



**HAL**  
open science

## Does rice breeding affect the ability of plants to interact with earthworms in nutrient-depleted Ferralsols?

O. Ratsiatosika, E. Blanchart, T. Razafimbelo, M. Razafindrakoto, K. Vom Brocke, T.-V. Cao-Hamadou, J.-M. Rakotomalala Andriamarosata, A. Ramanantsoanirina, J. Trap

### ► To cite this version:

O. Ratsiatosika, E. Blanchart, T. Razafimbelo, M. Razafindrakoto, K. Vom Brocke, et al.. Does rice breeding affect the ability of plants to interact with earthworms in nutrient-depleted Ferralsols?. Applied Soil Ecology, 2021, 163, 10.1016/j.apsoil.2021.103958 . hal-03259287

**HAL Id: hal-03259287**

**<https://hal.inrae.fr/hal-03259287v1>**

Submitted on 9 May 2023

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial 4.0 International License

1 Does rice breeding affect the ability of plants to interact with earthworms in nutrient-depleted  
2 Ferralsols?

3

4 O. Ratsiatosika<sup>a</sup>, E. Blanchart<sup>b</sup>, T. Razafimbelo<sup>a</sup>, M. Razafindrakoto<sup>a</sup>, K. vom Brocke<sup>c</sup>, T-V.  
5 Cao-Hamadou<sup>d</sup>, J-M. Rakotomalala Andriamarosata<sup>c</sup>, A. Ramanantsoanirina<sup>e</sup>, J. Trap<sup>b\*</sup>

6

7 Affiliations

8 <sup>a</sup>Laboratoire des Radio-Isotopes, University of Antananarivo, 101 Antananarivo, Madagascar

9 <sup>b</sup>Eco&Sols, Univ Montpellier, CIRAD, INRAe, IRD, Institut Agro, Montpellier, France

10 <sup>c</sup>CIRAD, UMR AGAP, SRR FOFIFA, BP 230, 110 Antsirabe, Madagascar

11 <sup>d</sup>CIRAD, UMR AGAP, TA A-108 / 01 - Avenue Agropolis - 34398 Montpellier Cedex 5

12 France

13 <sup>e</sup>SRR FOFIFA, BP 230, 110 Antsirabe, Madagascar

14

15 \*Corresponding author: [jean.trap@ird.fr](mailto:jean.trap@ird.fr)

16

17 **Abstract**

18 Agroecology is based on the optimization of ecological processes occurring in agrosystems,  
19 especially by improving beneficial interactions among organisms. Recent studies have  
20 provided evidence that the ability of plants to interact with free-living soil organisms can be  
21 influenced by plant breeding. The upland rice breeding programme in the Highlands of  
22 Madagascar aims to develop cultivars that are adapted to local climatic, soil fertility and pest  
23 constraints. However, it is still unknown whether this genetic improvement alters rice cultivar  
24 abilities to interact with free-living soil plant-mutualistic organisms such as earthworms. We  
25 assessed in mesocosms the response of six upland rice cultivars commonly used by farmers in  
26 the Highlands of Madagascar to the presence of the endogeic earthworm *Pontoscolex*  
27 *corethrurus*. Plant nutrition and growth-related traits were measured after eight weeks of  
28 growth. These traits were significantly improved in the presence of *P. corethrurus* for all  
29 cultivars compared to the control treatment. However, the magnitudes of earthworm effects  
30 on plant traits were strongly variable and were dependent on specific rice cultivars.  
31 Agronomic and phylogenetic distance matrices were computed using agronomic data and  
32 available phylogenetic data of the rice cultivars. We did not detect significant correlations  
33 between cultivar responses to earthworm inoculation and agronomic or phylogenetic  
34 distances. Our results suggest that (i) the ability of rice to exploit beneficial interactions  
35 involving free-living soil organisms is influenced by its genetic background, but (ii) the loss  
36 of earthworm-interactive abilities of rice crops is independent of the genetic distance among  
37 cultivars and breeders' agronomic criteria. Our findings are significant for attempts to use rice  
38 cultivars that optimize plant-soil interactions within agroecological practices and highlight the  
39 need to integrate free-living soil partners as life-long plant partners in future strategies for  
40 plant breeding.

41

42 Keywords: Agroecology, plant-mutualistic free-living organisms, plant-soil interactions,  
43 breeding programme

44

45

## 46 **1. Introduction**

47 Plant breeding programmes mainly focus on crop adaptation to environmental and biotic  
48 stresses to increase yields. In the Highlands of Madagascar, breeding was used to develop  
49 high yielding and adapted upland rice varieties due to (i) cold climate at the high altitudes  
50 (Raboin et al., 2014), (ii) poor mineral fertility of Ferralsols (Raminoarison et al., 2019), and  
51 (iii) blast disease caused by the ascomycete fungus *Pyricularia oryzae* Cavara (Dusserre et al.,  
52 2017). However, because wild plants have evolved over time with specific soil biota as their  
53 plant-mutualistic partners, plant breeding is also a practice that may affect interactions  
54 between cultivated plants and soil organisms (Barot et al., 2010; Litrico and Violle, 2015).  
55 Indeed, by developing genetically homogenous crops with higher yields, resistance to pests  
56 and higher tolerance to climatic stresses, plant breeding may have considerable effects on  
57 plant-soil interactions (Bakker et al., 2012; Mariotte et al., 2018). For instance, the response  
58 of crop plants to soil life is known to vary with the presence of bacteria (Briones et al., 2002;  
59 Hardoim et al., 2011), mycorrhizal fungi (Declerck et al., 1995; Hetrick et al., 1996; Vallino  
60 et al., 2009), protists (Somasundaram et al., 2008), or earthworms (Barot et al., 2010; Junaidi  
61 et al., 2018; Noguera et al., 2011). To date, plant breeding strategies with mutualistic free-  
62 living soil organisms as direct selection targets are poorly developed (Gopal and Gupta, 2016;  
63 Wei and Jousset, 2017). Knowing that soil organisms can significantly promote plant  
64 functions, this lack of consideration is seen as a significant failure (Altieri, 1999; Barrios,  
65 2007).

66 Earthworms present a worldwide-distributed soil biota (Phillips et al., 2019) and are involved  
67 in key soil functions (Lavelle, 1997; Lavelle et al., 2006; Jouquet et al., 2006, 2008). They are  
68 known to increase plant growth via different mechanisms (Brown et al., 2004; Scheu, 2003):  
69 (i) increased mineralization of soil organic matter (Bernard et al., 2012; Coq et al., 2007); (ii)  
70 production of plant growth substances via stimulation of microbial activity (Blouin, 2018);

71 (iii) control of pests and parasites (Blanchart et al., 2019; Loranger-Merciris et al., 2012); (iv)  
72 stimulation of symbionts (Huhta, 2006) and (v) modifications of soil porosity, aggregation  
73 and water retention (Blanchart et al., 1999). Breeding modern high-yielding cultivars takes  
74 place under high input conditions (i.e., mineral fertilizers and chemical treatment of pests) in  
75 soils that can be unfavourable to earthworms (Doran and Zeiss, 2000; Ponge et al., 2013).  
76 Therefore, it is possible that these cultivars have lost some of the plant functional traits  
77 involved in interactions with earthworms, which may result in contrasting responses of plants  
78 to earthworm presence. For instance, Noguera et al. (2011) showed that the effects of  
79 *Pontoscolex corethrurus* on rice growth varied significantly for different cultivars and to the  
80 presence of biochar. However, the factors e.g., genetic background, agronomic selection  
81 criteria and targeted plant traits that determine this variability in plant responses to  
82 earthworms remain unidentified.

83 In the Ferralsols of Madagascar, upland crops face several edaphic constraints. Ferralsols are  
84 very infertile with numerous limiting nutrients, especially P but also N, Ca, Mg  
85 (Raminoarison et al., 2020). They have very low carbon contents and a low biological activity  
86 leading to a very poor productivity of the native vegetation. In the Highlands, the main  
87 earthworm species (i.e. *P. corethrurus* (Rhinodrilidae), *Dichogaster bolau*, *Dichogaster*  
88 *saliens* (Acanthodrilidae), and *Amyntas corticis* (Megascolecidae)) can deeply modify soil  
89 structure and soil C and N cycles and P availability (Trap et al., 2021). This last function is  
90 especially important for P-fixing soils such as the Ferralsols.

