hamelin, F. M., et al., 2016. Mate Finding, Sexual Spore Production, and the Spread of Fungal Plant  
607 Parasites. *Bull Math Biol.* 78, 695-712.
- 608 Hill, M. O., 1973. Diversity and evenness: a unifying notation and its consequences. *Ecology.* 54, 427-  
609 432.
- 610 loos, R., et al., 2011. First Report of Black Sigatoka Disease in Banana Caused by *Mycosphaerella*  
611 *fijiensis* on Martinique Island. *Plant Disease.* 95, 359-359.
- 612 Jacobsen, K., et al., 2019. Seed degeneration of banana planting materials: strategies for improved  
613 farmer access to healthy seed. *Plant Pathology.* 68, 207-228.
- 614 Jones, D. R., 2019. *Handbook of diseases of banana, abaca and enset.* CABI, Wallingford.
- 615 Kamvar, Z. N., et al., 2014. Poppr: an R package for genetic analysis of populations with clonal,  
616 partially clonal, and/or sexual reproduction. *PeerJ.* 2, e281.

617 Lavergne, S., Molofsky, J., 2007. Increased genetic variation and evolutionary potential drive the  
618 success of an invasive grass. *Proc Natl Acad Sci U S A.* 104, 3883-8.

619 McDonald, B. A., Linde, C., 2002. Pathogen population genetics, evolution potential, and durable  
620 resistance. *Ann. Rev. Phytopathol.* 40, 349-379.

621 McDonald, B. A., Stukenbrock, E. H., 2016. Rapid emergence of pathogens in agro-ecosystems: global  
622 threats to agricultural sustainability and food security. *Philos Trans R Soc Lond B Biol Sci.* 371.

623 Mundt, C. C., 2014. Durable resistance: a key to sustainable management of pathogens and pests.  
624 *Infect Genet Evol.* 27, 446-55.

625 Nei, M., 1978. Estimation of average heterozygosity and genetic distances from a small number of  
626 individuals. *Genetics.* 89, 583-590.

627 Nei, M., 1987. *Molecular Evolutionary Genetics.* Columbia University Press, NY.

628 Parnell, S., et al., 2017. Surveillance to Inform Control of Emerging Plant Diseases: An Epidemiological  
629 Perspective. *Annual Review of Phytopathology, Vol 55.* 55, 591-610.

630 Peery, M. Z., et al., 2012. Reliability of genetic bottleneck tests for detecting recent population  
631 declines. *Molecular Ecology.* 21, 3403-3418.

632 Pennisi, E., 2010. Armed and Dangerous. *Science.* 327, 804-805.

633 Piry, S., et al., 1999. Bottleneck: A computer program for detecting recent reductions in the effective  
634 population size using allele frequency data. *The Journal of Heredity.* 90, 502-503.

635 Pritchard, J. K., et al., 2000. Inference of population structure using multilocus genotype data.  
636 *Genetics.* 155, 945-959.

637 Pudlo, P., et al., 2016. Reliable ABC model choice via random forests. *Bioinformatics.* 32, 859-66.

638 Qazi, J., 2015. Banana bunchy top virus and the bunchy top disease. *Journal of General Plant  
639 Pathology.* 82, 2-11.

640 Rieux, A., et al., 2014. Long-distance wind-dispersal of spores in a fungal plant pathogen: estimation  
641 of anisotropic dispersal kernels from an extensive field experiment. *PLoS One.* 9, e103225.

642 Rius, M., Darling, J. A., 2014. How important is intraspecific genetic admixture to the success of  
643 colonising populations? *Trends Ecol Evol.* 29, 233-42.

644 Robert, S., et al., 2012. Contrasting introduction scenarios among continents in the worldwide  
645 invasion of the banana fungal pathogen *Mycosphaerella fijiensis*. *Mol Ecol.* 21, 1098-114.

646 Robert, S., et al., 2010. Optimized genotyping with microsatellite markers in the fungal banana  
647 pathogen *Mycosphaerella fijiensis* (*Mycosphaerellaceae*). *Am J Bot.* 97, e130-2.

648 Robert, S., et al., 2015. MULTIPLE INTRODUCTIONS AND ADMIXTURE AT THE ORIGIN OF THE  
649 CONTINENTAL SPREAD OF THE FUNGAL BANANA PATHOGEN MYCOSPHAERELLA FIJIENSIS IN  
650 CENTRAL AMERICA: A STATISTICAL TEST USING APPROXIMATE BAYESIAN COMPUTATION.  
651 *Revue D Ecologie-La Terre Et La Vie.* 70, 127-138.

