



GSK3 β , a Master Kinase in the Regulation of Adult Stem Cell Behavior

Claire Racaud-Sultan, Nathalie Vergnolle

► To cite this version:

Claire Racaud-Sultan, Nathalie Vergnolle. GSK3 β , a Master Kinase in the Regulation of Adult Stem Cell Behavior. Cells, 2021, 10 (2), 10.3390/cells10020225 . hal-03181357

HAL Id: hal-03181357

<https://hal.inrae.fr/hal-03181357>

Submitted on 25 Mar 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License

Review

GSK3 β , a Master Kinase in the Regulation of Adult Stem Cell Behavior

Claire Racaud-Sultan * and Nathalie Vergnolle

IRSD, INSERM, INRAE, ENVT, Université de Toulouse, UPS, CHU Purpan, Place du Dr Baylac, CEDEX 3, 31024 Toulouse, France; nathalie.vergnolle@inserm.fr

* Correspondence: claire.racaud@inserm.fr

Abstract: In adult stem cells, Glycogen Synthase Kinase 3 β (GSK3 β) is at the crossroad of signaling pathways controlling survival, proliferation, adhesion and differentiation. The microenvironment plays a key role in the regulation of these cell functions and we have demonstrated that the GSK3 β activity is strongly dependent on the engagement of integrins and protease-activated receptors (PARs). Downstream of the integrin $\alpha_5\beta_1$ or PAR₂ activation, a molecular complex is organized around the scaffolding proteins RACK1 and β -arrestin-2 respectively, containing the phosphatase PP2A responsible for GSK3 β activation. As a consequence, a quiescent stem cell phenotype is established with high capacities to face apoptotic and metabolic stresses. A protective role of GSK3 β has been found for hematopoietic and intestinal stem cells. Latters survived to de-adhesion through PAR₂ activation, whereas formers were protected from cytotoxicity through $\alpha_5\beta_1$ engagement. However, a prolonged activation of GSK3 β promoted a defect in epithelial regeneration and a resistance to chemotherapy of leukemic cells, paving the way to chronic inflammatory diseases and to cancer resurgence, respectively. In both cases, a sexual dimorphism was measured in GSK3 β -dependent cellular functions. GSK3 β activity is a key marker for inflammatory and cancer diseases allowing adjusted therapy to sex, age and metabolic status of patients.



Citation: Racaud-Sultan, C.; Vergnolle, N. GSK3 β , a Master Kinase in the Regulation of Adult Stem Cell Behavior. *Cells* **2021**, *10*, 225. <https://doi.org/10.3390/cells10020225>

Academic Editor:
Hagit Eldar-Finkelman
Received: 30 November 2020
Accepted: 22 January 2021
Published: 24 January 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

The Glycogen Synthase Kinase 3 β (GSK3 β) is an ancestral protein kinase, ubiquitously expressed, which controls many cellular functions [1]. Inhibited in basal conditions, GSK3 β is activated under stress and can drive cells to death or to survival depending on its sub-cellular localization and specific partners. GSK3 β has been found activated in pathologies such as inflammation and cancer where the deregulation of adult stem cells plays a critical role [2]. Here, we will summarize some key GSK3 β -dependent regulations of adult stem cells and their consequences on the behavior of hematopoietic and intestinal stem cells, as examples. We will highlight the importance of adhesion and protease-activated receptors engagement in the activation of signaling pathways conducting to GSK3 β activation. Finally, we will discuss the positive and negative aspects of GSK3 β activation in pathological conditions and the benefit to consider GSK3 β as diagnostic marker and therapeutic target for a precision medicine.

2. GSK3 β as a Sensor of the Adult Stem Cell Niche

Adult stem cells are in charge of tissue regeneration along the life of organisms. This is the reason why they must be protected from stressors leading to cell death or exhaustion. The microenvironment of adult stem cells, namely “niche”, supports this protective role offering in particular an adhesive anchorage and proper metabolic exchanges [3]. However, aging and exogenous aggressions decrease the niche capacities to control stem cell behavior and thus favor pathogenesis.

Facing stress, adult stem cells could undergo apoptosis, or exhaustion through accelerated differentiation, or senescence, or tumor transformation. As a first step, stem cells must survive and cope with nutrient or oxidative stress. Through its strategic locations in the cell (plasma membrane, endoplasmic reticulum, mitochondria, nucleus), and in response to various signals, GSK3 β is able to control death-signaling pathways at the membranes, energetic metabolism and gene transcription [1]. Thus, a resistant quiescent stem cell phenotype is established. It is linked to a strong anchorage to extracellular matrix and supporting cells in the niche, as well as a low energetic metabolism [4,5].

GSK3 β acts as a key sensor of cell metabolism and its activation allows energetic supply and anti-oxidant defenses in a low-glucose microenvironment [6]. Indeed, in basal conditions, Insulin receptor-signaling complex inhibits GSK3 β to release glycogenesis and Nrf2-dependent anti-oxidant systems. In addition to its role in survival of adult stem cells, active GSK3 β prevents ROS-induced differentiation or senescence [7].

As recapitulated in Figure 1, GSK3 β plays a central role in the adaptation of adult stem cells to their microenvironment allowing their long-term maintenance during the whole life of the organism.

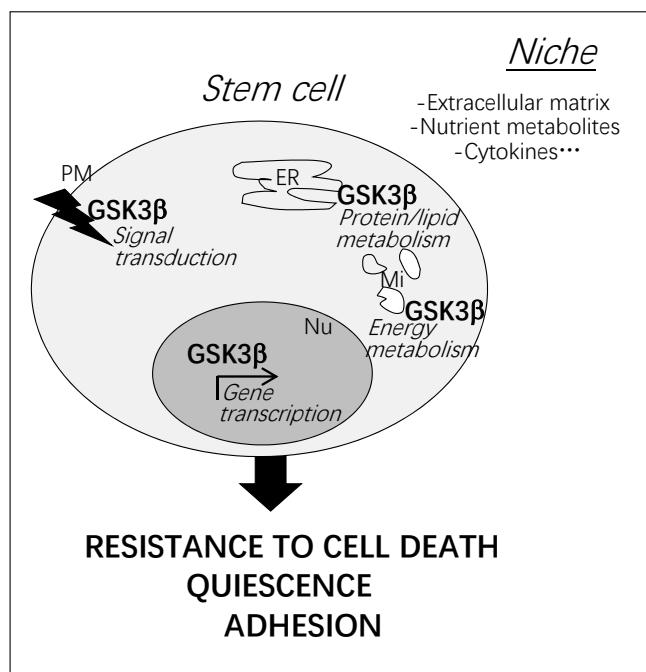


Figure 1. Multiple cellular localization of GSK3 β activation and consequences on stem cell status. At the plasma membrane, GSK3 β is critical to counteract death-signaling pathways induced by cytokines and adhesion changes in injured microenvironment. In case of nutrient or oxidative stresses, GSK3 β can adjust metabolic activities in the endoplasmic reticulum and in mitochondria. Cell survival and metabolism are tightly linked and their cross-regulation is particularly important in subcellular compartments where GSK3 β is found. In the nucleus, GSK3 β –modulated gene transcription contributes to shape the resistant phenotype of stem cells. Localized signaling activities responsible for GSK3 β activation are complex as well as the relationship between them [1]. PM: Plasma membrane; ER: Endoplasmic reticulum; Mi: Mitochondria; Nu: Nucleus.

