

# Waste-to-nutrition: a review of current and emerging conversion pathways

U Javourez, Michael O'Donohue, Lorie Hamelin

# • To cite this version:

U Javourez, Michael O'Donohue, Lorie Hamelin. Waste-to-nutrition: a review of current and emerging conversion pathways. Biotechnology Advances, 2021, 53, 26 p. 10.1016/j.biotechadv.2021.107857 . hal-03426554

# HAL Id: hal-03426554 https://hal.inrae.fr/hal-03426554v1

Submitted on 5 Jan 2024

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés. 1

Waste-to-nutrition: a review of current and emerging conversion pathways

U. Javourez<sup>1</sup>, M. O'Donohue<sup>1</sup>, L. Hamelin<sup>1\*</sup>

2 3

4

<sup>1</sup> TBI, Université de Toulouse, CNRS, INRAE, INSA, Toulouse, France

5 \*Corresponding author (hamelin@insa-toulouse.fr)

6

# 7 Abstract

8 Residual biomass is acknowledged as a key sustainable feedstock for the transition 9 towards circular and low fossil carbon economies to supply whether energy, chemical, 10 material and food products or services. The latter is receiving increasing attention, in 11 particular in the perspective of decoupling nutrition from arable land demand. 12

13 In order to provide a comprehensive overview of the technical possibilities to convert residual biomasses into edible ingredients, we reviewed over 950 scientific and industrial 14 15 records documenting existing and emerging waste-to-nutrition pathways, involving over 150 16 different feedstocks here grouped under 10 umbrella categories: (i) wood-related residual 17 biomass, (ii) primary crop residues, (iii) manure, (iv) food waste, (v) sludge and wastewater, (vi) green residual biomass, (vii) slaughterhouse by-products, (viii) agrifood co-products, (ix) 18 19  $C_1$  gases and (x) others. The review includes a detailed description of these pathways, as 20 well as the processes they involve. As a result, we proposed four generic building blocks to 21 systematize waste-to-nutrition conversion sequence patterns, namely enhancement, extraction and bioconversion. We further introduce a multidimensional 22 cracking. 23 representation of the biomasses suitability as potential as nutritional sources according to (i) 24 their content in anti-nutritional compounds, (ii) their degree of structural complexity and (iii) 25 their concentration of macro- and micronutrients. Finally, we suggest that the different 26 pathways can be grouped into eight large families of approaches: (i) insect biorefinery, (ii) 27 green biorefinery, (iii) lignocellulosic biorefinery, (iv) non-soluble protein recovery, (v) gasintermediate biorefinery, (vi) liquid substrate alternative, (vii) solid-substrate fermentation and 28 29 (viii) more-out-of-slaughterhouse by-products. The proposed framework aims to support 30 future research in waste recovery and valorization within food systems, along with stimulating reflections on the improvement of resources' cascading use. 31

32

# 33 Abbreviations

DAC, Direct air capture; DHA, Docosahexaenoic acid; DM, Dry matter; EC, Electrochemical
cell; HB, Haber-Bosch process; HOB, hydrogen-oxidizing bacteria; LCA: Life cycle
assessment; LPC, Leaf protein concentrate; MOB, Methane-oxidizing bacteria; MPF,
Macronutrient-poor feedstuff; MRC, Macronutrient-rich concentrate; SSF, Solid-substrate
fermentation; TC, Targeted compound; TSE, Transmissible spongiform encephalopathies;
VFA, Volatile fatty acids; WW, Wastewater.

# 40 Keywords

residual biomass, biorefinery, circular economy, microbial protein, insect, novel food,
 alternative feed, bioeconomy

43

44

#### 45 **1. Introduction**

The urgency to rethink the food system is increasingly attracting attention (IPCC, 46 2019; Searchinger et al., 2018; Willett et al., 2019). The current agrifood system consumes 47 about 70% of the world's freshwater (Sims et al., 2017), is responsible for at least 80% of 48 deforestation (Ramankutty et al., 2018), 30% of the overall energy consumption, annually 49 50 generates an estimated 20-30% of the anthropogenic greenhouse gas emissions (Rosenzweig et al., 2020) and is currently depleting non-renewable phosphorous ores 51 (Schoumans et al., 2015). One underlying flaw of the current agrifood system is its overall 52 inefficiency (Alexander et al., 2017b). For example, Schramski et al. (2020) showed that 53 today, roughly six calories of input energy yield just one calorie of consumable food-based 54 energy. Projected climate trends combined with forecasted demographic changes (including 55 overall population growth and its dietary preferences) depict a challenging picture for food 56 production capacities in 2050. For instance, the World Resources Institute estimates that 57 food demand will grow by 56% compared to 2010 levels, leading to an additional agricultural 58 land twice the size of India for food production to keep apace with demand (Searchinger et 59 al., 2018). On the other hand, Gerten et al. (2020) calculated that if the current agrifood 60 61 system was to operate within the safe zone of all planetary boundaries, it would only feed 3.4 billion people. Yet, because food is not a luxury, it is vital to identify solutions to sustain the 62 future food demand of the world's population. 63

There is consensus that resolving the food conundrum requires simultaneous actions 64 aimed at regulating food demand and consumption, improving production efficiency and 65 diminishing food losses (Billen et al., 2021; Clark et al., 2020; van der Goot et al., 2016). To 66 67 achieve these goals, it is possible to sustainably intensify agriculture, increasing production while lowering environmental impacts (Hamelin et al., 2021; Pretty, 2018; Rockström et al., 68 2017). Moreover, the emerging digitalization of agrifood systems is foreseen to boost 69 70 outputs, increase nutritional quality and enhance environmental performance (Herrero et al., 2020; World Economic Forum, 2018). Finally, the adoption of advanced technologies, such 71 as biotechnology, can procure novel foods and feed ingredients, providing nutritional 72 73 services comparable to those of current food- and feedstuffs (Alexander et al., 2017a; Ercili-Cura and Barth, 2021; Parodi et al., 2018; Torres-Tiji et al., 2020). The present study focuses 74 75 on the latter strategy, and particularly on solutions allowing to loop residual biomasses back 76 into the food chain, thereby decoupling food production from the demand of additional arable land. In this work, such solutions are generically referred to using the term "waste-to-77 78 nutrition", and "residual biomass" denominates both unused and underused biogenic wastes, 79 residues and co-products.

Residual biomass is a constrained resource (Hamelin et al., 2019), and many streams 80 are already supplying energy (e.g. via the biogas from anaerobic digestion), materials (e.g. 81 82 woodchips panel) and food-related services, either indirectly through agronomic valorization (e.g. manure and straws used as organic fertilizers) or direct use in animal diets (e.g. meals 83 84 from vegetable oil extraction) when feed standards are met (Mottet et al., 2017). Given the foreseen importance of residual biomass in future development narratives such as 85 86 bioeconomy (Muscat et al., 2021), competing value chains for these streams are emerging 87 (e.g. fine chemicals for cosmetics and pharmaceutics), prompting the need to ensure the implementation of the most efficient and cascading uses of these resources (Baldoni et al., 88 2021; Duque-Acevedo et al., 2020; Venkata Mohan et al., 2016). Valorization hierarchies 89 90 constitute a useful framework to tackle this challenge (EC Directive, 2008; Teigiserova et al., 2020). These suggest to privilege pathways where functional properties (e.g. proteins) are 91 safeguarded and directly valorized (e.g. into ingredients) before the implementation of lower-92

value cascading recovery pathways (e.g. nutrients or energy) (Garcia-Bernet et al., 2020;
Gómez-García et al., 2021).

95 The potential of residual biomass to directly (i.e. not through an agronomic valorization) produce food and feed ingredients has already been explored for specific cases such as 96 space travels (Clauwaert et al., 2017), agricultural catastrophes (Denkenberger and Pearce, 97 98 2015) or through livestock recycling (i.e. direct inclusion in farmed animal diets) (Rajeh et al., 2021; Van Hal, 2020; Van Zanten et al., 2019). Yet, to our knowledge, no comprehensive 99 attempts have been made to collate data and identify the multiple conversion pathways that 100 allow this. In an endeavor to fill this gap, the aim of the present work is to provide an 101 extensive overview of current and emerging waste-to-nutrition pathways. Overall, 660 102 scientific papers and 270 records from industrial literature, including patents, were reviewed 103 (review methodology available in Supplementing information; SI). The approach developed 104 does not allow to directly conclude on the environmental or economic relevance of the 105 reviewed value chains. Indeed, those aspects are beyond the scope of the present work and 106 are context-specific, requiring specialized assessment methodologies (e.g. life cycle or cost-107 108 benefit assessments). Nevertheless, as a stepping stone, this work is the first to detail, 109 classify and systematize in a single framework the main waste-to-nutrition pathways, facilitating their further comparisons. 110

#### 111 **2.** Scope: ingredients and biomasses considered

The umbrella categories of residual biomasses considered in this study are illustrated 112 113 in figure 1. The terminology used (i.e. waste, residue, co-product and by-product) for these categories, as well as throughout this study, is carefully chosen and based on the EU 114 legislation as further detailed in the SI. The scope of categories is an expansion and 115 harmonization of the streams described in Hamelin et al. (2019), also described in the SI and 116 briefly reported here. The category "Wood-related residual biomass" shown in figure 1 117 118 includes primary forestry residues (defined in Karan and Hamelin, 2020), pruning residues, wood-processing wastes (e.g. sawdust) and some packaging waste (e.g. cardboard). The 119 120 "Primary crop residues" category only includes straws, stalks and corn stover; tuber's top fractions (e.g. potato leaves) are included within the "Green residual biomass" category along 121 with garden- and park wastes (e.g. mowed grass). "Food waste" (discarded food stemming 122 from households or the service sector such as restaurants, etc.) and "Manure" (including all 123 124 types of livestock excreta, whether managed as slurry, solid or deep litter) are stand-alone categories. "Sludge" and "Wastewater" (WW) are grouped in a single category that includes 125 streams coming from both industrial (e.g. potato WW) and municipal (e.g. sewage sludge) 126 origins. The "Agrifood co-products" category encompasses streams from primary 127 transformation (e.g. bran, pulp, peels, spent grains, etc.) and secondary transformation (e.g. 128 fruit canning, bakery, etc.) of agrifood industries. Despite their frequent valorization as feed 129 (Chapoutot et al., 2019), these co-products host upgrading valorization potentials (Garcia-130 Bernet et al., 2020) and are therefore included in the scope of waste-to-nutrition pathways. 131 Because of their specific composition and of the regulation they are subjected to, 132 "Slaughterhouse by-products" (e.g. feather, carcass, bristle, etc.) are gathered in a distinct 133 category. Additionally, bioeconomy-related wastes (e.g. insect frass, digestates, etc.) and 134 135 specific wastes not stemming from the food sector (e.g. scrap newspapers) are added under the category "Others". Finally, nutrient looping pathways building on C<sub>1</sub> gaseous feedstocks 136 (CH<sub>4</sub>, CO<sub>2</sub>) are considered given their potential to decouple food production from land use 137 138 (Pikaar et al., 2018), whether these stem from a biogenic (e.g. resulting from biomass processing) or fossil origin. 139

140 Three general categories of ingredients supplying nutritional services are 141 distinguished. The first category encompasses macronutrient-rich concentrates (MRC) that

include energy- and/or protein-rich products, considered as high nutritional quality 142 ingredients. The macronutrient-poor feedstuff (MPF) category includes cellulose-rich 143 agricultural biomass (e.g. straw, grass, etc.) commonly used in animal husbandry as fodder 144 and roughage (Dale et al., 2009). The third category of ingredients, referred to as targeted 145 compounds (TC), are simple molecules used as additives to balance diets. These include a 146 147 wide variety of compounds from minerals and vitamins to amino acids. For the purpose of this review, the term TC describes compounds that confer direct nutritional benefits. 148 149 Consequently, functional additives such as antioxidants or enzymes are excluded.

For animals, the specific nutritional characteristics of MRC, MPF and TC are 150 combined in formulations to furnish balanced meals, often referred to as compound feed. 151 Formulation is performed based on precise knowledge of the animal's gut physiology, 152 nutritional requirements, health needs, legislative constraints, and an endeavor to minimize 153 the overall cost (Saxe et al., 2018). In the case of animals farmed for human consumption, 154 optimization also includes specific performance parameters (e.g., carcass lean meat, milk 155 yield, feed conversion ratio, etc.). On the other hand, human food is mainly restricted to MRC 156 and TC, even if the inclusion of ingredients in human diets also depends on cultural and 157 158 social habits (Teigiserova et al., 2020), with consumer acceptability being a key factor in relation to the commercialization of novel food ingredients (Aschemann-Witzel and Peschel, 159 2019; Rumpold and Langen, 2020). This present review does not explicitly address 160 161 consumer acceptability (figure 1), although it is reported when the issue is raised in the literature. Similarly, regulation aspects related to the entry of specific streams into food- and 162 feed-related markets are not specifically covered in this work. 163

164

#### 165

#### Figure 1. Scope of the literature review

Residual biomass categories, here illustrated by icons, are further detailed in the SI. Agronomic valorization (e.g. as fertilizer) is not part of the scope as this study focuses on the direct recovery of edible ingredients only.

168

#### 169 **3. Bridging the gap between waste and nutrition**

170 From a physicochemical perspective, a resource is considered as a food or feed ingredient when its "composition-structure characteristics" (Axelos et al., 2020) enter the 171 safety perimeter of the digestive tract, i.e. when the nutrients contained within the ingested 172 173 ingredients are released and assimilated without adverse effects. This safety perimeter is determined by the inherent features of digestive tracts and thus varies across different 174 species (Godon et al., 2013). The edibility, or nutritional quality of an ingredient is 175 multidimensional, but is often characterized by three main factors: (i) the absence of anti-176 nutritional compounds, (ii) the degree of structural complexity (i.e., biodegradability) and (iii) 177 the concentration of macro- and micronutrients. These determine to which extent an 178 179 ingredient can be considered as food grade (figure 2).

The term anti-nutritional compound refers to a substance that may damage the organism or prevent (or severely diminish) proper nutrient absorption (Makkar, 1993). These are quite variable in nature, ranging from heavy metals to plants secondary metabolites and mycotoxins (Salami et al., 2019). Generally, (human) food regulations explicitly require exhaustive proof of the absence of anti-nutritional compounds in novel ingredients (EFSA Panel on Dietetic Products, Nutrition and Allergies (NDA), 2016).

186 Structural complexity is particularly prevalent in fibrous biomass, such as 187 lignocellulose or keratin. These structures are highly robust and resist the chemical and 188 enzymatic reactions of common digestive processes. Therefore, the ability of monogastric animals to degrade MPF is quite limited, while the multiple intestinal tracts of ruminants (and termites gut microbiomes) are adapted to the digestion of cellulose-based structures (Godon et al., 2013). However, even ruminants only display very limited ability to digest lignin (Chapoutot et al., 2010; Moore and Jung, 2001) which constitute a highly resistant, hydrophobic barrier that survives most biodegradation processes (Triolo, 2013).

Finally, the nutritional quality of an ingredient is shaped by the presence and quantity of digestible nutrients such as proteins, lipids and carbohydrates, often correlated with the absence of structurally complex macromolecules, anti-nutritional compounds, inorganic compounds (e.g., ashes) and moisture. The latter particularly affects the stability of the nutritional quality over time, given the interrelation between moisture levels, fermentable compounds and microbial growth (Teigiserova et al., 2019).

200 As illustrated in figure 2, most residual biomasses cannot be considered as food grade. Anti-nutritional factors often arise because of increased heterogeneity and/or 201 biological activity (e.g. in food waste) when not intrinsically linked to the feedstock (e.g. 202 activated sludge hosting toxic organic and inorganic substances, see section 5.3.2.). 203 Moreover, streams from agricultural and forestry activities (primary crop residues, wood-204 205 related and to some extent green residual biomass) are rich in lignin, cellulose and hemicellulose, which together form composite, recalcitrant matrices that are incompatible 206 with direct edibility. Accordingly, to bridge the gap between waste and nutrition, pathways 207 require to implement a sequence of operations that breakdown structural barriers, remove 208 noxious compounds and, if required, enrich the assimilable nutrient content. In other words, 209 the initial composition-structure (i.e. position in figure 2) are defining properties that not only 210 211 determine the direct nutritional value of waste, co-products and residues, but also determine their technical and economic potential as nutritional sources. 212

- 213
- 214

#### Fig. 2. Waste-to-nutrition gap

Ternary diagram representing food grade quality perimeter (gray right corner), and approximating relative location of the studied solid residual biomass streams (colored circles). Phenolic lignin acts as both structural complexity and anti-nutritional proxies, but the latter is here privileged to differentiate wood-related residual biomass from green residual biomass and primary crops residues. MPF ingredients perimeter is not represented for tractability reasons. For the same reason, agrifood co-products and slaughterhouse by-products are gathered within the same broad circle (dotted line).
\*Albeit some slaughterhouse by-products (e.g. offal) are directly edible (within the gray right corner), others are mainly composed by keratin which is here considered as structural content (top corner), see section 5.2.2.

mainly composed by keratin which is here considered as structural content (top corner), see section 5.2.2. Background data is available in the SI database and icons are as defined in figure 1.

224

223

# **4. Describing waste-to-nutrition pathways using four building blocks**

The systematic analysis of reviewed literature (see database in SI) revealed that the 226 series of unit operations and processes implemented in waste-to-nutrition pathways vary 227 according to two main considerations: (i) the degree to which the feedstock input 228 composition-structure is altered, and (ii) the targeted nutrient recovery ratio (i.e., input versus 229 230 desired output). These are represented as a scale gradient in the X-Y axis of figure 3. Along this scale, four generic processes are proposed as pathway building blocks: (i) 231 enhancement, (ii) cracking, (iii) extraction and (iv) bioconversion. These are one key result 232 arising from the transversal interpretation of the present review, and were defined with the 233 aim to provide an interpretative framework highlighting common trends in waste-to-nutrition 234 235 pathways. In fact, conversion pathways can be described as a sequential workflow of processes using these modular building blocks. Notably, the four building blocks proposed 236

herein are in line with previous classification works (Colonna, 2020). Importantly, these blocks do not relate to specific technologies, but rather reflect the main macroscopic changes resulting from a set specific process when applied on a feedstock or product. Finally, the modular block representation (e.g. similarly used in Verstraete et al. 2016) reveals how different unit operations can be combined to generate varying degrees of nutritional quality and process intensity.

#### 243 **4.1. Enhancement: low composition-structure change, high nutrient recovery**

Nutritional enhancement refers in this study to the application of one or several unit 244 245 operations to augment the accessibility, preservation or quantity of nutrients, without the 246 removal of any components. Accessibility is mostly increased by inducing structural changes that render the nutrient more attainable (i.e. macro-fractionation), whereas preservation is 247 usually achieved through stabilization (e.g. water removal, homogenization, etc.). Nutrient 248 enrichment can be realized using methods such as solid substrate fermentation (SSF). In this 249 case, nutrient enrichment is mainly the result of either the partial degradation of fibers, the 250 251 development of microbial proteins (e.g. mycelium colonization), or both.

252 Albeit minor losses might occur (typically ranging between 5-30% of the dry matter during ensiling (Borreani et al., 2018) or other SSF (Castoldi et al., 2014; Rajesh et al., 253 254 2010)), enhancement processes recover a major share of the initial nutrients in the final product, while safeguarding the global composition-structure characteristics. Indeed, 255 modifications are generally limited to the macro- and mesoscopic scales (e.g. for 256 comminution or drying), although some impacts at the microscopic scale are possible. The 257 latter particularly applies for SSF as this process is based on microflora activities, hence 258 generating microscopic changes (e.g. lignin mineralization). However, these induced 259 changes remain partial and limited provided that the fermentation process is stopped before 260 significant quantities of nutrients are converted. When this condition is fulfilled, the product 261 262 displays compositional and structural properties that resemble that of the feedstock residue (e.g., ensiled grass versus raw grass), and therefore enters the enhancement building block. 263

264

# **4.2. Cracking: high composition-structure change, high nutrient recovery**

266 Through the literature reviewed, the release of nutritional compounds entangled in extremely recalcitrant structures and/or locked chemically into macromolecules (e.g., glucose 267 in cellulose) are only achieved through biomass deconstruction, hereafter referred to as 268 269 cracking (Axelos et al., 2020). Cracking requires several process steps. It typically involves a first physico-mechanical pretreatment (e.g., hydrothermal, steam-explosion) which denatures 270 organized macromolecular networks. Afterwards, macromolecules are subjected to some 271 degree of lysis (e.g., enzymatic, hydrothermal, chemical), thus yielding smaller platform 272 molecules (Farmer and Mascal, 2015) and unleashing chemical functions (Colonna, 2020; 273 De Jong et al., 2020). Although the removal of nutrients is not a desired outcome of cracking 274 processes, minor losses do occur. In recent examples, cracking led to the recovery of 73% of 275 276 amino acids present in bristle keratin (Falco et al., 2019), while commonly more than 90% of 277 sugars are recovered from wood-based cellulose (Wyman et al., 2009).

278

#### 4.3. Extraction: low composition-structure change, low nutrient recovery

Extraction includes all unit operations and processes that selectively solubilize and/or separate a target fraction from a matrix, while safeguarding its initial functional properties (Jimenez et al., 2015). Some extraction processes are hybrid, combining the features of both extraction and cracking processes (e.g., alkaline extraction). However, extraction processes differ from cracking in as much that the targeted compound or fraction is not necessarily a structural component of the feedstock and is generally a minor fraction (e.g., proteins in tomato seeds). Unlike cracking, extraction does not induce generalized molecular-scale disruptions (Gençdağ et al., 2020; Rodriguez-Lopez et al., 2020). Extraction often involves a sequence of separation processes (e.g., precipitation and filtration) and isolation processes, all included within the extraction building block.

290 According to the literature reviewed, the recovery potential of an extraction step is limited by (i) the amount of the targeted TC available in the feedstock and (ii) the maximum 291 achievable yield using the extraction technique. The latter is heavily dependent on the 292 compound-structure interactions and inversely correlated to the desired purity (Colonna, 293 2020; Tamayo Tenorio et al., 2018). For example, considering proteins present in green 294 residual biomasses (<20%DM), only a fraction (5-45% of total) is recovered using the 295 extraction techniques described in section 5.3.3. (Santamaría-Fernández and Lübeck, 2020). 296 Similarly, all common downstream separation and purification processes used to obtain TC 297 such as valuable fatty acids (e.g. docosahexaenoic acid: DHA) compliant with market 298 299 specifications imply mass and nutrient losses. For example, in weight terms, 1 to 15 units of 300 DHA is obtained per 100 units of microalgae or aquatic protists (Russo et al., 2021; Xiangping et al., 2019). The reviewed literature often highlighted this particular point: 301 302 extraction processes typically generate significant quantities of side streams whose 303 synergetic valorization is key to ensure economic sustainability (Teekens et al., 2016).

304

#### 305 **4.4 Bioconversion: high composition-structure change, low nutrient recovery**

306 Both microorganisms and animals retain and concentrate the nutrients they ingest, 307 integrating them into a variety of products, including their own cellular or body mass. This is achieved through bioconversion, which refers in this study to the conversion of feedstocks 308 into nutritional ingredients using the metabolic processes of living organisms. Bioconversion 309 yields a relatively low nutrient recovery, intrinsic to the fact that part of the feedstock is 310 converted to non-edible biomass or oxidized to gases instead of being recovered in the 311 edible product (i.e., meat, mushroom, etc.). Major losses are due to respiration (carbon-rich 312 gases), nitrogen-rich excretions and heat (El Abbadi and Criddle, 2019; Parodi et al., 2020; 313 314 Wirsenius, 2000). To provide concrete examples, a benchmark of bioconversion efficiency figures was derived from the literature review, including both livestock, insects and 315 microorganisms-related products (background data in SI). It indicates that even for optimized 316 317 species and farming conditions, hardly more than 50% and 30% of respectively proteins and calories invested as feedstuff are recovered within animal-based food products. Reported 318 319 values are slightly higher for insect farming in ideal conditions, yielding up to 70% of proteins 320 recovery and 30% for calories recovery into insect biomass. Finally, benchmarked values for 321 edible microorganisms, despite being highly dependent on specific strain, culture conditions and metabolic pathway, suggest that their energy conversion into edible calories ratio is 322 generally below 30-40%. The aforementioned values illustrate the highest bioconversion 323 efficiencies encountered; however, it should be highlighted that these efficiencies are closely 324 325 tight to the nutritional quality of input feedstock. Bioconversion efficiencies shrink rapidly as 326 the input feedstock's nutritional quality (or nutrient availability) decreases (details in SI).

For convenience, microbial and farmed animal bioconversion sub-groups are further distinguished. Microbial bioconversion encompasses the use of microorganisms both as biocatalysts that produce enzymes and nutritional TC such as vitamins, amino acids or flavor compounds (Specht and Crosser, 2020; Sun et al., 2021; Yang and Xu, 2016), and as final

standalone nutritional MRC themselves. Currently, this latter function is encountering 331 renewed interest, especially regarding so-called "single cell proteins" i.e., microbial proteins 332 and fungal proteins (Ciani et al., 2021; Hüttner et al., 2020; Linder, 2019; Singh et al., 2020; 333 Tubb and Seba, 2019) which cover organisms that generate high protein (up to 70-75%DM) 334 content (Pikaar et al., 2018). Similarly, single cell oils (Ochsenreither et al., 2016) and all 335 microbial-based fermentation and cell-culturing aiming at substituting common food products 336 (Crosser et al., 2019; Lv et al., 2021; Specht and Crosser, 2020) are also considered within 337 338 this bioconversion building block.

Farmed animal bioconversion refers to the use of livestock to produce food from nonedible biomass (Boland et al., 2013; Smith et al., 2013). This includes ruminants (e.g., that convert lignocellulosic biomass into milk), but also monogastric livestock (e.g., swine) whose potential role in upcycling residual biomass into foodstuff is also highlighted in the literature reviewed (ten Caat et al., 2021; Van Zanten et al., 2019). The use of insect (entomo-) farming as waste-to-nutrition bioconversion strategies is also part of this sub-category.

345

346

#### Fig. 3. Waste-to-nutrition pathways in four building blocks

Four generic families of conversion processes, illustrated with examples from the literature. Icons represent
 residual biomass categories as defined in figure 1 From top-down and left-right: (i) Fermented olive press-cake as
 fodder. (ii) Brewer' spent grains milled into bakery flour. (iii) Extraction of proteins from grass. (iv) Carbohydrates
 recovery from organic wastewater. (v) Recovery of cellulosic sugars. (vi) Feather processed with keratinases
 releasing amino acids. (vii) Insects farming on food waste and (viii) Microalgae cultured on aquaculture
 wastewater.

353

#### 354 **5. Waste-to-nutrition pathways debunked**

355 Each waste-to-nutrition pathway derived from the literature review is expressed as a combination of building blocks, representing the different process steps. Advantageously, 356 this provides a means to detect common patterns among the different waste-to-nutrition 357 pathways, while also representing their diversity. Most pathways are built upon a core 358 conversion unit, with accessory units being usually referred to as pre- and post-treatments. 359 The choice of these accessory units varies depending on the exact nature of the initial 360 361 feedstock and the target nutritional market i.e., the waste-to-nutrition gap as defined in figure 2. Accordingly, in this section, waste-to-nutrition pathways are grouped on the basis of the 362 common core building block. Each sub-section is illustrated with (i) a table reporting a 363 selection of related examples from the literature and (ii) a figure representing the unit 364 operations sequence pattern (table 1-3 and figure 5; other figures and tables available in SI). 365

366

#### 367 **5.1. Direct upgrading: nutritional enhancement pathways**

Nutritional enhancement pathways upgrade residual biomass into food and feed ingredients using a single, or multiple enhancement steps, as illustrated in figure S1 of the S1. Concrete examples are provided in table 1.

For animal feed, enhancement pathways typically increase the digestibility of fibrous materials by disrupting the complex plant cell wall matrix and releasing macromolecular structures, such as polysaccharides in lignocellulosic feedstocks. In addition to breaking down structural barriers, enhancement strategies sometimes achieve a net nutritional enrichment of the feedstock. This is the case for ammonia-fiber expansion treatments increasing the total nitrogen content of ruminant forage (MPF), but also for SSF treatments

provoking protein and/or lipid enrichment. SSF treatments are often applied after a first 377 mechanical enhancement step. Used on lignocellulosic residual biomass, SSF allows to 378 selectively degrade lignin while avoiding microbial polysaccharide consumption (e.g., using 379 white rot fungi) (van Kuijk et al., 2015; Villas-Boas et al., 2002), and thus provides access to 380 a wider range of lignocellulosic biomass for use as MPF. In the case of monogastrics, 381 enhancement pathways are employed to substitute MRC, using residual biomass with low 382 lignin content as feedstock for SSF aiming to convert part of the polysaccharide fibers into 383 assimilable nutrients (e.g., free sugars and proteins) (Patil et al., 2020; Villas-Boas et al., 384 2002; Wongputtisin et al., 2014). Consequently, the product is characterized by higher 385 nutrient availability and content and displays improved palatability. As conventional 386 387 aquaculture uses high quality feed products (high protein and lipid digestibility), the inclusion of mildly treated fibrous materials is often avoided (Leduc, 2018). Nevertheless, SSF 388 transformation units are used to generate alternative feed products for aquaculture, by 389 390 improving digestibility of non-lignocellulosic feedstocks such as feather meal and isolated plant-based proteins (Dawood and Koshio, 2020; Hamidoghli et al., 2020). 391

392 For food markets, nutritional enhancement can be used to tailor organoleptic 393 properties (e.g., texture, taste) of streams that are edible, but unappealing to consumers. These feedstocks mainly enter the agrifood co-products category (e.g., apple pomace, 394 bakery surplus) (Gmoser, 2021; Sabater et al., 2020; Souza Filho, 2018). The combination of 395 mechanical and/or SSF enhancement steps render these co-products suitable for direct 396 397 consumption or for inclusion in processed food products (e.g., as flour) in bakeries, drinks or meat-alternatives (Torres-León et al., 2018), often with unlocked bioactivity properties 398 399 (Leonard et al., 2021).