91 In the present study, by using a mesocosm greenhouse experimental approach, we aimed to  
92 assess the effects of earthworms on the growth and nutrition of six locally widespread upland  
93 rice cultivars that are adapted to the nutrient-depleted Ferralsols in Madagascar. We also  
94 tested whether agronomic selection criteria or the genetic signals among cultivars may explain  
95 the various responses of rice cultivars to earthworm inoculation. For this purpose, the

96 relationships between the response of rice cultivars to earthworm inoculation and agronomic  
97 or phylogenetic dissimilarity among the cultivars were assessed.

98

## 99 **2. Materials and methods**

### 100 ***2.1 Soil sampling***

101 The upper 10 cm layer of a Ferralsol (FAO classification) in a natural grassland was collected  
102 from the experimental station of the “Laboratoire des Radio-Isotopes” (LRI) in Lazaina  
103 (18°46'55 59° S, 47°32'46 3°N, 1,274 m altitude, Madagascar) in October 2016 from a 2 m<sup>2</sup>  
104 area. In the laboratory, the soil sample was dried at room temperature for 7 days and was then  
105 mixed using a shovel and then by hand to homogenize the sample. Most of the roots and  
106 vegetation debris were removed. The soil was then sieved to 2 mm and stored at room  
107 temperature in the dark for further analysis of soil properties.

108

### 109 ***2.2 Soil analyses***

110 Total soil organic carbon and nitrogen were measured by dry combustion in a Flash 2000  
111 CHNS analyser (Flash 2000 Series, CHNS/O 122 Analyser Thermo Scientific, IRCOF,  
112 France). The Olsen method (Olsen 1954) was used to determine the plant-available P content.  
113 Total P was evaluated by digestion (4 h at 600°C) with perchloric acid (HClO<sub>4</sub>) for extraction  
114 and by using a spectrophotometer and the colorimetry method (molybdenum blue) to analyse  
115 dosage. Cation exchange capacity (CEC), exchangeable K, exchangeable Mg, and  
116 exchangeable Ca were analysed using cobaltihexamine extraction and measured using an  
117 atomic absorption spectrophotometer (iCE 3000 Series AA spectrometer, Belgium). The  
118 exchangeable Al ions were extracted with a 1 N KCl solution and assayed by titrimetry with  
119 NaOH. The soil pH in water and in 1 M KCl were measured using a glass electrode pH metre  
120 (pH 211 microprocessor pH metre, Hanna Instruments, Romania) with a volume ratio of

121 1:2.5, and particle sizes were determined with the hydrometer method (Okalebo et al., 2002).  
122 The soil had a sandy-clay texture, pH (H<sub>2</sub>O) of 5.5, total organic C content of 20.8 g kg<sup>-1</sup>,  
123 total N-content of 1.3 g kg<sup>-1</sup>, C:N ratio of 16, total P content of 380 mg kg<sup>-1</sup>, Olsen P content  
124 of 7.12 mg kg<sup>-1</sup>, and exchangeable K, Ca and Mg contents of 30.7, 120.7 and 28.3 mg kg<sup>-1</sup>,  
125 respectively. The CEC was 1.34 cmol<sup>+</sup> kg<sup>-1</sup>, and the water-holding capacity was 0.49 g g<sup>-1</sup>  
126 dry soil.

127

### 128 **2.3. Biological materials**

129 Individuals of *P. corethrurus* (Rhinodrilidae) were used in the experiment. This endogeic,  
130 geophagous, peregrine species is dominant in all pedo-climatic regions in Madagascar  
131 (Razafindrakoto et al., 2010). It is a 5-8cm in length, weighing 0.7 g in average at the adult  
132 stage. This earthworm is able to live in an exceptionally substantial range of soil and climate  
133 environments. *P. corethrurus* individuals were collected near the experimental station in  
134 Lazaina and were stored in the same soil before use. Six upland rice (*Oryza sativa* L.)  
135 cultivars commonly used by farmers in the Highlands were studied: “B22”, “FOFIFA-161”,  
136 “FOFIFA-172”, “FOFIFA-182”, “NERICA-4” and “WAB 880-1-32-1” (Supplementary  
137 Table S1). Seeds were provided by the Madagascar National Centre for Applied Research on  
138 Rural Development (FOFIFA) and the French Agricultural Research Institute for  
139 Development (CIRAD)

140 All cultivars are fully described in the official catalogue of rainfed rice cultivars in  
141 Madagascar (Raboin et al., 2013; Radanielina et al., 2013). These rice cultivars cover the  
142 narrow current genetic diversity panel that can be found in the Highlands of Madagascar. To  
143 test the relationship between the ability of these cultivars to respond to earthworms and their  
144 agronomic characteristics, we collected agronomic data for each cultivar from the official  
145 catalogue. These agronomic data (e.g., optimal altitude for cropping, maximal grain yield,



146 tolerance to the root parasite *Striga asiatica* or the disease *Pyricularia oryzae*, cycle duration,  
147 grain weight, length of panicle and cold tolerance) of each cultivar described in the catalogue  
148 are the result of several years of field research on the genetic improvement of rainfed rice in  
149 the Highlands of Madagascar as part of the plant genetic programme led by CIRAD and  
150 FOFIFA (Supplementary Table S1) (Raboin et al., 2012; Raboin et al., 2013; Raboin et al.,  
151 2014; Raboin et al., 2010; Radanielina et al., 2013). The rice agronomic data from the  
152 programme were averaged over 6 years of upland rice cropping under high mineral  
153 fertilization levels (e.g., 150 kg ha<sup>-1</sup> of N:P:K = 11:22:16 and 80 kg ha<sup>-1</sup> of urea) and organic  
154 fertilization levels (5 t ha<sup>-1</sup> of cattle manure).

155

#### 156 **2.4. Experimental design**

157 The experiment was conducted at the Radio-Isotopes Laboratory (LRI) of the University of  
158 Antananarivo (Madagascar). Two treatments were considered: (i) an earthworm treatment  
159 including two modalities, i.e., without earthworms (-EW) and with earthworms (+EW) and  
160 (ii) a rice cultivar treatment that included six modalities. Each treatment was replicated five  
161 times.

162

#### 163 **2.5. Mesocosm setup**

164 The mesocosms consisted of 1 L plastic pots that were previously pierced (2 holes) at the  
165 bottom to avoid water accumulation. Each hole was filled with a cotton mesh to facilitate  
166 water flow. To prevent earthworms from escaping from the mesocosm, a mosquito net was  
167 placed at the bottom of the mesocosms. A total of 60 mesocosms (2 earthworm modalities × 6  
168 rice cultivars × 5 replicates) were filled with 1 kg of sieved dry soil. At the beginning of the  
169 experiment, mesocosms were watered with 350 ml of rainwater to mimic field conditions.  
170 Three seeds of each rice cultivar were sown. Three adults of *P. corethrurus* with a well-

171 developed clitellum and similar fresh mass (mean fresh weight of adults ~0.8 g) were  
172 introduced into each mesocosm. This earthworm density (~150 individuals per m<sup>2</sup>)  
173 corresponds to field density (Rakotomanga et al., 2016). During the experiment, the  
174 mesocosms were placed outside under natural weather conditions. The soil moisture content  
175 was maintained at 80% water holding capacity by weekly regular weighing. The mesocosm  
176 placements were changed every two weeks and used a new randomization. The experiment  
177 lasted 8 weeks.

178

## 179 ***2.6. Measurements of earthworm survival rates and rice traits***

180 At the end of the experiment, the soil was gently removed from the mesocosms to check for  
181 earthworm presence. All earthworm individuals (adults, juveniles, cocoons) were collected by  
182 hand sorting. We calculated the earthworm survival rate based on abundance, expressed as the  
183 initial percentage. To evaluate the photosynthetic activity of rice leaves, the photochemical  
184 reflectance index (PRI) and normalized difference vegetation index (NDVI) of each rice  
185 cultivar were measured using PlantPen PRI 200 & NDVI 300 (Gamon et al., 2015). Five  
186 measurements were randomly performed on the largest leaf in each mesocosm every week  
187 and at the end of the experiment. Only the maximum values among the five were considered  
188 for data analysis. We also scanned the same largest leaf at the end of the experiment at 600  
189 ppi to obtain the leaf area using the “Easy Leaf Area” software (Department of Plant  
190 Sciences, University of California). The final leaf dry matter content (LDMC) was determined  
191 after drying the leaves at 65°C for 48 h, and the specific leaf area (SLA) was calculated as the  
192 ratio of leaf area to leaf dry mass. Roots were separated from the soil by carefully washing the  
193 root system under flowing water to remove soil particles. The shoots and roots were dried at  
194 65°C for 48 h and weighed. Total N contents in shoots and roots were measured by a  
195 CHNS/O elemental analyser (Flash 2000 Series, CHNS/O 122 Analysers Thermo Scientific,

196 IRCOF, France). P contents in shoots and roots were determined after digestion of plant  
197 tissues with nitric acid 65%. P was measured in the digested solutions by colorimetry. The  
198 totals of N and P amounts in rice tissues were calculated by multiplying the N and P contents  
199 by biomass. N and P uptake efficiencies (NUE and PUE) were calculated as the ratio of P  
200 amounts in shoots by root biomass.