3. GSK3 β and Key Functions of Adult Stem Cells

The implication of GSK3 β activation in adult stem cell regulation has first been identified for self-renewal function. Indeed, as a serine-threonine kinase, GSK3 β is able to phosphorylate β -catenin, targeting it to the proteasome for degradation and blocking its translocation to the nucleus, normally required for stem cell renewal [8]. In the nucleus, β -catenin is a partner of the transcription factor TCF which promotes self-renewal, a

proliferative mode maintaining tissue stem cells. Thus the activation of the GSK3 β pool associated with β -catenin in the cytoplasm can push stem cells towards a quiescent state.

Another important aspect of GSK3 β -dependent regulation of stem cells is the control of cell differentiation. Active GSK3 β can influence the balance between energetic/proliferative and differentiation pathways through the activation of mTOR negative regulators [9] and the degradation of transcription factors promoting lineage commitment [10]. As a result, undifferentiated state of tissue progenitors is favored. Here, it is important to note that GSK3 β inhibition has a different impact in adult stem cells and their progeny [11–13], suggesting that the regulation of GSK3 β is tightly dependent on microenvironment specificities.

Migration of adult stem cells is crucial to tissue repair and tumor transformation. Active GSK3 β participates to the turnover of focal adhesions built by adhesion receptors (integrins, cadherins), promoting cell migration [14]. This could contribute to cell plasticity allowing transitions between different lineages and epithelial-mesenchymal transition [15].

Advantages conferred by a GSK3 β -dependent plasticity could also be critical for stem cell survival. At both plasma membrane and nucleus, active GSK3 β promotes a switch in death receptor and nuclear factor- κ B (NF- κ B) activities, respectively. Under the influence of active GSK3 β , a signaling complex (DDX3/cIAP-1) is established downstream of death receptors, promoting cell survival instead of cell death [16]. In addition, GSK3 β phosphorylates the p65 subunit, inducing an I κ B-independent activation of NF- κ B [17]. The GSK3 β -dependent activation of NF- κ B triggers the transcription of a specific set of genes [18]. Furthermore, NF- κ B target genes are under a GSK3 β -dependent epigenetic control [19].

All these GSK3 β -dependent functions could be critical in stem cells and depend on different pools of GSK3 β (Figure 2). It should be noted that certain types of stem cells require a very tight control of GSK3 β activity, such as neural stem cells pushed to cell death by autophagy in case of over-activation of GSK3 β [20].

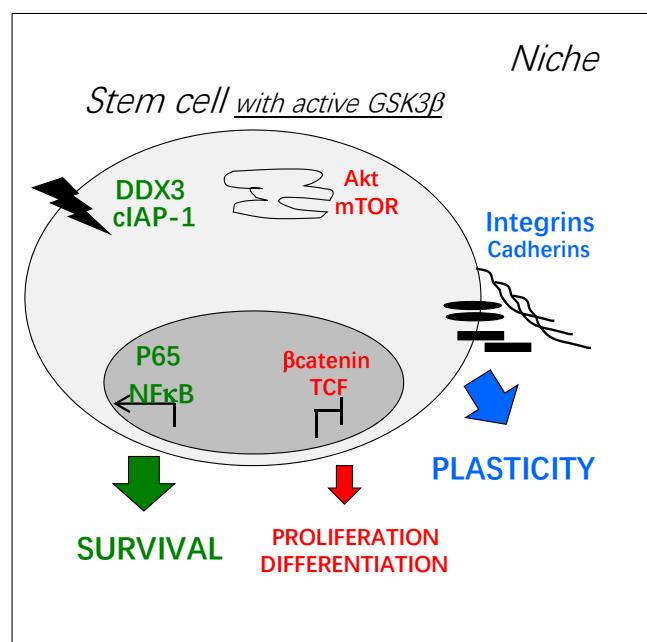


Figure 2. Key cellular functions potentially regulated by active GSK3 β in adult stem cells: Survival (green) and plasticity (blue) are promoted and metabolism/proliferation/differentiation (red) are decreased. Depending on the environment, signal transduction at the plasma membrane can trigger GSK3 β activation, by the inhibition of its kinase-dependent phosphorylation or the activation of its phosphatase-dependent de-phosphorylation, for example [1]. Inside the cell, active GSK3 β can be found in different molecular complexes and its trafficking between them is poorly known. These pools of GSK3 β can be independently activated or inhibited and little is known on their coordinated activation.

4. Integrin-Dependent Activation of GSK3 β in Leukemia

A number of works have demonstrated the major role of GSK3 β in hematopoietic stem cells homeostasis [21]. By regulating both β -catenin and mTOR signaling, GSK3 β controls hematopoietic stem cell renewal and differentiation, respectively. As a result, inhibition of GSK3 β promotes the expansion of hematopoietic stem cells and their better engraftment [22], whereas activation of GSK3 β is associated with differentiation blockade and egress from the hematopoietic niche [13].

GSK3 β has been found activated in many cancers, among them leukemia [23]. In leukemia stem cells, GSK3 β controls survival, proliferation and differentiation [21]. However, GSK3 β inhibition impacts leukemic stem cells survival in an opposite way to their normal counterparts [22]. We have found GSK3 β activation in a subset of leukemic cells independent from Akt kinase for their survival, after adhesion onto fibronectin [24–26]. Importantly, adhesion of leukemic progenitors to the hematopoietic niche (osteoblasts), has triggered both GSK3 β activation and resistance to cytotoxic drugs [24,26]. GSK3 β was found associated in a complex with the α_5 integrin, the scaffolding protein RACK1, and the phosphatase PP2A responsible for its activation through serine 9-dephosphorylation [25].

Adhesion-dependent activation of GSK3 β controls the survival of leukemic progenitors through multiple pathways such as the activation of NF- κ B independently from I κ B variations [24], the resistance to tumor necrosis factor- α [25] and the modulation of Wnt pathway through the up-regulation of secreted Frizzled-related protein-1 [24]. As a result, a very resistant and quiescent phenotype of cancer stem cell is established.

A major observation is that leukemic stem cells (CD34 $^+$ CD38 $^-$ CD123 $^+$, acute myeloid leukemia) surviving through the GSK3 β pathway are from female patients [26]. Interestingly, GSK3 β -dependent survival has also been measured in leukemic stem cells from male diabetic patients [unpublished results]. Leukemic stem cells from female patients have been characterized by an up-regulation of the expression of RACK1 [26]. Strikingly, normal hematopoietic stem cells from male donors, but not those from females, have been found dependent on the GSK3 β pathway to survive [26]. This is a demonstration of the strong capacities of cancer stem cells to hijack plasticity to develop tumors in a physiological niche [27]. However, other acute myeloid leukemia subsets do not depend on GSK3 β to survive and are capable to transform their microenvironment in a leukemic niche with benefits for their development [28].

