

Potential favourable health effects of some dietary uncommon fatty acids

Charles Coudray, Erwann Durand, Laurence Balas, Ariane Sultan, François Casas, Christine Feillet-coudray

► **To cite this version:**

Charles Coudray, Erwann Durand, Laurence Balas, Ariane Sultan, François Casas, et al.. Potential favourable health effects of some dietary uncommon fatty acids. OCL Oilseeds and fats crops and lipids, EDP, 2021, 28, 10.1051/ocl/2021028 . hal-03321938

HAL Id: hal-03321938

<https://hal.inrae.fr/hal-03321938>

Submitted on 18 Aug 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Potential favourable health effects of some dietary uncommon fatty acids

Charles Coudray¹, Erwann Durand^{2,3}, Laurence Balas⁴, Ariane Sultan^{5,6}, François Casas¹ and Christine Feillet-Coudray^{1,*}

¹ DMEM, Univ Montpellier, INRAE Montpellier, Montpellier, France

² CIRAD, UMR Qualisud, 34398 Montpellier, France

³ Qualisud, Univ Montpellier, Avignon Université, CIRAD, Institut Agro, IRD, Université de La Réunion, Montpellier, France

⁴ Institut des Biomolécules Max Mousseron, Univ Montpellier, UMR 5247, CNRS, ENSCM, Montpellier, France

⁵ Université de Montpellier, PhyMedExp, Inserm, CNRS, 34000 Montpellier, France

⁶ Montpellier University Hospital, Department of Endocrinology-Diabetes-Nutrition, 34295 Montpellier Cedex 5, France

Received 3 May 2021 – Accepted 13 July 2021

Abstract – In addition to the major fatty acids widely studied, our diet contains many bioactive fatty acids less frequently investigated such as n-3 docosapentaenoic acid (n-3 DPA), natural *trans* fatty acids, conjugated fatty acids (CLAs), furan fatty acids (FuFAs), branched chain fatty acids (BCFAs) and fatty acid esters of hydroxyl fatty acids (FAHFAs). Many of them may have beneficial health effects, particularly in the prevention of cardiovascular diseases, inflammation and metabolic disorders such as diabetes. This review aims to give a brief overview of the current knowledge on these lipids. Thus, information about biosynthesis, food and tissue content, daily intake, biological and potential health effects of these fatty acids is provided.

Keywords: bioactive lipids / biosynthesis / food content / dietary intake / biological and health effects

Résumé – Effets potentiellement bénéfiques sur la santé de certains acides gras peu courants de l'alimentation. Parallèlement aux acides gras prépondérants qui sont largement étudiés, notre alimentation contient de nombreux acides gras bioactifs moins fréquemment considérés tels que l'acide docosapentaénoïque n-3 (n-3 DPA), des acides gras *trans* naturels, des acides gras conjugués (CLAs), des acides gras furaniques (FuFAs), des acides gras à chaîne ramifiée (BCFAs) et des esters d'acides gras hydroxylés (FAHFAs). Nombreux d'entre eux peuvent avoir des effets bénéfiques sur la santé, notamment dans la prévention des maladies cardiovasculaires, de l'inflammation et des troubles métaboliques tels que le diabète. Cette revue vise à donner un bref aperçu des connaissances actuelles sur ces lipides. Ainsi, des informations sur la biosynthèse, les teneurs tissulaires et dans les aliments, l'apport alimentaire quotidien, les effets biologiques et les effets santé potentiels de ces acides gras sont rapportés.

Mots clés : lipides bioactifs / biosynthèse / composition des aliments / apport alimentaire / effets biologiques et sur la santé

Food plays an important role not only for an optimal growth and development but also in the maintenance of a good health. While the role of nutrients and micronutrients and the needs of the organism in these components are widely studied, there are many molecules in our diet whose roles are yet to be investigated more extensively. These include many bioactive lipids with potential beneficial health effects such as n-3 docosapentaenoic acid (n-3 DPA), natural *trans* fatty acids, conjugated fatty acids (CLAs), furan fatty acids (FuFAs), branched chain fatty acids (BCFAs) and fatty acid

hydroxylated fatty acid esters (FAHFAs). This review aims to give a brief overview of the current knowledge regarding these lipids. We present their structure and their biosynthesis (Fig. 1 for precursors and structure of these lipids), their content in food and their daily intake (Tab. 1), their tissue content, their biological and potential health effects. Some dietary lipids can be toxic (cyclopropene-containing lipids, mono-unsaturated long-chain fatty acids as erucic acid, *trans*-unsaturated fatty acids from industrial hydrogenated fats, and lipid peroxides) (Gurr *et al.*, 2002). However, except *trans* fatty acids of industrial origin, they are not discussed in this article.

*Correspondence: christine.coudray@inrae.fr

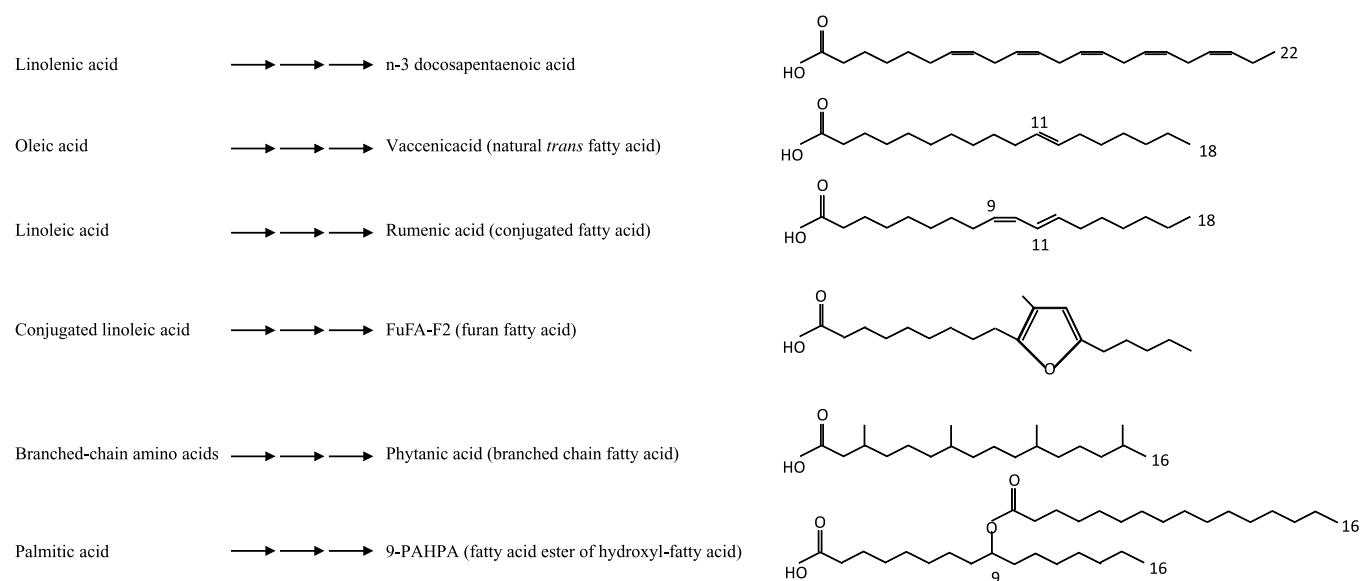


Fig. 1. Examples of some uncommon fatty acids and their precursors.