201

## 202 **2.7. Statistical analyses**

### 203 *2.7.1. Ecological distance*

204 To obtain a statistical distance among cultivars that can be used as the response to earthworm  
205 inoculation, we used the Cohen's d effect size (Cohen, 1988):  $d = (M_{i+EW} - M_{i-EW}) / (SD_{i-pooled})$  for the "i" cultivar, where  $M_{+EW}$  is the mean of the rice trait for the +EW  
206 treatment,  $M_{-EW}$  is the mean of the rice trait for the -EW treatment and  $SD_{pooled}$  is the  
207 pooled standard deviation of the rice trait. The Cohen's d effect size was thus calculated for  
208 each rice trait and for each cultivar using the five replications to determine the means and  
209 standard deviations (n= 5). We then computed the ecological distance using Cohens' d-values  
210 between all pairs of rice cultivars based on the Euclidean distance using the "dist" function  
211 from the R package "stats" (Supplementary Table S2).

212

### 213 *2.7.2. Phylogenetic distances*

214 We assessed the phylogenetic distance among all pairs from the six cultivars (Supplementary  
215 Table S2). The molecular information is stored and managed through a web-based tool,  
216 Gigwa-Genotype investigator, for genome-wide analyses (Sempéré et al., 2019). In all cases,  
217 the complete sequence of the Nipponbare rice cultivar (Os-Nipponbare-Reference-IRGSP-  
218 1.0) was used as a reference for alignments. The Tassel GBS pipeline v3.0 (Glaubitz et al.,  
219 2014) allowed us to filter the raw data. Data were filtered for minor allele frequencies (MAF  
220

221  $\geq 5\%$ ) and missing rates ( $< 20\%$ ). A total of 36,744 Single Nucleotide Polymorphisms (SNPs)  
222 were obtained. TASSEL software version 5.0 (Bradbury et al., 2007) was used to estimate the  
223 genetic distance matrix. These analyses considered only 7,338 non-missing loci.

224

### 225 2.7.3. Agronomic distances

226 We computed the agronomic distance matrix (Supplementary Table S2) by using data  
227 collected in previous plant genetic programmes performed in Madagascar (Supplementary  
228 Table S1). We used the following agronomic criteria: optimal altitude ranges for cropping  
229 (m), maximal grain yield in fertile field situations ( $\text{t ha}^{-1}$ ), tolerance to the root parasite *Striga*  
230 *asiatica*, cycle duration (days), grain weight (g), panicle length (cm) and cold tolerance from  
231 spikelet fertility (%). All data were collected from the cultivar catalogue that was created by  
232 the rice genetic programme of CIRAD and FOFIFA. The data were available in published  
233 studies (Raboin et al., 2013; Raboin et al., 2014; Raboin et al., 2010) or from unpublished  
234 technical sources but with free access on the internet ([www.dp-](http://www.dp-spada.org/productions2/documents-techniques)  
235 [spada.org/productions2/documents-techniques](http://www.dp-spada.org/productions2/documents-techniques)). The Euclidean distances for all cultivar pairs  
236 were calculated with the “dist” function from the R package “stats”. We did not weight the  
237 different agronomic parameters.

238

### 239 2.7.4. Data analyses

240 Means and standard deviations ( $n=5$ ) for each variable were computed for each treatment. The  
241 normality of the data was checked using the Shapiro-Wilk test. Data were transformed with  
242 the Box-Cox transformation when required. Two-way analyses of variance (ANOVA) were  
243 then conducted with the two main factors and their interactions: earthworm inoculation  
244 (“EW”) including two modalities (absent or present) and rice cultivars covering six  
245 modalities. The significance threshold was set to 5%. We also computed the eta-squared ( $\eta^2$ )

246 from ANOVA as a measure of effect size (% of variance explained) (Cohen, 1973). We  
247 detected the significant differences among the cultivars using the Tukey HSD post hoc test.  
248 The agronomic, phylogenetic and ecological distances among all pairs of cultivars were  
249 transformed to fit between 0.1 and 1.0 using a homothetic transformation;  $Y = 0.1 + ((x - b)/(a$   
250  $- b)) \times 0.9$ , where Y is the value of a variable after transformation, x is the distance to  
251 transform, a is the maximum distance and b is the minimum distance among the cultivars.  
252 Then, the linear relationships between the ecological, phylogenetic and agronomic distances  
253 among cultivars were tested using parametric Pearson's product-moment correlation  
254 coefficients at the 5% significance threshold.

255

### 256 **3. Results**

#### 257 ***3.1. Earthworm survival rate***

258 At the end of the experiment, the earthworm survival rate ranged on average from 93% to  
259 107% for all rice cultivar treatments (Supplementary Fig. S1). We found no significant  
260 differences ( $P > 0.05$ ) in the earthworm survival rates among rice cultivars.

261

#### 262 ***3.2. Effect of cultivars on rice traits***

263 After eight weeks of growth, all leaf traits, except for NDVI, were significantly different  
264 among rice cultivars (Table 1). On average, F161 had the highest leaf area, LDMC and SLA,  
265 regardless of earthworm treatments (Table 2). A significant effect for rice cultivars was found  
266 for shoot biomass and shoot P and N amounts (Table 1). The highest values for shoot biomass  
267 and shoot P and N amounts were found for cultivars B22 and F161, respectively (Table 2).  
268 The lowest values were observed for F172 and N4.

269

#### 270 ***3.3. Effect of earthworms on rice traits***

271 The inoculation of *P. corethrurus* (+EW) had a strong significant effect on rice traits, except  
272 for LDMC, SLA, shoot:root ratio and PUE (Table 1). On average, rice cultivars had higher  
273 PRI (+57%), NDVI (+34%), shoot biomass (+99%), root biomass (+93%), total biomass  
274 (+96%), shoot P amount (+87%), root P amount (82%), total rice P amount (+84%) and shoot  
275 N amount (+267%) in the presence of earthworms than in their absence (Table 2).

276

### 277 **3.4. Interaction effects of earthworms and cultivars on rice traits**

278 We found significant interactive effects of earthworms and rice cultivars on root biomass,  
279 shoot:root ratio, and N and P acquisition efficiency (Table 1). Considering the  $\eta^2$  of the  
280 different sources of variation from the ANOVA models, we identified eight rice traits that  
281 were mostly affected by earthworms ( $\eta^2$  ranged from 49% to 68%), including physiological  
282 traits (e.g., SLA, PRI, and NDVI), growth traits (e.g., shoot biomass and total biomass) and  
283 nutrition traits (e.g., shoot P amount, total rice P amount and shoot N amount). For instance,  
284 in the presence of earthworms, SLA increased by 146% and 126% for F161 and F172,  
285 respectively, whereas it increased only by 28% for W880 (Table 2) (Fig. 1). Likewise, the  
286 positive effect of *P. corethrurus* on rice root biomass exhibited a 323% increase for N4 while  
287 this effect exhibited only a 19% increase for B22. A similar trend was observed for root P  
288 levels (Table 2). Regarding the shoot:root ratio, the effect of earthworms was highly variable  
289 and depended on specific rice cultivars (Fig. 1). It was either positive (B22, F161, and F182),  
290 neutral (F172) or negative (N4 and W880). Similarly, the response of rice cultivars to  
291 earthworms in terms of P acquisition efficiency was positive (B22, F161, and F172) or  
292 negative (F182, N4, and W880). It is interesting to note that, even if not significant according  
293 to two-way ANOVA, the magnitude of the increase induced by earthworms on shoot biomass,  
294 shoot P and shoot N amount also varied depending on rice cultivar (Fig. 1). For instance, the  
295 increase in shoot biomass induced by earthworms ranged from 40% (W880) to 129% (B22)

296 according to the cultivars being examined. Likewise, the increase in shoot P amount in the  
297 presence of earthworms ranged from 37% (W880) to 145% (F161), and the increase in shoot  
298 N amount varied from 116% (W880) to 355% (B22) in the presence of earthworms.  
299 Rice cultivars in the present experiment can be classified into three categories (Fig. 1): (i) rice  
300 cultivars with high average Cohen's d-values (response to earthworms), such as N4 and F182,  
301 with average d-values of 0.95 and 0.64, respectively; (ii) rice cultivars with intermediate  
302 responses, such as F172, B22, and F161, with average d-values of 0.52, 0.43 and 0.32,  
303 respectively; and (iii) rice cultivars with low average d-values, such as W880 ( $d = 0.20$ ).

