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Prevalence, risk factors and metabolic predictors associated with subclinical ketosis on dairy cattle in Batna, Eastern of Algeria

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Abstract

Economic impact of subclinical ketosis (SCK) in dairy herds resulted from a decreased productivity, reproductive performances, and health condition. In Algeria, data availability regarding the prevalence of SCK in dairy cows were limited. The present study is a contribution in that direction and aimed to determine the prevalence of SCK between 2 and 50 days postpartum whereas investigating herd- and cow-level risk factors and the metabolic profile associated, on two bovine populations imported and native. The study was conducted with 16 typical representative local dairy cow herds ($n = 100$; 2–12 years old; located at Merouana, Batna Province) with data collected in a period comprised between August and February of the following year. Concentration of cows' plasma beta-hydroxybutyrate (BHBA) was used for the diagnosis, with values equal or higher than 1.2 mmol/L chosen as the threshold for SCK detection. Other key associated zootechnical parameters such as body condition score (BCS) were monitored at the same sampling points. Logistic regression analyses were carried out to characterize association between parameters. The average SCK prevalence was 9% and resulted highest during first- and second-week postcalving, with a peak in the first week. The herd size (< 30 lactating cows) and BCS (≤ 3) were risk factors associated with SCK prevalence. At the cow-level, risks of SCK increased in thin individuals ($P = 0.058$; odds ratio (OR) = 5.00, 95% confidence interval (CI) = 0.95–26.32). At the herd-level, risks of SCK increased in smaller frame size herds ($P = 0.047$; OR = 8.76, 95% CI = 1.03–74.49). Most cases of SCK were observed in native cows compared to imported. Interestingly, linear stepwise regression showed a good relationship between BHBA and total bilirubin ($P = 0.0001$; $R^2 = 0.59$). There were significant differences between healthy and ketotic cows regarding plasma concentration of bilirubin ($P = 0.003$), glucose ($P = 0.001$), alanine aminotransferase (ALT) ($P = 0.008$), creatinine ($P = 0.007$), and sodium ($P = 0.02$). According to receiver operating characteristic (ROC) curve analysis, the best parameter for SCK prediction is total bilirubin with more than 4.09 g/L as a cutoff point (77.78% sensitivity and 94.51% specificity). According to metabolic profile and feeding data we suggest that most cases of SCK observed in our study are type I. Further studies are required to validate other metabolic predictors for SCK and investigate adaptation of imported cows in semiarid regions in Algeria.

Keywords Subclinical ketosis · Dairy cattle · Prevalence · Risk factors · Metabolic profile

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Introduction

The cow peripartum or transition period goes from the 3 weeks before and 3 weeks after the calving date. This key period is a strong determinant of the full lactation success regarding cows' health and performance (Duffield et al. 2009). In early lactation, negative energy balance (NEB) is a common phenomenon in high yielding cattle, in which energy requirement for milk production increase and exceed energy intake. The normal adaptive response of cows through the peripartum involves complex and synchronized physiological adjustments in energy intake and basal energy requirements for maintenance. In contrast, a poor or a limited adaptive capacity response might alter homeostasis and induce NEB, which responsible of increased risks of possible periparturient diseases (Herdt 2000). A prolonged NEB deemed to be associated with ketosis development (Ingvarstsen 2006).

Blood BHBA (beta-hydroxybutyrate) determination was considered as the gold standard test for ketosis diagnosis because of its predominance and stability in blood compared to other ketone bodies (acetone or acetoacetate) (Oetzel 2004). The common BHBA analysis in laboratory used for ketosis diagnosis are, however, time consuming and expensive for large sample sizes, as it's the case of carrying out diagnosis in herds belonging to real-producing farms. Recently, electronic hand-held devices, initially developed for human BHBA determination has been used as cowside ketosis diagnosis, and showed excellent test results (Iwersen et al. 2009; Voyvoda a Erdogan 2010).

Two different forms of hyperketonemia differing in their clinical, frequency and importance has been commonly reported, where clinical and subclinical ketosis (SCK) both result in increased concentrations of ketone bodies in the tissues and milk of the cows (Enjalbert et al. 2001). The clinical form of ketosis is generally easy to diagnose and has lesser incidence, whereas the subclinical form is more prevalent and even more difficult to detect.

The most commonly used cut-off for SCK definition in dairy cattle is blood, plasma, or serum BHBA concentration at the threshold value equal or exceed 1.2 mmol/L (Ospina et al. 2010; McArt et al. 2012; Suthar et al. 2013; Garro et al. 2014; Tatone et al. 2016). Previous research reported an overall prevalence of cow's SCK ranges from 8.9 to 34% in the first two months of lactation (Duffield 2000). Recently, the peak prevalence of SCK was observed in the first two weeks of lactation, ranging from 20 to 43% (Duffield et al. 2009; Chapinal et al. 2011; McArt et al. 2012; Suthar et al. 2013; Berge and Vertenten 2014; Tatone et al. 2016).

The SCK provokes significant direct and indirect economic losses in dairy herds, respectively by decreasing milk production and increasing the occurrence of peripartum diseases (Oetzel 2004). Furthermore, increased

concentration of BHBA can affect milk production and reproduction, and increase the risk of subsequent peripartum diseases (Duffield et al. 2009; McArt et al. 2013; Suthar et al. 2013).

Currently, limited data on the prevalence of SCK in dairy cows' herds in Algeria are available. The present study is a contribution in that direction and aimed to determine the prevalence of SCK in postpartum whereas investigating herd- and cow-level risk factors and the metabolic profile associated, on two bovine populations imported and native.

Materials and methods

Study area

The study was carried out in Merouana, District of Batna Province, located in the eastern of Algeria, between 35° 38' N latitude and 5° 55' E longitude, with area of 509.97 km² (Fig. 1). Merouana has a semiarid climate with an annual average of pluviometry varies between 416.7 and 376.8 mm. The veterinary services of the district record 120 herds owners, having approximately 2810 dairy cows, composed genetically of native or crossbred and exotic or purebred (Direction des Services Agricoles de la Wilaya de Batna 2020).