Residual biomass	Barriers to direct edibility	Transformation units	Results	Final use	Status and references
Sunflower shell	High lignin content (50%DM)	Drying, milling, mixing	Unlock carbohydrates, added directly in pellets	Ruminant forage	Lab-experiment: (Osman et al., 2018)
Almond hulls	Total phenolics content (106g/kgDM)	3-5% urea solution moisturizing, covering for several weeks	Can substitute alfalfa in diets without adverse effects	Ruminant forage	Feeding trial: (Rad et al., 2016)
Sugarcane crop residues	Recalcitrance to digestion due to lignocellulose	Ammonia-fiber expansion (ammonia and steam, 100- 130°C)	Digestibility improved for ruminant (true digestibility, metabolizable energy, total nitrogen content)	Ruminant forage	Lab-experiment: (Mokomele et al., 2018)
Olive cake	Anti-nutritional content: phenols	<ol> <li>Ground, sieved 5mm, sterilized 20min, 121°C, moistened</li> <li>SSF: <i>Fusarium flocciferum</i> fungal strain: 2 weeks, 25°C</li> </ol>	Increase of protein content up to 94%, decrease of phenolic content by 70%	Ruminant feed	Lab-experiment: (Chebaibi et al., 2019)
Groundnut shells, pigeon pea husk, wheat bran	Anti-nutritional content and lignin structure	1. Pre-washed, sundried and pulverized residues 2. Supplementation with 2% ammonium nitrate and glucose and SSF with fungal strain <i>Colletotrichum</i> <i>spp</i> , 21 days, 30°C	Cellulose, hemicellulose, lignin, tanins and phytates contents were reduced and carbohydrates and proteins increased. Successfully added in poultry diets.	Monogastric feed	Lab-experiment: (Patil et al., 2020)
Cheese whey, molasse, fruits pulp, spent grains and rootlets	Mixed biowaste (heterogeneity)	<ol> <li>Solid substrate autoclaved 15min, 120°C</li> <li>SSF with <i>Kluyveromyces marxianus</i> for 4 days, 30°C</li> <li>Optional lipids extraction</li> </ol>	Protein and lipid content doubled. Extraction of lipids and press-cake as protein-rich animal feed ingredients	Animal feed	Lab-experiment: (Aggelopoulos et al., 2014)
Feather meal	Keratin content: low digestibility	1. Autoclaved 100°C, 15min 2. SSF with <i>Bacillus subtilis</i> bacterial strain 50°C, pH8, 72h	Can substitute fish meal up to 20% in silver pompano diets	Aquaculture feed	Lab-experiment: (Adelina et al., 2020)
Fish by- products	Acceptability	Cleaning, drying, milling	Increased acceptance of edibility, and enhancement of conservation	Human food	Lab-experiment: (Abbey et al., 2017)
Apple pomace	High perishability and moisture	Cleaning, drying, milling	Can substitute flour in bakery with dietary improvements	Human food	Lab-experiment: (Lyu et al., 2020; Zlatanović et al., 2019)
Brewer's spent grain	Organoleptic properties: bitter taste and unpleasant mouthfeel	Cleaning, drying, milling	Can substitute flour in snacking and pasta formulation	Human food	Lab-experiment: (Nocente et al., 2019), patent: (McHugh et al., 2020) with early commercial use (ReGrained, 2020)
Fruits bagasse and peels	Acceptability	<ol> <li>Sanitized, dried 55°C and ground</li> <li>SSF: Saccharomyces cerevisiae strain, 30°C, 70% moisture</li> <li>Homogenization and dried 55°C.</li> </ol>	Protein content increased 11 times. Can be included in cereal bars with improved censorial attributes and purchase intention	Human food	Lab-experiment: (Muniz et al., 2020), Similar process of (Villas-Boas and Granucci, 2018) under commercial development (Green Spot, 2020)

#### 400 Table 1. Selection of inventoried nutritional enhancement pathways – reported with wording used by original references

401 SSF: Solid Substrate Fermentation

#### 402 **5.2. Cracking pathways: Unlocking nutrients in structural biomass**

To supply nutritional services, the literature reveals that cracking processes form the core of two conversion pathway categories: (i) the recovery of edible sugars and fibers from lignocellulosic streams and (ii) the recovery of amino acids and bioactive peptides from slaughterhouse by-products. The proposed unit operation pattern and a selection of relevant examples of cracking pathways are illustrated in the SI (figure S2 and table S1).

408

#### 409 **5.2.1. Lignocellulosic feedstock to sugars and dietary fibers**

Lignocellulosic feedstocks are characterized by three interlinked macromolecules: (i) cellulose, (ii) hemicellulose and (iii) lignin representing 38-52%, 15-30% and 10-40% of the dry matter, respectively (Kapu and Trajano, 2014). Cellulose is a homopolymer composed of glucose, while hemicellulose is a generic term for  $\beta$ -1,4-linked non-cellulosic plant-based polymers (Scheller and Ulvskov, 2010). The most abundant class of hemicelluloses are xylans that are mainly composed of pentoses (i.e., C<sub>5</sub> sugars like xylose and arabinose).

416 Prior to cracking, lignocellulosic materials must be pretreated, using comminution methods to reduce particle size, partially disintegrate the plant cell wall matrix and promote 417 lignin removal. Afterwards, within the cracking process hemicelluloses and cellulose are 418 hydrolyzed, procuring "wood molasses". These products were originally used as nutritional 419 ingredients (Harris, 1947), but more recently have been driven towards chemical and energy 420 markets (Reese et al., 1972). When cracking is coupled to downstream separation and 421 purification, exploiting the different solubilities of cellulose and hemicellulose, it is possible to 422 isolate pure sugar streams (Ingle et al., 2020). Purified cellulose can be used to supply the 423 glucose or starch markets (You et al., 2013). However, wood-based glucose is currently 424 425 uncompetitive compared to sugar-beet or sugarcane, regarding both economic and environmental aspects (Bello et al., 2021; Denkenberger et al., 2019). Finding markets for 426 427 pentose sugars is less straight-forward, because their nutritional and fermentable value is 428 lower than that of hexose sugars (Huntley and Patience, 2018; Rolston and Mathan, 1989). 429 However, partial hydrolysis of hemicelluloses procures pentooligosaccharides (e.g. xylooligosaccharides) that can be used as prebiotic food ingredients (Poletto et al., 2020). 430 431 Moreover, further functionalization of monomeric pentoses yields molecules such as the lowcalorie sweetener, xylitol (Chandel et al., 2018; Franceschin et al., 2011). Finally, although 432 the nutritional value of polyphenolic lignin is rather marginal, it has limited use in food 433 industry as a texturizer or emulsifier (Bhat et al., 2020; Tenlep, 2020). 434

For food applications, product purity is of prime importance, because high severity 435 436 (e.g. high temperature, pH changes) cracking processes often generate undesirable products 437 and neoformed chemical species, such as furfural or acetic acid (Venkateswar Rao et al., 438 2016). The hydrolysates detoxification stage (i.e., removal of anti-nutritional compounds) is 439 thus often a prerequisite for commercialization (Domingos et al., 2020; Dupoiron et al., 2017). In this regard, the use of alternative strategies, such as preventive pretreatment 440 441 and/or enzyme-mediated hydrolysis might be advantageous and provide economically more 442 viable routes to target food ingredients from lignocellulose (Paës et al., 2019; You et al., 443 2013). Enzymes act as highly selective catalysts that operate in mild (e.g., low temperature 444 and near neutral pH) conditions and do not generate neoformed compounds, thus potentially positively influencing costs (Ingle et al., 2020; Roth et al., 2020). Consolidated bioprocesses 445 446 involving the *in-situ* production of enzymes are often preferred for economic reasons, albeit requiring an additional stage of bioconversion to produce them. 447

448

#### 449 **5.2.2.** Amino acids recovery from slaughterhouse by-products

Slaughterhouse by-products are here defined as low value animal body parts (offal, carcass, bristle, etc.) generated during meat production, which are included in the category 3 of the EU Animal By-products Regulation (see SI). These represent 35-50% of the animal body in weight (Alao et al., 2017; Ferraro, 2020). Aquaculture trimmings (30-75% of the fish in weight) are also included in this category (Leduc, 2018). These by-products share the fact that they are protein and/or lipid-rich, despite their quite different physical and chemical properties.

457 Meat scraps, offal, blood, bones and assimilates group into a single category of mostly edible (albeit unappealing to certain populations) slaughterhouse by-products that can 458 be consumed directly or after organoleptic enhancement (Said, 2019; Toldrá et al., 2016). 459 460 Being mostly devoid of structural barriers, this category of slaughterhouse by-products is commonly transformed using rendering processes into protein-rich meal for livestock and 461 pets (e.g., bone meal, meat meal), fats and oils, while recovering functional compounds such 462 as gelatin and collagens (Paul et al., 1962). Rendering mostly involves heat and mechanical 463 treatments, like the processes used to generate fish meal and oil (Aspevik et al., 2017). 464 465 Therefore, rendering is an enhancement technology. Unfortunately, health and safety concerns related to the use of slaughterhouse by-products (e.g., the risk of transmissible 466 spongiform encephalopathies or TSE diseases) mean that these protein-rich co-products are 467 currently forbidden for use as animal feed in several countries. For this reason, cracking 468 processes are appropriate to treat slaughterhouse waste, because these can mitigate risk 469 and satisfy regulatory bodies. For example, protein hydrolysate produced from 470 471 slaughterhouse by-products is authorized for the feed market under specific conditions (Aspevik et al., 2017; European Commission, 2020). The second category of slaughterhouse 472 by-products consists of inedible fibrous residues (e.g., animal horn and bristle) mainly 473 composed by keratin (Ferraro et al., 2016; Perta-Crisan et al., 2021). When mildly 474 processed, keratin-based residues provides low grade feed ingredients (e.g., feather meal), 475 providing that it is not banned by TSE-derived regulations (Heuzé V. et al., 2020). 476

477 Because the direct use of animal by-products proteins is hampered by safety, health concerns and/or low digestibility, cracking processes are often used to obtain amino acids. 478 479 For edible slaughterhouse by-products, enzymatic proteolysis is the preferred method to 480 achieve this, because enzyme selectivity provides the means to generate not only amino acids, but also peptides displaying specific biological activities (Ferraro et al., 2016; 481 Martínez-Alvarez et al., 2015), without the risk of generating unwanted substances. However, 482 releasing amino acids contained in keratinous slaughterhouse by-products requires higher 483 severity methods (e.g. higher temperatures) to breakdown the keratin polymer network 484 (Chaitanya Reddy et al., 2021; Holkar et al., 2018). Such cracking processes are both energy 485 486 demanding and poorly selective, leading to the denaturation of certain amino acids/peptides and reduction of the potential nutritional value (Falco, 2018; Tasaki, 2020). Fortunately, 487 488 recent research on enzymes has revealed keratin-specific proteases that can convert keratin into highly digestible nutrients for animals (Chaudhary et al., 2021; de Menezes et al., 2021; 489 490 Prajapati et al., 2021). The addition of a specific bioconversion stage to produce the 491 enzymes is not necessarily required, as illustrated by a recent experience which performed a simultaneous in-situ keratinase production and keratin hydrolysis using the filamentous fungi 492 Amycolatopsis keratiniphila on bristle (Falco et al., 2019). Irrespective of the slaughterhouse 493 494 starting material, the generation of protein hydrolysates generally requires a subsequent purification step to produce the final, marketable product that meets food and feed standards 495 (Martínez-Alvarez et al., 2015; Tasaki, 2020). 496

#### 498 **5.3. Extraction of nutritional compounds from residual biomass: pathways**

499 Several waste-to-nutrition pathways reviewed are based on the extraction of TC or 500 MRC ingredients trapped within residual biomass. Reported TC extractions from residual biomass are mainly targeting secondary metabolites additives (e.g. tannins, polyphenols or 501 bioactive fibers) (Ben-Othman et al., 2020; Hussain et al., 2020; Rodríguez García and 502 503 Raghavan, 2021; Saha and Basak, 2020; Tlais et al., 2020) and thus are not included in the scope of this work. Regarding MRC, the extraction pathways reviewed mainly target protein 504 recovery (Pojić et al., 2018; Sari et al., 2015b; Tamayo Tenorio et al., 2018). Indeed, from 505 506 both economic and energetic standpoints proteins are the costliest macronutrients obtained from photosynthesis (Bentsen and Møller, 2017). Considering that they generate underused 507 508 protein-rich streams, three residual biomass categories are the focus of growing attention: (i) 509 agrifood co-products, (ii) (activated) sludge and (iii) green residual biomass. The waste-tonutrition pathways required to upgrade these different streams are similar in as much that 510 they all involve a series of extraction unit operations and are devoid of bioconversion and 511 cracking steps (proposed unit operation patterns and relevant examples are displayed in the 512 513 SI, figure S3 and table S2).

514

#### 515 **5.3.1. Protein recovery from agrifood co-products**

Some agrifood co-products, mostly from cereal (e.g., wheat bran, 13% DM proteins) 516 517 and oilseed co-products (e.g., canola press-cake, 40% DM proteins) contain proteins 518 enmeshed in lignocellulosic matrices (Contreras et al., 2019). While these co-products are already widely used as animal feed (section 5.5.), the application of extraction technologies 519 can extend their nutritional potential up to food-grade markets. The first extraction step is 520 521 employed to release the proteins from the residual matrix, for example through alkaline or 522 enzyme-based extraction (Baker and Charlton, 2020; Kamal et al., 2021; Sari et al., 2015a). Afterwards, target purity is often achieved using a sequence of precipitation and membrane 523 524 filtration. In the case of co-products already under a liquid form (e.g., dairy industry), recovery pathways mainly involve membrane-based extraction sequences (Lakra et al., 2021; Shahid 525 et al., 2021). Overall, the reviewed literature reveals two main extraction strategies: 526

- 527 (i) A stepwise method providing the means to recover proteins from specific
  528 single protein-rich feedstocks, such as canola, sunflower (Subaşı et al., 2021;
  529 Tan et al., 2011), distiller's grains (Roth et al., 2019) and lupine meal (Prolupin
  530 GmbH, 2020). This approach is already close to the commercial scale
  531 (Mupondwa et al., 2018).
- (ii) Cascade methods that allow the recovery of a single protein extract from several feedstock streams within a process that simultaneously isolates a variety of products. These methods can be applied to low-protein content materials such as primary crop residues (e.g., wheat straw with a protein content around 4%DM). This is advantageous because the low protein content of such feedstocks is offset by their high availability (Contreras et al., 2019; Hamelin et al., 2019).

539 Current extraction methods need further environmental and cost optimization to allow 540 effective recovery (Baker and Charlton, 2020) and lead to competitive production. One 541 solution highlighted in the literature lies in pairing conventional extraction processes with 542 microwave and ultrasound technologies: besides often increasing protein extraction yields, 543 these technics can also reduce operational expenditure (Franca-Oliveira et al., 2021).

544

#### 545 **5.3.2. Protein recovery from activated sludge**

Activated sludge is the solid fraction remaining after the biological treatment of WW. 546 547 Activated sludge is mainly composed of microbial biomass and is rich in proteins (up to 60%DM). Albeit sometimes directly reused for example in "sewage fish farms" (Verstraete et 548 al., 2016), the presence of noxious compounds (e.g., pathogenic bacteria or detergent) 549 550 usually prevent its direct use as animal feed (Vriens et al., 1989). Therefore, several studies have investigated the means to extract proteins implementing "sludge-to-proteins route" 551 (Xiao and Zhou, 2020). In addition to the high moisture content of sludge, another 552 disadvantage is that sludge proteins are contained within microbial cells. Therefore, it is 553 necessary to use a first stage, such as hydrothermal and ultrasound treatments, to 554 disintegrate microbial cells and release proteins. Afterwards, the recovery stage yields up to 555 556 90% of the proteins and the removal of most noxious compounds, such as heavy metals (Gao et al., 2020; Liang et al., 2020; Zhang et al., 2018). The resulting sludge protein 557 concentrates display a complete amino acids profile and are investigated to supply the 558 animal feed markets (Belyaev et al., 1978; Hwang et al., 2008; Markham and Reid, 1988). 559 However, to drive sludge proteins into the food sector, it is necessary to implement more 560 561 intensive, cost prohibitive purification methods (Xiao and Zhou, 2020). Nevertheless, to obtain food-grade proteins, it might be more reasonable to focus on sludge derived 562 exclusively from WW treatment facilities associated with food transformations units (e.g., 563 564 brewer effluent, bakery effluent), with the drawback of limited available volumes (Vriens et al., 1989). 565

566

#### 567 **5.3.3. Protein recovery from green residual biomass**

568 Green biomass refers to all photosynthetic organs of plants, such as grass, vegetable 569 tops and leaves. Those all harbor significant quantities of Rubisco, which is the key CO<sub>2</sub>fixing enzyme in plants. Although crude protein levels vary among species and as function of 570 571 pedoclimatic conditions, they nevertheless represent 10-25% DM of green biomass, of which up to 50% is soluble (Solati et al., 2018). A specificity of green biomass compared to general 572 plant-based biomass, is their low lignin content (<10%DM) coupled to high moisture, typically 573 well above 70% of total weight (Tamayo Tenorio et al., 2018; Triolo, 2013). These 574 characteristics are compatible with the mechanical separation of the freshly harvested green 575 576 biomass into two fractions: a nutrient-rich juice and a fiber-rich cake, each harboring around 50% of the initial proteins (Kromus et al., 2008). This first mechanical extraction process is 577 578 the starting point of most protein recovery in green residual biomass conversion pathways.

579 To maximize protein recovery, common extraction units such as thermal coagulation 580 followed by centrifugation and drying are commonly performed on the green juice. The resulting leaf protein concentrate (LPC) (Davys et al., 2011; Pirie, 1971) targets monogastric 581 582 livestock market, as a substitute for soy meal (Stødkilde et al., 2019). The implementation of 583 additional refining steps can lead to food-grade extracts (Di Stefano et al., 2018; Martin et al., 584 2019). The fiber-rich cake mostly contains non-soluble proteins, hence preventing their direct 585 recovery. However, this cake can be used as a substitute for raw grass in ruminant diets, while additionally providing the means to reduce nitrogen excretions (Damborg et al., 2020; 586 587 Lucci et al., 2019). Advantageously for this fraction, the initial mechanical fractionation of 588 e.g., grass increases the overall accessibility of grass proteins and thus offsets the absence 589 of soluble proteins present in the green juice fraction (Damborg et al., 2018).

590 While a wide panel of residual biomass for LPC production had been historically 591 screened (Pirie, 1971; Rosas Romero and Diaz, 1983), to-date commercial-stage 592 developments are only based on premium green crops such as alfalfa. Monogastric-grade alfalfa LPC is already commercialized (Andurand et al., 2010) while alfalfa food-grade 593 extracts are recently entering markets (Luzixine, 2020; Tereos, 2020). However, numerous 594 595 European-based consortia attach to widen the panel of LPC production feedstocks, such as LPC production from raw grasses (Agroväst, 2020; Go Grass, 2020), green cuttings 596 (GrasGoed, 2020) and vegetable tops such as sugar-beet (Green Protein Project, 2020; 597 598 Tamayo Tenorio, 2017). The seasonality, heterogeneity and perishability of green biomass, 599 coupled to energy-intensive technologies that procure insufficiently high protein yields are increasingly driving LPC production towards green biorefinery schemes in which LPC is just 600 one of several added-value products (Corona et al., 2018b; Djomo et al., 2020; Santamaría-601 602 Fernández et al., 2020). Depending on local contexts, diverse green biorefinery setups and 603 schemes exist, each aiming to supply energy, material and chemicals (e.g. lactic acids) from the LPC co-products (i.e. fiber-rich cake and supernatant "brown" juice) (Corona et al., 604 605 2018a; Kamm et al., 2016; Kiskini, 2017; Parajuli et al., 2018). Similarly, focus has also been 606 put on the recovery of proteins in cellulosic bioethanol biorefineries (Bayat et al., 2021), using for this purpose non-residual biomass streams, such as switchgrass (Bals and Dale, 2011; 607 Kammes et al., 2011; Laser et al., 2009). 608

609

#### 610 **5.4. Microbial bioconversion pathways**

The literature survey revealed that the use microorganisms to recover and 611 concentrate nutritional products from residual biomass is a well-studied route. Obviously, the 612 term "microorganism" embraces an extraordinarily large number of species. Therefore, 613 waste-to-nutrition pathways reviewed herein are classified according to the main metabolic 614 processes involved, consistent with previous works (Jones et al., 2020; Spalvins et al., 615 2018). Resulting sub-categories are displayed in figure 4, and mainly differ regarding the 616 617 preferred carbon and energy sources of the microbes. The categories are indicative, because some microorganisms are mixotrophs, being capable of several metabolic 618 619 processes (e.g., purple bacteria), while other conversion pathways involve simultaneously 620 more than one microbial culture (Alloul et al., 2021a; Rasouli et al., 2018; Yang et al., 2019; Zhu et al., 2020). 621

Overall, two main trends are apparent for microbial bioconversion pathways. The first (direct) approach involves direct microbial bioconversion of raw or mildly processed residual biomass, while the second (indirect approach) involves a preliminary sequence of enhancement, cracking and extraction units. These processes convert the feedstock into a form assimilable by the targeted microorganism.

- 627
- 628

#### Fig. 4. Waste-to-nutrition microbial bioconversion pathways

Metabolic pathways are adapted from (Alloul et al., 2021b; Choi et al., 2021; Linder, 2019) and complemented to
 capture the diversity of inventoried waste-to-nutrition microbial bioconversions. Key nutrients such as nitrogen and
 phosphorus are not represented to ensure visual tractability. Chemo(auto)trophic carbon-monoxide-oxidizing
 bacteria pathways are not represented here due to the scarcity of reported information on these.

633

#### 634 **5.4.1. Direct microbial bioconversion pathways**

635 Direct microbial bioconversion pathways are heavily dependent on the characteristics 636 of the feedstock. Accordingly, two main approaches are distinguished, in which the substrate is either under a solid or a liquid (i.e., wastewater) form. The proposed unit operation patternis displayed in SI (figure S4) and corresponding examples are presented in table 2.

639

# 640 **5.4.1.1. Fungiculture**

In the review, SSF processes to generate harvestable nutritional metabolites targeting 641 both TC and MRC ingredients were reported. Regarding TC production, identified pathways 642 mainly focus on enzymes and flavors (Aggelopoulos et al., 2014; Sharma et al., 2020; 643 Teigiserova, 2020) which are not the purpose of the present study. On the other hand, the 644 production of MRC is principally achieved through fungiculture, as further detailed. The 645 fruiting bodies of fungi (i.e., mushrooms) are commonly cultivated on moisturized 646 lignocellulosic materials, meaning that food grade foodstuff (e.g., Pleurotus spp. ranging 15-647 45%DM protein content) is obtained in a single unit operation from non-food feedstock 648 (Bellettini et al., 2019; Chanakya et al., 2015; Ritota and Manzi, 2019). The fungiculture unit 649 operation can be decomposed into two main sub-stages corresponding to mycelium 650 651 colonization of the substrate, followed by fructification (cf., table 2). The spent mushroom substrate displays a reduced lignin content, and is enriched in proteins and lipids (Khan et 652 al., 2015; Wang et al., 2001). Therefore, depending on the initial feedstock, it is possible to 653 use the spent substrate in animal feed regimes (Mhlongo et al., 2021; Wanzenböck et al., 654 2017), mix it with fresh feedstock to generate new SSF substrate (Hamed et al., 2020), or 655 implement subsequent transformation steps. In this case fungiculture acts as a pretreatment 656 for cellulosic sugar recovery for example (Chen et al., 2021). The time period required to 657 convert residual biomass into mushrooms is generally at least three to four weeks when 658 using optimized strains and culture conditions, but this often extends to several months, 659 especially when it is necessary to first compost the feedstock (i.e., a prolonged bioconversion 660 661 process).

662

# 663 5.4.1.2. Wastewater to nutrition

WW has been successfully used as microbial bioconversion medium, mostly using 664 WW from: (i) food (e.g., vegetable oil) and beverage (e.g., brewing) processing 665 (Amenorfenyo et al., 2019; Marchão et al., 2018; Patsios et al., 2020), (ii) animal effluents 666 from farms and slaughterhouses (Li et al., 2019) and (iii) certain non-food industries, such as 667 fertilizer manufacture (Chavan and Mutnuri, 2020) or paper pulp mills (Romantschuk, 1975). 668 669 These WW have in common to host rich organic loads and a low carbon-to-nitrogen ratio (Spalvins et al., 2018; Vethathirri et al., 2021). However, the presence of toxic compounds 670 can vary greatly depending on their specific origin. The services supplied by microbial 671 672 bioconversion on such residual liquid substrate are thus two-fold: (i) production of microbial 673 biomass from WW and (ii) removal of the nutrient charge (i.e., WW treatment) (Muys et al., 2020; Tomlinson, 1976). Mixed-culture mixotrophic microalgae and purple bacteria are 674 particularly adapted to such diluted media (Cao et al., 2020; Capson-Tojo et al., 2020; 675 Solovchenko et al., 2020). They combine atmospheric carbon capture with the use of soluble 676 677 organics and nutrients, reducing the need for additional inputs (Hülsen et al., 2018; Shahid et 678 al., 2020).

The aquaculture market is a prime target for wastewater-to-nutrition pathways, because microorganisms are already part of the fish trophic chain (Glencross et al., 2020; Milhazes-Cunha and Otero, 2017). However, if not deployed *in-situ* (e.g., through activated sludge-derived biofloc technologies) (Alloul et al., 2018; Bossier and Ekasari, 2017), the requisite harvesting and dewatering of low concentration microbial biomass using a series of

energy-intensive extraction steps rapidly becomes cost ineffective. Furthermore, the final 684 market is currently dependent on the quality of the initial WW, because technical 685 specifications (e.g., fecal contamination) drive the requirements of additional intensive 686 prior/post purification treatment steps (Verstraete et al., 2016). In this respect, food-grade 687 applications are mainly limited to high quality food-processing WW (e.g., from starch 688 production, table available limited 689 2), which is only volumes. in

#### 690 Table 2. Selection of inventoried direct microbial bioconversion pathways – reported with wording used by original references

Residual biomass	Transformation units	Results	Potential	Status and references
Wastewater				
wastewater	<ol> <li>Digestion and sterilization with ozone 30min</li> <li>Microalgae <i>Chlorella pyrenoidosa</i> and yeast <i>Rhodotorula glutinis</i> cultured at 28°C for 5-7 days. Addition of glucose and yeast extract, pH7-7.5.</li> <li>Decantation, centrifugation and washing (sodium hydroxide, 47°C)</li> <li>Ultrasonic processing (25min at 47°C), then centrifugated washed and lyophilized</li> </ol>	Recovery of a microbial protein concentrate	Feed protein additive	Lab-pilot-experiment : (Li et al., 2019)
Alcoholic beverage wastewater	1. Direct aerobic submerged fermentation with microalgae (undisclosed strain)	Omega-3 rich biomass	Aquaculture feed	Commercial development: (MiAlgae, 2020)
Fishpond wastewater	1. Direct aerobic submerged fermentation with microalgae (undisclosed strain)	Microalgae rich stream recirculated back	Aquaculture feed	Commercial pilot: (Microterra, 2020)
Fishpond wastewater	<ol> <li>Add C source to equilibrate C:N ratio in wastewater</li> <li>Aerobic heterotrophic bacteria growth in the form of bioflocs in-situ or in dedicated reactor</li> <li>Direct recirculation of bioflocs in fishpond, or pelletizing.</li> </ol>	Microbial protein rich stream recirculated back	Aquaculture feed	Lab experiments (Crab et al., 2012)
nermeste	<ol> <li>Steam treatment to remove SO₂ and sterilization</li> <li>Cooling, aerobic fermentation by <i>Paecilomyces varioti</i> fungi 3-4h, pH4.5-4.7, 38-39°C with addition of NPK.</li> <li>Filtering and washed, dried and ground mechanically</li> </ol>	Recovery of a microbial protein concentrate (55- 60%DM)	Animal feed	Discontinued commercial Pekilo process (Halme et al., 1977) with renewed interests <sup>1</sup>
	<ol> <li>Anaerobic fermentation (formation of fatty acids, sugars and oligosaccharides)</li> <li>Aerobic fermentation with edible strains (undisclosed)</li> <li>Dewatering and drying (various technologies)</li> </ol>	Recovery of protein concentrates (60-80%DM)	Human food and animal feed	Patents: (Logan et al., 2011; Verstraete et al., 2020), commercial developments (Avecom, 2020; iCell Sustainable Nutrition Co., 2020)
digested organic	<ol> <li>Microfiltration (0,2 μm) to concentrate P, coupled with ultra- and nano-filtration to concentrate N</li> <li>Culture of heterotrophic microalgae (here <i>Chlorella vulgaris</i>, 28 days)</li> <li>Harvesting with microfiltration</li> <li>Wet biomass metabolite extraction</li> </ol>	Algal biomass derived bioproducts	To assess	Conceptual formulation by (Stiles et al., 2018) with pilot test (Fernandes et al., 2020)
Solid biomass				
bran busks	<ol> <li>Addition of carbohydrates (e.g corn, seeds) and steam sterilization</li> <li>SSF with fungal strains (<i>Shiitake, Pleurotus sp</i>) in bags for 3-20 weeks until filaments colonized the whole substrate</li> <li>Harvesting of fruiting bodies few days after transfer in culture room</li> </ol>	Recovery of mushrooms (diverse)	Human food	Commercial production: (Biopilz, 2020)
(75%) and straws (25%)	<ol> <li>Composting for 20 days, then pasteurization 5-6 days, at 50-60°C</li> <li>When T°&lt;25°C, SSF with Agaricus bisporus strain for 2-3 weeks (22-25°C, high moisture) until the mycelium develop</li> <li>Mycelium block tapped with soil mixture to keep humidity.</li> <li>Harvesting cycles through a 4-6 weeks period</li> </ol>	Recovery of common white mushrooms	Human food	Commercial production: (Roulleau, 2020)
	1. Homogenization and SSF with <i>Pleurotus</i> : 2 weeks without light, 20-24°C until white foam appears. 2. Position substrate outside air-exposed, high moisture (85-95%) until fruiting bodies reaches 4-7cm diameter.	Home-made mushroom culture	Human food	Commercial production : (La Boîte à Champignons, 2020)

691 <sup>1</sup>: (Eniferbio, 2020); C : Carbon; K : Potassium; N : Nitrogen; P : Phosphorous; SSF : Solid Substrate Fermentation

#### 692 **5.4.2. Indirect microbial bioconversion pathways**

In the field of nutritional services, direct microbial bioconversion is restricted to small 693 number of microorganisms and the use of quality feedstocks (cf., 5.4.1. and 5.1.). The 694 delivery of nutritional services from residual biomass through microorganisms can be 695 extended by adapting the feedstock to the specific requirements of the fermentative microbial 696 697 cultures. The common objective of these indirect bioconversion pathways is the substitution of high-grade nutrients (commonly used to formulate fermentation medium) with lower grade 698 materials. However, the scientific literature mainly focuses on the use of mixed feedstocks 699 700 where only a part of the nutrient requirement is furnished from lower grade materials and completed with quality ingredients. Nonetheless, the conversion pathways included in this 701 702 section all share the feature of at least partially supplying the energy, carbon and nutrients 703 required for microbial bioconversion from residual biomass or C<sub>1</sub> gases.