304

### 305 ***3.5. Phylogenetic, agronomic and ecological relationships***

306 We did not find significant linear relationships for the ecological distances among rice  
307 cultivars (i.e., their ability to respond to earthworm inoculation), agronomic ( $R^2 = 0.15$ , P-  
308 value 0.144), and phylogenetic ( $R^2 = 0.10$ , P-value 0.242) distances (Fig. 2). However, the  
309 agronomic and phylogenetic distances were significantly correlated ( $R^2 = 0.56$ , P-value  
310 0.001).

311

## 312 **4. Discussion**

### 313 ***4.1. Earthworms as plant promoters irrespective of rice cultivars***

314 The earthworm survival rate, production of cocoons (1 to 4 cocoons per mesocosm) and  
315 surface casts in the mesocosms indicated that the conditions were favourable for the  
316 development and activity of earthworms during the experiment. Under these conditions, we  
317 demonstrated that the earthworm *P. corethrurus* strongly increased plant growth and nutrition  
318 regardless of rice cultivar. Increased rice growth was expressed by higher shoot biomass (99%  
319 increase), root biomass (93% increase) and total biomass (96% increase). We also found  
320 positive earthworm effects on leaf physiological traits, which indicated that earthworms

321 enhanced photosynthetic light use efficiency and chlorophyll content in plants. The only plant  
322 traits that exhibited negative Cohen's d values in some cultivars were the leaf dry matter  
323 content, specific leaf area and P acquisition efficiency. Brown et al. (1999; 2004) reviewed  
324 246 experiments conducted in tropical countries and concluded that in 43% of these studies,  
325 earthworms enhanced biomass production by more than 20%. The same observation was  
326 noted by van Groenigen et al. (2014) in a meta-analysis of 58 studies published within one  
327 century (1910–2013). These authors pointed out that increased N mineralization was the main  
328 pathway for the positive effects of earthworms on plant growth. Likewise, in the present  
329 study, earthworm inoculation strongly increased shoot N amounts (+267%), and this plant  
330 trait had the highest percentage of variance explained by earthworms (79%).

331 A multiple nutrient-omission experiment that was conducted with the same soil showed that P  
332 was the main nutrient which limited rice growth (Raminoarison et al., 2020). It is thus more  
333 likely that the positive effects of earthworms on rice growth are mainly driven by improved P  
334 availability (Chapuis-Lardy et al., 1998; Ros et al., 2017; Vos et al., 2019). This hypothesis is  
335 supported by higher shoot P (87% increase) when *P. corethrurus* was inoculated. We also  
336 observed significant effects of earthworms on plant P acquisition efficiency. In our  
337 experiment, a large number of casts were observed (not quantified) at the end of the  
338 experiment, which support the hypothesis formulated by Vos et al., (2014) and suggest that  
339 the increased plant P uptake was probably induced by the increase in available P in earthworm  
340 casts. The positive effect of earthworms on plant growth via increased soil P availability has  
341 been reported many times (Graff, 1970; Kuczak et al., 2006; Ros et al., 2017; Sharpley and  
342 Syers, 1976; Sharpley and Syers, 1977; Van Groenigen et al., 2019; Vos et al., 2019; Vos et  
343 al., 2014), including in Ferralsols (Chapuis-Lardy et al., 1998; Chapuis-Lardy et al., 2009).

344

345 ***4.2. Plant breeding and rice ability to interact with earthworms***



346 Despite the overall positive effects of earthworms on plants, the ability of rice to interact with  
347 earthworms varied by cultivar. These findings are in agreement with Noguera et al. (2011),  
348 who conducted a mesocosm experiment in Colombia with the same earthworm species but in  
349 a nutrient-rich soil and with other rice cultivars.

350 From our data, it is difficult to identify the mechanisms involved in the response variability of  
351 rice cultivars to earthworms. For instance, the P acquisition efficiency (PUE) was the plant  
352 trait for which the percentage of variance explained by earthworm-rice cultivar interaction  
353 was greatest (29%). The role of P, as mentioned earlier, thus appears to be central in  
354 earthworm-rice interactions in nutrient-depleted tropical soils (Coulis et al., 2014; Le Bayon  
355 and Milleret, 2009). It would then be judicious to examine this trait. Vos et al. (2019) listed  
356 the potential pathways for earthworm-induced enhanced soil P availability: (i) elevated pH  
357 (higher pH in casts) that affects concentrations of P ions in solution, (ii) dissolved organic  
358 carbon production by worms with similar effects through competitive adsorption, (iii)  
359 stimulated mineralization of organic P (enzymes) and (iv) selective feeding. In our study,  
360 given the equal survival rates of earthworms for the various treatments, it is very unlikely that  
361 the differences observed in the plant response to earthworms could arise from these  
362 mechanisms, which are specific to worms. In contrast, we believe that these differences could  
363 come from root trait variability, which is known to be affected by earthworms (Arnone III and  
364 Zaller, 2014; Junaidi et al., 2018). We can thus suppose that the colonization of casts by roots  
365 and their fungal endomycorrhizal symbionts could affect plant nutrition (Milleret et al., 2009;  
366 Van Groenigen et al., 2014; Zaller et al., 2013), especially plant P uptake (Bolan, 1991;  
367 George et al., 1995; Smith et al., 2011). These root traits could vary according to rice  
368 cultivars. Knowing the crucial roles of these parameters in plant P acquisition (Bolan, 1991;  
369 George et al., 1995) and their interaction with earthworms (Milleret et al., 2009), it would be  
370 interesting to further investigate these root traits, e.g., root surface, length, number of tips or

371 mycorrhizal fungal colonization, in further studies to identify the mechanisms involved in the  
372 variability in earthworm-plant interactions with plant cultivars. It would also be interesting to  
373 monitor the P nutrition of rice during its growth, in the presence or absence of earthworms,  
374 for a reduced number of contrasting varieties. This could allow us to test whether the response  
375 of varieties to the presence of rice takes place at key phenological periods.

376 Considering the agronomic characteristics of rice cultivars (Table 2), we did not find evidence  
377 of relationships between some agronomic criteria and rice response to earthworms.

378 Accordingly, we did not observe any significant relationship between the agronomic distances  
379 among cultivars and their ability to interact with inoculated earthworms. These results suggest  
380 that, regardless of the agronomic criteria focused on in the breeding programme, plant  
381 breeding may affect the ability of rice crops to exploit the beneficial functions achieved by  
382 earthworms. As plant breeding affects the genetic inheritance of rice cultivars with contrasting  
383 NUE or PUE, the cultivars can therefore exploit the presence of earthworms differently.

384 However, before taking this for granted, it would be beneficial to test this hypothesis with a  
385 larger panel of rice cultivars.

386

#### 387 ***4.3. Ecological implications for agroecology and limits of the study***

388 This experimental study showed that the upland rice cultivars commonly used by farmers in  
389 the Highlands of Madagascar have varying abilities to interact with *P. corethrurus*. This result  
390 is noteworthy because the cross effects of earthworms and rice cultivars were highly  
391 significant despite the relatively short duration of the study. To confirm these patterns and to  
392 measure rice yield, it would be beneficial to continue with an earthworm inoculation  
393 experiment in the field. Also, the low genetic diversity within our narrow cultivar panel could  
394 also be a factor that limited our conclusions. However, the criteria for choosing rice cultivars  
395 in this study were constrained by the genetic pool that was adapted to the environmental and

396 agronomic conditions (climate, soil, uses and producer preferences) of the Highlands of  
397 Madagascar (Raboin et al., 2014). This genetic diversity is low and confers significant risks  
398 for the production of rainfed rice, particularly against pests. In view of the major role of  
399 biological interactions between soil organisms and rice for the tolerance of rice to pests and  
400 diseases (Blanchart et al., 2019), considering the capability of varieties to interact with  
401 earthworms seems to be essential. In these conditions, however, we succeeded in identifying  
402 the rice cultivars that exhibited strong (N4 and F182), intermediate (F172, B22, and F161)  
403 and low (W880) abilities for interacting with earthworms. Integrating the most appropriate  
404 rice cultivars into relevant agroecological practices has been a concern of several soil  
405 ecologists and agronomists (Loeuille et al., 2013; Noguera et al., 2011). We therefore propose  
406 to consider the plant genetic material that best interacts with earthworms to intensify soil  
407 ecological processes in low-input tropical agrosystems, either for direct use with relevant  
408 agro-ecological practices or for inclusion in plant breeding programs. Such innovative  
409 practices may be a suitable approach to meet the constraints of the low input production  
410 systems of smallholder farmers in the Highlands of Madagascar.

