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In silico QTL mapping in an oil palm breeding program reveals a quantitative and complex genetic resistance to *Ganoderma boninense*

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1 **Title Page**

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3 *In silico* QTL mapping in an oil palm breeding program reveals a quantitative and complex genetic resistance to
4 *Ganoderma boninense*

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31

32 **Abstract**

33 Basal stem rot caused by *Ganoderma boninense* is the major threat to oil palm cultivation in South-East Asia,
34 which accounts for 80% of palm oil production worldwide, and this disease is increasing in Africa. The use of
35 resistant planting material as part of an integrated pest management of this disease is one sustainable solution.
36 However, breeding for *Ganoderma* resistance requires long-term and costly research, which could greatly benefit
37 from marker assisted selection (MAS). In this study, we evaluated the effectiveness of an *in silico* genetic mapping
38 approach that took advantage of extensive data recorded in an ongoing breeding program. A pedigree-based QTL
39 mapping approach applied to more than 10 years' worth of data collected during pre-nursery tests revealed the
40 quantitative nature of *Ganoderma* resistance and identified underlying loci segregating in genetic diversity that is
41 directly relevant for the breeding program supporting the study. To assess the consistency of QTL effects between
42 pre-nursery and field environments, information was collected on the disease status of the genitors planted in
43 genealogical gardens and modeled with pre-nursery-based QTL genotypes. In the field, individuals were less likely
44 to be infected with *Ganoderma* when they carried more favorable alleles at the pre-nursery QTL. Our results pave
45 the way for a MAS of *Ganoderma* resistant and high yielding planting material and the provided proof-of-concept
46 of this efficient and cost-effective approach could motivate similar studies based on diverse breeding programs.

47

48 **Keywords**

49 Oil palm, basal stem rot disease, nursery screening test, breeding population, QTL mapping, pedigree-
50 based analysis.

51

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62

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70 Florence Jacob, Norbert Billotte, Sébastien Tisné ; Visualization: Aurélie Daval, Sébastien Tisné.

71

72 **1. Introduction**

73 The African oil palm (*Elaeis Guineensis* Jacq.) is the leading oil crop worldwide with a global annual production
74 of around 73 Mt, and accounts for more than 35% of all the edible vegetable oil produced worldwide (USDA
75 statistics, 2019). Oil palm is expected to be able to respond to the global increase in the demand for vegetable oil
76 projected to be 240 Mt in 2050, even higher if its non-food uses are included (Corley 2009). The oil palm sector
77 has agreed on sustainability goals to reach this global demand (Rochmyaningsih 2019), in particular through the
78 certification of sustainable produced palm oil (the Roundtable on Sustainable Palm Oil, RSPO, <https://rspo.org/>).
79 However, pests and diseases threaten palm oil production in all areas of cultivation and contribute to the current
80 yield gap (Woittiez et al. 2017). If it is to achieve the zero-deforestation goal in high conservation value forests
81 included in the RSPO commitments, oil palm will inevitably be cultivated on existing arable lands under increasing
82 pathogen pressure. The integrated pest management (IPM) covers sustainable solutions to this problem including
83 improved plant disease resistance. Oil palm breeders thus needs to focus on developing resistant planting material,
84 while maintaining or even improving oil yield.

85 The basal stem rot disease caused by *Ganoderma boninense* is a major threat in South East Asia, with projections
86 worsening due to climate change (Paterson 2019). This pathogenic fungus is a soil-borne basidiomycete that
87 mainly infects the oil palm when its roots come into contact with infected debris or with the roots of neighbor
88 palms (Rees et al. 2009). *Ganoderma* stem rot disease has a significant effect on oil yield even when only 10-20%
89 of palm trees are infected, and 30-70% of the trees may have died over a typical 25-year planting cycle (Durand-
90 Gasselin et al. 2005; Cooper et al. 2011). To date, no specific interaction and/or complete resistance have been
91 identified in oil palm/*Ganoderma* pathosystem, which is consistent with its hemibiotrophic pathogenic lifestyle.
92 However, observations of contrasted levels of resistance in diverse genetic backgrounds suggest that breeding for
93 quantitative disease resistance (QDR) is a promising solution (Franqueville et al. 2001). Typically, research on
94 perennial plant disease resistance is based on large scale costly field experiments, even more so when investigating
95 QDR. When possible, *ex situ* experiments with controlled inoculation of the pathogen are powerful tools that offer
96 more repeatability and increase both speed and throughput, especially in genetic surveys. In oil palm, such pre-
97 nursery tests were first developed for research on vascular wilt (De Franqueville and Renard 1990), followed by
98 *Ganoderma* in the 2000s (Idris et al. 2004; Breton et al. 2006b; Rees et al. 2007) and are now widely used.
99 However, transferring results to the field can be problematic because of a more complex biotic context, the age
100 specificity of the QDR mechanisms, or the effects of cultural practice management on disease epidemiology.
101 Despite these challenges, by combining field and pre-nursery approaches in long-term works in the framework of
102 an oil palm breeding program, Cirad, its subsidiary PalmElit, and their partners have managed to release planting
103 material that is highly resistant to vascular wilt and intermediate resistant to basal stem rot caused by *Ganoderma*
104 (De Franqueville and Renard 1990; Franqueville et al. 2001; Durand-Gasselin et al. 2005; Breton et al. 2009).
105 Information on the genetic architecture and molecular determinisms of traits of interest could help shorten the long
106 breeding cycle of oil palm, which currently exceeds 20 years, and would be particularly useful in the case of
107 *Ganoderma* disease given the cumbersome nature of field and nursery trials. Marker assisted selection (MAS)
108 based on this information would increase the annual genetic gain thanks to both accelerated evaluation of selection
109 candidates and increased selection intensity by enabling surveys of wider genetic diversity at the same cost (Cros
110 et al. 2015, 2017). Moreover, identification of the genetic bases of resistance to *Ganoderma* could resolve the
111 challenge of breeding for both QDR and yield related traits (Nelson et al. 2018) by using simulation and prediction
112 tools (Tisné et al. 2019). Most molecular studies on *Ganoderma* disease to date have been based on inoculated vs
113 non-inoculated seedlings at the pre-nursery stage, with no or low genetic diversity. The first investigations focused
114 on *a priori* selection of candidate resistance genes to fungal diseases (Yeoh et al. 2012, 2013; Tan et al. 2013).

115 Next the genes, proteins and pathways affected by *Ganoderma* infection were identified using broader
116 transcriptomic (Tee et al. 2013; Ho et al. 2016; Bahari et al. 2018; Faizah et al. 2020; Sakeh et al. 2020), proteomic
117 (Al-Obaidi et al. 2014) and metabolomic (Nusaibah et al. 2016) approaches. Considering that *Ganoderma* is a
118 white rot fungus (Paterson 2007), lignin related traits were investigated as putative QDR mechanisms by surveying
119 the response of lignin content and composition to *Ganoderma* infection together with the associated genes
120 (Govender et al. 2017). Lignin related traits and nutritional traits were found to differ in progenies with different
121 levels of resistance to *Ganoderma* (Govender et al. 2020) but the restrained genetic design confounds the effects
122 of genetic and resistance variation.

123 QTL mapping offers an alternative approach that provides information on the genetic architecture based on a
124 relevant genetic diversity, with no *a priori* biological knowledge. The detected loci form the basis of the MAS
125 strategy but also provide insights into the mechanisms and genes involved in the QDR. The first published QTL
126 study reported the analysis of 79 individuals from one resistant and two susceptible families based on 58 simple
127 sequence repeat markers and found alleles associated with *Ganoderma* symptoms (Hama-Ali et al. 2015). More
128 conclusive insights would require much more data, but QTL analyses of oil palm crosses are typically not
129 sufficiently effective due to biological and cost constraints (Jeennor and Volkaert 2014; Lee et al. 2015; Pootakham
130 et al. 2015). This is even more problematic for field studies that are indispensable to assess genetic diversity in an
131 agronomic context, whose implementation is very costly and would result in lower production income due to the
132 disease context. A powerful and cost-effective approach is to directly use the databases compiled in ongoing
133 breeding programs, which are typically large and obtained from diverse relevant genetic backgrounds, to map *in*
134 *silico* the QTLs for the traits of interest (Parrisieux and Bernardo 2004). Despite the potential of this approach,
135 data from breeding programs are unique, mainly because of a complex genetic design that may be biased due to
136 selection, or unbalanced phenotyping coverage. Thus, they require appropriate statistical models for their
137 development and evaluation in contrasted contexts, which are currently an active research topic (Würschum 2012;
138 Garin et al. 2017; Korontzis et al. 2020). In oil palm, an *in silico* QTL mapping approach based on the two step
139 variance component approach considering identity by descent (IBD) information (George et al. 2000; van Eeuwijk
140 et al. 2010) yielded promising results on production traits recorded in large scale evaluation genetic trials (Tisné
141 et al. 2015). This approach was successfully extended to survival data and applied to a multi-parent population to
142 detect *Ganoderma* resistance QTLs in the field, allowing to identify two QTL related to the occurrence of the first
143 disease symptoms, and two related to the death due to *Ganoderma* (Tisné et al. 2017). A Bayesian approach to
144 pedigree based QTL mapping using IBD information was also developed in the 2000s and implemented in the

145 FlexQTL software (van de Weg et al. 2004; Bink et al. 2008). This made it possible to carry out increasing numbers
146 of studies in several crops that share the constraints and potential described above for oil palm, in particular for
147 disease resistance in strawberry (Mangandi et al. 2017; Anciro et al. 2018) or in apple (van de Weg et al. 2018).
148 In this study, we evaluated the potential of an *in silico* approach based on the large existing databases of a long-
149 term oil palm breeding program for the study of *Ganoderma* resistance. We genotyped an existing DNA bank
150 primarily established for identity checking purpose and performed a pedigree-based QTL mapping using data
151 recorded in *Ganoderma* pre-nursery trials over a period of more than ten years. We then assessed the consistency
152 of pre-nursery QTL effects in natural field conditions using a database recording the *Ganoderma* infection status
153 over years for the palms planted in genealogical gardens. Thus, using a cost-effective approach that is directly
154 relevant to the breeding program, we were able to study two major issues, i.e. the genetic architecture and
155 consistency between pre-nursery and field results, paving the way for the implementation of MAS for *Ganoderma*
156 resistant planting material.

157

158 **2. Material and methods**

159 **2.1. Plant material**

160 The palm trees used in this study belong to the oil palm breeding program of Cirad, its subsidiary PalmElit and
161 their partner PT Socfin Indonesia (Indonesia). This breeding program is conducted in a recurrent reciprocal
162 selection scheme with two heterotic groups A and B (GA and GB to produce superior GA×GB hybrid crosses used
163 as commercial planting material (Gascon and De Berchoux 1964; Meunier and Gascon 1972). Individuals from
164 different heterotic groups have complementary yield component traits, with low fruit bunch number and high
165 bunch weight in GA and reciprocally in GB. GA×GB hybrids consequently show a heterosis effect on fruit bunch
166 yield. Moreover, individuals included in GA are Dura palms, homozygous for the thick alleles of the shell gene
167 (Singh et al. 2013) while individuals included in GB are Pisifera (homozygous alternative alleles), the hybrid
168 GA×GB being Tenera which is the most productive form with thin shell. The parental population studied for the
169 *Ganoderma* resistance included only individuals from GB, grouping genetic origins of La Mé (LM, Ivory Coast)
170 and Yangambi (YBI, Republic Democratic of Congo). The GB pedigree used in the pre-nursery analysis comprised
171 372 individuals including founders, with 246/126 from LM/YBI genetic origin respectively and 240/93 genotyped
172 (Supp. Table 1). Among them 200 LM and 83 YBI parents were directly progeny tested for *Ganoderma* resistance
173 in a pre-nursery screening test (Fig. 1). The individuals were distributed over many full-sib families derived from
174 a small number of founders through consecutive crosses or self-pollinations in the framework of the ongoing

175 breeding program (Fig. 1). Among the 372 individuals in the whole pedigree, 219 LM individuals were planted
176 between 1970s and 2000s at the same location (Bangun Bandar, Indonesia) and were used for subsequent field
177 analysis.

178 **2.2. Phenotypic data**

179 2.2.1. Pre-nursery screening tests

180 An early pre-nursery screening test was developed in the 2000s by Cirad and Socfin Indonesia in the Tanah
181 Gambus estate, Indonesia. The first objective was to speed up the evaluation of genetic resistance to *Ganoderma*
182 of commercial oil palm planting material, using controlled and standardized inoculation of germinated seeds
183 (Breton et al., 2006). The inoculation of germinated seed was performed using a 12 week-old *Ganoderma*-
184 colonized rubber wood block (108 cm³) as inoculum source, that was previously deposited in the nursery polybag
185 before the seeds were planted.

186 A pure dikaryotic *Ganoderma boninense* isolate was used in all the trials (NJ), previously harvested from an
187 infected oil palm planted in Bangun Bandar, SOCFINDO estate (Mercière et al. 2015). This isolate was
188 successively regenerated from the bole of young infected seedlings in consecutive pre-nursery trials to provide
189 several dikaryotic clonal lines (CL, n=7) over the 10 years of testing. These reactivating steps of the isolate made
190 it possible to avoid the loss of pathogenicity often observed after successive sub-cultures on artificial fungi growth
191 media (Butt et al. 2006). A single pathogen CL was used for all the crosses tested in a single trial. Around 100
192 crosses were assessed simultaneously in each pre-nursery trial. Among them, 20% were control crosses from
193 susceptible, intermediate and resistant genetic backgrounds and were included in all the trials performed. Of the
194 remaining 80% of crosses representing the tested crosses, 50% overlapped two consecutive trials, leading to at
195 least two independent tests per tested cross. Each cross was represented by 100 inoculated germinated seeds
196 clustered in five replicates following the protocol described by Breton et al. (2009). Inoculated seedlings were
197 observed every four weeks for the appearance of the first external disease symptom, on average between 8 and 12
198 weeks after inoculation of the germinated seeds, after which the disease symptoms were recorded at two weekly
199 intervals as (1) infected and (0) if not infected. The trial was stopped when the average percentage of infected
200 seedlings within the group of control crosses reached 30%, usually around 34 weeks after inoculation of the
201 germinated seeds. This 30% threshold was determined to have the best “discriminating power” between the
202 resistant and sensitive control crosses, and so among the tested progenies (Breton et al. 2009).

203 This study included 102 *Ganoderma* pre-nursery screening test trial, covering 10 years of data recording. The trials
 204 performed between 2007 and 2017 represented the evaluation of 4,017 unique crosses, from either GA×GA,
 205 GA×GB or GB×GB genetic background. Considering that the purpose of this study was to assess the genetic bases
 206 of *Ganoderma* resistance in the commercial genetic material, only the GA×GB crosses were taken into
 207 consideration (n=3,792), derived from 2,037 and 340 individuals from the GA and GB respectively. Each parent
 208 from GB included in the analysis was progeny tested in an average of 20.5 GA×GB crosses.

