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1 Effects of high-pressure treatment on the muscle structure of salmon (*Salmo*
2 *salar*)

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11

12 **Abstract**

13 High pressure (HP) is a non-thermal treatment that is generally used to reduce the
14 microbiological contamination of food products, such as Atlantic salmon (*Salmo salar*).

15 However, HP is known to alter the stability of proteins and can therefore affect the quality
16 of salmon flesh. In this study, the effects of HP treatment for 5 min at 200, 400 and 600

17 MPa on the structure of Atlantic salmon were investigated. Transversal histological

18 sections revealed a decrease in the fibre size from 200 MPa associated with an expansion

19 of the extracellular spaces. Connective tissue was found to be modified from 400 MPa,

20 resulting in an increase in its surface area. Fourier transform infrared (FT-IR)

21 microspectroscopy revealed a reduction in the α -helix content and an increase in the

22 aggregated β -sheet structure content with increasing pressure, reflecting a change in the

23 secondary structure of proteins from 200 MPa.

24

25 **Keywords**

26 High pressure, Salmon, Histology, Infrared microspectroscopy, Protein secondary
27 structure, Muscle structure

28

29 **1. Introduction**

30 Atlantic salmon (*Salmo salar*) is the third most consumed fish in the European Union, with
31 an apparent consumption per capita in 2018 of 2.24 kg live weight equivalent (Directorate-
32 General for Maritime Affairs and Fisheries (European Commission), 2020). However, due
33 to microbial, chemical and enzymatic activity, salmon is a highly perishable product with a
34 shelf life of approximately ten days on ice (Fogarty et al., 2019). Among the numerous
35 technologies that are being investigated to increase this shelf life, de Alba et al. (2019) and
36 Park et al. (2015) have shown a significant effect of high pressure (HP) on microbial
37 reduction. High-pressure treatments have been used since 1990 in Japan to process jams,
38 jellies and sauces and are currently used for processing vegetables, meat and fish
39 products, as well as juices and beverages (Oliveira et al., 2017). This technology
40 represents a particularly interesting preservation technique because it tackles numerous
41 issues of the food industry related to consumer expectations: it does not require the use of
42 food additives to improve the product shelf life, it is more energy efficient than traditional
43 technologies such as cooking (Truong et al., 2015) and it preserves fish nutrients, such as
44 unsaturated fatty acids (Yagiz et al., 2009).

45 Nonetheless, this technology leads to modifications of food biopolymers, especially
46 proteins, as a result of the thermodynamic effects of pressure. Indeed, according to the Le
47 Chatelier principle, the application of HP favours changes causing a decrease in volume
48 (Mozhaev et al., 1994). Consequently, phenomena such as disruption of hydrophobic and
49 electrostatic interactions are promoted by HP, leading to changes in quaternary, tertiary
50 and secondary structures. Covalent bonds, hydrogen bonds and disulphide bridges, on the
51 other hand, have low compressibility and are expected to be resistant to pressure

52 (Aubourg, 2018; Knorr et al., 2006; Mozhaev et al., 1994; Oliveira et al., 2017).

53 Fish muscles are composed of approximately 20% proteins in total and are therefore
54 particularly sensitive to HP. Fish meat consists mainly of myofibrillar proteins, which are
55 responsible for muscle contraction; sarcoplasmic proteins, which are water soluble; and
56 proteins from connective tissue such as collagen, which are insoluble extracellular
57 proteins.

58 HP has been proven interesting to improve the quality of fish by reducing its susceptibility
59 to oxidation (Yagiz et al., 2009) and by enhancing its sensory attributes such as taste,
60 odour and texture (Truong et al., 2015). Moreover, HP has been used to ameliorate the
61 freezing and thawing of different fish species, as it increases the kinetics of ice-water
62 transitions, thus avoiding the formation of large ice crystals (Aubourg, 2018). The protein
63 modifications caused by the pressure have also been used to improve the quality of fish
64 gels (Truong et al., 2017; Uresti et al., 2004). However, several studies have shown
65 unwanted effects of HP on the quality of fish fillets, such as an increased hardness and a
66 change in colour with increased lightness after treatment at 150 MPa (Arnaud et al., 2018;
67 Aubourg et al., 2013), giving the product a cooked appearance.-Even though the effects of
68 HP on the sensory attributes of salmon have been widely described, the molecular
69 mechanisms causing these modifications are still unclear. *In situ* approaches are therefore
70 a powerful tool, as they allow precise analysis of the microscopic and biochemical
71 alterations induced by treatment of the product while taking into account the complexity
72 and high level of organization of the biological structures. The knowledge of the effects of
73 HP on a microscopic and molecular scale should therefore help understanding the
74 changes of fish quality and determine the ideal parameters of treatment to minimize the
75 undesirable impacts of HP.

76 The purpose of this study was to investigate *in situ* the effects of a high-pressure treatment
77 on both the microstructure and secondary structure of salmon muscle proteins.

78

79 **2. Materials & Methods**

80

81 **2.1. Biological material**

82 Twelve whole salmon (*Salmo salar*, farmed in Scotland) were purchased from a local
83 supplier (Jean LEBEAUPIN, Nantes, France) one day after the harvesting date. The fish
84 weighed 2.5 to 3 kg and were of “Label Rouge” quality, which is an official French label
85 guaranteeing strict farming conditions and final product quality such as a lipid rate between
86 10 and 16%.

87 The collagen content in the muscle of the salmon, which was evaluated by an external
88 laboratory (Inovalys, Nantes, France) and calculated as [(L)-hydroxyproline content x 8],
89 was equal to 0.3 %. From each fish, a slice approximately 5 cm wide was excised and cut
90 into four 40 g parts (see the supplementary material), which were each vacuum packed
91 (98% vacuum) in individual polyamide/polyethylene bags (La Bovida, Paris, France) and
92 kept on ice before processing. The remaining parts of the fish were stored at -20°C until
93 further biochemical analysis.

94

95 **2.2. Reagents and chemicals**

96 For the HES staining, the following chemicals and reagents were used: Harris
97 Haematoxylin (Harris Hematoxylin Surgipath, Leica biosystem, Buffalo Grove, IL, USA),
98 Eosin 0.5% (eosin Y 0.5% w/v aqueous solution, Sigma Aldrich, St Louis, MO, USA),
99 Powdered saffron (Technical, VWR International, Radnor, PA, USA), Methylcyclohexane
100 (for synthesis, Merck, Darmstadt, Germany), Eukitt (Eukitt® Quick-hardening mounting
101 medium, Sigma Aldrich, St Louis, MO, USA) and Ethanol (99%, Carlo Erba Reagents, Val
102 de Reuil, France).

103 For the Picrosirius red staining, the following chemicals and reagents were used: Acetone

104 (99%, Sigma Aldrich, St Louis, MO, USA), Picro-Sirius red (Sirius red F3B, Gurr BDH
105 laboratory Supplies, Poole, England), Hydrochloric acid 0.01M (37%, VWR International,
106 Radnor, PA, USA), Ethanol (99%, Carlo Erba Reagents, Val de Reuil, France) and
107 Methycyclohexane (for synthesis, Merck, Darmstadt, Germany). A Picro-formalin solution
108 was prepared with ethanol 95% and formaldehyde (37 %, Sigma Aldrich, St Louis, MO,
109 USA) (180/25 v/v) and picric acid to saturation (Powdered, Sigma Aldrich, St Louis, MO,
110 USA).