411

## 412 **Acknowledgements**

413 We want to thank Eric Randrianasolo, Andrianantenaina Hilaire Damase Razafimahafaly and  
414 Marie-Paule Razafimanantsoa (Laboratoire des RadioIsotopes) for their technical assistance.  
415 This study was funded by the Agropolis Foundation (Investissements d’Avenir Programme,  
416 ANR-10-LABX-0001-01) under the framework of the SECuRE (ID 1605-007) project. This  
417 work was also supported by the French Ministry Europe and Foreign Affairs under the  
418 programme ‘Make Our Planet Great Again’ for postdoctoral researchers.

419

420

421 **References**

422

423 Altieri, M.A., 1999. The ecological role of biodiversity in agroecosystems. *Agric. Ecosyst.*

424 *Environ.* 74, 19-31.

425

426 Arnone III, J.A., Zaller, J.G., 2014. Earthworm effects on native grassland root system

427 dynamics under natural and increased rainfall. *Front. Plant. Sci.* 5:152.

428

429 Bakker, M.G., Manter, D.K., Sheflin, A.M., Weir, T.L., Vivanco, J.M., 2012. Harnessing the

430 rhizosphere microbiome through plant breeding and agricultural management. *Plant. Soil.*

431 360, 1-13.

432

433 Barot, S., Noguera, D., Laossi, K., de Carvalho, M., 2010. Breeding rice cultivars to enhance

434 positive feedbacks between soil and crops? In: *Breeding for resilience: a strategy for organic*

435 *and low-input farming systems? EUCARPIA 2nd Conference of the Organic and Low-Input*

436 *Agriculture Section, Paris, France, 1-3 December 2010, Institut National de la Recherche*

437 *Agronomique*, pp. 55-58.

438

439 Barrios, E., 2007. Soil biota, ecosystem services and land productivity. *Ecol. Econ.* 64, 269-

440 285.

441

442 Bernard, L., Chapuis-Lardy, L., Razafimbelo, T., Razafindrakoto, M., Pablo, A.L., Legname,

443 E., Poulain, J., Bröls, T., O'Donohue, M., Brauman, A., Chotte, J.L., Blanchart, E., 2012.

444 Endogeic earthworms shape bacterial functional communities and affect organic matter

445 mineralization in a tropical soil. *ISME J.* 6, 213-222.

446

447 Blanchart, E., Albrecht, A., Alegre, J., Duboisset, A., Villenave, C., Pashani, B., Lavelle, P.,  
448 Brussaard, L., 1999. Effects of earthworms on soil structure and physical properties, in:  
449 Lavelle, P., Brussaard, L., Hendrix, P. (Eds.), *Earthworm Management in Tropical*  
450 *Agroecosystems*. CABI Publishing, pp. 149-172.

451

452 Blanchart, E., Ratsiatosika, O., Raveloson, H., Razafimbelo, T., Razafindrakoto, M., Sester,  
453 M., Becquer, T., Bernard, L., Trap, J., 2020. Nitrogen supply reduces the earthworm-silicon  
454 control on rice blast disease in a Ferralsol. *Appl. Soil. Ecol.* 145, 103341.

455

456 Blouin, M., 2018. Chemical communication: An evidence for co-evolution between plants  
457 and soil organisms. *Appl. Soil. Ecol.*, 123, 409-415.

458

459 Bolan, N., 1991. A critical review on the role of mycorrhizal fungi in the uptake of  
460 phosphorus by plants. *Plant. Soil.* 134, 189-207.

461

462 Bradbury, P.J., Zhang, Z., Kroon, D.E., Casstevens, T.M., Ramdoss, Y., Buckler, E.S., 2007.  
463 TASSEL: software for association mapping of complex traits in diverse samples.  
464 *Bioinformatics.* 23, 2633-2635.

465

466 Briones, A.M., Okabe, S., Umemiya, Y., Ramsing, N.B., Reichardt, W., Okuyama, H., 2002.  
467 Influence of different cultivars on populations of ammonia-oxidizing bacteria in the root  
468 environment of rice. *Appl. Environ. Microbiol.* 68, 3067-3075.

469

470 Brown, G.G., Edwards, C.A., Brussaard, L., 2004. How earthworms affect plant growth:  
471 burrowing into the mechanisms, in: Edwards, C.A. (Ed.), Earthworm ecology, 2nd edition,  
472 CRC Press, pp. 13-49.

473

474 Brown, G.G., Pashanasi, B., Villenave, C., Patron, J.C., Senapati, B.K., Giri, S., Barois, I.,  
475 Lavelle, P., Blanchart, E., Blakemore, R.J., Spain, A.V., Boyer, J. 1999. Effects of  
476 earthworms on plant production in the tropics, in: Lavelle P, Brussaard L, Hendrix P (Eds.)  
477 Earthworm Management in Tropical Agroecosystems. CABI Publishing, pp 87-147.

478

479 Chapuis-Lardy, L., Brossard, M., Lavelle, P., Schouller, E., 1998. Phosphorus transformations  
480 in a ferralsol through ingestion by *Pontoscolex corethrurus*, a geophagous earthworm. Eur. J.  
481 Soil. Biol., 34, 61-67.

482

483 Chapuis-Lardy, L., Ramiandrisoa, R., Randriamanantsoa, L., Morel, C., Rabeharisoa, L.,  
484 Blanchart, E., 2009. Modification of P availability by endogeic earthworms  
485 (*Glossoscolecidae*) in Ferralsols of the Malagasy Highlands. Biol. Fertil. Soils. 45, 415-422.

486

487 Cohen, J., 1973. Eta-squared and partial eta-squared in fixed factor ANOVA designs. Educ.  
488 Psychol. Meas. 33, 107-112.

489

490 Cohen, J., 1988. Statistical power analysis for the behavioral sciences. Lawrence Earlbaum  
491 Associates, Hillsdale, NJ, USA.

492

493 Coq, S., Barthes, B.G., Oliver, R., Rabary, B., Blanchart, E., 2007. Earthworm activity affects  
494 soil aggregation and organic matter dynamics according to the quality and localization of crop  
495 residues - An experimental study (Madagascar). *Soil. Biol. Biochem.* 39, 2119-2128.  
496

497 Coulis, M., Bernard, L., Gerard, F., Hinsinger, P., Plassard, C., Villeneuve, M., Blanchart, E.,  
498 2014. Endogeic earthworms modify soil phosphorus, plant growth and interactions in a  
499 legume–cereal intercrop. *Plant. Soil.* 379, 149-160.  
500

501 Declerck, S., Plenchette, C., Strullu, D., 1995. Mycorrhizal dependency of banana (*Musa*  
502 *acuminata*, AAA group) cultivar. *Plant. Soil.* 176, 183-187.  
503

504 Doran, J.W., Zeiss, M.R., 2000. Soil health and sustainability: managing the biotic component  
505 of soil quality. *Appl. Soil. Ecol.* 15, 3-11.  
506

507 Dusserre, J., Raveloson, H., Michellon, R., Gozé, E., Auzoux, S., Sester, M., 2017.  
508 Conservation agriculture cropping systems reduce blast disease in upland rice by affecting  
509 plant nitrogen nutrition. *Field. Crops. Res.* 204, 208-221.  
510

511 Gamon, J., Kovalchuck, O., Wong, C., Harris, A., Garrity, S., 2015. Monitoring seasonal and  
512 diurnal changes in photosynthetic pigments with automated PRI and NDVI sensors.  
513 *Biogeosciences.* 12, 4149-4159.  
514