Indirect microbial bioconversion pathways can be subdivided into two categories. The first group relates to bioconversion of  $C_1$  gases, while the second group targets the elaboration and bioconversion of alternative soluble fermentable compounds ( $C_1$  to  $C_6$ ).

707

#### 708 **5.4.2.1. Gas-to-nutrition conversion pathways**

The gas-to-nutrition pathways employ  $C_1$  gases (mainly  $CH_4$  or  $CO_2$ ) as carbon source for microbial biomass production. The generic unit operation pattern is illustrated in SI (figure S5). The energy source defines the exact nature of the pathway: phototrophic bioconversion fixes carbon dioxide using light (representative examples in SI, table S3), while chemotrophic bioconversion converts high-energy gases such as dihydrogen into biomass (table 3).

Phototrophic pathways are similar to wastewater-to-nutrition pathways (section 715 5.4.1.2.), especially regarding post-treatment. However, phototrophic pathways usually focus 716 on the capture of concentrated CO<sub>2</sub> streams (figure 4). Accordingly, phototrophic organisms 717 (mainly microalgae and cyanobacteria) are operated in reactors optimized for light 718 penetration and gas transfer and fed with essential nutrients (e.g., N, P, K, etc.) in order to 719 produce proteins and omega-3 fatty acids. According to the literature, these nutrients are 720 721 often sourced from high grade products (e.g., glutamate, peptone, fertilizers) instead of being upcycled from residual biomass streams. However, in addition to the use of waste C<sub>1</sub> gases, 722 some gas-to-nutrition pathways also harness nutrients from underused streams (e.g., 723 724 predominant N leakages such as animal and human effluents) to satisfy the nutritional requirements of the microorganism (Matassa et al., 2015; Yang et al., 2021). The versatility 725 726 of bacterial strains, which can use different forms of nitrogen such as  $NH_3$ ,  $NH_4^+$  (Dou et al., 2019) and N<sub>2</sub> (Hu et al., 2020; Pfluger et al., 2011) provide the basis for a whole range of 727 728 pathway variants.

729 Chemotrophic pathways mainly rely on either hydrogen-oxidizing bacteria (HOB) or methane-oxidizing bacteria (MOB), both of which use gases as their carbon and energy 730 sources and hence provide the basis for "full-gas" pathways (Matassa et al., 2020). 731 Importantly, chemotrophic processes are characterized by high conversion efficiencies 732 733 (Claassens et al., 2016; Liu et al., 2016; Pander et al., 2020) in large part because, unlike phototrophic processes, they are not limited by energy availability. Additionally, chemotrophic 734 bacteria can fix between 80% and 100% of their N supply (Pikaar et al., 2017) into microbial 735 biomass, which yields a protein content of approximately 70%-80% and an amino acids 736 737 profile analogous to that of fish meal (Matassa et al., 2015; Øverland et al., 2010). For these 738 reasons, HOB and MOB strains have been arousing interest for some time, with their

proteins being studied for human nutrition, especially in the context of space travel (Alvarado 739 740 et al., 2021; Foster and Litchfield, 1964; Steinberg et al., 2017). The suitability of MOB meals as a substitute for conventional high-protein ingredients in monogastric animal diets has 741 been demonstrated (Øverland et al., 2011, 2010) and these are already close to 742 commercialization unlike HOB routes. However, one cause for concern is the significant 743 presence of anti-nutritional RNA/DNA and endotoxins in bacterial biomass. Therefore, if not 744 mitigated through genetic engineering, this often must be eliminated before the microbial 745 746 biomass is considered fit for consumption. Nucleic acids removal can be achieved through 747 heat treatments, as already performed during the industrial production of F. venenatum (RNA 748 content reduced from 8-9% down to 1% in weight) (Whittaker et al., 2020).

749 The origin of  $C_1$  gases is key to the economy of chemotrophic pathways (García Martínez et al., 2021; Huizing, 2005; Verbeeck et al., 2020). While scaling projects currently 750 focus on the use of fossil-based methane and electrolysis-based hydrogen, syngas and 751 752 biogas are also investigated (cf., table 3). The inherent variability and heterogeneity of these biogenic gases are attenuated through scrubbing, removing undesirable gas components, 753 such as carbon monoxide and hydrogen sulfide (Tsapekos et al., 2019; Xu et al., 2020). 754 755 Accordingly, gases for chemotrophic pathways can be derived from residual biomass through 756 a sequence of cracking and/or bioconversion units (e.g., anaerobic digestion, gasification) followed by refining units. 757

The low solubility of H<sub>2</sub> (De Vrieze et al., 2020) and the safety issues related to the 758 simultaneous presence of H<sub>2</sub> and O<sub>2</sub> for aerobic HOB production (Molitor et al., 2019) are 759 also questions that are addressed by current research work. Often, to meet these 760 761 challenges, additional processes are envisaged for chemotrophic pathways (Sakarika et al., 2020). For example, gases are first converted into organic compounds (e.g., acetic acid or 762 methanol) using either biocatalysis (e.g., fermentation using an acetogenic bacteria) or 763 764 physico-chemistry (e.g., hydrogenation) (Linder, 2019; Mishra et al., 2020). The resulting organic compound is then used as substrate to support the growth of a common 765 heterotrophic organisms such as yeast. Alternatively, the overall process can be achieved in 766 767 a single step using the co-culture of several microorganisms (Du et al., 2020). An emerging route is the use of volatile fatty acids (VFAs) as platform intermediates instead of end-gases. 768 This novel approach, described in Alloul et al. (2018) targets acidogenic fermentation on 769 dissolved carbon sources (obtained through prior cracking or conversion of residual biomass) 770 to recover VFAs. These are further converted into edible biomass through flexible 771 772 microorganisms such as purple bacteria (Capson-Tojo et al., 2020; Lu et al., 2021) or 773 filamentous fungi (Uwineza et al., 2021).

774

#### 775 **5.4.2.2.** Alternatives to common microbial bioconversion medium

776 The previous section describes processes that require the use of novel substrate-777 microbe associations regarding nutrition. However, axenic microbial bioconversion is already 778 a core technology of the feed and food industries, being widely used to produce amino acids, lipids or mycoproteins from well-defined substrates such as raw glucose or methanol. 779 Nevertheless, with increasing pressure to deliver cost-competitive carbohydrates in a 780 781 framework of environmental sustainability and food/feed security, analysis of the literature 782 reveals that considerable focus is put on the investigation of alternative carbon sources (Siben et al., 2018; Specht, 2020; Specht and Crosser, 2020). Accordingly, this section 783 provides a description of attempts to drive low-cost residual biomass into common microbial 784 bioconversion pathways, using a series of conversion processes to ensure that nutrient and 785

end-product quality and safety are maintained, as represented in figure 5. A selection ofexamples from the literature is available in SI, table S4.

The use of sugar- and lipid-rich streams has been previously reviewed (Spalvins et al., 2018). These pathways rely on quite homogeneous agrifood co-products, such as whey or molasses (Caporusso et al., 2021), and are used to upgrade their current animal feed value to food quality TC and MRC ingredients production. Considering the quantities available, certain biofuel co-products (e.g., glycerol) are also included because these can be used, for example, to implement *in-situ* microbial protein production (Fazenda et al., 2017; Tracy et al., 2020).

795 Cracking and/or extraction processes have been developed to breakdown structural complexity and deal with feedstock heterogeneity characteristic of (for example) urban food 796 797 waste. Common cracking operations, such as hydrolysis and saccharification, are deployed 798 either in stand-alone single product processes (Kwan et al., 2018; Pleissner et al., 2014) or in 799 multi-production platforms to solubilize compounds that serve as nutritional feedstock for 800 microbial bioconversion. In multi-production platforms, co-products (e.g., arising from protein extraction pathways) such as supernatants and residual fibers are used as sources of sugars 801 802 to sustain bioconversion (Øverland et al., 2019; Thomsen et al., 2008). Lignocellulosic biomass is also a source of fermentable sugars (cf., section 5.2.1.) and can sustain for 803 804 example straw- or wood-to-protein pathways (Upcraft et al., 2021). This type of strategy has already been implemented at industrial scale (e.g., Tornesch plant producing 20,000 tons of 805 yeast per year in the 1930's) in response to wartime (Harris, 1951). However, renewed 806 807 interest in this route has been prompted by recent advances in selective and food-grade 808 lignocellulose cracking (Asim et al., 2021; Tenlep, 2020; Voutilainen et al., 2021).

809 Finally, synthetic pathways to produce fermentable TC are reported in the literature. Also called power-to-food or power-to-protein (Mishra et al., 2020; Sillman et al., 2020), 810 811 these approaches use electricity to produce the reducing power further converted by a target microorganism. A first application is to extend the possibilities of gas-to-nutrition pathways, 812 for example through the "CO<sub>2</sub>-to-CH<sub>4</sub>-to-protein" route (Xu, 2021) or converting hydrogen into 813 814 methanol and acetic acid as mentioned in section 5.4.2.1.. Similarly, synthetic pathways are also engineered to fix CO<sub>2</sub> through "biological-inorganic" (Nangle et al., 2017) or "microbial 815 electrosynthesis" (Dessì et al., 2020) processes into non-gaseous fermentable TC such as 816 817 formate or methanol (Mishra et al., 2020; Sakarika et al., 2020).

818

819 820

# Fig. 5. Producing alternative fermentation mediums from residual biomass: unit operations pattern

821 The indicative ranking of residual biomass families in the nutritional quality scale is derived from figure 2., and 822 allows to visualize the estimated chain of unit processes required to bridge the gap between the initial 823 composition-structure of a feedstock and the composition-structure which is adequate to deliver a nutritional 824 service. Icons are as defined in figure 1. The identified conversion pathways are rather straightforward when 825 starting from sugar- or lipid-rich residual biomass, but can be more complex, involving prior cracking and 826 extraction operations to release fermentable compounds. Albeit limited by economic considerations (Kwan et al., 827 2019), purification technologies are often required to detoxify feedstocks and bring them up to nutritional 828 specifications. C1 gases can either have a fossil or biogenic origin, as represented in figure S5.

#### 829 Table 3: Inventoried examples of biomass recovery into food and feed through chemotrophic gas-to-proteins pathways – reported with wording used by original references

Residual stream targeted	Technological requirements for the carbon source	Technological requirements for the nitrogen source	Bioconversion conditions (incl. energy source)	Results (incl. potential)	Status and references
		NH <sub>3</sub>	<ol> <li>Renewable electricity allows DAC, Haber-Bosch process, and water electrolysis</li> <li>Submerged fermentation of <i>Curpiavidus necator</i> HOB bacterial strain</li> <li>Centrifugation and evaporation</li> </ol>	Protein-rich biomass for humans and animals	Concept: (Givirovskiy et al., 2019; Sillman et al., 2019), lab-pilot-experiment by (Solar Foods, 2020) with patent (Pitkänen, 2020)
Industrial combustion flue gases (e.g. power plant)	Cooling and direct pumping of flue gases in the bioreactor if reduced impurities		1. Electrolysis to supply $H_2$ to the bioreactor 2. Submerged anaerobic fermentation of HOB strain 3. Centrifuging and drying.	Aquaculture feed and human food	Commercial development: (Deep Branch Biotechnology, 2020; Kiverdi, Inc., 2020; NovoNutrients, 2020), patent: (Reed, 2019)
digested sewage	separate $CO_2$ (here carbon	direct numering into hisroceter	<ol> <li>Electrolysis or CH<sub>4</sub> reforming to produce H<sub>2</sub></li> <li>Submerged fermentation of HOB strain with recovered CO<sub>2</sub></li> <li>Transformation to edible products</li> </ol>	Protein-rich ingredients and prebiotics	Concept: (Matassa et al., 2016), Demo-pilot-plant: (Power-to-Protein, 2020)
Undiluted source- separated urine	electrochemical cell (EC)	<ol> <li>Autohydrolysis of urine (28°C)</li> <li>Hydrolysate supplied to the EC: recovery of NH<sub>3</sub> at the cathode.</li> <li>NH<sub>3</sub> recovered through air stripping and absorption</li> </ol>	1. $H_2$ and $O_2$ are produced in the EC (respectively cathode and anode) 2. All gases are supplied to a bubble column reactor for HOB production	Microbial protein production (no specified market)	Lab-experiment: (Christiaens et al., 2017)
Fecal WW			Axenic submerged fermentation of edible MOB and microalgae (aerobic) with recovered gases and nutrients	Microbial protein (no specified market)	Concept: (Verstraete et al., 2016)
biowaste,	Pyrolysis/Gasification to produce syngas with proportions of CO and CO <sub>2</sub>		Submerged fermentation of edible HOB or CO- oxidizing bacteria (aerobic)	Microbial protein (no specified market)	Concept: (Matassa et al., 2020)
Human solid and liquid waste	<ol> <li>Homogenezation</li> <li>Fixed-film, flow-through anaerobic digester</li> <li>Inorganic removal, and remaining effluents treated with strong base to remove CO<sub>2</sub></li> </ol>	•		MOB proteins: astronaut's foodstuff and denitrifying biomass as feed	Lab-experiment: (Steinberg et al., 2017)
Urban blowaste		<ol> <li>Centrifugation and filtration (0,2 μm) of liquid digestate</li> <li>Pasteurization (70°C 1h) and dilution</li> </ol>	Submerged fermentation with a mixed-MOB culture of Methylophilus sp, or Methylococcales and Methylophilales	Microbial protein (presumably for animal feed markets)	Proof-of-concept experiences: (Khoshnevisan et al., 2019; Tsapekos et al., 2019)
manure	1. Anaerobic digestion 2. Biogas upgraded with EC: resulting cathode off-gases are $CH_4$ , $O_2$ and $H_2$ , anode off- gases are $O_2$ , $CO_2$ , $H_2$	l uract use at row materials		Microbial protein (presumably for animal feed markets)	Proof-of-concept experiment: (Acosta et al., 2020)

(urine, sewage	CO <sub>2</sub> concentrated from flue gases, gasification exhaust or biogas upgrading	Not detailed	2 Submerged termentation of Clostrigium III Ingganiii	(presumably for human food	Proof-of-concept-experiment: (Molitor et al., 2019), concept: (Mishra et al., 2020)
----------------	--	--------------	---	----------------------------	---

<sup>1</sup>: As in Khoshnevisan et al. (2018), includes pulper, separator and dewatering; COD: Chemical Oxygen Demand; DAC: Direct Air Capture; EC: Electrochemical cell; HOB: Hydrogen-Oxidizing Bacteria; MOB: Methane-Oxidizing Bacteria; WW: Wastewater

831

#### 832 **5.5. Farmed animal bioconversion pathways**

A wide range of residual biomass has already been tested as direct animal feed, ranging from fruit wastes (Wadhwa and Bakshi, 2013) to manure (Mueller, 1980). Not only limited to circumstances of extreme necessity (De Groot and Bogdanski, 2013; Makkar et al., 2018), the wide adoption of residual biomass to sustain animal husbandry has been explored in recent literature (te Pas et al., 2021). Animal farming, or rather the farmed animal themselves can then be considered as a bioconversion process in waste-to-food pathways.

839

#### 840 **5.5.1. Waste-to-meat, milk and eggs**

Two recent studies revealed that supplying all food waste, agricultural residues and grasslands available in Europe to a combination of swine, laying hens and dairy cattle could provide 9 to 31g of proteins per person per day for the continent (Van Hal et al., 2019; Van Zanten et al., 2018). This is significant if one considers that the daily recommended protein intake for an adult is 50-60g (EFSA Panel on Dietetic Products, Nutrition and Allergies (NDA), 2012).

Provided that the use of residues and co-products does not affect performance and 847 food safety, and that it does not require intensive pretreatments, the use of livestock as direct 848 bioconverters is a viable option and indeed is already widely implemented (Mottet et al., 849 2017; Wilfart et al., 2019). Residual feedstocks still need to meet feed standards, and 850 therefore mainly consist of agrifood industry co-products, which on average represent 10% 851 and 20% of compound feeds used in France for monogastric and ruminants respectively 852 (Agreste, 2019, 2017). Monogastric animals are restricted to low-lignin feedstocks 853 (Chapoutot et al., 2019), but ruminants can also convert green biomass and pretreated crop 854 855 residues. Direct bioconversion of slaughterhouse by-products is also reported, especially for aquaculture in some regions, but in others this is considered unacceptable (Leduc, 2018; 856 Meeker, 2006). Besides direct enhancement techniques such as grass ensiling (cf., section 857 858 5.1.), a common strategy involves the inclusion of specific enzymes in diets. These degrade anti-nutritional compounds (e.g., mycotoxins) and increase the digestibility of fibrous 859 components (Nunes, 2018). 860

861 Less homogeneous residual biomass, such as food waste, are already used in livestock diets, not only in small-holder farming or periurban systems (Cesaro et al., 2019), 862 but also in industrial production after enhancement-like pretreatments (e.g., heat-treated 863 urban food waste in Eastern-Asia) (Dou et al., 2018; Georganas et al., 2020; Zu Ermgassen 864 et al., 2016). Such practices are banned in other parts of the world, mainly because of TSE-865 related risks (Castrica et al., 2018). An option to ensure safety and acceptance lies in only 866 867 considering pre-consumer food losses (e.g., surplus vegetables) (Luyckx et al., 2019; Pinotti 868 et al., 2021; San Martin et al., 2016). In addition to animal health concerns, livestock industries are guite risk-adverse and take precautions to preserve meat/milk/egg guality 869 (Research and Innovation, 2017; Salami et al., 2019). An example of the risk/benefit analysis 870 of novel feed is the inclusion of some plant-based residues in animal diets which would 871 872 positively furnish functionally valuable phytochemicals (e.g., antibiotic substitute) but will 873 negatively alter organoleptic properties of meat (Achilonu et al., 2018; Valenzuela-Grijalva et 874 al., 2017). Further issues also include human health problems arising from imbalances in 875 animal diets such as the omega-6/omega-3 fatty acids ratio correlated with the prevalence of 876 chronic diseases in humans (Duru and Magrini, 2017).

The main challenge of using livestock as residual biomass bioconverters is the fact that the current key determinant of stakeholder practices and choices is performance, mainly

measured as a feed-to-food ratio (Shepon et al., 2016; Wirsenius et al., 2010). Including 879 biomass of lower nutritional value into diets is detrimental to livestock's bioconversion 880 efficiency (details and benchmark in SI), and less-productive animals, better adapted for the 881 direct digestion of residual biomass, are not included in current farming strategies (Van 882 Zanten et al., 2019). Therefore, the extension of livestock bioconversion to a wider range of 883 residual biomass will involve reconsidering the choice of animal breeds and/or the 884 development of strategies to deal with the anti-nutritional and fibrous components of low-885 grade feed ingredients (Peyraud et al., 2020; te Pas et al., 2021). For the latter, this can be 886 dealt with using different waste-to-feed pathways yielding MPF, MCR and TC ingredients, as 887 those inventoried through the past sections. Regarding human nutrition, the consumption of 888 889 animal-based food (i.e., meat, milk, etc.) involves the addition of a livestock bioconversion 890 building block at the end of the biomass conversion pathway. In this respect, in the representation scheme proposed in this work, the slaughterhouse should be considered as a 891 combination of enhancement and extraction blocks that intervene before human 892 893 consumption.

894

#### 895 **5.5.2. Insect bioconversion pathways**

896 The use of insects to convert residual biomass into nutritional ingredients for both 897 animals and humans has been extensively reviewed in the literature (FAO, 2020; Rumokov et al., 2019; Van Huis, 2020). Among the numerous known species, focus is given herein to 898 those that are the subject of commercial projects for nutrition in Europe, especially Hermetia 899 Illucens (black soldier fly), Musca domestica (common housefly) and Tenebrio molitor (yellow 900 901 mealworm) (Cadinu et al., 2020). Depending on their diets and stage of life (e.g., larvae), these insects accumulate proteins and transform sugars into lipids (Colonna, 2020) to 902 903 achieve contents in the range 40-70%DM and 10-40%DM respectively (Makkar et al., 2014).

The "waste-to-protein" insect pathway (Zurbrugg et al., 2018) follows a pattern that is 904 905 almost identical to that of livestock bioconversion (illustrated in SI figure S6, S7 and table S5). First, common enhancement processes such as grinding, mixing, moisturizing are used 906 907 to improve nutrient uptake by the insects. The intensity of such pretreatment depend on insects specific characteristics, flies requiring an almost liquefied substrate while termites 908 directly degrade lignocellulose (Hubert and Berezina, 2020). The bioconversion itself takes 909 910 place in either fully automated or low-tech facilities, depending on the context (Cortes Ortiz et al., 2016; Dortmans et al., 2017; Kröncke et al., 2020; Melgar-Lalanne et al., 2019). Once the 911 insects reach maturity (i.e., bioconversion culminates), they are harvested, sanitized and 912 913 usually dried when not directly fed fresh. The final MRC is either the whole insect or a fraction of it (Pippinato et al., 2020). Fractionation often targets the generation of a protein-914 rich meal and thus involves lipid extraction (i.e., generation of insect oil). Once missing 915 916 nutrients (e.g., methionine) have been added, insect meals are good candidates to substitute sova meal or fish meal (Azagoh et al., 2016; Pleissner and Smetana, 2020) in compound 917 feed formulations. Initially only permitted for use in aquaculture and petfood, the inclusion of 918 insect protein meal in animal diets has been recently extended to swine and poultry in 919 Europe (IPIFF, 2021). Regarding waste-to-nutrition conversion pathways, the inclusion of 920 921 insect-based products in livestock feed corresponds to a successive sequence of bioconversion (i.e., insect followed by livestock). 922

Additionally, insects (or corresponding meal) are also ground and used as protein-rich ingredients for ready-to-eat products for humans (e.g., snacks) through common enhancement processes (García-Segovia et al., 2020; Lamsal et al., 2019), this strategy being preferential for many populations that are so far reluctant to consume whole insects

(Orsi et al., 2019; Poortvliet et al., 2019). Like other common protein-rich ingredients, insect 927 proteins can further enter a series of extraction units to refine and isolate precise 928 functionalities (Gravel and Doyen, 2020; Smetana et al., 2018). The main co-product, insect 929 oil, also has the potential to directly enter food and feed markets (Phan Van PhI et al., 2020; 930 Smetana et al., 2020). However, nutritional composition of the insects was found to be 931 variable, being correlated to diet quality (Gold et al., 2020; Oonincx et al., 2015), and 932 933 relatively few background data are available regarding biological and chemical risks 934 associated with such residual substrates (ANSES, 2015; Bessa et al., 2020; EFSA Scientific Committee, 2015). Consequently, European authorities have so far only authorized 935 commercial insect farming fed with "feed grade materials", thus creating potential competition 936 with conventional animal feeds (Gasco et al., 2020). Entomofarming facilities currently 937 938 operating in Europe mainly use industrial agrifood co-products (e.g. bran, peels) or homogeneous surplus food (e.g. unprocessed expired food) (IPIFF, 2019). Mainly intended 939 940 for feed markets, the latest legislation recognizes Tenebrio Molitor larvae, Locusta Migratoria and whole house crickets Acheta Domesticus as the first safe insect foodstuffs in Europe, 941 under the condition that these are fed with feed-grade materials (EFSA Panel on Nutrition, 942 Novel Foods and Food Allergens (NDA) et al., 2021c, 2021a, 2021b). Of particular interest 943 for stakeholders is the expansion of the authorized rearing substrates for insects and the 944 945 wider development of substrate-enhancement techniques (Raksasat et al., 2020). Food waste is particularly mentioned in the literature because this represents significant volumes 946 besides its nutritional potential (Jensen et al., 2021; Ojha et al., 2020; Varelas, 2019). 947

948

#### 949 6. A comprehensive overview of waste-to-nutrition pathways

#### 950 6.1. Building block framework: implications and limits

Overall, waste-to-nutrition pathways involve diverse technologies and different degree 951 of nutrients and energy circularity, using different schemes of unit operations of increasing 952 intensity and complexity. In general, the less edible the feedstock (as defined in section 3), 953 the more processing is required to derive an edible foodstuff from it, as it can be visualized in 954 figures 5 and S1-S7. Bioconversion can sometimes be used to circumvent this, providing 955 foodstuff in a reduced number of steps (e.g. figures S4, S6 and S7). This is because living 956 organisms can be considered as complex reactors performing a series of unit operations 957 958 (Godon et al., 2013). However, in the case of highly structured and chemically complex plantbased feedstock, most organisms display only limited direct conversion capabilities. As a 959 result, a pretreatment sequence to render the feedstock suitable for subsequent 960 961 bioconversion is often needed (e.g. figures 5 and S7). One notable exception to this rule are fungi, as these secrete complex arsenals of biomass-degrading enzymes (Kuyper et al., 962 2021; Souza Filho, 2018). Mushrooms cultivated on wood residues (figure S4) exemplify 963 964 such shortened bioconversion pathways that bridge the gap between an inedible feedstock 965 and edible nutrient production.

966 Often, the production of edible nutrients is the result of sequential bioconversions. In this case, animal-animal bioconversions are rather inefficient (e.g., 4-5 pelagic fishes 967 bioconverted in 1 salmon) (Tacon and Metian, 2008), while microorganism-animal (e.g., 968 969 microbial proteins to animals) or microorganism-microorganism (e.g., first strain freeing a TC 970 in turn processed by a second strain of interest as in figure 5) sequences are more efficient. 971 This is because the inherent losses associated with bioconversion may to some extent be compensated by two aspects. First, microorganisms are able to convert non-carbohydrates 972 energy (including light) into carbohydrates and convert non-protein nitrogen into proteins. 973 974 Particularly, some bacteria and microalgae report nitrogen fixation ratio higher than 70%

975 (details in SI). Second, animals are able to autonomously capture and concentrate diluted nutrients. For example, some insects efficiently concentrate residual biomass proteins (<50-976 60% recovery efficiency, see SI) and are themselves easily harvestable. As a second 977 example, farming fishes within microalgae production systems is an indirect way of collecting 978 microalgae biomass (as fish) otherwise diluted in production pond/reactor (Verstraete et al., 979 2016). In this case, conventional energy-intensive microalgae harvesting and drying 980 techniques are replaced by fishing techniques, yet at the expense of a significantly lower 981 982 overall nutritional output yield. It is also important to point out the ability of some edible microorganisms to grow in biomats at the liquid-air interface, removing the need of energy-983 intensive extraction stage (Kozubal et al., 2020). 984

985 Besides initial feedstock composition-structure considerations, the target market also strongly defines the intensity of conversion process strategies. Indeed, most conversion 986 987 pathways involve extensive post-treatment (extraction and enhancement of nutritional/organoleptic properties) to polish the final product's functionalities (e.g. 988 microalgae-based meat-analogs) (Bernaerts et al., 2019; Fu et al., 2021). Currently, these 989 steps are mostly implemented to concentrate/purify nutrients (up to TC) and attenuate anti-990 991 nutritional and safety issues.

As the effects and drawbacks of each unit operation are cumulative, it is still unclear 992 to what extent the foreseen advantages associated with waste-to-nutrition strategies would 993 offset their drawbacks if massively implemented (Guthman and Biltekoff, 2020; Helliwell and 994 Burton, 2021; Van Eenennaam and Werth, 2021). The building blocks framework proposed 995 996 herein is intended to simplify the representation of processes involved in waste-to-nutrition 997 pathways, systematizing their transformation sequence patterns and grouping these into 998 large categories. Accordingly, this framework is a stepping stone to further assessments, providing a convenient way to visualize available options when comparing different 999 1000 feedstocks. However, as key aspects such as energy demand or greenhouse gas emissions are technology- and operation-specific, further state-of-the-art life cycle assessments (LCA) 1001 accounting for all inputs, outputs including wastes and co-products generated during a 1002 1003 process are required to quantitatively estimate their full environmental performance. Accounting for services provided by co-products in waste-to-nutrition pathways is key, 1004 1005 because these are potential sources of energy, chemicals, materials and even feed. When 1006 feedstock give rise to multiple pathways and multiple products and services it is appropriate to use the term biorefinery. 1007

1008

#### 1009 **6.2. Waste-to-nutrition pathways into eight large families**

Earlier (cf., section 5) waste-to-nutrition pathways were grouped based on their core building block. To refine the analysis, the classification is here extended by grouping pathways into large families on the basis of belonging to the same biorefinery scheme or addressing the same challenge (figure 6). These families are not defined based on the use of specific feedstocks, nor do they deliver specific services. Moreover, livestock-based bioconversion was not considered as a waste-to-nutrition family, being rather a plug-in often present at the end of the other families.