111

112 **2.3 High-pressure processing**

113 The packed samples (approximately 40 g each) were inserted in the stainless steel
114 chamber of a custom-made high-pressure device (NOVA SWISS, Cesson, France). The
115 chamber, equipped with a water jacket and a temperature regulator device, was filled with
116 water at 20°C and pressure was generated by addition of water in the chamber using a
117 hydrostatic pump. For each fish, the four pieces were processed with a balanced
118 experimental design for 5 min at 200 MPa, 400 MPa, 600 MPa or unpressurized (control),
119 meaning that for each level of pressure, different locations of the salmon were used. These
120 pressure rates were chosen in order to investigate the process effect up to 600 MPa which
121 is generally the maximum HP used in industry and was the maximum limit of the
122 equipment. The compression rate was 3 MPa/s, and the depression rates were 4 MPa/s, 8
123 MPa/s and 10 MPa/s respectively. During the pressurization process, the maximal
124 temperatures reached were 24.7°C, 29.4°C and 33.6°C for the samples pressurized at
125 200, 400 and 600 MPa, respectively. The samples were kept on ice as soon as the
126 treatment ended and until histological sampling.

127

128 **2.4. Microstructure**

129 **2.4.1. Histochemical analysis**

130 A 10 mm wide cube was cut from each sample to identify the muscle fibre direction. The
131 cubes were plunged and stirred for 30 seconds in 2-methyl-butane (-160°C) cooled with
132 liquid nitrogen (-196°C). The cryofixed samples were then kept at -80°C until analysis.
133 A cryomicrotome (Leica CM 1950, Leica Biosystems, Nussloch, Germany) was used to cut
134 10 µm slices from each cube, transversally to the muscle fibre direction. The sections were
135 mounted on SuperFrost microscope slides, air-dried (20°C) and stained with haematoxylin-
136 eosin-saffron (HES) to contrast the tissue and visualize the general structure and with
137 picosirius red to reveal connective tissue. The stained sections were mounted with
138 synthetic resin (Eukitt, Kindler GmbH & Co, Freiburg, Germany) and protected by covering
139 with a glass coverslip. Images were acquired using an Olympus BX 61 microscope
140 coupled with a high-resolution digital camera (Olympus DP 71) and Olympus Cell Sens
141 software (Olympus France SAS, Rungis, France).
142 For each sample, at least ten images of x100-magnified HES staining and picosirius
143 staining were captured.

144

145 **2.4.2. Image analysis**

146 For each of the four conditions, ten images of HES staining and picosirius staining were
147 analysed using open-source ImageJ software (<https://imagej.nih.gov/ij/>). Approximately 50
148 muscle fibres were counted per optical field, which means that approximately 500 fibres
149 per sample were analysed. For each picture, the software was used to calculate both the
150 average size of the fibres and the percentage of surface occupied by the fibres, the
151 connective tissue and the extracellular spaces (ECSs), per optical field.

152

153 **2.5. Infrared spectroscopy data acquisition**

154 Histological cross-sections (6 µm thick) were obtained from the cryofixed muscle sample,
155 collected on a BaF₂ window with IR spectroscopy compatibility (CRYSTRAN, Poole,

156 United Kingdom) and air-dried at room temperature. IR spectra were collected using an
157 FT-IR microscope (Thermo Scientific, Nicolet iN10) scanning from 4000 to 675 cm^{-1} with a
158 spectral resolution of 4 cm^{-1} and an aperture size set at 30 X 30 μm . For each section, 10
159 spectra were acquired in 10 different muscle fibres. Each spectrum resulted from 64
160 accumulated scans. The accumulated spectra were averaged and subtracted from a
161 background spectrum obtained at the start of the scan by accumulating 128 scans.

162

163

164 **2.6. Infrared spectra pre-treatment**

165 The spectra acquired were analysed using TQAnalyst software (V 9.5.0, Thermo Fisher
166 Scientific Inc., Waltham, MA, USA). All spectra were first submitted to multiplicative signal
167 correction (MSC) so that analytical artefacts would not be taken into account. The second
168 derivative of the spectra was calculated, and a Savistky-Golay filter with 9 points and a
169 third-degree polynomial was applied to improve the data resolution.

170

171 **2.7. Statistical analysis**

172 The analysis on the infrared spectra was focused on the 1600-1700 cm^{-1} region mostly
173 assigned to the amide I band of proteins (Barth & Zscherp, 2002). Principal component
174 analysis (PCA) was performed on all spectra to explore similarities between the samples
175 without prior assumptions, using the absorbance at each measured wavenumber as a
176 variable. Principal component representation was used to determine which variables had
177 the most weight to describe the variability between the samples and the pressure
178 conditions. Measurement of the peak heights at 1655 cm^{-1} and 1628 cm^{-1} was performed,
179 and the resulting data were shifted to improve the graphical representation.

180

181 For histological data, statistical analysis was carried out using Statgraphics Centurion 18

182 software (Statgraphics Technologies, Inc., The Plains, VA, USA). Multivariate analysis of
183 variance (ANOVA) was performed on the data to determine significant differences between
184 the levels of pressure applied while taking intraspecies variability into account. Multiple
185 range tests were used to identify which means were significantly different according to
186 Fisher's least significant difference (LSD) test ($p < 0.05$).

187

188 The data in all the figures are expressed as the means \pm standard error.

189

190 **3. Results**

191

192 **3.1. Effect of HP on the microstructure of salmon muscle**

193 Figure 1 shows representative histological images of the HES- and Picrosirius red stained
194 cross-sections.

195 HES staining (Figure 1A) allowed us to assess the effects of HP on salmon microstructure.

196 The HES-stained control fibres appeared tight, with narrow spaces between cells. At 200

197 MPa, the ECS increased compared to that of the control. Similar results were found on

198 cold-smoked Atlantic salmon treated with HP above 400 MPa for 30 and 60 s

199 (Gudbjornsdottir et al., 2010) and on raw salmon muscle processed at 300 MPa

200 (Gudmundsson & Hafsteinsson, 2001). Conversely, Chéret et al. (2005) observed the

201 opposite effect on sea bass (*Dicentrarchus labrax L.*) fillet, with a decrease in the ECS with

202 increasing pressure from 200 to 500 MPa, which they attributed to gelation of the

203 myofibrillar proteins.

204 After pressurization at 400 and 600 MPa, the appearance of the fibres changes and

205 becomes smoother with brighter red staining, which could be a consequence of an

206 increase in protein density and/or protein coagulation.

207 Image analysis with ImageJ software was used to precisely quantify the changes in

208 microstructure. Collagen, which appears in purple in Figure 1A, was calculated as part of
209 the ECS.

210 The high-pressure treatment resulted in shrinkage of the fibres (Figure 2A) and an
211 increase in the ECS (Figure 2B), suggesting that this increase was at least partly a
212 consequence of lateral shrinkage of the muscle fibres. Although significant differences in
213 fibre size were demonstrated between the 200 MPa and 400 MPa samples, overall, the
214 treated samples showed little cross-sectional area (CSA) variation among them. However,
215 the treatment at 600 MPa resulted in a large increase in ECS, even though the average
216 fibre size was not modified compared to that at 400 MPa. Similarly, Gudbjornsdottir et al.
217 (2010) observed a significant increase in the ECS for samples pressurized to 600 MPa or
218 more. However, the increase in ECS was related to the shrinkage of muscle fibres,
219 contrary to our results. Our results could be explained by a disruption of the connective
220 tissue, which no longer holds the fibres together, as suggested by Figure 1B.

221

222 **3.1.2 Picrosirius red staining**

223 The images of the picrosirius red-stained sections are shown in Figure 1B: the fibres are
224 coloured yellow, and the connective tissue is coloured red. For the HES-stained sections,
225 the control fibres appeared close to each other, with a thin layer of connective tissue
226 between the cells. The fibre colour also changes with increasing pressure, becoming more
227 orange. From 400 MPa, connective tissue appears thicker and brighter red.

228 Compared to HES staining, which was used to precisely evaluate the fibre characteristics,
229 picrosirius red staining allowed estimating the amount of space occupied distinctively by
230 the fibres, connective tissue and ECSs, as presented in Figure 3A, 3B and 3C,
231 respectively.