209 2.2.2. Statistical modeling of pre-nursery data

210 The resistance of the GB individuals was progeny-tested through several GA×GB crosses involving them as GB
 211 parents. The response variable Y considered in this study was the proportion of affected progenies per cross at the
 212 end of the trial. A first step of statistical modeling of Y was necessary to obtain a single value per genotype required
 213 for the QTL analysis while accounting for nuisance effects due to the long-term data. Y was modeled using
 214 generalized linear mixed models (GLMM). Briefly, in a GLMM, Y is assumed to be generated by a particular
 215 distribution in the exponential family. The conditional mean of the distribution μ is linked to a linear predictor η
 216 which contains fixed and random effects, through the inverse link function g^{-1} :

$$217 \quad g(\mu) = \eta = X\beta + Z_T u_T + Z_A u_A + Z_B u_B + Z_C u_C$$

218 where X is a $n \times m$ design matrix relating observations to *Ganoderma boninense* CL fixed effects β where β is
 219 a $m \times 1$ vector ($m = 7$), Z_T is a $n \times t$ design matrix relating observations to trial random effects $u \sim N(0, I\sigma_T^2)$
 220 with u is a $t \times 1$ vector ($t = 102$), Z_C is a $n \times c$ design matrix relating observations to specific combining ability
 221 (SCA) random effects $g_C \sim N(0, I\sigma_C^2)$ where g_C is a $c \times 1$ vector ($c = 3,792$), Z_A and Z_B are $n \times q_A$ and $n \times q_B$
 222 design matrices relating observations to general combining ability (GCA) random effects for GA and GB,
 223 $g_A \sim N(0, A_A\sigma_A^2)$ and $g_B \sim N(0, A_B\sigma_B^2)$ respectively, where g_A and g_B are $q_A \times 1$ and $q_B \times 1$ vectors,
 224 respectively ($q_A = 2,037$ and $q_B = 340$). A_A and A_B are the pedigree-based kinship matrices of GA and GB,
 225 respectively.

226 In our work, we explored two types of distributions: binomial distribution, which is the appropriate one for
 227 proportional data, and normal distribution, for which more derived genetic parameters can be estimated.

228 The first model considers a binomial distribution such as:

$$229 \quad Y_{C,t} | u_t, u_A, u_B, u_C \sim Bin(n_{C,t}, \pi_{C,t})$$

230 where $Y_{c,t}$ is the number of affected progenies in the cross (c) and the trial (t) among the number of inoculated
231 progenies $n_{c,t}$, and $\pi_{c,t}$ is the associated probability.

232 The link function g is the logit such as:

$$233 \quad g(\pi_{c,t}) = \log\left(\frac{\pi_{c,t}}{1-\pi_{c,t}}\right) = \eta_{c,t}$$

234 The second model considers a normal distribution such as:

$$235 \quad Y_{c,t} | u_T, u_A, u_B, u_C \sim N(\eta_{c,t}, \sigma^2)$$

236 where $Y_{c,t}$ is the proportion of affected progenies in the cross (c) and the trial (t), σ^2 is the residual variance, and
237 the link function is the identity. Note that this second model is a linear mixed model (LMM).

238 Both models enabled prediction of the best linear unbiased predictor (BLUP) for each GB individual used in the
239 QTL mapping, A_B being replaced by an identity matrix in order to avoid using the pedigree information that was
240 subsequently used in the QTL analysis. Both statistical models were performed using ASReml-R software (Butler
241 et al. 2007, V4) and resulted in two vectors of BLUP for group B individuals that were used in subsequent QTL
242 mapping analysis.

243

244 **2.3. Molecular data and genetic map construction**

245 The 334 freeze-dried oil palm leaf samples available at the Cirad DNA-bank for the GB individuals included in
246 the analysis were genotyped with 199 SSR markers developed in different studies. Among the 199 markers, 177
247 markers were developed by Cirad (Billotte et al. 2005), two by the Lee et al. (2015), four markers by the Malaysian
248 Palm Oil Board (MPOB) (Zaki et al. 2012) and 18 expressed sequence tags markers were developed by IRD
249 (*Institut de Recherche pour le Développement*) and Cirad (Tranbarger et al. 2012). These markers were selected
250 based on a previous integrated pedigree-based genetic map constructed from a population of related individuals
251 (Cochard et al. 2015). Selection was for a uniform distribution in the genome and the highest level of
252 polymorphism in both LM and YBI genetic backgrounds. The information concerning markers was gathered in
253 the supp. Table 2. DNA extraction, evaluation of the DNA concentrations and microsatellite fragment
254 amplification were performed using the protocol described in Cochard et al. (2015). Genemapper© V4.1 (Applied
255 Biosystems, USA) software was used to determine the size of the alleles.

256 Three genetic maps were constructed, one for each of LM and YBI population and one integrated map using the
257 pedigree-based linkage mapping software CRI-MAP v2.4 (Green et al. 1990), as described in Cochard et al. (2015).

258 Consistency of marker calling across pedigrees and absence of spurious rates of double recombination events were
259 checked using both CRI-MAP and FlexQTL™, and data were improved where necessary. Genetic maps were
260 drawn using MapChart v2.0 software (Voorrips 2002) and are presented in Supporting Information Figure S1.

261 **2.4. Pre-nursery QTL mapping approach**

262 QTL mapping of *Ganoderma* disease resistance in pre-nursery conditions followed two main steps. The first step
263 was carried out using a Bayesian approach and a multiple QTL model implemented in FlexQTL™ (Bink et al.
264 2002, 2014, 2008; www.flexqtl.nl) on the pre-nursery data after modeling, in order to identify putative QTL
265 positions and predict the QTL genotypes. The second step consisted in stepwise QTL model selection on the raw
266 pre-nursery data using the predicted QTL genotypes as fixed effects in the LMM.

267 2.4.1. QTL region identification and QTL genotype prediction

268 Six separate QTL analyses, corresponding to the two vectors of GB individual BLUP (see Phenotypic data section)
269 with three different starting random seeds were performed using FlexQTL™. The six QTL analyses were based
270 on a model with additive QTL effects, with the parameters MaximQTL and priorQTL set at 20 and 5 respectively
271 for the Markov chain Monte Carlo simulation. The length of the Markov chains were set at 1 000 000 with a
272 thinning value of 1 000. Using these parameters, the convergence indicators reached satisfying values for each
273 parameter assessed (overall mean, μ , the residual variance, σ_e^2 , the number of QTLs, N_{QTL} , and the variance of
274 QTLs, v_{QTL}). QTL regions were marked from the marginal posterior distributions of the six simulations and
275 consensus QTL positions identified at the peaks of the summed posterior intensities profiles over the six
276 simulations. QTL regions were named by the concatenation of population ID (LM, YBI or GB which refers to the
277 grouped LM and YBI populations), the linkage group and the peaks separated by “@”. For each consensus QTL,
278 QTL genotypes for all individuals in the pedigree were predicted based on the vectors of QTL genotype posterior
279 probabilities extracted from the FlexQTL output “MQTRegionsGTP.csv”. QTL genotypes values were calculated
280 as $[(0 * P_{qq}) + (1 * P_{Qq}) + (2 * P_{QQ})]$, with P the probability associated with the qq , qQ and QQ QTL genotypes,
281 q being the favorable allele in this case. The continuous [0,2] values of the QTL genotypes were converted into
282 discrete values {0,1,2} using the following threshold: values in the ranges [0,0.7],]0.7,1.3[and [1.3,2], were
283 assigned to 0, 1 and 2 respectively, corresponding to individuals carrying homozygous favorable, heterozygous or
284 homozygous unfavorable disease resistance alleles at the respective QTL regions considered.

285 2.4.2. Stepwise QTL model selection

286 In order to obtain a full QTL model fitted on the raw phenotypic data, QTL results from different modeling and
 287 random seeds were aggregated using stepwise model selection. The stepwise approach was applied on QTL
 288 genotypes vectors tested in the LMM model (see Phenotypic data section), following the procedure of the
 289 stepwiseqtl function of the R/qtl package (Broman and Sen 2009). First, a main effect QTL model was selected by
 290 testing the QTL genotype vectors in the LMM model with sequentially, a forward selection and a backward
 291 elimination. Model selection was based on the Akaike information criterion (AIC, Akaike 1998) using the full
 292 loglikelihood (Verbyla 2019). Similarly, the main effect QTL model was extended to the complete QTL model by
 293 first testing the interactions between QTLs and both QTL and CL (fixed effects), and second with the GA genetic
 294 background (random effect). Stepwise model selection was performed using ASReml-R software (Butler et al.
 295 2007, V4).

296 **2.5. Field evaluation of pre-nursery QTL**

297 The relationships between *Ganoderma* genetic resistance in pre-nursery and field conditions were investigated
 298 using the census of disease status of the La Mé parents planted in genealogical gardens (see plant material section).
 299 The *Ganoderma* infection status was recorded biannually on 219 LM individuals planted in 1974 (5), 1976 (11),
 300 1996 (5), 1997(107), 1998 (1), 1999 (47), 2001 (20) and 2003 (23) in six different blocks at Bangun Bandar estate,
 301 Indonesia. The disease status recording began within the three years after planting in the case of plantation after
 302 1990 and in the 2000s for older plantings, and the last observation was recorded in 2018. *G. boninense* disease
 303 symptoms were scored blindly based on a six-level scale as described in Tisné et al. (2017). The appearance of the
 304 first *Ganoderma* symptom (T1S, first observation of score 2–6) was recorded and the associated time was
 305 considered as survival time, i.e., time from planting to the time the event occurred. The survival data were analyzed
 306 using the Cox model integrating a fixed effect for the date of planting:

$$307 \lambda(t, X) = \lambda_0(t)e^{X\beta} \quad (1)$$

308 where t is the time to the event or censoring, λ_0 denotes the baseline hazard function, X is the $n \times d$ design
 309 matrix relating the survival outcome for individuals to date of planting effects ($d = 8$) and $\beta = (\beta_1, \dots, \beta_d)$ is a
 310 $d \times 1$ unknown vector.

311 The effects of pre-nursery QTL were evaluated using the likelihood ratio test, for which the limiting distribution
 312 follows a chi-squared distribution, between the model (1) and the following model (2):

$$313 \lambda(t, X) = \lambda_0(t)e^{X\beta + X_q q} \quad (2)$$

314 with X_q being the {0,1,2} vector of pre-nursery-based QTL genotypes for the individuals and q the QTL effect.
315 The analysis was performed with R software version 3.2.3 (Team 2012) and the *survival* package (Therneau 2015).

316

317 **3. Results**

318 3.1. Segregation of *Ganoderma* resistance in the GB population

319 Resistance to *Ganoderma* disease was tested in pre-nursery trials on 3,792 GA×GB crosses. On average, 30.8%
320 of oil palm seedlings per cross presented disease symptoms at the end of the trial, ranging from 3 to 92.5% among
321 the different crosses (Fig. 2a). Both LMM or GLMM models led to very similar predictions of GCA for the GB
322 parents ($r=0.97$). Predictions of GCA were higher in YBI genetic background compared to LM, indicating higher
323 susceptibility of the YBI background tested in this study (Fig 2b-c). Within genetic backgrounds, the distribution
324 of GCA indicated segregation of quantitative resistance among founders, with mainly additive effects. Indeed, in
325 LM genetic background, LM_1 self-pollinated individuals were the most resistant, and all the combinations of
326 LM_1 and the alternative founders LM_2 or LM_3 showed higher resistance than the populations derived from
327 self-pollinations of LM_2 and LM_3 (Fig. 2b-c). Similarly in YBI, YBI_3 was the least resistant genetic
328 background, but its combination with YBI_2 improved the resistance of derived individuals. Even in narrow
329 genetic bases, i.e. self-pollinated progenies of the most recent generation, there was still segregation of the
330 resistance supporting the quantitative nature of *Ganoderma* resistance (Fig. 2b-c).

331 3.2. Genetic bases of *Ganoderma* resistance in pre-nursery trials

332 QTL mapping of the *Ganoderma* disease resistance in the GB population was performed using a Bayesian
333 approach. Cumulating both modeling and the three random seeds per model, the number of QTLs was 125
334 considering all the marked QTL regions found by FlexQTL, regardless the $2\ln BF$ threshold (supp. Table 3). These
335 125 QTL corresponded to around 20 QTLs on average per simulation. The QTLs were distributed in 30 consensus
336 regions covering every linkage group (LG), with overall, a similar pattern between the different simulations (Fig.
337 3). Among these 29 QTL regions, 11 located on LG 1, 5, 6, 8, 9, 10, 12, 13 and 16 were identified consistently in
338 the six simulations. The QTL mapping performed separately in LM and YBI revealed different QTL patterns
339 between them: consistent QTL regions on LG 1, 6, 10, 12 and 13 segregated in the LM genetic background while
340 the regions were located on LG 5, 8, 9 and 10 in the YBI genetic background (Supporting Information Figure S2).
341 The average length of the QTL interval was around 25 cM (4-107 cM). Considering QTL genotypes in the 30

342 consensus QTL regions, there were on average, 35, 41 and 24% of *QQ*, *Qq* and *qq* genotypes respectively, in the
343 GB population, q being the favorable allele in this case.

344 Stepwise model selection was performed based on the QTL genotype vectors calculated for the 30 consensus QTL
345 regions. The first step fitted the LMM and indicated that the components related to the genetic effects represented
346 21% of total phenotypic variation, while 6% corresponded to the GCA of the GB individuals (Fig. 4). The final
347 QTL model retained four main effect QTL on LG 8, 9, 10 and 16, and one in interaction with the GA genetic
348 background on LG 6 (Fig. 4, supp. Table 4). Adding either the main effect or interacting QTLs in the LMM in the
349 different steps did not change the values of the non-genetic components, whereas the GCA_{GB} was reduced to 1%.
350 Including the interaction between the QTL on LG6 and the GA genetic background reduced both the values of the
351 SCA and the GCA_{GA} components. The partial determination coefficients computed for each QTL ranged from
352 0.05-2% of the total phenotypic variance, corresponding to 3-9% of genetic variance.

353 3.3. Effects of pre-nursery-based QTL on field *Ganoderma* resistance in the La Mé parents

354 The effects of the QTL identified using the pre-nursery data on GA×GB crosses were evaluated in the field where
355 219 LM parents included in the pre-nursery study were planted and underwent natural, uncontrolled *Ganoderma*
356 infection. The time of the first *Ganoderma* symptom appearance (T1S) was modeled using Cox regression with
357 the date of planting as covariate ($P < 0.01$). The effect of the percentage of favorable alleles per individual among
358 the 21 QTL regions identified in the LM genetic background (range 28-75%) was first assessed to evaluate the
359 global trend between pre-nursery and field conditions. The percentage of favorable alleles effect was not found to
360 be significant ($P=0.2$), but Kaplan-Meier estimates of survival showed consistency between the pre-nursery and
361 field QTL effects, a higher percentage of favorable alleles increased the probability of survival (Fig. 5a). Hence,
362 the individuals with less than 50% of favorable alleles were twice more affected by *Ganoderma* 20 years after
363 planting than individuals with more than 50% of favorable alleles (Fig. 5a). Then QTL genotype vectors, predicted
364 either GB or LM populations, were tested one at a time as covariates in the Cox model. The level of statistical
365 evidence of QTL effects between pre-nursery and field data was not correlated and significant QTL effects were
366 found for both a high (LG 9) or low (LG 4, 15) level of evidence in pre-nursery conditions (Fig. 5b). However the
367 direction of effects between field and pre-nursery effects was consistent for 78% of the QTLs, and for 89% when
368 a P -value=0.05 threshold was applied in the Cox model (Fig. 5b, Supporting Information Figure S3).