Sample size determination

The total sample size for the present study is determined using the formula of prevalence studies as follows: $n = \frac{Z^2 P(1-P)}{d^2}$ (Pourhoseingholi et al. 2013);

where n is the sample size, Z is the statistic corresponding to level of confidence, P is the expected prevalence obtained from previous study or a pilot study, and d is the precision corresponding to effect size. The confidence level and the precision used in our study were, respectively, 95% and 10%. The prevalence of SCK (50%) in the eastern of Algeria reported by Tlidi et al. (2004) was used as the expected prevalence value. By applying these values in the formula, the sample size calculated was 96, which was rounded to 100 cows to be selected in our study.

Study population

The sampled cows were checked clinically for the presence of metabolic and infectious peripartum diseases (clinical ketosis, displaced abomasum, mastitis, retained placenta, metritis, milk fever, and lameness). The study was conducted from August to February of the following year. The experimental cows ($n = 100$ clinically healthy; 2–12 years old) were randomly selected from 16 dairy herds around the District of Merouana (Batna Province, Algeria) and were finally



Fig. 1 Geographical location of Merouana District, Batna Province, Algeria

composed by native or crossbred cows ($n = 56$) and imported or exotic European purebred cows ($n = 44$; 23 Holstein and 21 Fleckvieh). The study cows were composed of 37 primiparous and 63 multiparous. The lactation number ranged from one to 10. All herds were overall composed by less than 50 cows without calving concentration strategy (the calving occurring throughout the year). During lactation, cows were milked mechanically and have their concentrate portions, twice a day.

Data collection

The body condition score (BCS) was determined at the sampling day by a single observer, using a 1 to 5 scale with 0.25 increments as described by Edmonson et al. (1989). At that visit, data related to animals and herds were also collected and organized respectively, based on the origin or the genetic of the cow (imported purebred or native crossbred), the breed, the number of lactations, the day in milk (DIM), and the herd size and feeding.

Feeding

Data about feeding components are summarized in Table 1. There are two types of diet composition formula according to the herd size. Medium herd owners prepare their concentrate formulation and distribute a variety of forage; however, small herds use a prepared concentrate-based diet and distribute only straw as forage along the year with additional hay offered in winter. The grazing system of small herds performed in two seasons: stubble grazing and dry pasture in summer and early autumn and green grazing in

spring; however, in medium herds, cows are raised indoors. All herds distribute forage and concentrate separately. Cows were fed 8 kg of concentrate, offered at two equally daily portions after milking.

Study design

The present work is a cross-sectional prevalence study, considering the individual cow as experimental unit and the prevalence of SCK as main outcome measurement. Hyperketonemia was defined as suggested by Ospina et al. (2010), and cows were considered as having SCK, if blood BHBA concentrations equal or exceed 1.2 mmol/L. Each cow was tested only once for BHBA blood level, between 2 and 50 DIM. 16 dairy herds were selected from the list of dairy herds recorded in the veterinary services of Merouana's district based on their willingness to participate and the number of cows.

Table 1 Components of feeding in the studied herds

Ingredient	Medium herds	Small herds
Concentrate composition	40 kg of corn	30 kg of corn
Quantity in 100 kg	15 kg of soy 22 kg of bran 22 kg of barley 1 kg of VMC	6 kg of soy 63 kg of bran 1 kg of VMC
Forage composition	15 kg of lucerne 15 kg of corn silage or oat Straw ad libitum	Hay of lucerne added in winter Straw ad libitum

VMC vitamin mineral complex

The SCK diagnosis

Two types of whole blood samples were collected from the caudal and the jugular vein of each cow, for respectively, BHBA and metabolic parameter determination. The samples were performed always at the same time, approximately 4 to 5 h after morning concentrate distribution. The first one was performed using a 19-gauge needle attached to a syringe of 2.5 mL draw. Immediately after collection, blood BHBA concentration was determined in situ using a β -ketone test strip coupled to a cow-side electronic meter (FreeStyle Optium®; Abbott Diabetes Care Ltd., Witney, UK) according to the manufacturer's instructions. The second one was collected in a vacutainer tube and let to clot for 60 min at room temperature prior to centrifugation. After a centrifugation at 3000 rpm for 10 min, the serum was aliquoted and stored at $-20\text{ }^{\circ}\text{C}$ until biochemical analysis.

Biochemical analysis

The stored serum was used to determine different parameters of metabolism (glucose, triglycerides, cholesterol, total protein, albumin, total bilirubin, urea, and creatinine), enzymes (aspartate aminotransferase (AST), alanine aminotransferase (ALT), and alkaline phosphatase (ALP)), and minerals (calcium, phosphorus, sodium, potassium, and chloride). Biochemical analyses were carried out using specific commercial diagnostic kits as follows: glucose, triglycerides, cholesterol, total protein, albumin, total bilirubin, urea, and creatinine (Cypress Diagnostics, Belgium); total bilirubin (Roche Diagnostics, Germany); ALT, AST, and PAL (Cypress Diagnostics, Belgium); and calcium and phosphorus (Spinreact, Spain).

The biochemical analyses were performed through an automated chemistry analyzer Mindray BS 200 (Bio-Medical Electronics Co., Ltd, China) according to protocols of the manufacturer. Serum sodium, potassium, and chloride were determined using Easylyte plus Na/K/Cl electrolyte analyzer (Medica Corporation, USA), a fully automated system based on ion selective electrode (ISE) method, using solution pack kit.

Statistical analysis

The data were analyzed using the statistical software SPSS version 20.0. All variables were presented as mean and standard of deviation (SD) or median and interquartile range (IQR) according to data distribution, normally and non-normally, respectively. A value of $P \leq 0.05$ was considered significant. The BHBA concentration was considered as a continuous variable. Student *t* test was performed to compare the variation of metabolic blood values between cows with and without SCK. Pearson's correlation analyses were also conducted to evaluate relationships between blood BHBA and other metabolic parameters.

For logistic regression analyses, qualitative variables were dichotomised. The DIM was categorized as weeks. The origin or genetic of cows was categorized as imported or purebred ($n=44$) and native or crossbred ($n=56$), and the breed of cows as Holstein ($n=23$) and Fleckvieh ($n=21$). The rang of lactation was categorized as 1 (primiparous) and ≥ 2 (multiparous). The BCS of cows was categorized as thin (≤ 3.0) and fair (> 3.0). The herds were classified according to their size into two herd-size classes: small (10–29), medium (30–50). Moreover, for linear regression analyses, blood BHBA values were converted to mg/dL (initial unit was mmol/L) to avoid values of zero points, which are difficult to normalize. Afterwards, the data were normalized using a two-step approach for transforming continuous variables to normal (Templeton 2011).