652 Rogers, A. R., 2014. How population growth affects linkage disequilibrium. *Genetics.* 197, 1329-41.

653 Rousset, F., 2008. genepop'007: a complete re-implementation of the genepop software for  
654 Windows and Linux. *Mol Ecol Resour.* 8, 103-6.

655 Stokstad, E., 2019. Banana fungus puts Latin America on alert. *Science.* 365, 207-208.

656 Stukenbrock, E. H., McDonald, B. A., 2008. The origins of plant pathogens in agro-ecosystems. *Annual  
657 Review of Phytopathology.* 46, 75-100.

658 Teycheney, P.-Y., et al., Development of plant epidemiological surveillance networks, data exchanges  
659 and joint response strategies in the Caribbean: the french experience. In: G. B. R. Cfc, (Ed.),  
660 Annual Meeting of the Caribbean Food Crops Society. s.n., Basseterre, Saint-Kitts-et-Nevis,  
661 2009, pp. 8 p.

662 Varlas, G., et al., 2018. An analysis of the synoptic and dynamical characteristics of hurricane Sandy  
663 (2012). *Meteorology and Atmospheric Physics.* 131, 443-453.

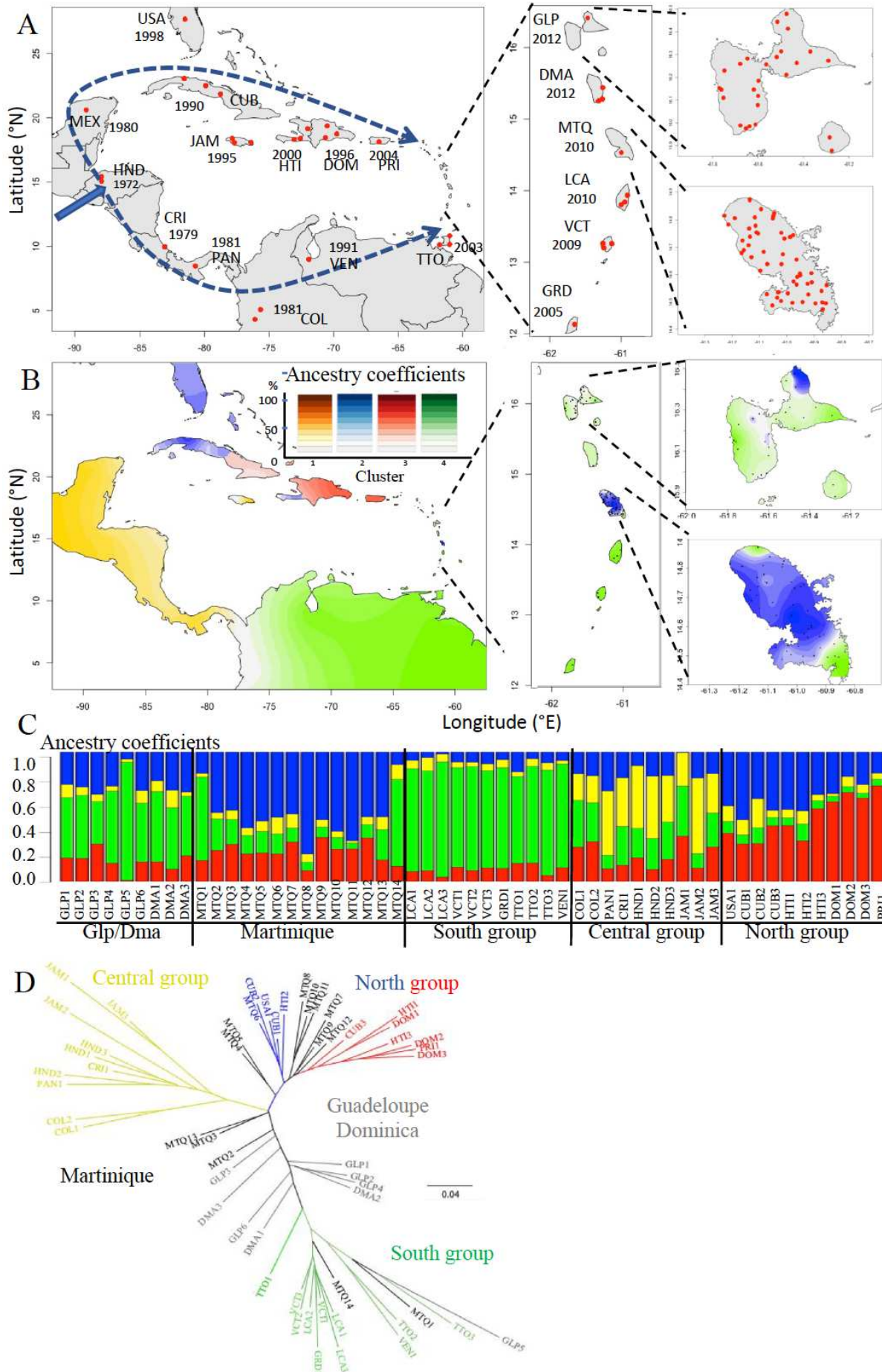
664 Weir, B. S., Cockerham, C. C., 1984. Estimating F-statistics for the analysis of population structure.  
665 *Evolution.* 38, 1358-1370.

