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1 **Effects of conservation agriculture maize-based cropping systems on soil health and crop performance in**  
2 **New Caledonia**

3  
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19 **Abstract**

20 Conservation agriculture (CA) is one strategy with which both sustainability and productivity can be achieved by  
21 improving soil health. However, linkages between practices, soil health and cropping system performance  
22 remain poorly disentangled. We assessed the relationships between soil health and cropping system performance  
23 for three maize-based cropping systems in New Caledonia. Two CA systems, one with direct seeding into a  
24 mixed species dead mulch (CA-DM) and one into a stylo living mulch (CA-LM), were compared to a  
25 conventional tillage (CT) system. CA vs. CT experiment started in 2011, whereas the differentiation between  
26 CA-DM and CA-LM was initiated in 2017 only. In 2018, soil health was evaluated using Biofunctool®, a set of  
27 ten in-field tools that assess soil carbon transformation, structure maintenance and nutrient cycling functions.  
28 The performance of the three cropping systems were assessed by monitoring weeds, maize growth and yield  
29 components. Structural equation modelling (SEM) was used to disentangle the links between agricultural  
30 management, soil health and cropping system performance. Soil structure maintenance and nutrient cycling  
31 functions were higher under CA-DM and CA-LM than under CT, and carbon transformation function was higher  
32 under CA-DM than under CT and CA-LM. Overall, the soil health index (SHI) was 1.3-fold higher under CA  
33 systems than under CT. Cropping system management had both direct and indirect effects on soil functioning  
34 and crop productivity leading to a 1.3-fold higher yield under CA than under CT. The direct and indirect effects  
35 of CA systems on soil health had positive impacts on ecosystem services (*i.e.*, productivity, weed regulation and  
36 soil ecosystem services). Such integrative approaches that account for the relationships and possible trade-offs  
37 between cropping system components enable a better understanding of the effects and the performance of  
38 practices, and support adaptive agricultural management.

39

40 **Keywords** Cover crop; Living mulch; Magnesian fluvisol; No tillage; Soil functions; Systemic approach

## 41 1. Introduction

42 Agricultural practices are key drivers of agroecosystem functions and their negative impacts have increased in  
43 recent decades. Land use changes, intensive use of chemical inputs, and fragmentation of habitats have  
44 contributed to the depletion of soil fertility, biodiversity, water quality and availability, and to the magnitude of  
45 climate change (Foley et al., 2011; Rockström et al., 2017). These rapid changes have also had positive effects  
46 including increasing food production at global scale, but significant trade-offs have been observed, to preserve  
47 environmental integrity (Tilman et al., 2011). Soil is one of the key components of ecosystems and is under  
48 serious pressure from human activities. To mitigate the negative impacts of agricultural systems, some  
49 approaches promote agronomic technical levers such as soil conservation practices or agroforestry (Altieri and  
50 Nicholls, 2013; Wezel and Soldat, 2009).

51 Agriculture represents less than two per cent of the gross domestic product of New Caledonia where the  
52 economy is mainly driven by the nickel industry and the service sector (ISEE, 2016). However, islands in the  
53 South Pacific need to increase their agricultural production to respond to population growth and to increasing  
54 demand from the commercial sector (Murray, 2001; Naidu, 2010). Like in many developing countries,  
55 agricultural intensification in these islands has had positive impacts on agricultural production and food security  
56 (Naidu, 2010; van der Velde et al., 2007). Unfortunately, agricultural intensification has also had detrimental  
57 impacts on soil and water resources, including significant soil erosion (Dugain, 1953; Losfeld et al., 2015),  
58 especially in New Caledonia, a hotspot of biodiversity (Myers et al., 2000).

59 Conservation agriculture (CA) is a farming system that promotes minimum soil disturbance (*i.e.*, no tillage),  
60 maintenance of a permanent soil cover, and diversification of plant species (FAO, 2014). Through the  
61 application of these three principles, the maintenance and improvement of soil functioning is driven by (i) high  
62 and continuous production of above and belowground biomass, (ii) a permanent soil cover which supports a  
63 continuous flow of nutrients and organic compounds and improves the water balance, and (iii) enhanced soil  
64 biological activity which regulates carbon transformation, soil structure maintenance, and improved nutrient  
65 cycling (FAO, 2014; Hobbs et al., 2008; Scopel et al., 2013). CA is being promoted to improve the resilience of  
66 cropping systems and reduce their negative externalities (Hobbs et al., 2008; Lal, 2015a; Séguy et al., 2006). CA  
67 can help reduce physical, chemical and biological soil depletion and production costs (Palm et al., 2014; Scopel  
68 et al., 2013; Sithole et al., 2016; Thierfelder and Wall, 2012). CA practices could thus be a promising way to  
69 reduce the negative impacts of agriculture, especially on soil, while conserving production and ecosystem  
70 services (Pittelkow et al., 2015; Verhulst et al., 2010).



71 The relationships among soil and crop management practices, soil health, crop performance and ecosystem  
72 services under CA practices are poorly described in the literature (Palm et al., 2014; Ranaivoson et al., 2017;  
73 Verhulst et al., 2010). Appropriate and sensitive indicators should be selected to assess agrosystem  
74 multifunctionality. Soil health is defined as “the capacity of a soil to produce a good quantity and quality food  
75 and fibre together with the delivery of other ecosystem services” (Kibblewhite et al., 2008). Although many  
76 approaches are available to assess soil health, Thoumazeau et al. (2019b) proposed an integrative,  
77 multifunctional, and easily transferable approach, named Biofunctool<sup>®</sup>. Biofunctool<sup>®</sup> makes it possible to assess  
78 the three main soil functions linked to soil biological activities identified by Kibblewhite et al. (2008): (i) carbon  
79 transformation, (ii) nutrient cycling, and (iii) soil structure maintenance with a core set of ten in-field and low-  
80 tech indicators. Weeds and crop development are key aspects to assess cropping system performance. Weeds are  
81 indeed a major factor affecting yields (Teasdale et al., 2007) and weed control is one of the farmer’s main  
82 concerns in agricultural systems (Hobbs, 2007; Nichols et al., 2015; van Heemst, 1985). On the other hand, grain  
83 yield is the main indicator used by farmers to assess the performance of their system. Combining these  
84 measurements should help understand the synergies and trade-offs between the components that may affect  
85 cropping system performance.

86 We hypothesise that CA practices have both direct and indirect effects on weeds and crop productivity by  
87 influencing soil health, thereby increasing the performance of CA compared to that of CT. The overall objective  
88 of the study was to conduct an integrative and quantified assessment of the relationships between contrasted  
89 maize-based cropping management (*i.e.*, conventional plough-based tillage (CT), and CA with a diversity of  
90 cover crops and managements), soil health and cropping system performance in New Caledonia.

## 91 **2. Materials and methods**

### 92 2.1. Site description

93 The study site is located at the Adecap Technopole Ouenghi experimental station in Boulouparis, South province,  
94 New Caledonia (21°53'50" S, 166°06'45" E). The west coast of New Caledonia is characterised by a semi-arid  
95 subtropical climate with a cool, dry season from May to September, and a warm, wet season from December to  
96 April. Intense rainfall associated with thunderstorms peaking in austral summer are usually followed by recurrent  
97 drought periods from October to November. Data from the Ouenghi Meteo-France station (21°55'42"S,  
98 166°05'00"E; 3.5 km from the study site) were used to characterise the meteorological conditions. Mean annual  
99 precipitation between 2011 and 2018 was 909 mm with most of the rainfall occurring from February to April. In  
100 the same period, the monthly average minimum and maximum temperatures were 17 °C and 29 °C, respectively.

101 Soil is classified as a silty loam soil according to the USDA classification with 33.6% sand, 51.6% silt and  
102 14.8% clay (Euro-analyse laboratory soil analysis, 2011). It is a magnesian alkaline soil ( $\text{pH}_{\text{water}} = 8.1$ ) with high  
103 concentrations of  $\text{Mg}^{2+}$  (exchangeable magnesium accounts for 76% of cation exchange capacity) and  $\text{Ca/Mg} =$   
104 0.3 ( $\text{K/Mg} = 0.01$ ). The average bulk density (in the 0-10 cm layer) was  $1.01 \pm 0.08 \text{ g cm}^{-3}$  and soil organic  
105 carbon (0-20 cm depth) was  $28.1 \pm 1.1 \text{ g kg}^{-1}$  (LAMA laboratory soil analysis, 2017).

## 106 2.2. Experimental design

107 The experiment was set up in 2011 to study contrasted cropping systems representative of cereal production  
108 along the west coast of New Caledonia characterised by short rotations and maize (*Zea mays* L.) grain as main  
109 crop production. Two main periods characterize the experiment (Supplementary information, Table A.1). From  
110 2011-2016, the cropping sequence was based on a succession cowpea-maize and cowpea-maize-sorghum under  
111 two type of management: (i) conventional plough-based management (CT), and (ii) CA management based on  
112 dead mulch. Cowpea (*Vigna unguiculata* L.) was used as a cover crop before maize in all treatments. The second  
113 period started in 2017, when the cropping pattern was updated with a maize-based cropping system under three  
114 different managements: (i) maize under CT, which is the main practice in the region, which represented a  
115 continuation of the CT management of the first period, (ii) maize under CA with direct seeding in a dead mulch  
116 (CA-DM), and (iii) maize under CA with direct seeding in a living mulch (CA-LM). CA-DM and CA-LM  
117 represented the continuation of the plots under CA management in the first period. Crop residues were not  
118 exported in all the cropping systems, and under CT, the soil was ploughed once a year to a depth of 25-30 cm  
119 with a mouldboard plough. A randomised block design experiment was used consisting in the three treatments  
120 with three replicates of plots measuring 1200 m<sup>2</sup> (50 m x 24 m) for each system (Supplementary information,  
121 Fig. A.1).

122 In 2018, all cover crops were sown on the 24<sup>th</sup> of January with a no-till seeder (Semeato PD 17) (Supplementary  
123 information, Table A.2). The cover crop used under CA-DM consisted of a mix of four species: sorghum  
124 (*Sorghum bicolor* L. Moench, cv. sweet jumbo; sowing density 15 kg ha<sup>-1</sup>), sunnhemp (*Crotalaria juncea* L., cv.  
125 crescent sunn; 10 kg ha<sup>-1</sup>), cowpea (*Vigna unguiculata* L. Walp., cv. ebony; 10 kg ha<sup>-1</sup>), and lablab (*Lablab*  
126 *purpureus* L. Sweet, cv. highworth; 15 kg ha<sup>-1</sup>). The cover crop used under CA-LM was stylo (*Stylosanthes*  
127 *guianensis* Aubl. Sw.; 10 kg ha<sup>-1</sup>). Under CT, the mouldboard plough was used on the 19<sup>th</sup> of March 2018 to a  
128 depth of 25-30 cm, and the rotary cultivator on the 27<sup>th</sup> of April 2018 to a depth of 5-10 cm, before maize  
129 sowing. Under CA-DM, the cover crop was terminated by rolling combined with herbicide spraying on the 20<sup>th</sup>  
130 of April 2018, 15 days before the maize was sown. Under CA-LM, the maize was sown directly in standing

131 green stylo. The aboveground biomass of the cover crops was assessed before maize was sown and ranged from  
132  $22.6 \pm 8.8$  t<sub>dry matter (DM)</sub> ha<sup>-1</sup> to  $2.5 \pm 0.8$  t<sub>DM</sub> ha<sup>-1</sup> under CA-DM and CA-LM, respectively. Under CA-DM, 100%  
133 of the soil surface was covered by mulch at sowing and about 80% under CA-LM.