369

370 4. Discussion

371 Marker assisted selection (MAS) has a great potential for plant breeding and has been widely used for many crops
372 with substantial achievements, especially for resistance to biotic stresses (Muranty et al. 2014). MAS should be
373 particularly useful for perennial crops with a long breeding cycle and high phenotyping costs like oil palm, despite
374 the identified biological, socioeconomic or technical issues (Muranty et al. 2014). In this paper, we report the proof
375 of concept of an efficient *in silico* QTL mapping approach based on data collected in an ongoing breeding program.
376 This allowed us to gain valuable insights into the genetic architecture of *Ganoderma* resistance and the
377 transferability between field and pre-nursery results, as a basis for a future MAS.

378 **4.1. Opportunities and issues of QTL mapping using data from breeding programs**

379 Breeding programs for perennials are inherently geared towards long-term work with extensive data recording.
380 This make them highly suited to the *in silico* approach, which is likely to improve the statistical properties of QTL
381 detection through the increase in population size and diversity compared to conventional biparental populations.
382 However, the specificity of the data from breeding programs, such as the extent of non-genetic effects due to long-
383 term data or the genetic and phenotypic design unbalances due to the selection process, could reduce the expected
384 benefits of QTL detection, namely its power and the accuracy of QTL location and QTL effect estimation
385 (Würschum 2012). Hence, these datasets require a first stage of statistical modeling to account for several non-
386 genetic effects and to obtain genotypic values. Thanks to their flexibility, mixed models are ideal tools to handle
387 several types of data and effects (Smith et al. 2005). We used two types of mixed models, LMM and GLMM that
388 enabled us to predict the GCA of genotyped individuals while accounting for confounding effects. We
389 subsequently used these GCA values in FlexQTL because this software requires only one value per genotyped
390 individual whereas they were progeny tested in the pre-nursery trials. Such a two-stage approach could affect QTL
391 results so one-stage approaches are preferred when possible (Xue et al. 2017; Barrasso et al. 2019). The two types
392 of mixed model used in this study did not lead to major differences in the QTLs identified, and a one-stage IBD-
393 based variance component approach previously reported for production traits (IBD-VC, Tisné et al. 2015) that we
394 used on pre-nursery *Ganoderma* data also produced similar results (data not shown). However, the calculation
395 time requirement for the IBD-VC is an obstacle to a proper estimation of the significance threshold by permutation
396 and a multi-QTL mapping procedure, which made us favor the approach presented.

397 Few studies have assessed the effects of the dataset features on QTL detection. In barley, using GWAS with an
398 unbalanced dataset, the false positive rate was increased, whereas one-stage analysis performed better (Wang et
399 al. 2012). In durum wheat, a GWAS performed both on an unbalanced and balanced dataset from a breeding

400 program showed major overlapping of selected SNP (Johnson et al. 2019). In diploid potato, a dataset grouping
401 F3 families under selection was analyzed using either GWAS, stratified linkage or IBD based approaches that led
402 to consistent QTL detection, but revealed issues concerning the QTL allele frequencies that could affect the results
403 (Korontzis et al. 2020). In our study, the population studied could be genetically biased due to prior selection of
404 the crosses tested for *Ganoderma* resistance based on yield related traits. However, inspection of QTL genotype
405 frequencies showed that there were no depleted allelic classes among the QTL retained in the stepwise model
406 selection. Moreover, the QTL genotype vectors predicted at the QTL regions were not correlated for the different
407 linkage groups, indicating little segregation distortion that could have arisen due to the selection process.

408 Concerning the accuracy of QTL location, the increased population size allowed by the *in silico* approach should
409 reduce the QTL interval thanks to the increased number of recombinations. In this proof of concept study, we
410 chose to genotype the population using well characterized SSR markers in order to be able to connect the results
411 with previous ones obtained with related populations. However, the QTL intervals were much larger than in other
412 studies using FlexQTL on populations of similar size but with thousands of markers, indicating that the density
413 was insufficient to mark them accurately. The large QTL regions could probably be considerably reduced thanks
414 to the favorable genetic design and we are currently performing high-density SNP genotyping to achieve this
415 objective. Beyond this limitation, the use of FlexQTL was particularly interesting: the use of IBD information
416 mitigates the effect of low density genotyping, and the prediction of QTL genotypes offers the opportunity to use
417 them in subsequent analyses. Hence, we were able to select a full QTL model using the raw data by testing main
418 and interaction effects, and to assess the effects of pre-nursery QTL in the field. As reported by Verma and
419 Whitaker (2018), QTL genotypes have great potential in the breeding context, for example, to predict QTL alleles
420 for unobserved individuals in the breeding program based only on their marker and pedigree information, and then
421 their expected resistance level.

422 **4.2. Insights into the genetic architecture of *Ganoderma* resistance in oil palm**

423 A first insight into genetic architecture came from the variance decomposition using the sire and dam mixed model
424 designed for the analysis of the data on GA×GB hybrids. The genetic component, i.e. GCA in both heterotic groups
425 and SCA, represented around 20% of the total phenotypic variance, which was expected due to the consistent
426 genetic resistances identified in contrasted crosses or clones, balanced by the moderate repeatability of the
427 screening tests (Durand-Gasselin et al. 2018). More surprising, the variance assigned to the GA pedigree was
428 double that for the GB pedigree, while the pure parental GB genetic backgrounds are both more resistant and

429 exhibit more resistance variability than GA backgrounds (Durand-Gasselin et al. 2018). This could be an artefact
430 of the unbalanced number of parents screened between heterotic groups and further investigation is needed to
431 accurately estimate their relative contribution to the GA×GB resistance. The variance associated with SCA effect
432 was 20% of the genetic variance and one QTL×genetic background interaction was retained, while well supported
433 previous observations indicated that resistance was mainly additive, both in pre-nursery and field trials (Durand-
434 Gasselin et al. 2018). Again, this could be an artefact, as only the GB pedigree was genotyped for this study but
435 further analyses using both heterotic groups will allow us to estimate the proportion of variance due to GA×GB
436 interaction and identifying underlying QTL.

437 The distributions of the GCA of GB individuals showed segregation of the *Ganoderma* resistance throughout the
438 pedigree, even in the most inbred generations. Consequently, we identified a large number of putative QTL regions
439 using FlexQTL, with weak to moderate effects. This partially reflects the composition of the GB that grouped two
440 contrasted populations, LM and YBI, which displayed distinct QTL patterns when analyzed separately. However,
441 even when we focused on a restricted genetic background, the large number of putative QTL found despite the
442 reduced population size confirm the quantitative nature of *Ganoderma* resistance (quantitative disease resistance,
443 QDR). Thus, the marked difference in *Ganoderma* resistance consistently observed between the four full-sib
444 founders of the studied LM pedigree (Durand-Gasselin et al. 2018) is rather the consequence of a better
445 combination of many favorable alleles than of a limited number of major QTLs. The numerous QTL found and
446 the dissimilarity of QTL patterns between the LM and YBI genetic backgrounds is likely due to either the
447 *Ganoderma* bio-trophic pathogenesis that induce contrasted transcriptomic responses (Bahari et al. 2018) or the
448 multiple mechanisms involved in the QDR (Poland et al. 2009). This could explain the few discrepancies observed
449 for some pre-nursery QTL with no effect in the field, and even a QTL with an opposite effect on LG12, considering
450 that such QDR mechanisms are more prone to depend on the age of palms, on the environmental conditions, or on
451 the genetic background surveyed.

452 Inspection of QTL colocalization may validate putative QTL when found for similar traits in independent
453 experiments and inform QTL pleiotropy or linkage for different traits. Pleiotropy is especially worth investigating
454 for QDR to obtain insights into possible underlying mechanisms and, together with linkage, on the resulting trade-
455 off with other traits of interest (Nelson et al. 2018). To date, only two genetic mapping studies have been reported
456 on *Ganoderma* resistance. The first analyzed data from a nursery test involving one resistant and two susceptible
457 progenies, with a similar genetic background (Deli×YBI) and common markers to our study (Hama-Ali et al.
458 2015). Despite the limited scope of the study, i.e. involving only 79 individuals genotyped with 58 SSRs, Hama-

459 Ali et al. (2015) identified two significant markers on LG2 and seven in the same QTL regions as in our study,
460 what is more, in equivalent populations, YBI and GB respectively. The second study used field data recorded on
461 a multi-parental GA×GB population involving four GB founders that were the same as in the present study (*Eg9PP*
462 population, Tisné *et al.* 2017). Four *Ganoderma* resistance loci were identified, two controlling the occurrence of
463 the first *Ganoderma* symptoms (T1S), and two the death of palm trees (TD). Among them, the T1S QTL at the
464 bottom of LG1 collocated with a QTL identified in GB and LM populations in the present study. The *Eg9PP*
465 population and a large-scale genetic trial involving GB parents related to the founder of the present study (NGP
466 population, Tisné *et al.* 2015, Tisné *et al.* 2019) were evaluated in the framework of the breeding program. Hence,
467 data for fruit bunch production, oil extraction rate, and height increment traits were stored in databases, and both
468 populations as well as the population from the present study were genotyped with the same SSR markers from a
469 reference genetic map (Cochard et al. 2015) allowing QTL detection. We observed that among the six *Ganoderma*
470 QTL regions with higher statistical support found in the GB, LM or YBI populations, most collocated with a large
471 number of QTL for other agronomic traits (Tisné, personal communication). The colocalizations were more
472 frequent in the LM population (33) than in the YBI one (15), while they were mostly found with oil extraction rate
473 related traits and bunch number in LM genetic background in contrast with bunch weight and height increment in
474 the YBI one (Tisné, personal communication). These preliminary findings now require further support, in
475 particular by using a high-density SNP genotyping that is currently in progress, but already provide interesting
476 insights into the possible diverse mechanisms underlying the QDR, which could differ considering the genetic
477 backgrounds. This also highlights the benefits of the *in silico* approach assessed in this study that makes it possible
478 to gather information from the entire breeding program for a more comprehensive description of the genetic
479 architecture of traits of interest.

480 **4.3. Advances towards a MAS of *Ganoderma* resistance in oil palm breeding programs**

481 No complete resistance to *Ganoderma* has been identified to date and the results of the present study corroborate
482 previous observations to indicate its quantitative nature (Franqueville et al. 2001; Idris et al. 2004; Durand-Gasselín
483 et al. 2005). Despite the increasing use of QDR to improve the sustainability of disease resistance (Poland et al.
484 2009; Roux et al. 2014) the high number of loci and mechanisms involved makes its selection challenging. This is
485 more acute in the case of oil palm with its long breeding cycle, worsened by the slow *Ganoderma* disease
486 progression. Pre-nursery testing accelerated the screening of genetic material and revealed a genetic component
487 that accounted for about 20% of phenotypic variance, which is generally a favorable level for a MAS perspective
488 (Muranty et al. 2014). A first concern is to insure the consistency of QTL effects between the pre-nursery and field

489 results, like in conventional selection (Durand-Gasselín et al. 2018). We attempted to assess this at the QTL level
490 with the extensive use of the data from the breeding program, including the *Ganoderma* census routinely recorded
491 on seed and genealogical gardens. Following the previous study assessing the *Ganoderma* resistance in field we
492 used a survival analysis approach that provides several advantages (Tisné et al. 2017). Despite the limitations of
493 specific to the data recorded in seed gardens, i.e. mature palms of pure genetic backgrounds in the field vs GA×GB
494 seedlings in pre-nursery and spatio-temporal heterogeneity in the field, the accumulation of favorable pre-nursery
495 QTL alleles improved field resistance. Interestingly, the majority of QTL effect directions were consistent
496 regardless the statistical evidence in pre-nursery. Thus, the many QTL that would not have been detected in the
497 field setup because of a lack of statistical power, were identified in the pre-nursery study and are valuable for a
498 marker-assisted *Ganoderma* resistance selection.

499 Secondly, the quantitative nature of *Ganoderma* resistance identified could hamper the conventional QTL
500 pyramiding approach due to the high number of loci involved, especially considering the long generation time in
501 oil palm. In such a QDR context, the MAS approaches developed for other agronomic quantitative traits are
502 probably more suitable, especially the genomic selection (GS) approach (Poland and Rutkoski 2016). In oil palm,
503 GS has emerged as an efficient MAS method and is being increasingly evaluated for yield improvement (Nyouma
504 et al. 2019). Thus GS statistical models and implementation modes already assessed in oil palm could be
505 transferred or adapted to *Ganoderma* disease related data from the breeding program (Cros et al. 2015, 2017).
506 However, the qualitative/quantitative nature of disease resistance is a continuum (Poland et al. 2009). Despite a
507 large number of QTL regions identified using FlexQTL, only 5 QTL with weak to moderate effects explained
508 almost all the GB GCA component based on pre-nursery data. GS models including information on QTL or genes
509 have been proposed to improve prediction capacity in such situations (Bernardo 2014; Zhang et al. 2014) and
510 should be considered for a GS of implementation in light of the emerging insights into the genetic architecture of
511 *Ganoderma* resistance.

512 A final issue is that selection for *Ganoderma* resistance will need to be combined with resistance to other diseases
513 and cannot be at the expense of other traits of interests. The cost of disease resistance through negative trade-off
514 with performance or fitness was a long-lasting question in model plants but was less investigated in plant breeding
515 (Brown 2002). In the former section, we described colocalization of *Ganoderma* resistance QTL with yield related
516 ones, with a genetic background specificity of these complex patterns. Dealing with multiple traits and multiple
517 genetic background is challenging and the QTL information provided by the *in silico* approach assessed in the
518 present study is very valuable for comprehensive modeling of a MAS strategy. Hence, a recent study in oil palm

519 simulated the outcomes of alternative selection strategies on yield and its components based on their global genetic
520 architecture, including the pleiotropy/linkage and phases between the underlying QTL (Tisné et al. 2019). Virtual
521 individuals and crosses were simulated from the actual founders via meiosis simulations based on the QTL
522 positions identified with FlexQTL, which thus integrated their recombination frequencies. The QTL genotypes
523 predicted in FlexQTL enabled prediction of their multiple trait values and their incorporation in yield based on the
524 QTL effects. This use of QTL genotypes is of prime interest as QTL genotypes can be predicted based on markers
525 alone in any related individual, whether phenotyped or not. In the MAS perspective for *Ganoderma* resistance,
526 this approach would help attenuate possible trade-offs with other traits of interest and optimize the combination of
527 QDR from diverse genetic backgrounds.

