



HAL
open science

Early Phenotype Programming in Birds by Temperature and Nutrition: A Mini-Review

Charlotte Andrieux, Angélique Petit, Anne Collin, Marianne Houssier, Sonia Metayer-Coustard, Stéphane Panserat, Frederique Pitel, Vincent Coustham

► To cite this version:

Charlotte Andrieux, Angélique Petit, Anne Collin, Marianne Houssier, Sonia Metayer-Coustard, et al.. Early Phenotype Programming in Birds by Temperature and Nutrition: A Mini-Review. *Frontiers in Animal Science*, 2022, 2, 10.3389/fanim.2021.755842 . hal-03509411

HAL Id: hal-03509411

<https://hal.inrae.fr/hal-03509411>

Submitted on 4 Jan 2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License



Early Phenotype Programming in Birds by Temperature and Nutrition: A Mini-Review

Charlotte Andrieux^{1†}, Angélique Petit^{2†}, Anne Collin², Marianne Houssier¹, Sonia Métayer-Coustard², Stéphane Panserat¹, Frédérique Pitel³ and Vincent Coustham^{1,2*}

¹ INRAE, Université de Pau et des Pays de l'Adour, E2S UPPA, NUMEA, Saint-Pée-sur-Nivelle, France, ² INRAE, Université de Tours, BOA, Nouzilly, France, ³ INRAE, Université de Toulouse, ENVT, GenPhySE, Castanet-Tolosan, France

OPEN ACCESS

Edited by:

Pascale Chavatte-Palmer,
INRA UMR 1198 Biologie du
Développement et
Reproduction, France

Reviewed by:

Dana L. M. Campbell,
Commonwealth Scientific and
Industrial Research Organisation
(CSIRO), Australia
Christi Swaggerty,
United States Department of
Agriculture (USDA), United States

*Correspondence:

Vincent Coustham
vincent.coustham@inrae.fr

†These authors have contributed
equally to this work

Specialty section:

This article was submitted to
Animal Physiology and Management,
a section of the journal
Frontiers in Animal Science

Received: 09 August 2021

Accepted: 14 December 2021

Published: 04 January 2022

Citation:

Andrieux C, Petit A, Collin A,
Houssier M, Métayer-Coustard S,
Panserat S, Pitel F and Coustham V
(2022) Early Phenotype Programming
in Birds by Temperature and Nutrition:
A Mini-Review.
Front. Anim. Sci. 2:755842.
doi: 10.3389/fanim.2021.755842

Early development is a critical period during which environmental influences can have a significant impact on the health, welfare, robustness and performance of livestock. In oviparous vertebrates, such as birds, embryonic development takes place entirely in the egg. This allows the effects of environmental cues to be studied directly on the developing embryo. Interestingly, beneficial effects have been identified in several studies, leading to innovative procedures to improve the phenotype of the animals in the long term. In this review, we discuss the effects of early temperature and dietary programming strategies that both show promising results, as well as their potential transgenerational effects. The timing, duration and intensity of these procedures are critical to ensure that they produce beneficial effects without affecting animal survival or final product quality. For example, cyclic increases in egg incubation temperature have been shown to improve temperature tolerance and promote muscular growth in chickens or fatty liver production in mule ducks. *In ovo* feeding has also been successfully used to enhance digestive tract maturation, optimize chick development and growth, and thus obtain higher quality chicks. In addition, changes in the nutritional availability of methyl donors, for example, was shown to influence offspring phenotype. The molecular mechanisms behind early phenotype programming are still under investigation and are probably epigenetic in nature as shown by recent work in chickens.

Keywords: programming, bird, temperature, nutrition, *in ovo*, embryo

INTRODUCTION

Early development is a critical period during which the environment influences the health, welfare, robustness and performance of livestock (Ho et al., 2011; Reed and Clark, 2011). Long-term effects of the early environment, i.e., during embryogenesis or the first days of life, have been demonstrated in cattle (Reynolds and Vonnahme, 2017), sheep (Reynolds et al., 2010), pigs (Feeney et al., 2014), fish (Panserat et al., 2019), and birds (Feeney et al., 2014; Loyau et al., 2015) among others. The oviparous vertebrate model differs from the others due to an embryonic development outside of the dam. Therefore, the embryo can be easily manipulated, opening up new opportunities for phenotypic programming to improve poultry production.

Embryonic incubation conditions have been studied since the mid-20th century to find optimal incubation parameters for poultry production. The concept of programming has emerged more

recently with the demonstration of beneficial effects of different stimuli such as temperature (Loyau et al., 2015), nutrition (Uni et al., 2005; Cherian, 2011) or changes in maternal diet (Hynd et al., 2016; Baéza et al., 2017). The purpose of this review is to provide a concise description of the advancement of this concept in four poultry species (broiler chickens, quails, ducks and turkeys) with a particular interest on two main embryonic programming strategies, nutrition and temperature.

NUTRITIONAL PROGRAMMING STRATEGIES

The Maternal Nutrition as a Lever to Program the Progeny's Phenotype

Several maternal nutritional strategies have been developed to modulate egg nutrient content to obtain higher quality chicks in terms of robustness, growth and body composition. The links between hen nutrition and management, egg composition and subsequent animal behavior, performance, and disease susceptibility are well-established (Aigueperse et al., 2013). The breeder hen's diet can modulate the levels of other essential nutrients, which in turn can impact the fitness of hatched chicks and their phenotype later in life (for reviews, see Rühl, 2007; Morisson et al., 2017).

Fatty acids are essential for embryonic development, bird growth, the development of the central nervous system and the immune system (Noble et al., 1984; Ding and Lilburn, 1996; Wang et al., 2004; Cherian, 2015; Koppenol et al., 2015; Thanabalan and Kiarie, 2021). The balance of fatty acids (FA) can be modulated via the hen's diet. Ducklings from ducks fed with a FA ω 3-enriched diet have a higher live weight at hatch (D0), D28 and D56 and a lower feed conversion ratio for the growing period (Baéza et al., 2017). Reduced hyperactivity and stress responsiveness in ducklings were also observed. Supplementation with FA ω 3 LC also reduced the frequency and severity of pecking in ducklings.

In low protein feeding programs, not only are egg-laying rate and egg weight altered, but also the amount of leptin in the yolk sac and the expression of a number of genes in the yolk sac, hypothalamus, or muscle of the offspring (Rao et al., 2009). The chicks have lower hatch weight but faster post-hatch growth. More recently, it has been shown that feeding broiler breeders reduced protein diets has a negative impact on reproductive performance but improved offspring performance (Lesuisse et al., 2017), even in subsequent generations (Lesuisse et al., 2017, 2018). Studies testing different levels of digestible lysine (Ciacciariello and Tyler, 2013) or arginine (Fernandes et al., 2014) in hen diets have shown positive effects on offspring such as performance improvement, carcass yield, abdominal fat content, and bone quality for arginine supplementation.