232

233 For the unpressurized samples, the fibres represent 57.7% (\pm 0.6%) of the total area, 3.1%

234 for the connective tissue and 3.0% for the ECS. Only the fibres that were fully inside the
235 optical area were taken into account for the calculation, which explains why the sum was
236 not equal to 100%.

237 The pressurization of the salmons at 200 MPa resulted in a marked increase in the area
238 occupied by the ECS and a slight decrease in the fibre area, whereas no significant effect
239 was observed for the connective tissue. At 400 MPa, no additional effect was observed on
240 the fibres and ECS, but the percentage of area occupied by the connective tissue was
241 doubled compared to that at 200 MPa, which we assumed could be explained by a
242 loosening of the collagen structure.

243 At 600 MPa, significant decreases in the fibres and the connective tissue area were
244 measured, correlated with a large increase in the ECS.

245

246 **3.2. Effects of high-pressure treatment on the secondary structure of salmon** 247 **proteins**

248 Mid-infrared spectroscopy is a powerful tool to characterize protein secondary structures
249 based on the principle that the vibrational frequencies of a molecule are affected by the
250 strength of its bonds and the mass of its vibrating atoms (Barth & Zscherp, 2002). In the
251 present study, vibrational spectra were acquired from approximately 120 myofibres for
252 each pressure condition.

253 PCA was performed on all the whole spectra (675-4000 cm^{-1}) and allowed identifying the
254 1700-1600 cm^{-1} band as the most discriminant for HP treatments.

255 This band is assigned to the amide I band of proteins and is more specifically related to
256 the vibration of the stretching of C=O bonds. It is strongly related to the secondary
257 structure of the proteins (Barth & Zscherp, 2002; Jackson & Mantsch, 1995).

258 Consequently, the data analysis focused on the 1700-1600 cm^{-1} part of the spectra
259 (Figures 4 - 5).

260

261 With increasing pressure, the second-derivative spectra (Figure 4) clearly show a
262 decrease in absorbance at 1655 cm^{-1} assigned to α -helices (Barth, 2007; Jackson &
263 Mantsch, 1995) and an increase in absorbance in the $1640\text{-}1620\text{ cm}^{-1}$ range. The band
264 occurring at 1628 cm^{-1} is assigned to denatured aggregated β -sheet components
265 (intermolecular), while the band between 1630 and 1640 cm^{-1} is assigned to antiparallel β -
266 sheet structures (intramolecular) (Jackson and Mantsch, 1995; Bocker et al., 2006, Wu et
267 al. 2007).

268 PCA (Figure 5) confirms that the dissimilarities between the pressure groups were strongly
269 correlated to the first principal component, which was mainly explained by bands at 1655
270 cm^{-1} and 1628 cm^{-1} reflecting myofibre protein denaturation (Astruc et al. 2012).

271

272 Quantification of the peak heights at 1655 cm^{-1} and 1628 cm^{-1} was performed to study the
273 evolution of the secondary structure with pressure (figure 6). Previous studies conducted
274 on fish treated with HP (Larrea-Wachtendorff et al., 2015; Martínez et al., 2017) also
275 focused on the amide I band of the FT-IR spectra and found a similar evolution of the
276 secondary structure, i.e., a decrease in the α -helix content and an increase in the content
277 of aggregated β -sheet structures. Villamonte et al. (2015) focused on the effects of HP on
278 sarcoplasmic proteins extracted from hake and observed the same evolution of secondary
279 structure correlated to a decrease of their emulsifying properties. The changes of the
280 sarcoplasmic proteins structure also caused their aggregation, which can result in an
281 increase of the hardness of the fish meat. The decrease of α -helix and increase of
282 aggregated β -sheet structures was also measured by Cando et al. (2014) on myofibrillar
283 proteins isolated from hake and treated by HP. The pressure treatment induced the
284 formation of gels which were less rigid and softer than gels formed by thermal denaturation
285 of the proteins. The authors suggested that the increase of aggregated β -sheet structures

286 could be the consequence of a gelling process.
287 Pressure denaturation of muscle proteins resulted in similar modifications of their
288 secondary structure than after thermal treatment (Astruc et al., 2012; Ovissipour et al.,
289 2017). However, in the present study, the denaturation cannot be caused by temperature
290 as it did not exceed 33.6°C. Protein denaturation occurs during HP due to destabilisation
291 of non-covalent interactions in the tertiary and secondary structures, particularly
292 hydrophobic and ionic interactions because their disruption is associated with a negative
293 change of volume and thus a compaction of the molecule (Chapleau et al., 2004;
294 Heremans, 1982; Mozhaev et al., 1994). High pressure acts by altering the balance of
295 intermolecular and solvent–protein interactions. Contrary to thermal denaturation, HP
296 treatments do not lead to the formation of intermolecular hydrogen bonds (Chapleau et al.
297 2004).

298

299 **4. Discussion**

300

301 The results of the histological analysis show that HP causes a decrease in the fibre size
302 regardless of the level of pressure applied, which is likely to be responsible for the ECS
303 increase. The shrinkage of the fibres could be related to water flows between the muscle
304 cells and the ECS. Several authors have shown a decrease in the water holding capacity
305 of fish submitted to HP, which supports this hypothesis (Chéret et al., 2005; Jiranuntakul et
306 al., 2018; Lakshmanan et al., 2007; Ramirez-Suarez & Morrissey, 2006). Chéret et al.
307 (2005) additionally studied the exudation of sea bass fillets after pressurization but did not
308 observe any modification caused by HP.

309 Furthermore, we have shown that pressure above 400 MPa resulted in a change in
310 connective tissue appearance and size. This alteration could be due to a change in the
311 collagen structure. Collagen is formed of a triple helix mainly stabilized by hydrogen and

312 covalent bonds connecting hydroxyproline from one strand to amide carboxyl from another
313 strand (Benjakul et al., 2012) and should therefore be relatively resistant to HP (Truong et
314 al., 2015). However, Kaur et al. (2016) studied the effects of HP on the structure of bovine
315 meat and suggested that pressure, by disrupting the non-covalent bonds, could disturb the
316 molecule stability. Fish collagen contains smaller amounts of proline and hydroxyproline
317 and is therefore less stable than mammalian collagen which supports our hypothesis
318 (Benjakul et al., 2012).

319 Kaur et al. (2016) also observed a decrease in connective tissue on raw pork treated by
320 HP (600 MPa, 10 min), which they attributed to a possible rise in temperature during high-
321 pressure processing associated with a decrease in denaturation temperature of the
322 collagen caused by the destabilization of non-covalent bonds. During the processing of our
323 samples, we measured a rise in temperature from 20°C to 29.4°C and 33.6°C for the 400
324 MPa and 600 MPa treatments, respectively. The denaturation of fish collagen was reported
325 to occur at between 30°C and 40°C (Benjakul et al., 2012; Hastings et al., 1985), and the
326 slight increase in temperature might have participated in the loosening of the connective
327 tissue at 400 MPa and the partial denaturation at 600 MPa caused by the high-pressure
328 treatment.

329 It is worth noting that after pressurization at 600 MPa, the ECS size is substantially
330 increased even though the fibre cross-sectional area is not significantly modified. It is
331 unlikely that the overall volume of the samples was expanded by the treatment because
332 pressure rather favours molecular states which are related with a decreased volume
333 (Mozhaev et al., 1994). However, this phenomenon was correlated with the decrease of
334 the connective tissue area at 600 MPa compared to 400 MPa, and it is therefore likely that
335 the pressure induced denaturation of collagen caused a partial disruption of the connective
336 tissue which could no longer hold the fibres together, explaining the increase of the ECS.
337 This could have possibly happened after depressurization, during the preparation of the

338 samples.