Value chains built on the extraction of rubisco, or the cracking of plant fibers are wellstudied, constituting respectively the green and lignocellulosic biorefinery families. Similarly, insect farming is increasingly studied from the angle of biorefining, because use of the coproducts will form part of the business model (da Costa Rocha et al., 2021; Hubert and Berezina, 2020; Ravi et al., 2020), hence the insect biorefinery proposed herein. Likewise,

we propose a "gas-intermediate biorefinery" family, as gas-based proteins can be part of 1022 larger anaerobic digestion and gasification platforms (Matassa et al., 2020), but also because 1023 it positions C<sub>1</sub> gases as key basic bricks for nutrition. Besides proteins and fats, 1024 1025 slaughterhouse by-products already constitute a source of ingredients for cosmetics and medical products. However, a "more-out-of-slaughterhouse by-products" family is defined 1026 with the aim to convey the idea of valorizing such hitherto underused streams (cf., 5.2.2.). 1027 The remaining categories are not related to specific value chains, but are rather challenge-1028 1029 oriented. These include the development of (i) non-soluble protein extraction from diverse 1030 residual biomass feedstocks, (ii) solid substrate fermentation for nutrition and (iii) alternative liquid substrates for fermentation. Importantly, the eight families proposed herein can interact 1031 1032 and are likely complementary to achieve a better use of resources from a circular economy 1033 standpoint. For example, cellulosic sugars, or organics remaining in the liquid fraction of anaerobic digestion, can be used as alternative fermentation substrates. Furthermore, 1034 1035 because the families are not defined by specific technologies, they should be amenable to future technology innovations, for example biotechnology-based processes. 1036

1037

#### 1038 **Fig. 6. Identified waste-to-nutrition conversion pathway categories and current status** 1039 <sup>\*</sup> Technology Readiness Level (1-9): based on information available to date (SI).

1039

<sup>\*</sup> Technology Readiness Level (1-9): based on information available to date (SI). <sup>\*\*</sup> Valid for all pathways involving microorganisms.

1041

# 1042 **7. Criteria for waste-to-nutrition pathways assessments**

Obviously, the waste-to-nutrition pathways reviewed herein do not hold the same potential to supply novel food and feed resources and further studies are required to ascertain their true relevance and feasibility. To perform a quantitative prospective assessment, common criteria are required.

First, a clear hierarchy of residual biomass valorization must be established and respected. From a sustainability standpoint it is unacceptable to use avoidable wastes and residues as feedstock, especially when these are directly derived from food (Leipold et al., 2021; Mourad, 2016; Redlingshöfer et al., 2020). Similarly, residual biomass prioritization is essential to prevent the risk of rebound effects (i.e. encouraging additional waste and residues generation to ensure feedstock availability for biorefineries) (Teigiserova et al., 2020).

1054 A second criterion must consider potential drawbacks and disruptions of diverting biomass streams from their current value chain (Abel and Blanc, 2017; Hedegaard et al., 1055 1056 2008). In this respect, a sensitive point is related to biomass that already sustain the food production system either directly (e.g. co-products as animal feed) or indirectly through 1057 agronomic valorization (e.g., composting, ploughing, fertilizing). The latter, not dealt with in 1058 1059 this review, echoes the numerous incentives to close nutrient cycles through the promotion of organic fertilizers, among others those widening the scope of streams able to enter 1060 1061 agronomic valorization pathways (European Union, 2019). Examples of their uses in urban farming are increasingly put forward (Billen et al., 2021; Stoknes et al., 2016; Van Zanten et 1062 al., 2019). Therefore, not only is it necessary to carefully compare waste-to-nutrition 1063 pathways between themselves, but also with conventional scenarios, also referred to as 1064 counterfactuals in LCA involving constrained resources (Pehme et al., 2017). Another trade-1065 off of waste-to-nutrition pathways is the burden shifting between resource efficiency and 1066 utilities requirements. Indeed, previous analyses revealed that, depending on the scenario, 1067 novel ingredients do not necessarily always perform better (environmentally) than 1068

1069 conventional ones, mainly because of higher requirements in energy, water, chemical and 1070 material (Bohnes and Laurent, 2021; de Boer et al., 2014; Sillman et al., 2020; Smetana et 1071 al., 2015; Spiller et al., 2020).

A third criterion must capture the spatio-temporal and context-dependent dimension 1072 (Dries et al., 2020). The demand for alternative practices that reduce pressure on arable land 1073 use is urgent and cannot be delayed for an indefinite period. However, different waste-to-1074 nutrition pathways are characterized by different technology readiness and feasibility levels 1075 1076 (Tuomisto, 2019). Some technologies are immature, while other are perhaps quite mature, but face major regulatory hurdles, stakeholder risk aversion or consumer rejection (Cameron 1077 et al., 2019; Specht et al., 2019; Tubb and Seba, 2019). In this regard, while synthetic 1078 1079 "cultured" animal products (e.g. cultured meat) are still far from widespread commercialization (Post et al., 2020; Zhang et al., 2020), rapidly evolving socio-cultural 1080 considerations are likely to push innovation and accelerate development in the area (Crosser 1081 1082 et al., 2019). Similarly, novel waste-to-nutrition pathways will not be deployable everywhere in the same way, either because of local availability of feedstock (see below) and technical 1083 skills, and/or because of socio-cultural trends. It is generally recognized that neophobia, 1084 cultural values and disgust are the major barriers for the widespread consumer acceptance 1085 of novel food (Fischer and Van Loo, 2021; Siegrist and Hartmann, 2020; Tuorila and 1086 Hartmann, 2020). These barriers make market uptake projections quite challenging, 1087 particularly when genetic engineering is involved (Boccia et al., 2018; Lähteenmäki-Uutela et 1088 al., 2021). Therefore, the challenge is to motivate change in people's perception so that they 1089 can progressively accept such novel ingredients at their tables. For example, the inclusion of 1090 1091 bacterial meal in astronauts' diets could trigger a wider acceptance of microbial-based food (Verstraete et al., 2016). Similarly, many initiatives investigate the organoleptic 1092 1093 enhancements of novel food such as plant-based proteins and upcycled agrifood co-products (cf. table 1 and SI database), aiming to increase their attractiveness. Safety considerations 1094 1095 (see below) will also condition market uptake, especially in the case of ingredients stemming 1096 from feedstocks currently covered by waste regulations. About 75% of the pathways reviewed in this study target inclusion of novel ingredients in animal diets, suggesting a 1097 1098 foreseen trend of waste-to-nutrition approaches development preferentially towards feed 1099 rather than food markets.

1100 Another spatially related criterion concerns the potential impact and development pattern of a given waste-to-nutrition pathway. The feasibility of a pathway is intrinsically 1101 linked to the availability and processability of an appropriate residual feedstock. Many 1102 biomass feedstocks are characterized by high moisture content, which limits transport and 1103 increases perishability (e.g., sugar beet leaves). For these reasons, it is often preferable to 1104 process biomass close to the site of production. In the case of industrial streams, it might be 1105 even preferable to process them onsite. When waste-to-nutrition pathways are implemented 1106 1107 in stand-alone facilities, it is necessary to carefully balance biomass availability and storage 1108 constraints with considerations related to the economy of scale, either opting for small-scale decentralized facilities or large-scale centralized ones (Maity, 2015). In this regard, although 1109 technology considerations partially dictate which option is most appropriate, this is not 1110 always the case. For example, insect biorefineries are economically feasible at both scale 1111 (Chia et al., 2019; Kröncke et al., 2020). Moreover, biotechnology-based processes also offer 1112 1113 scope for downscaling and decentralization, with microbial bioconversion being feasible in transportable containers (Kernel.bio, 2020) and even domestic scales (Shojinmeat Project, 1114 2020). Therefore, it is crucial to understand the ways in which waste-to-nutrition pathways 1115 1116 are likely to be deployed, because evidence-based future narratives and scenarios on food transition are essentials to the proper context-dependent prospective comparison of different 1117 pathways (Antonsen and McGowan, 2021). Finally, one important future narrative to consider 1118

is the degree to which meat will still be produced and will coexist with its alternatives (plantbased, microbe-based). Change in meat production will have implications both on the feedstock side of waste-to-nutrition pathways, but also on the market side, eventually reducing the relevance of producing feed ingredients.

The final criterion relates to safety and the way this affects the substitution potential of 1123 1124 novel nutritional services. Safety is a complex issue because it covers potential and real risks and needs to be addressed using technical and regulatory means. The application of the 1125 principle of precaution means that the introduction of any novel nutritional services will be 1126 confronted by strict institutional barriers (Lähteenmäki-Uutela et al., 2021; Tzachor et al., 1127 2021). Overcoming these requires the compilation of large amounts of background data on 1128 risk assessment, so the "Generally recognized as safe" or "Novel food" labels can be granted 1129 (EFSA Panel on Dietetic Products, Nutrition and Allergies (NDA), 2016). In cases where the 1130 risk is patent (e.g., use of sludge or other feedstocks under the Waste Regulation), the 1131 hurdles will be higher and advanced technical solutions (e.g., sequence of cracking, refining 1132 and conversion units) will likely be required to attenuate the presence of noxious substances, 1133 viruses and microorganisms (Alloul et al., 2018; Verstraete et al., 2016). Therefore, safety-1134 1135 related barriers to implementation of novel waste-to-nutrition pathways must be assessed to properly compare these, establishing the plausible timeline to implementation and the likely 1136 technology cost involved. Additionally, because safety often involve purity and intense 1137 refining, it is necessary to assess a collateral risk, which is that of the market product being 1138 part of a cracking-building food chemistry pattern, likely leading to ultra-processed food 1139 ingredients (Fardet, 2018; van der Goot et al., 2016). 1140

1141

#### 1142 8. Conclusion

To enhance food system resilience and limit its environmental impacts, pathways 1143 transforming residual biomasses and C<sub>1</sub> gases into food and feed ingredients are gaining 1144 increasing attention. As a first step to assess their potential, this study classifies the main 1145 waste-to-nutrition pathways through the review of 950 literature records. The analysis reveals 1146 1147 that most nutritional services can be provided through pathways built on different residual feedstocks, and reversely, one residual biomass can lead to a variety of nutritional outputs. 1148 Identified waste-to-nutrition pathways employ a sequence of unit operations workflow to 1149 1150 adapt the initial composition-structure (i.e. nutritional, anti-nutritional and structural contents triangle) of the input to the end-market requirements while ensuring safety and regulations 1151 compliance. This study proposes a qualitative four-quadrant building block framework, 1152 1153 composed by bioconversion, cracking, extraction and enhancement processes to systematically compare value chains, and classifies the reviewed waste-to-nutrition 1154 1155 approaches into eight generic families.

1156 Waste-to-nutrition pathways directly target the reduction, closing and shortcutting of nutrients flows loops, and should therefore be integrated within the broader context of 1157 1158 transition towards low fossil carbon and planetary boundaries-compliant economies. In this regard, this comprehensive review highlights the wide span of the basket of solutions, where 1159 traditional residual biomass recycling approaches (e.g. livestock, compost) are likely 1160 complemented with emerging biotechnologies and extraction processes. Yet, further work 1161 1162 remains necessary to capture the economic- and environmental relevance of a wide deployment of waste-to-nutrition strategies, considering context-dependencies. 1163

1164

#### 1165 **CRediT authorship contribution statement**

- U. Javourez: Conceptualization, Data curation, Formal analysis, Investigation, Methodology,
   Visualization, Writing Original Draft.
- 1168 M. O'Donohue: Validation, Writing Review & Editing.
- 1169 L. Hamelin: Conceptualization, Funding acquisition, Resources, Supervision, Validation, 1170 Writing – Review & Editing.
- 1171

#### 1172 **Declaration of competing interests**

- 1173 The authors declare no conflict of interest.
- 1174

#### 1175 Acknowledgement

1176 This work was carried out within the framework of the research project Cambioscop 1177 (https://cambioscop.cnrs.fr), partly financed by the French National Research Agency, 1178 Programme Investissement d'Avenir (ANR-17-MGPA-0006) and Region Occitanie 1179 (18015981). U. Javourez was additionally funded by the Metaprogram GLOFOODS (INRAE-1180 CIRAD).

- 1181
- 1182 Credits

Figures of the present manuscript have been designed using free icons resources from
Flaticon.com (authors: Surang, Linector, Good Ware, Monkik, Smashicons, Smalllikeart,
Itim2101, DinosoftLabs, Roundicons, Vector markets, Darius Dan) and Wikipedia commons.

- 1187
- 1188 Bibliography
- Abbey, L., Glover-Amengor, M., Atikpo, M.O., Atter, A., Toppe, J., 2017. Nutrient content of fish
  powder from low value fish and fish byproducts. Food Sci. Nutr. 5, 374–379.
  https://doi.org/10.1002/fsn3.402
- 1192Abel, J.-D., Blanc, M., 2017. Aiming for a sustainable bioeconomy (Opinion and Report), Journal1193Officiel de la République Française. ESEC, Paris, France.
- Achilonu, M., Shale, K., Arthur, G., Naidoo, K., Mbatha, M., 2018. Phytochemical Benefits of
   Agroresidues as Alternative Nutritive Dietary Resource for Pig and Poultry Farming. J. Chem.
   https://doi.org/10.1155/2018/1035071
- Acosta, N., Sakarika, M., Kerckhof, F.-M., Law, C.K.Y., De Vrieze, J., Rabaey, K., 2020. Microbial protein
   production from methane via electrochemical biogas upgrading. Chem. Eng. J. 391, 123625.
   https://doi.org/10.1016/j.cej.2019.123625
- Adelina, A., Feliatra, F., Siregar, Y.I., Suharman, I., 2020. Utilization of feather meal fermented Bacillus
   subtilis to replace fish meal in the diet of silver pompano, Trachinotus blochii (Lacepede,
   1801). AACL Bioflux 13, 100–108.
- Aggelopoulos, T., Katsieris, K., Bekatorou, A., Pandey, A., Banat, I.M., Koutinas, A.A., 2014. Solid state
   fermentation of food waste mixtures for single cell protein, aroma volatiles and fat
   production. Food Chem. 145, 710–716. https://doi.org/10.1016/j.foodchem.2013.07.105
- 1206 Agreste, 2019. Chapitre 12 Produits agroalimentaires. Alimentation animale, in: Graph'Agri 2020,
- 1207 Chiffres Clés.

- 1208Agreste, 2017. Les matières premières de l'alimentation animale en 2015 (No. 181), Chiffres et1209données Agroalimentaire.
- 1210Agroväst, 2020. Green Valleys [WWW Document]. URL https://agrovast.se/eu-projekt/green-1211valleys/green-valleys-in-english/ (accessed 12.4.20).
- Alao, B., Falowo, A., Chulayo, A.Y., Muchenje, V., 2017. The Potential of Animal By-Products in Food
   Systems: Production, Prospects and Challenges. Sustainability 9, 1089.
   https://doi.org/10.3390/su9071089
- Alexander, P., Brown, C., Arneth, A., Dias, C., Finnigan, J., Moran, D., Rounsevell, M.D.A., 2017a.
   Could consumption of insects, cultured meat or imitation meat reduce global agricultural
   land use? Glob. Food Sec. 15, 22–32. https://doi.org/10.1016/j.gfs.2017.04.001
- Alexander, P., Brown, C., Arneth, A., Finnigan, J., Moran, D., Rounsevell, M.D.A., 2017b. Losses,
   inefficiencies and waste in the global food system. Agric. Syst. 153, 190–200.
   https://doi.org/10.1016/j.agsy.2017.01.014
- Alloul, A., Ganigué, R., Spiller, M., Meerburg, F., Cagnetta, C., Rabaey, K., Vlaeminck, S.E., 2018.
   Capture–Ferment–Upgrade: A Three-Step Approach for the Valorization of Sewage Organics as Commodities. Environ. Sci. Technol. 52, 6729–6742.
   https://doi.org/10.1021/acs.est.7b05712
  - 224 https://doi.org/10.1021/acs.est.7b05712 225 Alloul A Muys M Hertoghs N Kerckhof E M Vlaemin
- Alloul, A., Muys, M., Hertoghs, N., Kerckhof, F.-M., Vlaeminck, S.E., 2021a. Cocultivating aerobic
   heterotrophs and purple bacteria for microbial protein in sequential photo- and
   chemotrophic reactors. Bioresour. Technol. 319, 124192.
   https://doi.org/10.1016/j.biortech.2020.124192
- Alloul, A., Spanoghe, J., Machado, D., Vlaeminck, S.E., 2021b. Unlocking the genomic potential of
   aerobes and phototrophs for the production of nutritious and palatable microbial food
   without arable land or fossil fuels. Microb. Biotechnol. https://doi.org/10.1111/1751 7915.13747
- Alvarado, K.A., García Martínez, J.B., Matassa, S., Egbejimba, J., Denkenberger, D., 2021. Food in
   space from hydrogen-oxidizing bacteria. Acta Astronaut. 180, 260–265.
   https://doi.org/10.1016/j.actaastro.2020.12.009
- Amenorfenyo, D.K., Huang, X., Zhang, Y., Zeng, Q., Zhang, N., Ren, J., Huang, Q., 2019. Microalgae
   Brewery Wastewater Treatment: Potentials, Benefits and the Challenges. IJERPH.
   https://doi.org/10.3390/ijerph16111910
- Andurand, J., Coulmier, D., Despres, J.L., Rambourg, J.C., 2010. Extraction industrielle de protéines et de pigments chez la luzerne: état des lieux et perspectives, Innovations Agronomiques.
   INRAE.
- ANSES, 2015. Opinion of the French Agency for Food, Environmental and Occupational Health &
  Safety on "the use of insects as food and feed and the review of scientific knowledge on the
  health risks related to the consumption of insects" (No. 2014- SA- 0153), ANSES Opinion.
  ANSES, Maison-Alfort.
- Antonsen, T., McGowan, J.M., 2021. 25. Exploring alternative food futures through critical design, in:
   Justice and Food Security in a Changing Climate. Wageningen Academic Publishers, pp. 176–
   181. https://doi.org/10.3920/978-90-8686-915-2\_25
- 1249Aschemann-Witzel, J., Peschel, A.O., 2019. How circular will you eat? The sustainability challenge in1250food and consumer reaction to either waste-to-value or yet underused novel ingredients in1251food. Food Qual. Prefer. 77, 15–20. https://doi.org/10.1016/j.foodqual.2019.04.012
- Asim, A.M., Uroos, M., Muhammad, N., Hallett, J.P., 2021. Production of Food-Grade Glucose from
   Rice and Wheat Residues Using a Biocompatible Ionic Liquid. ACS Sustainable Chem. Eng.
   https://doi.org/10.1021/acssuschemeng.1c00022
- Aspevik, T., Oterhals, Å., Rønning, S.B., Altintzoglou, T., Wubshet, S.G., Gildberg, A., Afseth, N.K.,
   Whitaker, R.D., Lindberg, D., 2017. Valorization of Proteins from Co- and By-Products from
   the Fish and Meat Industry. Top. Curr. Chem. 375, 53. https://doi.org/10.1007/s41061-017 0143-6
- 1259 Avecom, 2020. Bioproducts & Apps [WWW Document]. URL https://avecom.be/ (accessed 10.22.20).

- Axelos, M.A.V., Bamière L., Colin F., Dourmad J.-Y., Duru M., Gillot S., Kurek B., Mathias J.-D., Méry J.,
   O'Donohue M., Recous S., Requillart V., Steyer J.-P., Thomas A., Thoyer S., De Vries H.,
   Wohlfahrt J., 2020. Réflexion prospective interdisciplinaire bioéconomie. Rapport de
   synthèse. INRAE 70. https://doi.org/10.15454/X30B-QD69
- Azagoh, C., Ducept, F., Garcia, R., Rakotozafy, L., Cuvelier, M.-E., Keller, S., Lewandowski, R.,
   Mezdour, S., 2016. Extraction and physicochemical characterization of Tenebrio molitor
   proteins. Food Res. Int. 88, 24–31. https://doi.org/10.1016/j.foodres.2016.06.010
- Baker, P.W., Charlton, A., 2020. A comparison in protein extraction from four major crop residues in
   Europe using chemical and enzymatic processes-a review. Innov. Food Sci. Emerg. Technol.
   59, 102239. https://doi.org/10.1016/j.ifset.2019.102239
- Baldoni, E., Reumerman, P., Parisi, C., Platt, R., Gonzalez Hermoso, H., Vilka, K., Vos, J., M'barek, R.,
  2021. Chemical and material driven biorefineries in the EU and beyond: database and
  dashboard visualisation. Publications Office of the European Union, Luxembourg.
- 1273Bals, B., Dale, B., 2011. Economic Comparison of Multiple Techniques for Recovering Leaf Protein in1274Biomass Processing. Biotechnol. Bioeng. 108, 530–537. https://doi.org/10.1002/bit.22973
- Bayat, H., Cheng, F., Dehghanizadeh, M., Brewer, C.E., 2021. Recovery of Nitrogen from Low-Cost
   Plant Feedstocks Used for Bioenergy: A Review of Availability and Process Order. Energy
   Fuels. https://doi.org/10.1021/acs.energyfuels.1c02140
- Bellettini, M.B., Fiorda, F.A., Maieves, H.A., Teixeira, G.L., Ávila, S., Hornung, P.S., Júnior, A.M., Ribani,
   R.H., 2019. Factors affecting mushroom Pleurotus spp. Saudi J. Biol. Sci. 26, 633–646.
   https://doi.org/10.1016/j.sjbs.2016.12.005
- Bello, S., Salim, I., Feijoo, G., Moreira, M.T., 2021. Inventory review and environmental evaluation of
   first- and second-generation sugars through life cycle assessment. Environ. Sci. Pollut. Res.
   28, 27345–27361. https://doi.org/10.1007/s11356-021-12405-y
- Belyaev, V.D., Chepigo, S.V., Korotchenko, N.I., Mezentsev, A.N., Samokhina, O.V., Krasinskaya, A.L.,
   Timkin, V.N., Shkop, Y.Y., Colubev, A.O., 1978. Method for processing activated sludge into
   useful products. US4119495A.
- 1287Ben-Othman, S., Jõudu, I., Bhat, R., 2020. Bioactives from Agri-Food Wastes: Present Insights and1288Future Challenges. Molecules 25, 510. https://doi.org/10.3390/molecules25030510
- Bentsen, N.S., Møller, I.M., 2017. Solar energy conserved in biomass: Sustainable bioenergy use and reduction of land use change. Renew. Sust. Energ. Rev. 71, 954–958.
   https://doi.org/10.1016/j.rser.2016.12.124
- Bernaerts, T.M.M., Gheysen, L., Foubert, I., Hendrickx, M.E., Van Loey, A.M., 2019. The potential of
   microalgae and their biopolymers as structuring ingredients in food: A review. Biotechnol.
   Adv. 37, 107419. https://doi.org/10.1016/j.biotechadv.2019.107419
- Bessa, L.W., Pieterse, E., Marais, J., Hoffman, L.C., 2020. Why for feed and not for human
   consumption? The black soldier fly larvae. Compr. Rev. Food Sci. Food Saf. 19, 2747–2763.
   https://doi.org/10.1111/1541-4337.12609
- Bhat, R., Ahmad, A., Jõudu, I., 2020. Applications of Lignin in the Agri-Food Industry, in: Sharma, S.,
   Kumar, A. (Eds.), Lignin: Biosynthesis and Transformation for Industrial Applications, Springer
   Series on Polymer and Composite Materials. Springer International Publishing, Cham, pp.
   275–298. https://doi.org/10.1007/978-3-030-40663-9\_10
- Billen, G., Aguilera, E., Einarsson, R., Garnier, J., Gingrich, S., Grizzetti, B., Lassaletta, L., Le Noë, J.,
   Sanz-Cobena, A., 2021. Reshaping the European agro-food system and closing its nitrogen
   cycle: The potential of combining dietary change, agroecology, and circularity. One Earth 4,
   839–850. https://doi.org/10.1016/j.oneear.2021.05.008
- 1306Biopilz, 2020. Fine Funghi [WWW Document]. URL https://biopilz.ch/fr/production.html (accessed130710.22.20).
- Boccia, F., Covino, D., Sarnacchiaro, P., 2018. Genetically modified food versus knowledge and fear: A
   Noumenic approach for consumer behaviour. Food Res. Int. 111, 682–688.
   https://doi.org/10.1016/j.foodres.2018.06.013

- Bohnes, F.A., Laurent, A., 2021. Environmental impacts of existing and future aquaculture
   production: Comparison of technologies and feed options in Singapore. Aquaculture 532,
   736001. https://doi.org/10.1016/j.aquaculture.2020.736001
- Boland, M.J., Rae, A.N., Vereijken, J.M., Meuwissen, M.P.M., Fischer, A.R.H., van Boekel, M.A.J.S.,
  Rutherfurd, S.M., Gruppen, H., Moughan, P.J., Hendriks, W.H., 2013. The future supply of
  animal-derived protein for human consumption. Trends Food Sci. Technol. 29, 62–73.
  https://doi.org/10.1016/j.tifs.2012.07.002
- Borreani, G., Tabacco, E., Schmidt, R.J., Holmes, B.J., Muck, R.E., 2018. Silage review: Factors affecting
  dry matter and quality losses in silages. J. Dairy Sci. 101, 3952–3979.
  https://doi.org/10.3168/jds.2017-13837
- Bossier, P., Ekasari, J., 2017. Biofloc technology application in aquaculture to support sustainable
   development goals. Microb. Biotechnol. 10, 1012–1016. https://doi.org/10.1111/1751 7915.12836
- Cadinu, L.A., Barra, P., Torre, F., Delogu, F., Madau, F.A., 2020. Insect Rearing: Potential, Challenges,
   and Circularity. Sustainability 12, 4567. https://doi.org/10.3390/su12114567
- Cameron, B., O'Neill, S., Bushnell, C., Weston, Z., Derbes, E., Szejda, K., 2019. Plant-based meat, eggs
   and dairy, State of the Industry Report. The Good Food Institute.
- 1328 Cao, K., Zhi, R., Zhang, G., 2020. Photosynthetic bacteria wastewater treatment with the production
  1329 of value-added products: A review. Bioresour. Technol. 299, 122648.
  1330 https://doi.org/10.1016/j.biortech.2019.122648
- Caporusso, A., Capece, A., Bari, I.D., 2021. Oleaginous Yeasts as Cell Factories for the Sustainable
   Production of Microbial Lipids by the Valorization of Agri-Food Wastes. Fermentation 7.
   https://doi.org/10.3390/fermentation7020050
- Capson-Tojo, G., Batstone, D.J., Grassino, M., Vlaeminck, S.E., Puyol, D., Verstraete, W., Kleerebezem,
   R., Oehmen, A., Ghimire, A., Pikaar, I., Lema, J.M., Hülsen, T., 2020. Purple phototrophic
   bacteria for resource recovery: Challenges and opportunities. Biotechnol. Adv. 43, 107567.
   https://doi.org/10.1016/j.biotechadv.2020.107567
- Castoldi, R., Bracht, A., de Morais, G.R., Baesso, M.L., Correa, R.C.G., Peralta, R.A., Moreira, R. de
  F.P.M., Polizeli, M. de L.T. de M., de Souza, C.G.M., Peralta, R.M., 2014. Biological
  pretreatment of Eucalyptus grandis sawdust with white-rot fungi: Study of degradation
  patterns and saccharification kinetics. Chem. Eng. J. 258, 240–246.
- 1342 https://doi.org/10.1016/j.cej.2014.07.090
- Castrica, M., Tedesco, D.E.A., Panseri, S., Ferrazzi, G., Ventura, V., Frisio, D.G., Balzaretti, C.M., 2018.
  Pet Food as the Most Concrete Strategy for Using Food Waste as Feedstuff within the
  European Context: A Feasibility Study. Sustainability 10, 2035.
  https://doi.org/10.3390/su10062035
- 1347 Cesaro, J.-D., Cantard, T., Leroy, M.-L.N., Peyre, M.-I., Huyen, L.T.T., Duteurtre, G., 2019. Les élevages 1348 recycleurs de déchets alimentaires à Hanoï : un service informel en transition. Flux N° 116 1349 117, 74–94.
- 1350 Chaitanya Reddy, C., Khilji, I.A., Gupta, A., Bhuyar, P., Mahmood, S., Saeed AL-Japairai, K.A., Chua,
   1351 G.K., 2021. Valorization of keratin waste biomass and its potential applications. J. Water
   1352 Process 40, 101707. https://doi.org/10.1016/j.jwpe.2020.101707
- 1353 Chanakya, H.N., Malayil, S., Vijayalakshmi, C., 2015. Cultivation of Pleurotus spp. on a combination of
   1354 anaerobically digested plant material and various agro-residues. Energy Sustain. Dev. 27, 84–
   1355 92. https://doi.org/10.1016/j.esd.2015.04.007
- Chandel, A.K., Antunes, F.A.F., Terán-Hilares, R., Cota, J., Ellilä, S., Silveira, M.H.L., dos Santos, J.C., da
   Silva, S.S., 2018. Bioconversion of Hemicellulose Into Ethanol and Value-Added Products:
   Commercialization, Trends, and Future Opportunities, in: Advances in Sugarcane Biorefinery.
   Elsevier Inc., pp. 97–134. https://doi.org/10.1016/B978-0-12-804534-3.00005-7
- 1360 Chapoutot, P., Dorleans, M., Sauvant, D., 2010. Études des cinétiques de dégradation dans le rumen
   1361 des constituants pariétaux des aliments concentrés et coproduits agroindustriels. INRA Prod.
   1362 Anim. 23, 285–304. https://doi.org/10.20870/productions-animales.2010.23.3.3309