528 **5. Conclusion**

529 The cost-effective and efficient *in silico* mapping approach assessed in this study has great potential for the
530 implementation of MAS of traits of interest in oil palm. Its application in the context of *Ganoderma* disease
531 resistance enabled us to use the considerable quantities of data generated in the framework of conventional
532 phenotypic selection to obtain valuable information in the MAS perspective. First, important information on the
533 genetic architecture of resistance to *Ganoderma* disease was obtained, confirming its quantitative nature and
534 identifying the loci involved. In addition, together with other ongoing works, this study sheds light on the
535 relationships between *Ganoderma* resistance and yield related traits that could produce undesirable trade-offs.
536 Second, the consistency between genetic resistance in pre-nursery conditions and in the field was assessed at the
537 QTL level and globally indicated satisfactory portability. However, a few loci deserve careful consideration due
538 to underlying mechanisms that could lead to contrasted phenotypic expression between pre-nursery and field
539 conditions. Finally, this proof-of-concept study provides guidelines for future works on *Ganoderma* disease
540 resistance and should encourage oil palm breeders to use this approach to collectively acquire a better
541 comprehension of its complex genetic architecture.

542 **Declaration of Competing Interest**

543 The authors declare that they have no conflict of interests.

544 **Data availability**

545 The datasets generated and analyzed during the current study are available from the corresponding author.

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556

557 **Supplementary Information**

558 Supporting Information Figure S1: Genetic map of the prenursery GB, LM and YBI oil palm populations.

559 Supporting Information Figure S2: QTL mapping of the *Ganoderma* resistance in the prenursery LM and YBI oil
560 palm populations.

561 Supporting Information Figure S3: Survival curves of the La Mé population in field conditions according to the
562 genotypes of QTL identified based on the pre-nursery data.

563

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783

784 **Figure captions**

785 **Fig. 1** Pedigree of the pre-nursery GB oil palm population. Boxes on the left represent the founders of the La Mé
786 (LM, panel A) and Yangambi (YBI, panel B) populations. Note that the La Mé founders LM_1:4 are full sibs.
787 Other boxes represent full-sib families whose color represents their relation to their genetic background, with the
788 number of individuals in parenthesis. The circled cross symbols represent progenies obtained through self-
789 pollination, and successive self-pollinated progenies keep the same color.

790 **Fig. 2** Distribution of *Ganoderma* disease resistance in the pre-nursery GB oil palm population. Distribution of
791 the percentage of affected individuals in crosses (A), BLUP obtained from random effect of the GCA in GB in a
792 GLMM (B) and LMM (C) for the La Mé (LM) and Yangambi (YBI) populations. Different colors represent
793 different genetic backgrounds.

794 **Fig. 3** QTL mapping of *Ganoderma* resistance in the pre-nursery GB oil palm population. QTL regions marked
795 by FlexQTL software in six independent simulations (LMM and GLMM models, three random starting seeds) (A)
796 and the averaged posterior intensity calculated at a 1 cM grid for the six simulations (B) are plotted along the
797 genome. In panel A, the yellow to red color code scale depict the value of intensity of the corresponding marked
798 QTL regions found in the “MQTRegions.new” FlexQTL output file. In panel B, a white to red color scale indicates
799 the number of marked QTL regions among the six simulations at the corresponding position in the genome.

800 **Fig. 4** Variance components of *Ganoderma* resistance in the pre-nursery screening tests. Variance components are
801 plotted as a percentage of the total phenotypic variance for each of the steps performed in the stepwise selection
802 model. GA/GB: heterotic group A and B; GCA: general combining ability; SCA: Specific combining ability; CL:
803 *Ganoderma* clonal lines; QTL names: see M&M section.

804 **Fig. 5** Pre-nursery QTL effects on *Ganoderma* resistance to natural field infection in the La Mé genetic
805 background. (A) Survival curves of the La Mé population according to the percentage of favorable alleles at the
806 21 La Mé QTL detected in the pre-nursery analysis, the red to green color scale indicates an increasing percentage.
807 Survival estimates are plotted at the time of the first observation of a *Ganoderma* symptom. (B) Scatterplot
808 showing the relationship between the statistical significances of QTL effects in the pre-nursery experiments
809 (posterior intensity, x-axis) and in the field (-log (P-value) from the Cox model, y-axis). QTL originate from QTL
810 mapping using the GB (squares) or LM (triangles) pedigree. Consistency between field and pre-nursery QTL
811 effects was defined for QTL alleles decreasing the number of affected progenies in the pre-nursery trials and
812 delaying the appearance of the first symptom of *Ganoderma*: inconsistent and consistent QTL effects are depicted
813 by green (+) or red (-) symbols, respectively. QTL for which one of the three allelic classes (*QQ*, *Qq* or *qq*) was
814 represented by less than ten individuals are depicted by shaded symbols. QTL names: see M&M section.

Figure 3

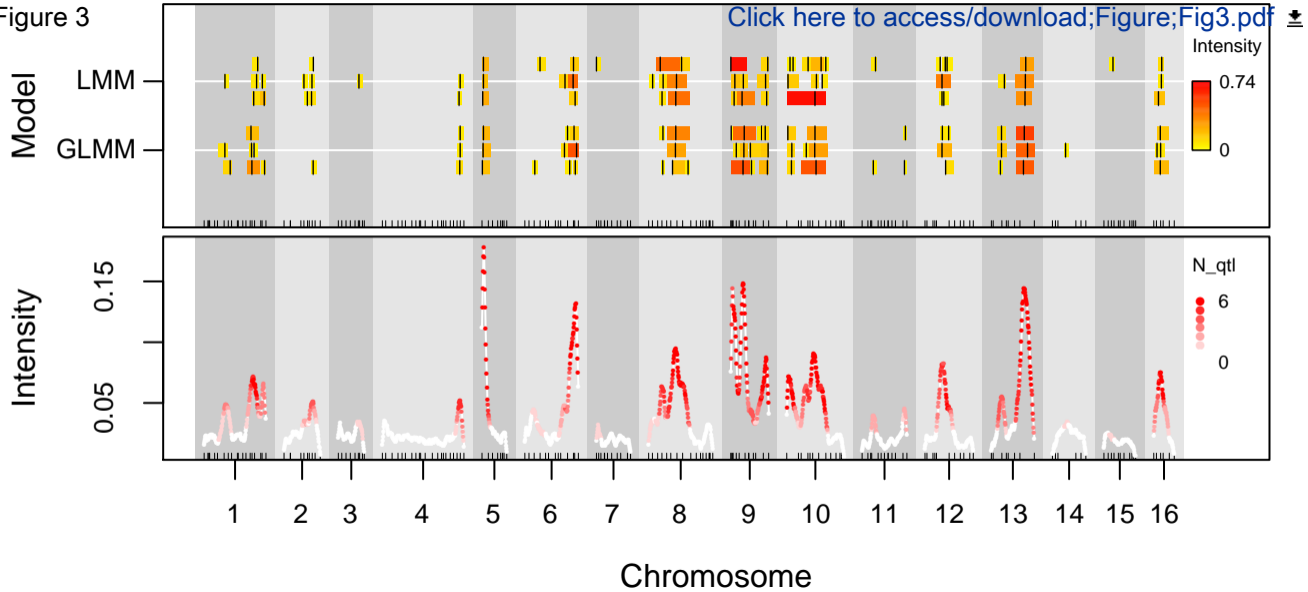


Figure 4

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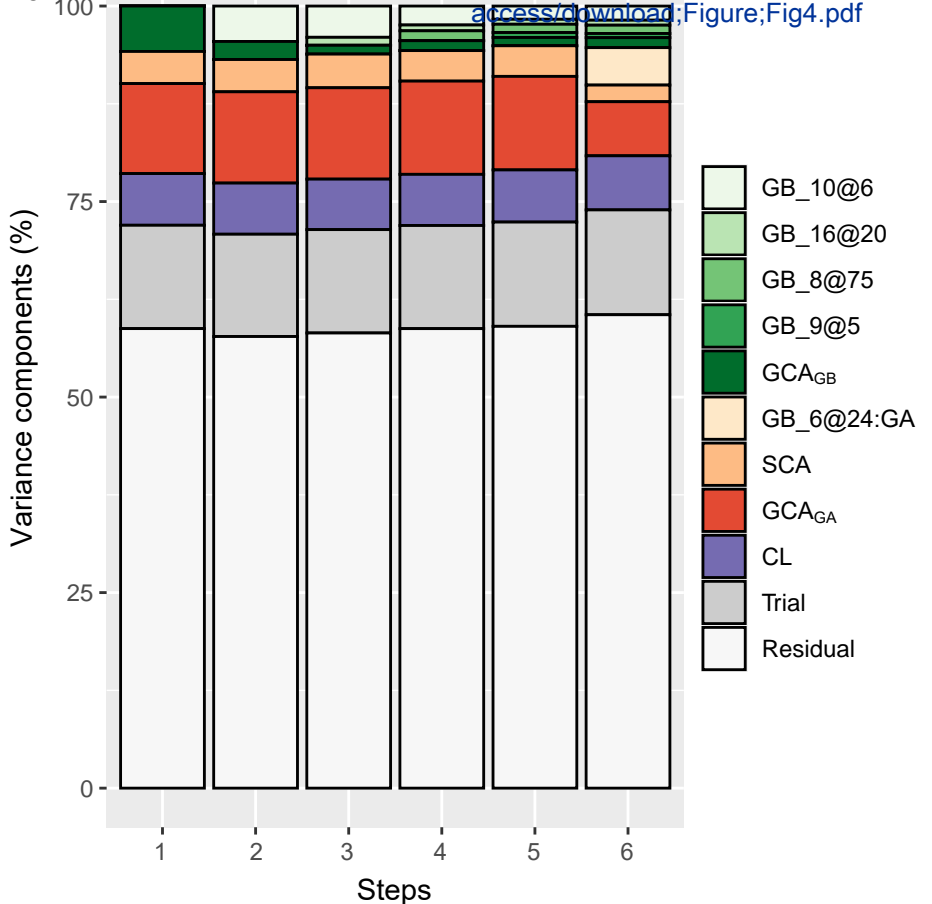
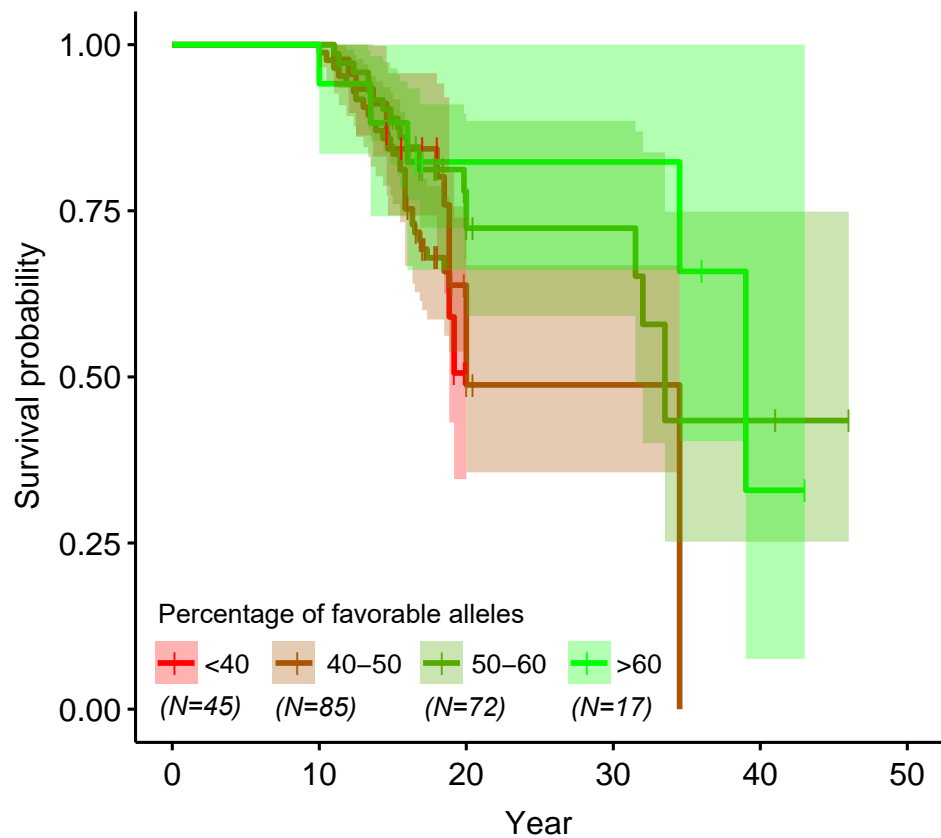


Figure 5
(a)

(b)

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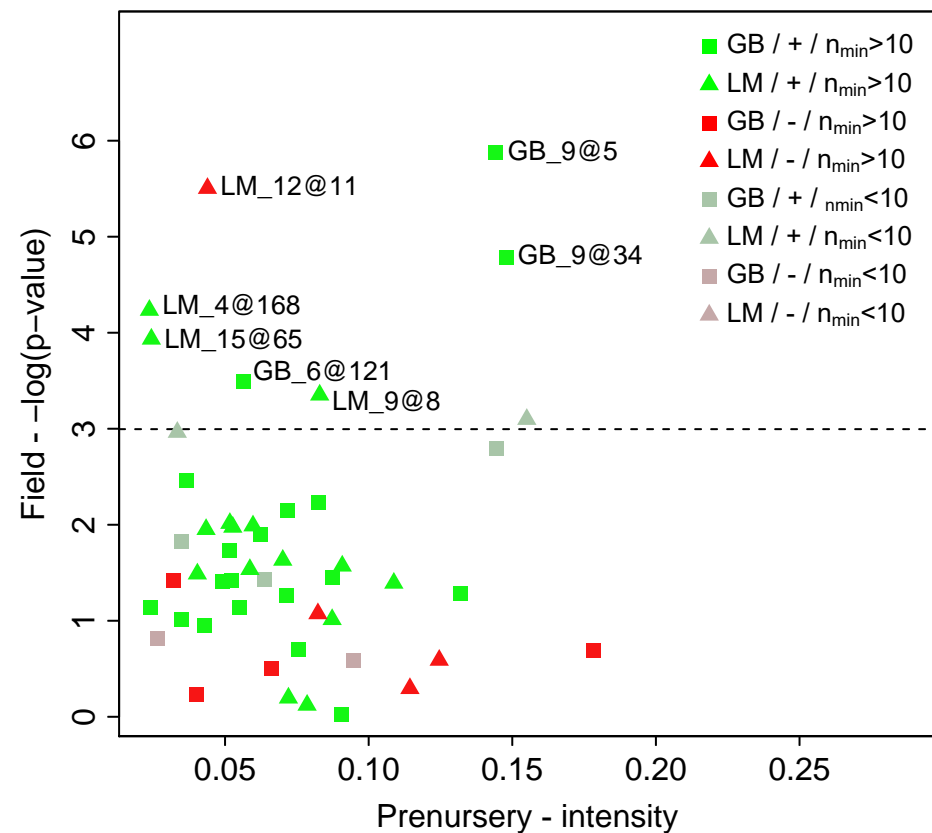