134 In all cropping systems, maize was grown during the dry, cool season (May-September) with 223 mm  
135 cumulative precipitation during the crop cycle. Maize (cv. CS Frontal) was sown at 108000 kernels ha<sup>-1</sup> in 76-cm  
136 rows on the 7<sup>th</sup> of May 2018, using a no-till seeder (Jumil JM3090 PD). A hose reel irrigation system was used  
137 on 13 occasions to supply 290 mm of water. The water balance method was used to determine water amounts,  
138 and irrigation uniformity was controlled by rain gauges. The nitrogen (N) fertilisation during the maize cycle  
139 included 350 kg ha<sup>-1</sup> of urea (46% N) and 300 kg ha<sup>-1</sup> of ammonium sulphate (21% N) applied 17 and 51 days  
140 after sowing (DAS), respectively. Herbicide treatments included pre- and post-emergence herbicides. Pre-  
141 emergence herbicides were applied immediately after sowing, while post-emergence herbicides were applied at  
142 10 and 31 DAS.

### 143 2.3. Soil monitoring and analysis

144 Biofunctool® consists in a set of ten functional indicators that assess three main soil functions with (i) carbon  
145 transformation, (ii) soil structure maintenance and (iii) nutrient cycling (Thoumazeau et al., 2019b). Four  
146 indicators were used to assess the changes of the carbon transformation including the labile fraction of the soil  
147 organic carbon (permanganate oxidizable carbon (POXC)) (Weil et al., 2003); the basal soil respiration  
148 (SituResp®) (Thoumazeau et al., 2017); and the soil biological activity using the bait lamina test (scored from 0  
149 [no degradation] to 1 [complete degradation]) (Törne, 1990; van Gestel et al., 2003) and the green tea bag (GTB)  
150 score (adapted from Keuskamp et al. (2013)). The bait lamina consists of a plastic strip, comprising 16 small  
151 holes, that was filled with an organic standard substrate, made of cellulose powder, bran flakes and active carbon  
152 (70:27:3). Bait laminas were vertically inserted in the soil for seven days. For the analysis, we used the average  
153 of lamina holes number 1 to 4 (0-2 cm) only, as it was the only depth that allowed us to significantly distinguish  
154 the treatments (Supplementary information, Fig. A.2). The GTB indicator consisted in the decomposed fraction  
155 of green tea after a burial period of 30 days.

156 We then used three indicators to study the impact of each cropping system on soil structure maintenance function  
157 by assessing soil aggregate water stability (AggSoil) at a depth of 0-10 cm (scored from 1 [poor] to 6 [high  
158 stability]) (Herrick et al., 2001), water infiltration (Beerkan) (Thoumazeau et al., 2019b), and soil structure  
159 (visual evaluation of soil structure (VESS)) in the 0-30 cm layer (scored from 1 [good] to 5 [poor soil structure])  
160 (Guimarães et al., 2011). The VESS consists of visually assessing the size and porosity of aggregates, the

161 strength of aggregates, the presence of roots and the colour of the soil. Finally, we used three indicators to study  
162 the impact of each cropping system on soil nutrient cycling function. We quantified available ammonium (N-  
163  $\text{NH}_4^+$ ) and nitrate (N- $\text{NO}_3^-$ ) in the soil after extraction with 1M KCl (Maynard et al., 1993; Thoumazeau et al.,  
164 2019b). Soil nitrate dynamics were evaluated using anion exchange membrane (AEM- $\text{NO}_3^-$ ) placed horizontally  
165 at a depth of 8 cm for a 10 days burial period (Qian and Schoenau, 2002; Thoumazeau et al., 2019b).  
166 Except for the VESS, soil samples were collected in June 2018 in the 0-10 cm soil layer. This soil layer was  
167 selected to fit with Biofunctool® approach that aims at integrating soil biological activities (Thoumazeau et al.,  
168 2019b). Also, early changes under CA mostly occur at the soil surface, making the top soil assessment highly  
169 relevant (de Moraes Sa and Lal, 2009). Three sampling points (internal replicates) were collected per plot giving  
170 a total of 27 soil samples for Biofunctool® analysis (except for available nitrogen (N- $\text{NH}_4^+$ , N- $\text{NO}_3^-$ ) for which  
171 only one replicate per plot was analysed).

#### 172 2.4. Agronomic data collection

173 Weed biomass was assessed using a quadrat sampling method at four maize stages: sowing, 6-leaf (25 DAS),  
174 flowering (80 DAS), and post-harvest. In each repetition (three repetitions per treatment), three quadrats of 0.25  
175  $\text{m}^2$  were delimited to count weeds. Weed aboveground biomass was then determined for each sampling period  
176 after drying at 80 °C until constant mass was reached. Cumulative weed biomass per treatment was determined  
177 by adding the dry matter of the four sampling periods.

178 Maize density was monitored weekly in three subplots per repetition (three repetitions per treatment) on two  
179 contiguous maize rows two meters in length (3.04  $\text{m}^2$ ) from emergence to the 8-leaf (35 DAS) stage. Maize  
180 density per treatment was the average of the maize counted during the successive sampling periods.

181 At harvest on the same subplots, thousand kernel weight (TKW) was measured at random from the grain lot of  
182 five maize plants per repetition (three repetitions per treatment). Three subsamples per repetition of one hundred  
183 kernels were dried at 80 °C until constant mass was reached and weighed. TKW was then standardized to 13%  
184 moisture content.

185 The yield was recorded from five plants randomly selected from three sub-plots per repetition (three repetitions  
186 per treatment) following methodologies from Echarte et al. (2006) and Daei et al. (2009). The ears were counted,  
187 and hand-shelled. The kernels of each ear were dried, and weighed. The grain yield was calculated as follows  
188 and standardized to 13% moisture content:

189  $\text{Maize yield (t ha}^{-1}\text{)} = \text{Maize density (plants m}^2\text{)} * \text{Number of ears per plant (ear plant}^{-1}\text{)} *$

190  $\text{Kernel weight per ear (g ear}^{-1}\text{)} * 10^{-2}$

## 191 2.5. Statistical analysis

192 All statistical analyses were performed using R software 3.6.0 (R Development Core Team, 2008).  
193 First, each Biofunctool® indicator was analysed separately using a linear-mixed effects model (package lme4,  
194 (Bates et al., 2015)). Treatment was defined as fixed factor and replicates (plots and internal replicates) as  
195 random factors. After checking the normality of the model residuals and the homoscedasticity of the variance  
196 residuals, ANOVAs were run using the car package (Fox and Weisberg, 2011). This was followed by a post-hoc  
197 mean comparison, using Tukey's test with Bonferroni adjustment (Hothorn et al., 2008).  
198 After analysing each indicator separately, indicators were computed within a principal component analysis  
199 (PCA) (FactoMineR package, (Lê et al., 2008)). The last step of analysis consisted in calculating the  
200 Biofunctool® soil health index (SHI), according to the methodology defined by Obriot et al. (2016) and  
201 Thoumazeau et al. (2019a). First, a weight was applied to the PCA variable to give the same weight to each soil  
202 function. The scoring function of the indicators was based on the "more is better" response curve, except for the  
203 VESS indicator where the "less is better" was used (Obriot et al., 2016). The SHI finally ranged from 0 (low) to  
204 1 (high soil health). After calculation of the index, a variance analysis of the contribution of each soil function to  
205 the final score was run using one-way ANOVA.  
206 Next, we used SEM (Grace et al., 2012, 2007) to explicit relationships from a web of possible causal pathways,  
207 including direct and indirect effects between practices (CT and CA systems), soil health and cropping systems  
208 performance. CA-DM and CA-LM were grouped into a single cropping system modality (CA). A combination  
209 of the aboveground biomass of the cover crops at maize sowing and the soil management practices (qualitative  
210 data) was used to characterize cropping system practices for the SEM. The three Biofunctool® aggregated  
211 functions (*i.e.*, structure maintenance, nutrient cycling, and carbon transformation) were used as soil health  
212 indicators. Cumulative weed aboveground biomass during the maize cycle, maize thousand kernel weight  
213 (TKW) and grain yield were used as cropping system performance parameters for the SEM. Weeds are a major  
214 factor that affects yields (Teasdale et al., 2007). TKW was used to assess maize growth performance, providing  
215 insight into the strength of late competition (Meynard and David, 1992). Grain yield expresses the overall  
216 conditions of the crop cycle, and is the main indicator used to assess system productivity. Strength and  
217 directionality (positive or negative) of the relationship between variables are indicated through the path  
218 coefficients. The SEM was performed using the piecewiseSEM package (Lefcheck, 2016).

## 219 3. Results

### 220 3.1. Effects of the cropping systems on soil health

221 For carbon transformation, labile fraction of the soil organic carbon (POXC), basal soil respiration (SituResp®)  
222 values as well as bait lamina scores were significantly higher under the two CA cropping systems than under CT  
223 (Table 1). The GTB score was significantly higher under CA-DM ( $0.46 \pm 0.03$ ) than under CT ( $0.43 \pm 0.02$ ) but  
224 did not significantly differ from CA-LM ( $0.45 \pm 0.02$ ).

225 Concerning structure maintenance, the same trend was recorded for the three indicators (Table 2). Mean VESS  
226 scores were significantly lower for soils under CA ( $1.45 \pm 0.3$  and  $1.28 \pm 0.3$  for CA-DM and CA-LM,  
227 respectively) indicating a better soil structure than under CT soil ( $2.11 \pm 0.4$ ). Mean AggSoil scores were  
228 significantly lower under CT soil ( $1.22 \pm 0.4$ ) than CA soils ( $2.00 \pm 0.8$  and  $2.15 \pm 0.9$  for CA-DM and CA-LM,  
229 respectively). Finally, water infiltration was two-fold lower in soil under CT ( $93.4 \pm 20.5 \text{ mL min}^{-1}$ ) than in soil  
230 under CA ( $176.5 \pm 71.5$  and  $226.0 \pm 117.3 \text{ mL min}^{-1}$  for CA-DM and CA-LM, respectively). No significant  
231 differences were found in VESS, AggSoil, and Beerkan scores between CA-DM and CA-LM.

232 For nutrient cycling, the mean AEM-NO<sub>3</sub><sup>-</sup> score was two-fold higher under CT than under CA ( $20.4 \pm 6.4$  vs.  
233  $10.5 \pm 4.0$  and  $9.8 \pm 5.0 \mu\text{g cm}^{-2} \text{ d}^{-1}$  for CA-DM and CA-LM, respectively) (Table 3). In contrast, the  
234 concentration of N-NH<sub>4</sub><sup>+</sup> was two-fold higher under CA-DM than under CT ( $6.1 \pm 0.2 \text{ mg kg}^{-1}$  vs.  $2.6 \pm 0.3 \text{ mg}$   
235  $\text{kg}^{-1}$ ). The concentration of N-NO<sub>3</sub><sup>-</sup> tended to be higher under CA than under CT but the differences were not  
236 statistically significant.

237 The PCA performed on the 10 functional indicators allowed to separate the treatments (Fig. 1). The differences  
238 between Biofunctool® indicators appeared mainly between the CT and CA cropping systems. Total variability  
239 was represented at 45.7% on the first axis and at 14.2% on the second axis. The difference in soil health between  
240 the two CA cropping systems and CT was mainly based on indicators linked with the first axis: AEM-NO<sub>3</sub><sup>-</sup> and  
241 N-NH<sub>4</sub><sup>+</sup> (nutrient cycling), VESS and AggSoil (structure maintenance), and POXC (carbon transformation).

242 Biofunctool® SHI values for CA treatments were about 1.3-fold higher than under CT (mean value of 0.7 vs. 0.5)  
243 (Fig. 2). For the nutrient cycling and the structure maintenance functions, the main differences were observed  
244 between CT and CA with mean CA scores (CA-DM and CA-LM) 20% and 46% higher than under CT,  
245 respectively. Concerning soil carbon transformation function, only the CA-DM score was significantly higher  
246 than CA-LM and CT, representing an increase of 12%.

### 247 3.2. Performance of the cropping systems

248 The cumulative aboveground weed biomass differed significantly among the three treatments with higher weed  
249 biomass under CT (mean value of  $1.4 \pm 0.7 \text{ t}_{\text{DM}} \text{ ha}^{-1}$ ) than under CA-LM ( $0.2 \pm 0.3 \text{ t}_{\text{DM}} \text{ ha}^{-1}$ ) and CA-DM ( $0.7 \pm$   
250  $0.3 \text{ t}_{\text{DM}} \text{ ha}^{-1}$ ) (Table 4).

