

Dynamics of bacterial and fungal rhizosphere communities during the infection of Brassica rapa roots by the protist Plasmodiophora brassicae

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Dynamics of bacterial and fungal rhizosphere communities during the infection of *Brassica rapa* roots by the protist *Plasmodiophora brassicae*

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This work put the bacterial and fungal diversities in relation with *Brassica rapa* – obligate parasite *Plasmodiophora brassicae* (clubroot disease) interaction in the rhizosphere (Fig 1).

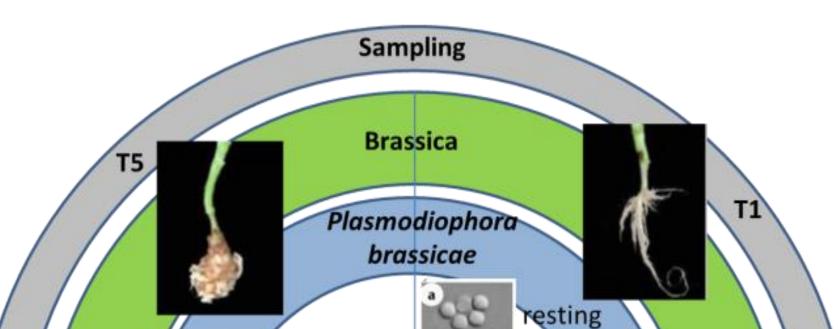


Diversity analysis allow the identification of potential key microbial players during the disease development

A GROWTH CHAMBER POT-EXPERIMENT

A split-plot experimental design

- 240 Brassica rapa cv. Granaat cultivated in a silty-clay soil and inoculated (I) or not (NI) with P. brassicae strain eH
- ➢ Growth chamber conditions: 16h light, 8h dark, 22°C/19°C



SOIL

ROOT HAIR

plasmode I

zoosporange

ROOT

zoospore II

plasmode II

spores

zoospore l 🕑

Fig1. Life cycle of Plasmodiophora brassicae (blue circle), the pathogen that causes clubroot, in relation with Brassica phenotype (green circle). The sampling strategy for plant trait measurements and molecular ecology is indicated in the grey circle

- 3 pools of 4 plants phenotyped at T1 (inoculation), and 7 (T2), 14 (T3), 23 (T4), 35 days (T5) after inoculation
- Plant traits : hypocotyl section and disease index

Microbial communities analysis

- Sampling of rhizosphere soil (RS) and bulk soil (BS) from inoculated (I) and not inoculated (NI) plants at each sampling date between T1 and T5
- DNA extraction following by a 16S and 18S rDNA amplicons sequencing performed by Illumina MiSeq platform
- Base trimming, assembly with a minimum of 30 bases overlapping 100% homology between reads
- Strict dereplication, clustering (97% similarity) and taxonomic assignation (SILVA databases) (GnS-PIPE workflow)
- Statistical analysis with Bioconductor package and edgeR under R

Plant traits related to disease development

- Differences of hypocotyl section from inoculated (I) and not inoculated (NI) plants starting 14 days after inoculation (Fig 2).
- Disease index of 15% at T3, 70% at T3 and 90% at T5 suggesting high sensibility of *B. rapa* cv. *Granaat* to clubroot.

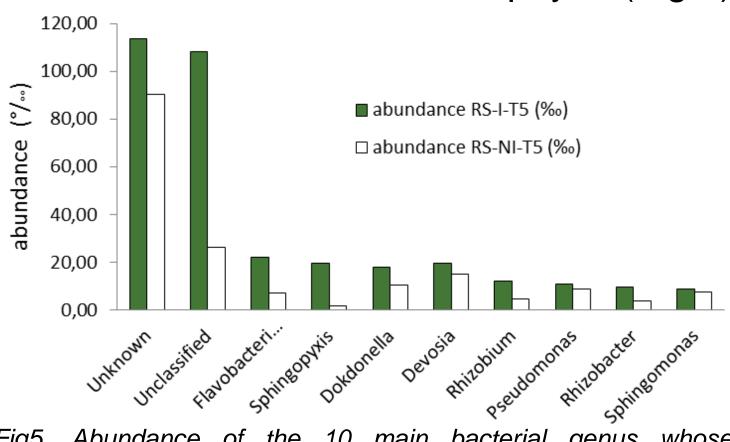
Bacterial and fungal communities diversity evolved during disease development