Figure 2

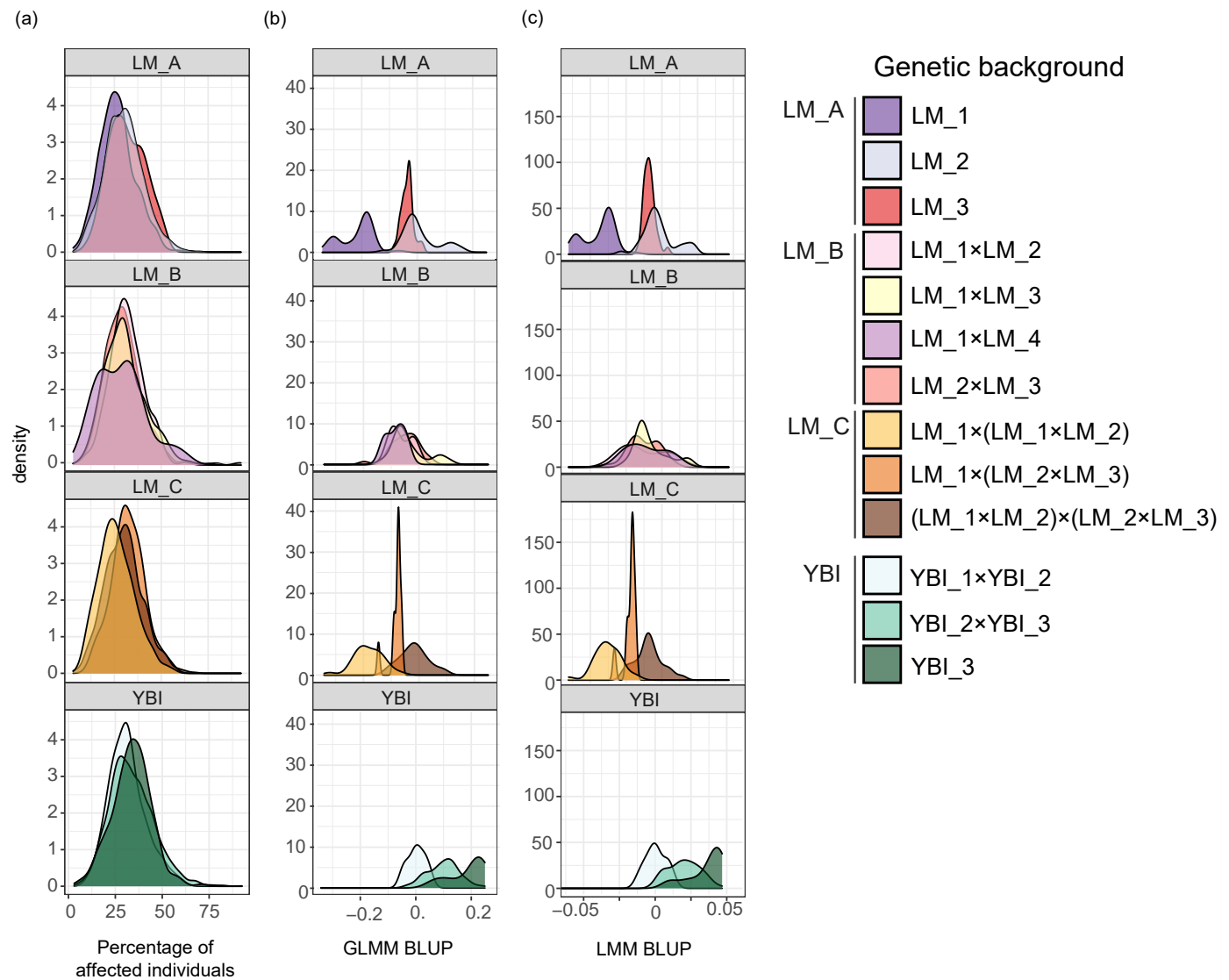
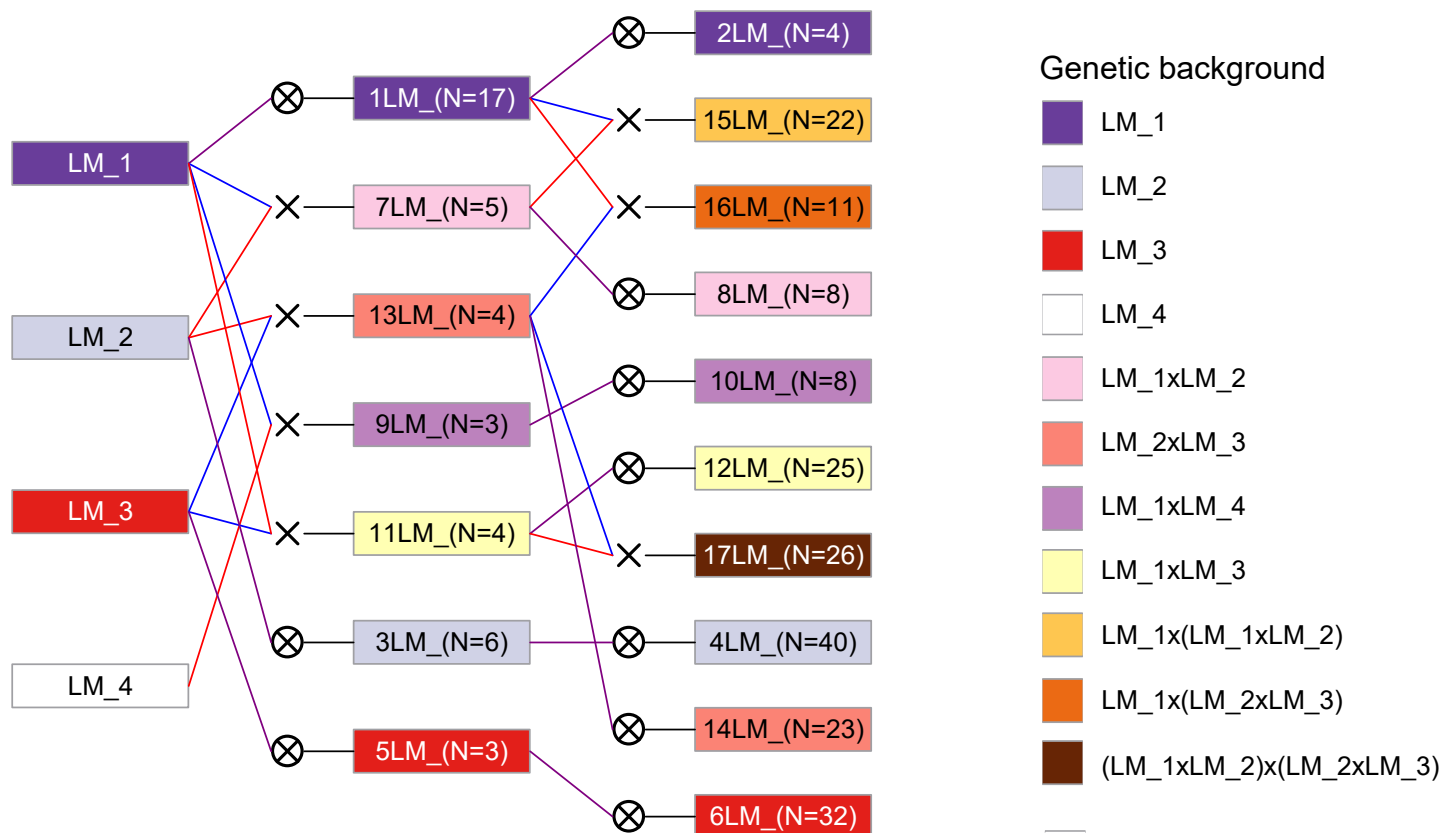
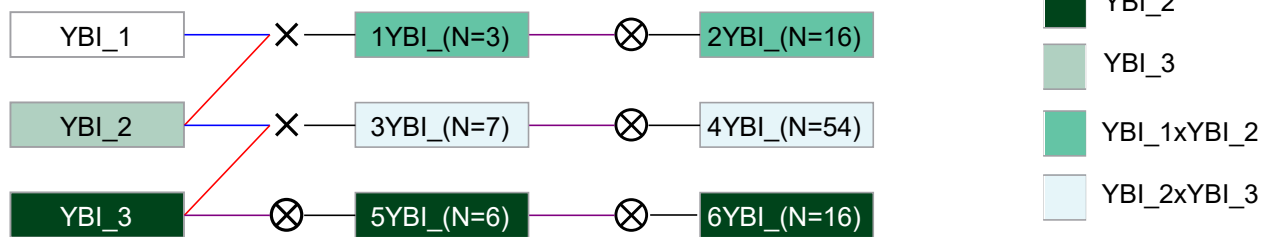
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Figure 1
(a)



(b)



1 **Title Page**

2 **Title**

3 *In silico* QTL mapping in an oil palm breeding program reveals a quantitative and complex genetic resistance to
4 *Ganoderma boninense*

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31

32 **Abstract**

33 Basal stem rot caused by *Ganoderma boninense* is the major threat to oil palm cultivation in South-East Asia,
34 which accounts for 80% of palm oil production worldwide, and this disease is increasing in Africa. The use of
35 resistant planting material as part of an integrated pest management of this disease is one sustainable solution.
36 However, breeding for *Ganoderma* resistance requires long-term and costly research, which could greatly benefit
37 from marker assisted selection (MAS). In this study, we evaluated the effectiveness of an *in silico* genetic mapping
38 approach that took advantage of extensive data recorded in an ongoing breeding program. A pedigree-based QTL
39 mapping approach applied to more than 10 years' worth of data collected during pre-nursery tests revealed the
40 quantitative nature of *Ganoderma* resistance and identified underlying loci segregating in genetic diversity that is
41 directly relevant for the breeding program supporting the study. To assess the consistency of QTL effects between
42 pre-nursery and field environments, information was collected on the disease status of the genitors planted in
43 genealogical gardens and modeled with pre-nursery-based QTL genotypes. In the field, individuals were less likely
44 to be infected with *Ganoderma* when they carried more favorable alleles at the pre-nursery QTL. Our results pave
45 the way for a MAS of *Ganoderma* resistant and high yielding planting material and the provided proof-of-concept
46 of this efficient and cost-effective approach could motivate similar studies based on diverse breeding programs.

47

48 **Keywords**

49 Oil palm, basal stem rot disease, nursery screening test, breeding population, QTL mapping, pedigree-
50 based analysis.

51

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62

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71

72 **1. Introduction**

73 The African oil palm (*Elaeis Guineensis* Jacq.) is the leading oil crop worldwide with a global annual production
74 of around 73 Mt, and accounts for more than 35% of all the edible vegetable oil produced worldwide (USDA
75 statistics, 2019). Oil palm is expected to be able to respond to the global increase in the demand for vegetable oil
76 projected to be 240 Mt in 2050, even higher if its non-food uses are included (Corley 2009). The oil palm sector
77 has agreed on sustainability goals to reach this global demand (Rochmyaningsih 2019), in particular through the
78 certification of sustainable produced palm oil (the Roundtable on Sustainable Palm Oil, RSPO, <https://rspo.org/>).
79 However, pests and diseases threaten palm oil production in all areas of cultivation and contribute to the current
80 yield gap (Woittiez et al. 2017). If it is to achieve the zero-deforestation goal in high conservation value forests
81 included in the RSPO commitments, oil palm will inevitably be cultivated on existing arable lands under increasing
82 pathogen pressure. The integrated pest management (IPM) covers sustainable solutions to this problem including
83 improved plant disease resistance. Oil palm breeders thus needs to focus on developing resistant planting material,
84 while maintaining or even improving oil yield.

85 The basal stem rot disease caused by *Ganoderma boninense* is a major threat in South East Asia, with projections
86 worsening due to climate change (Paterson 2019). This pathogenic fungus is a soil-borne basidiomycete that
87 mainly infects the oil palm when its roots come into contact with infected debris or with the roots of neighbor
88 palms (Rees et al. 2009). *Ganoderma* stem rot disease has a significant effect on oil yield even when only 10-20%
89 of palm trees are infected, and 30-70% of the trees may have died over a typical 25-year planting cycle (Durand-
90 Gasselin et al. 2005; Cooper et al. 2011). To date, no specific interaction and/or complete resistance have been
91 identified in oil palm/*Ganoderma* pathosystem, which is consistent with its hemibiotrophic pathogenic lifestyle.
92 However, observations of contrasted levels of resistance in diverse genetic backgrounds suggest that breeding for
93 quantitative disease resistance (QDR) is a promising solution (Franqueville et al. 2001). Typically, research on
94 perennial plant disease resistance is based on large scale costly field experiments, even more so when investigating
95 QDR. When possible, *ex situ* experiments with controlled inoculation of the pathogen are powerful tools that offer
96 more repeatability and increase both speed and throughput, especially in genetic surveys. In oil palm, such pre-
97 nursery tests were first developed for research on vascular wilt (De Franqueville and Renard 1990), followed by
98 *Ganoderma* in the 2000s (Idris et al. 2004; Breton et al. 2006b; Rees et al. 2007) and are now widely used.
99 However, transferring results to the field can be problematic because of a more complex biotic context, the age
100 specificity of the QDR mechanisms, or the effects of cultural practice management on disease epidemiology.
101 Despite these challenges, by combining field and pre-nursery approaches in long-term works in the framework of
102 an oil palm breeding program, Cirad, its subsidiary PalmElit, and their partners have managed to release planting
103 material that is highly resistant to vascular wilt and intermediate resistant to basal stem rot caused by *Ganoderma*
104 (De Franqueville and Renard 1990; Franqueville et al. 2001; Durand-Gasselin et al. 2005; Breton et al. 2009).
105 Information on the genetic architecture and molecular determinisms of traits of interest could help shorten the long
106 breeding cycle of oil palm, which currently exceeds 20 years, and would be particularly useful in the case of
107 *Ganoderma* disease given the cumbersome nature of field and nursery trials. Marker assisted selection (MAS)
108 based on this information would increase the annual genetic gain thanks to both accelerated evaluation of selection
109 candidates and increased selection intensity by enabling surveys of wider genetic diversity at the same cost (Cros
110 et al. 2015, 2017). Moreover, identification of the genetic bases of resistance to *Ganoderma* could resolve the
111 challenge of breeding for both QDR and yield related traits (Nelson et al. 2018) by using simulation and prediction
112 tools (Tisné et al. 2019). Most molecular studies on *Ganoderma* disease to date have been based on inoculated *vs*
113 non-inoculated seedlings at the pre-nursery stage, with no or low genetic diversity. The first investigations focused
114 on *a priori* selection of candidate resistance genes to fungal diseases (Yeoh et al. 2012, 2013; Tan et al. 2013).