Table 1. Daily intake and main dietary sources of some bioactive lipids.

	Daily intake	Main dietary sources
n-3 DPA	10 to 100 mg/d	Seafood
Natural <i>trans</i> fatty acids	0.5 to 0.9% total energy intake	Dairy products Beef and lamb meat
CLAs	Hundreds of milligrams/d	Dairy products Beef and lamb meat
FuFAs	Tens of milligrams/d	Fish
BCFAs	About 400 mg/d	Dairy products Meat and fish
FAHFAs	No data available	Animal products Plant products and milk

DPA: docosapentaenoic acid; CLAs: conjugated fatty acids; FuFAs: furan fatty acids; BCFAs: branched chain fatty acids; FAHFAs: fatty acid esters of hydroxyl fatty acids.

1 The n-3 docosapentaenoic acid DPA

1.1 Structure and biosynthesis

n-3 DPA (C22:5 n-3) belongs to the n-3 polyunsaturated fatty acid (PUFA) family. It is also called clupanodonic acid. It is an intermediate between eicosapentaenoic acid (EPA, C20:5 n-3) and docosahexaenoic acid (DHA, C22:6 n-3) in the conversion pathway of n-3 PUFA from α -linolenic acid (ALA, C18:3 n-3). n-3 DPA as a reservoir is metabolized into DHA, and further retro-converted back to EPA (Guo *et al.*, 2020).

1.2 Food content and daily intake

n-3 DPA is present in numerous foods in notable quantities, in particular in seafood (1 to 5% of total fatty acids in fish, i.e. up to 1/3 of EPA or DHA levels taken individually) as well as in human breast milk (0.2% of total fatty acids) (Drouin, 2018). For example, fish oils such as menhaden or sardine oil contain in percentage of total fatty acids: 10–13% EPA, 2–5% DPA and

9–11% DHA. Atlantic salmon contains 0.3 g of EPA, 0.3 g of DPA and 1.1 g of DHA per 100 g. (Kaur *et al.*, 2016).

The estimated average consumption of n-3 DPA is between 10 and 106 mg/d in Western countries and Japan (Richter *et al.*, 2019). n-3 DPA may represent a significant proportion of the total long-chain n-3 fatty acid intake, depending on the population, up to 30% of average total long-chain n-3 fatty acid intake in some cases (Richter *et al.*, 2019). However, the digestibility of n-3 DPA in rodents (digestibility reflecting net absorption in the digestive tract after enzymatic hydrolysis by digestive enzymes and microflora of the gastrointestinal tract) is lower than that of EPA and DHA (Drouin *et al.*, 2019a).

1.3 Tissue content and biological functions

The majority of the tissues present n-3 DPA levels in the order of 5% compared to DHA levels (Ghasemi Fard *et al.*, 2021). In the brain, n-3 DPA is the second n-3 PUFA found, although at levels 70 times lower than DHA (Drouin *et al.*, 2019b).

n-3 DPA is a source of EPA and to a lesser extent DHA in major metabolic tissues (liver, heart, lung, spleen and kidney), two fatty acids with numerous known health benefits. Moreover, n-3 DPA is the precursor of many major lipid mediators (protectins, resolvins, maresins, isoprostanooids), involved in the pro-resolution of inflammation, with specific effects compared to other n-3 PUFAs (Ghasemi Fard *et al.*, 2021).

1.4 Health effects

Low commercial availability of n-3 DPA in sufficient quantity, at high purity and at an affordable price (Drouin, 2018; Drouin *et al.*, 2019b) has limited its studies in rodents and humans. Thus, only 11 studies in animals and 2 studies in humans with n-3 DPA in pure form have been reported (Ghasemi Fard *et al.*, 2021). The effects of n-3 DPA on lipid parameters associated with the prevention of cardiovascular diseases are the most documented (anti-inflammatory properties, inhibition of cytokine synthesis, reduction of thrombosis and inhibition of atherosclerosis, ...) (von Schacky and Harris, 2018). The health effects of n-3 DPA could be both independent and shared with EPA and DHA (Richter *et al.*, 2019). It is important to note that the n-3 DPA could contribute to increasing the n-3 fatty acid status, as n-3 DPA is more present in meat than EPA or DHA and while the sources of fatty fish are limited; therefore, n-3 DPA could be helpful to maintain a suitable n-6/n-3 ratio which is the indicator of a preventive diet for the control of non-communicable diseases (Drouin *et al.*, 2019b). Drouin *et al.* recently published a comprehensive review on n-3 DPA (Drouin *et al.*, 2019b).

2 The *trans* fatty acids

2.1 Structure and biosynthesis

Trans fatty acids are fatty acids that have at least one double bond in the *trans* configuration while most of the naturally occurring unsaturated fatty acids contain *cis*-double bonds. Naturally occurring *trans* fatty acids are mostly monounsaturated fatty acids, mainly *trans* C18:1 n-7 (vaccenic acid), *trans*-C16:1 n-7 and all isomers of oleic acid (18:1 n-9) (Leray, 2013). There are also di-unsaturated *trans* fatty acids, derived from linoleic acid (18:2 n-6) or tri-unsaturated *trans* fatty acids, derived from linolenic acid (18:3 n-3). Conjugated linoleic acids (two conjugated double bonds, one of which is in a *trans* configuration) are discussed in Section 3.

Naturally occurring *trans* fatty acids come from a bacterial isomerisation of fatty acids in the digestive tract of ruminants (Leray, 2013). The eukaryotes are unable to synthesize them, however, it is possible that they can be synthesised by the action of intestinal microbiota on dietary fatty acids. *Trans* fatty acids of industrial origin are formed during the partial hydrogenation of vegetable or fish oils. Thermal treatments (frying, cooking, ...) can also produce *trans* fatty acids of (poly)unsaturated oils and fats.