For categorical variables, a chi-square test was performed to validate potential risk factors associated with SCK results; significant variables ($P \leq 0.05$) were subjected to binary logistic regression. The SCK disease (binary variable) was considered as a dependent variable and the factors as independent variables. The final model was verified with the Hosmer and Lemeshow goodness of fit test (Hosmer et al. 2013). However, a linear stepwise regression was performed for numerical variables. The normalized BHBA was included in the model as dependent variable and all other metabolic variables were included as independent variables. As a final step in stepwise regression (forward stepwise elimination), all variables selected previously with a $P \geq 0.05$ were excluded. Collinearity between independent variables was verified in the two previously cited models by a correlation analysis, where variables with a strong collinearity (correlation coefficient > 0.9) were excluded from the multiple analysis.

Finally, the cut-off points or critical thresholds of metabolic predictors for SCK diagnosis (dependent binary variable) were determined using receiver operating characteristic (ROC) analysis, where significant variables in correlation and regression analysis were subjected to this analysis. The area under the curve (AUC) was performed to evaluate the diagnostic precision of the selected parameters and only significant variables were selected for prediction (H_0 : $\text{AUC} = 0.5$ versus H_1 : $\text{AUC} \neq 0.5$; $P \leq 0.05$). Youden's index was calculated to determine the optimal thresholds of the significant parameters.

Results

Descriptive statistics

The mean \pm SD of determined BHBA across all cows was $0.65 (\pm 0.43)$ mmol/L ranging from 0.1 to 2.5 mmol/L. The blood BHBA concentration in cows with and without SCK were respectively, $1.72 (\pm 0.55)$ mmol/L and $0.55 (\pm 0.23)$ mmol/L. The median (IQR) day for SCK diagnosis was 7 (3)

Table 2 Univariate analysis of categorical variables for risk factors associated with subclinical ketosis (SCK)

Factors	Comparison level	Cows without SCK (total <i>n</i> =91)	Cows with SCK (total <i>n</i> =9)	<i>P</i> value
Parity	Primiparous	35	2	0.478
	Multiparous	56	7	
BCS	≤ 3.0 “thin”	37	7	0.041
	> 3.0 “fair”	54	2	
Genetic	Native	49	7	0.292
	Imported	42	2	
Breed	Holstein	21	2	0.489
	Fleckvieh	21	0	
Herd size	Small	43	8	0.031
	Medium	48	1	

*Variables selected and used in the multiple analysis ($P \leq 0.05$)

DIM. The median (IQR) lactation number was 2 (2) and 3 (3) in non-SCK and SCK cows, respectively. The median (IQR) BCS was 2.75 (1) in ketotic cows and 3 (1) in healthy cows.

Prevalence

The SCK was defined in the present study as BHBA concentrations higher or equal to 1.2 mmol/L. At this cutoff point, the prevalence of SCK was 9% between 2 and 50 DIM. Using alternative thresholds such as 1.0, 1.4, and 1.7 mmol/L for SCK definition, the prevalence became respectively, 14%, 6%, and 4%.

All diagnosed cases of SCK have been observed in the first three weeks post-calving. The peak prevalence of SCK was observed during the first week followed by the second week at 55.6% and 33.3%, respectively, while the remaining cases (11.1%) were detected in the third week.

Logistic regression analysis (risk factors for SCK)

The results of independence analysis are presented in Table 2 and those of logistic regression in Table 3. Variables subjected to risk analysis were: parity, BCS, breed, genetic

Table 3 Final logistic regression model of the association between BCS and herd size with the risk of SCK occurrence

Factors	Levels	Estimate	S.E	OR	95% CI	<i>P</i> value
BCS	Fair	Referent				
	Thin	1.61	0.85	5.00	0.95–26.32	0.058
Herd size	Medium	Referent				
	Small	2.17	1.09	8.76	1.03–74.49	0.047
Constant		−4.83	1.20	0.01		0.000

Hosmer and Lemeshow test: chi-square = 3.879; *df* = 2; $P = 0.144$; $R^2 = 0.224$

and herd size. In total, one herd-level factor and four animal-level factors. In the final model, the risk factors associated with SCK were: herd size < 30 lactating cows and BCS ≤ 3.

Parity

The chi-square test did not show any significant differences between multiparous and primiparous regarding the occurrence of SCK ($P = 0.041$; Table 2). As can be seen in Table 1, the prevalence of SCK seems increasing in multiparous cows (7%) compared to primiparous cows (2%).

BCS

The BCS was assigned to each cow once by visual assessment. Interestingly, there is a significant effect of SCK on BCS ($P = 0.478$; Table 2). Thin cows (BCS ≤ 3.0) were at significantly higher risk for developing SCK compared to fair cows ($P = 0.058$; OR = 5.00, 95% CI = 0.95–26.32; Table 3).

Breed

The breed of cow did not affect the occurrence of SCK ($P = 0.489$; Table 2). It can be seen from the data in Table 2 that all imported purebred cows diagnosed with SCK were of Holstein breed.

Genetic

The results did not detect any significant difference related to the origin of cows on the occurrence of SCK ($P = 0.292$; Table 2). The SCK seems to be more common in native cows (7 cases) than in cows imported (2 cases).