The Figure 3 recapitulates data on integrin-dependent GSK3 β activation in leukemic stem cells.

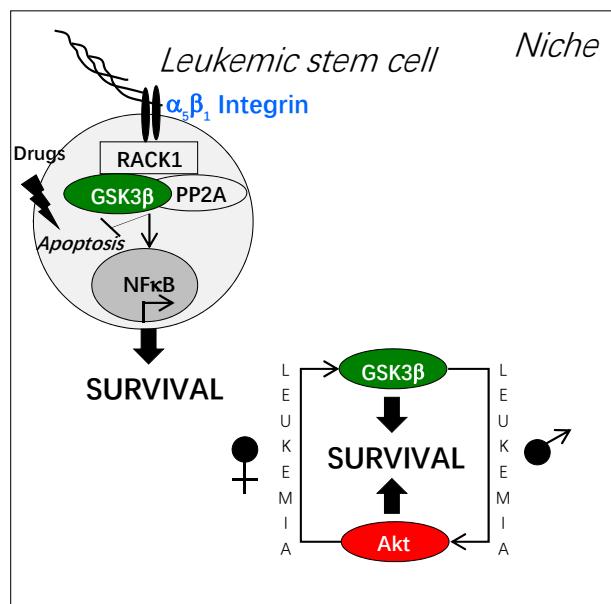


Figure 3. Integrin-dependent activation of GSK3 β and leukemic stem cell resistance. Adhesion of

leukemic stem cells to fibronectin or to osteoblasts triggers the formation of a molecular complex around the adaptor protein RACK1 associated to the cytoplasmic domain of integrin. In this molecular complex, GSK3 β is activated through its de-phosphorylation by the phosphatase PP2A. Thus, leukemic stem cell resistance to cytotoxic stress is promoted by different mechanisms, among them NF- κ B activation. Of note, the dependence of hematopoietic stem cells to GSK3 β displays a sexual dimorphism that is switched upon leukemogenesis.

5. GSK3 β in the Colon Crypt: Role and Controls by Proteases and Their Receptors

Is the protective role of GSK3 β in hematopoietic cells applicable to other adult stem cells? Pathologies such as inflammatory bowel diseases (IBD) and colorectal cancer (CRC) are characterized by a high activity of GSK3 β in epithelial cells and their microenvironment [29–31], but the etiology of this over-activity is poorly understood.

In IBD and CRC, epithelial stem cells, located in a cryptic niche, play a critical role since tissue regeneration is impaired either by down-regulation or by deregulation, respectively. Concomitant with their role in epithelial barrier, adhesive molecules are implicated in regeneration, and the remodeling of the adhesive support is critical for the behavior of stem cells [32]. The dual role of the GSK3 β -regulated β -catenin both in support of the cadherin-adhesive function and in TCF-transcription activity is critical for the colon stem cell homeostasis [33]. In colorectal cancer cells, active GSK3 β associated with mutated APC is not efficient to promote β -catenin degradation, resulting in Wnt signaling deregulation and increased proliferation. In addition, active GSK3 β promotes survival and drug resistance [34–36] and the degradation of Hafth1, transcription factor essential to the differentiation of the secretory lineage [10].

Proteases are expressed in large amounts during inflammation and cancer [37–41]. They can either modify the niche of stem cells through the release of growth factors or the proteolysis of extracellular matrix, or signal through specific receptors, namely protease-activated receptors (PARs) [41]. Our study by immunofluorescence has demonstrated the expression of PAR₂ and PAR₁, two members of the PAR family implicated in IBD and CRC, along the human or murine colon crypt and in colon stem cells (Lgr5 $^+$, Sox9 $^+$) [42]. Other groups have measured gene expression of PAR₂ in intestinal and colon stem cells [43,44]. In murine organoid assays, PAR₂ activation has been found critical to protect colon progenitors from anoikis [42]. PAR₂ activation has induced a decrease in proliferation of epithelial progenitors, by contrast with PAR₁ [42]. Downstream of PAR₂ activation, a signaling pathway implicating β arrestin-2, PP2A and GSK3 β has been triggered and regulated by the cytoskeleton organizer Rho, pushing colon stem cells towards a quiescent and resistant phenotype. This quiescence could result from GSK3 β -dependent β -catenin degradation and/or the increased expression of the DUSP6 phosphatase, a negative regulator of ERK signaling [42]. Interestingly, it has been shown that active GSK3 β could positively regulate the phosphatase PP1 [45] which stabilizes the co-transcription factor YAP downstream of PAR₂ to promote epithelial survival and regeneration in response to injury [46].

Importantly, as for hematopoietic stem cells, a sexual dimorphism in PAR₂-dependent GSK3 β activation in colon stem cells has been observed [47]. The PAR₂/GSK3 β pathway has been triggered in colon progenitors from male mice, whereas females have displayed a PAR₂/AKT pathway. Moreover, PAR₂ has been shown to control specific gene expression in males and females, i.e., Itga6 and Timp2, respectively.

Thus, active GSK3 β has a protective role for adult stem cells from both mesenchymal and epithelial origin, acting as a sensor of their niche, and under plastic regulation in both sexes.

The Figure 4 summarizes data on PAR₂-dependent regulation of GSK3 β in colon stem cells.

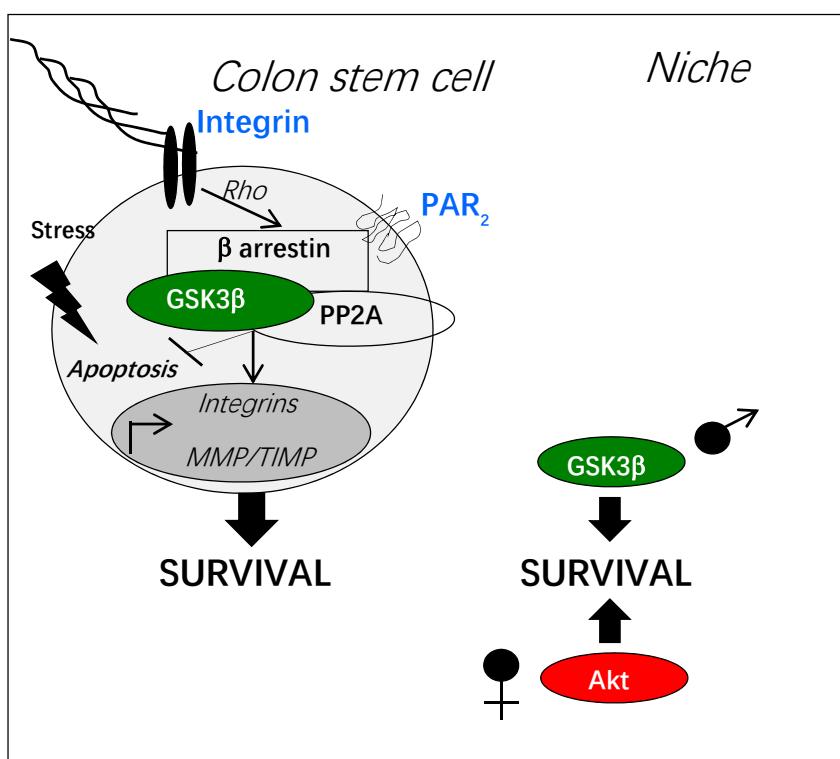


Figure 4. PAR₂-dependent activation of GSK3 β in normal colon stem cells. PAR₂ activation by protease-dependent cleavage can induce a β arrestin-dependent signaling pathway resulting in GSK3 β activation through its de-phosphorylation by the phosphatase PP2A. This signaling pathway is controlled by the Rho kinase activated by adhesive stress. Gene transcription of integrins, metalloproteases (MMP) and their inhibitors (TIMP), is regulated by the PAR₂/GSK3 β pathway. Active GSK3 β counteracts stress-induced apoptosis by multiple mechanisms (see Section 3, “GSK3 β and key functions of adult stem cells”). In the physiology of colon stem cells, the pro-survival signaling of PAR₂ is sexually dimorphic depending on GSK3 β or on Akt.