666 Zapater, M. F., et al., 2008. Microsatellite markers for the fungal banana pathogens *Mycosphaerella  
667 fijiensis*, *Mycosphaerella musicola* and *Mycosphaerella eumusae*. *Molecular Ecology  
668 Resources.* 8, 1121-1125.

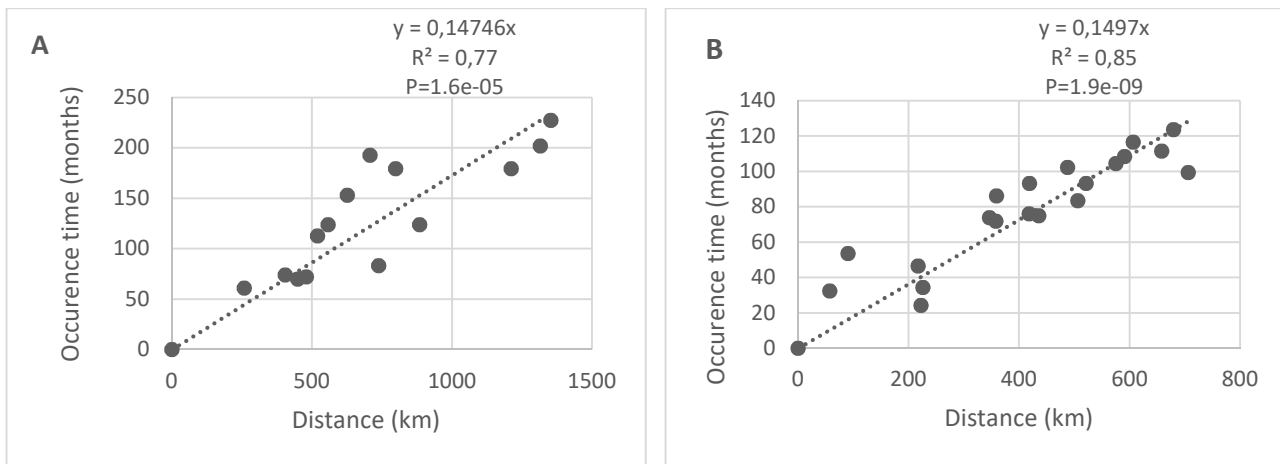
669

670

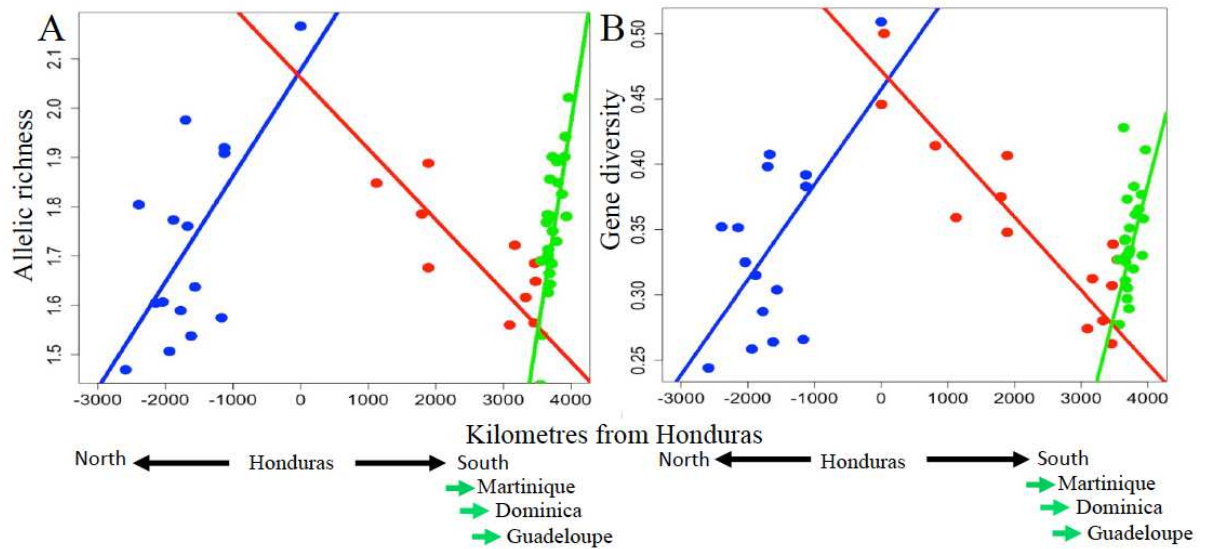




673 **Fig. 1.** Invasion and population genetic structure of *Pseudocercospora fijiensis* in the  
674 Americas. A, geographical locations of the samples analysed and year of first official  
675 detection of BLSD in the locations; B, interpolation of sNMF results with K=4 of the most  
676 represented cluster in all of the 2,188 isolates analysed; C, barplots with K=4 of the mean  
677 ancestry coefficients for the 55 samples with more than 18 isolates; D, unrooted neighbour-  
678 joining tree based on Cavalli-Sforza distances between the same 55 samples. Abbreviations:  
679 GLP = Guadeloupe, DMA = Dominica, MTQ = Martinique, LCA = Saint Lucia, VCT = Saint  
680 Vincent, GRD = Grenada, TTO = Trinidad, VEN = Venezuela, COL = Colombia, PAN =  
681 Panama, CRI= Costa Rica, HND= Honduras, MEX= Mexico, USA= United States of  
682 America, CUB=Cuba, JAM=Jamaica, HTI=Haiti, DOM= Dominican Republic, PRI= Puerto  
683 Rico.  
684



686 **Fig. 2.** Linear regression between occurrence times in Caribbean locations and Euclidian  
687 distances between these locations and a geographical point considered as the origin of the  
688 northern (A) or southern (B) migration front of *P. fijiensis* in the Caribbean.  
689



691  