2.2 Food content and daily intake

Trans fatty acids of natural origin are found in dairy products and beef and sheep meats. Butter contains 3 to 7 g/100 g of

natural *trans* fatty acids, cheeses from 1.3 to 2 g/100 g, whole milks around 0.15 g/100 g, vegetable oils between 0.5 and 2 g/100 g, beef and sheep meat from 0.1 to 0.5 g/100 g (Leray, 2013). *Trans* fatty acids from industrial process are used by the food industry as stabilizers and preservatives. Thus, they are found in many processed food products such as pastries, pizzas, quiches (Afssa, 2005).

According to the results of the INCA2 survey, the average and 95th percentile intake of the total *trans* fatty acids in the French population was estimated at 1–1.5% of total energy intake, regardless of age and sex (Afssa, 2009). More than half is of natural origin (0.5–0.9%), thus below the ANSES recommendations to limit the total *trans* fatty acid intake to less than 2% of the total energy intake (Afssa, 2009).

2.3 Tissue content and biological functions

In the 2000s, the total amount of *trans* fatty acids was $2.32 \pm 0.50\%$ of the total fatty acids in adipose tissue of French women (Boue *et al.*, 2000) and mean adipose tissue levels were lower in many European countries than in the USA (Arab, 2003).

The *trans* configuration impacts the physicochemical and functional properties of monounsaturated fatty acids. It makes them closer to the properties of the corresponding saturated fatty acids. Thus, high amounts of *trans* fatty acids may decrease the membrane fluidity and increase the oxidative stress (Leray, 2013), and may induce inflammation and apoptosis of the cells (Qiu *et al.*, 2018).

2.4 Health effects

A number of epidemiological studies have shown a relationship between *trans* fatty acid intake and cardiovascular diseases. Controlled feeding studies suggest that dietary *trans* fatty acids raise serum cholesterol concentrations to a very similar extent as saturated fatty acids (Gurr *et al.*, 2002; Leray, 2013). Numerous studies show that the risks of cardiovascular diseases with dietary *trans* fatty acids are attributable to industrial *trans* fatty acid (Oteng and Kersten, 2020). In contrast, no increase in cardiovascular risk has been observed with the consumption of naturally occurring *trans* fatty acids at the current consumption levels (Guillocheau *et al.*, 2019). Studies on cells, rodents and humans suggest physiological benefits on inflammation, type 2 diabetes and obesity (Guillocheau *et al.*, 2019), without knowing whether these naturally occurring *trans* fatty acids act directly or through their metabolites (Guillocheau *et al.*, 2019). Guillocheau *et al.* recently published a comprehensive review on natural *trans* fatty acids (Guillocheau *et al.*, 2019).

3 The conjugated linoleic acid CLAs

3.1 Structure and biosynthesis

Conjugated linoleic acid (CLA) is a collective term for a mixture of positional and geometrical isomers of linoleic acid (LA, C18:2 n-6) containing conjugated double bonds. Some linoleic acid isomers have conjugated doubles with one of them (at least) in a *trans* configuration (Leray, 2013).

Rumenic acid (9-*cis*, 11-*trans*18:2 n-6) is the most abundant CLA. There are other conjugated fatty acids such as conjugated linolenic acids (CLNAs), conjugated eicosapentaenoic acids (CEPAs) and conjugated docosahexaenoic acid (CDHAs) (Leray, 2013).

CLAs are produced naturally in the rumen of ruminant animals by fermentative bacteria (*Butyrivibrio fibrisolvens*) which isomerize linoleic acid into CLAs. Ruminants also synthesize CLAs by delta9-desaturase and from *trans*-11 18:1 (Leray, 2013).

3.2 Food content and daily intake

Of the possible isomers of CLA, about 20 have been identified in foods (Leray, 2013). Beef meat contains up to 120 mg/100 g of CLA and lamb meat about 80 mg/100 g. The main isomer present in milk fat is the rumenic acid, which accounts for 80% to 90% of the total CLA. Rumenic acid represents up to 700 mg/100 g in butter and up to 100 to 250 mg/100 g in cheese. Women's milk contains the same amount as cow's milk (10 mg/100 g). CLA, including rumenic acid, can also be found after heating vegetable oils and in certain food products. In fact, it is possible to obtain CLA through the partial hydrogenation of linoleic acid or by thermal treatments, and thus to find up to 0.5 g of CLA for 100 g of products in certain food products (industrial pasta, cookies) (Leray, 2013).

The intake of CLA from a typical diet is estimated at several 100 mg/d in various countries (Parodi, 2003). In the 2000s, mean daily intake of rumenic acid was 250 mg/d to 320 mg/d in female students in Germany (Fremann *et al.*, 2002), mean daily intake of CLA was 176 mg total CLA/d for men and 104 mg for women in the USA and was estimated to be almost 100 mg in the UK (Ritzenthaler *et al.*, 2001).

3.3 Tissue content and biological functions

To our knowledge, no data are found in the literature on CLA tissue content in human. It was demonstrated *in vitro* and *in vivo* in animal models that CLA plays a major role in lipid metabolism, especially as regards the oxidative cellular system. In fact, in conjugated fatty acids, the electrons become delocalized over conjugated double bonds, conferring to CLA unusual chemical properties (Gurr *et al.*, 2002). In addition to its role on lipid metabolism and lipid peroxidation, the impact of CLA on energy expenditure, insulin metabolism and inflammation were also observed (Lehnen *et al.*, 2015; Wang *et al.*, 2020a).

3.4 Health effects

Some animal studies show that CLA (and also CLNA, CEPA, CDHA) may have some beneficial health effects such as reduction of body fat, improved insulin resistance, anti-thrombogenic and anti-carcinogenic effects, reduction of atherosclerosis, improved lipid profile, modulation of the immune system and stimulation of bone mineralization (Wang *et al.*, 2020a). The most studied CLA supplementation effect is its capacity to alter the body composition, promoting an increase in lean mass and reduction of the fatty mass

(Lehnen *et al.*, 2015). However, in humans, the clinical evidence appears to be insufficient and not unanimous regarding the health effects of CLA (Ritzenthaler *et al.*, 2001; Lehnen *et al.*, 2015). Of the different isomers of CLA, rumenic acid has been reported to be the most bioactive CLA (Belury, 1995). Lehnen *et al.* recently published a comprehensive review on CLAs (Lehnen *et al.*, 2015).

4 The furan fatty acids FuFAs

4.1 Structure and biosynthesis

The furan fatty acids (FuFAs) are fatty acids with a furan ring. To date, thirty different structures have been identified (Glass *et al.*, 1974; Wang *et al.*, 2020b). The most common FuFAs are methylated or dimethylated forms. However, non-methylated furans have been also described (Yurawecz *et al.*, 1995).

Methylated FuFAs are formed from polyunsaturated fatty acids, in particular linoleic acid (Batna *et al.*, 1993) and non-methylated FuFAs are formed from conjugated dienes, in particular CLA (Yurawecz *et al.*, 1995). Currently, the biosynthetic pathway of FuFAs is not completely established and might depend on the species considered (plant, bacteria, animal, ...).