339

340 All levels of HP resulted in a modification of the salmon muscle secondary structure. After
341 pressurization at 200 MPa, myofibres contained a smaller amount of α -helices and more
342 aggregated β -sheet structures, associated with the compaction of the fibres and larger
343 ECS than that of the control. This modification of the secondary structure was enhanced at
344 higher levels of pressure. After pressurization at 400 MPa, connective tissue integrity was
345 altered and therefore occupied an increased surface area compared to that of the control
346 and 200 MPa samples. When pressurization at 600 MPa was applied, connective tissue
347 was found to be partially degraded, leading to an expansion of the ECS.

348 Myofibrillar proteins are the most abundant proteins in muscle cells, with 50-60% myosin
349 (Venugopal & Shahidi, 1996) and are mainly organized in α -helices (Wang et al., 2021; Xu
350 et al., 2020). Several authors (Angsupanich & Ledward, 1998; Arnaud et al., 2018; Larrea-
351 Wachtendorff et al., 2015) have studied the thermal denaturation of proteins from fishes
352 previously treated by HP, and all found that myofibrillar proteins were denatured by
353 pressures above 200 MPa, with denaturation enthalpies decreasing all the more that
354 pressure was increased. The protein secondary structure is stabilized by hydrogen bonds,
355 which are known to be resistant or even strengthened by HP (Mozhaev et al., 1994;
356 Oliveira et al., 2017) because of the negative volume changes that are associated with the
357 formation of these bonds. Boonyaratanakornkit et al. (2002) reported that these bonds
358 were, however, modified by HP, with a reduced length and a possible promotion of
359 intermolecular hydrogen bonds to the detriment of intramolecular hydrogen bonds. HP, by
360 unfolding myofibrillar proteins and changing the properties of the hydrogen bonds, could
361 therefore result in the destabilization of part of the α -helices and an increase in the content
362 of β -sheets. The consequent compression of myofibrillar proteins might explain the
363 observed decrease in myofibre size.

364 These alterations in muscle structure are linked to pressure-induced modifications of fish
365 sensory properties. Authors have reported significant changes in salmon colour after
366 pressurization, especially increased lightness (Gudbjornsdottir et al., 2010; Yagiz et al.,
367 2009). Although the mechanisms responsible for this discoloration are still unclear,
368 evidence shows that it could be due to the denaturation or coagulation of myofibrillar
369 proteins. The unfolding of these proteins could indeed modify their surface properties and
370 affect the way they absorb and reflect light (Kruk et al., 2011; Suemitsu & Cristianini,
371 2019). The decrease in α -helical structures that we observed as well as the histological
372 analysis, which show that the myofibre cross-sections become smoother with pressure,
373 support the hypothesis that the change in colour caused by HP is linked to the
374 denaturation of muscle proteins rather than the alteration of carotenoid pigments. These
375 molecules are responsible for the distinctive orange colour of salmon and are likely
376 unaltered by mechanical treatments such as HP (Martínez-Delgado et al., 2017).

377 In most cases, high-pressure treatments increase the hardness of fish muscles (Oliveira
378 et al., 2017), and this change in texture has been associated with a higher compaction of
379 the fibres after treatment and a preservation of collagen by pressure (Truong et al., 2015).
380 In contrast, we observed an increase in the intercellular space and an alteration in
381 connective tissue integrity in this study. Arnaud et al. (2018) suggested that the increased
382 hardness observed on salmon after pressurization at 300 MPa could be a result of protein
383 aggregation, which is consistent with the increase in the β -sheet structure that we
384 measured.

385 In the present study, FT-IR spectral analysis could not be performed on the connective
386 tissue because the spatial resolution of a laboratory FT-IR microspectrometer is too low
387 (best acquisition area of 10 μm x 10 μm) to precisely target the endomysium. Further
388 analyses using a synchrotron infrared beamline (Astruc et al., 2012) or Raman
389 microspectroscopy technique will allow us to achieve a sufficient spatial resolution (3x3 μm

390 for synchrotron-FTIR microspectroscopy; 1 x 1 μm for Raman microspectroscopy) to
391 investigate the intramuscular connective tissue in order to better understand the
392 modifications of the connective tissue affected by high-pressure treatment.

393

394 **5. Conclusion**

395

396 Most interpretations of the pressure-induced modifications of fish muscles are based on
397 physical rules such as Le Chatelier's principle (Mozhaev et al., 1994) or on experimental
398 data obtained from extracted proteins. However, these interpretations fail to take into
399 consideration the high level of organization and complexity of the muscular structure. The
400 combination of histological and FT-IR microspectroscopy analysis allowed us to
401 investigate the microstructure and myofibre protein macromolecular structure *in situ* and
402 concomitantly. Supplementary analysis of quality parameters of Atlantic salmon treated by
403 HP, such as colour, texture and water holding capacity, would provide helpful data to
404 explain the relations between the microstructure and the technological properties of
405 salmon meat.

406

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411

412 **Conflicts of interest**

413 The authors declare that they have no known competing financial interests or personal
414 relationships that could have appeared to influence the work reported in this paper.

415

416 **References**

417 Angsupanich, K., & Ledward, D. A. (1998). High pressure treatment effects on cod (*Gadus*
418 *morhua*) muscle. *Food Chemistry*, *63*(1), 39–50. <https://doi.org/10.1016/S0308->

419 8146(97)00234-3

420 Arnaud, C., Lamballerie, M. de, & Pottier, L. (2018). Effect of high pressure processing on
421 the preservation of frozen and re-thawed sliced cod (*Gadus morhua*) and salmon (*Salmo*
422 *salar*) fillets. *High Pressure Research*, *38*(1), 62–79.

423 <https://doi.org/10.1080/08957959.2017.1399372>

424 Astruc, T., Peyrin, F., Vénien, A., Labas, R., Abrantes, M., Dumas, P., & Jamme, F. (2012).
425 In situ thermal denaturation of myofibre sub-type proteins studied by

426 immunohistofluorescence and synchrotron radiation FT-IR microspectroscopy. *Food*

427 *Chemistry*, *134*(2), 1044–1051. <https://doi.org/10.1016/j.foodchem.2012.03.012>

428 Aubourg, S. P. (2018). Impact of high-pressure processing on chemical constituents and
429 nutritional properties in aquatic foods: A review. *International Journal of Food Science &*

430 *Technology*, *53*(4), 873–891. <https://doi.org/10.1111/ijfs.13693>

431 Aubourg, S. P., Rodríguez, A., Sierra, Y., Tabilo-Munizaga, G., & Pérez-Won, M. (2013).

432 Sensory and Physical Changes in Chilled Farmed Coho Salmon (*Oncorhynchus kisutch*):

433 Effect of Previous Optimized Hydrostatic High-Pressure Conditions. *Food and Bioprocess*

434 *Technology*, *6*(6), 1539–1549. <https://doi.org/10.1007/s11947-012-0799-4>

435 Barth, A., & Zscherp, C. (2002). What vibrations tell about proteins. *Quarterly Reviews of*

436 *Biophysics*, *35*(4), 369–430. <https://doi.org/10.1017/S0033583502003815>

437 Benjakul, S., Nalinanon, S., & Shahidi, F. (2012). Fish Collagen. In *Food Biochemistry and*

438 *Food Processing: Second Edition* (pp. 365–387).
439 <https://doi.org/10.1002/9781118308035.ch20>

440 Boonyaratankornkit, B. B., Park, C. B., & Clark, D. S. (2002). Pressure effects on intra-
441 and intermolecular interactions within proteins. *Biochimica et Biophysica Acta (BBA) -*
442 *Protein Structure and Molecular Enzymology*, 1595(1), 235–249.
443 [https://doi.org/10.1016/S0167-4838\(01\)00347-8](https://doi.org/10.1016/S0167-4838(01)00347-8)