 692 **Fig. 3.** Linear regression between cumulative step-by-step distance from Honduras with  
 693 allelic richness (A) or (B) Nei's gene diversity calculated for the 55 *P. fijiensis* samples with  
 694 more than 18 isolates. Correlation tests were: (A), northern direction ( $r=-0.67$ ,  $p=6 \cdot 10^{-3}$ ),  
 695 southern direction ( $r=-0.86$ ,  $p=2.9 \cdot 10^{-6}$ ), Martinique to Guadeloupe ( $r=-0.65$ ,  $p=4.6 \cdot 10^{-4}$ ); (B),  
 696 northern direction ( $r=-0.63$ ,  $p=0.011$ ), southern direction ( $r=-0.89$ ,  $p=6.4 \cdot 10^{-7}$ ), Martinique to  
 697 Guadeloupe ( $r=-0.44$ ,  $p=0.02$ ).

698

699

700

701

Country/Place	First official detection (month/year)	Sampling (Number)		Reference
		Locations	Isolates	
France/Guadeloupe	01/2012	27	397	This study
Dominica	06/2012	3	91	This study
France/Martinique	09/2010	58	680	This study
Saint Lucia	01/2010	3	78	This study
Saint Vincent	10/2009	3	94	This study
Grenada	11/2005	1	25	This study
Trinidad	11/2003	3	68	This study
Venezuela	1991*	1	26	Robert et al., 2012
Colombia	1981*	2	46	Robert et al., 2012
Panama	1981*	1	31	Robert et al., 2012
Costa Rica	1979*	1	30	Robert et al., 2012
Honduras	1972*	3	80	Robert et al., 2012
Mexico	1980*	1	11	Robert et al., 2012
USA/Florida	1998*	1	60	This study
Cuba	11/1989	3	138	This study
Jamaica	08/1995	3	95	This study
Haiti	01/2000	3	85	This study
Dominican Rep.	09/1996	3	127	This study
USA/Puerto Rico	08/2004	1	26	This study
Total		121	2188	

703 \*According to Jones (Jones, 2019)

704 **Table 2** Genetic statistics calculated in *Pseudocercospora fijiensis* populations.