4.2 Food content and daily intake

Fish are an important source of FuFAs, with 1 to 4% of total fatty acids in the form of FuFAs (Vetter *et al.*, 2012). Butter and dairy products contain 5 to 50 mg/100 g FuFAs (Vetter *et al.*, 2012; Wendlinger and Vetter, 2014). FuFAs have been found in wheat, rice, potatoes, cabbage, orange, lemon, raspberries, with levels ranging from 1 to 350 µg/g dry matter (Hannemann *et al.*, 1989). Soybeans contain FuFAs at levels of 30 to 300 µg/g (Guth and Grosch, 1991; Wu *et al.*, 1997) while the levels are not quantifiable in olives, sesame, nuts, grape seeds and sunflower (Wahl *et al.*, 1994).

Few data exist on the ingested amount of FuFAs. The estimated average consumption of FuFAs was estimated in Germany in 2014, and observed to be about 10 to 25 mg/day (6.6 to 16.5 mg via fish, 0.7–4.8 mg via milk fat, 1.4 to 2.5 mg via soybean oil, 0.2–0.5 mg via rapeseed oil and 0.008 mg via olive oil) (Wendlinger and Vetter, 2014).

4.3 Tissue content and biological functions

To our knowledge, no data are found in the literature on FuFAs tissue content in human. FuFAs possess antioxidant properties due to the presence of the furan ring (Okada *et al.*, 1990, 1996; Masuchi Buscato *et al.*, 2020). They also have anti-microbial (Knechtle *et al.*, 2014; Dasagrathi *et al.*, 2016; Kimura *et al.*, 2018) and anti-inflammatory (Wakimoto *et al.*, 2011; Khan *et al.*, 2018; Lauvai *et al.*, 2019) properties. FuFAs may thus participate to the anti-inflammatory effects of fish oils and fish-based diets.

4.4 Health effects

FuFAs present in fish may be involved in the beneficial effects of fish consumption on cardiovascular disease (Spiteller, 2005). Several *in vitro* studies support this hypothesis

(Graff *et al.*, 1984; Okada *et al.*, 1996; Fuchs and Spiteller, 1999) as well as studies conducted in humans (Wahl *et al.*, 1994; Zheng *et al.*, 2016; Tovar *et al.*, 2017). Moreover, *in vitro* FuFAs modulate lipid metabolism in adipose tissues (Lengler *et al.*, 2012; Lauvai *et al.*, 2019). The 3-carboxy-4-methyl-5-propyl-2-furanpropanoic acid (CMPF), a degradation product of FuFAs, also derived from the metabolism of n-3 PUFAs, could prevent or even reverse hepatic steatosis (Prentice *et al.*, 2018; Dai *et al.*, 2019; Mohan *et al.*, 2019). Alvarado *et al.* recently published a comprehensive review on FuFAs (Alvarado *et al.*, 2021).

5 The branched chain fatty acids BCFAs

5.1 Structure and biosynthesis

BCFAs are saturated fatty acids with one or more methyl groups in the linear carbon chain. There are two distinct series of BCFAs: the *iso*-series where the terminal group is $\begin{array}{c} \text{CH}_3 \\ | \\ \text{CH}_3\text{-CH-} \end{array}$ and the *anteiso*-series where the terminal group is $\begin{array}{c} \text{CH}_3 \\ | \\ \text{CH}_3\text{-CH}_2\text{-CH-} \end{array}$ (Gurr *et al.*, 2002). However, branch points can also be found in other positions. More than 50 BCFAs have been identified in ruminant-derived fats (Taormina *et al.*, 2020). Monomethyl BCFAs are the most abundant. Among multimethyl BCFAs, phytanic acid (3,7,11,15-tetramethylhexadecanoic acid) and pristanic acid (2,6,10,14-tetramethylpenta-decanoic acid) are predominant although in lesser amounts compared to monomethyl structures (Leray, 2013; Taormina *et al.*, 2020).

In ruminants, BCFAs are synthesized by microorganisms in the rumen, from dietary branched-chain amino acids such as valine, leucine and isoleucine (Taormina *et al.*, 2020). Wallace *et al.* (Wallace *et al.*, 2018) demonstrated also that BCFAs were synthesized *de novo* in adipose tissues from branched-chain amino acids catabolized in mitochondria, and then exported by carnitine acetyltransferase to the cytosol, where they were elongated by fatty acid synthase.

5.2 Food content and daily intake

BCFAs occur widely but mainly at low concentrations in animal fat and some marine oils (Gurr *et al.*, 2002). BCFAs are present in the milk and tissues of ruminants consumed by humans (beef, sheep, goat). In cow's milk, the concentration of phytanic acid ranges from 0.16 to 0.59 g/100 g of lipids and that of pristanic acid from 0.03 to 0.09 g/100 g of lipids (Leray, 2013). In some wild fishes, BCFAs were only 1% ± 0.5% (mean ± SD) of the total fatty acids, contributing only a small amount of BCFAs per serving to the diet. Consuming a standardized portion (70 g) of wild freshwater fish contributes to only small amounts of BCFAs (for instance 2.5–24.2 mg, in the American diet) (Wang *et al.*, 2016). Asian food, fermented soy known as natto and fermented shrimp paste have high BCFA levels, 1.71 ± 0.17% and 3.18 ± 0.14% BCFAs, respectively (Wang, 2017), relative to total fatty acids.

Few data exist on the ingested amount of BCFAs. In the USA in 2011, the consumption of milk, cheese and beef contributed to a daily dietary intake of about 400 mg of branched fatty acids (Ran-Ressler *et al.*, 2011). Consumption of chocolate contributed to about 6 mg BCFA/day (Ran-Ressler *et al.*, 2014).

5.3 Tissue content and biological functions

In mammalian tissues, BCFAs rarely constitute more than 1–2% of the total fatty acid pool (Pakiet *et al.*, 2020). BCFA are present in the gut from a very early age and throughout the human life cycle. BCFA are major components of the lipids of Gram-positive bacteria (such as *Bacillus* and *Lactobacillus*). They play an important regulatory role in fluidity and permeability of bacterial membrane (Taormina *et al.*, 2020). They have a positive influence on the development of commensal bacteria from birth, and on intestinal metabolism (Leray, 2013).