Herd size

There was a significant difference between SCK occurrence and herd size ($P = 0.031$; Table 2). Our findings also indicate

Table 4 Univariate analysis of metabolic profile parameters comparison between ketotic and healthy cows and correlation between blood BHBA concentration and other metabolic parameters

Metabolic parameters	Independent-samples <i>t</i> test			Pearson correlation test	
	SCK cows	Healthy cows	<i>P</i> value	Blood BHBA	
	Mean ± σ	Mean ± σ		Corr. coef	<i>P</i> value
Albumin (g/L)	27.19 ± 5.17	25.55 ± 4.44	0.33	-0.12	0.231
Total protein (g/L)	66.16 ± 12.19	69.44 ± 15.55	0.55	0.18	0.068
Total bilirubin (g/L)	2.71 ± 1.05	4.87 ± 1.62	0.003**	0.52	0.0001***
AST (IU/L)	73.09 ± 30.85	76.66 ± 19.18	0.62	0.09	0.359
ALT (IU/L)	21.27 ± 11.69	14.00 ± 6.14	0.008**	-0.05	0.634
ALP (IU/L)	48.91 ± 25.18	39.77 ± 12.29	0.08	0.04	0.701
Urea (g/L)	0.24 ± 0.12	0.27 ± 0.08	0.40	0.05	0.597
Creatinine (g/L)	13.02 ± 3.03	10.92 ± 1.74	0.007**	-0.08	0.413
Cholesterol (g/L)	0.94 ± 0.36	1.15 ± 0.38	0.15	-0.04	0.711
Triglycerides (g/L)	0.16 ± 0.04	0.20 ± 0.10	0.28	0.22	0.027*
Glucose (g/L)	0.61 ± 0.18	0.44 ± 0.10	0.001**	-0.04	0.711
Phosphorus (mg/L)	48.71 ± 10.61	48.33 ± 5.65	0.86	0.11	0.276
Calcium (mg/L)	83.81 ± 15.22	83.11 ± 31.02	0.94	-0.03	0.773
Sodium (mEq/L)	152.70 ± 18.46	145.25 ± 6.38	0.02*	-0.10	0.326
Potassium (mEq/L)	4.55 ± 0.74	4.53 ± 0.43	0.91	0.01	0.949
Chloride (mEq/L)	80.62 ± 10.67	82.00 ± 4.30	0.48	0.23	0.023*

Corr. coef. correlation coefficient

that the prevalence of SCK was lesser in medium-sized herds with 1 case against 8 cases recorded in small herds. Small herds have more risk for SCK compared with medium size herds (OR = 8.76, 95% CI = 1.03–74.49, $P = 0.047$; Table 3).

Mean's comparison (metabolic status comparison between ketotic and healthy cows).

Metabolic parameters differences between cows with and without SCK are summarized in Table 4. When cows with SCK were compared to healthy cows, there were significant differences regarding bilirubin ($P = 0.003$), glucose ($P = 0.001$), ALT ($P = 0.008$), creatinine ($P = 0.007$), and sodium ($P = 0.02$). Total bilirubin showed higher values in cows with SCK compared to those without SCK, dissimilar to other significant parameters (glucose, ALT, creatinine and sodium) ketotic cows presented the lowest values.

Pearson's correlation coefficients and regression analysis (metabolic parameters association with blood BHBA)

The results of correlational analysis are presented in Table 4 and those of linear regression in Table 5. There was a positive correlation between BHBA and total bilirubin, triglycerides and chlorides with respective coefficients of correlation 0.52 ($P = 0.000$), 0.22 ($P = 0.027$), and 0.23 ($P = 0.023$) (Table 4). After a forward stepwise elimination, four parameters statistically significant were retained in the finale model, which are: total bilirubin, chloride, sodium and phosphorus (Table 5). Interestingly, a good positive correlation is observed between BHBA and total bilirubin

($R^2 = 0.59$, $P = 0.000$; Table 5). In Fig. 2, there is a clear linear association between blood BHBA and total bilirubin.

ROC analysis (metabolic predictor selection and analysis)

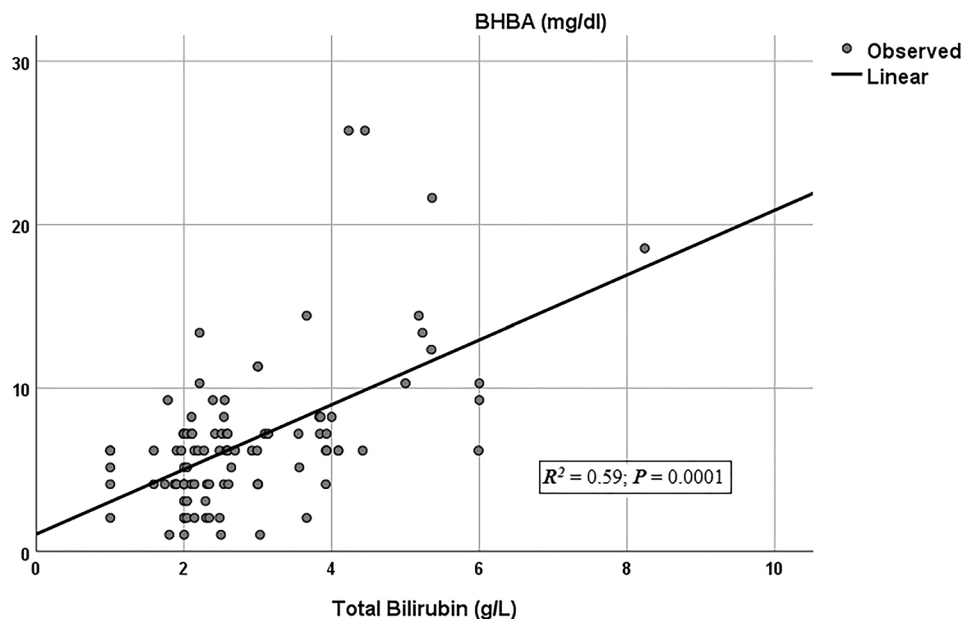
The metabolic predictors for SCK diagnosis were selected based on their coefficients in correlation and regression analysis, where the significant variables are selected. For prediction analysis only significant variables to AUC analysis (area > 5; $P \leq 0.05$) were retained. Total bilirubin was the only metabolic predictor (area = 0.890; $P = 0.0001$) with a discriminating ability for SCK detection. The optimal prediction threshold was determined by Youden's index ($J = 0.7228$; 95% CI = 0.3785 to 0.8559) for total bilirubin to be more than 4.09 g/L (95% CI = > 3.14 to > 5) with 77.78% sensitivity and 94.51% specificity (Table 6). Figure 3 presents the ROC plot of total bilirubin for SCK diagnosis.