515 George, E., Marschner, H., Jakobsen, I., 1995. Role of arbuscular mycorrhizal fungi in uptake  
516 of phosphorus and nitrogen from soil. *Crit. Rev. Biotechnol.* 15, 257-270.  
517

518 Glaubitz, J.C., Casstevens, T.M., Lu, F., Harriman, J., Elshire, R.J., Sun, Q., Buckler, E.S.,  
519 2014. TASSEL-GBS: a high capacity genotyping by sequencing analysis pipeline. PLoS.  
520 One. 9, 0090346.  
521  
522 Gopal, M., Gupta, A., 2016. Microbiome selection could spur next-generation plant breeding  
523 strategies. Front. Microbiol. 7, 1971.  
524  
525 Graff, O., 1970. The phosphorus content of earthworm casts. Landbauforschung. Volkenrode.  
526 20, 33-36.  
527  
528 Hardoim, P.R., Andreote, F.D., Reinhold-Hurek, B., Sessitsch, A., van Overbeek, L.S., van  
529 Elsas, J.D., 2011. Rice root-associated bacteria: insights into community structures across 10  
530 cultivars. FEMS. Microbiol. Ecol. 77, 154-164.  
531  
532 Hetrick, B., Wilson, G., Todd, T., 1996. Mycorrhizal response in wheat cultivars: relationship  
533 to phosphorus. Can. J. Bot, 74, 19-25.  
534  
535 Huhta, V., 2006. The role of soil fauna in ecosystems: A historical review. Pedobiologia. 50,  
536 489-495.  
537  
538 Junaidi, J., Kallenbach, C.M., Byrne, P.F., Fonte, S.J., 2018. Root traits and root biomass  
539 allocation impact how wheat genotypes respond to organic amendments and earthworms  
540 PLoS. One. doi.org/10.1371/journal.pone.0200646.  
541



542 Kuczak, C.N., Fernandes, E.C., Lehmann, J., Rondon, M.A., Luizao, F.J., 2006. Inorganic and  
543 organic phosphorus pools in earthworm casts (Glossoscolecidae) and a Brazilian rainforest  
544 Oxisol. *Soil. Biol. Biochem.* 38, 553-560.  
545  
546 Le Bayon, R.C., Milleret, R., 2009. Effects of earthworms on phosphorus dynamics—a review  
547 *Dynamic. Soil. Dynamic. Plant.* 3, 21-27.  
548  
549 Litrico, I., Violle, C., 2015. Diversity in plant breeding: a new conceptual framework. *Trends*  
550 *Plant. Sci.* 20, 604-613.  
551  
552 Loeuille, N., Barot, S., Georgelin, E., Kylafis, G., Lavigne, C., 2013. Eco-evolutionary  
553 dynamics of agricultural networks: implications for sustainable management, in: *Adv. Ecol.*  
554 *Res.* 49, 339-435.  
555  
556 Loranger-Merciris, G., Cabidoche, Y.M., Delone, B., Queneherve, P., Ozier-Lafontaine, H.  
557 2012. How earthworm activities affect banana plant response to nematodes parasitism? *Appl.*  
558 *Soil. Ecol.* 52, 1-8.  
559  
560 Mariotte, P., Mehrabi, Z., Bezemer, T.M., De Deyn, G.B., Kulmatiski, A., Drigo, B., Veen,  
561 G.F.C., van der Heijden, M.G.A., Kardol, P., 2018. Plant–soil feedback: bridging natural and  
562 agricultural sciences. *Trends. Ecol. Evol.* 33, 129-142.  
563  
564 Milleret, R., Le Bayon, R.C., Gobat, J.M., 2009. Root, mycorrhiza and earthworm  
565 interactions: their effects on soil structuring processes, plant and soil nutrient concentration  
566 and plant biomass. *Plant. Soil.* 316, 1-12.

567

568 Noguera, D., Laossi, K.R., Lavelle, P., De Carvalho, M.C., Asakawa, N., Botero, C., Barot,  
569 S., 2011. Amplifying the benefits of agroecology by using the right cultivars. *Ecol. Appl.* 21,  
570 2349-2356.

571

572 Okalebo, J.R., Gathua, K.W., Woomer, P.L., 2002. Laboratory methods of soil and plant  
573 analysis: a working manual, second edition. TSBF-CIAT and Sacred Africa, Nairobi.

574

575 Olsen, S.R., 1954. Estimation of available phosphorus in soils by extraction with sodium  
576 bicarbonate. United States Department of Agriculture, Washington, n°939.

577

578 Phillips, H.R. et al., 2019. Global distribution of earthworm diversity. *Science.* 366, 480-485.

579

580 Ponge, J.F, Peres, G., Guernion, M., Camacho, N.R., Cortet, J., Pernin, C., Villenave, C.,  
581 Chaussod, R., Martin-Laurent, F., Bispo, A., Cluzeau, D., 2013. The impact of agricultural  
582 practices on soil biota: A regional study. *Soil. Biol. Biochem.* 67, 271-284.

583

584 Raboin, L.M., Ramanantsoanirina, A., Dusserre, J., Razasolofonahary, F., Tharreau, D.,  
585 Lannou, C., Sester M.; 2012. Two-component cultivar mixtures reduce rice blast epidemics in  
586 an upland agrosystem. *Plant. Pathol.* 61, 1103-1111.

587

588 Raboin, L.M., Ramanantsoanirina, A., Dzido, J.L., Frouin, J., Radanielina, T., Tharreau D.,  
589 Dusserre, J., Ahmadi, N., 2013. Création variétale pour la riziculture pluviale d'altitude à  
590 Madagascar: bilan de 25 années de selection. *Cah. Agric.* 22, 450-458.

591

592 Raboin, L.M., Randriambololona, T., Radanielina, T., Ramanantsoanirina, A., Ahmadi, N.,  
593 Dusserre, J., 2014. Upland rice varieties for smallholder farming in the cold conditions in  
594 Madagascar's tropical highlands. *Field. Crops. Res.* 169, 11-20.  
595  
596 Raboin, L., Ramanantsoanirina, A., Dzido, J., Andriantsimialona, D., Tharreau, D.,  
597 Radanielina, T., Ahmadi, N., 2010. Upland (aerobic) rice breeding for the harsh environment  
598 of the High Plateau of Madagascar, in: *Innovation and Partnerships to Realize Africa's Rice*  
599 *Potential. Second Africa Rice Congress*, 26-27.  
600  
601 Radanielina, T., Ramanantsoanirina, A., Raboin, L.M., Frouin, J., Perrier, X., Brabant, P.,  
602 Ahmadi, N., 2013. The original features of rice (*Oryza sativa* L.) genetic diversity and the  
603 importance of within-variety diversity in the highlands of Madagascar build a strong case for  
604 in situ conservation. *Genet. Resour. Crop. Ev.* 60, 311-323.  
605  
606 Rakotomanga, D., Blanchart, E., Baraby, B., Randriamanantsoa, R., Razafindrakoto, M.,  
607 Autfray, P., 2016. Diversité de la macrofaune des sols cultivés sur les Hautes-Terres de  
608 Madagascar. *Biotechn. Agron. Soc.* 20, 495-507.  
609  
610 Raminoarison, M., Razafimbelo, T., Rakotoson, T., Becquer, T., Blanchart, E., Trap, J., 2020.  
611 Multiple-nutrient limitation of upland rainfed rice in ferralsols: a greenhouse nutrient-  
612 omission trial. *J. Plant. Nutr.* 43, 270-284.  
613  
614 Ranoarisoa, M. P., Blanchart, E., Vom Brocke, K., Ramanantsoanirina, A., Sester, M.,  
615 Plassard, C., Cournac, L., Trap, J., 2017. Attractancy of bacterivorous nematodes to root-  
616 adhering soils differs according to rice cultivars. *Rhizosphere.* 3, 128-131.

617

618 Razafindrakoto, M., Csuzdi, C., Rakotofiringa, S., Blanchart, E., 2010. New records of  
619 earthworms (Oligochaeta) from Madagascar. *Opusc. Zool. Budapest.* 41, 231-236.

620

621 Ros, M.B., Hiemstra, T., van Groenigen, J.W., Chareesri, A., Koopmans, G.F., 2017.,  
622 Exploring the pathways of earthworm-induced phosphorus availability. *Geoderma.* 3, 99-109.

623

624 Scheu, S., 2003. Effects of earthworms on plant growth: patterns and perspectives.  
625 *Pedobiologia.* 47, 846-856.