KEY RESULTS

- Means of 100000 and 45000 reads clustering in 10000 and 1600 OTUs for bacterial and fungal communities, respectively.
- Spatial (BS versus RS) and temporal effects on microbial diversity (Fig 3 and 4).



Among 416 genus, the frequencies of 90 genus were significantly modified between inoculated and not inoculated treatments at T5. The 8 main genus significantly modified belonged to the *Proteobacteria* and *Bacteroidetes* phyla (Fig 5).



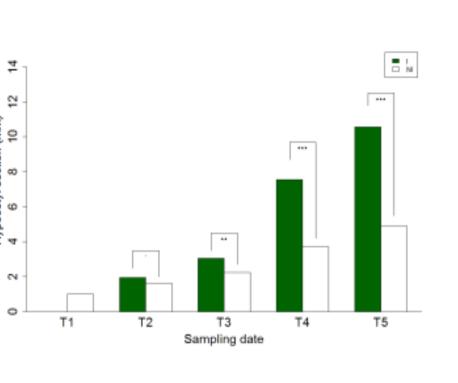


Fig 2 . Evolution of hypocotyl section from not inoculated (NI) and inoculated plants (I) between T1 to T5

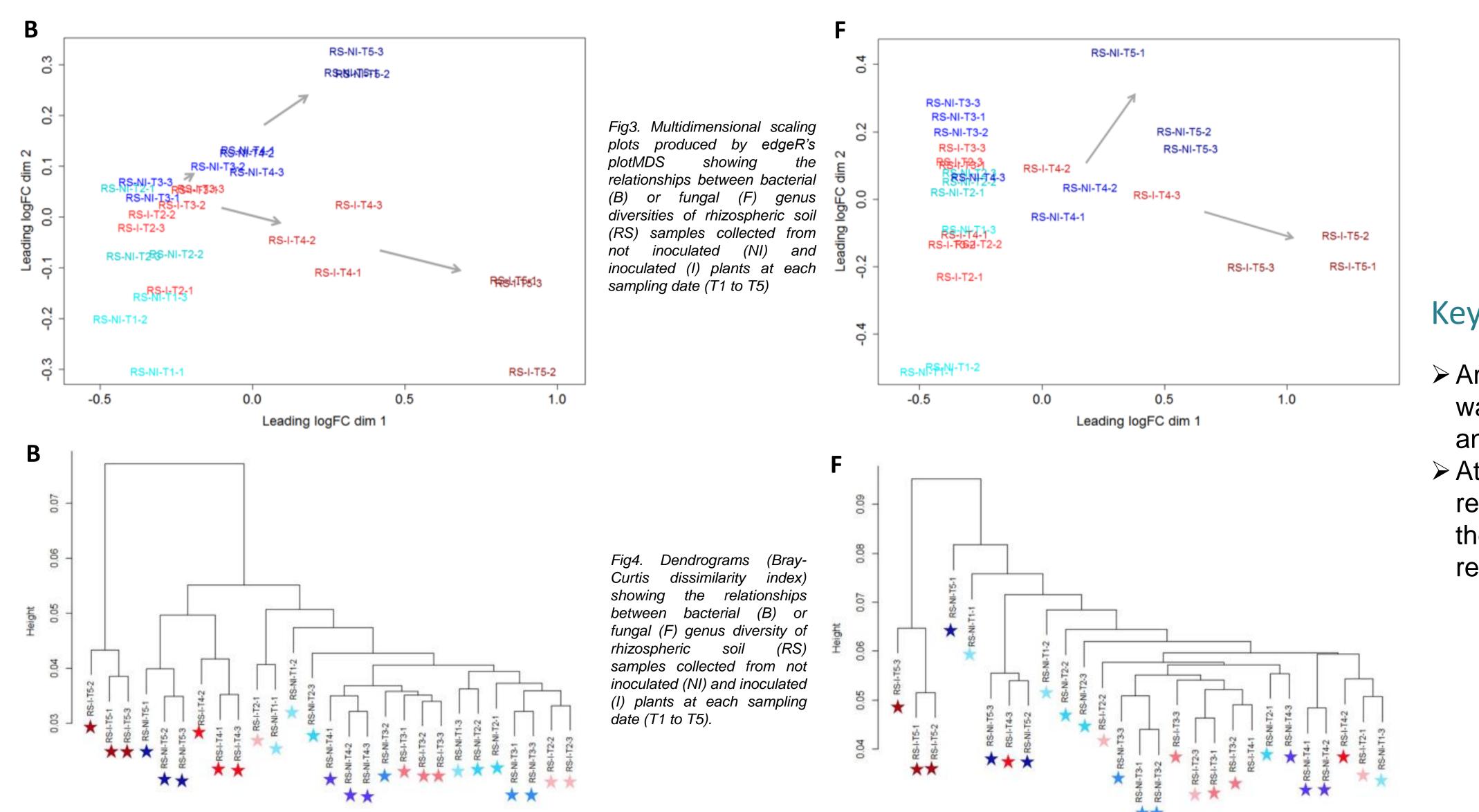
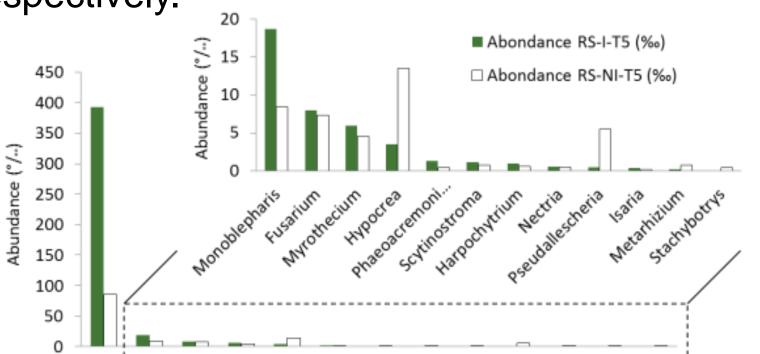