Mineral and vitamin supplementations have often been studied to solve defects of mineralization of the skeleton and legs problems. Vitamins can be enriched in the egg through the hen's diet. Vitamin A is produced by the hen from the carotenoids in the feed. They have antioxidant properties, which are essential for the embryo. Indeed, in the last stage of incubation, fatty

acid oxidation increases, as does the production of free radicals and oxidative stress. These processes mainly cause damage to the embryos (Surai et al., 2016). Vitamin D3 regulates the flow of calcium through the chorioallantoic membrane. A vitamin D-deficient diet leads to decreased Ca^{++} transport across the chorioallantoic membrane and decreased Ca^{++} accumulation in the embryo, as well as increased late embryonic mortality (malposition, beak unable to break through the shell). Minerals such as iodine, selenium, magnesium, zinc, copper, iron or manganese can also be enriched in eggs (Jiakui and Xiaolong, 2004; Chinrasri et al., 2013; Favero et al., 2013; Saunders-Blades and Korver, 2015; Torres and Korver, 2018; Xie et al., 2019).

Overall, maternal feeding approaches optimize the hen's diet through supplementation or restriction of a wide variety of nutrients. However, it is often difficult to assess whether the effects of maternal diet on offspring are direct or not.

In ovo Nutrition Programming Strategies

In ovo feeding is a more direct way to influence offspring phenotype. Several studies have reported the use of *in ovo* nutrient supplementation to reduce the hatch window and improve health, post-hatch immune status, hatchability, hatched chick weight, growth performance, and meat quality (Uni and Ferket, 2004; Wei et al., 2011; Kadam et al., 2013; Roto et al., 2016; Gao et al., 2017; Peebles, 2018; Taha-Abdelaziz et al., 2018; Jha et al., 2019; Kalantar et al., 2019; Ayansola et al., 2021). New *in ovo* strategies also aim to address new challenges such as finding alternatives to antibiotic use through probiotic injections (Oladokun and Adewole, 2020). Therefore, *in ovo* stimulation of chicken microflora offers a better approach in establishing intestinal microflora (Alagawany et al., 2021).

At hatching, chicks switch from a yolk FA-based diet to a complete diet. Injection of carbohydrates and amino acids (AA) during embryonic development allows chicks to adapt to their post-hatch diet. Carbohydrates are widely studied because their concentration within the egg is less than one percent of total nutrients (Campos et al., 2011). To limit the utilization of FA and proteolysis of muscle proteins for energy purposes, injections of carbohydrates alone or combined with other nutrients of interest have been performed *in ovo* to increase glycogen storage and modulate energy status of chicks (Retes et al., 2017). Results depended on the type of sugar injected, injection site, embryo developmental stage, and genetics. Smirnov et al. (2006) showed an effect of carbohydrate injection on intestinal epithelium development with a 27% increase in villus area at hatching.

Amino acid administration improves hatching weight (Ohta et al., 2001) which persists up to 56 days of age in some studies (Al-Murrani, 1982). *In ovo* injection of AA such as arginine, considered an essential amino acid in birds, has been used to improve post-hatch growth performance via regulation of protein synthesis through the mTOR pathway (Yu et al., 2018). Arginine also stimulated myogenin gene expression in cultured chicken tissues (Li et al., 2016b). Moreover, *in ovo* injection of sulfur AA (methionine plus cysteine) resulted in improved embryonic development, IGF-I and TLR4 gene expression, antioxidant status and jejunum histomorphometry of newly

hatched broiler chicks exposed to heat stress during incubation (Elnesr et al., 2019; Elwan et al., 2019).

In ovo injection of AA, FA, vitamins, and trace elements on early post-hatch growth may also impact the development of lymphoid organs (thymus, bursa, and spleen) and immune parameters in broilers (Bakyaraj et al., 2012). *In ovo* vitamin and mineral administration significantly augmented the hatchability percentage and body weight of chicks post hatching (Alagawany et al., 2020; Hassan et al., 2021). The efficacy of vitamins C, E, D3, and folic acid on embryonic health and development has been reported in the literature (Peebles, 2018). The use of the yolk by the embryo for energy purposes results in oxidative processes, leading to the degradation of polyunsaturated fatty acids in cell membranes. Vitamins, such as vitamin E or C, protect the embryo by limiting the negative effects of free radicals (Surai et al., 2016; Araújo et al., 2018; Peebles, 2018). Results may depend on doses, ages, and injection sites.

Determining the mechanisms by which egg nutrients regulate cellular metabolism, signaling, gene expression and function is critical to improving nutrient utilization, poultry production efficiency and animal robustness. In birds, most studies only report phenotypic results of *in ovo* injections. Only a few recent studies are beginning to decipher the mechanisms involved in these phenotypic changes. Epigenetic changes may be involved, especially when methyl group donors, such as methionine, are injected (Anderson et al., 2012; Donohoe and Bultman, 2012; Veron et al., 2018). Thus, manipulation of sulfur AA content can induce changes in cellular function that may have implications for the development, long-term growth, and health of the animal. The early utilization of nutrients like AA can influence disease resistance and embryo survival (Saeed et al., 2019). Folate supplementation improved growth performance, immune function, and folate metabolism of broilers through epigenetic regulation of immune genes by altering chromatin conformation and epigenetic marks such as histone methylation (Li et al., 2016a). Injection of betaine (a component of the methionine cycle), also considered an effective antioxidant agent and methyl donor, affects hepatic cholesterol metabolism through epigenetic gene regulation in newly hatched chicks (Hu et al., 2015).

TEMPERATURE PROGRAMMING STRATEGIES

Temperature Increases During Egg Incubation

Thermal manipulation (TM) during embryogenesis has been studied for over three decades. TM involves altering egg incubation temperature to improve post-hatch physiological responses of birds (Iqbal et al., 1990). In particular, fine-tuning egg incubation temperature has been used to develop strategies to help chickens better withstand heat later in life (Loyau et al., 2015). TM has since been studied in several other avian species, including turkeys for thermoregulation and muscle growth (Maltby et al., 2004; Piestun et al., 2015), ducks for the lipid metabolism and liver (Wang et al., 2014; Massimino et al., 2019),

and quail for growth, physiological and metabolic parameters (Vitorino Carvalho et al., 2020).

Cyclic increases in incubation temperature, mimicking naturally fluctuating conditions, have been found to improve thermal tolerance while minimizing hatching defects (Piestun et al., 2008; Loyau et al., 2015). Because of the interference between the thermoregulatory system and other body functions, TM has also been shown to alter a broader range of phenotypes. For instance, TM has been shown to affect growth in broiler chickens and quails (Loyau et al., 2013; Vitorino Carvalho et al., 2020), muscle development in broiler chickens (Collin et al., 2007; Piestun et al., 2009), skin vascularization in broiler chickens (Morita et al., 2016) and immunity in Pekin ducks (Shanmugasundaram et al., 2018). In Pekin ducks, TM positively impacted muscle fiber diameter and regulatory pathways of muscle development (Liu et al., 2015; Li et al., 2017). Interestingly, TM increased liver weight (Liu et al., 2015) and lipogenesis gene activity in Pekin ducks (Wang et al., 2014). Three different TM conditions were shown to result in increased fatty liver weight, lipid amount, and droplets size after the overfeeding period in mule ducks (Massimino et al., 2019).