251 Maize density differed significantly among the treatments: the maize plant population was higher under CA-LM  
252 ( $10.3 \pm 0.5$  plants  $m^{-2}$ ) than under CT ( $9.0 \pm 0.4$  plants  $m^{-2}$ ) and CA-DM ( $8.0 \pm 1.1$  plants  $m^{-2}$ ), with a decrease at  
253 emergence under CA-DM.

254 There was one ear per plant for all the maize plants sampled. The kernel weight per ear was significantly higher  
255 under CA-DM ( $158.6 \pm 25.5$  g) than under CA-LM and CT ( $125.8 \pm 18.2$  g and  $107.8 \pm 21.0$  g, respectively).  
256 The TKW followed the same trend and was significantly higher under CA-DM ( $388.2 \pm 7.5$  g) than under both  
257 CA-LM and CT ( $364.2 \pm 12.9$  g and  $355.1 \pm 16.3$  g, respectively).

258 Maize grain yields ranged from  $9.7 \pm 2.0$  t  $ha^{-1}$  under CT to  $12.7 \pm 2.9$  t  $ha^{-1}$  and  $12.9 \pm 1.8$  t  $ha^{-1}$  under CA-DM  
259 and CA-LM, respectively, and were significantly higher under the two CA treatments than under CT.

### 260 3.3. Links between practices, soil health, and cropping system performance

261 The SEM fitness index was significant (Fisher's test  $P = 0.255$ ), and six of the 21 relationships tested were  
262 significant (Fig. 3). SEM revealed significant links between agricultural practices and soil health: CT had a  
263 negative influence on soil structure maintenance (path coefficient = -0.55) while CA had positive effects on  
264 carbon transformation and nutrient cycling (path coefficient = 0.38 and 0.33, respectively). SEM also confirmed  
265 significant links between agricultural practices and cropping system performance: CT had a positive impact on  
266 weed development with higher biomass collected (path coefficient = 0.40) whereas CA had a positive influence  
267 on TKW (path coefficient = 0.46). Finally, SEM highlighted significant links between soil functions and  
268 cropping system performance with a positive correlation between nutrient cycling and weed development (path  
269 coefficient = 0.36). However, no significant indirect effects of soil health on maize crop performance emerged.

## 270 4. Discussion

271 It is worth noting that the results are based on the cumulative effects of the two distinct periods linked to changes  
272 in the experiment management strategy. The results of CT compared to CA are linked to a relatively long-term  
273 change (2011-2018), whereas the results that compare CA practices are linked to short-term changes (2017-  
274 2018).

### 275 4.1. Effects of CA cropping systems on soil functions

276 First, higher POXC and SituResp<sup>®</sup> scores were measured under CA treatments than under CT. POXC is sensitive  
277 to management practices, and mainly depends on the amount of residues returned to the soil (Bongiorno et al.,  
278 2019; Chan et al., 2002). Plant material including above- and below-ground biomass and living organisms  
279 mainly contribute to the labile carbon fraction. The higher basal soil respiration observed in soils under CA can  
280 be explained by the increased labile carbon fraction, which stimulated microbial pools and activity (Balota et al.,

281 2004; Bongiorno et al., 2019). Bait laminas and GTB bioindicators showed greater biological activity in CA  
282 cropping systems than under CT. Concerning laminas, feeding activity was mainly observed in the 0-2 cm layer.  
283 This vertical feeding pattern has already been reported in the literature and the 0-2 cm layer was mentioned as a  
284 key layer (Gongalsky et al., 2004; Hamel et al., 2007; Rozen et al., 2010). In our system, the vertical pattern can  
285 be explained by the effects of cover crop residues on the soil surface and root systems of dead and living  
286 mulches that may affect specific organisms such as earthworms (van Gestel et al., 2003) and soil mesofauna  
287 (Helling et al., 1998), and then reflected in the bait lamina score. Concerning the GTB indicator, only CA-DM  
288 had a higher score than CT. CA-DM thus enhanced decomposition of the green tea at a depth of 8 cm thanks to  
289 soil biological activity (Tóth et al., 2018). The larger quantity of mulch under CA-DM ( $22.6 \text{ t}_{\text{DM}} \text{ ha}^{-1}$ ) than under  
290 CA-LM ( $2.5 \text{ t}_{\text{DM}} \text{ ha}^{-1}$ ) may have had a short term positive effect on the environmental variables (*e.g.*, soil  
291 moisture) resulting in differences in soil biological activity (Arroita et al., 2013). The difference in mulch quality  
292 (N contents: 1.14% and 2.82% of DM for CA-DM and CA-LM, respectively) is also an important factor that  
293 may have influenced the activity under CA-DM compared with CA-LM (Lienhard et al., 2013; Nemergut et al.,  
294 2010; Pascault et al., 2010).

295 The VESS, Beerkan and AggSoil indicators were significantly improved by CA management. The absence of  
296 tillage combined with the presence of plant residues on the soil surface, and living or dead cover crop root  
297 systems globally improved the structure maintenance function (Indoria et al., 2017; Tivet et al., 2013). The  
298 addition of residues and mulches stimulated microbial activity, which, along with root exudates, enhanced  
299 aggregate stability (Lal, 2015b; Zuber et al., 2017). In contrast, tillage destroyed soil aggregates, thereby  
300 increasing slaking and pore clogging, which could reduce porosity and infiltration rates (Mitchell et al., 2017;  
301 Rosolem et al., 2016).

302 A higher concentration of  $\text{NH}_4^+$  and a trend (although not significant) of higher concentration of  $\text{NO}_3^-$  were  
303 observed under CA. These results were linked to a better soil structure (AggSoil) enabling diversified pH-redox  
304 (Eh) niches, and consequently diversified microbial communities (Husson et al., 2018). The soil nitrogen should  
305 have therefore operated in a variety of forms from nitrate to ammonium in the 0-10 cm layer. The better soil  
306 structure (AggSoil) explains the better water infiltration but also the fact that concentrations of both nitrate and  
307 ammonium were higher under CA. In their study on a Red Oxisol in Cambodia, Pheap et al. (2019) also reported  
308 higher concentrations of  $\text{NO}_3^-$  (although not significant) and  $\text{NH}_4^+$  under CA compared with CT. As ion  
309 exchange membranes aim at mimicking plant-rooting systems, measurement of the AEM- $\text{NO}_3^-$  indicator  
310 provided information on plant nutrient absorption and dynamics based on soil and crop management (Le Cadre



311 et al., 2018; Qian and Schoenau, 2002). Compared to other measurements such as nitrate and ammonium  
312 extracted from the soil, the quantity of nitrate adsorbed on the membrane was two-fold higher under CT than  
313 CA. Tillage may expose previously protected organic matter which may then serve as a substrate for microbial  
314 growth (Rovira and Greacen, 1957), stimulating mineralisation and nitrification under an oxidized environment  
315 (Calderón et al., 2001; Muruganandam et al., 2010), explaining higher nitrate dynamics under CT. However, this  
316 tillage-induced nitrogen dynamics can lead to N losses through denitrification and nitrate leaching especially  
317 under soil with poor soil structure, which could explain the smaller amounts of available  $\text{N-NH}_4^+$  and  $\text{N-NO}_3^-$   
318 from soil extraction measured under CT (Boulakia et al., 2019; Calderón et al., 2001; Chatskikh and Olesen,  
319 2007; Ruan and Robertson, 2013). In addition, the results of AEM- $\text{NO}_3^-$  can be analysed in accordance with a  
320 previous study conducted by Husson et al. (2018) who observed a reversed soil profile for the redox potential  
321 when comparing CA to CT for four soil types in France. The authors observed lower redox potential on the soil  
322 surface under CA which is likely to lead to a higher concentration of  $\text{NH}_4^+$ , while limiting N leaching. Under CT,  
323 they observed a higher redox potential on the soil surface (0-5 cm) and a strong decrease with depth creating an  
324 electrical force which pushes the negative charges from the soil surface to depth. The higher oxidation on the top  
325 soil under CT and the trend of Eh from the soil surface to depth may increase  $\text{NO}_3^-$  leaching. We can also note  
326 that the  $\text{NH}_4^+:\text{NO}_3^-$  ratio is 27-73% under CA (average of CA-DM and CA-LM) and 20-80% under CT which  
327 can lead to a physiological imbalance in the plant, alkalize the rhizosphere, promoting fungi, viruses, bacteria  
328 and insects (Husson et al., 2018). Considering these results and the key role of Eh to characterize soil health  
329 (Cottes et al., 2020; Husson, 2013), it would appear judicious to consider the assessment of the redox potential  
330 within the framework of Biofunctool®.

331 At multivariate and Biofunctool® index analysis scales, the results generally reflect the trend observed at  
332 indicator scale, *i.e.*, the improvement in soil functioning was mainly observed between CT and the two CA  
333 systems (CA-DM and CA-LM). The Biofunctool® index showed better soil health under CA than under CT. The  
334 three soil functions also mainly reflected the difference between CT and CA. However, the carbon  
335 transformation function under CA-LM did not differ significantly from that under CT. This may be directly  
336 linked to the quality and the larger quantity of the biomass inputs under CA-DM than under CA-LM and CT,  
337 although the living root biomass may have affected soil biological activity and carbon turnover under CA-LM.  
338 Thus, no significant differences in SHI were observed between CA-DM and CA-LM probably due to the  
339 relatively recent establishment of the CA-LM cropping system (2 years).

340 4.2. Effects of CA cropping systems on crop performance

341 CA has significant and positive effects on soil functions that are likely to produce similar or even higher crop  
342 yields than CT (Thierfelder et al., 2015; Triplett and Dick, 2008). In this study, regardless of the cropping  
343 system, maize yields were generally high compared to current average farm yield of 9 t ha<sup>-1</sup>. Moreover, maize  
344 yields were 1.3-fold higher under CA-DM and CA-LM than under CT. These results are consistent with those of  
345 other studies, in which the positive impact of CA on crop yield was also demonstrated (Lal, 2014; Pittelkow et  
346 al., 2015; Ranaivoson et al., 2019; Rusinamhodzi et al., 2011). At the same time, these results contrast with other  
347 studies with mixed conclusions (Erenstein et al., 2012; Pittelkow et al., 2015; Thierfelder et al., 2015) that may  
348 arise from geographical and environmental patterns of CA implementation, duration, quality and quantity of the  
349 biomass-C inputs (DeFelice et al., 2006; Fujisaki et al., 2018; Gruber et al., 2012; Thierfelder et al., 2015).

350 In the present experiment, the physical barrier of the high biomass input of the dead mulch under CA-DM has  
351 reduced seed-soil contact and promoted early season insect damage, decreasing final plant density. This  
352 observation is corroborated by previous studies, including those by Bezuidenhout et al. (2012) and Pantoja et al.  
353 (2015). In contrast, maize density with direct sowing in standing green stylo under CA-LM was higher than  
354 under CT because it avoids the formation of a slaking crust and provides better maize emergence conditions.

355 CA-DM produced higher yield as well as kernel weight and TKW. The large amount of cover crop residues  
356 under CA-DM provided better growth conditions at grain filling and enhanced available resources for maize due  
357 to less competition thanks to lower maize density and reduced weed development, increased soil water  
358 infiltration and water holding capacity (Ranaivoson et al., 2017). In comparison, higher yield was also observed  
359 under CA-LM compared with CT, while similar kernel weight and TKW values were observed for both  
360 treatments. This suggests the same late cycle crop conditions as CT with advantages in the early stages due to  
361 better weed control, reduced formation of a slaking crust (Scopel and Findeling, 2001; Sithole et al., 2016;  
362 Verhulst et al., 2010), with higher maize density and complementarity of stylo and maize during the growth  
363 period (Birteeb et al., 2011; Edye et al., 1977). Finally, the short period (2 cycles) of CA-LM practice may not be  
364 sufficient for the soil to reach a new equilibrium and thus may not provide all support and provisioning services  
365 (Gruber et al., 2012; He et al., 2011; Machado et al., 2008).