5.4 Health effects

BCFAs may contribute to the positive health effects attributed to dairy product consumption. Several *in vivo* studies show protective effects against inflammation, cancers and metabolic disorders (Ran-Ressler *et al.*, 2014; Taormina *et al.*, 2020). In an animal model, BCFAs play a beneficial role against inflammation in the premature intestine, modulate the microbiota and increase the expression of anti-inflammatory cytokines (Taormina *et al.*, 2020). To date, no data concerning the metabolic effects were reported in humans. However it was suggested that BCFAs may favourably influence insulin sensitivity, energy and glucose metabolism in human (Taormina *et al.*, 2020). Taormina *et al.* recently published a comprehensive review on BCFA (Taormina *et al.*, 2020).

6 The branched fatty acid esters of hydroxy fatty acids FAHFAs

6.1 Structure and biosynthesis

FAHFAs are fatty acid esters of hydroxy fatty acids. As multiple combinations of fatty acids (FA) and hydroxylated fatty acids (HFA) are possible, there are hundreds of FAHFAs (Yore *et al.*, 2014; Kuda *et al.*, 2016). Almost 50 families of FAHFAs have been identified, the esters of palmitic acid and hydroxy stearic acid (PAHSA) being the most studied. In each family of branched FAHFAs, several positional isomers are possible, with more than 300 regioisomers identified, and for each isomer, there are also 2 possible configurations (Kuda *et al.*, 2018).

Branched FAHFAs are synthesized *in vivo* (Yore *et al.*, 2014) or can be obtained exogenously from food. To date, only PAHSA biosynthesis pathway in adipocytes was elucidated, involving esterification of hydroxy fatty acids with acyl-CoA fatty acids by an acyltransferase (Kuda *et al.*, 2018), and storage in adipocyte as FAHFA-containing triacylglycerols (Tan *et al.*, 2019).

6.2 Food content and daily intake

Numerous branched FAHFA families have been detected in food of plant origin (fruits, vegetables and cereals) (Zhu *et al.*, 2018; Liberati-Cizmek *et al.*, 2019) and of animal origin (egg, chicken, beef, caribou, moose) (Yore *et al.*, 2014; Pham *et al.*, 2019). Abundance of each FAHFAs varies according to the type of food considered. Total FAHFAs range from 45 to 320 ng/g in fresh food. Branched FAHFAs were also detected in caribou meat and moose at very high doses (50 µg/g)

compared to other food sources. Branched FAHFAs are present in breast milk, although at very low concentrations (Kuda *et al.*, 2018).

To our knowledge, no data are available on the amount of FAHFAs ingested daily. Moreover, absorption and bioavailability of dietary FAHAs are unknown.

6.3 Tissue content and biological functions

FAHFAs are present in blood and in many tissues in rodent and humans. PAHSA content is around 100 ng/g in white adipose tissue, 150 ng/g in brown adipose tissue, and 10–20 ng in liver, kidney and pancreas (Yore *et al.*, 2014). In lung, kidney, thymus, liver and heart FAHFAs content is rather in pg/g (Zhu *et al.*, 2017). Short-chain FAHFAs are in a concentration range from 0.84 to 57 pmoles/mg in the large intestine (Gowda *et al.*, 2020b, 2020c).

Only a few FAHFAs have been studied. They modulate favourably insulin sensitivity and glucose metabolism. In particular, 5- and 9-PAHSA have been reported to improve glucose metabolism and insulin signalling (Yore *et al.*, 2014; Moraes-Vieira *et al.*, 2016; Smith and Kahn, 2016; Syed *et al.*, 2018). 9-PAHPA or 9-OAHPA increased insulin sensitivity, but without modifying glucose tolerance, and increased basal metabolism, both in healthy mice and in obese mice with lower insulin sensitivity (Benlebna *et al.*, 2020a, 2020b). Moreover, 9-PAHPA or 9-OAHPA induced a switch toward a more oxidative contractile phenotype of skeletal muscle, suggesting a muscular origin of the increase in insulin sensitivity observed (Benlebna *et al.*, 2020c). Surprisingly, 9-PAHPA or 9-OAHPA induced hepatic steatosis and fibrosis in some healthy mice but not in obese mice, likely because both FAHFAs had insulin-sensitized the healthy liver so much that *de novo* lipogenesis promoted steatosis/fibrosis (Benlebna *et al.*, 2020a, 2020b). FAHFAs activate GPR120 and GPR40 and increase GLP-1 secretion (Yore *et al.*, 2014; Hammarstedt *et al.*, 2018; Kimura *et al.*, 2020). The FAHFAs studied to date have anti-inflammatory effects, as demonstrated both *in vitro* and *in vivo* in chronic and acute inflammation models (Yore *et al.*, 2014; Kuda *et al.*, 2016; Lee *et al.*, 2016; Kolar *et al.*, 2019). At least some FAHFAs, notably from omega-3 fatty acid derived-FAHFAs family, may have antioxidant effects (Gowda *et al.*, 2020a).

6.4 Health effects

Metabolic dysfunction in adipose tissue of healthy moderately overweight humans is associated with reduced levels of PAHSAs in the same tissue (Hammarstedt *et al.*, 2012; Hammarstedt *et al.*, 2018). In addition, serum PAHSAs levels are reduced in obese patients and in diabetics (Yore *et al.*, 2014; Moraes-Vieira *et al.*, 2016). Thus, beneficial effects of PAHSAs were suggested in human in various metabolic disorders such as type 1 and type 2 diabetes, and in chronic inflammation (Brejchova *et al.*, 2020). Other beneficial health effects have been also suggested, in particular against some cancers (Rodriguez *et al.*, 2019). It is important to note that these health effects of FAHFAs have been demonstrated with pharmacological doses in animal models or linked to

circulating FAHFAs levels, but not associated to diet normal content. Several comprehensive reviews on branched FAHFAs have been recently published (Brejchova *et al.*, 2020; Benlebna *et al.*, 2021).

7 Conclusion

Human diet contains many uncommon fatty acids with bioactive properties such as n-3 DPA, natural *trans* fatty acids, CLAs, FuFAs, BCFAs and FAHFAs. Many of them may have favourable effects on health, in particular on prevention of cardiovascular diseases, inflammation and metabolic disorders such as diabetes. It is interesting to note that many of these lipids are found mainly in seafood and in dairy products. As bacteria are involved in the synthesis of some of these fatty acids, the role of intestinal microbiota in their metabolism in humans deserves to be explored. As not only food intake but also bioavailability is important to provide adequate nutrients status, and as bioavailability is still unknown for some of these uncommon fatty acids, this parameter needs to be investigated to better understand their health effects.

Acknowledgement. The authors acknowledge the financial support of the French Lipid Nutrition Group for the work conducted on 9-PAHPA and 9-OAHPA (Benlebna *et al.*, 2020a, 2020b, 2020c). We thank all colleagues who have contributed and/or who are co-authors of our publications cited in this review.

Conflicts of interest. The authors declare that they have no conflicts of interest in relation to this article.