Chapoutot, P., Rouillé, B., Sauvant, D., Renaud, B., 2019. Les coproduits de l'industrie agro-1363 alimentaire : des ressources alimentaires de qualité à ne pas négliger. INRA Prod. Anim. 31, 1364 1365 201-220. https://doi.org/10.20870/productions-animales.2018.31.3.2353 1366 Chaudhary, L., Siddiqui, M.H., Vimal, A., Bhargava, P., 2021. Biological Degradation of Keratin by 1367 Microbial Keratinase for Effective Waste Management and Potent Industrial Applications. 1368 Curr. Protein Pept. Sci. https://doi.org/10.2174/1389203722666210215151952 1369 Chavan, R., Mutnuri, S., 2020. Demonstration of pilot-scale integrative treatment of nitrogenous industrial effluent for struvite and algal biomass production. J. Appl. Phycol. 32, 1215–1229. 1370 1371 https://doi.org/10.1007/s10811-019-01978-4 1372 Chebaibi, S., Leriche Grandchamp, M., Burgé, G., Clément, T., Allais, F., Laziri, F., 2019. Improvement 1373 of protein content and decrease of anti-nutritional factors in olive cake by solid-state 1374 fermentation: A way to valorize this industrial by-product in animal feed. J. Biosci. Bioeng. 1375 128, 384-390. https://doi.org/10.1016/j.jbiosc.2019.03.010 Chen, F., Grimm, A., Eilertsen, L., Martín, C., Arshadi, M., Xiong, S., 2021. Integrated production of 1376 1377 edible mushroom (Auricularia auricular-judae), fermentable sugar and solid biofuel. Renew. 1378 Energy 170, 172-180. https://doi.org/10.1016/j.renene.2021.01.124 1379 Chia, S.Y., Tanga, C.M., van Loon, J.J.A., Dicke, M., 2019. Insects for sustainable animal feed: inclusive 1380 business models involving smallholder farmers. Curr. Opin. Environ. Sustain. https://doi.org/10.1016/j.cosust.2019.09.003 1381 Choi, K.R., Yu, H.E., Lee, S.Y., 2021. Microbial food: microorganisms repurposed for our food. Microb. 1382 1383 Biotechnol. https://doi.org/10.1111/1751-7915.13911 1384 Christiaens, M.E.R., Gildemyn, S., Matassa, S., Ysebaert, T., De Vrieze, J., Rabaey, K., 2017. 1385 Electrochemical Ammonia Recovery from Source-Separated Urine for Microbial Protein 1386 Production. Environ. Sci. Technol. 51, 13143–13150. 1387 https://doi.org/10.1021/acs.est.7b02819 1388 Ciani, M., Lippolis, A., Fava, F., Rodolfi, L., Niccolai, A., Tredici, M.R., 2021. Microbes: Food for the 1389 Future. Foods 10, 971. https://doi.org/10.3390/foods10050971 Claassens, N.J., Sousa, D.Z., dos Santos, V.A.P.M., de Vos, W.M., van der Oost, J., 2016. Harnessing 1390 1391 the power of microbial autotrophy. Nat. Rev. Microbiol. 14, 692–706. https://doi.org/10.1038/nrmicro.2016.130 1392 Clark, M.A., Domingo, N.G.G., Colgan, K., Thakrar, S.K., Tilman, D., Lynch, J., Azevedo, I.L., Hill, J.D., 1393 1394 2020. Global food system emissions could preclude achieving the 1.5° and 2°C climate 1395 change targets. Science 370, 705–708. https://doi.org/10.1126/science.aba7357 1396 Clauwaert, P., Muys, M., Alloul, A., De Paepe, J., Luther, A., Sun, X., Ilgrande, C., Christiaens, M.E.R., 1397 Hu, X., Zhang, D., Lindeboom, R.E.F., Sas, B., Rabaey, K., Boon, N., Ronsse, F., Geelen, D., 1398 Vlaeminck, S.E., 2017. Nitrogen cycling in Bioregenerative Life Support Systems: Challenges 1399 for waste refinery and food production processes. Prog. Aerosp. Sci. 91, 87–98. 1400 https://doi.org/10.1016/j.paerosci.2017.04.002 1401 Colonna, P., 2020. Bioraffineries. Techniques de l'ingénieur, Procédés chimie - bio - agro | Chimie 1402 verte. 1403 Contreras, M. del M., Lama-Muñoz, A., Manuel Gutiérrez-Pérez, J., Espínola, F., Moya, M., Castro, E., 1404 2019. Protein extraction from agri-food residues for integration in biorefinery: Potential 1405 techniques and current status. Bioresour. Technol. 280, 459–477. 1406 https://doi.org/10.1016/j.biortech.2019.02.040 1407 Corona, A., Ambye-Jensen, M., Vega, G.C., Hauschild, M.Z., Birkved, M., 2018a. Techno-1408 environmental assessment of the green biorefinery concept: Combining process simulation 1409 and life cycle assessment at an early design stage. Sci. Total Environ. 635, 100–111. https://doi.org/10.1016/j.scitotenv.2018.03.357 1410 1411 Corona, A., Parajuli, R., Ambye-Jensen, M., Hauschild, M.Z., Birkved, M., 2018b. Environmental 1412 screening of potential biomass for green biorefinery conversion. J. Clean. Prod. 189, 344-1413 357. https://doi.org/10.1016/j.jclepro.2018.03.316

- 1414 Cortes Ortiz, J.A., Ruiz, A.T., Morales-Ramos, J. A., Thomas, M., Rojas, M. G., Tomberlin, J.K., Yi, L.,
  1415 Han, R., Giroud, L., Jullien, R.L., 2016. Insect Mass Production Technologies, in: Dossey, A.T.,
  1416 Morales-Ramos, Juan A., Rojas, M. Guadalupe (Eds.), Insects as Sustainable Food Ingredients,
  1417 Production, Processing and Food Applications. Academic Press, San Diego, pp. 153–201.
  1418 https://doi.org/10.1016/B978-0-12-802856-8.00006-5
- Crab, R., Defoirdt, T., Bossier, P., Verstraete, W., 2012. Biofloc technology in aquaculture: Beneficial
   effects and future challenges. Aquaculture 356–357, 351–356.
- 1421 https://doi.org/10.1016/j.aquaculture.2012.04.046
- 1422 Crosser, N., Bushnell, C., Derbes, E., Friedrich, B., Lamy, J., Manu, N., Swartz, E., 2019. Cultivated
   1423 Meat, State of the Industry Report. The Good Food Institute.
- 1424da Costa Rocha, A.C., José de Andrade, C., de Oliveira, D., 2021. Perspective on integrated biorefinery1425for valorization of biomass from the edible insect Tenebrio molitor. Trends Food Sci. Technol.1426116, 480–491. https://doi.org/10.1016/j.tifs.2021.07.012
- 1427Dale, B.E., Allen, M.S., Laser, M., Lynd, L.R., 2009. Protein feeds coproduction in biomass conversion1428to fuels and chemicals. Biofuel Bioprod Bior 3, 219–230. https://doi.org/10.1002/bbb.132
- Damborg, V.K., Jensen, S.K., Weisbjerg, M.R., Adamsen, A.P., Stødkilde, L., 2020. Screw-pressed
   fractions from green forages as animal feed: Chemical composition and mass balances. Anim.
   Feed Sci. Technol. 261, 114401. https://doi.org/10.1016/j.anifeedsci.2020.114401
- Damborg, V.K., Stødkilde, L., Jensen, S.K., Weisbjerg, M.R., 2018. Protein value and degradation
   characteristics of pulp fibre fractions from screw pressed grass, clover, and lucerne. Anim.
   Feed Sci. Technol. 244, 93–103. https://doi.org/10.1016/j.anifeedsci.2018.08.004
- Davys, M.N.G., Richardier, F.C., Kennedy, D., Mathan, O. de, Collin, S.M., Subtil, J., Bertin, E., Davys,
  M.J., 2011. Leaf concentrate and other benefits of leaf fractionation., in: Combating
  Micronutrient Deficiencies: Food-Based Approaches. FAO, pp. 338–365.
  https://doi.org/10.1079/9781845937140.0338
- 1439Dawood, M.A.O., Koshio, S., 2020. Application of fermentation strategy in aquafeed for sustainable1440aquaculture. Rev. Aquacult. 12, 987–1002. https://doi.org/10.1111/raq.12368
- de Boer, H.C., van Krimpen, M.M., Blonk, H., Tyszler, M., 2014. Replacement of soybean meal in
  compound feed by European protein sources (No. 819), Wageningen UR Livestock Research.
  Wageningen, Lelystad.
- De Groot, L., Bogdanski, A., 2013. Bioslurry = Brown Gold ? A review of scientific literature on the co product of biogas production (Environmental and natural resources management working
   paper No. 55), Climate, Energy and Tenure Division (NRC). FAO.
- 1447De Jong, E., Stichnothe, H., Bell, G., Jorgensen, H., 2020. Bio-based Chemical, a 2020 Update,1448Technology Collaboration Programme. IEA Bioenergy.
- de Menezes, C.L.A., Santos, R. do C., Santos, M.V., Boscolo, M., da Silva, R., Gomes, E., da Silva, R.R.,
  2021. Industrial sustainability of microbial keratinases: production and potential applications.
  World J. Microbiol. Biotechnol. 37, 86. https://doi.org/10.1007/s11274-021-03052-z
- De Vrieze, J., Verbeeck, K., Pikaar, I., Boere, J., Van Wijk, A., Rabaey, K., Verstraete, W., 2020. The
  hydrogen gas bio-based economy and the production of renewable building block chemicals,
  food and energy. N. Biotechnol. 55, 12–18. https://doi.org/10.1016/j.nbt.2019.09.004
- 1455 Deep Branch Biotechnology, 2020. alternative protein, Simplified [WWW Document]. URL 1456 https://deepbranchbio.com/ (accessed 10.23.20).
- 1457Denkenberger, D., Pearce, J.M., 2015. Feeding everyone no matter what: Managing food security1458after global catastrophe, Academic Press. ed. Elsevier Science.
- 1459Denkenberger, D., Pearce, J.M., Taylor, A.R., Black, R., 2019. Food without sun: price and life-saving1460potential. Foresight 21, 118–129. https://doi.org/10.1108/FS-04-2018-0041
- 1461 Dessì, P., Rovira-Alsina, L., Sánchez, C., Dinesh, G.K., Tong, W., Chatterjee, P., Tedesco, M., Farràs, P.,
  1462 Hamelers, H.M.V., Puig, S., 2020. Microbial electrosynthesis: Towards sustainable
  1463 biorefineries for production of green chemicals from CO2 emissions. Biotechnol. Adv.
  1464 107675. https://doi.org/10.1016/j.biotechadv.2020.107675

- Di Stefano, E., Agyei, D., Njoku, E.N., Udenigwe, C.C., 2018. Plant RuBisCo: An Underutilized Protein 1465 for Food Applications. J. Am. Oil Chem. Soc. 95, 1063–1074. 1466 1467 https://doi.org/10.1002/aocs.12104
- 1468 Djomo, S.N., Knudsen, M.T., Martinsen, L., Andersen, M.S., Ambye-Jensen, M., Møller, H.B., 1469 Hermansen, J.E., 2020. Green proteins: An energy-efficient solution for increased self-1470 sufficiency in protein in Europe. Biofuels, Bioproducts and Biorefining 14, 605–619. 1471 https://doi.org/10.1002/bbb.2098
- Domingos, J.M.B., Teixeira, A.R.S., Dupoiron, S., Allais, F., Lameloise, M.-L., 2020. Simultaneous 1472 1473 recovery of ferulic acid and sugars from wheat bran enzymatic hydrolysate by 1474 diananofiltration. Sep. Purif. Technol. 242, 116755.
- 1475 https://doi.org/10.1016/j.seppur.2020.116755
- 1476 Dortmans, B.M.A., Diener, S., Verstappen, B.M., Zurbrügg, C., 2017. Black Soldier Fly Biowaste 1477 Processing - A Step-by-Step Guide. Eawag: Swiss Federal Institute of Aquatic Science and 1478 Technology, Dübendorf, Switzerland.
- 1479 Dou, J., Huang, Y., Ren, H., Li, Z., Cao, Q., Liu, X., Li, D., 2019. Autotrophic, Heterotrophic, and 1480 Mixotrophic Nitrogen Assimilation for Single-Cell Protein Production by Two Hydrogen-1481 Oxidizing Bacterial Strains. Appl. Biochem. Biotechnol. 187, 338–351. 1482 https://doi.org/10.1007/s12010-018-2824-1
- Dou, Z., Toth, J.D., Westendorf, M.L., 2018. Food waste for livestock feeding: Feasibility, safety, and 1483 1484 sustainability implications. Glob. Food Sec. 17, 154–161. 1485
  - https://doi.org/10.1016/j.gfs.2017.12.003
- 1486 Dries, L., Heijman, W., Jongeneel, R., Purnhagen, K., Wesseler, J., 2020. EU Bioeconomy: economics 1487 and policies: Volume II, Palgrave advances in bioeconomy: economics and policies. Springer 1488 Nature, Switzerland.
- 1489 Du, Y., Zou, W., Zhang, K., Ye, G., Yang, J., 2020. Advances and Applications of Clostridium Co-culture 1490 Systems in Biotechnology. Front. Microbiol. 11, 560223. 1491 https://doi.org/10.3389/fmicb.2020.560223
- 1492 Dupoiron, S., Lameloise, M.-L., Pommet, M., Bennaceur, O., Lewandowski, R., Allais, F., Teixeira, 1493 A.R.S., Rémond, C., Rakotoarivonina, H., 2017. A novel and integrative process: From 1494 enzymatic fractionation of wheat bran with a hemicellulasic cocktail to the recovery of ferulic 1495 acid by weak anion exchange resin. Ind. Crops Prod. 105, 148–155. 1496 https://doi.org/10.1016/j.indcrop.2017.05.004
- 1497 Duque-Acevedo, M., Belmonte-Ureña, L.J., Cortés-García, F.J., Camacho-Ferre, F., 2020. Agricultural 1498 waste: Review of the evolution, approaches and perspectives on alternative uses. Glob. Ecol. 1499 Conserv. 22, e00902. https://doi.org/10.1016/j.gecco.2020.e00902
- 1500 Duru, M., Magrini, M.-B., 2017. Composition en acides gras poly-insaturés de notre assiette et 1501 utilisation des matières premières agricoles en France : une amélioration lente, mais 1502 insuffisante. OCL 24, A201. https://doi.org/10.1051/ocl/2017007
- 1503 EC Directive, 2008. Directive 2008/98/EC of the European Parliament and of the Council of 19 1504 November 2008 on waste and repealing certain Directives (Text with EEA relevance), 312.
- 1505 EFSA Panel on Dietetic Products, Nutrition and Allergies (NDA), 2016. Guidance on the preparation 1506 and presentation of an application for authorisation of a novel food in the context of 1507 Regulation (EU) 2015/2283. EFSA Journal, Scientific Opinion 14. 1508 https://doi.org/10.2903/j.efsa.2016.4594
- 1509 EFSA Panel on Dietetic Products, Nutrition and Allergies (NDA), 2012. Scientific Opinion on Dietary 1510 Reference Values for protein. EFSA Journal, Scientific Opinion 10, 2557. 1511 https://doi.org/10.2903/j.efsa.2012.2557
- EFSA Panel on Nutrition, Novel Foods and Food Allergens (NDA), Turck, D., Bohn, T., Castenmiller, J., 1512 1513 Henauw, S.D., Hirsch-Ernst, K.I., Maciuk, A., Mangelsdorf, I., McArdle, H.J., Naska, A., Pelaez, 1514 C., Pentieva, K., Siani, A., Thies, F., Tsabouri, S., Vinceti, M., Cubadda, F., Frenzel, T., 1515 Heinonen, M., Marchelli, R., Neuhäuser-Berthold, M., Poulsen, M., Maradona, M.P.,
- 1516 Schlatter, J.R., Loveren, H. van, Goumperis, T., Knutsen, H.K., 2021a. Safety of frozen and

- 1517 dried formulations from whole house crickets (Acheta domesticus) as a Novel food pursuant
- 1518 to Regulation (EU) 2015/2283. EFSA Journal, Scientific Opinion 19, e06779.
- 1519 https://doi.org/10.2903/j.efsa.2021.6779
- EFSA Panel on Nutrition, Novel Foods and Food Allergens (NDA), Turck, D., Castenmiller, J., Henauw,
  S.D., Hirsch-Ernst, K.I., Kearney, J., Maciuk, A., Mangelsdorf, I., McArdle, H.J., Naska, A.,
  Pelaez, C., Pentieva, K., Siani, A., Thies, F., Tsabouri, S., Vinceti, M., Cubadda, F., Frenzel, T.,
  Heinonen, M., Marchelli, R., Neuhäuser-Berthold, M., Poulsen, M., Maradona, M.P.,
  Schlatter, J.R., Loveren, H. van, Azzollini, D., Knutsen, H.K., 2021b. Safety of frozen and dried
- 1525formulations from migratory locust (Locusta migratoria) as a Novel food pursuant to1526Regulation (EU) 2015/2283. EFSA Journal, Scientific Opinion 19, e06667.
- 1527 https://doi.org/10.2903/j.efsa.2021.6667
- EFSA Panel on Nutrition, Novel Foods and Food Allergens (NDA), Turck, D., Castenmiller, J., Henauw,
  S.D., Hirsch-Ernst, K.I., Kearney, J., Maciuk, A., Mangelsdorf, I., McArdle, H.J., Naska, A.,
  Pelaez, C., Pentieva, K., Siani, A., Thies, F., Tsabouri, S., Vinceti, M., Cubadda, F., Frenzel, T.,
  Heinonen, M., Marchelli, R., Neuhäuser-Berthold, M., Poulsen, M., Maradona, M.P.,
  Schlatter, J.R., Loveren, H. van, Ververis, E., Knutsen, H.K., 2021c. Safety of dried yellow
  mealworm (Tenebrio molitor larva) as a novel food pursuant to Regulation (EU) 2015/2283.
- 1534 EFSA Journal, Scientific Opinion 19, e06343. https://doi.org/10.2903/j.efsa.2021.6343
- 1535EFSA Scientific Committee, 2015. Risk profile related to production and consumption of insects as1536food and feed. EFSA Journal, Opinion 13, 4257. https://doi.org/10.2903/j.efsa.2015.4257
- El Abbadi, S.H., Criddle, C.S., 2019. Engineering the Dark Food Chain. Environ. Sci. Technol. 53, 2273–
   2287. https://doi.org/10.1021/acs.est.8b04038
- 1539 Eniferbio, 2020. Value Everything [WWW Document]. URL https://www.eniferbio.fi/ (accessed1540 10.22.20).
- Ercili-Cura, D., Barth, D., 2021. Cellular Agriculture: Lab Grown Foods, American Chemical Society. ed,
   Inaugural. American Chemical Society.
- 1543European Commission, 2020. Regulation (EU) No 142/2011 of 25 February 2011 implementing1544regulation (EC) No 1069/2009 of the European Parliament and of the Council laying down1545health rules as regards animal by-products and derived products not intended for human1546consumption and implementing Council Directive 97/78/EC as regards certain samples and1547items exempt from veterinary checks at the border under that Directive. Official J. Eur.1548Union.
- European Union, 2019. Regulation (EU) 2019/1009 of the European Parliament and of the council of
   5 June 2019 laying down rules on the making available on the market of EU fertilising
   products and amending Regulations (EC) No 1069/2009 and (EC) No 1107/2009 and
   repealing Regulation (EC) No 2003/2003 (Text with EEA relevance), OJ L.
- Falco, F.C., 2018. Microbial degradation of keratin-rich porcine by-products for protein hydrolysate
   production: a process engineering perspective. Technical University of Denmark, Kgs. Lyngby.
   Falco, F.C., Espersen, R., Svensson, B., Gernaey, K., V., Lantz, A.E., 2019. An integrated strategy for the
- raico, F.C., Espersen, K., Svensson, B., Gernaey, K., V., Lanz, A.E., 2019. An integrated strategy for the
   effective production of bristle protein hydrolysate by the keratinolytic filamentous bacterium
   Amycolatopsis keratiniphila D2. Waste Manag.
- 1558 https://doi.org/10.1016/j.wasman.2019.03.067
- 1559FAO, 2020. Insects for food and feed [WWW Document]. URL http://www.fao.org/edible-insects/en/1560(accessed 10.29.20).
- 1561 Fardet, A., 2018. Chapter Three Characterization of the Degree of Food Processing in Relation With
  1562 Its Health Potential and Effects, in: Toldrá, F. (Ed.), Advances in Food and Nutrition Research.
  1563 Academic Press, pp. 79–129. https://doi.org/10.1016/bs.afnr.2018.02.002
- 1564Farmer, T.J., Mascal, M., 2015. Platform Molecules, in: Introduction to Chemicals from Biomass. John1565Wiley & Sons, Ltd, pp. 89–155. https://doi.org/10.1002/9781118714478.ch4
- 1566 Fazenda, M., Johnston, C., Mcneil, B., 2017. Bioprocess for coproduction of ethanol and1567 mycoproteins. EP3209789A1.

- 1568 Fernandes, F., Silkina, A., Fuentes-Grünewald, C., Wood, E.E., Ndovela, V.L.S., Oatley-Radcliffe, D.L., Lovitt, R.W., Llewellyn, C.A., 2020. Valorising nutrient-rich digestate: Dilution, settlement and 1569 1570 membrane filtration processing for optimisation as a waste-based media for microalgal 1571 cultivation. Waste Manag. 118, 197–208. https://doi.org/10.1016/j.wasman.2020.08.037
- Ferraro, V., 2020. Valorisation des sous-produits de la filière viande (et poisson). Viandes & 1572 1573 Produits carnés 7.
- 1574 Ferraro, V., Anton, M., Santé-Lhoutellier, V., 2016. The "sisters" α-helices of collagen, elastin and keratin recovered from animal by-products: Functionality, bioactivity and trends of 1575 application. Trends Food Sci. Technol. 51, 65–75. https://doi.org/10.1016/j.tifs.2016.03.006 1576
- 1577 Fischer, A.R.H., Van Loo, E.J., 2021. Chapter 9 - Social acceptability of radical food innovations, in: 1578 Galanakis, C.M. (Ed.), Food Technology Disruptions. Academic Press, pp. 325–361. 1579 https://doi.org/10.1016/B978-0-12-821470-1.00002-1
- 1580 Foster, J.F., Litchfield, J.H., 1964. A continuous culture apparatus for the microbial utilization of 1581 hydrogen produced by electrolysis of water in closed-cycle space systems. Biotechnol. 1582 Bioeng. 6, 441-456. https://doi.org/10.1002/bit.260060406
- 1583 Franca-Oliveira, G., Fornari, T., Hernández-Ledesma, B., 2021. A Review on the Extraction and 1584 Processing of Natural Source-Derived Proteins through Eco-Innovative Approaches. Processes 1585 9, 1626. https://doi.org/10.3390/pr9091626
- 1586 Franceschin, G., Sudiro, M., Ingram, T., Smirnova, I., Brunner, G., Bertucco, A., 2011. Conversion of rye straw into fuel and xylitol: a technical and economical assessment based on experimental 1587 1588 data. Chem. Eng. Res. Des. 89, 631-640. https://doi.org/10.1016/j.cherd.2010.11.001
- 1589 Fu, Y., Chen, T., Yuet Chen, S.H., Liu, B., Sun, P., Sun, H., Chen, F., 2021. The potentials and challenges 1590 of using microalgae as an ingredient to produce meat analogues. Trends Food Sci. Technol. 1591 112, 188–200. https://doi.org/10.1016/j.tifs.2021.03.050
- 1592 Gao, J., Wang, Y., Yan, Y., Li, Z., Chen, M., 2020. Protein extraction from excess sludge by alkali-1593 thermal hydrolysis. Environ. Sci. Pollut. Res. 27, 8628–8637. https://doi.org/10.1007/s11356-1594 019-07188-2
- García Martínez, J.B., Egbejimba, J., Throup, J., Matassa, S., Pearce, J.M., Denkenberger, D.C., 2021. 1595 1596 Potential of microbial protein from hydrogen for preventing mass starvation in catastrophic scenarios. Sustain. Prod. Consum. 25, 234–247. https://doi.org/10.1016/j.spc.2020.08.011 1597
- Garcia-Bernet, D., Ferraro, V., Moscoviz, R., 2020. Coproduits des IAA : un vivier mondial sous-1598 1599 exploité de biomolécules d'intérêt, in: Chimie Verte et Industries Agroalimentaires – Vers 1600 Une Bioéconomie Durable, Sciences et Techniques Agroalimentaires. Lavoisier Tec & Doc, 1601 Paris, pp. 149–180.
- García-Segovia, P., Igual, M., Noguerol, A.T., Martínez-Monzó, J., 2020. Use of insects and pea 1602 1603 powder as alternative protein and mineral sources in extruded snacks. Eur. Food Res. 1604 Technol. 246, 703–712. https://doi.org/10.1007/s00217-020-03441-y
- 1605 Gasco, L., Biancarosa, I., Liland, N.S., 2020. From waste to feed: a review of recent knowledge on 1606 insects as producers of protein and fat for animal feeds. Curr. Opin. Green Sustain. Chem. 23, 1607 67-79. https://doi.org/10.1016/j.cogsc.2020.03.003
- 1608 Gençdağ, E., Görgüç, A., Yılmaz, F.M., 2020. Recent Advances in the Recovery Techniques of Plant-1609 Based Proteins from Agro-Industrial By-Products. Food Rev. Int. 37, 447–468. 1610 https://doi.org/10.1080/87559129.2019.1709203
- Georganas, A., Giamouri, E., Pappas, A.C., Papadomichelakis, G., Galliou, F., Manios, T., Tsiplakou, E., 1611 1612 Fegeros, K., Zervas, G., 2020. Bioactive Compounds in Food Waste: A Review on the 1613 Transformation of Food Waste to Animal Feed. Foods 9, 291. https://doi.org/10.3390/foods9030291 1614
- 1615 Gerten, D., Heck, V., Jägermeyr, J., Bodirsky, B.L., Fetzer, I., Jalava, M., Kummu, M., Lucht, W., 1616 Rockström, J., Schaphoff, S., Schellnhuber, H.J., 2020. Feeding ten billion people is possible 1617 within four terrestrial planetary boundaries. Nat Sustain 3, 200–208.
- 1618 https://doi.org/10.1038/s41893-019-0465-1

1619 Givirovskiy, G., Ruuskanen, V., Ojala, L.S., Kokkonen, P., Ahola, J., 2019. In Situ Water Electrolyzer Stack for an Electrobioreactor. Energies 12, 1904. https://doi.org/10.3390/en12101904 1620 1621 Glencross, B.D., Huyben, D., Schrama, J.W., 2020. The Application of Single-Cell Ingredients in 1622 Aquaculture Feeds—A Review. Fishes 5, 22. https://doi.org/10.3390/fishes5030022 1623 Gmoser, R., 2021. Circular bioeconomy through valorisation of agro-industrial residues by the edible 1624 filamentous fungus Neurospora intermedia. University of Boras, Boras, Sweden. 1625 Go Grass, 2020. Denmark - Organic Protein [WWW Document]. URL https://www.go-1626 grass.eu/denmark/ (accessed 12.4.20). Godon, J.-J., Arcemisbéhère, L., Escudié, R., Harmand, J., Miambi, E., Steyer, J.-P., 2013. Overview of 1627 1628 the Oldest Existing Set of Substrate-optimized Anaerobic Processes: Digestive Tracts. 1629 Bioenerg. Res. 6, 1063–1081. https://doi.org/10.1007/s12155-013-9339-y 1630 Gold, M., Cassar, C.M., Zurbrügg, C., Kreuzer, M., Boulos, S., Diener, S., Mathys, A., 2020. Biowaste 1631 treatment with black soldier fly larvae: Increasing performance through the formulation of biowastes based on protein and carbohydrates. Waste Manag. 102, 319–329. 1632 1633 https://doi.org/10.1016/j.wasman.2019.10.036 1634 Gómez-García, R., Campos, D.A., Aguilar, C.N., Madureira, A.R., Pintado, M., 2021. Valorisation of 1635 food agro-industrial by-products: From the past to the present and perspectives. J. Environ. 1636 Manage. 299, 113571. https://doi.org/10.1016/j.jenvman.2021.113571 1637 GrasGoed, 2020. GrasGoed zet maaisel in als innovative grondstof [WWW Document]. URL 1638 https://www.grasgoed.eu/over-grasgoed/ (accessed 12.4.20). 1639 Gravel, A., Doyen, A., 2020. The use of edible insect proteins in food: Challenges and issues related to 1640 their functional properties. Innov. Food Sci. Emerg. Technol. 59, 102272. 1641 https://doi.org/10.1016/j.ifset.2019.102272 1642 Green Protein Project, 2020. Valorisation of Vegetable Processing Industry Remnants into High-Value 1643 Functional Proteins and other Food Ingredients [WWW Document]. URL 1644 http://greenproteinproject.eu/ (accessed 12.4.20). 1645 Green Spot, 2020. Challenging the food waste paradigm to feed the future. [WWW Document]. URL https://greenspot-tech.com/en/ (accessed 10.22.20). 1646 1647 Guthman, J., Biltekoff, C., 2020. Magical disruption? Alternative protein and the promise of de-1648 materialization. Environ. Plan. E. https://doi.org/10.1177/2514848620963125 Halme, A., Holmberg A., Tiussa E., 1977. Modelling and Control of a Protein Fermentation Process 1649 1650 Utilizing the Spent Sulphite Liquor. IFAC Proc. Vol. 10, 817–825. 1651 https://doi.org/10.1016/S1474-6670(17)66916-7 1652 Hamed, H.A., Abdalla, M.F.M., Hosseney, M.H., Elshaikh, K.A., 2020. Upcycling of Oyster Mushroom 1653 Spent Through Reuse as Substrate in Sequential Production Cycles of Mushroom. Egyp. J. 1654 Hortic. 47, 69-79. https://doi.org/10.21608/ejoh.2020.25962.1129 1655 Hamelin, L., Borzęcka, M., Kozak, M., Pudełko, R., 2019. A spatial approach to bioeconomy: 1656 Quantifying the residual biomass potential in the EU-27. Renew. Sust. Energ. Rev. 100, 127-1657 142. https://doi.org/10.1016/j.rser.2018.10.017 1658 Hamelin, L., Møller, H.B., Jørgensen, U., 2021. Harnessing the full potential of biomethane towards 1659 tomorrow's bioeconomy: A national case study coupling sustainable agricultural 1660 intensification, emerging biogas technologies and energy system analysis. Renew. Sust. 1661 Energ. Rev. 138, 110506. https://doi.org/10.1016/j.rser.2020.110506 Hamidoghli, A., Won, S., Farris, N.W., Bae, J., Choi, W., Yun, H., Bai, S.C., 2020. Solid state fermented 1662 plant protein sources as fish meal replacers in whiteleg shrimp Litopaeneus vannamei. Anim. 1663 1664 Feed Sci. Technol. 264, 114474. https://doi.org/10.1016/j.anifeedsci.2020.114474 1665 Harris, E.E., 1951. Molasses and Yeast From Wood. US Dept. Agr. Yearbook 887–890. Harris, E.E., 1947. Wood-sugar molasses from wood waste. Sth. Lumberm., Forest Products 1666 1667 Laboratory 175, 157–161. 1668 Hedegaard, K., Thyø, K.A., Wenzel, H., 2008. Life Cycle Assessment of an Advanced Bioethanol 1669 Technology in the Perspective of Constrained Biomass Availability. Environ. Sci. Technol. 42, 1670 7992–7999. https://doi.org/10.1021/es800358d