#### 366 4.3. Systemic approach of CA cropping systems

367 SEM confirmed direct causal relationships of management practices on soil functioning revealed by  
368 Biofunctool®. In the long term, CT exhibited negative effects on soil health impacting soil structure  
369 maintenance, disrupting soil aggregation, exposing the labile carbon pool encapsulated within the aggregates to  
370 microbial oxidation and reducing water infiltration (Mitchell et al., 2017). By contrast, CA positively influenced

371 carbon transformation and nutrient cycling functions. Several studies emphasized that CA systems contribute to  
372 an accumulation of soil organic carbon (Cheesman et al., 2016; Lal, 2015c; Powlson et al., 2016), primarily due  
373 to the continuous inputs of biomass (above and belowground), the quality of the inputs, and the protection of the  
374 labile carbon pool from microbial transformation (Fujisaki et al., 2018; Virto et al., 2012). Concomitantly, a  
375 higher soil available nitrogen concentration ( $\text{N-NO}_3^-$ ,  $\text{N-NH}_4^+$ ) was assessed under CA systems, promoting crop  
376 growth supported by a higher structure maintenance function, and consequently limiting nitrogen losses  
377 compared to CT (Calderón et al., 2001; Chatskikh and Olesen, 2007; Husson et al., 2018).

378 In the short term, management practices had direct effects on the performance of the cropping systems. During  
379 the early stages of maize growth, more weeds were recorded under CT while the physical barrier and the  
380 allelopathy effect of dead or living mulch under CA systems reduced weed pressure (Altieri et al., 2011; Burgos  
381 and Talbert, 1996; Murphy et al., 2006). On the other hand, SEM highlighted a positive effect of CA systems on  
382 TKW. The period from flowering to grain filling is highly sensitive to water stress, and the higher kernel weight  
383 was the result of better conditions under CA (Bolaños and Edmeades, 1996; NeSmith and Ritchie, 1992). Mulch  
384 was shown to be an effective way to reduce soil evaporation and to moderate the temperature at the surface of  
385 the soil, which, along with the higher infiltration rate, improved water-use efficiency notably during the maize  
386 grain filling period (Hartkamp et al., 2004).

#### 387 4.4. Toward the quantification of linkages between soil health, productivity, and ecosystem services

388 The comprehensive links between agricultural practices, soil functions and ecosystem services (*i.e.*, productivity,  
389 weed regulation, and soil ecosystem services) were analysed with the SEM approach. In our study, the link  
390 between soil health and plant productivity was not significant and cropping system management was the main  
391 direct factor explaining differences in yield components. However, with same fertilisation and irrigation  
392 management, the CA cropping systems improved the overall crop conditions leading to a higher yield than under  
393 CT. Further understanding of the indirect effects of agricultural practices and soil health on crop productivity are  
394 needed. Long-term agronomic trial would make it possible to apply such a systemic approach and would be  
395 particularly helpful in quantifying the links between system management, soil functioning and crop productivity.  
396 Finally, we focussed on the links between soil functions, productivity, and weed regulation, but other ecosystem  
397 services also need to be tackled, for example, pest regulation, pollination, or biodiversity maintenance (Chabert  
398 and Sarthou, 2020).

399

## 400 5. Conclusions

401 The effects of three annual cropping systems (*i.e.*, CT, CA-DM and CA-LM) on soil functioning were evaluated  
402 using an integrative assessment of soil health. Higher structure maintenance (*i.e.*, soil aggregation, water  
403 infiltration, VESS) and nutrient cycling functions (*i.e.*,  $\text{NO}_3^-$ ,  $\text{NH}_4^+$ ) were recorded under CA-DM and CA-LM,  
404 and a higher carbon transformation function (*i.e.*, labile-C, soil respiration, baits lamina, GTB) was assessed  
405 under CA-DM. Overall, the soil health index (SHI) was 1.3-fold higher under CA systems than under CT  
406 although it did not differ between CA-DM and CA-LM, probably because the two CA management practices  
407 were recently established. By combining these results with the application of structural equation modelling  
408 (SEM), we identified relationships between soil functions and cropping system performance that are sensitive to  
409 cover crops and tillage practices. CA practices had both direct and indirect influence on soil health, thereby  
410 improving yield system performance when compared to CT. These findings indicate that CA systems are  
411 promising alternatives to the conventional plough-based system in the magnesian Fluvisol context of the west  
412 coast of New Caledonia.

413

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- 419 Altieri, M.A., Lana, M.A., Bittencourt, H.V., Kieling, A.S., Comin, J.J., Lovato, P.E., 2011. Enhancing crop  
420 productivity via weed suppression in organic no-till cropping systems in Santa Catarina, Brazil. *J.*  
421 *Sustain. Agric.* 35, 855–869. <https://doi.org/10.1080/10440046.2011.588998>
- 422 Altieri, M.A., Nicholls, C.I., 2013. The adaptation and mitigation potential of traditional agriculture in a  
423 changing climate. *Clim. Change* 140, 33–45. <https://doi.org/10.1007/s10584-013-0909-y>
- 424 Arroita, M., Causapé, J., Comín, F.A., Díez, J., Jimenez, J.J., Lacarta, J., Lorente, C., Merchán, D., Muñoz, S.,  
425 Navarro, E., Val, J., Elozegi, A., 2013. Irrigation agriculture affects organic matter decomposition in  
426 semi-arid terrestrial and aquatic ecosystems. *J. Hazard. Mater.* 263, 139–145.  
427 <https://doi.org/10.1016/j.jhazmat.2013.06.049>
- 428 Balota, E.L., Colozzi Filho, A., Andrade, D.S., Dick, R.P., 2004. Long-term tillage and crop rotation effects on  
429 microbial biomass and C and N mineralization in a Brazilian Oxisol. *Soil Tillage Res.* 77, 137–145.  
430 <https://doi.org/10.1016/j.still.2003.12.003>
- 431 Bates, D., Mächler, M., Bolker, B., Walker, S., 2015. Fitting linear mixed-effects models using lme4. *J. Stat.*  
432 *Softw.* 67. <https://doi.org/10.18637/jss.v067.i01>
- 433 Bezuidenhout, S.R., Reinhardt, C.F., Whitwell, M.I., 2012. Cover crops of oats, strolling rye and three annual  
434 ryegrass cultivars influence maize and *Cyperus esculentus* growth. *Weed Res.* 52, 153–160.  
435 <https://doi.org/10.1111/j.1365-3180.2011.00900.x>
- 436 Birteeb, P.T., Addah, W., Jakper, N., Addo-Kwafo, A., 2011. Effects of intercropping cereal-legume on biomass  
437 and grain yield in the savannah zone. *Livest. Res. Rural Dev.* 23.
- 438 Bolaños, J., Edmeades, G.O., 1996. The importance of the anthesis-silking interval in breeding for drought  
439 tolerance in tropical maize. *Field Crops Res.* 48, 65–80. [https://doi.org/10.1016/0378-4290\(96\)00036-6](https://doi.org/10.1016/0378-4290(96)00036-6)
- 440 Bongiorno, G., Bünemann, E.K., Oguejiofor, C.U., Meier, J., Gort, G., Comans, R., Mäder, P., Brussaard, L., de  
441 Goede, R., 2019. Sensitivity of labile carbon fractions to tillage and organic matter management and  
442 their potential as comprehensive soil quality indicators across pedoclimatic conditions in Europe. *Ecol.*  
443 *Indic.* 99, 38–50. <https://doi.org/10.1016/j.ecolind.2018.12.008>
- 444 Boulakia, S., Tivet, F., Husson, O., Seguy, L., 2019. Nutrient management practices and benefits in  
445 Conservation Agriculture systems, in: *Advances in Conservation Agriculture*. Amir Kassam,  
446 Cambridge, UK.
- 447 Burgos, N.R., Talbert, R.E., 1996. Weed control and sweet corn (*Zea mays* var. *rugosa*) response in a no-till  
448 system with cover crops. *Weed Sci.* 44, 355–361.
- 449 Calderón, F.J., Jackson, L.E., Scow, K.M., Rolston, D.E., 2001. Short-term dynamics of nitrogen, microbial  
450 activity, and phospholipid fatty acids after tillage. *Soil Sci. Soc. Am. J.* 65, 118–126.  
451 <https://doi.org/10.2136/sssaj2001.651118x>
- 452 Chabert, A., Sarthou, J.-P., 2020. Conservation agriculture as a promising trade-off between conventional and  
453 organic agriculture in bundling ecosystem services. *Agric. Ecosyst. Environ.* 292, 106815.  
454 <https://doi.org/10.1016/j.agee.2019.106815>
- 455 Chan, K.Y., Heenan, D.P., Oates, A., 2002. Soil carbon fractions and relationship to soil quality under different  
456 tillage and stubble management. *Soil Tillage Res.* 63, 133–139. [https://doi.org/10.1016/S0167-1987\(01\)00239-2](https://doi.org/10.1016/S0167-1987(01)00239-2)
- 457 Chatskikh, D., Olesen, J.E., 2007. Soil tillage enhanced CO<sub>2</sub> and N<sub>2</sub>O emissions from loamy sand soil under  
458 spring barley. *Soil Tillage Res.* 97, 5–18. <https://doi.org/10.1016/j.still.2007.08.004>
- 459 Cheesman, S., Thierfelder, C., Eash, N.S., Kassie, G.T., Frossard, E., 2016. Soil carbon stocks in conservation  
460 agriculture systems of Southern Africa. *Soil Tillage Res.* 156, 99–109.  
461 <https://doi.org/10.1016/j.still.2015.09.018>
- 462 Cottes, J., Saquet, A., Palayret, L., Husson, O., Beghin, R., Allen, D., Scheiner, J., Cabanes, C., Guiresse, M.,  
463 2020. Effects of soil redox potential (Eh) and pH on growth of sunflower and wheat. *Arch. Agron. Soil*  
464 *Sci.* 66, 473–487.
- 465 Daei, G., Ardekani, M.R., Rejali, F., Teimuri, S., Miransari, M., 2009. Alleviation of salinity stress on wheat  
466 yield, yield components, and nutrient uptake using arbuscular mycorrhizal fungi under field conditions.  
467 *J. Plant Physiol.* 166, 617–625.
- 468 de Moraes Sa, J.C., Lal, R., 2009. Stratification ratio of soil organic matter pools as an indicator of carbon  
469 sequestration in a tillage chronosequence on a Brazilian Oxisol. *Soil Tillage Res.* 103, 46–56.
- 470 DeFelice, M.S., Carter, P.R., Mitchell, S.B., 2006. Influence of tillage on corn and soybean yield in the United  
471 States and Canada. *Crop Manag.* 5. <https://doi.org/10.1094/CM-2006-0626-01-RS>
- 472 Dugain, F., 1953. Premières observations sur l'érosion en Nouvelle-Calédonie. *Agron. Trop.* 466–475.
- 473 Echarte, L., Andrade, F.H., Sadras, V.O., Abbate, P., 2006. Kernel weight and its response to source  
474 manipulations during grain filling in Argentinean maize hybrids released in different decades. *Field*  
475 *Crops Res.* 96, 307–312.