References

- Afssa. 2005. Risques et bénéfices pour la santé des acides gras *trans* apportés par les aliments. Afssa, p. 221.
- Afssa. 2009. Avis de l'Agence française de sécurité sanitaire des aliments sur l'estimation des apports en acides gras trans de la population française S.n. 2007-SA-0220, ed. Afssa.
- Alvarado K, Durand E, Vaysse L, *et al.* 2021. Effets bénéfiques potentiels des acides gras furaniques, des lipides alimentaires bioactifs-Potential beneficial effects of furan fatty acids, bioactive food lipids. *Cah Nutr Diet* 56: 117–125.
- Arab L. 2003. Biomarkers of fat and fatty acid intake. *J Nutr* 133 (Suppl. 3): 925S–932S.
- Batna A, Scheinkonig J, Spitteller G. 1993. The occurrence of furan fatty acids in *Isochrysis* sp. and *Phaeodactylum tricornutum*. *Biochim Biophys Acta* 1166: 171–176.
- Belury M.A. 1995. Conjugated dienoic linoleate: a polyunsaturated fatty acid with unique chemoprotective properties. *Nutr Rev* 53: 83–89.
- Benlebna M, Balas L, Bonafos B, *et al.* 2020a. Long-term intake of 9-PAHPA or 9-OAHPA modulates favorably the basal metabolism and exerts an insulin sensitizing effect in obesogenic diet-fed mice. *Eur J Nutr*.
- Benlebna M, Balas L, Bonafos B, *et al.* 2020b. Long-term high intake of 9-PAHPA or 9-OAHPA increases basal metabolism and insulin sensitivity but disrupts liver homeostasis in healthy mice. *J Nutr Biochem* 79: 108361.
- Benlebna M, Balas L, Pessemesse L, *et al.* 2020c. FAHFAs Regulate the Proliferation of C2C12 Myoblasts and Induce a Shift toward a More Oxidative Phenotype in Mouse Skeletal Muscle. *Int J Mol Sci* 21.

- Benlebna M, Balas L, Gaillet S, *et al.* 2021. Potential physiopathological effects of branched fatty acid esters of hydroxy fatty acids. *Biochimie* 182: 13–22.
- Boue C, Combe N, Billeaud C, *et al.* 2000. Trans fatty acids in adipose tissue of French women in relation to their dietary sources. *Lipids* 35: 561–566.
- Brejchova K, Balas L, Paluchova V, Brezinova M, Durand T, Kuda O. 2020. Understanding FAHFAs: From structure to metabolic regulation. *Prog Lipid Res* 79: 101053.
- Dai J, Yi J, Zhang S, *et al.* 2019. Serum 3-carboxy-4-methyl-5-propyl-2-furanpropanoic acid is associated with lipid profiles and might protect against non-alcoholic fatty liver disease in Chinese individuals. *J Diabetes Investig* 10: 793–800.
- Dasagrandhi C, Ellamar JB, Kim YS, Kim HR. 2016. Antimicrobial activity of a novel furan fatty acid, 7, 10-epoxyoctadeca-7, 9-dienoic acid against methicillin-resistant *Staphylococcus aureus*. *Food Sci Biotechnol* 25: 1671–1675.
- Drouin G. 2018. Métabolisme et intérêt nutritionnel de l'acide docosapentaénoïque n-3. Modulation du statut tissulaire en acides gras n-3 par les lipides laitiers alimentaires chez le rat. Rennes 1.
- Drouin G, Catheline D, Guillocheau E, *et al.* 2019a. Comparative effects of dietary n-3 docosapentaenoic acid (DPA), DHA and EPA on plasma lipid parameters, oxidative status and fatty acid tissue composition. *J Nutr Biochem* 63: 186–196.
- Drouin G, Rioux V, Legrand P. 2019b. The n-3 docosapentaenoic acid (DPA): A new player in the n-3 long chain polyunsaturated fatty acid family. *Biochimie* 159: 36–48.
- Fremann D, Linseisen J, Wolfram G. 2002. Dietary conjugated linoleic acid (CLA) intake assessment and possible biomarkers of CLA intake in young women. *Public Health Nutr* 5: 73–80.
- Fuchs CT, Spiteller G. 1999. 9-(3, 4-Dimethyl-5-pentyl-furan-2-yl) nonanoic Acid and 9-(3, 4-Dimethyl-5-propyl-furan-2-yl) nonanoic Acid: New Naturally Occurring Peroxidase Inhibitors. *Z Naturforsch* 54: 932–936.
- Ghasemi Fard S, Cameron-Smith D, Sinclair AJ. 2021. n-3 Docosapentaenoic acid: the iceberg n-3 fatty acid. *Curr Opin Clin Nutr Metab Care* 24: 134–138.
- Glass RL, Krick TP, Echaradt AE. 1974. New series of fatty acids in northern pike (*Esox lucius*). *Lipids* 9: 1004–1008.
- Gowda SGB, Fuda H, Tsukui T, Chiba H, Hui, SP. 2020a. Discovery of eicosapentaenoic acid esters of hydroxy fatty acids as potent Nrf2 activators. *Antioxidants* 9.
- Gowda SGB, Liang C, Gowda D, *et al.* 2020b. Identification of short chain fatty acid esters of hydroxy fatty acids (SFAHFAs) in murine model by nontargeted analysis using ultra-high-performance liquid chromatography/linear trap quadrupole-Orbitrap mass spectrometry. *Rapid Commun Mass Spectrom*.