6. Consequences of Prolonged GSK3 β Activation in Inflammation and Cancer

GSK3 β activation has a protective role for adult stem cells through its capacity to switch them towards a quiescent and resistant phenotype. However, in a deregulated microenvironment, prolonged or iterative activation of GSK3 β can be deleterious. Factors responsible for deregulated microenvironment include chronic inflammation with high levels of cytokines and proteases, persistent matrix remodeling and metabolic changes. It is thus clear that the restoration of a stem cell niche with physiological functions of integrins and protease-activated receptors is critical to control GSK3 β activity.

In inflammation, long-term GSK3 β activation should be deleterious through a decrease of regenerative capacities. Indeed, after acute inflammation, stem cells maintain their quiescent phenotype showing decreased proliferation and migration, as well as poor differentiation. In addition to immunity control, tissue regeneration is a major aim in IBD therapy [48].

Prolonged GSK3 β activation in inflammatory niche could maintain abnormal stem cells, avoiding apoptosis (i.e., mitotic catastrophe). As a result, stem cells accumulating mutations or chromosomal aberrations reside in the tissue until the release of their dormant state by epigenomic or genomic changes inducing proliferation [4]. These pre-cancerous stem cells are characterized by genomic instability and re-expression of embryonic genes, as well as dependence to the normal niche [49]. Such pre-leukemic stem cells with high GSK3 β activity have been described in transition to cancer stem cells with deregulated β -catenin [50]. Importantly, due to the different pools of GSK3 β , an aggressive cancer stem cell can cumulate both active GSK3 β -dependent survival and β -catenin-dependent

self-renewal deregulation. Indeed, we have measured higher clonogenic capacities of leukemic progenitors displaying GSK3 β -dependent drug resistance [26].

The capacity to activate GSK3 β in adult stem cells could be the property of specific receptors after their binding to matrix proteins (α_5 , α_2 integrins, [25,51]) or their protease-dependent cleavage (PAR₂, [42]). Also, the GSK3 β activation could result from the dialog between integrins and PAR₂ [42,52]. The α_5 and α_2 integrin-binding proteins, fibronectin and collagen respectively, and PAR₂ have been implicated in cell plasticity [53–55]. Prolonged GSK3 β activation could induce a switch in stem cell identity that influences the responses to the microenvironment, paving the way to tumor transformation [56] (Figure 5).

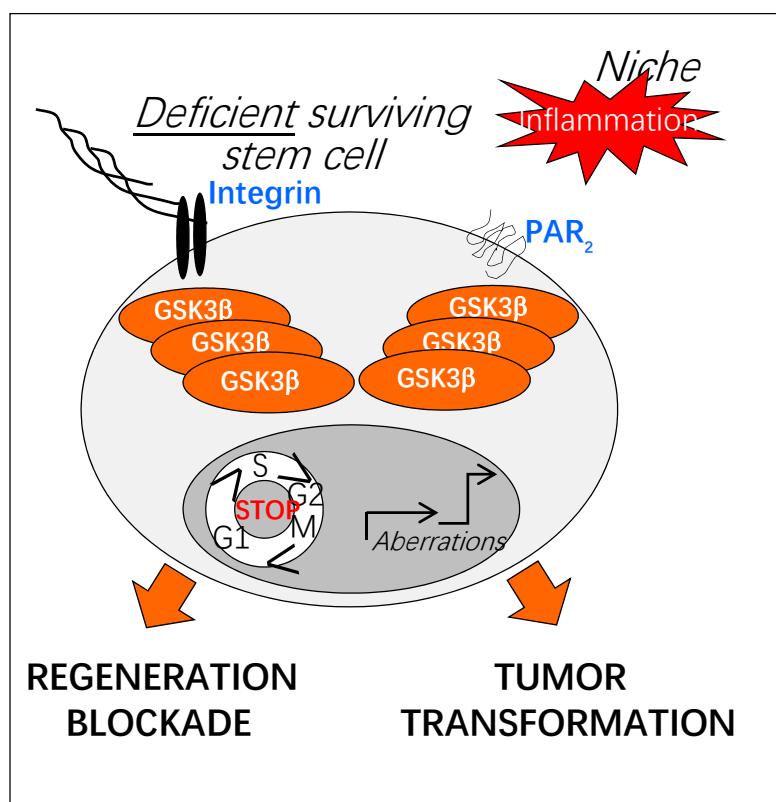


Figure 5. Potential impact of prolonged GSK3 β activation on regenerative functions of adult stem cells. Upon chronic inflammation, increased activation of GSK3 β could result in a deregulation of the regenerative properties of adult stem cells (defect in tissue repair, tumor transformation). As sensors of the injured stem cell niche, adhesion and protease-activated receptors (Integrins, PARs) could play a crucial role in the sustained activation of GSK3 β .

7. GSK3 β as a Target for Precision Medicine

GSK3 β has an important theranostic potential in pathologies where stem cells are deregulated, such as inflammation [57] and cancer [58]. It seems that the activated GSK3 β signature could involve subsets of pathological progenitor/stem cells depending both on their membrane receptors and the adhesive and protease activities in their niche. For example, GSK3 β has a key oncogenic role in leukemia with MLL mutations [50] and intestinal neuroendocrine tumors [59]. Interestingly, the cancer stem cells in those pathologies could be developed from early progenitors [60] through the dialog with an inflammatory [61,62] and neurologic-deficient [63,64] microenvironment.

Clinical parameters as gender, age and metabolic status of patients should strongly influence therapeutic decisions aiming to target GSK3 β in inflammatory and cancer diseases. We have seen above that the sexual dimorphism occurring in GSK3 β -dependent regulation of adult stem cells could be inverted during the transition from inflammation to cancer.

Also, aging and obesity could modify cell responses to insulin and to stresses. Therefore, therapeutic targeting of GSK3 β must be thought as a personalized medicine.

Measurement of the activity of GSK3 β is complex due to its different cellular pools with independent regulations. However, when human samples are available, 2D or 3D stem cell cultures with controlled addition of growth factors represent good investigating tools for pre-clinical assays with adhesive conditions akin to the tissue architecture [26,42]. Also, active GSK3 imaging agents are in development for positron emission tomography as a diagnostic tool but require yet a sufficient knowledge of pathways governing GSK3 β regulation [65].