Country	Code	Number of isolates	Number of MLG <sup>1</sup>	P <sup>2</sup>	Ar <sup>3</sup>	λ <sup>4</sup>	He <sup>5</sup>	Test of He excess <sup>6</sup>	$\bar{r}_d$ <sup>7</sup>
France/ Guadeloupe	GLP1	32	31	93.8	2.02	0.97	0.41	**	0.00
	GLP2	20	17	87.5	1.94	0.94	0.33	NS <sup>8</sup>	0.02
	GLP3	25	24	87.5	1.90	0.96	0.38	**	0.02

	GLP4	38	37	87.5	1.78	0.97	0.36	NS	0.01
	GLP5	18	16	31.3	1.28	0.93	0.21	NS	0.08**
	GLP6	32	29	81.3	1.83	0.96	0.37	**	0.00
Dominica	DMA1	37	37	81.3	1.85	0.97	0.36	**	0.01
	DMA2	26	25	87.5	1.89	0.96	0.38	*	0.01
	DMA3	28	26	81.3	1.73	0.96	0.32	NS	0.03*
France/ Martinique	MTQ1	23	17	43.8	1.36	0.93	0.20	NS	0.10**
	MTQ2	21	20	81.3	1.75	0.95	0.33	NS	0.05**
	MTQ3	29	28	93.8	1.90	0.96	0.35	NS	0.00
	MTQ4	36	29	75.0	1.68	0.96	0.29	NS	0.04**
	MTQ5	27	26	75.0	1.78	0.96	0.33	*	0.02
	MTQ6	18	18	87.5	1.86	0.94	0.37	NS	-0.01
	MTQ7	19	19	68.8	1.69	0.95	0.31	*	-0.02
	MTQ8	32	30	68.8	1.64	0.96	0.30	*	0.01
	MTQ9	18	18	68.8	1.66	0.94	0.33	NS	0.00
	MTQ10	47	41	87.5	1.71	0.97	0.33	NS	0.01
	MTQ11	20	19	81.3	1.70	0.95	0.31	NS	0.04*
	MTQ12	25	23	68.8	1.63	0.95	0.34	NS	0.05**
	MTQ13	19	17	81.3	1.78	0.93	0.34	NS	0.04*
	MTQ14	18	17	81.3	1.77	0.94	0.43	NS	0.06**
Saint Lucia	LCA1	23	22	62.5	1.54	0.95	0.28	NS	0.04*
	LCA2	31	31	68.8	1.69	0.97	0.33	*	0.02
	LCA3	24	20	56.3	1.44	0.94	0.22	NS	0.07**
Saint Vincent	VCT1	46	44	68.8	1.65	0.98	0.34	NS	0.02*
	VCT2	24	24	68.8	1.56	0.96	0.26	NS	0.01
	VCT3	24	24	68.8	1.68	0.96	0.31	**	-0.01
Grenada	GRD1	25	25	62.5	1.62	0.96	0.28	NS	0.01
Trinidad	TTO1	20	20	75.0	1.72	0.95	0.31	NS	0.00
	TTO2	26	23	56.3	1.56	0.95	0.27	NS	0.01
	TTO3	22	17	37.5	1.32	0.92	0.23	NS	0.12**