Table 5 Association (final stepwise linear regression model) between BHBA and total bilirubin, chloride, chloride, and phosphorus

Variables	Estimate	S.E	Beta	<i>P</i> value
Total bilirubin	2.06	0.29	0.59	0.0001
Chloride	0.07	0.03	0.18	0.043
Sodium	-0.04	0.02	-0.17	0.034
Phosphorus	0.08	0.04	0.18	0.042
Constant	-2.43	4.05		0.549

Model summary: $F = 14.978$; $df = 4$; $P = 0.000$; $R^2 = 0.387$

Fig. 2 Association (fitted line) between blood BHBA and total bilirubin



Discussion

Literature data showed a wide variation in the reported prevalence of bovine ketosis. Previous studies indicate that prevalence of SCK ranges from 6.9 to 14.1% in the first 2 months of lactation (Dohoo and Martin 1984; Andersson and Emanuelson 1985; Nielen et al. 1994; Duffield et al. 1997), whereas, recently, the peak prevalence of SCK in dairy cattle was noticed in the first 2 weeks of lactation, ranging from 20 to 43% (Herdt 2000; Duffield et al. 2009; Chapinal et al. 2011; McArt et al. 2012; Suthar et al. 2013). Furthermore, a recent study covering wider geographic areas from different continents reported that SCK prevalence ranged approximately from 8 to 40% (Brunner et al. 2019). This variation of SCK prevalence across different countries is related to numerous factors, such as, the test characteristics (sensitivity and specificity), the threshold used for defining SCK, the screening period in post-calving (e.g., first week or month), the scheme of monitoring (testing once, twice or more) and test intervals (e.g., week, more or less).

The overall prevalence of SCK detected in our research was 9% between 2 to 50 DIM. Recently, it has been shown that the median time for SCK resolution is about 5 days; consequently, the weekly monitoring might underestimate the real incidence of SCK (McArt et al. 2012). Assessing the real incidence of SCK require testing twice or more in a week, because the weekly screening can potentially lead to development and resolution of SCK between test intervals.

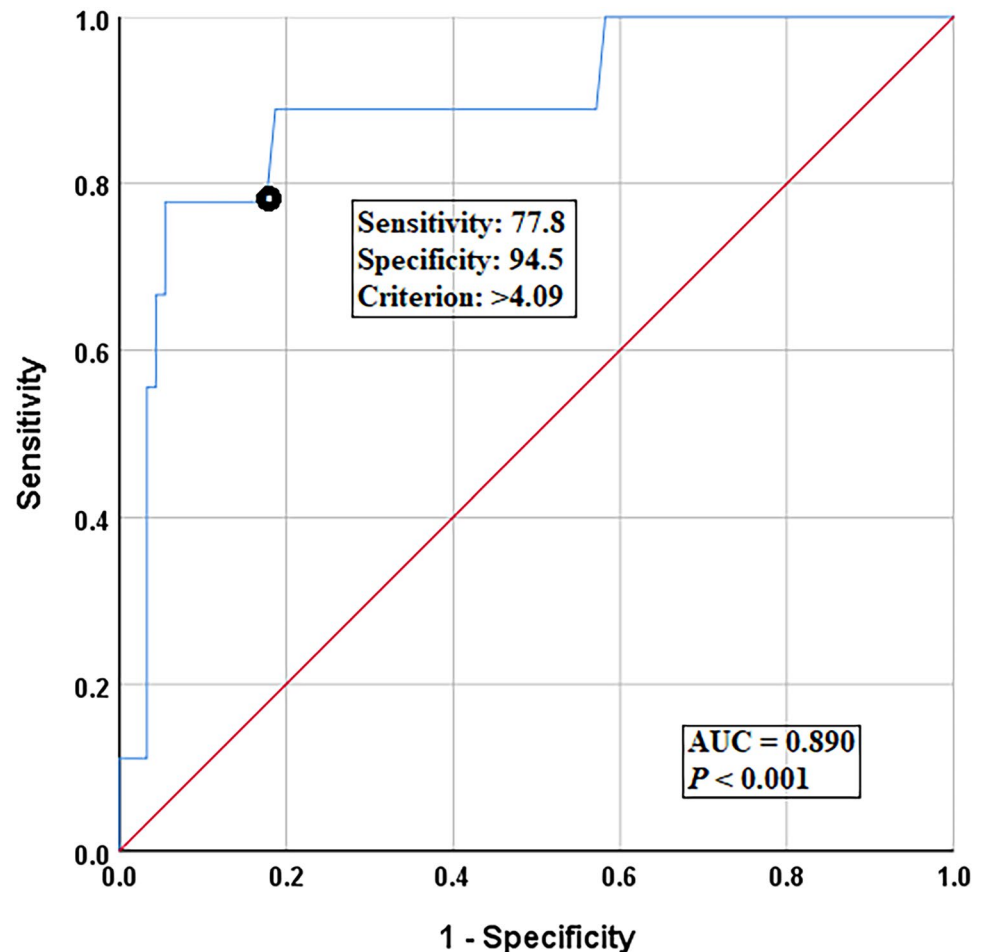
Reported prevalence rates range from 11.2 to 36.6% in Western Europe (Suthar et al. 2013), from 26 to 56% in North America (McArt et al. 2012), and from 8 to 40% in 12 different countries worldwide (Brunner et al. 2019). This variation is related to the difference in the scheme of sampling, the period of monitoring and the definition of SCK. Further explanations for the elevated incidence observed in Western productions systems is the higher milk production objectives. Garro et al. (2014) and Samiei et al. (2013) reported SCK prevalence of 10.3% and 12.2%, respectively, which are about the same as reported in our study and share the similar protocol for sampling and defining SCK. Similarly, a recent study reported a

Table 6 Prediction threshold, Se, Sp, +LR, -LR, NPV, PPV, Acc, and area under the ROC curve for total bilirubin in cow SCK diagnosis

Parameter	Statistic	Value	95% CI
Total bilirubin threshold (> 4.09 g/L)	Sensitivity (Se)	77.78%	39.99 to 97.19%
	Specificity (Sp)	94.51%	87.64 to 98.19%
	Positive likelihood ratio (+LR)	14.16	5.64 to 35.55
	Negative likelihood ratio (-LR)	0.24	0.07 to 0.80
	Positive predictive value (PPV)	58.33%	35.79 to 77.86%
	Negative predictive value (NPV)	97.73%	92.68 to 99.32%
	Accuracy (Acc)	93.00%	86.11 to 97.14%

AUC=0.890; SE=0.060; Z=6.157; P=0.001; 95% CI=0.812 to 0.944

Fig. 3 Curve receiver operating characteristic (ROC) plot of total bilirubin for diagnosis of SCK in dairy cattle



SCK prevalence of less than 10% in 2 countries (Australia and Colombia) out of 12 (Brunner et al. 2019). This later explain this difference in SCK prevalence at the level of countries by feeding and genetic variation, milk production objectives and management differences in dairy herds.