626

627 Sempéré, G., Pétel, A., Rouard, M., Frouin, J., Hueber, Y., De Bellis, F., Larmande, P., 2019.  
628 Gigwa v2—Extended and improved genotype investigator. *GigaScience.* 8.  
629 [doi.org/10.1093/gigascience/giz051](https://doi.org/10.1093/gigascience/giz051).

630

631 Sharpley, A., Syers J., 1976. Potential role of earthworm casts for the phosphorus enrichment  
632 of run-off waters. *Soil. Biol. Biochem.* 8, 341-346.

633

634 Sharpley, A., Syers, J., 1977. Seasonal variation in casting activity and in the amounts and  
635 release to solution of phosphorus forms in earthworm casts *Soil. Biol. Biochem.* 9, 227-231.

636

637 Smith, S.E., Jakobsen, I., Grønlund, M., Smith, F.A., 2011. Roles of arbuscular mycorrhizas  
638 in plant phosphorus nutrition: interactions between pathways of phosphorus uptake in  
639 arbuscular mycorrhizal roots have important implications for understanding and manipulating  
640 plant phosphorus acquisition. *Plant. Physiol.* 156, 1050-1057.

641

642 Somasundaram, S., Bonkowski, M., Iijima, M., 2008. Functional role of mucilage-border  
643 cells: a complex facilitating protozoan effects on plant growth. *Plant. Prod. Sci.* 11, 344-351.  
644

645 Trap, J., Blanchart, E., Ratsiatosika, O., Razafindrakoto, M., Becquer, T., Andriamananjara,  
646 A., Morel, C., 2021. Effects of the earthworm *Pontoscolex corethrurus* on rice P nutrition and  
647 plant-available soil P in a tropical Ferralsol. *Appl. Soil. Ecol.* 160, 103867.  
648

649 Vallino, M., Greppi, D., Novero, M., Bonfante, P., Lupotto, E., 2009. Rice root colonisation  
650 by mycorrhizal and endophytic fungi in aerobic soil. *Ann. Appl. Biol.* 154, 195-204.  
651

652 Van Groenigen, J.W., Lubbers, I.M., Vos, H.M., Brown, G.G., De Deyn, G.B., Van  
653 Groenigen, K.J., 2014. Earthworms increase plant production: a meta-analysis. *Sci. Rep.* 4,  
654 6365.  
655

656 Van Groenigen, J.W., Van Groenigen, K.J., Koopmans, G.F., Stokkermans, L., Vos, H.M.,  
657 Lubbers, I.M., 2019. How fertile are earthworm casts? A meta-analysis. *Geoderma.* 338, 525-  
658 535.  
659

660 Vos, H.M., Koopmans, G.F., Beezemer, L., de Goede, R.G., Hiemstra, T., van Groenigen,  
661 J.W., 2019. Large variations in readily-available phosphorus in casts of eight earthworm  
662 species are linked to cast properties. *Soil. Biol. Biochem.* 138, 107583.  
663

664 Vos, H.M., Ros, M.B., Koopmans, G.F., van Groenigen, J.W., 2014. Do earthworms affect  
665 phosphorus availability to grass? A pot experiment. *Soil. Biol. Biochem.* 79, 34-42.  
666

667 Wei, Z., Jousset A., 2017. Plant breeding goes microbial. *Trends. Plant. Sci.* 22, 555-558.

668

669 Zaller, J.G., Wechselberger, K.F., Gorfer, M., Hann, P., Frank, T., Wanek, W., Drapela, T.,

670 2013. Subsurface earthworm casts can be important soil microsites specifically influencing

671 the growth of grassland plants. *Biol. Fert. Soils.* 49, 1097-1107.

672

673

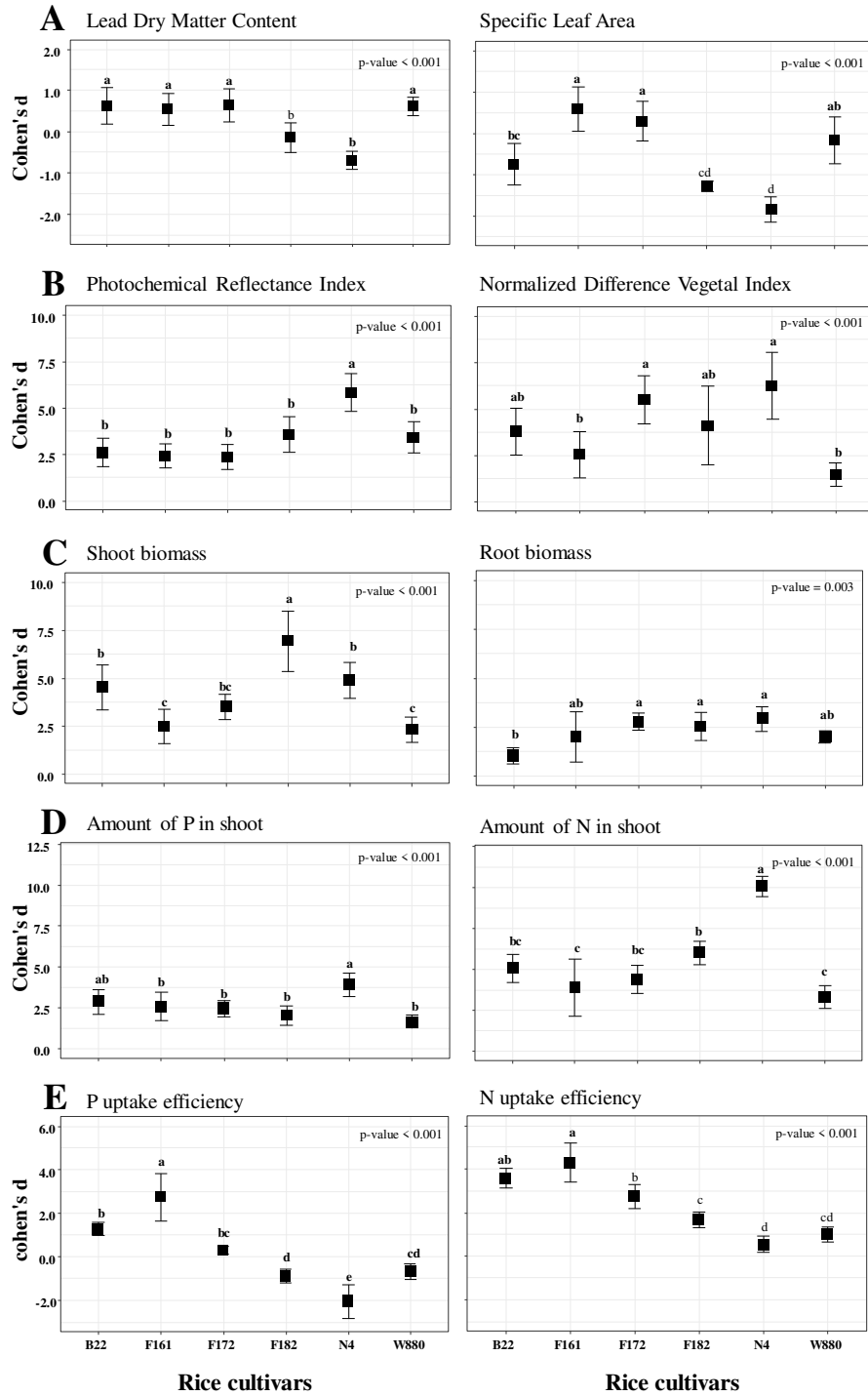
674 **Fig. 1.** Effect sizes (Cohen's d) of earthworms on different plant traits according to rice  
675 cultivars. (A) Leaf traits, (B) leaf physiological traits, (C) biomass traits, (D) nutrition traits,  
676 and (E) nutrient acquisition efficiencies. Whiskers correspond to standard deviation (SD).  
677 Different letters (a, b, c and d) indicate significant differences between rice cultivars  
678 according to Tukey HSD test ( $P$ -value < 0.05).