Natural and synthetic GSK3 inhibitors with different modes of action are already commercialized and some of them are in clinical trials for the treatment of neurodegenerative diseases and cancer [65,66]. We and others have found that natural compounds and their derivatives are potential drugs to kill cancer stem cells through GSK3 β inhibition [67,68]. It is important to note that GSK3 inhibitors represent also interesting therapeutic tools to restore normal niche functions [69]. Thus, targeting both stem cells and their niche is crucial to restore physiological tissue regeneration (Figure 6).

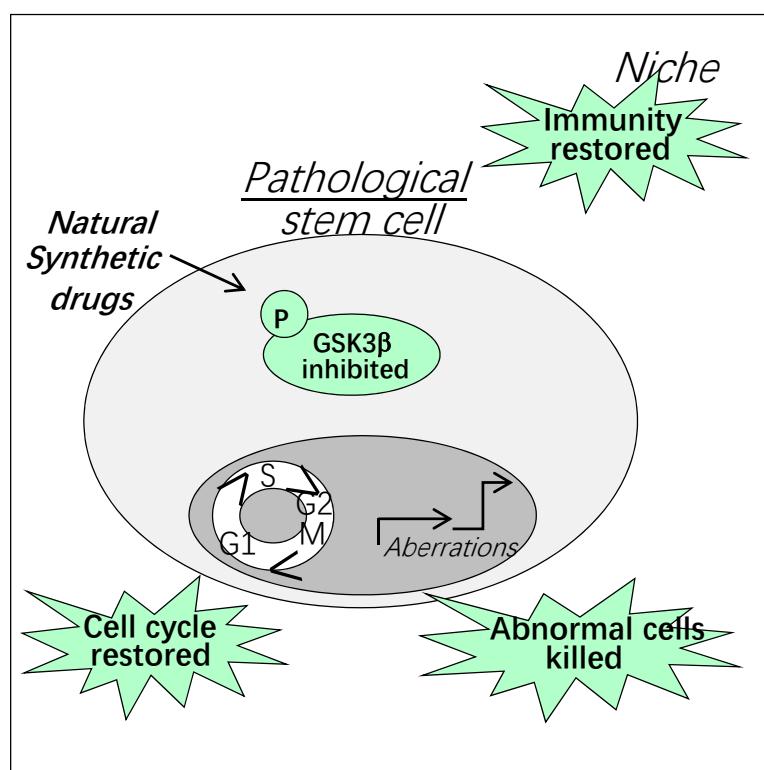


Figure 6. Therapeutic targeting of GSK3 β in pathologic stem cells and their niche.

8. Conclusions and Future Directions

GSK3 β is a master kinase in the regulation of adult stem cells through the control of common mechanisms in different cells such as hematopoietic and intestinal stem cells. It is a critical sensor of the microenvironment allowing important phenotypic changes in stem cells for their adaptation and their maintenance. Our previous work demonstrated that the GSK3 β activation is supervised by a balanced control between adhesion and protease-activated receptors in the regulation of adult stem cell behavior.

However, the protective role of GSK3 β can be perverted in pathogenesis by the maintenance of stem cells with functional deficits and genetic aberrations. This is the reason why the interest for GSK3 β as a therapeutic target continues to grow with increased rate of publications and pre-clinical trials. In inflammation and cancer, targeting of GSK3 β could restore a normal interaction of stem cells with their microenvironment and consequently

homeostatic tissue regeneration. An increasing number of studies now strongly suggests that therapeutic targeting of GSK3 β must take into account clinical parameters such as gender and metabolic status of patients.

Progress is required both in fundamental and clinical research to improve our knowledge of GSK3 β . As an ancestral kinase of the stress response in eukaryotes, attention should be drawn towards works on its orthologue shaggy in drosophila and other primitive organisms. Indeed, to cope with stress, vertebrate adult stem cells develop mechanisms based on ancient roots. Also, for a theranostic purpose, diagnostic and therapeutic tools specific for GSK3 β versus its isoform GSK3 α are necessary. Rapid advances in the research about neurodegenerative disorders should offer opportunities in this field.

Author Contributions: C.R.-S., writing and original draft preparation; C.R.-S. and N.V., review and editing. All authors have read and agreed to the published version of the manuscript.

Funding: This work was funded by grants from the Institut National du Cancer (Contract No. 07/3D1616/IABC-23-8/NC-NG), Association de Recherche contre le Cancer (Contract Nos. 3638 and 8407) and DGRSRT/Inserm cooperation to C. R-S., the European Research Council (ERC-310973 PIPE) and the region Occitanie to N. V.

Acknowledgments: The authors thank Michèle Allouche for critical reading of the manuscript.

Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

References

1. Beurel, E.; Grieco, S.F.; Jope, R.S. Glycogen synthase kinase-3 (GSK3): Regulation, actions and diseases. *Pharm. Ther.* **2015**, *148*, 114–131. [[CrossRef](#)] [[PubMed](#)]
2. Murata, M. Inflammation and cancer. *Environ. Health Prev. Med.* **2018**, *23*, 50. [[CrossRef](#)]
3. Chacon-Martinez, A.C.; Koester, J.; Wickström, S.A. Signaling in the stem cell niche: Regulating cell fate, function and plasticity. *Development* **2018**, *145*, dev165399. [[CrossRef](#)] [[PubMed](#)]
4. Fiore, A.P.Z.P.; Ribeiro, P.d.F.; Bruni-Cardoso, A. Sleeping beauty and the microenvironment enchantment: Microenvironmental regulation of the proliferation-quiescence decision in normal tissues and in cancer development. *Front. Cell Dev. Biol.* **2018**, *6*, 59. [[CrossRef](#)]
5. Dimova, N.; Wysoczynski, M.; Rokosh, G. SDF1 promotes CSPC quiescence via CK1 α and GSK3 β . *Stem Cells* **2014**, *32*, 487–499.
6. Hayes, J.D.; Dinkova-Kostova, A.T. The Nrf2 regulatory network provides an interface between redox and intermediary metabolism. *Trends Biochem. Sci.* **2014**, *39*, 199–218. [[CrossRef](#)]
7. Kwon, S.M.; Hong, S.M.; Lee, Y.-K.; Min, S.; Yoon, G. Metabolic features and regulation in cell senescence. *BMB Rep.* **2019**, *52*, 5–12. [[CrossRef](#)]
8. Patel, P.; Woodgett, J.R. Glycogene synthase kinase 3: A kinase for all pathways? *Curr. Top. Dev. Biol.* **2017**, *123*, 277–302.
9. Huang, J.; Zhang, Y.; Bersenev, A.; O'Brien, W.T.; Tong, W.; Emerson, S.G.; Klein, P.S. Pivotal role of glycogen synthase kinase-3 in hematopoietic stem cell homeostasis in mice. *J. Clin. Investig.* **2009**, *119*, 3519–3529. [[CrossRef](#)]
10. Tsuchiya, K.; Nakamura, T.; Okamoto, R.; Takanori, K.; Watanabe, M. Reciprocal targeting of Hath1 and β -catenin by Wnt Glycogen synthase kinase 3 β in human colon cancer. *Gastroenterology* **2007**, *132*, 208–220. [[CrossRef](#)]
11. Gao, X.; Li, X.; Qian, C.; Li, F.; Zhang, Y.; Dang, L.; Xiao, X.; Liu, F.; Li, H.; Zhang, X. MiR-21 functions oppositely in proliferation and differentiation of neural stem/precursor cells via regulating AKT and GSK-3 β . *Cell. Mol. Biol.* **2016**, *62*, 144–149. [[CrossRef](#)] [[PubMed](#)]
12. Trowbridge, J.J.; Xenocostas, A.; Moon, R.T.; Bhatia, M. Glycogen synthase kinase-3 is an vivo regulator of hematopoietic stem cell repopulation. *Nat. Med.* **2006**, *12*, 89–98. [[CrossRef](#)] [[PubMed](#)]
13. Lapid, K.; Itkin, T.; D'Uva, G.; Ovadya, Y.; Ludin, A.; Caglio, G.; Kalinkovich, A.; Golan, K.; Porat, Z.; Zollo, M.; et al. GSK3b regulates physiological migration of stem/progenitor cells via cytoskeletal rearrangement. *J. Clin. Investig.* **2013**, *123*, 1705–1717. [[CrossRef](#)] [[PubMed](#)]
14. Kobayashi, T.; Hino, S.; Oue, N.; Asahara, T.; Zollo, M.; Yasui, W.; Kikuchi, A. Glycogen synthase kinase 3 and h-prune regulate cell migration by modulating focal adhesions. *Mol. Cell. Biol.* **2006**, *26*, 898–911. [[CrossRef](#)]
15. Turano, M.; Costabile, V.; Cerasuolo, A.; Duratoro, F.; Liccardo, R.; Delrio, P.; Pace, U.; Rega, D.; Dodaro, C.A.; Milone, M.; et al. Characterisation of mesenchymal colon tumour-derived cells in tumourspheres as a model for colorectal cancer progression. *Int. J. Oncol.* **2018**, *53*, 2379–2396. [[CrossRef](#)]
16. Sun, M.; Song, L.; Li, Y.; Zhou, T.; Jope, R.S. Identification of an antiapoptotic protein complex at death receptors. *Cell Death Differ.* **2008**, *15*, 1887–1900. [[CrossRef](#)]

17. Schwabe, R.F.; Brenner, D.A. Role of glycogen synthase kinase-3 in TNF- α -induced NF- κ B activation and apoptosis in hepatocytes. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2002**, *283*, G204–G211. [[CrossRef](#)]
18. Steinbrecher, K.A.; Wilson III, W.; Cogswell, P.C.; Baldwin, A.C. Glycogen synthase kinase 3 β functions to specify gene-specific, NF- κ B-dependent transcription. *Mol. Cell. Biol.* **2005**, *25*, 8444–8455. [[CrossRef](#)]
19. Ougolkov, A.V.; Bone, N.D.; Fernandez-Zapico, M.E.; Kay, N.E.; Billadeau, D.D. Inhibition of glycogen synthase kinase-3 activity leads to epigenetic silencing of nuclear factor kB target genes and induction of apoptosis in chronic lymphocytic leukemia B cells. *Blood* **2007**, *110*, 735–742. [[CrossRef](#)]
20. Ha, S.; Ryu, H.Y.; Chung, K.M.; Baek, S.H.; Kim, E.K.; Yu, S.W. Regulation of autophagic cell death by glycogen synthase kinase-3 β in adult hippocampal neural stem cells following insulin withdrawal. *Mol. Brain* **2015**, *8*, 30–42. [[CrossRef](#)]
21. McCubrey, J.A.; Steelman, L.S.; Bertrand, F.E.; Davis, N.M.; Abrams, S.L.; Montalto, G.; D’Assoro, A.B.; Libra, M.; Nicoletti, F.; Maestro, R.; et al. Multifaceted roles of GSK-3 and Wnt/ β -catenin in hematopoiesis and leukemogenesis: Opportunities for therapeutic intervention. *Leukemia* **2014**, *28*, 15–33. [[CrossRef](#)] [[PubMed](#)]
22. Holmes, T.; O’Brien, T.; Knight, R.; Lindeman, R.; Shen, S.; Song, E.; Symonds, G.; Dolnikov, A. Glycogen synthase kinase-3 β inhibition preserves hematopoietic stem cell activity and inhibits leukemic cell growth. *Stem Cells* **2008**, *26*, 1288–1297. [[CrossRef](#)] [[PubMed](#)]
23. Duda, P.; Akula, S.M.; Abrams, S.L.; Steelman, L.S.; Martelli, A.M.; Cocco, L.; Ratti, S.; Candido, S.; Libra, M.; Montalto, G.; et al. Targeting GSK3 and associated signaling pathways involved in cancer. *Cells* **2020**, *9*, 1110. [[CrossRef](#)] [[PubMed](#)]
24. De Toni, F.; Racaud-Sultan, C.; Chicanne, G.; Mansat-De Mas, V.; Cariven, C.; Mesange, F.; Salles, J.P.; Demur, C.; Allouche, M.; Payrastre, B.; et al. A crosstalk between the Wnt and the adhesion-dependent signaling pathways governs the chemosensitivity of acute myeloid leukemia. *Oncogene* **2006**, *25*, 3113–3122. [[CrossRef](#)] [[PubMed](#)]
25. De Toni-Costes, F.; Despeaux, M.; Bertrand, J.; Bourogaa, E.; Ysebaert, L.; Payrastre, B.; Racaud-Sultan, C. A new $\alpha_5\beta_1$ integrin-dependent survival pathway through GSK3 β activation in leukemic cells. *PLoS ONE* **2010**, *5*, e9807. [[CrossRef](#)]
26. Bertrand, J.; Despeaux, M.; Joly, S.; Bourogaa, E.; Gallay, N.; Demur, C.; Bonnevialle, P.; Louache, F.; Maguer-Satta, V.; Vergnolle, N.; et al. Sex differences in the GSK3 β -mediated survival of adherent leukemic progenitors. *Oncogene* **2012**, *31*, 694–705. [[CrossRef](#)]
27. Ge, Y.; Fuchs, E. Stretching the limits: From homeostasis to stem cell plasticity in wound healing and cancer. *Nat. Rev. Genet.* **2018**, *19*, 311–325. [[CrossRef](#)]
28. Despeaux, M.; Labat, E.; Gadelorge, M.; Prade, N.; Bertrand, J.; Demur, C.; Recher, C.; Bonnevialle, P.; Payrastre, B.; Bourin, P.; et al. Critical features of FAK-expressing AML bone marrow microenvironment through leukemia stem cell hijacking of mesenchymal stromal cells. *Leukemia* **2011**, *25*, 1789–1793. [[CrossRef](#)]
29. Larabi, A.; Barnich, N.; Nguyen, H.T.T. New insights into the interplay between autophagy, gut microbiota and inflammatory responses in IBD. *Autophagy* **2020**, *16*, 38–51. [[CrossRef](#)]
30. Obermeier, F.; Hofmann, C.; Falk, W. Inflammatory bowel diseases: When natural friends turn into enemies-The importance of CpG motifs of bacterial DNA in intestinal homeostasis and chronic intestinal inflammation. *Int. J. Inflam.* **2010**, *2010*, 641910. [[CrossRef](#)]
31. Vidri, R.J.; Fitzgerald, T.L. GSK-3: An important kinase in colon and pancreatic cancers. *Biochim. Biophys. Acta Mol. Cell Res.* **2020**, *1867*, 118626. [[CrossRef](#)] [[PubMed](#)]
32. Hong, A.W.; Meng, Z.; Guan, K.-L. The Hippo pathway in intestinal regeneration and disease. *Nat. Rev. Gastroenterol. Hepatol.* **2016**, *13*, 324–337. [[CrossRef](#)] [[PubMed](#)]
33. Huels, D.J.; Ridgway, R.A.; Radulescu, S.; Leushacke, M.; Campbell, A.D.; Biswas, S.; Leedham, S.; Serra, S.; Chetty, R.; Moreaux, G.; et al. E-cadherin can limit the transforming properties of activating β -catenin mutations. *EMBO J.* **2015**, *34*, 2321–2333. [[CrossRef](#)] [[PubMed](#)]
34. Mai, W.; Kawakami, K.; Shakoori, A.; Kyo, S.; Miyashita, K.; Yokoi, K.; Jin, M.; Shimasaki, T.; Motoo, Y.; Minamoto, T. Deregulated GSK3b sustains gastrointestinal cancer cells survival by modulating human telomerase reverse transcriptase and telomerase. *Clin. Cancer Res.* **2009**, *15*, 6810–6819. [[CrossRef](#)] [[PubMed](#)]
35. Ghosh, J.C.; Altieri, D.C. Activation of p53-dependent apoptosis by acute ablation of glycogen synthase kinase-3beta in colorectal cancer cells. *Clin. Cancer Res.* **2005**, *11*, 4580–4588. [[CrossRef](#)]
36. Grassilli, E.; Narloch, R.; Federzoni, E.; Lanzano, L.; Pisano, F.; Giovanonni, R.; Romano, G.; Masiero, L.; Leone, B.E.; Bonin, S.; et al. Inhibition of GSK3B bypass drug resistance of p53-null colon carcinomas by enabling necroptosis in response to chemotherapy. *Clin. Cancer Res.* **2013**, *19*, 3820–3831. [[CrossRef](#)]
37. Motta, J.P.; Magne, L.; Descamps, D.; Rolland, C.; Squarzoni-Dale, C.; Rousset, P.; Martin, L.; Cenac, N.; Balloy, V.; Huerre, M.; et al. Modifying the protease, antiprotease pattern by elafin overexpression protects mice from colitis. *Gastroenterology* **2011**, *140*, 1272–1282. [[CrossRef](#)]
38. Motta, J.P.; Bermudez-Humaran, L.G.; Deraison, C.; Martin, L.; Rolland, C.; Rousset, P.; Boue, J.; Dietrich, G.; Chapman, K.; Kharrat, P.; et al. Food-grade bacteria expressing elafin protect against inflammation and restore colon homeostasis. *Sci. Transl. Med.* **2012**, *4*, 158ra144. [[CrossRef](#)]
39. Denadai-Souza, A.; Bonnart, C.; Tapias, N.S.; Marcellin, M.; Gilmore, B.; Alric, L.; Bonnet, D.; Burlet-Schiltz, O.; Hollenberg, M.D.; Vergnolle, N.; et al. Functional proteomic profiling of secreted serine proteases in health and inflammatory bowel disease. *Sci. Rep.* **2018**, *8*, 7834–7843. [[CrossRef](#)]