115 Next the genes, proteins and pathways affected by *Ganoderma* infection were identified using broader
116 transcriptomic (Tee et al. 2013; Ho et al. 2016; Bahari et al. 2018; Faizah et al. 2020; Sakeh et al. 2020), proteomic
117 (Al-Obaidi et al. 2014) and metabolomic (Nusaibah et al. 2016) approaches. Considering that *Ganoderma* is a
118 white rot fungus (Paterson 2007), lignin related traits were investigated as putative QDR mechanisms by surveying
119 the response of lignin content and composition to *Ganoderma* infection together with the associated genes
120 (Govender et al. 2017). Lignin related traits and nutritional traits were found to differ in progenies with different
121 levels of resistance to *Ganoderma* (Govender et al. 2020) but the restrained genetic design confounds the effects
122 of genetic and resistance variation.

123 QTL mapping offers an alternative approach that provides information on the genetic architecture based on a
124 relevant genetic diversity, with no *a priori* biological knowledge. The detected loci form the basis of the MAS
125 strategy but also provide insights into the mechanisms and genes involved in the QDR. The first published QTL
126 study reported the analysis of 79 individuals from one resistant and two susceptible families based on 58 simple
127 sequence repeat markers and found alleles associated with *Ganoderma* symptoms (Hama-Ali et al. 2015). More
128 conclusive insights would require much more data, but QTL analyses of oil palm crosses are typically not
129 sufficiently effective due to biological and cost constraints (Jeennor and Volkaert 2014; Lee et al. 2015; Pootakham
130 et al. 2015). This is even more problematic for field studies that are indispensable to assess genetic diversity in an
131 agronomic context, whose implementation is very costly and would result in lower production income due to the
132 disease context. A powerful and cost-effective approach is to directly use the databases compiled in ongoing
133 breeding programs, which are typically large and obtained from diverse relevant genetic backgrounds, to map *in*
134 *silico* the QTLs for the traits of interest (Parrisieux and Bernardo 2004). Despite the potential of this approach,
135 data from breeding programs are unique, mainly because of a complex genetic design that may be biased due to
136 selection, or unbalanced phenotyping coverage. Thus, they require appropriate statistical models for their
137 development and evaluation in contrasted contexts, which are currently an active research topic (Würschum 2012;
138 Garin et al. 2017; Korontzis et al. 2020). In oil palm, an *in silico* QTL mapping approach based on the two step
139 variance component approach considering identity by descent (IBD) information (George et al. 2000; van Eeuwijk
140 et al. 2010) yielded promising results on production traits recorded in large scale evaluation genetic trials (Tisné
141 et al. 2015). This approach was successfully extended to survival data and applied to a multi-parent population to
142 detect *Ganoderma* resistance QTLs in the field, allowing to identify two QTL related to the occurrence of the first
143 disease symptoms, and two related to the death due to *Ganoderma* (Tisné et al. 2017). A Bayesian approach to
144 pedigree based QTL mapping using IBD information was also developed in the 2000s and implemented in the

145 FlexQTL software (van de Weg et al. 2004; Bink et al. 2008). This made it possible to carry out increasing numbers
146 of studies in several crops that share the constraints and potential described above for oil palm, in particular for
147 disease resistance in strawberry (Mangandi et al. 2017; Anciro et al. 2018) or in apple (van de Weg et al. 2018).
148 In this study, we evaluated the potential of an *in silico* approach based on the large existing databases of a long-
149 term oil palm breeding program for the study of *Ganoderma* resistance. We genotyped an existing DNA bank
150 primarily established for identity checking purpose and performed a pedigree-based QTL mapping using data
151 recorded in *Ganoderma* pre-nursery trials over a period of more than ten years. We then assessed the consistency
152 of pre-nursery QTL effects in natural field conditions using a database recording the *Ganoderma* infection status
153 over years for the palms planted in genealogical gardens. Thus, using a cost-effective approach that is directly
154 relevant to the breeding program, we were able to study two major issues, i.e. the genetic architecture and
155 consistency between pre-nursery and field results, paving the way for the implementation of MAS for *Ganoderma*
156 resistant planting material.

157

158 **2. Material and methods**

159 **2.1. Plant material**

160 The palm trees used in this study belong to the oil palm breeding program of Cirad, its subsidiary PalmElit and
161 their partner PT Socfin Indonesia (Indonesia). This breeding program is conducted in a recurrent reciprocal
162 selection scheme with two heterotic groups A and B (GA and GB to produce superior GA×GB hybrid crosses used
163 as commercial planting material (Gascon and De Berchoux 1964; Meunier and Gascon 1972). Individuals from
164 different heterotic groups have complementary yield component traits, with low fruit bunch number and high
165 bunch weight in GA and reciprocally in GB. GA×GB hybrids consequently show a heterosis effect on fruit bunch
166 yield. **Moreover, individuals included in GA are Dura palms, homozygous for the thick alleles of the shell gene**
167 **(Singh et al. 2013) while individuals included in GB are Pisifera (homozygous alternative alleles), the hybrid**
168 **GA×GB being Tenera which is the most productive form with thin shell.** The parental population studied for the
169 *Ganoderma* resistance included only individuals from GB, grouping genetic origins of La Mé (LM, Ivory Coast)
170 and Yangambi (YBI, Republic Democratic of Congo). The GB pedigree used in the pre-nursery analysis comprised
171 372 individuals including founders, with 246/126 from LM/YBI genetic origin respectively and 240/93 genotyped
172 (Supp. Table 1). Among them 200 LM and 83 YBI parents were directly progeny tested for *Ganoderma* resistance
173 in a pre-nursery screening test (Fig. 1). The individuals were distributed over many full-sib families derived from
174 a small number of founders through consecutive crosses or self-pollinations in the framework of the ongoing

175 breeding program (Fig. 1). Among the 372 individuals in the whole pedigree, 219 LM individuals were planted
176 between 1970s and 2000s at the same location (Bangun Bandar, Indonesia) and were used for subsequent field
177 analysis.

178 2.2. Phenotypic data

179 2.2.1. Pre-nursery screening tests

180 An early pre-nursery screening test was developed in the 2000s by Cirad and Socfin Indonesia in the Tanah
181 Gambus estate, Indonesia. The first objective was to speed up the evaluation of genetic resistance to *Ganoderma*
182 of commercial oil palm planting material, using controlled and standardized inoculation of germinated seeds
183 (Breton et al., 2006). The inoculation of germinated seed was performed using a 12 week-old *Ganoderma*-
184 colonized rubber wood block (108 cm³) as inoculum source, that was previously deposited in the nursery polybag
185 before the seeds were planted.

186 A pure dikariotic *Ganoderma boninense* isolate was used in all the trials (NJ), previously harvested from an
187 infected oil palm planted in Bangun Bandar, SOCFINDO estate (Mercière et al. 2015). This isolate was
188 successively regenerated from the bole of young infected seedlings in consecutive pre-nursery trials to provide
189 several dikariotic clonal lines (CL, n=7) over the 10 years of testing. These reactivating steps of the isolate made
190 it possible to avoid the loss of pathogenicity often observed after successive sub-cultures on artificial fungi growth
191 media (Butt et al. 2006). A single pathogen CL was used for all the crosses tested in a single trial. Around 100
192 crosses were assessed simultaneously in each pre-nursery trial. Among them, 20% were control crosses from
193 susceptible, intermediate and resistant genetic backgrounds and were included in all the trials performed. Of the
194 remaining 80% of crosses representing the tested crosses, 50% overlapped two consecutive trials, leading to at
195 least two independent tests per tested cross. Each cross was represented by 100 inoculated germinated seeds
196 clustered in five replicates following the protocol described by Breton et al. (2009). Inoculated seedlings were
197 observed every four weeks for the appearance of the first external disease symptom, on average between 8 and 12
198 weeks after inoculation of the germinated seeds, after which the disease symptoms were recorded at two weekly
199 intervals as (1) infected and (0) if not infected. The trial was stopped when the average percentage of infected
200 seedlings within the group of control crosses reached 30%, usually around 34 weeks after inoculation of the
201 germinated seeds. **This 30% threshold was determined to have the best “discriminating power” between the**
202 **resistant and sensitive control crosses, and so among the tested progenies (Breton et al. 2009).**

203 This study included 102 *Ganoderma* pre-nursery screening test trial, covering 10 years of data recording. The trials
 204 performed between 2007 and 2017 represented the evaluation of 4,017 unique crosses, from either GA×GA,
 205 GA×GB or GB×GB genetic background. Considering that the purpose of this study was to assess the genetic bases
 206 of *Ganoderma* resistance in the commercial genetic material, only the GA×GB crosses were taken into
 207 consideration (n=3,792), derived from 2,037 and 340 individuals from the GA and GB respectively. Each parent
 208 from GB included in the analysis was progeny tested in an average of 20.5 GA×GB crosses.

209 2.2.2. Statistical modeling of pre-nursery data

210 The resistance of the GB individuals was progeny-tested through several GA×GB crosses involving them as GB
 211 parents. The response variable Y considered in this study was the proportion of affected progenies per cross at the
 212 end of the trial. A first step of statistical modeling of Y was necessary to obtain a single value per genotype required
 213 for the QTL analysis while accounting for nuisance effects due to the long-term data. Y was modeled using
 214 generalized linear mixed models (GLMM). Briefly, in a GLMM, Y is assumed to be generated by a particular
 215 distribution in the exponential family. The conditional mean of the distribution μ is linked to a linear predictor η
 216 which contains fixed and random effects, through the inverse link function g^{-1} :

$$217 \quad g(\mu) = \eta = X\beta + Z_T u_T + Z_A u_A + Z_B u_B + Z_C u_C$$

218 where X is a $n \times m$ design matrix relating observations to *Ganoderma boninense* CL fixed effects β where β is
 219 a $m \times 1$ vector ($m = 7$), Z_T is a $n \times t$ design matrix relating observations to trial random effects $u \sim N(0, I\sigma_T^2)$
 220 with u is a $t \times 1$ vector ($t = 102$), Z_C is a $n \times c$ design matrix relating observations to specific combining ability
 221 (SCA) random effects $g_C \sim N(0, I\sigma_C^2)$ where g_C is a $c \times 1$ vector ($c = 3,792$), Z_A and Z_B are $n \times q_A$ and $n \times q_B$
 222 design matrices relating observations to general combining ability (GCA) random effects for GA and GB,
 223 $g_A \sim N(0, A_A\sigma_A^2)$ and $g_B \sim N(0, A_B\sigma_B^2)$ respectively, where g_A and g_B are $q_A \times 1$ and $q_B \times 1$ vectors,
 224 respectively ($q_A = 2,037$ and $q_B = 340$). A_A and A_B are the pedigree-based kinship matrices of GA and GB,
 225 respectively.

226 In our work, we explored two types of distributions: binomial distribution, which is the appropriate one for
 227 proportional data, and normal distribution, for which more derived genetic parameters can be estimated.

228 The first model considers a binomial distribution such as:

$$229 \quad Y_{C,t} | u_t, u_A, u_B, u_C \sim Bin(n_{C,t}, \pi_{C,t})$$

230 where $Y_{c,t}$ is the number of affected progenies in the cross (c) and the trial (t) among the number of inoculated
231 progenies $n_{c,t}$, and $\pi_{c,t}$ is the associated probability.

232 The link function g is the logit such as:

$$233 \quad g(\pi_{c,t}) = \log\left(\frac{\pi_{c,t}}{1-\pi_{c,t}}\right) = \eta_{c,t}$$

234 The second model considers a normal distribution such as:

$$235 \quad Y_{c,t} | u_T, u_A, u_B, u_C \sim N(\eta_{c,t}, \sigma^2)$$

236 where $Y_{c,t}$ is the proportion of affected progenies in the cross (c) and the trial (t), σ^2 is the residual variance, and
237 the link function is the identity. Note that this second model is a linear mixed model (LMM).

238 Both models enabled prediction of the best linear unbiased predictor (BLUP) for each GB individual used in the
239 QTL mapping, A_B being replaced by an identity matrix in order to avoid using the pedigree information that was
240 subsequently used in the QTL analysis. Both statistical models were performed using ASReml-R software (Butler
241 et al. 2007, V4) and resulted in two vectors of BLUP for group B individuals that were used in subsequent QTL
242 mapping analysis.

243

244 **2.3. Molecular data and genetic map construction**

245 The 334 freeze-dried oil palm leaf samples available at the Cirad DNA-bank for the GB individuals included in
246 the analysis were genotyped with 199 SSR markers developed in different studies. Among the 199 markers, 177
247 markers were developed by Cirad (Billotte et al. 2005), two by the Lee et al. (2015), four markers by the Malaysian
248 Palm Oil Board (MPOB) (Zaki et al. 2012) and 18 expressed sequence tags markers were developed by IRD
249 (*Institut de Recherche pour le Développement*) and Cirad (Tranbarger et al. 2012). These markers were selected
250 based on a previous integrated pedigree-based genetic map constructed from a population of related individuals
251 (Cochard et al. 2015). Selection was for a uniform distribution in the genome and the highest level of
252 polymorphism in both LM and YBI genetic backgrounds. **The information concerning markers was gathered in**
253 **the supp. Table 2.** DNA extraction, evaluation of the DNA concentrations and microsatellite fragment
254 amplification were performed using the protocol described in Cochard et al. (2015). Genemapper© V4.1 (Applied
255 Biosystems, USA) software was used to determine the size of the alleles.

256 Three genetic maps were constructed, one for each of LM and YBI population and one integrated map using the
257 pedigree-based linkage mapping software CRI-MAP v2.4 (Green et al. 1990), as described in Cochard et al. (2015).

258 Consistency of marker calling across pedigrees and absence of spurious rates of double recombination events were
259 checked using both CRI-MAP and FlexQTL™, and data were improved where necessary. Genetic maps were
260 drawn using MapChart v2.0 software (Voorrips 2002) and are presented in Supporting Information Figure S1.

261 2.4. Pre-nursery QTL mapping approach

262 QTL mapping of *Ganoderma* disease resistance in pre-nursery conditions followed two main steps. The first step
263 was carried out using a Bayesian approach and a multiple QTL model implemented in FlexQTL™ (Bink et al.
264 2002, 2014, 2008; www.flexqtl.nl) on the pre-nursery data after modeling, in order to identify putative QTL
265 positions and predict the QTL genotypes. The second step consisted in stepwise QTL model selection on the raw
266 pre-nursery data using the predicted QTL genotypes as fixed effects in the LMM.