679

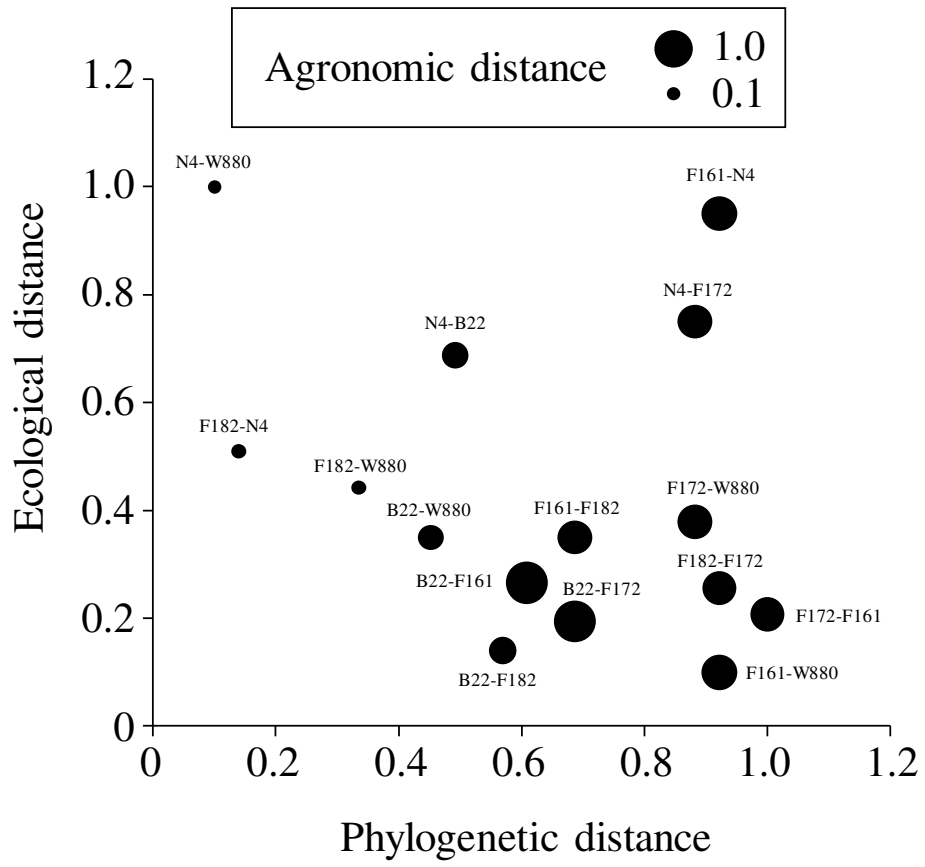
680 **Fig. 2.** Relationships between phylogenetic, agronomic and ecological distances. The  
681 phylogenetic distance among cultivars were computed using TASSEL software based on  
682 Single Nucleotide Polymorphisms. The Euclidean agronomic distance is based on seven  
683 unweighted agronomic criteria (Table S2). The ecological distance is the Euclidean distance  
684 computed from the response of cultivars to earthworm inoculation using Cohen's d effect size  
685 for each rice trait (Fig. 1). All distances were bounded between 0.1 and 1 using homothetic  
686 transformation. N4, W880, F161, B22, N172 and F182 refers to rice cultivars (Table S1).

687

688







**Table 1.** Results of two-way ANOVA (eta<sup>2</sup> in % and p-value) for the main effects (rice cultivars and earthworms) and their interactions for each plant variable.

	<b>Rice cultivars</b>		<b>Earthworms</b>		<b>Interaction</b>		<b>Residuals</b>
	Eta <sup>2</sup> <sup>s</sup>	p-value	Eta <sup>2</sup>	p-value	Eta <sup>2</sup>	p-value	Eta <sup>2</sup>
Leaf Dry Matter Content	34	***	1	ns	4	ns	61
Specific Leaf Area	20	*	1	ns	9	ns	71
Photochemical Reflectance Index	7	*	64	***	3	ns	25
Normalized Difference Vegetation Index	4	ns	68	***	5	ns	23
Shoot biomass	18	***	59	***	3	ns	20
Root biomass	3	ns	40	***	12	*	44
Shoot:root ratio	8	ns	2	ns	26	**	64
Shoot P amount	13	**	52	***	6	ns	30
Shoot N amount	7	***	79	***	2	ns	12
P acquisition efficiency	8	ns	0	ns	29	**	63
N acquisition efficiency	1	ns	33	***	17	*	49

<sup>s</sup>Eta-squared (eta<sup>2</sup>) is a measure of effect size from ANOVA (% of variance explained)

P-value thresholds: \*\*\* < 0.001; \*\* < 0.01; \* < 0.05, ns: not significant

**Table 2.** Mean (+SD) for each plant variable according to treatments without earthworms (-EW) and with earthworms (+EW) after eight weeks of growth.

Variables	Unit	Rice cultivars															
		B22		F161		F172		F182		N4		W880					
		-EW	+EW	-EW	+EW	-EW	+EW	-EW	+EW	-EW	+EW	-EW	+EW				
Leaf Dry Matter Content	mg.g <sup>-1</sup>	2.4 (0.7)	2.8 (0.5)	2.7 (0.2)	2.9 (0.5)	2.1 (0.5)	2.4 (0.3)	2.1 (0.6)	2.0 (0.5)	2.3 (0.4)	2.1 (0.3)	2.0 (0.5)	2.2 (0.3)				
Specific Leaf Area	mm <sup>2</sup> .mg <sup>-1</sup>	11.9 (1.4)	10.7 (1.9)	14.3 (1.5)	17.9 (8.9)	12 (2.2)	12.7 (2.7)	14.2 (1.6)	12.3 (1.4)	14.0 (2.5)	10.4 (1.6)	14.7 (3.9)	13.9 (3.0)				
Normalized Difference Vegetation Index	Abs	0.025 (0.009)	0.039 (0.003)	0.029 (0.005)	0.039 (0.004)	0.029 (0.009)	0.043 (0.005)	0.030 (0.007)	0.049 (0.005)	0.028 (0.006)	0.051 (0.002)	0.030 (0.005)	0.047 (0.005)				
NDVI	Abs	0.55 (0.1)	0.76 (0.02)	0.54 (0.07)	0.70 (0.08)	0.49 (0.04)	0.7 (0.04)	0.51 (0.08)	0.71 (0.04)	0.50 (0.05)	0.75 (0.03)	0.61 (0.08)	0.70 (0.07)				
Shoot biomass	mg	59.4 (4.7)	136 (29.2)	47.4 (8.3)	104.0 (40.4)	44.6 (10.8)	89.1 (15.7)	38.7 (5.9)	81.9 (11.0)	35.0 (9.5)	73.7 (5.6)	61.9 (16.1)	86.7 (8.0)				
Root biomass	mg	83.9 (29.2)	99.6 (9.8)	54.0 (5.5)	90.6 (35.8)	48.3 (18.5)	94.1 (18.5)	48.3 (4.4)	107.0 (44.7)	33.9 (2.7)	143.2 (73.7)	66.7 (18.5)	113.8 (30.3)				
Shoot:root ratio	ratio	0.78 (0.25)	1.37 (0.3)	0.85 (0.13)	1.16 (0.22)	0.98 (0.2)	0.98 (0.27)	0.81 (0.18)	0.9 (0.4)	1.03 (0.26)	0.68 (0.41)	0.96 (0.22)	0.81 (0.24)				
Shoot P amount	μg-P.pot <sup>-1</sup>	46.7 (4.5)	95.8 (31.1)	35.2 (8.1)	86.1 (34.0)	30.0 (7.9)	66.7 (23.3)	39.3 (9.3)	56.0 (8.2)	31.2 (3.0)	58.6 (11.7)	42.8 (8.4)	58.4 (11.4)				
Shoot N amount	mg-N.pot <sup>-1</sup>	0.65 (0.06)	2.94 (0.88)	0.58 (0.16)	2.39 (0.88)	0.48 (0.16)	1.88 (0.52)	0.45 (0.06)	1.72 (0.37)	0.40 (0.15)	1.68 (0.11)	0.82 (0.31)	1.78 (0.29)				
P uptake efficiency	μg-P.g <sup>-1</sup> of dry root	0.61 (0.23)	0.97 (0.33)	0.64 (0.11)	0.97 (0.15)	0.66 (0.18)	0.73 (0.25)	0.83 (0.27)	0.60 (0.24)	0.92 (0.04)	0.54 (0.34)	0.67 (0.14)	0.55 (0.18)				
N uptake efficiency	mg-N.g <sup>-1</sup> of dry root	8.56 (3.07)	29.68 (8.82)	10.56 (2.55)	27.50 (5.68)	10.20 (1.86)	20.42 (5.81)	9.46 (2.02)	18.92 (9.28)	11.72 (4.15)	15.52 (9.80)	12.38 (2.95)	16.84 (6.22)				