40. Motta, J.P.; Palese, S.; Giorgio, C.; Chapman, K.; Denadai-Souza, A.; Rousset, P.; Sagnat, D.; Guiraud, L.; Edir, A.; Seguy, C.; et al. Increased mucosal thrombin is associated with Crohn's disease and causes inflammatory damage through protease-activated receptors activation. *J. Crohns Colitis* **2020**, *jcaa229*. [[CrossRef](#)]
41. Vergnolle, N. Protease inhibition as a new therapeutic strategy for GI diseases. *Gut* **2016**, *65*, 1215–1224. [[CrossRef](#)] [[PubMed](#)]
42. Nasri, I.; Bonnet, D.; Zwarycz, B.; d'Aldebert, E.; Khou, S.; Mezghani-Jarraya, R.; Quaranta, M.; Rolland, C.; Bonnart, C.; Mas, E.; et al. PAR₂-dependent activation of GSK3β regulates the survival of colon stem/progenitor cells. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2016**, *311*, G221–G236. [[CrossRef](#)] [[PubMed](#)]
43. Haber, A.L.; Biton, M.; Rogel, N.; Herbst, R.H.; Shekhar, K.; Smillie, C.; Burgin, G.; Delorey, T.M.; Howitt, M.R.; Katz, Y.; et al. A single-cell survey of the small intestinal epithelium. *Nature* **2017**, *551*, 333–342. [[CrossRef](#)] [[PubMed](#)]
44. Parikh, K.; Antanaviciute, A.; Fawkner-Corbett, D.; Jagielowicz, M.; Aulicino, A.; Lagerholm, C.; Davis, S.; Kinchen, J.; Chen, H.H.; Alham, N.K.; et al. Colonic epithelial cell diversity in health and inflammatory bowel disease. *Nature* **2019**, *567*, 49–55. [[CrossRef](#)] [[PubMed](#)]
45. Xu, D.; Song, R.; Wang, G.; Jeyabal, P.V.S.; Weiskoff, A.M.; Ding, K.; Shi, Z.-Z. Obg-like ATPase 1 regulates global protein serine/threonine phosphorylation in cancer cells by suppressing the GSK3β-inhibitor 2-PP1 positive feedback loop. *Oncotarget* **2016**, *7*, 3427–3439. [[CrossRef](#)]
46. He, L.; Ma, Y.; Li, W.; Han, W.; Zhao, X.; Wang, H. Protease-activated receptor 2 signaling modulates susceptibility of colonic epithelium to injury through stabilization of YAP in vivo. *Cell Death Dis.* **2018**, *9*, 949–960. [[CrossRef](#)]
47. Noguerol, J.; Roustan, P.J.; N'Taye, M.; Delcombel, L.; Rolland, C.; Guiraud, L.; Sagnat, D.; Edir, A.; Bonnart, C.; Denadai-Souza, A.; et al. Sexual dimorphism in PAR₂-dependent regulation of primitive colonic cells. *Biol. Sex. Differ.* **2019**, *10*, 47. [[CrossRef](#)]
48. Okamoto, R.; Shimizu, H.; Suzuki, K.; Kawamoto, A.; Takahashi, J.; Kawai, M.; Nagata, S.; Hiraguri, Y.; Takeoka, S.; Sugihara, H.Y.; et al. Organoid-based regenerative medicine for inflammatory bowel disease. *Regen. Ther.* **2020**, *13*, 1–6. [[CrossRef](#)]
49. Gao, J.X. Cancer stem cells: The lessons from pre-cancerous stem cells. *J. Cell. Mol. Med.* **2008**, *12*, 67–96. [[CrossRef](#)]
50. Yeung, J.; Esposito, M.T.; Gandillet, A.; Zeisig, B.B.; Griessinger, E.; Bonnet, D.; So, C.W.E. β-catenin mediates the establishment and drug resistance of MLL leukemic stem cells. *Cancer Cell* **2010**, *18*, 606–618. [[CrossRef](#)]
51. Ivaska, J.; Nissinen, L.; Immonen, N.; Eriksson, J.E.; Kahari, V.M.; Heino, J. Integrinα₂β₁ promotes activation of protein phosphatase 2A and dephosphorylation of Akt and glycogen synthase kinase 3β. *Mol. Cell. Biol.* **2002**, *22*, 1352–1359. [[CrossRef](#)] [[PubMed](#)]
52. Miyata, S.; Koshikawa, N.; Yasumitsu, H.; Miyazaki, K. Trypsin stimulates integrin α₅β₁-dependent adhesion to fibronectin and proliferation of human gastric carcinoma cells through activation of proteinase-activated receptor-2. *J. Biol. Chem.* **2000**, *275*, 4592–4598. [[CrossRef](#)] [[PubMed](#)]
53. Choi, J.S.; Harley, B.A.C. Marrow-inspired matrix cues rapidly affect early fate decisions of hematopoietic stem and progenitors cells. *Sci. Adv.* **2017**, *3*, e1600455. [[CrossRef](#)] [[PubMed](#)]
54. Hussey, G.S.; Cramer, M.C.; Badylak, S.F. Extracellular matrix bioscaffolds for building gastrointestinal tissue. *Cell Mol. Gastroenterol. Hepatol.* **2018**, *5*, 1–13. [[CrossRef](#)]
55. Piran, R.; Lee, S.H.; Kuss, P.; Hao, E.; Newlin, R.; Millan, J.L.; Levine, F. PAR2 regulates regeneration, transdifferentiation, and death. *Cell Death Dis.* **2016**, *7*, e2452. [[CrossRef](#)]
56. Li, Z.; Guo, X.; Huang, H.; Wang, C.; Yang, F.; Zhang, Y.; Wang, J.; Han, L.; Jin, Z.; Cai, T.; et al. A switch in tissue stem cell identity causes neuroendocrine tumors in drosophila gut. *Cell Rep.* **2020**, *30*, 1724–1734. [[CrossRef](#)]
57. Jope, R.S.; Cheng, Y.; Lowell, J.; Worthen, R.J.; Sitbon, Y.H.; Beurel, E. Stressed and inflamed, can GSK3 be blamed? *Trends Biochem. Sci.* **2017**, *42*, 180–192. [[CrossRef](#)]
58. Nagini, S.; Sophia, J.; Mishra, R. Glycogen synthase kinases: Moonlighting proteins with theranostic potential in cancer. *Semin. Cancer Biol.* **2019**, *56*, 25–36. [[CrossRef](#)]
59. Aristizabal Prada, E.T.; Weis, C.; Orth, M.; Lauseker, M.; Spöttl, G.; Maurer, J.; Grabowski, P.; Grossman, A.; Auernhammer, C.J.; Nölting, S. GSK3α/β: A novel therapeutic target for neuroendocrine tumors. *Neuroendocrinology* **2018**, *106*, 335–351. [[CrossRef](#)]
60. Cleary, M.L. Regulating the leukemia stem cell. *Best Pract. Res. Clin. Haematol.* **2009**, *22*, 483–487. [[CrossRef](#)]
61. Corrigan, D.J.; Luchsinger, L.L.; Justino De Almeida, M.; Williams, L.J.; Strikoudis, A.; Snoeck, H.-W. PRDM16 isoforms differentially regulate normal and leukemic hematopoiesis and inflammatory gene signature. *J. Clin. Investig.* **2018**, *128*, 3250–3264. [[CrossRef](#)] [[PubMed](#)]
62. Vitale, G.; Carra, S.; Ferrau, F.; Guadagno, E.; Faggiano, A.; Colao, A. Gastroenteropancreatic neuroendocrine neoplasms and inflammation: A complex cross-talk with relevant clinical implications. *Crit. Rev. Oncol. Hematol.* **2020**, *146*, 102840. [[CrossRef](#)] [[PubMed](#)]
63. Hanoun, M.; Zhang, D.; Mizoguchi, T.; Pinho, S.; Pierce, H.; Kunisaki, Y.; Lacombe, J.; Armstrong, S.A.; Dührsen, U.; Frenette, P.S. Acute myelogenous leukemia-induced sympathetic neuropathy promotes malignancy in an altered hematopoietic stem cell niche. *Cell Stem Cell* **2014**, *15*, 365–375. [[CrossRef](#)]
64. Sinha, S.; Fu, Y.-Y.; Grimont, A.; Ketcham, M.; Lafaro, K.; Saglimbeni, J.A.; Askan, G.; Bailey, J.M.; Melchor, J.P.; Zhong, Y.; et al. PanIN neuroendocrine cells promote tumorigenesis via neuronal crosstalk. *Cancer Res.* **2017**, *77*, 1868–1879. [[CrossRef](#)] [[PubMed](#)]
65. Pandey, M.K.; DeGrado, T.R. Glycogen synthase kinase-3 (GSK-3)-targeted therapy and imaging. *Theranostics* **2016**, *6*, 571–593. [[CrossRef](#)]