444 Cando, D., Moreno, H. M., Tovar, C. A., Herranz, B., & Borderias, A. J. (2014). Effect of
445 High Pressure and/or Temperature over Gelation of Isolated Hake Myofibrils. *Food and*
446 *Bioprocess Technology*, 7(11), 3197–3207. <https://doi.org/10.1007/s11947-014-1279-9>

447 Chapleau, N., Mangavel, C., Compoin, J.-P., & Lamballerie-Anton, M. de. (2004). Effect of
448 high-pressure processing on myofibrillar protein structure. *Journal of the Science of Food*
449 *and Agriculture*, 84(1), 66–74. <https://doi.org/10.1002/jsfa.1613>

450 Chéret, R., Chapleau, N., Delbarre-Ladrat, C., Verrez-Bagnis, V., & Lamballerie, M. de.
451 (2005). Effects of High Pressure on Texture and Microstructure of Sea Bass (*Dicentrarchus*
452 *labrax* L.) Fillets. *Journal of Food Science*, 70(8), e477–e483.
453 <https://doi.org/10.1111/j.1365-2621.2005.tb11518.x>

454 de Alba, M., Pérez-Andrés, J. M., Harrison, S. M., Brunton, N. P., Burgess, C. M., & Tiwari,
455 B. K. (2019). High pressure processing on microbial inactivation, quality parameters and
456 nutritional quality indices of mackerel fillets. *Innovative Food Science & Emerging*
457 *Technologies*, 55, 80–87. <https://doi.org/10.1016/j.ifset.2019.05.010>

458 Directorate-General for Maritime Affairs and Fisheries (European Commission). (2020).
459 The EU fish market: 2020 edition. Publications Office of the European Union.
460 <https://data.europa.eu/doi/10.2771/664425>

461 Fogarty, C., Whyte, P., Brunton, N., Lyng, J., Smyth, C., Fagan, J., & Bolton, D. (2019).
462 Spoilage indicator bacteria in farmed Atlantic salmon (*Salmo salar*) stored on ice for 10
463 days. *Food Microbiology*, 77, 38–42. <https://doi.org/10.1016/j.fm.2018.08.001>

464 Gudbjornsdottir, B., Jonsson, A., Hafsteinsson, H., & Heinz, V. (2010). Effect of high-
465 pressure processing on *Listeria* spp. And on the textural and microstructural properties of
466 cold smoked salmon. *LWT - Food Science and Technology*, 43(2), 366–374.
467 <https://doi.org/10.1016/j.lwt.2009.08.015>

468 Gudmundsson, M., & Hafsteinsson, H. (2001). Effect of electric field pulses on
469 microstructure of muscle foods and roes. *Trends in Food Science & Technology*, 12(3),
470 122–128. [https://doi.org/10.1016/S0924-2244\(01\)00068-1](https://doi.org/10.1016/S0924-2244(01)00068-1)

471 Hastings, R. J., Rodger, G. W., Park, R., Matthews, A. D., & Anderson, E. M. (1985).
472 Differential Scanning Calorimetry of Fish Muscle: The Effect of Processing and Species
473 Variation. *Journal of Food Science*, 50(2), 503–506. [https://doi.org/10.1111/j.1365-](https://doi.org/10.1111/j.1365-2621.1985.tb13437.x)
474 [2621.1985.tb13437.x](https://doi.org/10.1111/j.1365-2621.1985.tb13437.x)

475 Heremans, K. (1982). High pressure effects on proteins and other biomolecules. Annual
476 Review of Biophysics and Bioengineering, 11, 1–21.
477 <https://doi.org/10.1146/annurev.bb.11.060182.000245>

478 Jackson, M., & Mantsch, H. H. (1995). The Use and Misuse of FTIR Spectroscopy in the
479 Determination of Protein Structure. *Critical Reviews in Biochemistry and Molecular*
480 *Biology*, 30(2), 95–120. <https://doi.org/10.3109/10409239509085140>

481 Jiranuntakul, W., Nakwiang, N., Berends, P., Kasemsuwan, T., Saetung, T., & Devahastin,
482 S. (2018). Physicochemical, Microstructural, and Microbiological Properties of Skipjack
483 Tuna (*Katsuwonus pelamis*) After High-Pressure Processing. *Journal of Food Science*,

484 83(9), 2324–2336. <https://doi.org/10.1111/1750-3841.14318>

485 Kaur, L., Astruc, T., Vénien, A., Loison, O., Cui, J., Irastorza, M., & Boland, M. (2016). High
486 pressure processing of meat: Effects on ultrastructure and protein digestibility. *Food &*
487 *Function*, 7(5), 2389–2397. <https://doi.org/10.1039/c5fo01496d>

488 Knorr, D., Heinz, V., & Buckow, R. (2006). High pressure application for food biopolymers.
489 *Biochimica et Biophysica Acta (BBA) - Proteins and Proteomics*, 1764(3), 619–631.
490 <https://doi.org/10.1016/j.bbapap.2006.01.017>

491 Kruk, Z. A., Yun, H., Rutley, D. L., Lee, E. J., Kim, Y. J., & Jo, C. (2011). The effect of high
492 pressure on microbial population, meat quality and sensory characteristics of chicken
493 breast fillet. *Food Control*, 22(1), 6–12. <https://doi.org/10.1016/j.foodcont.2010.06.003>

494 Lakshmanan, R., Parkinson, J. A., & Piggott, J. R. (2007). High-pressure processing and
495 water-holding capacity of fresh and cold-smoked salmon (*Salmo salar*). *LWT - Food*
496 *Science and Technology*, 40(3), 544–551. <https://doi.org/10.1016/j.lwt.2005.12.003>

497 Larrea-Wachtendorff, D., Tabilo-Munizaga, G., Moreno-Osorio, L., Villalobos-Carvajal, R.,
498 & Pérez-Won, M. (2015). Protein Changes Caused by High Hydrostatic Pressure (HHP): A
499 Study Using Differential Scanning Calorimetry (DSC) and Fourier Transform Infrared
500 (FTIR) Spectroscopy. *Food Engineering Reviews*, 7(2), 222–230.
501 <https://doi.org/10.1007/s12393-015-9107-1>

502 Martínez, M. A., Velazquez, G., Cando, D., Núñez-Flores, R., Borderías, A. J., & Moreno,
503 H. M. (2017). Effects of high pressure processing on protein fractions of blue crab
504 (*Callinectes sapidus*) meat. *Innovative Food Science & Emerging Technologies*, 41, 323–
505 329. <https://doi.org/10.1016/j.ifset.2017.04.010>

506 Martínez-Delgado, A. A., Khandual, S., & Villanueva-Rodríguez, S. J. (2017). Chemical

507 stability of astaxanthin integrated into a food matrix: Effects of food processing and
508 methods for preservation. *Food Chemistry*, 225, 23–30.
509 <https://doi.org/10.1016/j.foodchem.2016.11.092>

510 Mozhaev, V. V., Heremans, K., Frank, J., Masson, P., & Balny, C. (1994). Exploiting the
511 effects of high hydrostatic pressure in biotechnological applications. *Trends in*
512 *Biotechnology*, 12(12), 493–501. [https://doi.org/10.1016/0167-7799\(94\)90057-4](https://doi.org/10.1016/0167-7799(94)90057-4)

513 Oliveira, F. A. de, Neto, O. C., Santos, L. M. R. dos, Ferreira, E. H. R., & Rosenthal, A.
514 (2017). Effect of high pressure on fish meat quality – A review. *Trends in Food Science &*
515 *Technology*, 66, 1–19. <https://doi.org/10.1016/j.tifs.2017.04.014>

516 Ovissipour, M., Rasco, B., Tang, J., & Sablani, S. (2017). Kinetics of Protein Degradation
517 and Physical Changes in Thermally Processed Atlantic Salmon (*Salmo salar*). *Food and*
518 *Bioprocess Technology*, 10(10), 1865–1882. <https://doi.org/10.1007/s11947-017-1958-4>