Several factors must be considered when implementing a thermal incubation strategy, the most important being the timing and the cyclicity of the treatment and the level of temperature increase (Loyau et al., 2015). For example, early days of embryogenesis and continuous temperature increases should be avoided, as they are associated with hatching defects (Massimino et al., 2019; Vitorino Carvalho et al., 2020). Breeding age and genetics contribute to the effectiveness of TM (Yalçın et al., 2005). Increasing the relative humidity in the incubator is another important parameter to prevent dehydration during temperature elevation (Loyau et al., 2015). Therefore, incubation parameters must be finely tuned to tip the balance toward positive rather than negative effects. This may explain why this seemingly straightforward procedure is not yet widely used in hatcheries.

One way to refine practices is to understand the mechanisms underlying the effects of TM. With advances in next-generation sequencing, genome-wide gene expression and epigenetic data have shed the light on some central and peripheral molecular effects of TM. In 35-day-old chickens and quails, TM has been shown to have a limited effect on gene expression in muscle and hypothalamus under normal rearing conditions (Loyau et al., 2016; Vitorino Carvalho et al., 2021). However, when animals were subjected to heat exposure at the same age, a much stronger gene expression response was found in the TM group compared to the control group. This may be explained by the involvement of epigenetic marks that are imprinted during the embryonic heat exposure and may trigger a differential response when the animals are again exposed to heat. This hypothesis is supported by the identification of several hundred differential peaks of histone marks altered by TM in the hypothalamus of 35-day-old chickens (David et al., 2019). In ducks, TM has been shown to affect the gene expression level of methylation enzymes (Yan et al., 2015), suggesting that incubation temperature may influence DNA methylation in ducks during early development. In addition, several studies have shown the involvement of heat shock proteins (HSP) and factors (HSF) that protect cells from

deleterious effects of stress such as misfolding and apoptosis (Costa et al., 2020). Interestingly, a recent study showed that TM altered the basal expression of HSP108, HSP90, HSF-1 and HSF-2 during late embryogenesis and the first week of life, but also the mRNA expression dynamics of these HSPs and HSFs during heat stress (Al-Zghoul and El-Bahr, 2019). HSPs were also identified as differentially expressed in genome-wide studies (Loyau et al., 2016; Vitorino Carvalho et al., 2021) but did not appear to be altered at the epigenetic level (David et al., 2019), suggesting that other mechanisms may be involved in TM lifelong memory.

Temperature Decreases During the Egg Incubation

Exposure of eggs to low incubation temperatures has several impacts on chick physiology, but also on long-term health and welfare traits. The young broiler chick is particularly sensitive to cold after hatching (Collin et al., 2003), and later in life, fast-growing broilers placed in a cold environment may develop an accumulation of fluid in the peritoneal cavity called ascites (Decuyper et al., 2000). While continuous incubation of eggs at low temperatures below 36°C results in degraded hatchability and increased pre-hatch incubation time (Kühn et al., 1982; Black and Burggren, 2004a,b), fine decreases in incubation temperatures have been proposed to stimulate subsequent cold tolerance in birds (Nichelmann and Tzschentke, 2002; Shinder et al., 2009; Akşit et al., 2013). Exposure to cold at the end of incubation did not alter hatchability but resulted in an increase in internal temperature at 3 days of age compared to control broilers chicks. This improved performance with a 5–10% increase in body weight at 14 and 35 days of age in standard temperature rearing (Shinder et al., 2011) and a 4% increase in female weight at 40 days of age, whereas no such change was observed in males (Nyuiadzi et al., 2020). The authors demonstrated beneficial effects of embryonic thermal programming on broiler health, with 19 and 26% reductions in mortality and incidence of ascites, respectively, compared to control chickens under ascites-inducing conditions. Less intense but cyclic cold embryonic thermal programming decreased mortality and ascites incidence during growth of chicks from old breeders (Shinder et al., 2011). Such treatment induced an increase in body weight but a degradation in feed efficiency and a better cold tolerance of broilers when subsequently subjected to cold (Akşit et al., 2013). Loyau et al. (2014) reported that at hatching, the same embryonic cold exposure conditions resulted in a 9-fold increase in catalase activity in the liver of treated chicks compared to controls. This suggests that cyclic embryonic cold exposure stimulated antioxidant defenses in chicks, presumably in response to a transient increase in cold-induced tissue oxidation risk during incubation (Mujahid and Furuse, 2009).

These medium- to long-term effects of short cold exposures during incubation have been shown to trigger heat production through modifications in thermoregulatory mechanisms via a change in neuronal receptors sensitivity in the hypothalamus (Nichelmann and Tzschentke, 2002), and an increase in plasma triiodothyronine T3 concentration (Kamanli et al., 2015). Finally, the impacts of cold exposure during incubation on subsequent

chick behavior were reported by Bertin et al. (2018). The authors analyzed the effect of acute decreases in temperature during days 12–19 of incubation on the expression of fear-related behaviors in broilers. At hatching, this treatment affected neurodevelopmental plasticity in the brain with higher expression of corticotropin-releasing factor in nuclei of the amygdala, altering the chicks' social behavior, novelty perception, and increasing their fear behavior. However, cold exposures during incubation under these conditions impaired the health and welfare of chickens reared in postnatal cold (Nyuiadzi et al., 2020).

TRANSGENERATIONAL PROGRAMMING

A growing number of studies have suggested that environmental exposures may be transmitted beyond exposed generations via “transgenerational epigenetic inheritance” (Jablonka and Raz, 2009). Non-genetic transgenerational inheritance has recently been shown to occur in birds (Brun et al., 2015; Leroux et al., 2017). For instance, Brun et al. (2015) showed that the Muscovy duck diet is capable of affecting traits related to growth and lipid metabolism in the grand-offspring, via the sire. In quail, *in ovo* injection of genistein, a phytoestrogen, impacted reproductive and behavioral traits after 3 generations without further injection (Leroux et al., 2017). In a 3-generation study in broilers, a reduced balanced protein diet induced transgenerational effects, including feather condition, polydipsia and frustration-related behavior (Buyse et al., 2020).

While these examples illustrate the existence of non-genetic inheritance of embryonic exposure in birds, the magnitude of these effects remains to be assessed in most cases. Although “epigenetic heritability” has been estimated at very low values for several egg quality traits in meat-type quails (Paiva et al., 2018b), an epigenetic heritability of 0.10 for the weight at 7 days of age has been reported (Paiva et al., 2018a). Concurrently, very little is known about the molecular mechanisms involved, especially in poultry (Guerrero-Bosagna et al., 2018). These may include alterations in sperm miRNAs and lncRNAs (Wu et al., 2019), or putative RNA modifications, DNA methylation, and retained histones (Matsushima et al., 2019).