- 477 Edye, L., Williams, W., Burt, R., Grof, B., Stillman, S., Winter, W., 1977. The assessment of seasonal yield  
478 using some *Stylosanthes guyanensis* accessions in humid tropical and sub-tropical environments. *Aust.*  
479 *J. Exp. Agric.* 17, 425. <https://doi.org/10.1071/EA9770425>
- 480 Erenstein, O., Sayre, K., Wall, P., Hellin, J., Dixon, J., 2012. Conservation agriculture in maize-and wheat-based  
481 systems in the (sub) tropics: lessons from adaptation initiatives in South Asia, Mexico, and Southern  
482 Africa. *J. Sustain. Agric.* 36, 180–206. <https://doi.org/10.1080/10440046.2011.620230>
- 483 FAO, 2014. Conservation Agriculture. [WWW Document]. URL [http://www.fao.org/conservation-](http://www.fao.org/conservation-agriculture/en/)  
484 [agriculture/en/](http://www.fao.org/conservation-agriculture/en/) (accessed 8.17.19).
- 485 Foley, J.A., Ramankutty, N., Brauman, K.A., Cassidy, E.S., Gerber, J.S., Johnston, M., Mueller, N.D.,  
486 O’Connell, C., Ray, D.K., West, P.C., Balzer, C., Bennett, E.M., Carpenter, S.R., Hill, J., Monfreda, C.,  
487 Polasky, S., Rockström, J., Sheehan, J., Siebert, S., Tilman, D., Zaks, D.P.M., 2011. Solutions for a  
488 cultivated planet. *Nature* 478, 337–342. <https://doi.org/10.1038/nature10452>
- 489 Fox, J., Weisberg, S., 2011. An {R} Companion to Applied Regression.
- 490 Fujisaki, K., Chevallier, T., Chapuis-Lardy, L., Albrecht, A., Razafimbelo, T., Masse, D., Ndour, Y.B., Chotte,  
491 J.-L., 2018. Soil carbon stock changes in tropical croplands are mainly driven by carbon inputs: A  
492 synthesis. *Agric. Ecosyst. Environ.* 259, 147–158. <https://doi.org/10.1016/j.agee.2017.12.008>
- 493 Gongalsky, K.B., Pokarzhevskii, A.D., Filimonova, Z.V., Savin, F.A., 2004. Stratification and dynamics of bait-  
494 lamina perforation in three forest soils along a north–south gradient in Russia. *Appl. Soil Ecol.* 25, 111–  
495 122. <https://doi.org/10.1016/j.apsoil.2003.09.001>
- 496 Grace, J.B., Michael Anderson, T., Smith, M.D., Seabloom, E., Andelman, S.J., Meche, G., Weiher, E., Allain,  
497 L.K., Jutila, H., Sankaran, M., Knops, J., Ritchie, M., Willig, M.R., 2007. Does species diversity limit  
498 productivity in natural grassland communities? *Ecol. Lett.* 10, 680–689. [https://doi.org/10.1111/j.1461-](https://doi.org/10.1111/j.1461-0248.2007.01058.x)  
499 [0248.2007.01058.x](https://doi.org/10.1111/j.1461-0248.2007.01058.x)
- 500 Grace, J.B., Schoolmaster, D.R., Guntenspergen, G.R., Little, A.M., Mitchell, B.R., Miller, K.M., Schweiger,  
501 E.W., 2012. Guidelines for a graph-theoretic implementation of structural equation modeling.  
502 *Ecosphere* 3, art73. <https://doi.org/10.1890/ES12-00048.1>
- 503 Gruber, S., Pekrun, C., Möhring, J., Claupein, W., 2012. Long-term yield and weed response to conservation and  
504 stubble tillage in SW Germany. *Soil Tillage Res.* 121, 49–56. <https://doi.org/10.1016/j.still.2012.01.015>
- 505 Guimarães, R.M.L., Ball, B.C., Tormena, C.A., 2011. Guimarães, R. M. L., Ball, B. C., & Tormena, C. A.  
506 (2011). Improvements in the visual evaluation of soil structure. *Soil Use and Management*, 27(3), 395-  
507 403. *Soil Use Manag.* 395–403.
- 508 Hamel, C., Schellenberg, M.P., Hanson, K., Wang, H., 2007. Evaluation of the “bait-lamina test” to assess soil  
509 microfauna feeding activity in mixed grassland. *Appl. Soil Ecol.* 36, 199–204.  
510 <https://doi.org/10.1016/j.apsoil.2007.02.004>
- 511 Hartkamp, A.D., White, J.W., Rossing, W.A.H., van Ittersum, M.K., Bakker, E.J., Rabbinge, R., 2004. Regional  
512 application of a cropping systems simulation model: crop residue retention in maize production systems  
513 of Jalisco, Mexico. *Agric. Syst.* 82, 117–138. <https://doi.org/10.1016/j.agsy.2003.12.005>
- 514 He, J., Li, H., Rasaily, R.G., Wang, Q., Cai, G., Su, Y., Qiao, X., Liu, L., 2011. Soil properties and crop yields  
515 after 11 years of no tillage farming in wheat–maize cropping system in North China Plain. *Soil Tillage*  
516 *Res.* 113, 48–54. <https://doi.org/10.1016/j.still.2011.01.005>
- 517 Helling, B., Pfeiff, G., Larink, O., 1998. A comparison of feeding activity of collembolan and enchytraeid in  
518 laboratory studies using the bait-lamina test. *Appl. Soil Ecol.* 7, 207–212.  
519 [https://doi.org/10.1016/S0929-1393\(97\)00065-6](https://doi.org/10.1016/S0929-1393(97)00065-6)
- 520 Herrick, J.E., Whitford, W.G., de Soyza, A.G., Van Zee, J.W., Havstad, K.M., Seybold, C.A., Walton, M., 2001.  
521 Field soil aggregate stability kit for soil quality and rangeland health evaluations. *CATENA* 44, 27–35.  
522 [https://doi.org/10.1016/S0341-8162\(00\)00173-9](https://doi.org/10.1016/S0341-8162(00)00173-9)
- 523 Hobbs, P.R., 2007. Conservation agriculture: what is it and why is it important for future sustainable food  
524 production? *J. Agric. Sci.* 145, 127. <https://doi.org/10.1017/S0021859607006892>
- 525 Hobbs, P.R., Sayre, K., Gupta, R., 2008. The role of conservation agriculture in sustainable agriculture. *Philos.*  
526 *Trans. R. Soc. B Biol. Sci.* 363, 543–555. <https://doi.org/10.1098/rstb.2007.2169>
- 527 Hothorn, T., Bretz, F., Westfall, P., 2008. Simultaneous inference in general parametric models. *Biom. J.* 50,  
528 346–363. <https://doi.org/10.1002/bimj.200810425>
- 529 Husson, O., 2013. Redox potential (Eh) and pH as drivers of soil/plant/microorganism systems: a  
530 transdisciplinary overview pointing to integrative opportunities for agronomy. *Plant Soil* 362, 389–417.
- 531 Husson, O., Brunet, A., Babre, D., Charpentier, H., Durand, M., Sarthou, J.-P., 2018. Conservation agriculture  
532 systems alter the electrical characteristics (Eh, pH and EC) of four soil types in France. *Soil Tillage*  
533 *Res.* 176, 57–68.
- 534 Indoria, A.K., Rao, S., Sharma, K.L., Sammi Reddy, K., 2017. Conservation Agriculture - A Panacea to Improve  
535 Soil Physical Health. *Curr. Sci.* 112, 52. <https://doi.org/10.18520/cs/v112/i01/52-61>

536 ISEE [WWW Document], 2016. . www.isee.nc. URL [https://www.isee.nc/economie-entreprises/entreprises-](https://www.isee.nc/economie-entreprises/entreprises-secteurs-d-activites/agriculture-peche-aquaculture#analysesresultats-commentes-2)  
537 [secteurs-d-activites/agriculture-peche-aquaculture#analysesresultats-commentes-2](https://www.isee.nc/economie-entreprises/entreprises-secteurs-d-activites/agriculture-peche-aquaculture#analysesresultats-commentes-2) (accessed 8.28.20).

538 Keuskamp, J.A., Dingemans, B.J.J., Lehtinen, T., Sarneel, J.M., Hefting, M.M., 2013. Tea Bag Index: a novel  
539 approach to collect uniform decomposition data across ecosystems. *Methods Ecol. Evol.* 4, 1070–1075.  
540 <https://doi.org/10.1111/2041-210X.12097>

541 Kibblewhite, M.G., Ritz, K., Swift, M.J., 2008. Soil health in agricultural systems. *Philos. Trans. R. Soc. B Biol.*  
542 *Sci.* 363, 685–701. <https://doi.org/10.1098/rstb.2007.2178>

543 Lal, R., 2015a. Sequestering carbon and increasing productivity by conservation agriculture. *J. Soil Water*  
544 *Conserv.* 70, 55A-62A. <https://doi.org/10.2489/jswc.70.3.55A>

545 Lal, R., 2015b. Sequestering carbon and increasing productivity by conservation agriculture. *J. Soil Water*  
546 *Conserv.* 70, 55A-62A. <https://doi.org/10.2489/jswc.70.3.55A>

547 Lal, R., 2015c. Restoring soil quality to mitigate soil degradation. *Sustainability* 7, 5875–5895.  
548 <https://doi.org/10.3390/su7055875>

549 Lal, R., 2014. Soil conservation and ecosystem services. *Int. Soil Water Conserv. Res.* 2, 36–47.  
550 [https://doi.org/10.1016/S2095-6339\(15\)30021-6](https://doi.org/10.1016/S2095-6339(15)30021-6)

551 Le Cadre, E., Kinkondi, M., Koutika, L., Epron, D., Mareschal, L., 2018. Anionic exchange membranes, a  
552 promising tool to measure distribution of soil nutrients in tropical multispecific plantations. *Ecol. Indic.*  
553 254–256.

554 Lê, S., Josse, J., Husson, F., 2008. FactoMineR: an R package for multivariate analysis. *J. Stat. Softw.* 25, 1–18.  
555 <https://doi.org/10.18637/jss.v025.i01>

556 Lefcheck, J.S., 2016. PiecewiseSEM: Piecewise structural equation modelling in R for ecology, evolution, and  
557 systematics. *Methods Ecol. Evol.* 7, 573–579. <https://doi.org/10.1111/2041-210X.12512>

558 Lienhard, P., Tivet, F., Chabanne, A., Dequiedt, S., Lelièvre, M., Sayphoummie, S., Leudphanane, B., Prévost-  
559 Bouré, N.C., Séguy, L., Maron, P.-A., Ranjard, L., 2013. No-till and cover crops shift soil microbial  
560 abundance and diversity in Laos tropical grasslands. *Agron. Sustain. Dev.* 33, 375–384.  
561 <https://doi.org/10.1007/s13593-012-0099-4>

562 Losfeld, G., L’Huillier, L., Fogliani, B., Jaffré, T., Grison, C., 2015. Mining in New Caledonia: environmental  
563 stakes and restoration opportunities. *Environ. Sci. Pollut. Res.* 22, 5592–5607.  
564 <https://doi.org/10.1007/s11356-014-3358-x>

565 Machado, S., Petrie, S., Rhinhart, K., Ramig, R.E., 2008. Tillage effects on water use and grain yield of winter  
566 wheat and green pea in rotation. *Agron. J.* 100, 154–162. <https://doi.org/10.2134/agronj2006.0218>

567 Maynard, D.G., Kalra, Y.P., Crumbaugh, J.A., 1993. Nitrate and exchangeable ammonium nitrogen. *Soil Sampl.*  
568 *Methods Anal.* 1.

569 Meynard, J., David, G., 1992. Diagnostic de l’élaboration du rendement des cultures. *Cahiers agricultures* 1, 9–  
570 19.

571 Mitchell, J.P., Shrestha, A., Mathesius, K., Scow, K.M., Southard, R.J., Haney, R.L., Schmidt, R., Munk, D.S.,  
572 Horwath, W.R., 2017. Cover cropping and no-tillage improve soil health in an arid irrigated cropping  
573 system in California’s San Joaquin Valley, USA. *Soil Tillage Res.* 165, 325–335.  
574 <https://doi.org/10.1016/j.still.2016.09.001>

575 Murphy, S.D., Clements, D.R., Belaoussoff, S., Kevan, P.G., Swanton, C.J., 2006. Promotion of weed species  
576 diversity and reduction of weed seedbanks with conservation tillage and crop rotation. *Weed Sci.* 54,  
577 69–77. <https://doi.org/10.1614/WS-04-125R1.1>

578 Murray, W.E., 2001. The second wave of globalisation and agrarian change in the Pacific Islands. *J. Rural Stud.*  
579 17, 135–148. [https://doi.org/10.1016/S0743-0167\(00\)00042-5](https://doi.org/10.1016/S0743-0167(00)00042-5)

580 Muruganandam, S., Israel, D.W., Robarge, W.P., 2010. Nitrogen transformations and microbial communities in  
581 soil aggregates from three tillage systems. *Soil Sci. Soc. Am. J.* 74, 120–129.  
582 <https://doi.org/10.2136/sssaj2009.0006>

583 Myers, N., Mittermeier, R.A., Mittermeier, C.G., da Fonseca, G.A.B., Kent, J., 2000. Biodiversity hotspots for  
584 conservation priorities. *Nature* 403, 853–858. <https://doi.org/10.1038/35002501>

585 Naidu, V., 2010. Modernisation and development in the South Pacific, in: Jowitt, A., Cain, T.N. (Eds.), *Passage*  
586 *of Change, Law, Society and Governance in the Pacific.* ANU Press, pp. 7–32.