Venezuela	VEN1	26	26	62.5	1.68	0.96	0.35	NS	0.05**
Colombia	COL1	23	23	75.0	1.79	0.96	0.38	NS	0.01
	COL2	23	23	81.3	1.89	0.96	0.41	NS	0.00
Panama	PAN1	31	31	93.8	1.85	0.97	0.36	NS	0.00
Costa Rica	CRI1	30	30	100.0	1.91	0.97	0.41	NS	0.03*
Honduras	HND1	30	30	100.0	2.11	0.97	0.50	**	0.01
	HND2	23	22	87.5	2.01	0.95	0.45	**	0.01
	HND3	27	27	100.0	2.17	0.96	0.51	**	0.01
USA	USA1	60	60	87.5	1.80	0.98	0.35	**	0.00
Cuba	CUB1	46	43	81.3	1.77	0.98	0.32	*	0.00
	CUB2	40	40	87.5	1.98	0.98	0.40	**	0.01
	CUB3	52	52	62.5	1.64	0.98	0.30	*	0.01*
Jamaica	JAM1	32	27	87.5	1.57	0.96	0.27	NS	0.06**
	JAM2	30	30	93.8	1.91	0.97	0.38	NS	0.03*
	JAM3	33	33	87.5	1.92	0.97	0.39	NS	0.03**
Haiti	HTI1	25	25	75.0	1.54	0.96	0.26	NS	-0.01
	HTI2	28	28	75.0	1.76	0.96	0.41	*	0.01
	HTI3	32	27	68.8	1.59	0.96	0.29	*	0.01
Dominican	DOM1	45	41	68.8	1.51	0.97	0.26	NS	0.04**
Republic	DOM2	46	46	68.8	1.61	0.98	0.33	NS	0.04**
	DOM3	36	34	68.8	1.60	0.97	0.35	NS	0.02*
USA/Puerto Rico	PRI1	26	22	50.0	1.47	0.95	0.24	NS	0.07**

705 <sup>1</sup> Number of multi-locus genotypes (MLG), <sup>2</sup> Percentage of polymorphic loci, <sup>3</sup> Allelic richness, <sup>4</sup> Simpson's  
706 index (Hill, 1973), <sup>5</sup> Nei's gene diversity (Nei, 1978), <sup>6</sup> Wilcoxon test implemented in Bottleneck software (Piry  
707 et al., 1999), <sup>7</sup> Standardized index of association (Agapow and Burt, 2001), <sup>8</sup> Not significant, \* Significant at a  
708 false discovery rate (FDR) of 5%, \*\* Significant at a false discovery rate (FDR) of 1%.

709

710 **Table 3** Genetic differentiation between *Pseudocercospora fijiensis* populations.

Comparisons	<i>F</i> -statistic <sup>1</sup>	CI-95% <sup>2</sup>	Test
Global analysis: AMOVA <sup>3</sup>			
Between populations	0.36	0.30/0.43	Perm <sup>4</sup> ***

Between populations within groups <sup>5</sup>	0.16	0.14/0.19	Perm***
Between populations within countries	0.12	0.06/0.15	Perm***
Between countries	0.27	0.22/0.33	Perm***
Between countries within groups	0.05	0.01/0.10	Perm***
Between groups	0.23	0.18/0.32	Perm***
Guadeloupe/Dominica: pairwise estimate <sup>6</sup>			
Min	0.00	-0.025/0.01	Fet <sup>7</sup> NS <sup>8</sup>
Max	0.14	0.06/0.21	Fet***
Martinique: pairwise estimate			
Min	0.00	-0.03/0.00	Fet NS
Max	0.51	0.31/0.68	Fet***
MTQ1 vs others	0.31-0.51		Fet***
MTQ14 vs others	0.21-0.39		Fet***
MTQ1 vs MTQ14	0.21		Fet***
Combinations excluding			
MTQ1 and MTQ14	0.00-0.11		Fet NS to Fet***

711 <sup>1</sup> Hierarchical *F*-statistics from the AMOVA or pairwise estimate of Weir and Cockerham (1984), <sup>2</sup>  
712 95% confidence interval, <sup>3</sup> AMOVA carried out without samples from Guadeloupe, Martinique,  
713 Dominica, <sup>4</sup> Test using permutations, <sup>5</sup> Groups corresponding to those defined in figure 2C: central  
714 group (Colombia, Panama, Costa Rica, Honduras, Jamaica), northern group (Florida, Cuba, Haiti,  
715 Dominican Republic, Porto Rico), southern group (Saint Lucia, Saint Vincent, Grenada, Trinidad,  
716 Venezuela), <sup>6</sup> range of *F*-statistics estimated between population pairs. Estimates for all population  
717 pairs are given in Data S1. <sup>7</sup> Fisher exact test between population pairs, <sup>8</sup> Not significant, \*\*\*  $p <$   
718 0.0001.