CONCLUSIONS AND PERSPECTIVES

To face future challenges, including fluctuations due to climate change and changing farming systems, breeders are under pressure to increase performance and productivity, but also to ensure resilience and reduce resource use and environmental impact. In this context, epigenetic programming is an underestimated lever, as maternal or embryonic nutritional and thermal programming offers promising prospects to improve poultry performance and welfare. Programming the environment of animals (e.g., optimizing the way they are housed and fed) can indeed promote non-genetic factors that may be passed on the subsequent generations. This aspect is particularly important in the poultry industry where,

generally, production farms are located all over the world, including in warm climate regions, while breeding farms are concentrated in a few temperate locations. Identifying environmental changes in ancestors that affect offspring traits through the transmission of epigenetic marks would therefore allow breeders to produce commercial animals better adapted to local production environments. In order to implement such fine-tuned practices in the field, additional research is needed in this challenging area to account for the potential variability of breeders and the response of their offspring.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding author.

ETHICS STATEMENT

Ethical review and approval was not required for the animal study because this review cites approved studies.

REFERENCES

- Aigueperse, N., Calandreau, L., and Bertin, A. (2013). Maternal diet influences offspring feeding behavior and fearfulness in the precocial chicken. *PLoS ONE* 8:e77583. doi: 10.1371/journal.pone.0077583
- Akşit, M., Yalçın, S., Siegel, P. B., Yenisey, Ç., Özdemir, D., and Özkan, S. (2013). Broilers respond to cooler ambient temperatures after temperature acclimation during incubation and early postnatal age. *J. Appl. Poult. Res.* 22, 298–307. doi: 10.3382/japr.2012-00675
- Alagawany, M., Elnesr, S. S., Farag, M. R., Abd El-Hack, M. E., Barkat, R. A., Gabr, A. A., et al. (2021). Potential role of important nutraceuticals in poultry performance and health - A comprehensive review. *Res. Vet. Sci.* 137, 9–29. doi: 10.1016/j.rvsc.2021.04.009
- Alagawany, M., Elnesr, S. S., Farag, M. R., Tiwari, R., Yattoo, M. I., Karthik, K., et al. (2020). Nutritional significance of amino acids, vitamins and minerals as nutraceuticals in poultry production and health—a comprehensive review. *Vet. Q.* 41, 1–29. doi: 10.1080/01652176.2020.1857887
- Al-Murrani, W. K. (1982). Effect of injecting amino acids into the egg on embryonic and subsequent growth in the domestic fowl. *Br. Poult. Sci.* 23, 171–174. doi: 10.1080/00071688208447943
- Al-Zghoul, M. B., and El-Bahr, S. M. (2019). Basal and dynamics mRNA expression of muscular HSP108, HSP90, HSF-1 and HSF-2 in thermally manipulated broilers during embryogenesis. *BMC Vet. Res.* 15:83. doi: 10.1186/s12917-019-1827-7
- Anderson, O. S., Sant, K. E., and Dolinoy, D. C. (2012). Nutrition and epigenetics: an interplay of dietary methyl donors, one-carbon metabolism and DNA methylation. *J. Nutr. Biochem.* 23, 853–859. doi: 10.1016/j.jnutbio.2012.03.003
- Araújo, I. C. S., Café, M. B., Noleto, R. A., Martins, J. M. S., Ulhoa, C. J., Guareshi, G. C., et al. (2018). Effect of vitamin E in ovo feeding to broiler embryos on hatchability, chick quality, oxidative state, and performance. *Poult. Sci.* 98, 3652–3661. doi: 10.3382/ps/pey439
- Ayansola, H., Liao, C., Dong, Y., Yu, X., Zhang, B., and Wang, B. (2021). Prospect of early vascular tone and satellite cell modulations on white striping muscle myopathy. *Poult. Sci.* 100:100945. doi: 10.1016/j.psj.2020.12.042
- Baéza, E., Chartrin, P., Bordeau, T., Lessire, M., Thoby, J. M., Gigaud, V., et al. (2017). Omega-3 polyunsaturated fatty acids provided during embryonic development improve the growth performance and welfare of Muscovy ducks (*Cairina moschata*). *Poult. Sci.* 96, 3176–3187. doi: 10.3382/ps/pex147
- Bakyraraj, S., Bhanja, S. K., Majumdar, S., and Dash, B. (2012). Modulation of post-hatch growth and immunity through in ovo supplemented nutrients in broiler chickens. *J. Sci. Food Agric.* 92, 313–320. doi: 10.1002/jsfa.4577
- Bertin, A., Calandreau, L., Meurisse, M., Georgelin, M., Palme, R., Lumineau, S., et al. (2018). Incubation temperature affects the expression of young precocial birds' fear-related behaviours and neuroendocrine correlates. *Sci. Rep.* 8, 1–10. doi: 10.1038/s41598-018-20319-y
- Black, J. L., and Burggren, W. W. (2004a). Acclimation to hypothermic incubation in developing chicken embryos (*Gallus domesticus*) I. Developmental effects and chronic and acute metabolic adjustments. *J. Exp. Biol.* 207, 1543–1552. doi: 10.1242/jeb.00909
- Black, J. L., and Burggren, W. W. (2004b). Acclimation to hypothermic incubation in developing chicken embryos (*Gallus domesticus*) II. Hematology and blood O₂ transport. *J. Exp. Biol.* 207, 1553–1561. doi: 10.1242/jeb.00910
- Brun, J. M., Bernadet, M. D., Cornuez, A., Leroux, S., Bodin, L., Basso, B., et al. (2015). Influence of grand-mother diet on offspring performances through the male line in Muscovy duck. *BMC Genet.* 16, 1–11. doi: 10.1186/s12863-015-0303-z
- Buyse, J., Collin, A., Coustham, V., De Haas, E., and Pitel, F. (2020). The use of epigenetics in poultry breeding. *Adv. Poult. Genet. Genomics.* doi: 10.19103/AS.2020.0065.28
- Campos, A. M., de, A., Rostagno, H. S., Gomes, P. C., da Silva, E. A., Albino, L. F. T., et al. (2011). Efeito da inoculação de soluções nutritivas *in ovo* sobre a eclodibilidade e o desempenho de frangos de corte. *Rev. Bras. Zootec.* 40, 1712–1717. doi: 10.1590/S1516-35982011000800013
- Cherian, G. (2011). Essential fatty acids and early life programming in meat-type birds. *Worlds. Poult. Sci. J.* 67, 599–614. doi: 10.1017/S0043933911000705
- Cherian, G. (2015). Nutrition and metabolism in poultry: Role of lipids in early diet. *J. Anim. Sci. Biotechnol.* 6:28. doi: 10.1186/s40104-015-0029-9
- Chinrasri, O., Chantiratikul, P., Maneetong, S., Chookhampaeng, S., and Chantiratikul, A. (2013). Productivity and selenium concentrations in egg and tissue of laying quails fed selenium from hydroponically produced selenium-enriched kale sprout (*Brassica oleracea* var. *alboglabra* L.). *Biol. Trace Elem. Res.* 155, 381–386. doi: 10.1007/s12011-013-9824-3
- Ciacchiariello, M., and Tyler, N. C. (2013). The effects of maternal dietary lysine intake on offspring performance to 21 days of age. *J. Appl. Poult. Res.* 22, 238–244. doi: 10.3382/japr.2012-00625

AUTHOR CONTRIBUTIONS

All authors contributed to the writing of the review.