519 Park, D.-H., Jung, J.-G., Jung, B.-R., Kim, G., Lee, H., Kim, H.-A., & Bang, M.-A. (2015).
520 Changes in Salmon (*Oncorhynchus keta*) Flesh Quality Following Ultra-High Pressure
521 Treatment and 30 d of Chilled Storage. *Journal of Food Science*, 80(1), M142–M146.
522 <https://doi.org/10.1111/1750-3841.12714>

523 Ramirez-Suarez, J. C., & Morrissey, M. T. (2006). High Hydrostatic Pressure and Heat
524 Treatment Effects on Physicochemical Characteristics of Albacore Tuna (*Thunnus*
525 *alalunga*) Minced Muscle. *Journal of Aquatic Food Product Technology*, 15(1), 5–17.
526 https://doi.org/10.1300/J030v15n01_02

527 Suemitsu, L., & Cristianini, M. (2019). Effects of high pressure processing (HPP) on quality
528 attributes of tilapia (*Oreochromis niloticus*) fillets during refrigerated storage. *LWT*, 101,
529 92–99. <https://doi.org/10.1016/j.lwt.2018.11.028>

530 Truong, B. Q., Buckow, R., Nguyen, M. H., & Furst, J. (2017). Gelation of barramundi
531 (*Lates calcarifer*) minced muscle as affected by pressure and thermal treatments at low
532 salt concentration. *Journal of the Science of Food and Agriculture*, 97(11), 3781–3789.
533 <https://doi.org/10.1002/jsfa.8242>

534

535 Truong, B. Q., Buckow, R., Stathopoulos, C. E., & Nguyen, M. H. (2015). Advances in
536 High-Pressure Processing of Fish Muscles. *Food Engineering Reviews*, 7(2), 109–129.
537 <https://doi.org/10.1007/s12393-014-9084-9>

538 Uresti, R. M., Velazquez, G., Ramírez, J. A., Vázquez, M., & Torres, J. A. (2004). Effect of
539 high-pressure treatments on mechanical and functional properties of restructured products
540 from arrowtooth flounder (*Atheresthes stomias*). *Journal of the Science of Food and*
541 *Agriculture*, 84(13), 1741–1749. <https://doi.org/10.1002/jsfa.1876>

542 Venugopal, V., & Shahidi, F. (1996). Structure and composition of fish muscle. *Food*
543 *Reviews International*, 12(2), 175–197. <https://doi.org/10.1080/87559129609541074>

544 Wang, Y.-Y., Tayyab Rashid, M., Yan, J.-K., & Ma, H. (2021). Effect of multi-frequency
545 ultrasound thawing on the structure and rheological properties of myofibrillar proteins from
546 small yellow croaker. *Ultrasonics Sonochemistry*, 70, 105352.
547 <https://doi.org/10.1016/j.ultsonch.2020.105352>

548 Xu, Y., Wang, R., Zhao, H., Yin, Y., Li, X., Yi, S., & Li, J. (2020). Effect of heat treatment
549 duration on the interaction between fish myosin and selected flavor compounds. *Journal of*
550 *the Science of Food and Agriculture*, 100(12), 4457–4463.
551 <https://doi.org/10.1002/jsfa.10486>

552 Yagiz, Y., Kristinsson, H. G., Balaban, M. O., Welt, B. A., Ralat, M., & Marshall, M. R.
553 (2009). Effect of high pressure processing and cooking treatment on the quality of Atlantic

554 salmon. *Food Chemistry*, 116(4), 828–835.

555 <https://doi.org/10.1016/j.foodchem.2009.03.029>

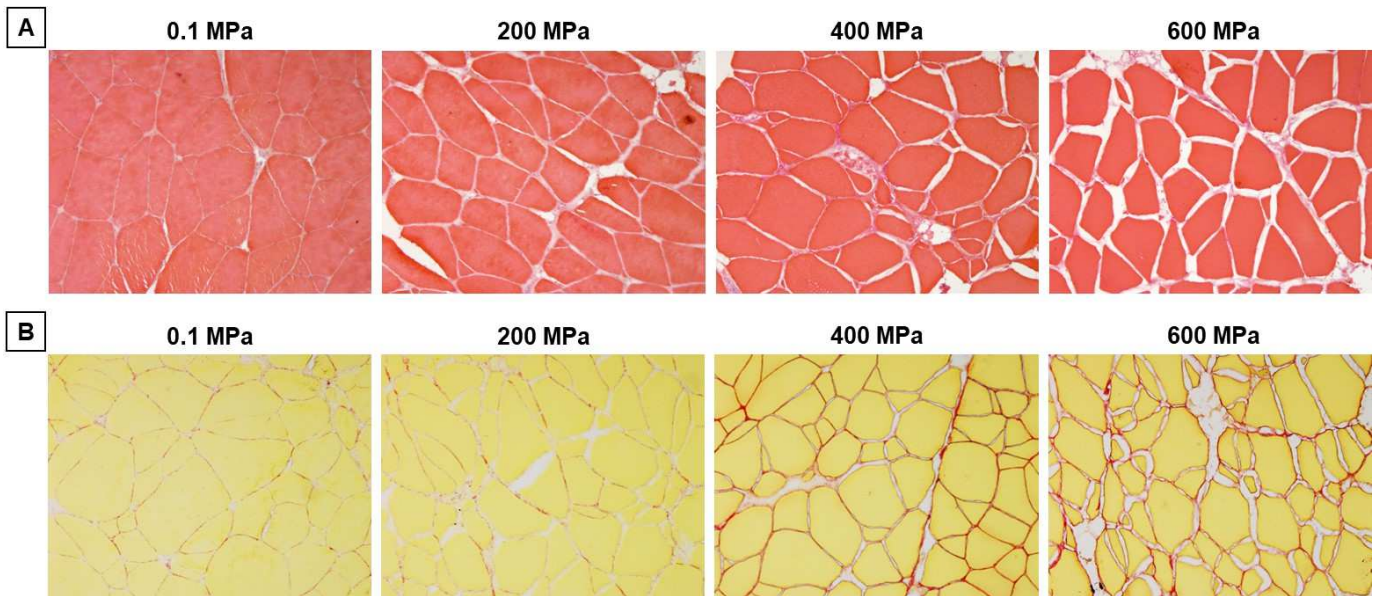


Figure 1: Histological sections of non-pressurized (0.1 MPa) and pressurized (200, 400 and 600 MPa, 5 min) salmon muscle. HES staining (A) colours the fibres red. Picrosirius red staining (B) colours the fibres yellow and the connective tissue red. Scale: 50 mm = 100 μ m