- 1671 Helliwell, R., Burton, R.J.F., 2021. The promised land? Exploring the future visions and narrative 1672 silences of cellular agriculture in news and industry media. J. Rural Stud. 84, 180–191. 1673 https://doi.org/10.1016/j.jrurstud.2021.04.002
- 1674 Herrero, M., Thornton, P.K., Mason-D'Croz, D., Palmer, J., Benton, T.G., Bodirsky, B.L., Bogard, J.R., Hall, A., Lee, B., Nyborg, K., Pradhan, P., Bonnett, G.D., Bryan, B.A., Campbell, B.M., 1675
- 1676 Christensen, S., Clark, M., Cook, M.T., de Boer, I.J.M., Downs, C., Dizyee, K., Folberth, C.,
- Godde, C.M., Gerber, J.S., Grundy, M., Havlik, P., Jarvis, A., King, R., Loboguerrero, A.M., 1677
- Lopes, M.A., McIntyre, C.L., Naylor, R., Navarro, J., Obersteiner, M., Parodi, A., Peoples, M.B., 1678
- 1679 Pikaar, I., Popp, A., Rockström, J., Robertson, M.J., Smith, P., Stehfest, E., Swain, S.M., Valin,
- 1680 H., van Wijk, M., van Zanten, H.H.E., Vermeulen, S., Vervoort, J., West, P.C., 2020. Innovation 1681 can accelerate the transition towards a sustainable food system. Nat Food 1, 266–272. 1682 https://doi.org/10.1038/s43016-020-0074-1
- 1683 Heuzé V., Tran G., Nozière P., Bastianelli D., Lebas F., 2020. Feather meal [WWW Document]. 1684 Feedipedia, a programme by INRAE, CIRAD, AFZ and FAO. URL 1685 https://www.feedipedia.org/node/213 (accessed 12.4.20).
- 1686 Holkar, C.R., Jain, S.S., Jadhav, A.J., Pinjari, D.V., 2018. Valorization of keratin based waste. Process 1687 Saf Environ Prot, Biowaste for Energy Recovery and Environmental Remediation 115, 85–98. 1688 https://doi.org/10.1016/j.psep.2017.08.045
- Hu, X., Kerckhof, F.-M., Ghesquière, J., Bernaerts, K., Boeckx, P., Clauwaert, P., Boon, N., 2020. 1689 Microbial Protein out of Thin Air: Fixation of Nitrogen Gas by an Autotrophic Hydrogen-1690 1691 Oxidizing Bacterial Enrichment. Environ. Sci. Technol. 54, 3609–3617. 1692
  - https://doi.org/10.1021/acs.est.9b06755
- Hubert, A., Berezina, N., 2020. Une future bioraffinerie des insectes, in: Chimie Verte et Industries 1693 1694 Agroalimentaires – Vers Une Bioéconomie Durable, Sciences et Techniques Agroalimentaires. 1695 Lavoisier Tec & Doc, Paris, pp. 327–391.
- 1696 Huizing, H.J., 2005. Single Cell Protein (SCP) als alternatief voor soja: een haalbaarheidsstudie. 1697 InnovatieNetwerk.
- 1698 Hülsen, T., Hsieh, K., Lu, Y., Tait, S., Batstone, D.J., 2018. Simultaneous treatment and single cell 1699 protein production from agri-industrial wastewaters using purple phototrophic bacteria or 1700 microalgae – A comparison. Bioresour. Technol. 254, 214–223. 1701 https://doi.org/10.1016/j.biortech.2018.01.032
- 1702 Huntley, N.F., Patience, J.F., 2018. Xylose: absorption, fermentation, and post-absorptive metabolism 1703 in the pig. J. Anim. Sci. Biotechnol. 9. https://doi.org/10.1186/s40104-017-0226-9
- 1704 Hussain, S., Jõudu, I., Bhat, R., 2020. Dietary Fiber from Underutilized Plant Resources—A Positive 1705 Approach for Valorization of Fruit and Vegetable Wastes. Sustainability 12, 5401. 1706 https://doi.org/10.3390/su12135401
- 1707 Hüttner, S., Ramkumar, N., Johansson, A., Teixeira, P., Achterberg, P., 2020. Recent Advances in the 1708 Intellectual Property Landscape of Filamentous Fungi. Fungal Biol. Biotechnol. 7. 1709 https://doi.org/10.1186/s40694-020-00106-z
- 1710 Hwang, J., Zhang, L., Seo, S., Lee, Y.-W., Jahng, D., 2008. Protein recovery from excess sludge for its 1711 use as animal feed. Bioresour. Technol. 99, 8949-8954. 1712 https://doi.org/10.1016/j.biortech.2008.05.001
- 1713 iCell Sustainable Nutrition Co., 2020. iCell Sustainable [WWW Document]. URL 1714 http://www.icellsustainable.com/en/company (accessed 10.22.20).
- 1715 Ingle, A.P., Chandel, A.K., Silva, S.S. da (Eds.), 2020. Lignocellulosic biorefining technologies, John 1716 Wiley&Sons Ltd. ed. Wiley-Blackwell, West Sussex.
- 1717 IPCC, 2019. Summary for Policymakers, in: Climate Change and Land: An IPCC Special Report on Climate Change, Desertification, Land Degradation, Sustainable Land Management, Food 1718 1719 Security and Greenhouse Gasfluxesin Terrestrial Ecosystems. [P.R. Shukla, J. Skea, E. Calvo 1720 Buendia, V. Masson-Delmotte, H.- O. Pörtner, D. C. Roberts, P. Zhai, R. Slade, S. Connors, R. 1721 van Diemen, M. Ferrat, E. Haughey, S. Luz, S. Neogi, M. Pathak, J. Petzold, J. Portugal Pereira, 1722 P. Vyas, E. Huntley, K. Kissick, M. Belkacemi, J. Malley, (eds.)]. In press.
  - 41

- 1723 IPIFF, 2021. Press Release The European Union authorises insect proteins in poultry and pig feed.
- 1724 IPIFF, 2019. The European insect sector today: challenges, opportunities and regulatory landscape.
   1725 IPIFF vision paper on the future of the insect sector towards 2030.
- 1726Jensen, H., Elleby, C., Domínguez, I.P., Chatzopoulos, T., Charlebois, P., 2021. Insect-based protein1727feed: from fork to farm. J. Insects as Food Feed. 1–16. https://doi.org/10.3920/JIFF2021.0007
- Jimenez, J., Aemig, Q., Doussiet, N., Steyer, J.-P., Houot, S., Patureau, D., 2015. A new organic matter
   fractionation methodology for organic wastes: Bioaccessibility and complexity
   characterization for treatment optimization. Bioresour. Technol. 194, 344–353.
   https://doi.org/10.1016/j.biortech.2015.07.037
- Jones, S.W., Karpol, A., Friedman, S., Maru, B.T., Tracy, B.P., 2020. Recent advances in single cell
  protein use as a feed ingredient in aquaculture. Curr. Opin. Biotechnol. 61, 189–197.
  https://doi.org/10.1016/j.copbio.2019.12.026
- Kamal, H., Le, C.F., Salter, A.M., Ali, A., 2021. Extraction of protein from food waste: An overview of
  current status and opportunities. Compr. Rev. Food Sci. Food Saf. 20, 2455–2475.
  https://doi.org/10.1111/1541-4337.12739
- 1738 Kamm, B., Schönicke, P., Hille, Ch., 2016. Green biorefinery Industrial implementation. Food Chem.
   1739 197, 1341–1345. https://doi.org/10.1016/j.foodchem.2015.11.088
- Kammes, K.L., Bals, B.D., Dale, B.E., Allen, M.S., 2011. Grass leaf protein, a coproduct of cellulosic
  ethanol production, as a source of protein for livestock. Anim. Feed Sci. Technol. 164, 79–88.
  https://doi.org/10.1016/j.anifeedsci.2010.12.006
- Kapu, N.S., Trajano, H.L., 2014. Review of hemicellulose hydrolysis in softwoods and bamboo.
   Biofuels, Bioprod. Bioref. 8, 857–870. https://doi.org/10.1002/bbb.1517
- 1745 Karan, S.K., Hamelin, L., 2020. Towards local bioeconomy: A stepwise framework for high-resolution
  1746 spatial quantification of forestry residues. Renewable and Sustainable Energy Reviews 134,
  1747 110350. https://doi.org/10.1016/j.rser.2020.110350
- Kernel.bio, 2020. Decentralization [WWW Document]. URL https://www.kernel.bio/decentralization(accessed 11.20.20).
- Khan, N.A., Hussain, S., Ahmad, N., Alam, S., Bezabih, M., Hendriks, W., Yu, P., Cone, J., 2015.
   Improving the feeding value of straws with Pleurotus ostreatus. Anim. Prod. Sci. 55.
   https://doi.org/10.1071/AN14184
- 1753 Khoshnevisan, B., Tsapekos, P., Alvarado-Morales, M., Rafiee, S., Tabatabaei, M., Angelidaki, I., 2018.
  1754 Life cycle assessment of different strategies for energy and nutrient recovery from source
  1755 sorted organic fraction of household waste. J. Clean. Prod. 180, 360–374.
  1756 https://doi.org/10.1016/j.jclepro.2018.01.198
- 1757 Khoshnevisan, B., Tsapekos, P., Zhang, Y., Valverde-Pérez, B., Angelidaki, I., 2019. Urban biowaste
   1758 valorization by coupling anaerobic digestion and single cell protein production. Bioresour.
   1759 Technol. 290, 121743. https://doi.org/10.1016/j.biortech.2019.121743
- Kiskini, A., 2017. Sugar beet leaves: from biorefinery to techno-functionality. Wageningen University,
   Wageningen, the Netherlands. https://doi.org/10.18174/421994
- 1762Kiverdi, Inc., 2020. We Are Remaking How Things Are Made With Carbon Transformation [WWW1763Document]. URL https://www.kiverdi.com (accessed 10.23.20).
- Kozubal, M.A., Macur, R.E., Avniel, Y.C., 2020. Filamentous fungal biomats, methods of their
   production and methods of their use. US10787638B2.
- Kromus, S., Kamm, B., Kamm, M., Fowler, P., Narodoslawsky, M., 2008. Green Biorefineries: The
   Green Biorefinery Concept Fundamentals and Potential, in: Biorefineries-Industrial
   Processes and Products: Status Quo and Future Directions. John Wiley & Sons, Ltd, pp. 253–
   294. https://doi.org/10.1002/9783527619849.ch12
- Kröncke, N., Baur, A., Böschen, V., Demtröder, S., Benning, R., Delgado, A., 2020. Automation of
   Insect Mass Rearing and Processing Technologies of Mealworms (Tenebrio molitor), in: Adam
   Mariod, A. (Ed.), African Edible Insects As Alternative Source of Food, Oil, Protein and
   Bioactive Components. Springer International Publishing, Cham, pp. 123–139.
- 1774 https://doi.org/10.1007/978-3-030-32952-5\_8

- 1775 Kuyper, T., van Peer, A., Baars, J., 2021. Coprophilous fungi : Closing the loop: improving circularity
   1776 with manure-loving mushrooms (No. Report WPR-2021-1). Wageningen.
   1777 https://doi.org/10.18174/539315
- 1778 Kwan, T.H., Ong, K.L., Haque, M.A., Kulkarni, S., Lin, C.S.K., 2019. Biorefinery of food and beverage
   1779 waste valorisation for sugar syrups production: Techno-economic assessment. Process Saf.
   1780 Environ. Prot. 121, 194–208. https://doi.org/10.1016/j.psep.2018.10.018
- 1781 Kwan, T.H., Ong, K.L., Haque, M.A., Kwan, W.H., Kulkarni, S., Lin, C.S.K., 2018. Valorisation of food
  1782 and beverage waste via saccharification for sugars recovery. Bioresour. Technol. 255, 67–75.
  1783 https://doi.org/10.1016/j.biortech.2018.01.077
- La Boîte à Champignons, 2020. Comment faire pousser des pleurotes avec le mycélium 5L ? [WWW
   Document]. URL
- 1786https://laboiteachampignonshelp.zendesk.com/hc/fr/articles/360016484453 (accessed178710.22.20).
- 1788Lähteenmäki-Uutela, A., Rahikainen, M., Lonkila, A., Yang, B., 2021. Alternative proteins and EU food1789law. Food Control 130, 108336. https://doi.org/10.1016/j.foodcont.2021.108336
- Lakra, R., Choudhury, S., Basu, S., 2021. Recovery of protein and carbohydrate from dairy wastewater
   using ultrafiltration and forward osmosis processes. Mater. Today.
   https://doi.org/10.1016/j.matpr.2021.02.702
- Lamsal, B.P., Wang, H., Pinsirodom, P., Dossey, A.T., 2019. Applications of Insect-Derived Protein
  Ingredients in Food and Feed Industry. J. Am. Oil Chem. Soc. 96, 105–123.
  https://doi.org/10.1002/aocs.12180
- Laser, M., Jin, H., Jayawardhana, K., Dale, B.E., Lynd, L.R., 2009. Projected mature technology
  scenarios for conversion of cellulosic biomass to ethanol with coproduction thermochemical
  fuels, power, and/or animal feed protein. Biofuels, Bioprod. Bioref. 3, 231–246.
  https://doi.org/10.1002/bbb.131
- 1800 Leduc, A., 2018. Développement d'hydrolysats destinés à la formulation d'aliments pour
   1801 l'aquaculture : normalisation structurale et optimisation fonctionnelle (Alimentation et
   1802 Nutrition). Normandie Université.
- Leipold, S., Weldner, K., Hohl, M., 2021. Do we need a 'circular society'? Competing narratives of the
   circular economy in the French food sector. Ecol. Econ. 187, 107086.
   https://doi.org/10.1016/j.ecolecon.2021.107086
- Leonard, W., Zhang, P., Ying, D., Adhikari, B., Fang, Z., 2021. Fermentation transforms the phenolic
   profiles and bioactivities of plant-based foods. Biotechnol. Adv. 49, 107763.
   https://doi.org/10.1016/j.biotechadv.2021.107763
- Li, H., Zhong, Y., Lu, Q., Zhang, X., Wang, Q., Liu, Huifan, Diao, Z., Yao, C., Liu, Hui, 2019. Co-cultivation
   of Rhodotorula glutinis and Chlorella pyrenoidosa to improve nutrient removal and protein
   content by their synergistic relationship. RSC Advances 9, 14331–14342.
   https://doi.org/10.1039/C9RA01884K
- Liang, X., Zhao, Y., Si, H., Hua, D., Dong, T., Liao, W., Zhang, B., Liu, S., 2020. Optimization of Protein
   Extraction from the Low Organic Residual Municipal Sludge Waste. IOP Conf. Ser.: Earth
   Environ. Sci. 450, 012048. https://doi.org/10.1088/1755-1315/450/1/012048
- 1816 Linder, T., 2019. Making the case for edible microorganisms as an integral part of a more sustainable
  1817 and resilient food production system. Food Sec. 11, 265–278.
  1818 https://doi.org/10.1007/s12571-019-00912-3
- Liu, C., Colón, B.C., Ziesack, M., Silver, P.A., Nocera, D.G., 2016. Water splitting-biosynthetic system
   with CO2 reduction efficiencies exceeding photosynthesis. Science 352, 1210–1213.
   https://doi.org/10.1126/science.aaf5039
- Logan, A.J., Terry, S.S., Swenson, R., 2011. Wastewater treatment method and apparatus, biosolids based food additive, and business application. US7931806B2.
- Lu, H., Zhang, G., He, S., Zhao, R., Zhu, D., 2021. Purple non-sulfur bacteria technology: a promising
   and potential approach for wastewater treatment and bioresources recovery. World J.
   Microbiol. Biotechnol. 37, 161. https://doi.org/10.1007/s11274-021-03133-z

- Lucci, G.M., Henchion, M.M., Lange, L., Ledgard, S.F., Collie, S.R., Cosgrove, G.P., Meyer, A.S.,
   Graichen, F.H.M., Barth, S., Lenehan, J.J., 2019. Beyond ruminants: discussing opportunities
   for alternative pasture uses in New Zealand. J. New Zeal. Grass 217–222.
   https://doi.org/10.33584/jnzg.2019.81.401
- Luyckx, K., Bowman, M., Woroniecka, K., Taillard, D., Broeze, J., 2019. Technical Guidelines Animal
   Feed Final The safety, environmental and economic aspects of feeding treated surplus food
   to omnivorous livestock (No. 6.7), Refresh Deliverable. Brussels.
- 1834 Luzixine, 2020. Luzixine, a concentrate of benefits [WWW Document]. URL
- 1835 https://luzixine.fr/en/homepage/ (accessed 12.4.20).
- 1836 Lv, X., Wu, Y., Gong, M., Deng, J., Gu, Y., Liu, Y., Li, J., Du, G., Ledesma-Amaro, R., Liu, L., Chen, J.,
   2021. Synthetic biology for future food: research progress and future directions. Future
   1838 Foods 3, 100025. https://doi.org/10.1016/j.fufo.2021.100025
- Lyu, F., Luiz, S.F., Azeredo, D.R.P., Cruz, A.G., Ajlouni, S., Ranadheera, C.S., 2020. Apple Pomace as a
   Functional and Healthy Ingredient in Food Products: A Review. Processes 8, 319.
   https://doi.org/10.3390/pr8030319
- Maity, S.K., 2015. Opportunities, recent trends and challenges of integrated biorefinery: Part I.
   Renew. Sust. Energ. Rev. 43, 1427–1445. https://doi.org/10.1016/j.rser.2014.11.092
- Makkar, H.P.S., 1993. Antinutritional factors in foods for livestock. BSAP Occasional Publication 16,
   69–85. https://doi.org/10.1017/S0263967X00031086
- Makkar, H.P.S., Addonizio, E., Gizachew, L., 2018. Emergency animal feed and feed strategies for dry
   areas. Broadening Horizons 49.
- Makkar, H.P.S., Tran, G., Heuzé, V., Ankers, P., 2014. State-of-the-art on use of insects as animal feed.
   Anim. Feed Sci. Technol. 197, 1–33. https://doi.org/10.1016/j.anifeedsci.2014.07.008
- Marchão, L., da Silva, T.L., Gouveia, L., Reis, A., 2018. Microalgae-mediated brewery wastewater
   treatment: effect of dilution rate on nutrient removal rates, biomass biochemical
   composition, and cell physiology. J. Appl. Phycol. 30, 1583–1595.
   https://doi.org/10.1007/s10811-017-1374-1
- 1854 Markham, W.M., Reid, J.H., 1988. Conversion of biological sludge and primary float sludge to animal 1855 protein supplement. US4728517A.
- Martin, A.H., Castellani, O., Jong, G.A. de, Bovetto, L., Schmitt, C., 2019. Comparison of the functional properties of RuBisCO protein isolate extracted from sugar beet leaves with commercial whey protein and soy protein isolates. J. Sci. Food Agric. 99, 1568–1576.
   https://doi.org/10.1002/jsfa.9335
- Martínez-Alvarez, O., Chamorro, S., Brenes, A., 2015. Protein hydrolysates from animal processing
   by-products as a source of bioactive molecules with interest in animal feeding: A review.
   Food Res. Int. 73, 204–212. https://doi.org/10.1016/j.foodres.2015.04.005
- Matassa, S., Boon, N., Verstraete, W., 2015. Resource recovery from used water: The manufacturing
  abilities of hydrogen-oxidizing bacteria. Water Res. 68, 467–478.
  https://doi.org/10.1016/j.watres.2014.10.028
- Matassa, S., Papirio, S., Pikaar, I., Hülsen, T., Leijenhorst, E., Esposito, G., Pirozzi, F., Verstraete, W.,
  2020. Upcycling of biowaste carbon and nutrients in line with consumer confidence: the "full
  gas" route to single cell protein. Green Chem. 22, 4912–4929.
  https://doi.org/10.1039/D0GC01382J
- Matassa, S., Verstraete, W., Pikaar, I., Boon, N., 2016. Autotrophic nitrogen assimilation and carbon
   capture for microbial protein production by a novel enrichment of hydrogen-oxidizing
   bacteria. Water Res. 101, 137–146. https://doi.org/10.1016/j.watres.2016.05.077
- 1873 McHugh, T.H., Bustillos, R.D.A., Olson, D.A., Pan, Z., Kurzrock, D.J., Schwartz, J.L., 2020. Intermittent 1874 infrared drying for brewery-spent grain. US10578358B2.
- 1875 Meeker, D.L. (Ed.), 2006. Essential rendering: all about the animal by-products industry. National
   1876 Renderers Association : Fats and Proteins Research Foundation : Animal Protein Producers
   1877 Industry, Alexandria, Va.

- 1878 Melgar-Lalanne, G., Hernández-Álvarez, A.-J., Salinas-Castro, A., 2019. Edible Insects Processing:
   1879 Traditional and Innovative Technologies. Compr. Rev. Food Sci. Food Saf. 18, 1166–1191.
   1880 https://doi.org/10.1111/1541-4337.12463
- 1881 Mhlongo, G., Mnisi, C.M., Mlambo, V., 2021. Cultivating oyster mushrooms on red grape pomace
   1882 waste enhances potential nutritional value of the spent substrate for ruminants. PLoS One
   1883 16, e0246992. https://doi.org/10.1371/journal.pone.0246992
- 1884 MiAlgae, 2020. Nexte Generation Agriculture [WWW Document]. URL https://www.mialgae.com/
   1885 (accessed 10.22.20).
- 1886 Microterra, 2020. Affordable, scalable and sustainable plant-based protein [WWW Document]. URL
   1887 https://www.microterra.com.mx (accessed 10.22.20).
- Milhazes-Cunha, H., Otero, A., 2017. Valorisation of aquaculture effluents with microalgae: The
   Integrated Multi-Trophic Aquaculture concept. Algal Res. 24, 416–424.
   https://doi.org/10.1016/j.algal.2016.12.011
- Mishra, A., Ntihuga, J.N., Molitor, B., Angenent, L.T., 2020. Power-to-Protein: Carbon Fixation with
   Renewable Electric Power to Feed the World. Joule 4, 1142–1147.
   https://doi.org/10.1016/j.joule.2020.04.008
- Mokomele, T., Sousa, L. da C., Bals, B., Balan, V., Goosen, N., Dale, B.E., Görgens, J.F., 2018. Using
   steam explosion or AFEX<sup>™</sup> to produce animal feeds and biofuel feedstocks in a biorefinery
   based on sugarcane residues. Biofuels, Bioprod. Bioref. 12, 978–996.
   https://doi.org/10.1002/bbb.1927
- Molitor, B., Mishra, A., Angenent, L.T., 2019. Power-to-protein: converting renewable electric power
   and carbon dioxide into single cell protein with a two-stage bioprocess. Energy Environ. Sci.
   12, 3515–3521. https://doi.org/10.1039/C9EE02381J
- Moore, K.J., Jung, H.J.G., 2001. Lignin and fiber digestion. Rangeland Ecol. Manag. 54, 420–430.
   https://doi.org/10.2307/4003113
- Mottet, A., de Haan, C., Falcucci, A., Tempio, G., Opio, C., Gerber, P., 2017. Livestock: On our plates or
   eating at our table? A new analysis of the feed/food debate. Glob. Food Sec., Food Security
   Governance in Latin America 14, 1–8. https://doi.org/10.1016/j.gfs.2017.01.001
- Mourad, M., 2016. Recycling, recovering and preventing "food waste": competing solutions for food
   systems sustainability in the United States and France. J. Clean. Prod. 126, 461–477.
   https://doi.org/10.1016/j.jclepro.2016.03.084
- Mueller, Z.O., 1980. Feed from animal wastes: State of knowledge, FAO Animal Production and
   Health Papers. FAO, Rome.
- Muniz, C.E.S., Santiago, Â.M., Gusmão, T.A.S., Oliveira, H.M.L., Conrado, L. de S., Gusmão, R.P. de,
   2020. Solid-state fermentation for single-cell protein enrichment of guava and cashew by products and inclusion on cereal bars. Biocatal. Agric. Biotechnol. 25, 101576.
   https://doi.org/10.1016/j.bcab.2020.101576
- Mupondwa, E., Li, X., Wanasundara, J.P.D., 2018. Technoeconomic Prospects for Commercialization
   of Brassica (Cruciferous) Plant Proteins. Journal of the American Oil Chemists' Society 95,
   903–922. https://doi.org/10.1002/aocs.12057
- 1918Muscat, A., de Olde, E.M., Ripoll-Bosch, R., Van Zanten, H.H.E., Metze, T.A.P., Termeer, C.J.A.M., van1919Ittersum, M.K., de Boer, I.J.M., 2021. Principles, drivers and opportunities of a circular1920bioeconomy. Nat. Food 2, 561–566. https://doi.org/10.1038/s43016-021-00340-7
- Muys, M., Papini, G., Spiller, M., Sakarika, M., Schwaiger, B., Lesueur, C., Vermeir, P., Vlaeminck, S.E.,
   2020. Dried aerobic heterotrophic bacteria from treatment of food and beverage effluents:
   Screening of correlations between operation parameters and microbial protein quality.
   Bioresour. Technol. 307, 123242. https://doi.org/10.1016/j.biortech.2020.123242
- Nangle, S.N., Sakimoto, K.K., Silver, P.A., Nocera, D.G., 2017. Biological-inorganic hybrid systems as a
   generalized platform for chemical production. Curr. Opin. Chem. Biol. 41, 107–113.
   https://doi.org/10.1016/j.cbpa.2017.10.023