FUNDING

CA was funded by the Comité Départemental des Landes (CD40), France. AP was funded by the University of Tours, France. Some results on which this review is based on were part of studies funded by INRAE (Department Animal Physiology and Livestock Systems, Eval_Adapt project) and carried out within the UMT framework Integrative Biology Research and Development (BIRD) associating ITAVI and INRAE, by the French Agence Nationale de la Recherche (ANR), Programs ANR-09-JCJC-0015-01 THERMOCHICK, ANR-09-GENM-004 EPIBIRD and ANR-15-CE02-0009-01 QuailHeatE, and by the Comité Interprofessionnel des Palmipèdes à Foie Gras (CIFOG).

ACKNOWLEDGMENTS

The authors thank Mireille Morisson (UMR GenPhySE, Toulouse, France) for initial discussions on the review.

- Collin, A., Berri, C., Tesseraud, S., Requena Rodón, F. E., Skiba-Cassy, S., Crochet, S., et al. (2007). Effects of thermal manipulation during early and late embryogenesis on thermotolerance and breast muscle characteristics in broiler chickens. *Poult. Sci.* 86, 795–800. doi: 10.1093/ps/86.5.795
- Collin, A., Buyse, J., Van As, P., Darras, V. M., Malheiros, R. D., Moraes, V. M. B., et al. (2003). Cold-induced enhancement of avian uncoupling protein expression, heat production, and triiodothyronine concentrations in broiler chicks. *Gen. Comp. Endocrinol.* 130, 70–77. doi: 10.1016/S0016-6480(02)00571-3
- Costa, B. T. A., Lopes, T. S. B., Mesquita, M. A., Lara, L. J. C., and Araújo, I. C. S. (2020). Thermal manipulations of birds during embryogenesis. *Worlds. Poult. Sci. J.* 76, 843–851. doi: 10.1080/00439339.2020.1823302
- David, S. A., Vitorino Carvalho, A., Gimonet, C., Brionne, A., Hennequet-Antier, C., Piégu, B., et al. (2019). Thermal manipulation during embryogenesis impacts H3K4me3 and H3K27me3 histone marks in chicken hypothalamus. *Front. Genet.* 10, 1–11. doi: 10.3389/fgene.2019.01207
- Decuyper, E., Buyse, J., and Buys, N. (2000). Ascites in broiler chickens: exogenous and endogenous structural and functional causal factors. *Worlds. Poult. Sci. J.* 56, 374–377. doi: 10.1079/WPS20000025
- Ding, S. T., and Lilburn, M. S. (1996). Characterization of changes in yolk sac and liver lipids during embryonic and early posthatch development of turkey poults. *Poult. Sci.* 75, 478–483. doi: 10.3382/ps.0750478
- Donohoe, D. R., and Bultman, S. J. (2012). Metaboloepigenetics: Interrelationships between energy metabolism and epigenetic control of gene expression. *J. Cell. Physiol.* 227, 3169–3177. doi: 10.1002/jcp.24054
- Elnesr, S. S., Elwan, H. A. M., Xu, Q. Q., Xie, C., Dong, X. Y., and Zou, X. T. (2019). Effects of in ovo injection of sulfur-containing amino acids on heat shock protein 70, corticosterone hormone, antioxidant indices, and lipid profile of newly hatched broiler chicks exposed to heat stress during incubation. *Poult. Sci.* 98, 2290–2298. doi: 10.3382/ps/pey609
- Elwan, H. A. M., Elnesr, S. S., Xu, Q., Xie, C., Dong, X., and Zou, X. (2019). Effects of in ovo methionine-cysteine injection on embryonic development, antioxidant status, IGF-I and TLR4 gene expression, and jejunal histomorphometry in newly hatched broiler chicks exposed to heat stress during incubation. *Animals* 9, 1–13. doi: 10.3390/ani9010025
- Favero, A., Vieira, S. L., Angel, C. R., Bess, F., Cemin, H. S., and Ward, T. L. (2013). Reproductive performance of Cobb 500 breeder hens fed diets supplemented with zinc, manganese, and copper from inorganic and amino acid-complexed sources. *J. Appl. Poult. Res.* 22, 80–91. doi: 10.3382/japr.2012-00607
- Feeney, A., Nilsson, E., and Skinner, M. K. (2014). Epigenetics and transgenerational inheritance in domesticated farm animals. *J. Anim. Sci. Biotechnol.* 5, 1–7. doi: 10.1186/2049-1891-5-48
- Fernandes, J. I. M., Murakami, A. E., de Souza, L. M. G., Ospina-Rojas, I. C., and Rossi, R. M. (2014). Effect of arginine supplementation of broiler breeder hens on progeny performance. *Can. J. Anim. Sci.* 94, 313–321. doi: 10.4141/cjas2013-067
- Gao, T., Zhao, M., Zhang, L., Li, J., Yu, L., Lv, P., et al. (2017). Effect of in ovo feeding of L-arginine on the hatchability, growth performance, gastrointestinal hormones, and jejunal digestive and absorptive capacity of posthatch broilers. *J. Anim. Sci.* 95, 3079–3092. doi: 10.2527/jas.2016.0465
- Guerrero-Bosagna, C., Morisson, M., Liaubet, L., Rodenburg, T. B., de Haas, E. N., Košťál, L., et al. (2018). Transgenerational epigenetic inheritance in birds. *Environ. Epigenetics* 4, 0–8. doi: 10.1093/eep/dvy008
- Hassan, H. A., Arafat, A. R., Farroh, K. Y., Bahnas, M. S., El-wardany, I., and Elnesr, S. S. (2021). Effect of in ovo copper injection on body weight, immune response, blood biochemistry and carcass traits of broiler chicks at 35 days of age. *Anim. Biotechnol.* 1–8. doi: 10.1080/10495398.2021.1874964
- Ho, D. H., Reed, W. L., and Burggren, W. W. (2011). Egg yolk environment differentially influences physiological and morphological development of broiler and layer chicken embryos. *J. Exp. Biol.* 214, 619–628. doi: 10.1242/jeb.046714