587 Nemergut, D.R., Cleveland, C.C., Wieder, W.R., Washenberger, C.L., Townsend, A.R., 2010. Plot-scale  
588 manipulations of organic matter inputs to soils correlate with shifts in microbial community  
589 composition in a lowland tropical rain forest. *Soil Biol. Biochem.* 42, 2153–2160.  
590 <https://doi.org/10.1016/j.soilbio.2010.08.011>

591 NeSmith, D.S., Ritchie, J.T., 1992. Maize (*Zea mays* L.) response to a severe soil water-deficit during grain-  
592 filling. *Field Crops Res.* 29, 23–35. [https://doi.org/10.1016/0378-4290\(92\)90073-I](https://doi.org/10.1016/0378-4290(92)90073-I)

593 Nichols, V., Verhulst, N., Cox, R., Govaerts, B., 2015. Weed dynamics and conservation agriculture principles:  
594 A review. *Field Crops Res.* 183, 56–68. <https://doi.org/10.1016/j.fcr.2015.07.012>

595 Obriot, F., Stauffer, M., Goubard, Y., Cheviron, N., Peres, G., Eden, M., Revallier, A., Vieublé-Gonod, L.,  
596 Houot, S., 2016. Multi-criteria indices to evaluate the effects of repeated organic amendment  
597 applications on soil and crop quality. *Agric. Ecosyst. Environ.* 232, 165–178.  
598 <https://doi.org/10.1016/j.agee.2016.08.004>

599 Palm, C., Blanco-Canqui, H., DeClerck, F., Gatere, L., Grace, P., 2014. Conservation agriculture and ecosystem  
600 services: An overview. *Agric. Ecosyst. Environ.* 187, 87–105.  
601 <https://doi.org/10.1016/j.agee.2013.10.010>

602 Pantoja, J.L., Woli, K.P., Sawyer, J.E., Barker, D.W., 2015. Corn nitrogen fertilization requirement and corn-  
603 soybean productivity with a rye cover crop. *Soil Sci. Soc. Am. J.* 79, 1482–1495.  
604 <https://doi.org/10.2136/sssaj2015.02.0084>

605 Pascault, N., Cécillon, L., Mathieu, O., Hénault, C., Sarr, A., Lévêque, J., Farcy, P., Ranjard, L., Maron, P.-A.,  
606 2010. In situ dynamics of microbial communities during decomposition of wheat, rape, and alfalfa  
607 residues. *Microb. Ecol.* 60, 816–828. <https://doi.org/DOI.10.1007/s00248-010-9705-7>

608 Pheap, S., Lefèvre, C., Thoumazeau, A., Leng, V., Boulakia, S., Koy, R., Hok, L., Lienhard, P., Brauman, A.,  
609 Tivet, F., 2019. Multi-functional assessment of soil health under Conservation Agriculture in  
610 Cambodia. *Soil Tillage Res.* 194, 104349. <https://doi.org/10.1016/j.still.2019.104349>

611 Pittelkow, C.M., Linnquist, B.A., Lundy, M.E., Liang, X., van Groenigen, K.J., Lee, J., van Gestel, N., Six, J.,  
612 Venterea, R.T., van Kessel, C., 2015. When does no-till yield more? A global meta-analysis. *Field*  
613 *Crops Res.* 183, 156–168. <https://doi.org/10.1016/j.fcr.2015.07.020>

614 Powlson, D.S., Stirling, C.M., Thierfelder, C., White, R.P., Jat, M.L., 2016. Does conservation agriculture  
615 deliver climate change mitigation through soil carbon sequestration in tropical agro-ecosystems? *Agric.*  
616 *Ecosyst. Environ.* 220, 164–174. <https://doi.org/10.1016/j.agee.2016.01.005>

617 Qian, P., Schoenau, J.J., 2002. Practical applications of ion exchange resins in agricultural and environmental  
618 soil research. *Can. J. Soil Sci.* 82, 9–21. <https://doi.org/10.4141/S00-091>

619 R Development Core Team, 2008. R: The R Project for Statistical Computing.

620 Ranaivoson, L., Naudin, K., Ripoché, A., Affholder, F., Rabeharisoa, L., Corbeels, M., 2017. Agro-ecological  
621 functions of crop residues under conservation agriculture. A review. *Agron. Sustain. Dev.* 37, 26.  
622 <https://doi.org/10.1007/s13593-017-0432-z>

623 Ranaivoson, L., Naudin, K., Ripoché, A., Rabeharisoa, L., Corbeels, M., 2019. Effectiveness of conservation  
624 agriculture in increasing crop productivity in low-input rainfed rice cropping systems under humid  
625 subtropical climate. *Field Crops Res.* 239, 104–113. <https://doi.org/10.1016/j.fcr.2019.05.002>

626 Rockström, J., Williams, J., Daily, G., Noble, A., Matthews, N., Gordon, L., Wetterstrand, H., DeClerck, F.,  
627 Shah, M., Steduto, P., 2017. Sustainable intensification of agriculture for human prosperity and global  
628 sustainability. *Ambio* 46, 4–17.

629 Rosolem, C.A., Li, Y., Garcia, R.A., 2016. Soil carbon as affected by cover crops under no-till under tropical  
630 climate. *Soil Use Manag.* 32, 495–503. <https://doi.org/10.1111/sum.12309>

631 Rovira, A.D., Greacen, E.L., 1957. The effect of aggregate disruption on the activity of microorganisms in the  
632 soil. *Aust. J. Agric. Res.* 8, 659–673. <https://doi.org/10.1071/ar9570659>

633 Rożen, A., Sobczyk, Ł., Liszka, K., Weiner, J., 2010. Soil faunal activity as measured by the bait-lamina test in  
634 monocultures of 14 tree species in the Siemianice common-garden experiment, Poland. *Appl. Soil Ecol.*  
635 45, 160–167. <https://doi.org/10.1016/j.apsoil.2010.03.008>

636 Ruan, L., Robertson, G.P., 2013. Initial nitrous oxide, carbon dioxide, and methane costs of converting  
637 conservation reserve program grassland to row crops under no-till vs. conventional tillage. *Glob.*  
638 *Change Biol.* 19, 2478–2489. <https://doi.org/10.1111/gcb.12216>

639 Rusinamhodzi, L., Corbeels, M., van Wijk, M.T., Rufino, M.C., Nyamangara, J., Giller, K.E., 2011. A meta-  
640 analysis of long-term effects of conservation agriculture on maize grain yield under rain-fed conditions.  
641 *Agron. Sustain. Dev.* 31, 657–673. <https://doi.org/10.1007/s13593-011-0040-2>

642 Scopel, E., Findeling, A., 2001. Conservation tillage impact on rainfed maize production in semi-arid zones of  
643 western Mexico. Importance of runoff reduction, in: *Conservation Agriculture, a Worldwide*  
644 *Challenge : Ist World Congress on Conservation Agriculture.*

645 Scopel, E., Triomphe, B., Affholder, F., Da Silva, F.A.M., Corbeels, M., Xavier, J.H.V., Lahmar, R., Recous, S.,  
646 Bernoux, M., Blanchart, E., de Carvalho Mendes, I., De Tournonnet, S., 2013. Conservation agriculture  
647 cropping systems in temperate and tropical conditions, performances and impacts. A review. *Agron.*  
648 *Sustain. Dev.* 33, 113–130. <https://doi.org/10.1007/s13593-012-0106-9>

649 Ségué, L., Bouzinac, S., Husson, O., 2006. Direct-seeded tropical soil systems with permanent soil cover:  
650 learning from Brazilian experience, in: *Biological Approaches to Sustainable Soil Systems.* CRC Press,  
651 pp. 323–342.

652 Sithole, N.J., Magwaza, L.S., Mafongoya, P.L., 2016. Conservation agriculture and its impact on soil quality and  
653 maize yield: A South African perspective. *Soil Tillage Res.* 162, 55–67.  
654 <https://doi.org/10.1016/j.still.2016.04.014>



655 Teasdale, J.R., Brandsaeter, L.O., Calegari, A., Neto, F.S., Upadhyaya, M.K., Blackshaw, R.E., 2007. Cover  
656 crops and weed management, in: *Non-Chemical Weed Management: Principles, Concepts and*  
657 *Technology*. pp. 49–64.

658 Thierfelder, C., Matemba-Mutasa, R., Rusinamhodzi, L., 2015. Yield response of maize (*Zea mays* L.) to  
659 conservation agriculture cropping system in Southern Africa. *Soil Tillage Res.* 146, 230–242.  
660 <https://doi.org/10.1016/j.still.2014.10.015>

661 Thierfelder, C., Wall, P.C., 2012. Effects of conservation agriculture on soil quality and productivity in  
662 contrasting agro-ecological environments of Zimbabwe. *Soil Use Manag.* 28, 209–220.  
663 <https://doi.org/10.1111/j.1475-2743.2012.00406.x>

664 Thoumazeau, A., Bessou, C., Renevier, M.-S., Panklang, P., Puttaso, P., Peerawat, M., Heepngoen, P., Polwong,  
665 P., Koonklang, N., Sdoodee, S., Chantuma, P., Lawongsa, P., Nimkingrat, P., Thaler, P., Gay, F.,  
666 Brauman, A., 2019a. Biofunctool®: a new framework to assess the impact of land management on soil  
667 quality. Part B: investigating the impact of land management of rubber plantations on soil quality with  
668 the Biofunctool® index. *Ecol. Indic.* 97, 429–437. <https://doi.org/10.1016/j.ecolind.2018.10.028>

669 Thoumazeau, A., Bessou, C., Renevier, M.-S., Trap, J., Marichal, R., Mareschal, L., Decaëns, T., Bottinelli, N.,  
670 Jaillard, B., Chevallier, T., Suvannang, N., Sajjaphan, K., Thaler, P., Gay, F., Brauman, A., 2019b.  
671 Biofunctool®: a new framework to assess the impact of land management on soil quality. Part A:  
672 concept and validation of the set of indicators. *Ecol. Indic.* 97, 100–110.  
673 <https://doi.org/10.1016/j.ecolind.2018.09.023>

674 Thoumazeau, A., Gay, F., Alonso, P., Suvannang, N., Phongjinda, A., Panklang, P., Chevallier, T., Bessou, C.,  
675 Brauman, A., 2017. SituResp®: A time- and cost-effective method to assess basal soil respiration in the  
676 field. *Appl. Soil Ecol.* 121, 223–230. <https://doi.org/10.1016/j.apsoil.2017.10.006>

677 Tilman, D., Balzer, C., Hill, J., Befort, B.L., 2011. Global food demand and the sustainable intensification of  
678 agriculture. *Proc. Natl. Acad. Sci.* 108, 20260–20264. <https://doi.org/10.1073/pnas.1116437108>

679 Tivet, F., de Moraes Sá, J.C., Lal, R., Briedis, C., Borszowski, P.R., dos Santos, J.B., Farias, A., Eurich, G.,  
680 Hartman, D. da C., Nadolny Junior, M., Bouzinac, S., Séguy, L., 2013. Aggregate C depletion by  
681 plowing and its restoration by diverse biomass-C inputs under no-till in sub-tropical and tropical  
682 regions of Brazil. *Soil Tillage Res.* 126, 203–218. <https://doi.org/10.1016/j.still.2012.09.004>

683 Törne, E. von, 1990. Assessing feeding activities of soil-living animals. I. Bait-lamina-tests. *Pedobiologia* 34,  
684 89–101.