All cases of SCK have been diagnosed in the first three weeks post-calving, with a peak during the first and second weeks, followed by the third week. As reported previously, the peak prevalence of SCK occurred during the third and fourth week (Dohoo and Martin 1984; Andersson and Emanuelson 1985). Recently, it has been reported that the peak prevalence of SCK was mostly seen in the first two weeks postpartum (McArt et al. 2012; Suthar et al. 2013; Tatone et al. 2016). This recent finding corroborate with our outcomes and with those of Duffield (2000), who stated that advanced metabolic challenge closer to calving is a consequence of progress in genetics and feeding management.

Important risk factors influencing the occurrence of SCK include parity, BCS, genetics, herd and season. Herd-level differences regarding the prevalence of ketosis has been well documented (Duffield 2000).

In this study, the SCK prevalence was higher in multiparous cows but without significant effect. Most studies reported an increased risk of SCK occurrence in cows with higher parity (≥ 3) (McArt et al. 2012; Suthar et al. 2013; Berge and Vertenten 2014; Garzón-Audor and Oliver-Espinosa 2019). This may be related to the higher NEB and lipid mobilization in multiparous than primiparous cows, in which energy depletion is related to both concomitant physiologic stages (gestation and lactation) (Berge and Vertenten 2014).

The present results suggest that thin cows (BCS ≤ 3.0) are more prone to develop hyperketonemia than fair cows. Dissimilar to our outcomes, cows leading to overcondition (fatty), with BCS ≥ 3.5 , have an increased risk of ketosis in early lactation (Gillund et al. 2001; McArt et al. 2015; Seifi et al. 2011). Moreover, it has been reported recently that cows with BCS ≥ 3.0 at calving have an increased risk of ketosis associated with enhanced lipolysis, leading to greater plasma concentrations of BHBA (Garzón-Audor and Oliver-Espinosa 2019). However, in early lactation, it's natural to think that thin cows have low-fat reserves, which make them unable to manage the higher energy and nutrient requirements made by milk production. Also, cows highly

supplemented with simple sugars, could produce more BHBA not because of lipid mobilization (they don't need because they are not in NEB) but because of feeding (feed source BHBA).

The prevalence of SCK was significantly higher in small sized herds compared to medium size herds. Similarly, Berge and Vertenten (2014) reported a negative association between the size of lactating herd and the prevalence of ketosis. According to the same authors, herds with greater size have more pens related to lactation stages with specific dietary recommendations and higher nutritional services. Recently, it has been shown that large lactating herds were at higher risk for ketosis development (Garzón-Audor & Oliver-Espinosa 2019). This later explains that large herds raise high lactating cows to maximize milk production as a consequence SCK risk increases. According to these contradictory findings we suppose that herd size can be a confounding factor because it is more related to herd management and feeding as well as individual cow variation (genetic selection).

The cow's breed did not affect the occurrence of SCK contrary to a previous (Andersson and Emanuelson 1985), and a more recent study who demonstrated that Jersey breed was more susceptible to ketosis compared to Holstein (Tatone et al. 2016). Similar to our results, other authors found no association between breed and ketosis occurrence (Berge and Vertenten 2014; Garzón-Audor and Oliver-Espinosa 2019).

The present research demonstrated that SCK was more common in native cows than in cows imported but without significant effect. All cases of SCK in native cows were observed in small herds and the most of imported cows were raised in medium herds with a good feeding and housing compared to small herds. We especulate that this genetic variation in SCK susceptibility could be more related to feeding practices. Further explanation for lower cases of SCK in exotic cows is their adaptability to local environmental conditions (heat stress and lack of forage). As we hypothesized, van der Drift et al. (2012) evokes that environmental factors affect more the ethology of hyperketonemia in herds compared to genetic aspects. We think that under harsh conditions, cows with high production potential might adapt by reducing milk than expected based on their breeding value, but further investigations are needed.

The concentration of glucose was significantly lower in ketotic cows than healthy. Similarly, other authors have reported a significant lower glucose level in ketotic cows and compared to healthy (Sun et al. 2015; Cao et al. 2017; Yang et al. 2019). In the same view, a recent study showed that hypoglycemia affects roughly one-third of hyperketonemic cows (Dubuc and Buczinski 2018). Moreover, statistically significant higher blood levels of total bilirubin were found in ketotic cows. In the same vein, the blood total bilirubin levels were higher in cows with SCK compared to those

without (Sun et al. 2015; Yang et al. 2019). The significant difference observed in glucose and total bilirubin may be related in part to anorexia and in other part to the stressing effect of lipomobilization and ketone bodies accumulation on liver; this hepatic accumulation may decrease the processes of gluconeogenesis for glucose and reduce the excretory capacity for total bilirubin. However, the blood ALT, creatinine and sodium values recorded in the present study were slightly lower in hyperketonemic cows but within physiological limits and without pathophysiological signification.

Pearson's correlation coefficients and linear regression analysis demonstrated a reliable association between BHBA and total bilirubin. This finding supports previous research, which reported a good positive correlation between acetoacetate and total bilirubin (Kauppinen 1984; Ropstad et al. 1989). Our findings regarding relations between BHBA and bilirubin but not between BHBA and ASAT are consistent with results reported by Pechová and Nečasová (2018).

An important finding in our work was the excellent discriminating ability of total bilirubin for SCK diagnosis using the threshold 4.09 g/L (77.78% sensitivity and 94.51% specificity). Sun et al. (2015) reported that total bilirubin concentration more than 3.3 mmol/L indicates abnormal liver function and can also be considered as a predictor for a high risk of ketosis (58% sensitivity and 83% specificity). This later considered total bilirubin as indicator reflecting the capacity of the liver for excretion, secretion and detoxication, which is in agreement with our results.