- Hu, Y., Sun, Q., Li, X., Wang, M., Cai, D., Li, X., et al. (2015). In ovo injection of betaine affects hepatic cholesterol metabolism through epigenetic gene regulation in newly hatched chicks. *PLoS ONE* 10:e0122643. doi: 10.1371/journal.pone.0122643
- Hynd, P. I., Weaver, S., Edwards, N. M., Heberle, N. D., and Bowling, M. (2016). Developmental programming: a new frontier for the poultry industry? *Anim. Prod. Sci.* 56, 1233–1238. doi: 10.1071/AN15373
- Iqbal, A., Decuyper, E., Abd El Azim, A., and Kühn, E. R. (1990). Pre- and post-hatch high temperature exposure affects the thyroid hormones and corticosterone response to acute heat stress in growing chicken (*Gallus domesticus*). *J. Therm. Biol.* 15, 149–153. doi: 10.1016/0306-4565(90)90032-D
- Jablónka, E., and Raz, G. (2009). Epigenetic inheritance: prevalence, mechanisms, and implications for the study of heredity and evolution. *Q. Rev. Biol.* 84, 131–176. doi: 10.1086/598822
- Jha, R., Singh, A. K., Yadav, S., Berrococo, J. F. D., and Mishra, B. (2019). Early nutrition programming (in ovo and post-hatch feeding) as a strategy to modulate gut health of poultry. *Front. Vet. Sci.* 6:82. doi: 10.3389/fvets.2019.00082
- Jiakui, L., and Xiaolong, W. (2004). Effect of dietary organic versus inorganic selenium in laying hens on the productivity, selenium distribution in egg and selenium content in blood, liver and kidney. *J. Trace Elem. Med. Biol.* 18, 65–68. doi: 10.1016/j.jtemb.2004.04.002
- Kadam, M. M., Barekatin, M. R., K., Bhanja, S., and Iji, P. A. (2013). Prospects of in ovo feeding and nutrient supplementation for poultry: the science and commercial applications—a review. *J. Sci. Food Agric.* 93, 3654–3661. doi: 10.1002/jsfa.6301
- Kalantar, M., Hosseini, S. M., Hosseini, M. R., Kalantar, M. H., Farmanullah, and Yang, L. G. (2019). Effects of in ovo injection of coenzyme Q10 on hatchability, subsequent performance, and immunity of broiler chickens. *Biomed Res. Int.* 2019:7167525. doi: 10.1155/2019/7167525
- Kamanli, S., Durmuş, I., Yalçın, S., Yıldırım, U., and Meral, Ö. (2015). Effect of prenatal temperature conditioning of laying hen embryos: hatching, live performance and response to heat and cold stress during laying period. *J. Therm. Biol.* 51, 96–104. doi: 10.1016/j.jtherbio.2015.04.001
- Koppenol, A., Buyse, J., Everaert, N., Willems, E., Wang, Y., Franssens, L., et al. (2015). Transition of maternal dietary n-3 fatty acids from the yolk to the liver of broiler breeder progeny via the residual yolk sac. *Poult. Sci.* 94, 43–52. doi: 10.3382/ps/peu006
- Kühn, E. R., Decuyper, E., Colen, L. M., and Michels, H. (1982). Posthatch growth and development of a circadian rhythm for thyroid hormones in chicks incubated at different temperatures. *Poult. Sci.* 61, 540–549. doi: 10.3382/ps.0610540
- Leroux, S., Gourichon, D., Leterrier, C., Labrune, Y., Coustham, V., Rivière, S., et al. (2017). Embryonic environment and transgenerational effects in quail. *Genet. Sel. Evol.* 49, 1–8. doi: 10.1186/s12711-017-0292-7
- Lesuisse, J., Li, C., Schallier, S., Leblois, J., Everaert, N., and Buyse, J. (2017). Feeding broiler breeders a reduced balanced protein diet during the rearing and laying period impairs reproductive performance but enhances broiler offspring performance. *Poult. Sci.* 96, 3949–3959. doi: 10.3382/ps/pep211
- Lesuisse, J., Schallier, S., Li, C., Bautil, A., Li, B., Leblois, J., et al. (2018). Multigenerational effects of a reduced balanced protein diet during the rearing and laying period of broiler breeders. 2. Zootechnical performance of the F1 broiler offspring. *Poult. Sci.* 97, 1666–1676. doi: 10.3382/ps/pey014
- Li, S., Zhi, L., Liu, Y., Shen, J., Liu, L., Yao, J., et al. (2016a). Effect of in ovo feeding of folic acid on the folate metabolism, immune function and epigenetic modification of immune effector molecules of broiler. *Br. J. Nutr.* 115, 411–421. doi: 10.1017/S0007114515004511
- Li, X., Qiu, J., Liu, H., Wang, Y., Hu, J., Gan, X., et al. (2017). Long-term thermal manipulation in the late incubation period can inhibit breast muscle development by activating endoplasmic reticulum stress in duck (*Anas platyrhynchos domestica*). *J. Therm. Biol.* 70, 37–45. doi: 10.1016/j.jtherbio.2017.10.008
- Li, Y., Wang, Y., Willems, E., Willemsen, H., Franssens, L., Buyse, J., et al. (2016b). In ovo L-arginine supplementation stimulates myoblast differentiation but negatively affects muscle development of broiler chicken after hatching. *J. Anim. Physiol. Anim. Nutr.* 100, 167–177. doi: 10.1111/jpn.12299
- Liu, H., Liu, J., Yan, X., Li, Q., Zhao, Y., Wang, Y., et al. (2015). Impact of thermal stress during incubation on gene expression in embryonic muscle of Peking ducks (*Anas platyrhynchos domestica*). *J. Therm. Biol.* 53, 80–89. doi: 10.1016/j.jtherbio.2015.08.013
- Louay, T., Bedrani, L., Berri, C., Métayer-Coustard, S., Praud, C., Coustham, V., et al. (2015). Cyclic variations in incubation conditions induce adaptive responses to later heat exposure in chickens: a review. *Animal* 9, 76–85. doi: 10.1017/S1751731114001931