685 Tóth, Z., Hornung, E., Báldi, A., 2018. Effects of set-aside management on certain elements of soil biota and  
686 early stage organic matter decomposition in a High Nature Value Area, Hungary. *Nat. Conserv.* 29, 1–  
687 26. <https://doi.org/10.3897/natureconservation.29.24856>

688 Triplett, G.B., Dick, W.A., 2008. No-tillage crop production: A revolution in agriculture! *Agron. J.* 100, S-153-  
689 S-165. <https://doi.org/10.2134/agronj2007.0005c>

690 van der Velde, M., Green, S.R., Vanclooster, M., Clothier, B.E., 2007. Sustainable development in small island  
691 developing states: Agricultural intensification, economic development, and freshwater resources  
692 management on the coral atoll of Tongatapu. *Ecol. Econ.* 61, 456–468.  
693 <https://doi.org/10.1016/j.ecolecon.2006.03.017>

694 van Gestel, C.A.M., Kruidenier, M., Berg, M.P., 2003. Suitability of wheat straw decomposition, cotton strip  
695 degradation and bait-lamina feeding tests to determine soil invertebrate activity. *Biol. Fertil. Soils* 37,  
696 115–123. <https://doi.org/10.1007/s00374-002-0575-0>

697 van Heemst, H.D.J., 1985. The influence of weed competition on crop yield. *Agric. Syst.* 18, 81–93.  
698 [https://doi.org/10.1016/0308-521X\(85\)90047-2](https://doi.org/10.1016/0308-521X(85)90047-2)

699 Verhulst, N., François, I.M., Govaerts, B., 2010. Conservation agriculture, improving soil quality for sustainable  
700 production systems. *Advances in soil science: food security and soil quality* 137–208.

701 Virto, I., Barré, P., Burlot, A., Chenu, C., 2012. Carbon input differences as the main factor explaining the  
702 variability in soil organic C storage in no-tilled compared to inversion tilled agrosystems.  
703 *Biogeochemistry* 108, 17–26. <https://doi.org/10.1007/s10533-011-9600-4>

704 Weil, R.R., Islam, K.R., Stine, M.A., Gruver, J.B., Samson-Liebig, S.E., 2003. Estimating active carbon for soil  
705 quality assessment: A simplified method for laboratory and field use. *Am. J. Altern. Agric.* 18, 3–17.  
706 <https://doi.org/10.1079/AJAA2003003>

707 Wezel, A., Soldat, V., 2009. A quantitative and qualitative historical analysis of the scientific discipline of  
708 agroecology. *Int. J. Agric. Sustain.* 7, 3–18. <https://doi.org/10.3763/ijas.2009.0400>

709 Zuber, S.M., Behnke, G.D., Nafziger, E.D., Villamil, M.B., 2017. Multivariate assessment of soil quality  
710 indicators for crop rotation and tillage in Illinois. *Soil Tillage Res.* 174, 147–155.  
711 <https://doi.org/10.1016/j.still.2017.07.007>

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713

714 **Figure captions**

715

716 **Fig. 1** Principal component analysis of the effects of the cropping system on soil health.

717 **a** Variables factor map. POXC: Permanganate OXidizable Carbon, SituResp<sup>®</sup>: basal soil respiration, Laminas:  
718 lamina bait degradation, GTB: fraction of Green Tea Bag decomposed, VESS: Visual Evaluation of Soil  
719 Structure, Beerkan: water infiltration, AggSoil: soil aggregate water stability, AEMNO<sub>3</sub>: nitrate evaluated with  
720 anion exchange membrane, NNH<sub>4</sub>, NNO<sub>3</sub>: available ammonium and nitrate.

721 **b** Individual factor map. CT: Conventional Tillage, CA: Conservation Agriculture with direct seeding in Dead  
722 Mulch (CA-DM) or Living Mulch (CA-LM).

723 *Note: AggSoil median score and 0-2cm depth laminas score were used to run the PCA.*

724

725 **Fig. 2** Biofunctool<sup>®</sup> Soil Health Index (SHI) per treatment. CT: Conventional Tillage, CA: Conservation  
726 Agriculture with direct seeding in Dead Mulch (CA-DM) or Living Mulch (CA-LM); n=9 for each treatment.  
727 Standard error of the index is given for each treatment. Different letters indicate significant differences at  $P<0.05$   
728 according to Tukey's test.

729

730 **Fig. 3** Structural Equation Modelling (SEM) linking the cropping system, soil health, and cropping system  
731 performance (Fisher's  $C=14.76$ ,  $df=12$ ,  $P=0.26$ ). CT: Conventional Tillage, CA: Conservation Agriculture  
732 systems (direct seeding in dead mulch and living mulch not differentiated): characterised by the aboveground  
733 biomass of the cover crops and the soil management practices. Weeds: Weed cumulative aboveground dry  
734 matter during the crop cycle, Maize Yield: grain yield, TKW: Maize Thousand Kernel Weight. The arrows  
735 indicate unidirectional relationships between the variables (direct effects of one variable on the others). Green  
736 arrows indicate significant positive effects, red arrows indicate significant negative effects, and grey arrows  
737 indicate non-significant relationships at  $P=0.05$ . Path coefficients are indicated adjacent to the corresponding  
738 arrows. Arrow widths are proportional to the path coefficients.

739

740 **Table 1** Biofunctool<sup>®</sup> indicators of soil carbon transformation per treatment. CT: Conventional Tillage, CA:  
 741 Conservation Agriculture with direct seeding in Dead Mulch (CA-DM) or Living Mulch (CA-LM). POXC:  
 742 Permanganate OXidizable Carbon, SituResp<sup>®</sup>: basal soil respiration, Laminas: lamina bait degradation, GTB:  
 743 fraction of Green Tea Bag decomposed. The analysis was conducted in the 0-10 cm layer, except for laminas (in  
 744 the 0-2 cm layer) and GTB (at a depth of 8 cm); n=9 for each treatment; sd: standard deviation. Different letters  
 745 indicate significant differences according to Tukey's test ( $P<0.05$ ).  
 746

Treatment	Carbon transformation							
	POXC		SituResp <sup>®</sup>		Laminas		GTB	
	(mg <sub>C</sub> kg <sub>soil</sub> <sup>-1</sup> )		(Absorbance difference)		(Score)		(Score)	
	mean	sd	mean	sd	mean	sd	mean	sd
CT	1071 <b>a</b>	27	0.87 <b>a</b>	0.05	4.91 <b>a</b>	4.0	0.43 <b>a</b>	0.02
CA-DM	1124 <b>b</b>	27	0.96 <b>b</b>	0.06	8.71 <b>b</b>	4.3	0.46 <b>b</b>	0.03
CA-LM	1122 <b>b</b>	34	0.95 <b>b</b>	0.06	7.17 <b>b</b>	4.0	0.45 <b>ab</b>	0.02
ANOVA	$P<0.001$		$P<0.001$		$P<0.001$		$P<0.001$	

747  
 748

749 **Table 2** Biofunctool® indicators of soil structure maintenance per treatment. CT: Conventional Tillage, CA:  
 750 Conservation Agriculture with direct seeding in Dead Mulch (CA-DM) or Living Mulch (CA-LM). VESS:  
 751 Visual Evaluation of Soil Structure, Beerkan: water infiltration, AggSoil: soil aggregate water stability. The  
 752 analysis was made in the 0-10 cm layer, except for VESS (in the 0-30 cm layer); n=9 for each treatment; sd:  
 753 standard deviation. Different letters indicate significant differences according to Tukey's test.  
 754

Treatment	Structure maintenance					
	VESS (Score)		Beerkan (mL min <sup>-1</sup> )		AggSoil (Score)	
	mean	sd	mean	sd	median	sd
CT	2.11 <b>b</b>	0.4	93.4 <b>a</b>	20.5	1.22 <b>a</b>	0.4
CA-DM	1.45 <b>a</b>	0.3	176.5 <b>b</b>	71.5	2.00 <b>b</b>	0.8
CA-LM	1.28 <b>a</b>	0.3	226.0 <b>b</b>	117.3	2.15 <b>b</b>	0.9
ANOVA	<i>P</i> <0.001		<i>P</i> <0.001		<i>P</i> <0.001	

755  
 756

757 **Table 3** Biofunctool® indicators of soil nutrient cycling per treatment. CT: Conventional Tillage, CA:  
 758 Conservation Agriculture with direct seeding in Dead Mulch (CA-DM) or Living Mulch (CA-LM). AEM-NO<sub>3</sub><sup>-</sup>:  
 759 nitrate evaluated with anion exchange membrane, N-NH<sub>4</sub><sup>+</sup>, N-NO<sub>3</sub><sup>-</sup>: available ammonium and nitrate. The  
 760 analysis was conducted in the 0-10 cm layer, except for AEM-NO<sub>3</sub><sup>-</sup> (at a depth of 8 cm); n=9 for each treatment  
 761 except for N-NH<sub>4</sub><sup>+</sup> and N-NO<sub>3</sub><sup>-</sup> where n=3 per treatment (no internal replicates); sd: standard deviation. Different  
 762 letters indicate significant differences according to Tukey's test.

763

Treatment	Nutrient cycling					
	AEM-NO <sub>3</sub> <sup>-</sup>		N-NO <sub>3</sub> <sup>-</sup>		N-NH <sub>4</sub> <sup>+</sup>	
	(μg <sub>N-NO<sub>3</sub><sup>-</sup></sub> cm <sup>-2</sup> d <sup>-1</sup> )		(mg kg <sup>-1</sup> )		(mg kg <sup>-1</sup> )	
	mean	sd	mean	sd	mean	sd
CT	20.4 <b>b</b>	6.4	10.9 <b>ns</b>	4.1	2.6 <b>a</b>	0.3
CA-DM	10.5 <b>a</b>	4.0	14.7 <b>ns</b>	2.2	6.1 <b>b</b>	0.2
CA-LM	9.8 <b>a</b>	5.0	14.7 <b>ns</b>	3.2	4.7 <b>ab</b>	1.3
ANOVA	<i>P</i> <0.001		<i>P</i> =0.4		<i>P</i> <0.001	