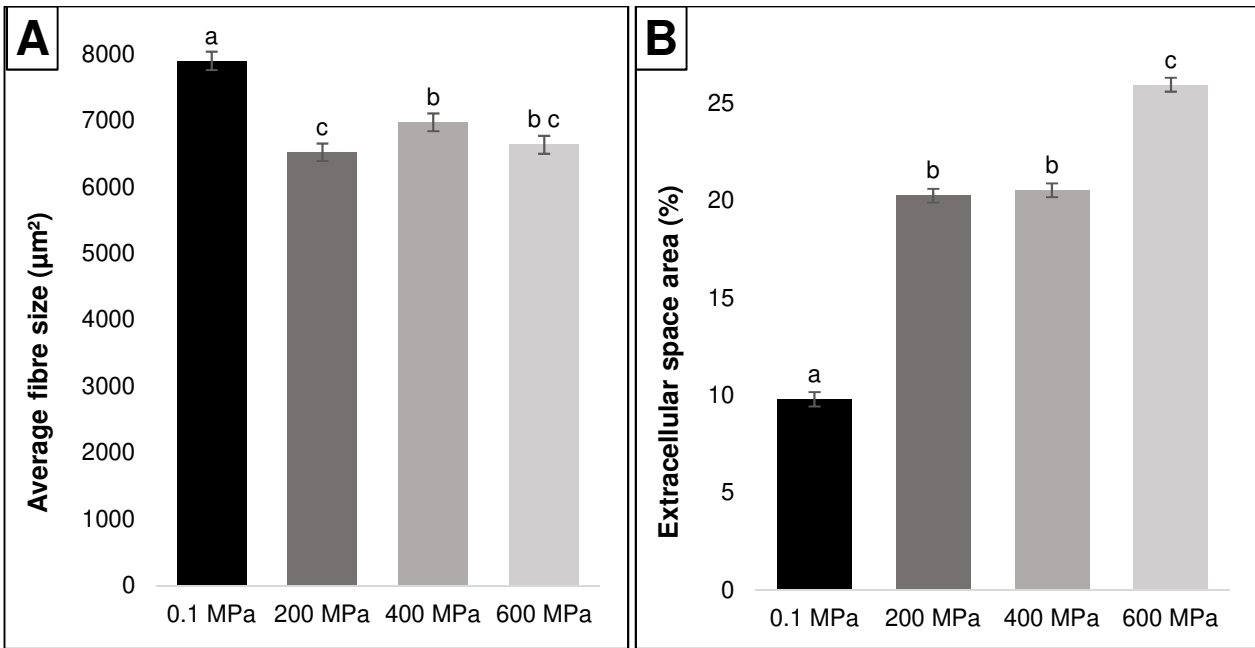


Figure 2: Average fibre size in μm^2 (A) and area from extracellular space in % (B) for non-pressurized (0.1 MPa) and pressurized (200, 400 and 600 MPa, 5 min) salmon (n=12) measured on HES-stained sections. The results are expressed as the mean \pm standard error. Means with different letters are significantly different ($p < 0.05$).

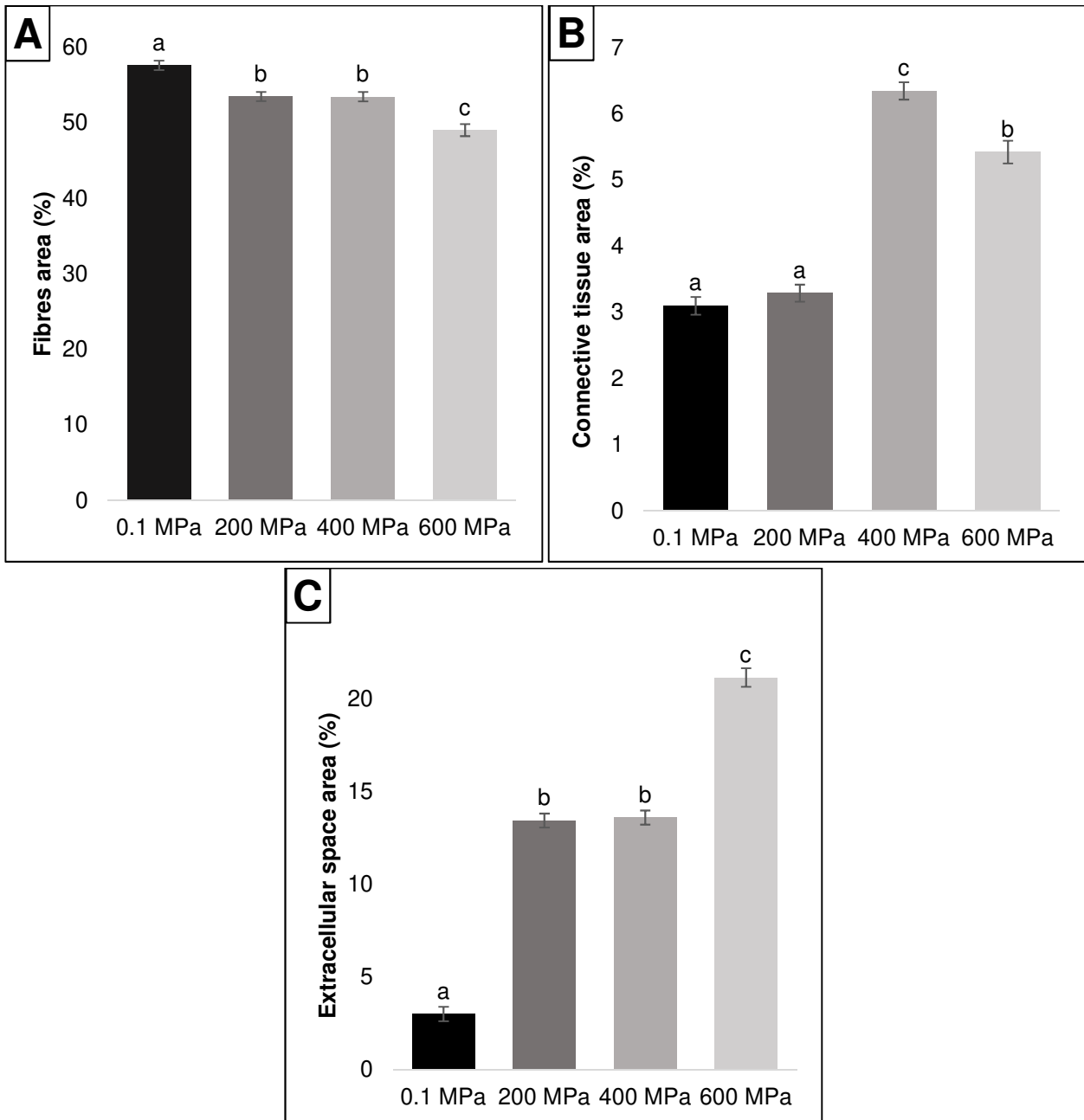


Figure 3: Percentage of the total section area from fibres (A), connective tissue (B) and extracellular space (C) for non-pressurized (0.1 MPa) and pressurized (200, 400 and 600 MPa, 5 min) salmon (n=12) measured on Picrosirius red-stained sections. The results are expressed as the mean \pm standard error. Means with different letters are significantly different ($p < 0.05$).