- Loyau, T., Berri, C., Bedrani, L., Métayer-Coustard, S., Praud, C., Duclos, M. J., et al. (2013). Thermal manipulation of the embryo modifies the physiology and body composition of broiler chickens reared in floor pens without affecting breast meat processing quality. *J. Anim. Sci.* 91, 3674–3685. doi: 10.2527/jas.2013-6445
- Loyau, T., Collin, A., Yenisey, Ç., Crochet, S., Siegel, P. B., Akşit, M., et al. (2014). Exposure of embryos to cyclically cold incubation temperatures durably affects energy metabolism and antioxidant pathways in broiler chickens. *Poult. Sci.* 93, 2078–2086. doi: 10.3382/ps.2014-03881
- Loyau, T., Hennequet-Antier, C., Coustham, V., Berri, C., Leduc, M., Crochet, S., et al. (2016). Thermal manipulation of the chicken embryo triggers differential gene expression in response to a later heat challenge. *BMC Genomics* 17:329. doi: 10.1186/s12864-016-2661-y
- Maltby, V., Somaiya, A., French, N. A., and Stickland, N. C. (2004). In ovo temperature manipulation influences post-hatch muscle growth in the turkey. *Br. Poult. Sci.* 45, 491–498. doi: 10.1080/00071660412331286190
- Massimino, W., Davail, S., Bernadet, M., Pioche, T., Ricaud, K., Gontier, K., et al. (2019). Impact of thermal manipulation during embryogenesis on hepatic metabolism in mule ducks. *Front. Physiol.* 10, 1–12. doi: 10.3389/fphys.2019.01495
- Matsushima, W., Brink, K., Schroeder, J., Miska, E. A., and Gapp, K. (2019). Mature sperm small-RNA profile in the sparrow: implications for transgenerational effects of age on fitness. *Environ. Epigenetics* 5, 1–11. doi: 10.1093/eep/dvz007
- Morisson, M., Coustham, V., Frésard, L., Collin, A., Zerjal, T., Métayer-coustard, S., et al. (2017). *Handbook of Nutrition, Diet, and Epigenetics*. Cham: Springer. Available online at: https://link.springer.com/referenceworkentry/10.1007%2F978-3-319-31143-2_40-1#citeas
- Morita, V. S., Almeida, V. R., Matos, J. B., Vicentini, T. I., van den Brand, H., et al. (2016). Incubation temperature during fetal development influences morphophysiological characteristics and preferred ambient temperature of chicken hatchlings. *PLoS ONE* 11:e0154928. doi: 10.1371/journal.pone.0154928
- Mujahid, A., and Furuse, M. (2009). Oxidative damage in different tissues of neonatal chicks exposed to low environmental temperature. *Comp. Biochem. Physiol. Mol. Integr. Physiol.* 152, 604–608. doi: 10.1016/j.cbpa.2009.01.011
- Nichelmann, M., and Tzschentke, B. (2002). Ontogeny of thermoregulation in precocial birds. *Comp. Biochem. Physiol. Mol. Integr. Physiol.* 131, 751–763. doi: 10.1016/S1095-6433(02)00013-2
- Noble, R. C., Connor, K., and Smith, W. K. (1984). The synthesis and accumulation of cholesteryl esters by the developing embryo of the domestic fowl. *Poult. Sci.* 63, 558–564. doi: 10.3382/ps.0630558
- Nyuiadz, D., Berri, C., Dusart, L., Travel, A., Méda, B., Bouvarel, I., et al. (2020). Short cold exposures during incubation and postnatal cold temperature affect performance, breast meat quality, and welfare parameters in broiler chickens. *Poult. Sci.* 99, 857–868. doi: 10.1016/j.psj.2019.10.024
- Ohta, Y., Kidd, M. T., and Ishibashi, T. (2001). Embryo growth and amino acid concentration profiles of broiler breeder eggs, embryos, and chicks after *in ovo* administration of amino acids. *Poult. Sci.* 80, 1430–1436. doi: 10.1093/ps/80.10.1430
- Oladokun, S., and Adewole, D. I. (2020). In ovo delivery of bioactive substances: an alternative to the use of antibiotic growth promoters in poultry production—a review. *J. Appl. Poult. Res.* 29, 744–763. doi: 10.1016/j.japr.2020.06.002
- Paiva, J. T., de Resende, M. D. V., Resende, R. T., de Oliveira, H. R., Silva, H. T., Caetano, G. C., et al. (2018a). Transgenerational epigenetic variance for body weight in meat quails. *J. Anim. Breed. Genet.* 135, 178–185. doi: 10.1111/jbg.12329
- Paiva, J. T., De Resende, M. D. V., Resende, R. T., Oliveira, H. R., Silva, H. T., Caetano, G. C., et al. (2018b). A note on transgenerational epigenetics affecting egg quality traits in meat-type quail. *Br. Poult. Sci.* 59, 624–628. doi: 10.1080/00071668.2018.1514582
- Panserat, S., Marandel, L., Seiliez, I., and Skiba-Cassy, S. (2019). New insights on intermediary metabolism for a better understanding of nutrition in teleosts. *Annu. Rev. Anim. Biosci.* 7, 195–220. doi: 10.1146/annurev-animal-020518-115250
- Peebles, E. D. (2018). *In ovo* applications in poultry: a review. *Poult. Sci.* 97, 2322–2338. doi: 10.3382/ps/pey081
- Piestun, Y., Harel, M., Barak, M., Yahav, S., and Halevy, O. (2009). Thermal manipulations in late-term chick embryos have immediate and longer term effects on myoblast proliferation and skeletal muscle hypertrophy. *J. Appl. Physiol.* 106, 233–240. doi: 10.1152/japplphysiol.91090.2008
- Piestun, Y., Shinder, D., Ruzal, M., Halevy, O., Brake, J., and Yahav, S. (2008). Thermal manipulations during broiler embryogenesis: Effect on the acquisition of thermotolerance. *Poult. Sci.* 87, 1516–1525. doi: 10.3382/ps.2008-00030
- Piestun, Y., Zimmerman, I., and Yahav, S. (2015). Thermal manipulations of turkey embryos: the effect on thermoregulation and development during embryogenesis. *Poult. Sci.* 94, 273–280. doi: 10.3382/ps/peu047
- Rao, K., Xie, J., Yang, X., Chen, L., Grossmann, R., and Zhao, R. (2009). Maternal low-protein diet programmes offspring growth in association with alterations in yolk leptin deposition and gene expression in yolk-sac membrane, hypothalamus and muscle of developing Langshan chicken embryos. *Br. J. Nutr.* 102, 848–857. doi: 10.1017/S0007114509276434
- Reed, W. L., and Clark, M. E. (2011). Beyond maternal effects in birds: responses of the embryo to the environment. *Integr. Comp. Biol.* 51, 73–80. doi: 10.1093/icb/ict032
- Retes, P. L., Clemente, A. H. S., Neves, D. G., Espósito, M., Makiyama, L., Alvarenga, R. R., et al. (2017). *In ovo* feeding of carbohydrates for broilers—a systematic review. *J. Anim. Physiol. Anim. Nutr.* 102, 361–369. doi: 10.1111/jpn.12807
- Reynolds, L. P., Borowicz, P. P., Caton, J. S., Vonnahme, K. A., Luther, J. S., Hammer, C. J., et al. (2010). Developmental programming: the concept, large animal models, and the key role of uteroplacental vascular development. *J. Anim. Sci.* 88, E61–72. doi: 10.2527/jas.2009-2359
- Reynolds, L. P., and Vonnahme, K. A. (2017). Livestock as models for developmental programming. *Anim. Front.* 7, 12–17. doi: 10.2527/af.2017-0123
- Roto, S. M., Kwon, Y. M., and Ricke, S. C. (2016). Applications of *in ovo* technique for the optimal development of the gastrointestinal tract and the potential influence on the establishment of its microbiome in poultry. *Front. Vet. Sci.* 3:63. doi: 10.3389/fvets.2016.00063
- Rühl, R. (2007). Effects of dietary retinoids and carotenoids on immune development: symposium on “nutritional influences on developmental immunology.” *Proc. Nutr. Soc.* 66, 458–469. doi: 10.1017/S002966510600509X
- Saeed, M., Babazadeh, D., Naveed, M., Alagawany, M., Abd El-Hack, M. E., Arain, M. A., et al. (2019). *In ovo* delivery of various biological supplements, vaccines and drugs in poultry: current knowledge. *J. Sci. Food Agric.* 99, 3727–3739. doi: 10.1002/jsfa.9593
- Saunders-Blades, J. L., and Korver, D. R. (2015). Effect of hen age and maternal vitamin D source on performance, hatchability, bone mineral density, and progeny *in vitro* early innate immune function. *Poult. Sci.* 94, 1233–1246. doi: 10.3382/ps/pev002
- Shanmugasundaram, R., Wick, M., and Lilburn, M. S. (2018). Effect of embryonic thermal manipulation on heat shock protein 70 expression and immune system development in Pekin duck embryos. *Poult. Sci.* 97, 4200–4210. doi: 10.3382/ps/pey298
- Shinder, D., Rusal, M., Giloh, M., and Yahav, S. (2009). Effect of repetitive acute cold exposures during the last phase of broiler embryogenesis on cold resistance through the life span. *Poult. Sci.* 88, 636–646. doi: 10.3382/ps.2008-00213
- Shinder, D., Ruzal, M., Giloh, M., Druyan, S., Piestun, Y., and Yahav, S. (2011). Improvement of cold resistance and performance of broilers by acute cold exposure during late embryogenesis. *Poult. Sci.* 90, 633–641. doi: 10.3382/ps.2010-01089
- Smirnov, A., Tako, E., Ferket, P. R., and Uni, Z. (2006). Mucin gene expression and mucin content in the chicken intestinal goblet cells are affected by *in ovo* feeding of carbohydrates. *Poult. Sci.* 85, 669–673. doi: 10.1093/ps/85.4.669
- Surai, P. F., Fisinin, V. I., and Karadas, F. (2016). Antioxidant systems in chick embryo development. Part 1. Vitamin E, carotenoids and selenium. *Anim. Nutr.* 2, 1–11. doi: 10.1016/j.aninu.2016.01.001
- Taha-Abdelaziz, K., Hodgins, D. C., Lammers, A., Alkie, T. N., and Sharif, S. (2018). Effects of early feeding and dietary interventions on development of lymphoid organs and immune competence in neonatal chickens: a review. *Vet. Immunol. Immunopathol.* 201, 1–11. doi: 10.1016/j.vetimm.2018.05.001
- Thanabalan, A., and Kiarie, E. G. (2021). Influence of feeding omega-3 polyunsaturated fatty acids to broiler breeders on indices of immunocompetence, gastrointestinal, and skeletal development in broiler chickens. *Front. Vet. Sci.* 8:653152. doi: 10.3389/fvets.2021.653152
- Torres, C. A., and Korver, D. R. (2018). Influences of trace mineral nutrition and maternal flock age on broiler embryo bone development. *Poult. Sci.* 97, 2996–3003. doi: 10.3382/ps/pey136