267 2.4.1. QTL region identification and QTL genotype prediction

268 Six separate QTL analyses, corresponding to the two vectors of GB individual BLUP (see Phenotypic data section)
269 with three different starting random seeds were performed using FlexQTL™. The six QTL analyses were based
270 on a model with additive QTL effects, with the parameters MaximQTL and priorQTL set at 20 and 5 respectively
271 for the Markov chain Monte Carlo simulation. The length of the Markov chains were set at 1 000 000 with a
272 thinning value of 1 000. Using these parameters, the convergence indicators reached satisfying values for each
273 parameter assessed (overall mean, μ , the residual variance, σ_e^2 , the number of QTLs, N_{QTL} , and the variance of
274 QTLs, v_{QTL}). QTL regions were marked from the marginal posterior distributions of the six simulations and
275 consensus QTL positions identified at the peaks of the summed posterior intensities profiles over the six
276 simulations. QTL regions were named by the concatenation of population ID (LM, YBI or GB which refers to the
277 grouped LM and YBI populations), the linkage group and the peaks separated by “@”. For each consensus QTL,
278 QTL genotypes for all individuals in the pedigree were predicted based on the vectors of QTL genotype posterior
279 probabilities extracted from the FlexQTL output “MQTRegionsGTP.csv”. QTL genotypes values were calculated
280 as $[(0 * P_{qq}) + (1 * P_{qQ}) + (2 * P_{QQ})]$, with P the probability associated with the qq , qQ and QQ QTL genotypes,
281 q being the favorable allele in this case. The continuous [0,2] values of the QTL genotypes were converted into
282 discrete values {0,1,2} using the following threshold: values in the ranges [0,0.7],]0.7,1.3[and [1.3,2], were
283 assigned to 0, 1 and 2 respectively, corresponding to individuals carrying homozygous favorable, heterozygous or
284 homozygous unfavorable disease resistance alleles at the respective QTL regions considered.

285 2.4.2. Stepwise QTL model selection

286 In order to obtain a full QTL model fitted on the raw phenotypic data, QTL results from different modeling and
 287 random seeds were aggregated using stepwise model selection. The stepwise approach was applied on QTL
 288 genotypes vectors tested in the LMM model (see Phenotypic data section), following the procedure of the
 289 stepwiseqtl function of the R/qtl package (Broman and Sen 2009). First, a main effect QTL model was selected by
 290 testing the QTL genotype vectors in the LMM model with sequentially, a forward selection and a backward
 291 elimination. Model selection was based on the Akaike information criterion (AIC, Akaike 1998) using the full
 292 loglikelihood (Verbyla 2019). Similarly, the main effect QTL model was extended to the complete QTL model by
 293 first testing the interactions between QTLs and both QTL and CL (fixed effects), and second with the GA genetic
 294 background (random effect). Stepwise model selection was performed using ASReml-R software (Butler et al.
 295 2007, V4).

296 **2.5. Field evaluation of pre-nursery QTL**

297 The relationships between *Ganoderma* genetic resistance in pre-nursery and field conditions were investigated
 298 using the census of disease status of the La Mé parents planted in genealogical gardens (see plant material section).
 299 The *Ganoderma* infection status was recorded biannually on 219 LM individuals planted in 1974 (5), 1976 (11),
 300 1996 (5), 1997(107), 1998 (1), 1999 (47), 2001 (20) and 2003 (23) in six different blocks at Bangun Bandar estate,
 301 Indonesia. The disease status recording began within the three years after planting in the case of plantation after
 302 1990 and in the 2000s for older plantings, and the last observation was recorded in 2018. *G. boninense* disease
 303 symptoms were scored blindly based on a six-level scale as described in Tisné et al. (2017). The appearance of the
 304 first *Ganoderma* symptom (T1S, first observation of score 2–6) was recorded and the associated time was
 305 considered as survival time, i.e., time from planting to the time the event occurred. The survival data were analyzed
 306 using the Cox model integrating a fixed effect for the date of planting:

$$307 \lambda(t, X) = \lambda_0(t)e^{X\beta} \quad (1)$$

308 where t is the time to the event or censoring, λ_0 denotes the baseline hazard function, X is the $n \times d$ design
 309 matrix relating the survival outcome for individuals to date of planting effects ($d = 8$) and $\beta = (\beta_1, \dots, \beta_d)$ is a
 310 $d \times 1$ unknown vector.

311 The effects of pre-nursery QTL were evaluated using the likelihood ratio test, for which the limiting distribution
 312 follows a chi-squared distribution, between the model (1) and the following model (2):

$$313 \lambda(t, X) = \lambda_0(t)e^{X\beta + X_q q} \quad (2)$$

314 with X_q being the {0,1,2} vector of pre-nursery-based QTL genotypes for the individuals and q the QTL effect.
315 The analysis was performed with R software version 3.2.3 (Team 2012) and the *survival* package (Therneau 2015).

316

317 **3. Results**

318 3.1. Segregation of *Ganoderma* resistance in the GB population

319 Resistance to *Ganoderma* disease was tested in pre-nursery trials on 3,792 GA×GB crosses. On average, 30.8%
320 of oil palm seedlings per cross presented disease symptoms at the end of the trial, ranging from 3 to 92.5% among
321 the different crosses (Fig. 2a). Both LMM or GLMM models led to very similar predictions of GCA for the GB
322 parents ($r=0.97$). Predictions of GCA were higher in YBI genetic background compared to LM, indicating higher
323 susceptibility of the YBI background tested in this study (Fig 2b-c). Within genetic backgrounds, the distribution
324 of GCA indicated segregation of quantitative resistance among founders, with mainly additive effects. Indeed, in
325 LM genetic background, LM_1 self-pollinated individuals were the most resistant, and all the combinations of
326 LM_1 and the alternative founders LM_2 or LM_3 showed higher resistance than the populations derived from
327 self-pollinations of LM_2 and LM_3 (Fig. 2b-c). Similarly in YBI, YBI_3 was the least resistant genetic
328 background, but its combination with YBI_2 improved the resistance of derived individuals. Even in narrow
329 genetic bases, i.e. self-pollinated progenies of the most recent generation, there was still segregation of the
330 resistance supporting the quantitative nature of *Ganoderma* resistance (Fig. 2b-c).

331 3.2. Genetic bases of *Ganoderma* resistance in pre-nursery trials

332 QTL mapping of the *Ganoderma* disease resistance in the GB population was performed using a Bayesian
333 approach. Cumulating both modeling and the three random seeds per model, the number of QTLs was 125
334 **considering all the marked QTL regions found by FlexQTL, regardless the 2lnBF threshold (supp. Table 3)**. These
335 125 QTL corresponded to around 20 QTLs on average per simulation. The QTLs were distributed in 30 consensus
336 regions covering every linkage group (LG), with overall, a similar pattern between the different simulations (Fig.
337 3). Among these 29 QTL regions, 11 located on LG 1, 5, 6, 8, 9, 10, 12, 13 and 16 were identified consistently in
338 the six simulations. The QTL mapping performed separately in LM and YBI revealed different QTL patterns
339 between them: consistent QTL regions on LG 1, 6, 10, 12 and 13 segregated in the LM genetic background while
340 the regions were located on LG 5, 8, 9 and 10 in the YBI genetic background (Supporting Information Figure S2).
341 The average length of the QTL interval was around 25 cM (4-107 cM). Considering QTL genotypes in the 30

342 consensus QTL regions, there were on average, 35, 41 and 24% of *QQ*, *Qq* and *qq* genotypes respectively, in the
343 GB population, q being the favorable allele in this case.

344 Stepwise model selection was performed based on the QTL genotype vectors calculated for the 30 consensus QTL
345 regions. The first step fitted the LMM and indicated that the components related to the genetic effects represented
346 21% of total phenotypic variation, while 6% corresponded to the GCA of the GB individuals (Fig. 4). The final
347 QTL model retained four main effect QTL on LG 8, 9, 10 and 16, and one in interaction with the GA genetic
348 background on LG 6 (Fig. 4, [supp. Table 4](#)). Adding either the main effect or interacting QTLs in the LMM in the
349 different steps did not change the values of the non-genetic components, whereas the GCA_{GB} was reduced to 1%.
350 Including the interaction between the QTL on LG6 and the GA genetic background reduced both the values of the
351 SCA and the GCA_{GA} components. The partial determination coefficients computed for each QTL ranged from
352 0.05-2% of the total phenotypic variance, corresponding to 3-9% of genetic variance.

353 3.3. Effects of pre-nursery-based QTL on field *Ganoderma* resistance in the La Mé parents

354 The effects of the QTL identified using the pre-nursery data on GA×GB crosses were evaluated in the field where
355 219 LM parents included in the pre-nursery study were planted and underwent natural, uncontrolled *Ganoderma*
356 infection. The time of the first *Ganoderma* symptom appearance (T1S) was modeled using Cox regression with
357 the date of planting as covariate ($P < 0.01$). The effect of the percentage of favorable alleles per individual among
358 the 21 QTL regions identified in the LM genetic background (range 28-75%) was first assessed to evaluate the
359 global trend between pre-nursery and field conditions. The percentage of favorable alleles effect was not found to
360 be significant ($P=0.2$), but Kaplan-Meier estimates of survival showed consistency between the pre-nursery and
361 field QTL effects, a higher percentage of favorable alleles increased the probability of survival (Fig. 5a). Hence,
362 the individuals with less than 50% of favorable alleles were twice more affected by *Ganoderma* 20 years after
363 planting than individuals with more than 50% of favorable alleles (Fig. 5a). Then QTL genotype vectors, predicted
364 either GB or LM populations, were tested one at a time as covariates in the Cox model. The level of statistical
365 evidence of QTL effects between pre-nursery and field data was not correlated and significant QTL effects were
366 found for both a high (LG 9) or low (LG 4, 15) level of evidence in pre-nursery conditions (Fig. 5b). However the
367 direction of effects between field and pre-nursery effects was consistent for 78% of the QTLs, and for 89% when
368 a P -value=0.05 threshold was applied in the Cox model (Fig. 5b, Supporting Information Figure S3).

369

370 4. Discussion

371 Marker assisted selection (MAS) has a great potential for plant breeding and has been widely used for many crops
372 with substantial achievements, especially for resistance to biotic stresses (Muranty et al. 2014). MAS should be
373 particularly useful for perennial crops with a long breeding cycle and high phenotyping costs like oil palm, despite
374 the identified biological, socioeconomic or technical issues (Muranty et al. 2014). In this paper, we report the proof
375 of concept of an efficient *in silico* QTL mapping approach based on data collected in an ongoing breeding program.
376 This allowed us to gain valuable insights into the genetic architecture of *Ganoderma* resistance and the
377 transferability between field and pre-nursery results, as a basis for a future MAS.

378 **4.1. Opportunities and issues of QTL mapping using data from breeding programs**

379 Breeding programs for perennials are inherently geared towards long-term work with extensive data recording.
380 This make them highly suited to the *in silico* approach, which is likely to improve the statistical properties of QTL
381 detection through the increase in population size and diversity compared to conventional biparental populations.
382 However, the specificity of the data from breeding programs, such as the extent of non-genetic effects due to long-
383 term data or the genetic and phenotypic design unbalances due to the selection process, could reduce the expected
384 benefits of QTL detection, namely its power and the accuracy of QTL location and QTL effect estimation
385 (Würschum 2012). Hence, these datasets require a first stage of statistical modeling to account for several non-
386 genetic effects and to obtain genotypic values. Thanks to their flexibility, mixed models are ideal tools to handle
387 several types of data and effects (Smith et al. 2005). We used two types of mixed models, LMM and GLMM that
388 enabled us to predict the GCA of genotyped individuals while accounting for confounding effects. We
389 subsequently used these GCA values in FlexQTL because this software requires only one value per genotyped
390 individual whereas they were progeny tested in the pre-nursery trials. Such a two-stage approach could affect QTL
391 results so one-stage approaches are preferred when possible (Xue et al. 2017; Barrasso et al. 2019). The two types
392 of mixed model used in this study did not lead to major differences in the QTLs identified, and a one-stage IBD-
393 based variance component approach previously reported for production traits (IBD-VC, Tisné et al. 2015) that we
394 used on pre-nursery *Ganoderma* data also produced similar results (data not shown). However, the calculation
395 time requirement for the IBD-VC is an obstacle to a proper estimation of the significance threshold by permutation
396 and a multi-QTL mapping procedure, which made us favor the approach presented.

397 Few studies have assessed the effects of the dataset features on QTL detection. In barley, using GWAS with an
398 unbalanced dataset, the false positive rate was increased, whereas one-stage analysis performed better (Wang et
399 al. 2012). In durum wheat, a GWAS performed both on an unbalanced and balanced dataset from a breeding

400 program showed major overlapping of selected SNP (Johnson et al. 2019). In diploid potato, a dataset grouping
401 F3 families under selection was analyzed using either GWAS, stratified linkage or IBD based approaches that led
402 to consistent QTL detection, but revealed issues concerning the QTL allele frequencies that could affect the results
403 (Korontzis et al. 2020). In our study, the population studied could be genetically biased due to prior selection of
404 the crosses tested for *Ganoderma* resistance based on yield related traits. However, inspection of QTL genotype
405 frequencies showed that there were no depleted allelic classes among the QTL retained in the stepwise model
406 selection. Moreover, the QTL genotype vectors predicted at the QTL regions were not correlated for the different
407 linkage groups, indicating little segregation distortion that could have arisen due to the selection process.

408 Concerning the accuracy of QTL location, the increased population size allowed by the *in silico* approach should
409 reduce the QTL interval thanks to the increased number of recombinations. In this proof of concept study, we
410 chose to genotype the population using well characterized SSR markers in order to be able to connect the results
411 with previous ones obtained with related populations. However, the QTL intervals were much larger than in other
412 studies using FlexQTL on populations of similar size but with thousands of markers, indicating that the density
413 was insufficient to mark them accurately. The large QTL regions could probably be considerably reduced thanks
414 to the favorable genetic design and we are currently performing high-density SNP genotyping to achieve this
415 objective. Beyond this limitation, the use of FlexQTL was particularly interesting: the use of IBD information
416 mitigates the effect of low density genotyping, and the prediction of QTL genotypes offers the opportunity to use
417 them in subsequent analyses. Hence, we were able to select a full QTL model using the raw data by testing main
418 and interaction effects, and to assess the effects of pre-nursery QTL in the field. As reported by Verma and
419 Whitaker (2018), QTL genotypes have great potential in the breeding context, for example, to predict QTL alleles
420 for unobserved individuals in the breeding program based only on their marker and pedigree information, and then
421 their expected resistance level.

422 **4.2. Insights into the genetic architecture of *Ganoderma* resistance in oil palm**

423 A first insight into genetic architecture came from the variance decomposition using the sire and dam mixed model
424 designed for the analysis of the data on GA×GB hybrids. The genetic component, i.e. GCA in both heterotic groups
425 and SCA, represented around 20% of the total phenotypic variance, which was expected due to the consistent
426 genetic resistances identified in contrasted crosses or clones, balanced by the moderate repeatability of the
427 screening tests (Durand-Gasselin et al. 2018). More surprising, the variance assigned to the GA pedigree was
428 double that for the GB pedigree, while the pure parental GB genetic backgrounds are both more resistant and

429 exhibit more resistance variability than GA backgrounds (Durand-Gasselin et al. 2018). This could be an artefact
430 of the unbalanced number of parents screened between heterotic groups and further investigation is needed to
431 accurately estimate their relative contribution to the GA×GB resistance. The variance associated with SCA effect
432 was 20% of the genetic variance and one QTL×genetic background interaction was retained, while well supported
433 previous observations indicated that resistance was mainly additive, both in pre-nursery and field trials (Durand-
434 Gasselin et al. 2018). Again, this could be an artefact, as only the GB pedigree was genotyped for this study but
435 further analyses using both heterotic groups will allow us to estimate the proportion of variance due to GA×GB
436 interaction and identifying underlying QTL.