- 1928 Nocente, F., Taddei, F., Galassi, E., Gazza, L., 2019. Upcycling of brewers' spent grain by production of 1929 dry pasta with higher nutritional potential. LWT 114, 108421. 1930 https://doi.org/10.1016/j.lwt.2019.108421
- 1931 NovoNutrients, 2020. Transforming industrial waste carbon dioxide emissions into food and feed 1932 ingredients, through industrial biotech [WWW Document]. URL
- 1933 https://www.novonutrients.com/ (accessed 10.23.20).
- 1934 Nunes, C.S., 2018. General perspectives of enzymes, environment preservation, and scarce natural resources—conclusions, in: Enzymes in Human and Animal Nutrition. Elsevier, pp. 515–526. 1935 1936 https://doi.org/10.1016/B978-0-12-805419-2.00027-7
- 1937 Ochsenreither, K., Glück, C., Stressler, T., Fischer, L., Syldatk, C., 2016. Production Strategies and 1938 Applications of Microbial Single Cell Oils. Front. Microbiol. 7, 1539. 1939
  - https://doi.org/10.3389/fmicb.2016.01539
- 1940 Ojha, S., Bußler, S., Schlüter, O.K., 2020. Food waste valorisation and circular economy concepts in 1941 insect production and processing. Waste Manag. 118, 600–609. 1942 https://doi.org/10.1016/j.wasman.2020.09.010
- 1943 Oonincx, D.G.A.B., Broekhoven, S. van, Huis, A. van, Loon, J.J.A. van, 2015. Feed Conversion, Survival 1944 and Development, and Composition of Four Insect Species on Diets Composed of Food By-1945 Products. PloS One 10, e0144601. https://doi.org/10.1371/journal.pone.0144601
- 1946 Orsi, L., Voege, L.L., Stranieri, S., 2019. Eating edible insects as sustainable food? Exploring the 1947 determinants of consumer acceptance in Germany. Food Res. Int. 125, 108573. 1948 https://doi.org/10.1016/j.foodres.2019.108573
- 1949 Osman, N.S., Sapawe, N., Sapuan, M.A., Fozi, M.F.M., Azman, M.H.I.F., Fazry, A.H.Z., Zainudin, M.Z.H., 1950 Hanafi, M.F., 2018. Sunflower shell waste as an alternative animal feed. Mater. Today. 1951 https://doi.org/10.1016/j.matpr.2018.07.049
- 1952 Øverland, M., Borge, G.I., Vogt, G., Schøyen, H.F., Skrede, A., 2011. Oxidative stability and sensory 1953 quality of meat from broiler chickens fed a bacterial meal produced on natural gas. Poult. Sci. 1954 90, 201-210. https://doi.org/10.3382/ps.2010-00784
- Øverland, M., Mydland, L.T., Skrede, A., 2019. Marine macroalgae as sources of protein and bioactive 1955 1956 compounds in feed for monogastric animals. J. Sci. Food Agric. https://doi.org/10.1002/jsfa.9143 1957
- Øverland, M., Tauson, A.-H., Shearer, K., Skrede, A., 2010. Evaluation of methane-utilising bacteria 1958 1959 products as feed ingredients for monogastric animals. Arch. Anim. Nutr. 64, 171–89. 1960 https://doi.org/10.1080/17450391003691534
- 1961 Paës, G., Navarro, D., Benoit, Y., Blanquet, S., Chabbert, B., Chaussepied, B., Coutinho, P.M., Durand, 1962 S., Grigoriev, I.V., Haon, M., Heux, L., Launay, C., Margeot, A., Nishiyama, Y., Raouche, S., 1963 Rosso, M.-N., Bonnin, E., Berrin, J.-G., 2019. Tracking of enzymatic biomass deconstruction by 1964 fungal secretomes highlights markers of lignocellulose recalcitrance. Biotechnol. Biofuels 12. 1965 https://doi.org/10.1186/s13068-019-1417-8
- Pander, B., Mortimer, Z., Woods, C., McGregor, C., Dempster, A., Thomas, L., Maliepaard, J., 1966 1967 Mansfield, R., Rowe, P., Krabben, P., 2020. Hydrogen oxidising bacteria for production of 1968 single-cell protein and other food and feed ingredients. Eng. Biol. 4, 21–24. 1969 https://doi.org/10.1049/enb.2020.0005
- Parajuli, R., Dalgaard, T., Birkved, M., 2018. Can farmers mitigate environmental impacts through 1970 1971 combined production of food, fuel and feed? A consequential life cycle assessment of 1972 integrated mixed crop-livestock system with a green biorefinery. Sci. Total Environ. 619–620, 1973 127–143. https://doi.org/10.1016/j.scitotenv.2017.11.082
- 1974 Parodi, A., De Boer, I.J.M., Gerrits, W.J.J., Van Loon, J.J.A., Heetkamp, M.J.W., Van Schelt, J., Elizabeth 1975 Bolhuis, J., Van Zanten, H.H.E., 2020. Bioconversion efficiencies, greenhouse gas and 1976 ammonia emissions during black soldier fly rearing – A mass balance approach. J. Clean. 1977 Prod. 271, 122488. https://doi.org/10.1016/j.jclepro.2020.122488
- 1978 Parodi, A., Leip, A., De Boer, I.J.M., Slegers, P.M., Ziegler, F., Temme, E.H.M., Herrero, M., Tuomisto, 1979 H., Valin, H., Van Middelaar, C.E., Van Loon, J.J.A., Van Zanten, H.H.E., 2018. The potential of

1980 future foods for sustainable and healthy diets. Nat. Sustain. 1, 782–789. 1981 https://doi.org/10.1038/s41893-018-0189-7 1982 Patil, R.H., Patil, M.P., Maheshwari, V.L., 2020. Microbial transformation of crop residues into a 1983 nutritionally enriched substrate and its potential application in livestock feed. SN Appl. Sci. 2, 1984 1140. https://doi.org/10.1007/s42452-020-2949-z 1985 Patsios, S.I., Dedousi, A., Sossidou, E.N., Zdragas, A., 2020. Sustainable Animal Feed Protein through 1986 the Cultivation of Yarrowia Lipolytica on Agro-Industrial Wastes and by-Products. 1987 Sustainability. https://doi.org/10.3390/su12041398 Paul, S., Charles, G., Mcavoy, D.J., 1962. Process for recovering fats and meat and bone scrap from 1988 1989 inedible slaughterhouse materials. US3046286A. 1990 Pehme, S., Veromann, E., Hamelin, L., 2017. Environmental performance of manure co-digestion with 1991 natural and cultivated grass – A consequential life cycle assessment. J. Clean. Prod. 162, 1992 1135–1143. https://doi.org/10.1016/j.jclepro.2017.06.067 1993 Perța-Crișan, S., Ursachi, C. Ștefan, Gavrilaș, S., Oancea, F., Munteanu, F.-D., 2021. Closing the Loop 1994 with Keratin-Rich Fibrous Materials. Polymers 13, 1896. 1995 https://doi.org/10.3390/polym13111896 1996 Peyraud, J.L., Aubin, J., Barbier, M., Baumont, R., Berri, C., Bidanel, J.P., Citti, Ch., Cotinot, C., Ducrot, 1997 C., Dupraz, P., Faverdin, P., Friggens, N., Houot, S., Nozières-Petit, M.O., Rogel-Gaillard, C., 1998 Santé-Lhoutellier, V., 2020. Réflexion prospective interdisciplinaire science pour les élevages 1999 de demain. Rapport de synthèse. INRAE 53. https://doi.org/10.15454/X83C-0674 2000 Pfluger, A.R., Wu, W.-M., Pieja, A.J., Wan, J., Rostkowski, K.H., Criddle, C.S., 2011. Selection of Type I 2001 and Type II methanotrophic proteobacteria in a fluidized bed reactor under non-sterile 2002 conditions. Bioresour. Technol. 102, 9919–9926. 2003 https://doi.org/10.1016/j.biortech.2011.08.054 2004 Phan Van PhI, C., Walraven, M., Bézagu, M., Lefranc, M., Ray, C., 2020. Industrial Symbiosis in Insect 2005 Production—A Sustainable Eco-Efficient and Circular Business Model. Sustainability 12, 2006 10333. https://doi.org/10.3390/su122410333 2007 Pikaar, I., Matassa, S., Bodirsky, B.L., Weindl, I., Humpenöder, F., Rabaey, K., Boon, N., Bruschi, M., 2008 Yuan, Z., van Zanten, H., Herrero, M., Verstraete, W., Popp, A., 2018. Decoupling Livestock 2009 from Land Use through Industrial Feed Production Pathways. Environ. Sci. Technol. 52, 7351-2010 7359. https://doi.org/10.1021/acs.est.8b00216 Pikaar, I., Matassa, S., Rabaey, K., Bodirsky, B., Popp, A., Herrero, M., Verstraete, W., 2017. Microbes 2011 2012 and the Next Nitrogen Revolution. Environ. Sci. Technol. 51. 2013 https://doi.org/10.1021/acs.est.7b00916 2014 Pinotti, L., Luciano, A., Ottoboni, M., Manoni, M., Ferrari, L., Marchis, D., Tretola, M., 2021. Recycling 2015 food leftovers in feed as opportunity to increase the sustainability of livestock production. J. 2016 Clean. Prod. 294, 126290. https://doi.org/10.1016/j.jclepro.2021.126290 2017 Pippinato, L., Gasco, L., Di Vita, G., Mancuso, T., 2020. Current scenario in the European edible-insect 2018 industry: a preliminary study. J. Insects as Food Feed. 6, 371–381. 2019 https://doi.org/10.3920/JIFF2020.0008 2020 Pirie, N.W., 1971. Leaf protein: its agronomy, preparation, quality and use, IBP Handbook. 2021 International Biological Programme. 2022 Pitkänen, J.-P., 2020. Bioreactors for growing micro-organisms. FI128391B. 2023 Pleissner, D., Kwan, T.H., Lin, C.S.K., 2014. Fungal hydrolysis in submerged fermentation for food 2024 waste treatment and fermentation feedstock preparation. Bioresour. Technol. 158, 48–54. 2025 https://doi.org/10.1016/j.biortech.2014.01.139 2026 Pleissner, D., Smetana, S., 2020. Estimation of the economy of heterotrophic microalgae- and insect-2027 based food waste utilization processes. Waste Manag. 102, 198–203. 2028 https://doi.org/10.1016/j.wasman.2019.10.031 2029 Pojić, M., Mišan, A., Tiwari, B., 2018. Eco-innovative technologies for extraction of proteins for 2030 human consumption from renewable protein sources of plant origin. Trends Food Sci. 2031 Technol. 75, 93-104. https://doi.org/10.1016/j.tifs.2018.03.010

- Poletto, P., Pereira, G.N., Monteiro, C.R.M., Pereira, M.A.F., Bordignon, S.E., de Oliveira, D., 2020.
   Xylooligosaccharides: Transforming the lignocellulosic biomasses into valuable 5-carbon
   sugar prebiotics. Process Biochem. 91, 352–363.
- 2035 https://doi.org/10.1016/j.procbio.2020.01.005
- Poortvliet, P.M., Van der Pas, L., Mulder, B.C., Fogliano, V., 2019. Healthy, but Disgusting: An
   Investigation Into Consumers' Willingness to Try Insect Meat. J. Econ. Entomol. 112, 1005–
   1010. https://doi.org/10.1093/jee/toz043
- Post, M.J., Levenberg, S., Kaplan, D.L., Genovese, N., Fu, J., Bryant, C.J., Negowetti, N., Verzijden, K.,
   Moutsatsou, P., 2020. Scientific, sustainability and regulatory challenges of cultured meat.
   Nat. Food 1, 403–415. https://doi.org/10.1038/s43016-020-0112-z
- Power-to-Protein, 2020. Closing the artificial nitrogen cycle by direct N upcycling as microbial protein
   (WWW Document]. URL https://www.powertoprotein.eu/ (accessed 10.23.20).
- Prajapati, S., Koirala, S., Anal, A.K., 2021. Bioutilization of Chicken Feather Waste by Newly Isolated
   Keratinolytic Bacteria and Conversion into Protein Hydrolysates with Improved
   Functionalities. Appl. Biochem. Biotechnol. https://doi.org/10.1007/s12010-021-03554-4
- Pretty, J., 2018. Intensification for redesigned and sustainable agricultural systems. Science 362,
   eaav0294. https://doi.org/10.1126/science.aav0294
- 2049 Prolupin GmbH, 2020. Production process [WWW Document]. URL
- 2050 https://www.prolupin.com/production-process.html (accessed 12.3.20).
- Rad, M.I., Rouzbehan, Y., Rezaei, J., 2016. Effect of dietary replacement of alfalfa with urea-treated
   almond hulls on intake, growth, digestibility, microbial nitrogen, nitrogen retention, ruminal
   fermentation, and blood parameters in fattening lambs. J. Anim. Sci. 94, 349–358.
   https://doi.org/10.2527/jas.2015-9437
- Rajeh, C., Saoud, I.P., Kharroubi, S., Naalbandian, S., Abiad, M.G., 2021. Food loss and food waste
   recovery as animal feed: a systematic review. J Mater Cycles Waste Manag 23, 1–17.
   https://doi.org/10.1007/s10163-020-01102-6
- Rajesh, N., Imelda-Joseph, Paul Raj, R., 2010. Value addition of vegetable wastes by solid-state
   fermentation using Aspergillus niger for use in aquafeed industry. Waste Manag. 30, 2223–
   2227. https://doi.org/10.1016/j.wasman.2009.12.017
- Raksasat, R., Lim, J.W., Kiatkittipong, W., Kiatkittipong, K., Ho, Y.C., Lam, M.K., Font-Palma, C., Mohd
   Zaid, H.F., Cheng, C.K., 2020. A review of organic waste enrichment for inducing palatability
   of black soldier fly larvae: Wastes to valuable resources. Environ. Pollut. 267, 115488.
   https://doi.org/10.1016/j.envpol.2020.115488
- Ramankutty, N., Mehrabi, Z., Waha, K., Jarvis, L., Kremen, C., Herrero, M., Rieseberg, L.H., 2018.
   Trends in Global Agricultural Land Use: Implications for Environmental Health and Food
   Security. Annu. Rev. Plant Biol. 69, 789–815. https://doi.org/10.1146/annurev-arplant 042817-040256
- Rasouli, Z., Valverde-Pérez, B., D'Este, M., De Francisci, D., Angelidaki, I., 2018. Nutrient recovery
   from industrial wastewater as single cell protein by a co-culture of green microalgae and
   methanotrophs. Biochem. Eng. J. 134, 129–135. https://doi.org/10.1016/j.bej.2018.03.010
- Ravi, H.K., Degrou, A., Costil, J., Trespeuch, C., Chemat, F., Vian, M.A., 2020. Larvae Mediated
   Valorization of Industrial, Agriculture and Food Wastes: Biorefinery Concept through
   Bioconversion, Processes, Procedures, and Products. Processes 8, 857.
   https://doi.org/10.3390/pr8070857
- 2076 Redlingshöfer, B., Barles, S., Weisz, H., 2020. Are waste hierarchies effective in reducing
   2077 environmental impacts from food waste? A systematic review for OECD countries. Resourc.
   2078 Conserv. Recy. 156, 104723. https://doi.org/10.1016/j.resconrec.2020.104723
- 2079 Reed, J.S., 2019. Biological and Chemical Process Utilizing Chemoautotrophic Microorganisms for the
   2080 Chemosynthetic Fixation of Carbon Dioxide and/or Other Inorganic Carbon Sources into
   2081 Organic Compounds and the Generation of Additional Useful Products. US20190382808A1.

- 2082 Reese, E.T., Mandels, M., Weiss, Alvin.H., 1972. Cellulose as a novel energy source, in: Advances in
   2083 Biochemical Engineering. Springer-Verlag, Berlin/Heidelberg, pp. 181–200.
   2084 https://doi.org/10.1007/BFb0006669
- 2085ReGrained, 2020. ReGrained [WWW Document]. URL https://www.regrained.com/ (accessed208610.21.20).
- 2087 Research and Innovation, 2017. Innovation and exploration through cutting-edge microbiome
   2088 research, European Union. ed, Publications about Food 2030. Publications Office of the
   2089 European Union, Luxembourg.
- 2090Ritota, M., Manzi, P., 2019. Pleurotus spp. Cultivation on Different Agri-Food By-Products: Example of2091Biotechnological Application. Sustainability 11, 5049. https://doi.org/10.3390/su11185049
- 2092 Rockström, J., Williams, J., Daily, G., Noble, A., Matthews, N., Gordon, L., Wetterstrand, H., DeClerck,
  2093 F., Shah, M., Steduto, P., de Fraiture, C., Hatibu, N., Unver, O., Bird, J., Sibanda, L., Smith, J.,
  2094 2017. Sustainable intensification of agriculture for human prosperity and global
  2095 sustainability. Ambio 46, 4–17. https://doi.org/10.1007/s13280-016-0793-6
- 2096 Rodríguez García, S.L., Raghavan, V., 2021. Green extraction techniques from fruit and vegetable
   2097 waste to obtain bioactive compounds—A review. Crit. Rev. Food. Sci. Nutr.
   2098 https://doi.org/10.1080/10408398.2021.1901651
- Rodriguez-Lopez, A.D., Melgar, B., Conidi, C., Barros, L., Ferreira, I.C.F.R., Cassano, A., Garcia-Castello,
   E.M., 2020. Food industry by-products valorization and new ingredients, in: Sustainability of
   the Food System. Elsevier, pp. 71–99. https://doi.org/10.1016/B978-0-12-818293-2.00005-7
- Rolston, D., Mathan, V., 1989. Xylose transport in the human jejunum. Dig. Dis. Sci. 34, 553–558.
   https://doi.org/10.1007/bf01536332
- Romantschuk, H., 1975. Pekilo process: protein from spent sulfite liquor, in: Single Cell Protein II.
   Presented at the International Conference on Single Cell Protein.
- Rosas Romero, A., Diaz, A.C., 1983. Composition of plantain leaves (musa paradisiaca L., subsp.
   normals O. kze). A possible source for leaf protein concentrate. Acta cient. venez 72–3.
- Rosenzweig, C., Mbow, C., Barioni, L.G., Benton, T.G., Herrero, M., Krishnapillai, M., Liwenga, E.T.,
  Pradhan, P., Rivera-Ferre, M.G., Sapkota, T., Tubiello, F.N., Xu, Y., Mencos Contreras, E.,
  Portugal-Pereira, J., 2020. Climate change responses benefit from a global food system
  approach. Nat. Food 1, 94–97. https://doi.org/10.1038/s43016-020-0031-z
- Roth, J.C.G., Hoeltz, M., Benitez, L.B., 2020. Current approaches and trends in the production of
   microbial cellulases using residual lignocellulosic biomass: a bibliometric analysis of the last
   10 years. Arch. Microbiol. 202, 935–951. https://doi.org/10.1007/s00203-019-01796-9
- Roth, M., Jekle, M., Becker, T., 2019. Opportunities for upcycling cereal byproducts with special focus
  on Distiller's grains. Trends Food Sci. Technol. 91, 282–293.
  https://doi.org/10.1016/j.tifs.2019.07.041
- Roulleau, J., 2020. Les Cycles De Culture Du Champignon De Paris [WWW Document]. La Cave vivante
   du champignon Le Puy-Notre-Dame (49). URL https://www.lechampignon.com/LES-CYCLES DE-CULTURE-DU-CHAMPIGNON-DE-PARIS\_a24.html (accessed 10.22.20).
- Rumokoy, L., Adiani, S., Kaunang, C., Kiroh, H., Untu, I., Toar, W.L., 2019. The Wisdom of Using Insects
   as Animal Feed on Decreasing Competition with Human Food. Anim. Sci. 62, 51–56.
- Rumpold, B.A., Langen, N., 2020. Consumer acceptance of edible insects in an organic waste-based
   bioeconomy. Curr. Opin. Green Sustain. Chem. 23, 80–84.
   https://doi.org/10.1016/j.cogsc.2020.03.007
- Russo, G.L., Langellotti, A.L., Oliviero, M., Sacchi, R., Masi, P., 2021. Sustainable production of food
   grade omega-3 oil using aquatic protists: reliability and future horizons. New Biotechnol. 62,
   32–39. https://doi.org/10.1016/j.nbt.2021.01.006
- Sabater, C., Ruiz, L., Delgado, S., Ruas-Madiedo, P., Margolles, A., 2020. Valorization of Vegetable
   Food Waste and By-Products Through Fermentation Processes. Front. Microbiol. 11, 2604.
   https://doi.org/10.3389/fmicb.2020.581997

- Saha, A., Basak, B.B., 2020. Scope of value addition and utilization of residual biomass from medicinal
  and aromatic plants. Ind. Crops Prod. 145, 111979.
  https://doi.org/10.1016/j.indcrop.2019.111979
- Said, M.I., 2019. Characteristics of by-product and animal waste: A Review. Large Anim. Rev. 25, 243–
  250.
- Sakarika, M., Candry, P., Depoortere, M., Ganigué, R., Rabaey, K., 2020. Impact of substrate and
   growth conditions on microbial protein production and composition. Bioresour. Technol.
   317, 124021. https://doi.org/10.1016/j.biortech.2020.124021
- Salami, S.A., Luciano, G., O'Grady, M.N., Biondi, L., Newbold, C.J., Kerry, J.P., Priolo, A., 2019.
  Sustainability of feeding plant by-products: A review of the implications for ruminant meat
  production. Anim. Feed Sci. Technol. 251, 37–55.
- 2143 https://doi.org/10.1016/j.anifeedsci.2019.02.006
- San Martin, D., Ramos, S., Zufía, J., 2016. Valorisation of food waste to produce new raw materials
   for animal feed. Food Chem. 198, 68–74. https://doi.org/10.1016/j.foodchem.2015.11.035
- Santamaría-Fernández, M., Lübeck, M., 2020. Production of leaf protein concentrates in green
   biorefineries as alternative feed for monogastric animals. Anim. Feed Sci. Technol. 268,
   114605. https://doi.org/10.1016/j.anifeedsci.2020.114605
- Santamaría-Fernández, M., Ytting, N.K., Lübeck, M., Uellendahl, H., 2020. Potential Nutrient Recovery
   in a Green Biorefinery for Production of Feed, Fuel and Fertilizer for Organic Farming. Waste
   Biomass Valor. 11, 5901–5911. https://doi.org/10.1007/s12649-019-00842-3
- Sari, Y.W., Mulder, W.J., Sanders, J.P.M., Bruins, M.E., 2015a. Towards plant protein refinery: Review
   on protein extraction using alkali and potential enzymatic assistance. Biotechnol. J. 10, 1138–
   1157. https://doi.org/10.1002/biot.201400569
- Sari, Y.W., Syafitri, U., Sanders, J.P.M., Bruins, M.E., 2015b. How biomass composition determines
   protein extractability. Ind. Crops Prod. 70, 125–133.
   https://doi.org/10.1016/j.indcrop.2015.03.020
- Saxe, H., Hamelin, L., Hinrichsen, T., Wenzel, H., 2018. Production of Pig Feed under Future
   Atmospheric CO2 Concentrations: Changes in Crop Content and Chemical Composition, Land
   Use, Environmental Impact, and Socio-Economic Consequences. Sustainability 10, 3184.
- 2161 https://doi.org/10.3390/su10093184
- Scheller, H.V., Ulvskov, P., 2010. Hemicelluloses. Annu. Rev. Plant Biol. 61, 263–289.
   https://doi.org/10.1146/annurev-arplant-042809-112315
- Schoumans, O.F., Bouraoui, F., Kabbe, C., Oenema, O., van Dijk, K.C., 2015. Phosphorus management
  in Europe in a changing world. Ambio 44, 180–192. https://doi.org/10.1007/s13280-0140613-9
- Schramski, J.R., Woodson, C.B., Brown, J.H., 2020. Energy use and the sustainability of intensifying
   food production. Nat. Sustain. 3, 257–259. https://doi.org/10.1038/s41893-020-0503-z
- Searchinger, T., Waite, R., Hanson, C., Ranganathan, J., Matthews, E., 2018. Creating a sustainable
   food future: A menu of solutions to feed nearly 10 billion people by 2050. Final Report.
   World Resource Institute, Washington (USA).
- Shahid, A., Malik, S., Zhu, H., Xu, J., Nawaz, M.Z., Nawaz, S., Asraful Alam, Md., Mehmood, M.A.,
  2020. Cultivating microalgae in wastewater for biomass production, pollutant removal, and
  atmospheric carbon mitigation; a review. Sci. Total Environ. 704, 135303.
  https://doi.org/10.1016/j.scitotenv.2019.135303
- Shahid, K., Srivastava, V., Sillanpää, M., 2021. Protein recovery as a resource from waste specifically
   via membrane technology—from waste to wonder. Environ. Sci. Pollut. Res. 28, 10262–
   10282. https://doi.org/10.1007/s11356-020-12290-x
- Sharma, A., Sharma, P., Singh, J., Singh, S., Nain, L., 2020. Prospecting the Potential of Agroresidues
   as Substrate for Microbial Flavor Production. Front. Sustain. Food Syst. 4.
   https://doi.org/10.3389/fsufs.2020.00018

Shepon, A., Eshel, G., Noor, E., Milo, R., 2016. Energy and protein feed-to-food conversion 2182 2183 efficiencies in the US and potential food security gains from dietary changes. Environ. Res. 2184 Lett. 11, 105002. https://doi.org/10.1088/1748-9326/11/10/105002 2185 Shojinmeat Project, 2020. Open-source cell-based meat: Cellular agriculture by citizens. URL 2186 https://shojinmeat.com/wordpress/en/ (accessed 11.20.20). 2187 Siben, C., Sportiche Maurice, Rupp-Dalhem Christophe, Lequime Catherine, Le Vely Didier, Monnet 2188 François, Schneider Didier, Sainsot Alain, 2018. Stratégie nationale bio-production en France - Comment développer la bio-production en France (No. V4-10). Conseil national de 2189 2190 l'industrie, Paris. 2191 Siegrist, M., Hartmann, C., 2020. Consumer acceptance of novel food technologies. Nat. Food 1, 343– 2192 350. https://doi.org/10.1038/s43016-020-0094-x 2193 Sillman, J., Nygren, L., Kahiluoto, H., Ruuskanen, V., Tamminen, A., Bajamundi, C., Nappa, M., 2194 Wuokko, M., Lindh, T., Vainikka, P., Pitkänen, J.-P., Ahola, J., 2019. Bacterial protein for food 2195 and feed generated via renewable energy and direct air capture of CO2: Can it reduce land 2196 and water use? Glob. Food Sec. 22, 25–32. https://doi.org/10.1016/j.gfs.2019.09.007 2197 Sillman, J., Uusitalo, V., Ruuskanen, V., Ojala, L., Kahiluoto, H., Soukka, R., Ahola, J., 2020. A life cycle 2198 environmental sustainability analysis of microbial protein production via power-to-food 2199 approaches. Int. J. Life Cycle Assess. 25, 2190–2203. https://doi.org/10.1007/s11367-020-2200 01771-3 2201 Sims, R., Bracco, S., Flammini, A., Santos, N., Pereira, L.D., Carita, A., Oze, D., 2017. Adoption of 2202 climate technologies in the agrifood sector. Methodology, Directions in Investments. FAO 2203 Investment centre, Rome. 2204 Singh, A.K., Prajapati, K.S., Shuaib, M., Kushwaha, P.P., Kumar, S., 2020. Microbial Proteins: A Potential Source of Protein, in: Egbuna, C., Dable Tupas, G. (Eds.), Functional Foods and 2205 2206 Nutraceuticals: Bioactive Components, Formulations and Innovations. Springer International 2207 Publishing, Cham, pp. 139-147. https://doi.org/10.1007/978-3-030-42319-3 8 2208 Smetana, S., Ashtari Larki, N., Pernutz, C., Franke, K., Bindrich, U., Toepfl, S., Heinz, V., 2018. 2209 Structure design of insect-based meat analogs with high-moisture extrusion. J. Food Eng. 2210 229, 83-85. https://doi.org/10.1016/j.jfoodeng.2017.06.035 2211 Smetana, S., Leonhardt, L., Kauppi, S.-M., Pajic, A., Heinz, V., 2020. Insect margarine: Processing, 2212 sustainability and design. J. Clean. Prod. 264, 121670. 2213 https://doi.org/10.1016/j.jclepro.2020.121670 2214 Smetana, S., Mathys, A., Knoch, A., Heinz, V., 2015. Meat alternatives: life cycle assessment of most 2215 known meat substitutes. Int. J. Life Cycle Assess. 20, 1254–1267. 2216 https://doi.org/10.1007/s11367-015-0931-6 2217 Smith, J., Sones, K., Grace, D., MacMillan, S., Tarawali, S., Herrero, M., 2013. Beyond milk, meat, and 2218 eggs: Role of livestock in food and nutrition security. Anim. Front. 3, 6–13. 2219 https://doi.org/10.2527/af.2013-0002 2220 Solar Foods, 2020. Food Out of Thin Air [WWW Document]. URL https://solarfoods.fi/ (accessed 2221 10.23.20). 2222 Solati, Z., Manevski, K., Jørgensen, U., Labouriau, R., Shahbazi, S., Lærke, P.E., 2018. Crude protein 2223 yield and theoretical extractable true protein of potential biorefinery feedstocks. Ind. Crops 2224 Prod. 115, 214–226. https://doi.org/10.1016/j.indcrop.2018.02.010 2225 Solovchenko, A., Lukyanov, A., Gokare Aswathanarayana, R., Pleissner, D., Ambati, R.R., 2020. Recent 2226 developments in microalgal conversion of organic-enriched waste streams. Curr. Opin. Green 2227 Sustain. Chem. 24, 61–66. https://doi.org/10.1016/j.cogsc.2020.03.006 2228 Souza Filho, P.F., 2018. Fungi-based biorefinery model for food industry waste: progress toward a 2229 circular economy. University of Boras, Boras, Sweden. 2230 Spalvins, K., Ivanovs, K., Blumberga, D., 2018. Single cell protein production from waste biomass: 2231 review of various agricultural by-products. Agric. Res. 16, 1493–1508. 2232 https://doi.org/10.15159/AR.18.129

- Specht, K., Zoll, F., Schümann, H., Bela, J., Kachel, J., Robischon, M., 2019. How Will We Eat and
  Produce in the Cities of the Future? From Edible Insects to Vertical Farming—A Study on the
  Perception and Acceptability of New Approaches. Sustainability 11, 4315.
  https://doi.org/10.3390/su11164315
- Specht, L., 2020. An analysis of culture medium costs and production volumes for cultivated meat.
   The Good Food Institute.
- Specht, L., Crosser, N., 2020. Fermentation: An Introduction to a Pillar of the Alternative Protein
   Industry, State of the Industry Report. The Good Food Institute.
- Spiller, M., Muys, M., Papini, G., Sakarika, M., Buyle, M., Vlaeminck, S.E., 2020. Environmental impact
   of microbial protein from potato wastewater as feed ingredient: Comparative consequential
   life cycle assessment of three production systems and soybean meal. Water Res. 171,
   115406. https://doi.org/10.1016/j.watres.2019.115406
- Steinberg, L.M., Kronyak, R.E., House, C.H., 2017. Coupling of anaerobic waste treatment to produce
   protein- and lipid-rich bacterial biomass. Life Sci. Space Res. 15, 32–42.
   https://doi.org/10.1016/j.lssr.2017.07.006
- Stiles, W.A.V., Styles, D., Chapman, S.P., Esteves, S., Bywater, A., Melville, L., Silkina, A., Lupatsch, I.,
   Fuentes Grünewald, C., Lovitt, R., Chaloner, T., Bull, A., Morris, C., Llewellyn, C.A., 2018. Using
   microalgae in the circular economy to valorise anaerobic digestate: challenges and
   opportunities. Bioresour. Technol. 267, 732–742.
- 2252 https://doi.org/10.1016/j.biortech.2018.07.100
- Stødkilde, L., Damborg, V.K., Jørgensen, H., Lærke, H.N., Jensen, S.K., 2019. Digestibility of
   fractionated green biomass as protein source for monogastric animals. Animal 13, 1817–
   1825. https://doi.org/10.1017/S1751731119000156
- Stoknes, K., Scholwin, F., Krzesiński, W., Wojciechowska, E., Jasińska, A., 2016. Efficiency of a novel
   "Food to waste to food" system including anaerobic digestion of food waste and cultivation
   of vegetables on digestate in a bubble-insulated greenhouse. Waste Manag. 56, 466–476.
   https://doi.org/10.1016/j.wasman.2016.06.027
- Subaşı, B.G., Vahapoğlu, B., Capanoglu, E., Mohammadifar, M.A., 2021. A review on protein extracts
   from sunflower cake: techno-functional properties and promising modification methods. Crit.
   Rev. Food. Sci. Nutr. https://doi.org/10.1080/10408398.2021.1904821
- Sun, L., Xin, F., Alper, H.S., 2021. Bio-synthesis of food additives and colorants-a growing trend in
   future food. Biotechnol. Adv. 47, 107694. https://doi.org/10.1016/j.biotechadv.2020.107694
- 2265Tacon, A.G.J., Metian, M., 2008. Global overview on the use of fish meal and fish oil in industrially2266compounded aquafeeds: Trends and future prospects. Aquaculture 285, 146–158.2267https://doi.org/10.1016/j.aquaculture.2008.08.015
- Tamayo Tenorio, A., 2017. Sugar beet leaves for functional ingredients. Wageningen University,
   Wageningen, the Netherlands.
- Tamayo Tenorio, A., Kyriakopoulou, K.E., Suarez-Garcia, E., van den Berg, C., van der Goot, A.J., 2018.
   Understanding differences in protein fractionation from conventional crops, and herbaceous
   and aquatic biomass Consequences for industrial use. Trends Food Sci. Technol. 71, 235–
   245. https://doi.org/10.1016/j.tifs.2017.11.010
- Tan, S.H., Mailer, R.J., Blanchard, C.L., Agboola, S.O., 2011. Canola Proteins for Human Consumption:
   Extraction, Profile, and Functional Properties. J. Food Sci. 76, R16–R28.
   https://doi.org/10.1111/j.1750-3841.2010.01930.x
- Tasaki, K., 2020. A novel thermal hydrolysis process for extraction of keratin from hog hair for
   commercial applications. Waste Manag. 104, 33–41.
   https://doi.org/10.1016/j.wasman.2019.12.042
- te Pas, M.F.W., Veldkamp, T., de Haas, Y., Bannink, A., Ellen, E.D., 2021. Adaptation of Livestock to
   New Diets Using Feed Components without Competition with Human Edible Protein
   Sources—A Review of the Possibilities and Recommendations. Animals 11, 2293.
   https://doi.org/10.3390/ani11082293