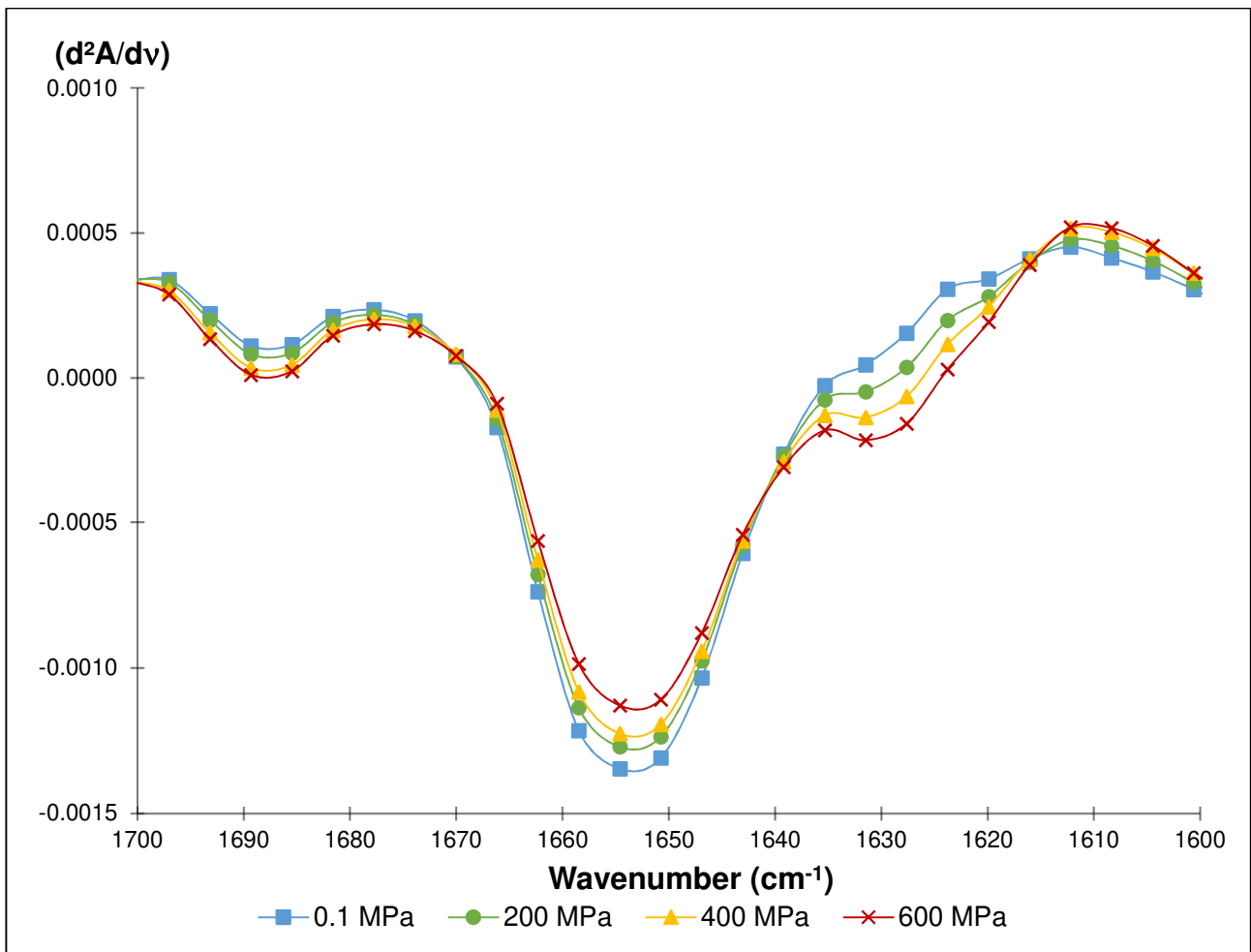


Figure 4: Second-derivative mean infrared spectra (1700-1600 cm^{-1}) obtained in muscle fibres of non-pressurized (0.1 MPa) and pressurized (200, 400 and 600 MPa, 5 min) salmons (n=12), each from approximately 120 spectra.



Figure 5: Principal component analysis (PCA) of the second derivative of the amide I (1600-1700 cm⁻¹) band (A). Pressure groups are separated along the first principal component (PC1).

The principal component graph (B) describes what wavenumbers contribute the most in describing each principal component. PC1 is mainly described by two bands with maximums at 1628 cm^{-1} and 1655 cm^{-1} and describes 56.77% of the variability between all samples.

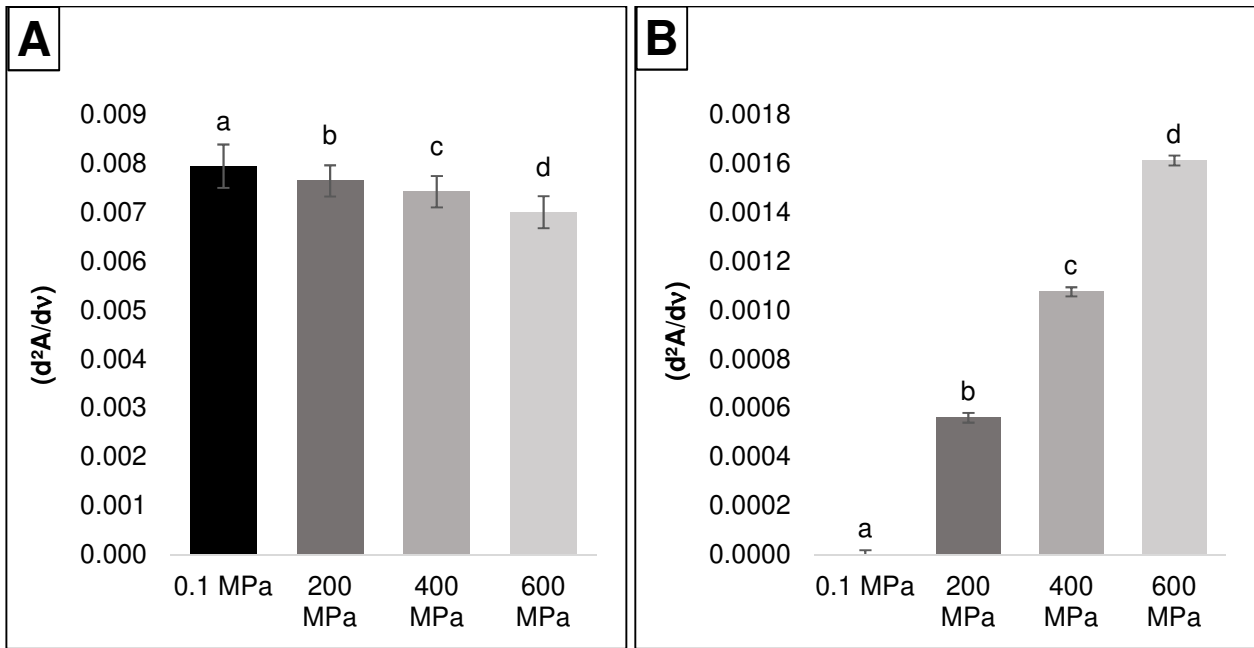
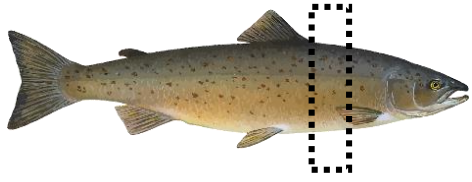


Figure 6: Evolution of α -helices (A) and aggregated β -sheets (B) in salmon (n=12) muscle fibres measured by the height of the second-derivative spectra peaks at 1655 cm^{-1} and 1628 cm^{-1} , respectively. The results are expressed as the mean \pm standard error. Means with different letters are significantly different ($p < 0.05$).

Effects of high-pressure treatment on the muscle structure of salmon (*Salmo salar*)



n = 12 salmons

High-pressure processing

Control, 200 MPa,
400 MPa, 600 MPa
5 min, 20°C

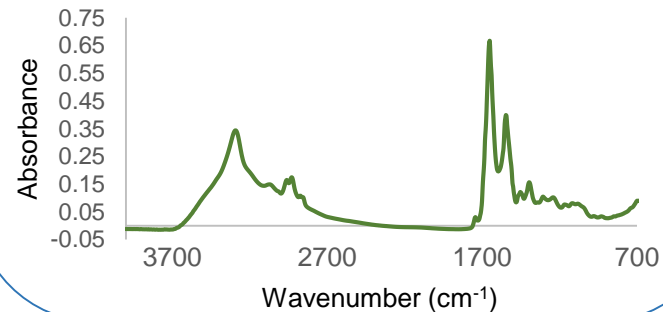


Histochemical analysis

- average fibre size
- amount of space occupied by :
 - the extracellular spaces (ECS)
 - the connective tissue

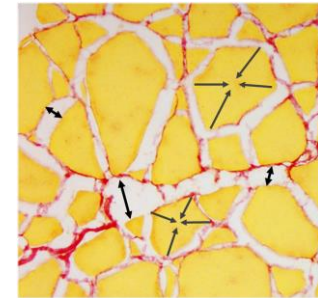
Infrared spectromicroscopy

- Infrared spectra acquired in the muscle fibres
- Evaluation of α -helix and aggregated β -sheet content



Results

- Shrinkage of the fibres from 200 MPa
- Increase of the ECS from 200 MPa
- Modification of the connective tissue at 400 MPa



- Decrease of the α -helix and increase of the aggregated β -sheet with pressure