437 The distributions of the GCA of GB individuals showed segregation of the *Ganoderma* resistance throughout the
438 pedigree, even in the most inbred generations. Consequently, we identified a large number of putative QTL regions
439 using FlexQTL, with weak to moderate effects. This partially reflects the composition of the GB that grouped two
440 contrasted populations, LM and YBI, which displayed distinct QTL patterns when analyzed separately. However,
441 even when we focused on a restricted genetic background, the large number of putative QTL found despite the
442 reduced population size confirm the quantitative nature of *Ganoderma* resistance (quantitative disease resistance,
443 QDR). Thus, the marked difference in *Ganoderma* resistance consistently observed between the four full-sib
444 founders of the studied LM pedigree (Durand-Gasselin et al. 2018) is rather the consequence of a better
445 combination of many favorable alleles than of a limited number of major QTLs. The numerous QTL found and
446 the dissimilarity of QTL patterns between the LM and YBI genetic backgrounds is likely due to either the
447 *Ganoderma* bio-trophic pathogenesis that induce contrasted transcriptomic responses (Bahari et al. 2018) or the
448 multiple mechanisms involved in the QDR (Poland et al. 2009). This could explain the few discrepancies observed
449 for some pre-nursery QTL with no effect in the field, and even a QTL with an opposite effect on LG12, considering
450 that such QDR mechanisms are more prone to depend on the age of palms, on the environmental conditions, or on
451 the genetic background surveyed.

452 Inspection of QTL colocalization may validate putative QTL when found for similar traits in independent
453 experiments and inform QTL pleiotropy or linkage for different traits. Pleiotropy is especially worth investigating
454 for QDR to obtain insights into possible underlying mechanisms and, together with linkage, on the resulting trade-
455 off with other traits of interest (Nelson et al. 2018). To date, only two genetic mapping studies have been reported
456 on *Ganoderma* resistance. The first analyzed data from a nursery test involving one resistant and two susceptible
457 progenies, with a similar genetic background (Deli×YBI) and common markers to our study (Hama-Ali et al.
458 2015). Despite the limited scope of the study, i.e. involving only 79 individuals genotyped with 58 SSRs, Hama-

459 Ali et al. (2015) identified two significant markers on LG2 and seven in the same QTL regions as in our study,
460 what is more, in equivalent populations, YBI and GB respectively. The second study used field data recorded on
461 a multi-parental GA×GB population involving four GB founders that were the same as in the present study (*Eg9PP*
462 population, Tisné *et al.* 2017). Four *Ganoderma* resistance loci were identified, two controlling the occurrence of
463 the first *Ganoderma* symptoms (T1S), and two the death of palm trees (TD). Among them, the T1S QTL at the
464 bottom of LG1 collocated with a QTL identified in GB and LM populations in the present study. The *Eg9PP*
465 population and a large-scale genetic trial involving GB parents related to the founder of the present study (NGP
466 population, Tisné *et al.* 2015, Tisné *et al.* 2019) were evaluated in the framework of the breeding program. Hence,
467 data for fruit bunch production, oil extraction rate, and height increment traits were stored in databases, and both
468 populations as well as the population from the present study were genotyped with the same SSR markers from a
469 reference genetic map (Cochard et al. 2015) allowing QTL detection. We observed that among the six *Ganoderma*
470 QTL regions with higher statistical support found in the GB, LM or YBI populations, most collocated with a large
471 number of QTL for other agronomic traits (Tisné, personal communication). The colocalizations were more
472 frequent in the LM population (33) than in the YBI one (15), while they were mostly found with oil extraction rate
473 related traits and bunch number in LM genetic background in contrast with bunch weight and height increment in
474 the YBI one (Tisné, personal communication). These preliminary findings now require further support, in
475 particular by using a high-density SNP genotyping that is currently in progress, but already provide interesting
476 insights into the possible diverse mechanisms underlying the QDR, which could differ considering the genetic
477 backgrounds. This also highlights the benefits of the *in silico* approach assessed in this study that makes it possible
478 to gather information from the entire breeding program for a more comprehensive description of the genetic
479 architecture of traits of interest.

480 **4.3. Advances towards a MAS of *Ganoderma* resistance in oil palm breeding programs**

481 No complete resistance to *Ganoderma* has been identified to date and the results of the present study corroborate
482 previous observations to indicate its quantitative nature (Franqueville et al. 2001; Idris et al. 2004; Durand-Gassel
483 et al. 2005). Despite the increasing use of QDR to improve the sustainability of disease resistance (Poland et al.
484 2009; Roux et al. 2014) the high number of loci and mechanisms involved makes its selection challenging. This is
485 more acute in the case of oil palm with its long breeding cycle, worsened by the slow *Ganoderma* disease
486 progression. Pre-nursery testing accelerated the screening of genetic material and revealed a genetic component
487 that accounted for about 20% of phenotypic variance, which is generally a favorable level for a MAS perspective
488 (Muranty et al. 2014). A first concern is to insure the consistency of QTL effects between the pre-nursery and field

489 results, like in conventional selection (Durand-Gasselín et al. 2018). We attempted to assess this at the QTL level
490 with the extensive use of the data from the breeding program, including the *Ganoderma* census routinely recorded
491 on seed and genealogical gardens. Following the previous study assessing the *Ganoderma* resistance in field we
492 used a survival analysis approach that provides several advantages (Tisné et al. 2017). Despite the limitations of
493 specific to the data recorded in seed gardens, i.e. mature palms of pure genetic backgrounds in the field vs GA×GB
494 seedlings in pre-nursery and spatio-temporal heterogeneity in the field, the accumulation of favorable pre-nursery
495 QTL alleles improved field resistance. Interestingly, the majority of QTL effect directions were consistent
496 regardless the statistical evidence in pre-nursery. Thus, the many QTL that would not have been detected in the
497 field setup because of a lack of statistical power, were identified in the pre-nursery study and are valuable for a
498 marker-assisted *Ganoderma* resistance selection.

499 Secondly, the quantitative nature of *Ganoderma* resistance identified could hamper the conventional QTL
500 pyramiding approach due to the high number of loci involved, especially considering the long generation time in
501 oil palm. In such a QDR context, the MAS approaches developed for other agronomic quantitative traits are
502 probably more suitable, especially the genomic selection (GS) approach (Poland and Rutkoski 2016). In oil palm,
503 GS has emerged as an efficient MAS method and is being increasingly evaluated for yield improvement (Nyouma
504 et al. 2019). Thus GS statistical models and implementation modes already assessed in oil palm could be
505 transferred or adapted to *Ganoderma* disease related data from the breeding program (Cros et al. 2015, 2017).
506 However, the qualitative/quantitative nature of disease resistance is a continuum (Poland et al. 2009). Despite a
507 large number of QTL regions identified using FlexQTL, only 5 QTL with weak to moderate effects explained
508 almost all the GB GCA component based on pre-nursery data. GS models including information on QTL or genes
509 have been proposed to improve prediction capacity in such situations (Bernardo 2014; Zhang et al. 2014) and
510 should be considered for a GS of implementation in light of the emerging insights into the genetic architecture of
511 *Ganoderma* resistance.

512 A final issue is that selection for *Ganoderma* resistance will need to be combined with resistance to other diseases
513 and cannot be at the expense of other traits of interests. The cost of disease resistance through negative trade-off
514 with performance or fitness was a long-lasting question in model plants but was less investigated in plant breeding
515 (Brown 2002). In the former section, we described colocalization of *Ganoderma* resistance QTL with yield related
516 ones, with a genetic background specificity of these complex patterns. Dealing with multiple traits and multiple
517 genetic background is challenging and the QTL information provided by the *in silico* approach assessed in the
518 present study is very valuable for comprehensive modeling of a MAS strategy. Hence, a recent study in oil palm

519 simulated the outcomes of alternative selection strategies on yield and its components based on their global genetic
520 architecture, including the pleiotropy/linkage and phases between the underlying QTL (Tisné et al. 2019). Virtual
521 individuals and crosses were simulated from the actual founders via meiosis simulations based on the QTL
522 positions identified with FlexQTL, which thus integrated their recombination frequencies. The QTL genotypes
523 predicted in FlexQTL enabled prediction of their multiple trait values and their incorporation in yield based on the
524 QTL effects. This use of QTL genotypes is of prime interest as QTL genotypes can be predicted based on markers
525 alone in any related individual, whether phenotyped or not. In the MAS perspective for *Ganoderma* resistance,
526 this approach would help attenuate possible trade-offs with other traits of interest and optimize the combination of
527 QDR from diverse genetic backgrounds.

528 **5. Conclusion**

529 The cost-effective and efficient *in silico* mapping approach assessed in this study has great potential for the
530 implementation of MAS of traits of interest in oil palm. Its application in the context of *Ganoderma* disease
531 resistance enabled us to use the considerable quantities of data generated in the framework of conventional
532 phenotypic selection to obtain valuable information in the MAS perspective. First, important information on the
533 genetic architecture of resistance to *Ganoderma* disease was obtained, confirming its quantitative nature and
534 identifying the loci involved. In addition, together with other ongoing works, this study sheds light on the
535 relationships between *Ganoderma* resistance and yield related traits that could produce undesirable trade-offs.
536 Second, the consistency between genetic resistance in pre-nursery conditions and in the field was assessed at the
537 QTL level and globally indicated satisfactory portability. However, a few loci deserve careful consideration due
538 to underlying mechanisms that could lead to contrasted phenotypic expression between pre-nursery and field
539 conditions. Finally, this proof-of-concept study provides guidelines for future works on *Ganoderma* disease
540 resistance and should encourage oil palm breeders to use this approach to collectively acquire a better
541 comprehension of its complex genetic architecture.

542 **Declaration of Competing Interest**

543 The authors declare that they have no conflict of interests.

544 **Data availability**

545 The datasets generated and analyzed during the current study are available from the corresponding author.

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556

557 **Supplementary Information**

558 Supporting Information Figure S1: Genetic map of the prenursery GB, LM and YBI oil palm populations.

559 Supporting Information Figure S2: QTL mapping of the *Ganoderma* resistance in the prenursery LM and YBI oil
560 palm populations.

561 Supporting Information Figure S3: Survival curves of the La Mé population in field conditions according to the
562 genotypes of QTL identified based on the pre-nursery data.

563

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784 **Figure captions**


785 **Fig. 1** Pedigree of the pre-nursery GB oil palm population. Boxes on the left represent the founders of the La Mé
786 (LM, panel A) and Yangambi (YBI, panel B) populations. Note that the La Mé founders LM_1:4 are full sibs.
787 Other boxes represent full-sib families whose color represents their relation to their genetic background, with the
788 number of individuals in parenthesis. The circled cross symbols represent progenies obtained through self-
789 pollination, and successive self-pollinated progenies keep the same color.

790 **Fig. 2** Distribution of *Ganoderma* disease resistance in the pre-nursery GB oil palm population. Distribution of
791 the percentage of affected individuals in crosses (A), BLUP obtained from random effect of the GCA in GB in a
792 GLMM (B) and LMM (C) for the La Mé (LM) and Yangambi (YBI) populations. Different colors represent
793 different genetic backgrounds.

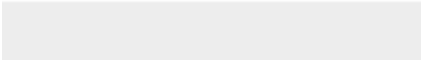

794 **Fig. 3** QTL mapping of *Ganoderma* resistance in the pre-nursery GB oil palm population. QTL regions marked
795 by FlexQTL software in six independent simulations (LMM and GLMM models, three random starting seeds) (A)
796 and the averaged posterior intensity calculated at a 1 cM grid for the six simulations (B) are plotted along the
797 genome. In panel A, the yellow to red color code scale depict the value of intensity of the corresponding marked
798 QTL regions found in the “MQTRegions.new” FlexQTL output file. In panel B, a white to red color scale indicates
799 the number of marked QTL regions among the six simulations at the corresponding position in the genome.

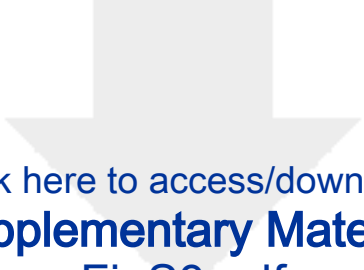
800 **Fig. 4** Variance components of *Ganoderma* resistance in the pre-nursery screening tests. Variance components are
801 plotted as a percentage of the total phenotypic variance for each of the steps performed in the stepwise selection
802 model. GA/GB: heterotic group A and B; GCA: general combining ability; SCA: Specific combining ability; CL:
803 *Ganoderma* clonal lines; QTL names: see M&M section.

804 **Fig. 5** Pre-nursery QTL effects on *Ganoderma* resistance to natural field infection in the La Mé genetic
805 background. (A) Survival curves of the La Mé population according to the percentage of favorable alleles at the
806 21 La Mé QTL detected in the pre-nursery analysis, the red to green color scale indicates an increasing percentage.
807 Survival estimates are plotted at the time of the first observation of a *Ganoderma* symptom. (B) Scatterplot
808 showing the relationship between the statistical significances of QTL effects in the pre-nursery experiments
809 (posterior intensity, x-axis) and in the field (-log (P-value) from the Cox model, y-axis). QTL originate from QTL
810 mapping using the GB (squares) or LM (triangles) pedigree. Consistency between field and pre-nursery QTL
811 effects was defined for QTL alleles decreasing the number of affected progenies in the pre-nursery trials and
812 delaying the appearance of the first symptom of *Ganoderma*: inconsistent and consistent QTL effects are depicted
813 by green (+) or red (-) symbols, respectively. QTL for which one of the three allelic classes (*QQ*, *Qq* or *qq*) was
814 represented by less than ten individuals are depicted by shaded symbols. QTL names: see M&M section.

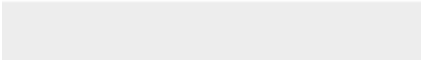


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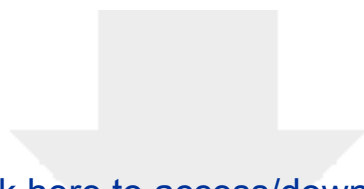




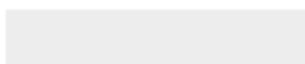
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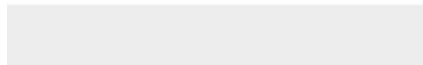


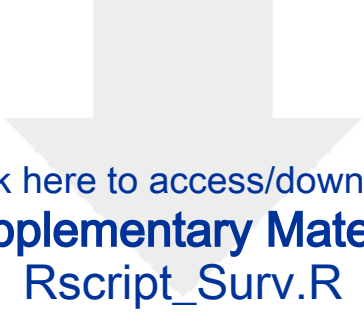
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