- Gowda SGB, Gowda D, Liang C, *et al.* 2020c. Chemical Labeling Assisted Detection and Identification of Short Chain Fatty Acid Esters of Hydroxy Fatty Acid in Rat Colon and Cecum Contents. *Metabolites* 10.
- Graff G, Gellerman JL, Sand DM, Schlenk H. 1984. Inhibition of blood platelet aggregation by dioxo-ene compounds. *Biochim Biophys Acta* 799: 143–150.
- Guillocheau E, Legrand P, Rioux V. 2019. Benefits of natural dietary trans fatty acids towards inflammation, obesity and type 2 diabetes: defining the n-7 trans fatty acid family. *OCL* 26: 9.
- Guo XF, Tong WF, Ruan Y, Sinclair AJ, Li D. 2020. Different metabolism of EPA, DPA and DHA in humans: A double-blind cross-over study. *Prostaglandins Leukot Essent Fatty Acids* 158: 102033.
- Gurr MI, Harwood JL, Frayn KN. 2002. *Lipid Biochemistry*. 5th ed. Blackwell Sciences.
- Guth H, Grosch W. 1991. Detection of furanoid fatty acids in soybean oil – cause for the light-induced off-flavour. *Eur J Lipid Sci Technol* 93: 249–255.
- Hammarstedt A, Graham TE, Kahn BB. 2012. Adipose tissue dysregulation and reduced insulin sensitivity in non-obese individuals with enlarged abdominal adipose cells. *Diabetol Metab Syndr* 4: 42.
- Hammarstedt A, Syed I, Vijayakumar A, *et al.* 2018. Adipose tissue dysfunction is associated with low levels of the novel Palmitic Acid Hydroxystearic Acids. *Sci Rep* 8: 15757.
- Hannemann K, Puchta V, Simon E, Ziegler H, Ziegler G, Spiteller G. 1989. The common occurrence of furan fatty acids in plants. *Lipids* 24: 296–298.
- Kaur G, Guo XF, Sinclair AJ. 2016. Short update on docosapentaenoic acid: a bioactive long-chain n-3 fatty acid. *Curr Opin Clin Nutr Metab Care* 19: 88–91.
- Khan MA, Pace-Asciak C, Al-Hassan JM, *et al.* 2018. Furanoid F-Acid F6 Uniquely Induces NETosis Compared to C16 and C18 Fatty Acids in Human Neutrophils. *Biomolecules* 8: 144–161.
- Kimura I, Ichimura A, Ohue-Kitano R, Igarashi M. 2020. Free Fatty Acid Receptors in Health and Disease. *Physiol Rev* 100: 171–210.
- Kimura T, Tajima A, Inahashi Y, *et al.* 2018. Mumiamicin: Structure and bioactivity of a new furan fatty acid from *Mumia* sp. YSP-2-79. *J Gen Appl Microbiol* 64: 62–67.
- Knechtel P, Diefenbacher M, Greve KB, *et al.* 2014. The natural diene-furan fatty acid EV-086 is an inhibitor of fungal delta-9 fatty acid desaturation with efficacy in a model of skin dermatophytosis. *Antimicrob Agents Chemother* 58: 455–466.
- Kolar MJ, Konduri S, Chang T, *et al.* 2019. Linoleic acid esters of hydroxy linoleic acids are anti-inflammatory lipids found in plants and mammals. *J Biol Chem* 294: 10698–10707.
- Kuda O, Brezinova M, Rombaldova M, *et al.* (2016). Docosahexaenoic Acid-Derived Fatty Acid Esters of Hydroxy Fatty Acids (FAHFAs) With Anti-inflammatory Properties. *Diabetes* 65: 2580–2590.
- Kuda O, Brezinova M, Silhavy J, *et al.* 2018. Nrf2-mediated antioxidant defense and peroxiredoxin 6 are linked to biosynthesis of palmitic acid ester of 9-hydroxystearic acid. *Diabetes* 67: 1190–1199.
- Lauvai J, Becker AK, Lehnert K, Schumacher M, *et al.* 2019. The furan fatty acid 9m5 acts as a partial ligand to peroxisome proliferator-activated receptor gamma and enhances adipogenesis in 3T3-L1 preadipocytes. *Lipids* 54: 277–288.
- Lee J, Moraes-Vieira PM, Castoldi A, *et al.* 2016. Branched Fatty Acid Esters of Hydroxy Fatty Acids (FAHFAs) protect against colitis by regulating gut innate and adaptive immune responses. *J Biol Chem* 291: 22207–22217.
- Lehnen TE, da Silva MR, Camacho A, Marcadenti A, Lehnen AM. 2015. A review on effects of conjugated linoleic fatty acid (CLA) upon body composition and energetic metabolism. *J Int Soc Sports Nutr* 12: 36.
- Lengler I, Buhre T, Scharmach E, Lampen A. 2012. *In-vitro* toxicological and proteomic analysis of furan fatty acids which are oxidative metabolites of conjugated linoleic acids. *Lipids* 47: 1085–1097.
- Leray C. 2013. Les lipides. Nutrition et santé. Lavoisier.
- Liberati-Cizmek AM, Bilus M, Brkic AL, *et al.* 2019) Analysis of Fatty Acid Esters of Hydroxyl Fatty Acid in Selected Plant Food. *Plant Foods Hum Nutr* 74: 235–240.