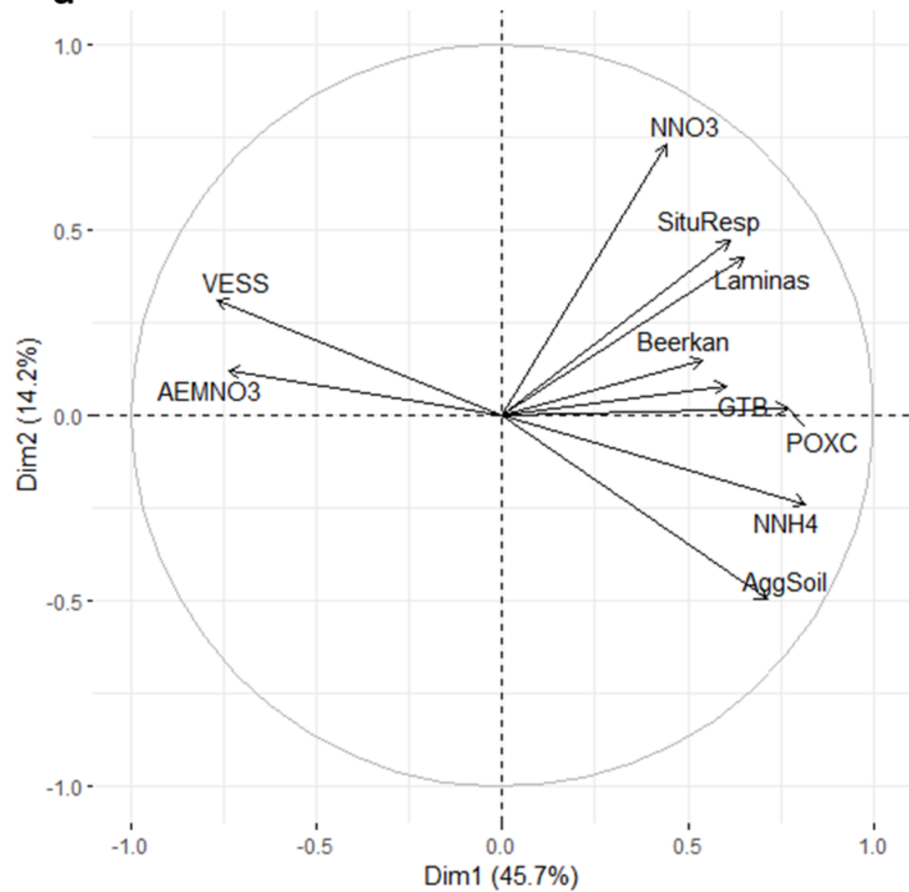
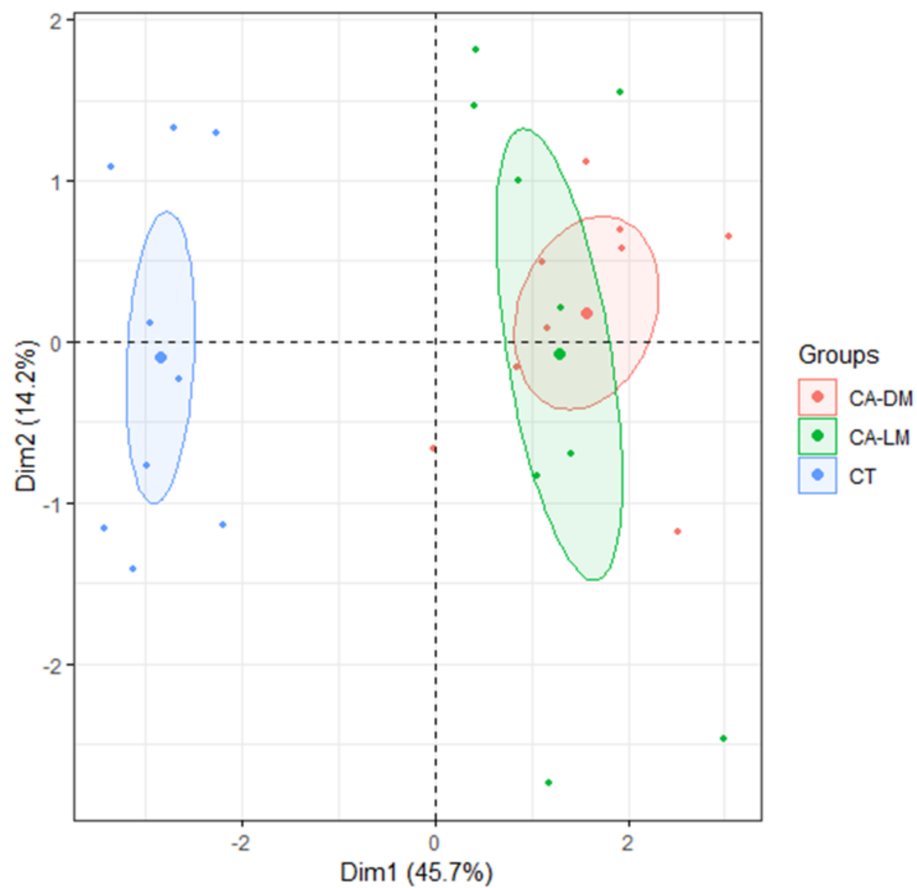
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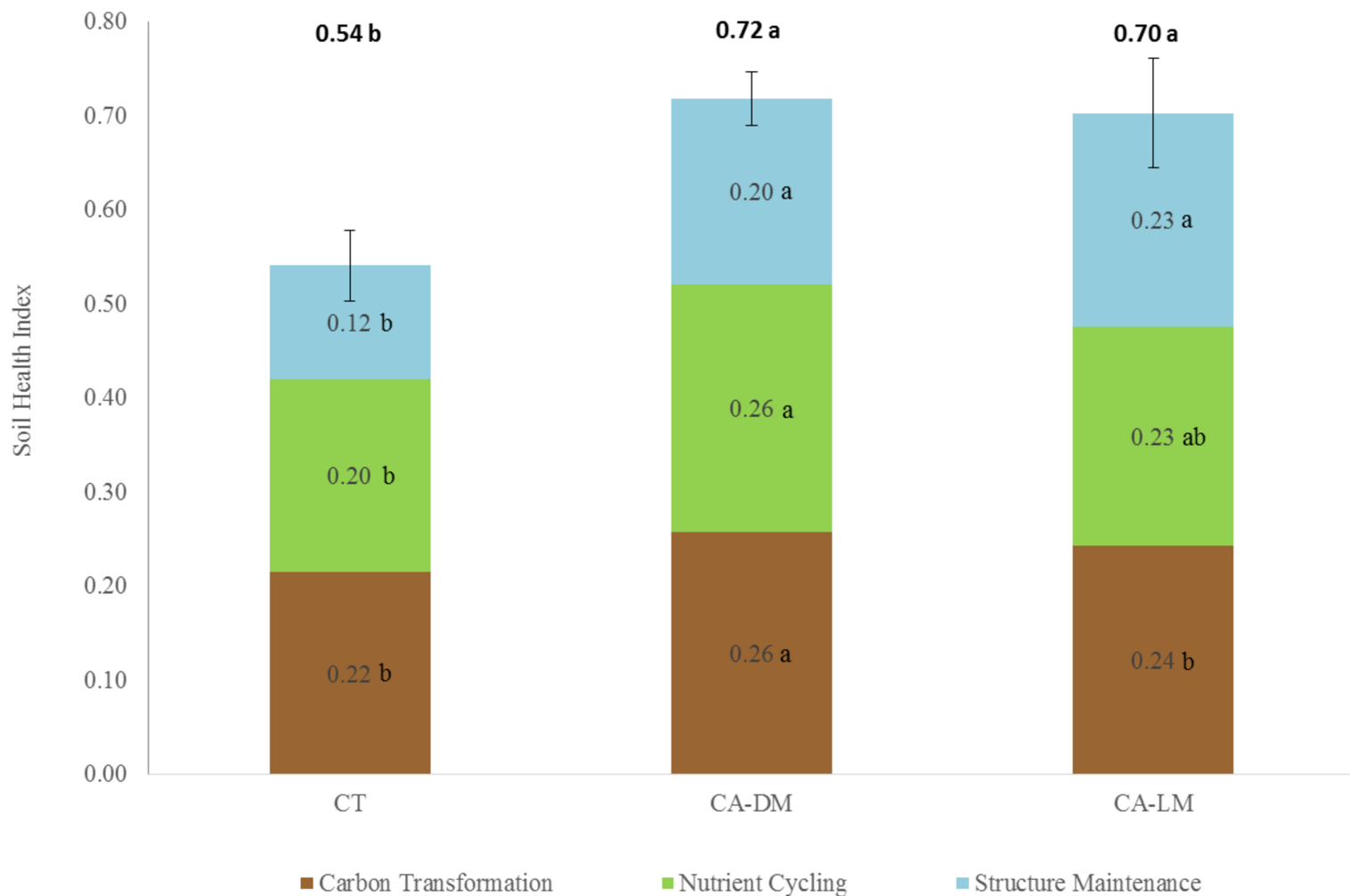
766 **Table 4** Cropping system performance indicators per treatment. CT: Conventional Tillage, CA: Conservation  
 767 Agriculture with direct seeding in Dead Mulch (CA-DM) or Living Mulch (CA-LM). Weeds: Weed cumulative  
 768 aboveground dry matter during crop cycle, Maize density: Maize plant population, Kernel weight: Total kernel  
 769 weight per maize ear, TKW: Maize Thousand Kernel Weight, Maize yield: grain yield; n=9 for each treatment;  
 770 sd: standard deviation. Different letters indicate significant differences according to Tukey's test.

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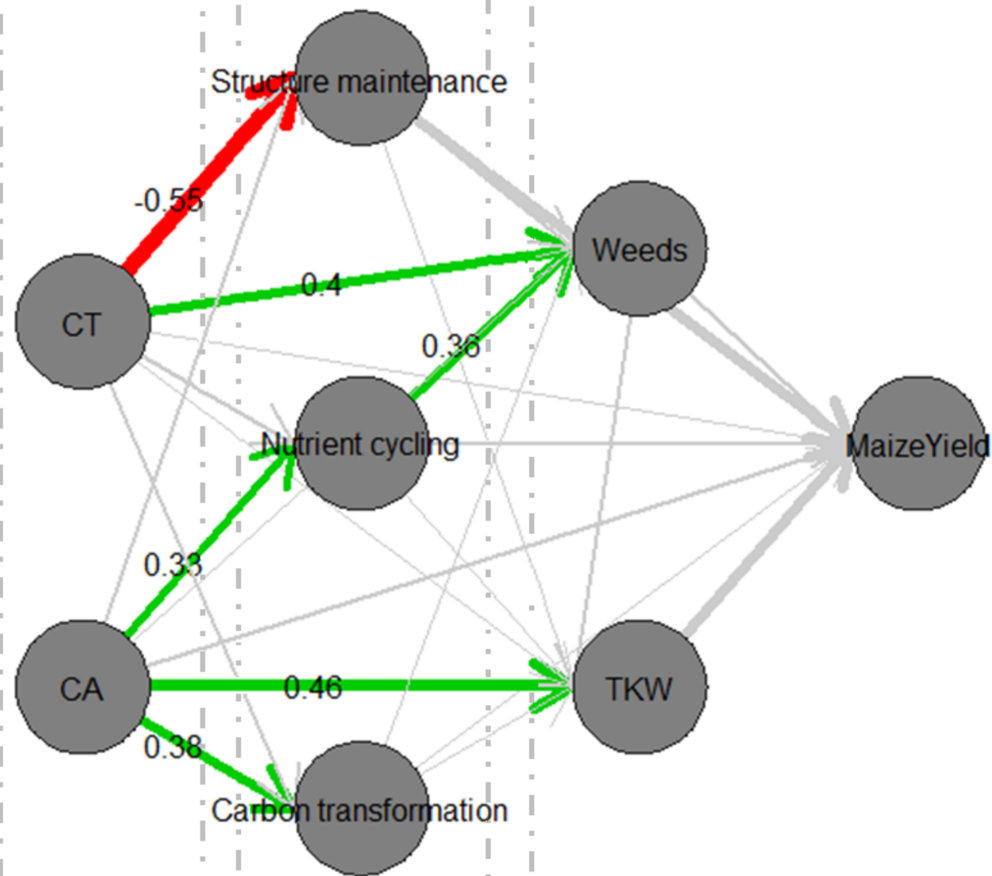
Treatment	Weeds		Maize density		Kernel weight		TKW		Yield	
	(t <sub>cumulative DM ha<sup>-1</sup></sub> )		(plants m <sup>-2</sup> )		(g ear <sup>-1</sup> )		(g)		(t ha <sup>-1</sup> )	
	mean	sd	mean	sd	mean	sd	mean	sd	mean	sd
CT	1.4 <b>c</b>	0.7	9.0 <b>b</b>	0.4	107.8 <b>a</b>	21.0	355.1 <b>a</b>	16.3	9.7 <b>a</b>	2.0
CA-DM	0.7 <b>b</b>	0.3	8.0 <b>a</b>	1.1	158.6 <b>b</b>	25.5	388.2 <b>b</b>	7.5	12.7 <b>b</b>	2.9
CA-LM	0.2 <b>a</b>	0.3	10.3 <b>c</b>	0.5	125.8 <b>a</b>	18.2	364.2 <b>a</b>	12.9	12.9 <b>b</b>	1.8
ANOVA	<i>P</i> <0.001		<i>P</i> <0.001		<i>P</i> <0.001		<i>P</i> <0.001		<i>P</i> <0.001	

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**a****b**







**Cropping system**

**Soil health**

**Cropping system performance**