- Uni, Z., Ferket, P. R., Tako, E., and Kedar, O. (2005). In ovo feeding improves energy status of late-term chicken embryos. *Poult. Sci.* 84, 764–770. doi: 10.1093/ps/84.5.764
- Uni, Z., and Ferket, R. P. (2004). Methods for early nutrition and their potential. *Worlds. Poult. Sci. J.* 60, 101–111. doi: 10.1079/WPS20038
- Veron, V., Marandel, L., Liu, J., Vélez, E. J., Lepais, O., Panserat, S., et al. (2018). DNA methylation of the promoter region of *bnip3* and *bnip3l* genes induced by metabolic programming 06 Biological Sciences 0604 Genetics. *BMC Genomics* 19, 1–14. doi: 10.1186/s12864-018-5048-4
- Vitorino Carvalho, A., Hennequet-antier, C., Brionne, A., Crochet, S., Jimenez, J., Couroussé, N., et al. (2021). Embryonic thermal manipulation impacts the postnatal transcriptome response of heat-challenged Japanese quails. *BMC Genomics* 22, 1–13. doi: 10.1186/s12864-021-07832-7
- Vitorino Carvalho, A., Hennequet-Antier, C., Crochet, S., Bordeau, T., Couroussé, N., Cailleau-Audouin, E., et al. (2020). Embryonic thermal manipulation has short and long-term effects on the development and the physiology of the Japanese quail. *PLoS ONE* 15:e0227700. doi: 10.1371/journal.pone.0227700
- Wang, G., Liu, J., Xiang, S., Yan, X., Li, Q., Cui, C., et al. (2014). Influence of *in ovo* thermal manipulation on lipid metabolism in embryonic duck liver. *J. Therm. Biol.* 43, 40–45. doi: 10.1016/j.jtherbio.2014.05.001
- Wang, Y. W., Sunwoo, H., Cherian, G., and Sim, J. S. (2004). Maternal dietary ratio of linoleic acid to α -linolenic acid affects the passive immunity of hatching chicks. *Poult. Sci.* 83, 2039–2043. doi: 10.1093/ps/83.12.2039
- Wei, X. J., Ni, Y. D., Lu, L. Z., Grossmann, R., and Zhao, R. Q. (2011). The effect of equol injection *in ovo* on posthatch growth, meat quality and antioxidation in broilers. *Animal* 5, 320–327. doi: 10.1017/S1751731110001771
- Wu, S., Guo, W., Li, X., Liu, Y., Li, Y., Lei, X., et al. (2019). Paternal chronic folate supplementation induced the transgenerational inheritance of acquired developmental and metabolic changes in chickens. *Proc. R. Soc. B Biol. Sci.* 286:20191653. doi: 10.1098/rspb.2019.1653
- Xie, C., Elwan, H. A. M., Elnesr, S. S., Dong, X. Y., and Zou, X. T. (2019). Effect of iron glycine chelate supplementation on egg quality and egg iron enrichment in laying hens. *Poult. Sci.* 98, 7101–7109. doi: 10.3382/ps/pez421
- Yalçın, S., Özkan, S., Çabuk, M., Buyse, J., Decuypere, E., and Siegel, P. B. (2005). Pre- and postnatal conditioning induced thermotolerance on body weight, physiological responses and relative asymmetry of broilers originating from young and old breeder flocks. *Poult. Sci.* 84, 967–976. doi: 10.1093/ps/84.6.967
- Yan, X. P., Liu, H. H., Liu, J. Y., Zhang, R. P., Wang, G. S., Li, Q. Q., et al. (2015). Evidence in duck for supporting alteration of incubation temperature may have influence on methylation of genomic DNA. *Poult. Sci.* 94, 2537–2545. doi: 10.3382/ps/pev201
- Yu, L. L., Gao, T., Zhao, M. M., Lv, P. A., Zhang, L., Li, J. L., et al. (2018). Effects of *in ovo* feeding of l-arginine on breast muscle growth and protein deposition in post-hatch broilers. *Animal* 12, 2256–2263. doi: 10.1017/S1751731118000241

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Andrieux, Petit, Collin, Houssier, Métayer-Coustard, Panserat, Pitel and Coustham. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.