66. Khan, I.; Tantray, M.A.; Alam, M.S.; Hamid, H. Natural and synthetic bioactive inhibitors of glycogen synthase kinase. *Eur. J. Med. Chem.* **2017**, *125*, 464–477. [[CrossRef](#)]
67. Bourogaa, E.; Bertrand, J.; Despeaux, M.; Jarraya, R.; Fabre, N.; Payrastre, L.; Demur, C.; Fournié, J.-J.; Damak, M.; El Feki, A.; et al. Hammada scoparia flavonoids and rutin kill adherent and chemoresistant leukemic cells. *Leuk. Res.* **2011**, *35*, 1093–1101. [[CrossRef](#)]
68. Nasri, I.; Chawech, R.; Girardi, C.; Mas, E.; Ferrand, A.; Vergnolle, N.; Fabre, N.; Mezghani-Jarraya, R.; Racaud-Sultan, C. Anti-inflammatory and anticancer effects of flavonol glycosides from Diplotaxis harra through GSK3 β regulation in intestinal cells. *Pharm. Biol.* **2017**, *55*, 124–131. [[CrossRef](#)]
69. Augello, G.; Emma, M.R.; Cusimano, A.; Azzolina, A.; Montalto, G.; McCubrey, J.A.; Cervello, M. The role of GSK-3 in cancer immunotherapy: GSK-3 inhibitors as a new frontier in cancer treatment. *Cells* **2020**, *9*, 1427. [[CrossRef](#)]