- Masuchi Buscato MH, Muller F, Vetter W, Weiss J, Salminen H. 2020. Furan fatty acids in enriched omega-3 fish oil: Oxidation kinetics with and without added monomethyl furan fatty acid as potential natural antioxidant. *Food Chem* 327: 127087.
- Mohan H, Brandt SL, Kim JH, *et al.* 2019. 3-carboxy-4-methyl-5-propyl-2-furanpropanoic acid (CMPF) prevents high fat diet-induced insulin resistance *via* maintenance of hepatic lipid homeostasis. *Diabetes Obes Metab* 21: 61–72.
- Moraes-Vieira PM, Saghatelian A, Kahn BB. 2016. GLUT4 Expression in Adipocytes Regulates De Novo Lipogenesis and Levels of a Novel Class of Lipids With Antidiabetic and Anti-inflammatory Effects. *Diabetes* 65: 1808–1815.
- Okada Y, Kaneko M, Okajima H. 1996. Hydroxyl radical scavenging activity of naturally occurring furan fatty acids. *Biol Pharm Bull* 19: 1607–1610.
- Okada Y, Okajima H, Konishi H, 1990. Antioxidant effect of naturally occurring furan fatty acids on oxidation of linoleic acid in aqueous dispersion. *JAOCS* 67: 858–862.
- Oteng AB, Kersten S. 2020. Mechanisms of Action of trans Fatty Acids. *Adv Nutr* 11: 697–708.
- Pakiet A, Wilczynski M, Rostkowska O, *et al.* 2020. The Effect of One Anastomosis Gastric Bypass on Branched-Chain Fatty Acid and Branched-Chain Amino Acid Metabolism in Subjects with Morbid Obesity. *Obes Surg* 30: 304–312.
- Parodi P.W. 2003. Conjugated linoleic acid in food. In: Sebedio JL, Christie WW, Adlof R, eds. *Advances in conjugated linoleic acid research*. Champaign: AOCS Press, pp. 101–122.
- Pham TH, Vidal NP, Manful CF. 2019. Moose and Caribou as Novel Sources of Functional Lipids: Fatty Acid Esters of Hydroxy Fatty Acids, Diglycerides and Monoacyldiglycerides. *Molecules* 24: 232–247.
- Prentice KJ, Wendell SG, Liu Y, *et al.* 2018. CMPF, a Metabolite Formed Upon Prescription Omega-3-Acid Ethyl Ester Supplementation, Prevents and Reverses Steatosis. *EBio Med* 27: 200–213.
- Qiu B, Wang Q, Liu W, *et al.* 2018. Biological effects of trans fatty acids and their possible roles in the lipid rafts in apoptosis regulation. *Cell Biol Int* 42: 904–912.
- Ran-Ressler R.R., Bae S, Lawrence P, Wang DH, Brenna JT. 2014. Branched-chain fatty acid content of foods and estimated intake in the USA. *Br J Nutr* 112: 565–572.
- Ran-Ressler RR, Sim D, O'Donnell-Megaró AM, Bauman DE, Barbano DM, Brenna JT. 2011. Branched chain fatty acid content of United States retail cow's milk and implications for dietary intake. *Lipids* 46: 569–576.
- Richter CK, Bisselou KS, Nordgren TM. 2019. n-3 Docosapentaenoic Acid Intake and Relationship with Plasma Long-Chain n-3 Fatty Acid Concentrations in the United States: NHANES 2003–2014. *Lipids* 54: 221–230.
- Ritzenthaler KL, McGuire MK, Falen R, Shultz TD, Dasgupta N, McGuire MA. 2001. Estimation of conjugated linoleic acid intake by written dietary assessment methodologies underestimates actual intake evaluated by food duplicate methodology. *J Nutr* 131: 1548–1554.
- Rodriguez JP, Guijas C, Astudillo AM, Rubio JM, Balboa MA, Balsinde J. 2019. Sequestration of 9-Hydroxystearic Acid in FAHFA (Fatty Acid Esters of Hydroxy Fatty Acids) as a Protective Mechanism for Colon Carcinoma Cells to Avoid Apoptotic Cell Death. *Cancers* 11.
- Smith U, Kahn BB. 2016. Adipose tissue regulates insulin sensitivity: role of adipogenesis, de novo lipogenesis and novel lipids. *J Intern Med* 280: 465–475.
- Spiteller G. 2005. The relation of lipid peroxidation processes with atherogenesis: a new theory on atherogenesis. *Mol Nutr Food Res* 49: 999–1013.
- Syed I, Lee J, Moraes-Vieira PM, *et al.* 2018. Palmitic Acid Hydroxystearic Acids Activate GPR40, Which Is Involved in Their Beneficial Effects on Glucose Homeostasis. *Cell Metab* 27: 419–427 e414.
- Tan D, Ertunc ME, Konduri S, *et al.* 2019. Discovery of FAHFA-Containing Triacylglycerols and Their Metabolic Regulation. *J Am Chem Soc* 141: 8798–8806.
- Taormina VM, Unger AL, Schiksnis MR, Torres-Gonzalez M, Kraft J. 2020. Branched-Chain Fatty Acids-An Underexplored Class of Dairy-Derived Fatty Acids. *Nutrients* 12: 28756–22872.
- Tovar J, de Mello VD, Nilsson A, *et al.* 2017. Reduction in cardiometabolic risk factors by a multifunctional diet is mediated *via* several branches of metabolism as evidenced by nontargeted metabolite profiling approach. *Mol Nutr Food Res* 61.
- Vetter W, Laure S, Wendlinger C, Mattes A, Smith AWT, Knight DW. 2012. Determination of Furan Fatty Acids in Food Samples. *J Am Oil Chem Soc* 89: 1501–1508.
- von Schacky C, Harris WS. 2018. Why docosapentaenoic acid is not included in the Omega-3 Index. *Prostaglandins Leukot Essent Fatty Acids* 135: 18–21.
- Wahl HG, Liebich HM, Hoffmann A. 1994. Identification of fatty acid methyl esters as minor components of fish oil by multidimensional GC-MSD: New furan fatty acids. *J Separation Sci* 17: 308–311.
- Wakimoto T, Kondo H, Nii H, *et al.* 2011. Furan fatty acid as an anti-inflammatory component from the green-lipped mussel *Perna canaliculus*. *Proc Natl Acad Sci U S A* 108: 17533–17537.
- Wallace M, Green CR, Roberts LS, *et al.* 2018. Enzyme promiscuity drives branched-chain fatty acid synthesis in adipose tissues. *Nat Chem Biol* 14: 1021–1031.
- Wang D. 2017. Branched chain fatty acids (BCFA) in nature: fish, fermented food and sea lion vernix caseosa R. Cornell University, 116 p.
- Wang DH, Jackson JR, Twining C, *et al.* 2016. Saturated Branched Chain, Normal Odd-Carbon-Numbered, and n-3 (Omega-3) Polyunsaturated Fatty Acids in Freshwater Fish in the Northeastern United States. *J Agric Food Chem* 64: 7512–7519.
- Wang J, Han L, Wang D, Li P, Shahidi F. 2020a. Conjugated Fatty Acids in Muscle Food Products and Their Potential Health Benefits: A Review. *J Agric Food Chem* 68: 13530–13540.
- Wang Y, Pritchard GJ, Kimber MC. 2020b. A General Convergent Strategy for the Synthesis of Tetra-Substituted Furan Fatty Acids (FuFAs). *Eur JOC* 2020: 2914–2922.
- Wendlinger C, Vetter W. 2014. High concentrations of furan fatty acids in organic butter samples from the German market. *J Agric Food Chem* 62: 8740–8744.
- Wu X, Hammond EG, White PJ, Fehr W. 1997. Analysis of furanoid esters in soybean oil and the effect of variety and environment on furanoid ester content. *JAOCS* 74: 1099–1103.
- Yore MM, Syed I, Moraes-Vieira PM, *et al.* 2014. Discovery of a class of endogenous mammalian lipids with anti-diabetic and anti-inflammatory effects. *Cell* 159: 318–332.
- Yurawecz MP, Hood JK, Mossoba MM, Roach JA, Ku Y. 1995. Furan fatty acids determined as oxidation products of conjugated octadecadienoic acid. *Lipids* 30: 595–598.
- Zheng JS, Lin M, Imamura F, *et al.* 2016. Serum metabolomics profiles in response to n-3 fatty acids in Chinese patients with type 2 diabetes: a double-blind randomised controlled trial. *Sci Rep* 6: 29522.

- Zhu QF, Yan JW, Gao Y, Zhang JW, Yuan BF, Feng YQ. 2017. Highly sensitive determination of fatty acid esters of hydroxyl fatty acids by liquid chromatography-mass spectrometry. *J Chromatogr B Analyt Technol Biomed Life Sci* 1061–1062: 34–40.
- Zhu QF, Yan JW, Zhang TY, Xiao HM, Feng YQ. 2018. Comprehensive Screening and Identification of Fatty Acid Esters of Hydroxy Fatty Acids in Plant Tissues by Chemical Isotope Labeling-Assisted Liquid Chromatography-Mass Spectrometry. *Anal Chem* 90: 10056–10063.

Cite this article as: Coudray C, Durand E, Balas L, Sultan A, Casas F, Feillet-Coudray C. 2021. Potential favourable health effects of some dietary uncommon fatty acids. *OCL* 28: 41.