According to the metabolic profile which indicates a lower glycemia and higher total bilirubin in ketotic cows and higher cases of SCK in small herds in which the quality of feeding is lower compared to medium herds, we suggest that observed hyperketonemia in our study is type I which related to low energy density of rations.

Conclusion

This study allowed obtaining first report on the prevalence of SCK and the associated risk factors in Algeria. The prevalence was 9% and lower BCS, advanced parity and decreased herd size were considered as major risk factors for developing SCK in dairy cows in the early lactation (2 to 50 DIM). Also, SCK occurs mostly within the first 2 weeks postpartum (with a peak at the first week), which make this period the best for monitoring to maximize detection and reduce negative economic impacts. At the metabolic level, cows with SCK presented a lower glycemia and a higher total bilirubin level compared to healthy ones. Interestingly, a good positive correlation is observed between BHBA and total bilirubin, that showed the best prediction potential of SCK. We suggest that ketosis type I is the dominated one in our study. Further studies are required to validate other

metabolic predictors for SCK diagnosis and investigate on feeding and adaptation of imported highly yielding cows in semiarid regions in the eastern of Algeria.

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Author contribution A. Azizi, K. Deghnouche and M. Tlidjane: conceptualization, methodology. A. Azizi and A. Achouri: farm visit and sampling. Azizi A. and K. Ghouguel: statistical analysis. K. Deghnouche: laboratory analysis. A. Azizi: writing—original draft preparation. A. Azizi and E. González-García: writing—reviewing and editing.

Availability of data and material The datasets generated during the current study are available from the corresponding author on reasonable request.

Code availability Not applicable.

Declarations

Ethical approval The sampling and data collection in this study was performed by certified veterinarians that respected the ethical recommendations of the Algerian government.

Consent to participate The authors get, from all animal owners, verbal informed consent prior to the sampling and the data recording at level of animal and farm.

Consent for publication All authors read and consent the publication of the manuscript.

Conflict of interest The authors declare no competing interests.

References

- Andersson L, Emanuelson U (1985) An epidemiological study of hyperketonaemia in Swedish dairy cows; determinants and the relation to fertility. *Prev Vet Med* 3(5):449–462
- Berge AC, Vertenten G (2014) A field study to determine the prevalence, dairy herd management systems, and fresh cow clinical conditions associated with ketosis in western European dairy herds. *J Dairy Sci* 97(4):2145–2154. <https://doi.org/10.3168/jds.2013-7163>
- Brunner N, Groeger S, Canelas Raposo J, Bruckmaier RM, Gross JJ (2019) Prevalence of subclinical ketosis and production diseases in dairy cows in Central and South America, Africa, Asia, Australia, New Zealand, and Eastern Europe. *Translational Animal Science* 3(1):84–92. <https://doi.org/10.1093/tas/txy102>
- Cao Y, Zhang J, Yang W, Xia C, Zhang H-Y, Wang Y-H, Xu C (2017) Predictive value of plasma parameters in the risk of postpartum ketosis in dairy cows. *Journal of Veterinary Research* 61(1):91–95. <https://doi.org/10.1515/jvetres-2017-0011>
- Chapinal N, Carson M, Duffield TF, Capel M, Godden S, Overton M, Santos JEP, LeBlanc SJ (2011) The association of serum metabolites with clinical disease during the transition period. *J Dairy Sci* 94(10):4897–4903. <https://doi.org/10.3168/jds.2010-4075>
- Direction des Services Agricoles de la Wilaya de Batna (2020) Rapport d'activité annuelle (Accessed on January 2021).
- Dohoo IR, Martin SW (1984) Subclinical ketosis: prevalence and associations with production and disease. *Can J Comp Med* 48(1):1
- Dubuc J, Buczinski S (2018) Short communication: Cow- and herd-level prevalence of hypoglycemia in hyperketonemic postpartum dairy cows. *J Dairy Sci* 101(4):3374–3379. <https://doi.org/10.3168/jds.2017-13773>
- Duffield T (2000) Subclinical ketosis in lactating dairy cattle. *The Veterinary clinics of North America. Food animal practice* 16(2):231–53, v.
- Duffield TF, Kelton DF, Leslie KE, Lissimore KD, Lumsden JH (1997) Use of test day milk fat and milk protein to detect subclinical ketosis in dairy cattle in Ontario. *The Canadian Vet J. La revue vétérinaire canadienne* 38(11):713–718.
- Duffield TF, Lissimore KD, McBride BW, Leslie KE (2009) Impact of hyperketonemia in early lactation dairy cows on health and production. *J Dairy Sci* 92(2):571–580. <https://doi.org/10.3168/jds.2008-1507>
- Edmonson AJ, Lean IJ, Weaver LD, Farver T, Webster G (1989) A body condition scoring chart for Holstein dairy cows. *J Dairy Sci* 72(1):68–78
- Enjalbert F, Nicot MC, Bayourthe C, Moncoulon R (2001) Ketone bodies in milk and blood of dairy cows: relationship between concentrations and utilization for detection of subclinical ketosis. *J Dairy Sci* 84(3):583–589. [https://doi.org/10.3168/jds.S0022-0302\(01\)74511-0](https://doi.org/10.3168/jds.S0022-0302(01)74511-0)
- Garro CJ, Mian L, Cobos Roldán M (2014) Subclinical ketosis in dairy cows: prevalence and risk factors in grazing production system. *J Anim Physiol Anim Nutr* 98(5):838–844
- Garzón-Audor A, Oliver-Espinosa O (2019) Incidence and risk factors for ketosis in grazing dairy cattle in the Cundi-Boyacencian Andean plateau. *Colombia Tropical Animal Health and Production* 51(6):1481–1487. <https://doi.org/10.1007/s11250-019-01835-z>
- Gillund P, Reksen O, Gröhn YT, Karlberg K (2001) Body condition related to ketosis and reproductive performance in Norwegian dairy cows. *J Dairy Sci* 84(6):1390–1396. [https://doi.org/10.3168/jds.S0022-0302\(01\)70170-1](https://doi.org/10.3168/jds.S0022-0302(01)70170-1)
- Herdth TH (2000) Ruminant adaptation to negative energy balance. Influences on the etiology of ketosis and fatty liver. *The Veterinary clinics of North America. Food animal practice* 16(2):215–30, v.
- Hosmer DW, Lemeshow S, Sturdivant RX editors (2013) Applied logistic regression. 3rd ed. Wiley series in probability and statistics. Wiley, Hoboken.
- Ingvarstsen KL (2006) Feeding- and management-related diseases in the transition cow. *Anim Feed Sci Technol* 126(3–4):175–213. <https://doi.org/10.1016/j.anifeedsci.2005.08.003>
- Iwersen M, Falkenberg U, Voigtsberger R, Forderung D, Heuwieser W (2009) Evaluation of an electronic cowside test to detect subclinical ketosis in dairy cows. *J Dairy Sci* 92(6):2618–2624. <https://doi.org/10.3168/jds.2008-1795>
- Kauppinen K (1984) ALAT, AP, ASAT, GGT, OCT activities and urea and total bilirubin concentrations in plasma of normal and ketotic dairy cows. *Zentralbl Veterinarmed [a]* 31(8):567–576. <https://doi.org/10.1111/j.1439-0442.1984.tb01316.x>
- McArt JA, Nydam DV, Oetzel GR (2012) Epidemiology of subclinical ketosis in early lactation dairy cattle. *J Dairy Sci* 95(9):5056–5066. <https://doi.org/10.3168/jds.2012-5443>
- McArt JA, Nydam DV, Oetzel GR (2013) Dry period and parturient predictors of early lactation hyperketonemia in dairy cattle. *J Dairy Sci* 96(1):198–209. <https://doi.org/10.3168/jds.2012-5681>
- McArt JA, Nydam DV, Overton MW (2015) Hyperketonemia in early lactation dairy cattle: a deterministic estimate of component and total cost per case. *J Dairy Sci* 98(3):2043–2054. <https://doi.org/10.3168/jds.2014-8740>