Fig5. Abundance of the 10 main bacterial genus whose frequencies were significantly increased at T5 in the rhizosphere of inoculated plants compared to healty plants (p < 0.05)

Key rhizosphere fungal taxa identified

- Among 100 genus, the occurrence of 13 genus was significantly modified between inoculated and not inoculated treatments at T5 (Fig 6).
- At T5, the dominant fungus is Olpidium sp. which represents 38 % and 10 % of total sequences in the healthy and diseased rhizosphere, respectively.



TAKE HOME MESSAGE

> First description of the dynamics of bacterial and fungal communities during clubroot development.

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- > The strongest difference between microbial diversity appeared at the latest stages of disease development.
- The occurrence of bacterial genus belonging to Proteobacteria and Bacteroidetes, and the incertae sedis fungus Olpidium was strongly increased in the diseased plant rhizosphere when clubroot severity is higher.
- Proteobacteria, Bacteroidetes and Olpidium were host-specific rhizosphere genus able to develop intimate interactions with plant and seem to be key players of Brassica-Plasmodiophora interaction. Future directions will precise functional basis of these interactions.

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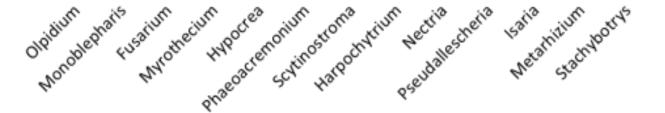


Fig6. Abundance of the 13 fungal genus whose frequencies were siginificantly modified at T5 in the rhizosphere of inoculated plants compared to healty plants (p < 0.05)