- Teekens, A.M., Bruins, M.E., Kasteren, J.M. van, Hendriks, W.H., Sanders, J.P., 2016. Synergy between
   bio-based industry and the feed industry through biorefinery. J. Sci. Food Agric. 96, 2603–
   2612. https://doi.org/10.1002/jsfa.7596
- 2287Teigiserova, D., 2020. Production of enzymes via solid state fermentation from cereals. Mendeley2288Data. https://doi.org/10.17632/k2xv3yss8m.1
- Teigiserova, D.A., Hamelin, L., Thomsen, M., 2020. Towards transparent valorization of food surplus,
   waste and loss: Clarifying definitions, food waste hierarchy, and role in the circular economy.
   Sci. Total Environ. 706, 136033. https://doi.org/DOI:
- 2292 http://dx.doi.org/10.1016/j.scitotenv.2019.136033
- Teigiserova, D.A., Thomsen, M., Hamelin, L., 2019. Review of high-value food waste and food
   residues biorefineries with focus on unavoidable wastes from processing. Resourc. Conserv.
   Recy. 149, 413–426. https://doi.org/10.1016/j.resconrec.2019.05.003
- ten Caat, N., Tillie, N., Tenpierik, M., 2021. Pig Farming vs. Solar Farming: Exploring Novel
   Opportunities for the Energy Transition, in: Roggema, R. (Ed.), TransFEWmation: Towards
   Design-Led Food-Energy-Water Systems for Future Urbanization, Contemporary Urban
   Design Thinking. Springer International Publishing, Cham, pp. 253–280.
   https://doi.org/10.1007/978-3-030-61977-0\_12
- 2301Tenlep, L., 2020. Upcycling wood into nutritional, sustainable protein source for people and the2302planet [WWW Document]. Arbiom. URL https://arbiom.com/wp-
- 2303content/uploads/2020/07/Upcycling-Wood-Into-Nutritional-Sustainable-Protein-Source-for-2304People-the-Planet\_Arbiom-wp\_Ltenlep.pdf (accessed 9.3.20).
- 2305Tereos, 2020. Alfalfa [WWW Document]. URL https://tereos.com/en/activities-and-products/raw-2306materials/alfalfa/ (accessed 12.4.20).
- Thomsen, M.H., Andersen, M., Kiel, P., 2008. Plant Juice in the Biorefinery Use of Plant Juice as
   Fermentation Medium, in: Biorefineries-Industrial Processes and Products. John Wiley &
   Sons, Ltd, pp. 295–314. https://doi.org/10.1002/9783527619849.ch13
- Tlais, A.Z.A., Fiorino, G.M., Polo, A., Filannino, P., Di Cagno, R., 2020. High-Value Compounds in Fruit,
   Vegetable and Cereal Byproducts: An Overview of Potential Sustainable Reuse and
   Exploitation. Molecules 25, 2987. https://doi.org/10.3390/molecules25132987
- 2313Toldrá, F., Mora, L., Reig, M., 2016. New insights into meat by-product utilization. Meat Sci. 120, 54–231459. https://doi.org/10.1016/j.meatsci.2016.04.021
- Tomlinson, E.J., 1976. The production of Single-cell protein from strong organic waste waters from
   the food and drink processing industries. Water Res. 10, 367–371.
   https://doi.org/10.1016/0043-1354(76)90053-1
- Torres-León, C., Ramírez-Guzman, N., Londoño-Hernandez, L., Martinez-Medina, G.A., Díaz-Herrera,
   R., Navarro-Macias, V., Alvarez-Pérez, O.B., Picazo, B., Villarreal-Vázquez, M., Ascacio-Valdes,
   J., Aguilar, C.N., 2018. Food Waste and Byproducts: An Opportunity to Minimize Malnutrition
   and Hunger in Developing Countries. Front. Sustain. Food Syst. 2.
- 2322 https://doi.org/10.3389/fsufs.2018.00052
- 2323Torres-Tiji, Y., Fields, F.J., Mayfield, S.P., 2020. Microalgae as a future food source. Biotechnology2324Advances 107536. https://doi.org/10.1016/j.biotechadv.2020.107536
- Tracy, B.P., Jones, S.W., Phillips, J.R., Mitchell, D.K., Eyal, A.M., 2020. Single cell protein products and
   an integrated method for the production of ethanol and single cell protein.
   US20200063091A1.
- 2328Triolo, J.M., 2013. Novel mathematical algorithms to predict energy potential and biodegradability of2329carbon sources for biogas production. University of Southern Denmark.
- Tsapekos, P., Khoshnevisan, B., Zhu, X., Zha, X., Angelidaki, I., 2019. Methane oxidising bacteria to
   upcycle effluent streams from anaerobic digestion of municipal biowaste. J. Environ.
   Manage. 251, 109590. https://doi.org/10.1016/j.jenvman.2019.109590
- Tubb, C., Seba, T., 2019. Rethinking Food & Agriculture 2020-2030, The second Domestication of
   Plants and Animals, the Disruption of the Cow, and the Collapse of Industrial Livestock
   Farming, Sector Disruption Report. RethinkX.

2336 Tuomisto, H.L., 2019. Vertical Farming and Cultured Meat: Immature Technologies for Urgent 2337 Problems. One Earth 1, 275–277. https://doi.org/10.1016/j.oneear.2019.10.024 2338 Tuorila, H., Hartmann, C., 2020. Consumer responses to novel and unfamiliar foods. Curr. Opin. Food 2339 Sci. 33, 1–8. https://doi.org/10.1016/j.cofs.2019.09.004 2340 Tzachor, A., Richards, C.E., Holt, L., 2021. Future foods for risk-resilient diets. Nat. Food 1–4. 2341 https://doi.org/10.1038/s43016-021-00269-x 2342 Upcraft, T., Tu, W.-C., Johnson, R., Finnigan, T., Hung, N.V., Hallett, J., Guo, M., 2021. Protein from renewable resources: mycoprotein production from agricultural residues. Green Chem. 23, 2343 2344 5150-5165. https://doi.org/10.1039/D1GC01021B 2345 Uwineza, C., Mahboubi, A., Atmowidjojo, A., Ramadhani, A., Wainaina, S., Millati, R., Wikandari, R., Niklasson, C., Taherzadeh, M.J., 2021. Cultivation of edible filamentous fungus Aspergillus 2346 2347 oryzae on volatile fatty acids derived from anaerobic digestion of food waste and cow 2348 manure. Bioresour. Technol. 337, 125410. https://doi.org/10.1016/j.biortech.2021.125410 Valenzuela-Grijalva, V.N., Pinelli-Saavedra, A., Muhlia-Almazan, A., Dominguez-Diaz, D., Gonzalez-2349 2350 Rios, H., 2017. Dietary inclusion effects of phytochemicals as growth promoters in animal 2351 production. J. Anim. Sci. Technol. https://doi.org/10.1186/s40781-017-0133-9 2352 van der Goot, A.J., Pelgrom, P.J.M., Berghout, J.A.M., Geerts, M.E.J., Jankowiak, L., Hardt, N.A., Keijer, 2353 J., Schutyser, M.A.I., Nikiforidis, C.V., Boom, R.M., 2016. Concepts for further sustainable 2354 production of foods. J. Food Eng. 168, 42–51. 2355 https://doi.org/10.1016/j.jfoodeng.2015.07.010 2356 Van Eenennaam, A.L., Werth, S.J., 2021. Animal board invited review: Animal agriculture and 2357 alternative meats – learning from past science communication failures. Animal 15, 100360. 2358 https://doi.org/10.1016/j.animal.2021.100360 2359 Van Hal, O., 2020. Upcycling biomass in a circular food system: the role of livestock and fish. 2360 Wageningen University, Wageningen, the Netherlands. 2361 Van Hal, O., de Boer, I.J.M., Muller, A., de Vries, S., Erb, K.-H., Schader, C., Gerrits, W.J.J., van Zanten, 2362 H.H.E., 2019. Upcycling food leftovers and grass resources through livestock: Impact of 2363 livestock system and productivity. J. Clean. Prod. 219, 485–496. 2364 https://doi.org/10.1016/j.jclepro.2019.01.329 Van Huis, A., 2020. Insects as food and feed, a new emerging agricultural sector: a review. J. Insects 2365 2366 as Food Feed. 6, 27–44. https://doi.org/10.3920/JIFF2019.0017 2367 van Kuijk, S.J.A., Sonnenberg, A.S.M., Baars, J.J.P., Hendriks, W.H., Cone, J.W., 2015. Fungal treated 2368 lignocellulosic biomass as ruminant feed ingredient: A review. Biotechnol. Adv. 33, 191–202. 2369 https://doi.org/10.1016/j.biotechadv.2014.10.014 2370 Van Zanten, H.H.E., Herrero, M., Van Hal, O., Röös, E., Muller, A., Garnett, T., Gerber, P.J., Schader, C., 2371 De Boer, I.J.M., 2018. Defining a land boundary for sustainable livestock consumption. Glob. 2372 Change Biol. 24, 4185–4194. https://doi.org/10.1111/gcb.14321 2373 Van Zanten, H.H.E., Ittersum, M., Boer, I.J.M., 2019. The role of farm animals in a circular food 2374 system. Glob. Food Sec. 21, 18–22. https://doi.org/10.1016/j.gfs.2019.06.003 2375 Varelas, V., 2019. Food Wastes as a Potential New Source for Edible Insect Mass Production for Food 2376 and Feed: A review. Fermentation 5, 81. https://doi.org/10.3390/fermentation5030081 2377 Venkata Mohan, S., Nikhil, G.N., Chiranjeevi, P., Nagendranatha Reddy, C., Rohit, M.V., Kumar, A.N., 2378 Sarkar, O., 2016. Waste biorefinery models towards sustainable circular bioeconomy: Critical 2379 review and future perspectives. Bioresour. Technol. 215, 2–12. 2380 https://doi.org/10.1016/j.biortech.2016.03.130 2381 Venkateswar Rao, L., Goli, J.K., Gentela, J., Koti, S., 2016. Bioconversion of lignocellulosic biomass to 2382 xylitol: An overview. Bioresour. Technol. 213, 299-310. https://doi.org/10.1016/j.biortech.2016.04.092 2383 Verbeeck, K., Vrieze, J.D., Pikaar, I., Verstraete, W., Rabaey, K., 2020. Assessing the potential for up-2384 cycling recovered resources from anaerobic digestion through microbial protein production. 2385 2386 Microb. Biotechnol. 14, 897–910. https://doi.org/10.1111/1751-7915.13600

- Verstraete, W., Clauwaert, P., Vlaeminck, S.E., 2016. Used water and nutrients: Recovery
  perspectives in a 'panta rhei' context. Bioresour. Technol. 215, 199–208.
  https://doi.org/10.1016/j.biortech.2016.04.094
- Verstraete, W., Windey, K.F.M.-L., Wambeke, M.A.L.A., 2020. Werkwijze voor het omzetten van
   zetmeelhoudende reststromen naar hoogwaardige proteïnes. BE1026952B1.
- Vethathirri, R.S., Santillan, E., Wuertz, S., 2021. Microbial community-based protein production from
   wastewater for animal feed applications. Bioresour. Technol. 341, 125723.
   https://doi.org/10.1016/j.biortech.2021.125723
- Villas-Boas, S., Esposito, E., Mitchell, D., 2002. Microbial conversion of lignocellulosic residues for
   production of animal feeds. Anim. Feed Sci. Technol. https://doi.org/10.1016/S0377 8401(02)00017-2
- Villas-Boas, S.G., Granucci, N., 2018. Process and composition for an improved flour product.
   US20180146688A1.
- Voutilainen, E., Pihlajaniemi, V., Parviainen, T., 2021. Economic comparison of food protein
   production with single-cell organisms from lignocellulose side-streams. Bioresour. Technol.
   Rep. 14, 100683. https://doi.org/10.1016/j.biteb.2021.100683
- Vriens, L., Nihoul, R., Verachtert, H., 1989. Activated sludges as animal feed: A review. Biol. Wastes
   27, 161–207. https://doi.org/10.1016/0269-7483(89)90001-3
- Wadhwa, M., Bakshi, M.P.S., 2013. Utilization of fruit and vegetable wastes as livestock feed and as
   substrates for generation of other value-added products, Rap Publication. FAO, Rome.
- 2407 Wang, D., Sakoda, A., Suzuki, M., 2001. Biological efficiency and nutritional value of Pleurotus
  2408 ostreatus cultivated on spent beer grain. Bioresour. Technol. 78, 293–300.
  2409 https://doi.org/10.1016/S0960-8524(01)00002-5
- Wanzenböck, E., Apprich, S., Tirpanalan, Ö., Zitz, U., Kracher, D., Schedle, K., Kneifel, W., 2017. Wheat
   bran biodegradation by edible Pleurotus fungi A sustainable perspective for food and feed.
   LWT 86, 123–131. https://doi.org/10.1016/j.lwt.2017.07.051
- Whittaker, J.A., Johnson, R.I., Finnigan, T.J.A., Avery, S.V., Dyer, P.S., 2020. The Biotechnology of
  Quorn Mycoprotein: Past, Present and Future Challenges, in: Nevalainen, H. (Ed.), Grand
  Challenges in Fungal Biotechnology, Grand Challenges in Biology and Biotechnology. Springer
  International Publishing, Cham, pp. 59–79. https://doi.org/10.1007/978-3-030-29541-7\_3
- Wilfart, A., Dusart, L., Méda, B., Gac, A., Espagnol, S., Morin, L., Dronne, Y., Garcia-Launay, F., 2019.
  Réduire les impacts environnementaux des aliments pour les animaux d'élevage. INRA Prod.
  Anim. 31, 289–306. https://doi.org/10.20870/productions-animales.2018.31.2.2285
- Willett, W., Rockström, J., Loken, B., Springmann, M., Lang, T., Vermeulen, S., Garnett, T., Tilman, D.,
  DeClerck, F., Wood, A., Jonell, M., Clark, M., Gordon, L.J., Fanzo, J., Hawkes, C., Zurayk, R.,
  Rivera, J.A., Vries, W.D., Sibanda, L.M., Afshin, A., Chaudhary, A., Herrero, M., Agustina, R.,
  Branca, F., Lartey, A., Fan, S., Crona, B., Fox, E., Bignet, V., Troell, M., Lindahl, T., Singh, S.,
- 2424 Cornell, S.E., Reddy, K.S., Narain, S., Nishtar, S., Murray, C.J.L., 2019. Food in the
  2425 Anthropocene: the EAT–Lancet Commission on healthy diets from sustainable food systems.
  2426 The Lancet 393, 447–492. https://doi.org/10.1016/S0140-6736(18)31788-4
- 2427 Wirsenius, S., 2000. Human Use of Land and Organic materials. Chalmers University of Technology 2428 and Göteborg University, Göteborg, Sweden.
- Wirsenius, S., Azar, C., Berndes, G., 2010. How much land is needed for global food production under
   scenarios of dietary changes and livestock productivity increases in 2030? Agric Syst 103,
   621–638. https://doi.org/10.1016/j.agsy.2010.07.005
- Wongputtisin, P., Khanongnuch, C., Kongbuntad, W., Niamsup, P., Lumyong, S., Sarkar, P.K., 2014.
  Use of Bacillus subtilis isolates from Tua-nao towards nutritional improvement of soya bean
  hull for monogastric feed application. Lett. Appl. Microbiol. 59, 328–333.
  https://doi.org/10.1111/lam.12279
- World Economic Forum, 2018. Innovation with a Purpose: The role of technology innovation in
   accelerating food systems transformation. World Economic Forum, Switzerland.

- Wyman, C.E., Dale, B.E., Elander, R.T., Holtzapple, M., Ladisch, M.R., Lee, Y.Y., Mitchinson, C.,
  Saddler, J.N., 2009. Comparative sugar recovery and fermentation data following
  pretreatment of poplar wood by leading technologies. Biotechnol. Prog. 25, 333–339.
  https://doi.org/10.1002/btpr.142
- Xiangping, L., Juping, L., Guanyi, C., Jianguang, Z., Chuanbin, W., Bin, L., 2019. Extraction and
   purification of eicosapentaenoic acid and docosahexaenoic acid from microalgae: A critical
   review. Algal Res. 43, 101619. https://doi.org/10.1016/j.algal.2019.101619
- Xiao, K., Zhou, Y., 2020. Protein recovery from sludge: A review. J. Clean. Prod. 249, 119373.
   https://doi.org/10.1016/j.jclepro.2019.119373
- Xu, M., 2021. Beyond the farm: Bioinorganic electro- synthesis of single cell protein from CO2 andgreen electricity. Technical University of Denmark.
- Xu, M., Zhou, H., Yang, X., Angelidaki, I., Zhang, Y., 2020. Sulfide restrains the growth of
   Methylocapsa acidiphila converting renewable biogas to single cell protein. Water Res. 184,
   116138. https://doi.org/10.1016/j.watres.2020.116138
- Yang, L., Li, H., Wang, Q., 2019. A novel one-step method for oil-rich biomass production and harvesting by co-cultivating microalgae with filamentous fungi in molasses wastewater.
  Bioresour. Technol. 275, 35–43. https://doi.org/10.1016/j.biortech.2018.12.036
- Yang, W., Xu, H., 2016. Industrial Fermentation of Vitamin C, in: Industrial Biotechnology of Vitamins,
  Biopigments, and Antioxidants. John Wiley & Sons, Ltd, pp. 161–192.
  https://doi.org/10.1002/9783527681754.ch7
- Yang, X., Xu, M., Zou, R., Angelidaki, I., Zhang, Y., 2021. Microbial protein production from CO2, H2,
  and recycled nitrogen: focusing on ammonia toxicity and nitrogen sources. J. Clean. Prod.
  125921. https://doi.org/10.1016/j.jclepro.2021.125921
- You, C., Chen, H., Myung, S., Sathitsuksanoh, N., Ma, H., Zhang, X.-Z., Li, J., Zhang, Y.-H.P., 2013.
  Enzymatic transformation of nonfood biomass to starch. PNAS 110.
  https://doi.org/10.1073/pnas.1302420110
- Zhang, G., Zhao, X., Li, X., Du, G., Zhou, J., Chen, J., 2020. Challenges and possibilities for biomanufacturing cultured meat. Trends Food Sci. Technol. 97, 443–450.
  https://doi.org/10.1016/j.tifs.2020.01.026
- Zhang, W., Alvarez-Gaitan, J.P., Dastyar, W., Saint, C.P., Zhao, M., Short, M.D., 2018. Value-Added
   Products Derived from Waste Activated Sludge: A Biorefinery Perspective. Water 10, 545.
   https://doi.org/10.3390/w10050545
- Zhu, W., He, Q., Gao, H., Nitayavardhana, S., Khanal, S.K., Xie, L., 2020. Bioconversion of yellow wine
   wastes into microbial protein via mixed yeast-fungus cultures. Bioresour. Technol. 299,
   122565. https://doi.org/10.1016/j.biortech.2019.122565
- Zlatanović, S., Kalušević, A., Micić, D., Laličić-Petronijević, J., Tomić, N., Ostojić, S., Gorjanović, S.,
  2019. Functionality and Storability of Cookies Fortified at the Industrial Scale with up to 75%
  of Apple Pomace Flour Produced by Dehydration. Foods 8, 561.
- 2476 https://doi.org/10.3390/foods8110561
- Zu Ermgassen, E.K.H.J., Phalan, B., Green, R.E., Balmford, A., 2016. Reducing the land use of EU pork
  production: where there's swill, there's a way. Food Policy 58, 35–48.
  https://doi.org/10.1016/j.foodpol.2015.11.001
- Zurbrugg, C., Dortmans, B., Fadhila, A., Vertsappen, B., Diener, S., 2018. From Pilot to Full Scale
   Operation of a Waste-to-Protein Treatment Facility. Detritus.
- 2482 https://doi.org/10.26403/detritus/2018.22
- 2483

## Fig. 1. Scope of the literature review

Residual biomass categories, here illustrated by icons, are further detailed in the SI. Agronomic valorization (e.g. as fertilizer) is not part of the scope as this study focuses on the direct recovery of edible ingredients only.

## Fig. 2. Waste-to-nutrition gap

Ternary diagram representing food grade quality perimeter (gray right corner), and approximating relative location of the studied solid residual biomass streams (colored circles). Phenolic lignin acts as both structural complexity and anti-nutritional proxies, but the latter is here privileged to differentiate wood-related residual biomass from green residual biomass and primary crops residues. MPF ingredients perimeter is not represented for tractability reasons. For the same reason, agrifood co-products and slaughterhouse by-products are gathered within the same broad circle (dotted line). Background data is available in the SI database and icons are as defined in figure 1.

\*Albeit some slaughterhouse by-products (e.g. offal) are directly edible (within the gray right corner), others are mainly composed by keratin which is here considered as structural content (top corner), see section 5.2.2.

## Fig. 3. Waste-to-nutrition pathways in four building blocks

Four generic families of conversion processes, illustrated with examples from the literature. Icons represent residual biomass categories as defined in figure 1 From top-down and leftright: (i) Fermented olive press-cake as fodder. (ii) Brewer' spent grains milled into bakery flour. (iii) Extraction of proteins from grass. (iv) Carbohydrates recovery from organic wastewater. (v) Recovery of cellulosic sugars. (vi) Feather processed with keratinases releasing amino acids. (vii) Insects farming on food waste and (viii) Microalgae cultured on aquaculture wastewater.

## Fig. 4. Waste-to-nutrition microbial bioconversion pathways

Metabolic pathways are adapted from (Alloul et al., 2021; Choi et al., 2021; Linder, 2019) and complemented to capture the diversity of inventoried waste-to-nutrition microbial bioconversions. Key nutrients such as nitrogen and phosphorus are not represented to ensure visual tractability. Chemo(auto)trophic carbon-monoxide-oxidizing bacteria pathways are not represented here due to the scarcity of reported information on these.

# Fig. 5. Producing alternative fermentation mediums from residual biomass: unit operations pattern

The indicative ranking of residual biomass families in the nutritional quality scale is derived from figure 2., and allows to visualize the estimated chain of unit processes required to bridge the gap between the initial composition-structure of a feedstock and the composition-structure which is adequate to deliver a nutritional service. Icons are as defined in figure 1. The identified conversion pathways are rather straightforward when starting from sugar- or lipid-rich residual biomass, but can be more complex, involving prior cracking and extraction operations to release fermentable compounds. Albeit limited by economic considerations (Kwan et al.,

2019), purification technologies are often required to detoxify feedstocks and bring them up to nutritional specifications.  $C_1$  gases can either have a fossil or biogenic origin, as represented in figure S5.

### Fig. 6. Identified waste-to-nutrition conversion pathway categories and current status

\* Technology Readiness Level (1-9): based on information available to date (SI).

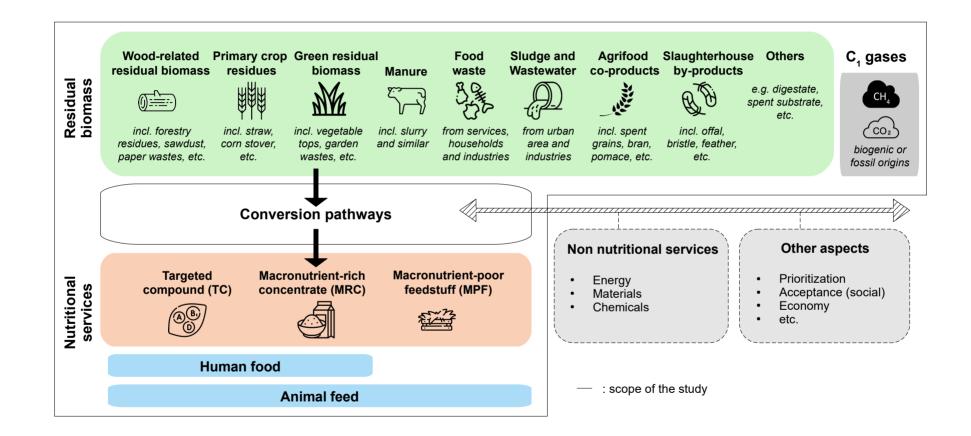
\*\* Valid for all pathways involving microorganisms.

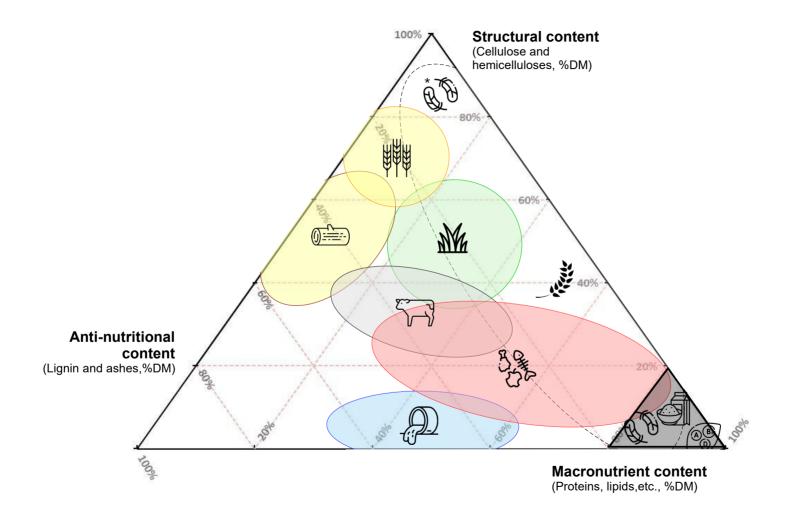
### References

Alloul, A., Spanoghe, J., Machado, D., Vlaeminck, S.E., 2021. Unlocking the genomic potential of aerobes and phototrophs for the production of nutritious and palatable microbial food without arable land or fossil fuels. Microb. Biotechnol. https://doi.org/10.1111/1751-7915.13747

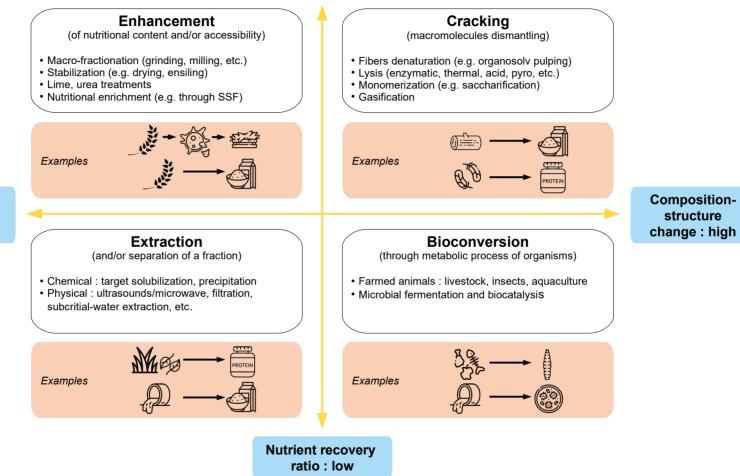
Choi, K.R., Yu, H.E., Lee, S.Y., 2021. Microbial food: microorganisms repurposed for our food. Microb. Biotechnol. https://doi.org/10.1111/1751-7915.13911

- Kwan, T.H., Ong, K.L., Haque, M.A., Kulkarni, S., Lin, C.S.K., 2019. Biorefinery of food and beverage waste valorisation for sugar syrups production: Techno-economic assessment. Process Saf. Environ. Prot. 121, 194–208. https://doi.org/10.1016/j.psep.2018.10.018
- Linder, T., 2019. Making the case for edible microorganisms as an integral part of a more sustainable and resilient food production system. Food Sec. 11, 265–278. https://doi.org/10.1007/s12571-019-00912-3





#### Nutrient recovery ratio : high



Compositionstructure change : low