- Nielen M, Aarts MG, Jonkers AG, Wensing T, Schukken YH (1994) Evaluation of two cowside tests for the detection of subclinical ketosis in dairy cows. *Can Vet J* 35(4):229
- Oetzel (GR) (2004) Monitoring and testing dairy herds for metabolic disease. *The Veterinary clinics of North America. Food animal practice* 20(3):651–674. <https://doi.org/10.1016/j.cvfa.2004.06.006>.
- Ospina PA, Nydam DV, Stokol T, Overton TR (2010) Association between the proportion of sampled transition cows with increased nonesterified fatty acids and beta-hydroxybutyrate and disease incidence, pregnancy rate, and milk production at the herd level. *J Dairy Sci* 93(8):3595–3601. <https://doi.org/10.3168/jds.2010-3074>
- Pechová A, Nečasová A (2018) The relationship between subclinical ketosis and ruminal dysfunction in dairy cows. *Annals of Animal Science* 18(4):955–971. <https://doi.org/10.2478/aoas-2018-0038>
- Pourhoseingholi MA, Vahedi M, Rahimzadeh M (2013) Sample size calculation in medical studies. *Gastroenterology and Hepatology from Bed to Bench* 6(1):14–17
- Ropstad E, Halse K, Refsdal AO (1989) Variations in parameters of liver function and plasma progesterone related to underfeeding and ketosis in a dairy herd. *Acta Veterin Scandi* 30(2).
- Samiei A, Liang JB, Ghorbani GR, Hirooka H, Ansari-Mahyari S, Sadri H (2013) Prevalence of ketosis and its correlation with lactation stage, parity and peak of milk yield in Iran. *Asian Journal of Animal and Veterinary Advances* 8(4):604–612
- Seifi HA, LeBlanc SJ, Leslie KE, Duffield TF (2011) Metabolic predictors of post-partum disease and culling risk in dairy cattle. *Vet J* 188(2):216–220. <https://doi.org/10.1016/j.tvjl.2010.04.007>
- Sun Y, Wang B, Shu S, Zhang H, Xu C, Wu L, Xia C (2015) Critical thresholds of liver function parameters for ketosis prediction in dairy cows using receiver operating characteristic (ROC) analysis. *Vet Q* 35(3):159–164. <https://doi.org/10.1080/01652176.2015.1028657>
- Suthar VS, Canelas-Raposo J, Deniz A, Heuwieser W (2013) Prevalence of subclinical ketosis and relationships with postpartum diseases in European dairy cows. *J Dairy Sci* 96(5):2925–2938. <https://doi.org/10.3168/jds.2012-6035>
- Tatone EH, Gordon JL, Hubbs J, LeBlanc SJ, DeVries TJ, Duffield TF (2016) A systematic review and meta-analysis of the diagnostic accuracy of point-of-care tests for the detection of hyperketonemia in dairy cows. *Prev Vet Med* 130:18–32. <https://doi.org/10.1016/j.prevetmed.2016.06.002>
- Templeton GF (2011) A two-step approach for transforming continuous variables to normal: implications and recommendations for IS research. *CAIS* 28. <https://doi.org/10.17705/1CAIS.02804>
- Tlidjane M, Alloui N, Deghnouche K, Alloui O (2004) Cas de cétose subclinique en Algérie. http://www.journees3r.fr/IMG/pdf/2004_Sante_14_Tlidjane.pdf. (Accessed 15 May 2014).
- van der Drift SGA, van Hulzen KJE, Teweldemedhn TG, Jorritsma R, Nielen M, Heuven HCM (2012) Genetic and nongenetic variation in plasma and milk β -hydroxybutyrate and milk acetone concentrations of early-lactation dairy cows. *J Dairy Sci* 95(11):6781–6787
- Voyvoda H, Erdogan H (2010) Use of a hand-held meter for detecting subclinical ketosis in dairy cows. *Res Vet Sci* 89(3):344–351. <https://doi.org/10.1016/j.rvsc.2010.04.007>
- Yang W, Zhang B, Xu C, Zhang H, Xia C (2019) Effects of ketosis in dairy cows on blood biochemical parameters, milk yield and composition, and digestive capacity. *Journal of Veterinary Research* 63(4):555–560. <https://doi.org/10.2478/jvetres-2019-0059>

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