



Une structure emboîtée universelle pour les interactions quantitatives entre les plantes et leurs parasites ?

Benoît Moury, Jean-Marc Audergon, Sylvie Baudracco-Arnas, Safa Ben Krima, François Bertrand, Nathalie Boissot, Mireille Buisson, Valérie Caffier, Melissa Cantet, Sylvia Chanéac, et al.

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Benoît Moury, Jean-Marc Audergon, Sylvie Baudracco-Arnas, Safa Ben Krima, François Bertrand, et al.. Une structure emboîtée universelle pour les interactions quantitatives entre les plantes et leurs parasites ?. Réunion annuelle du réseau E3GP3, Dec 2021, Visioconférence, France. hal-03553078

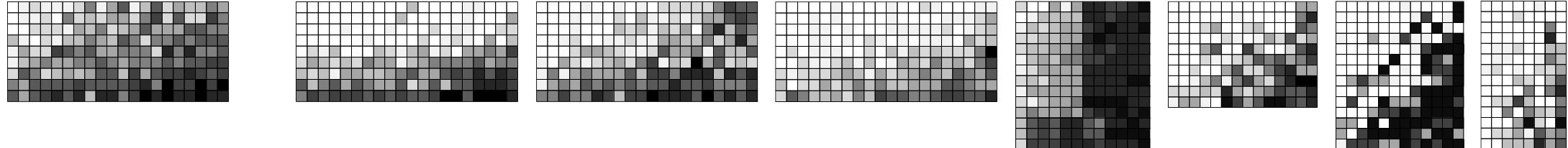
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Submitted on 3 Feb 2022

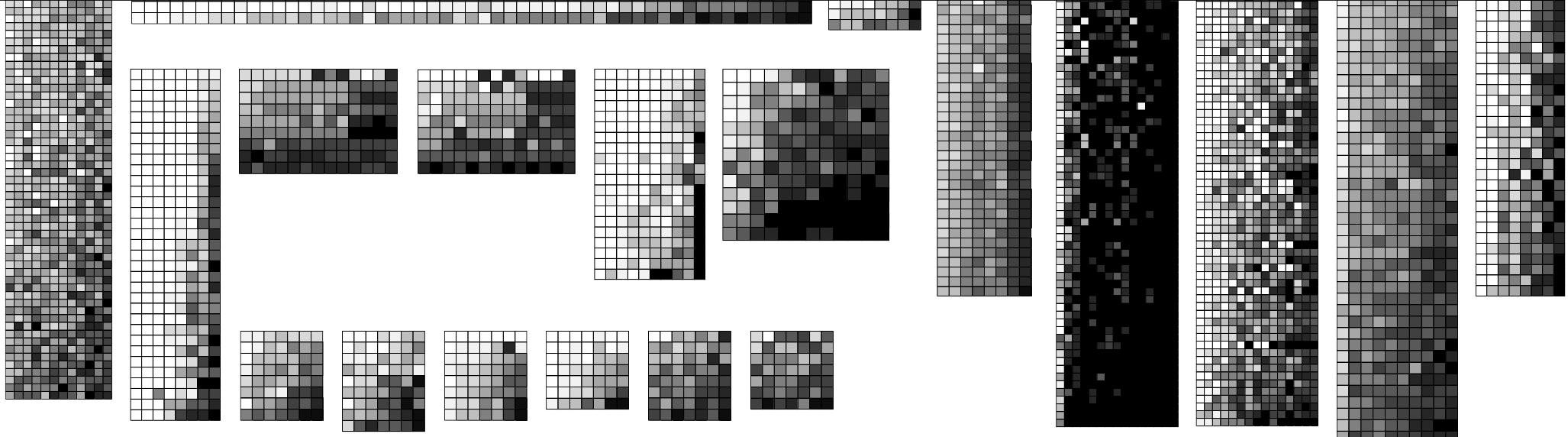
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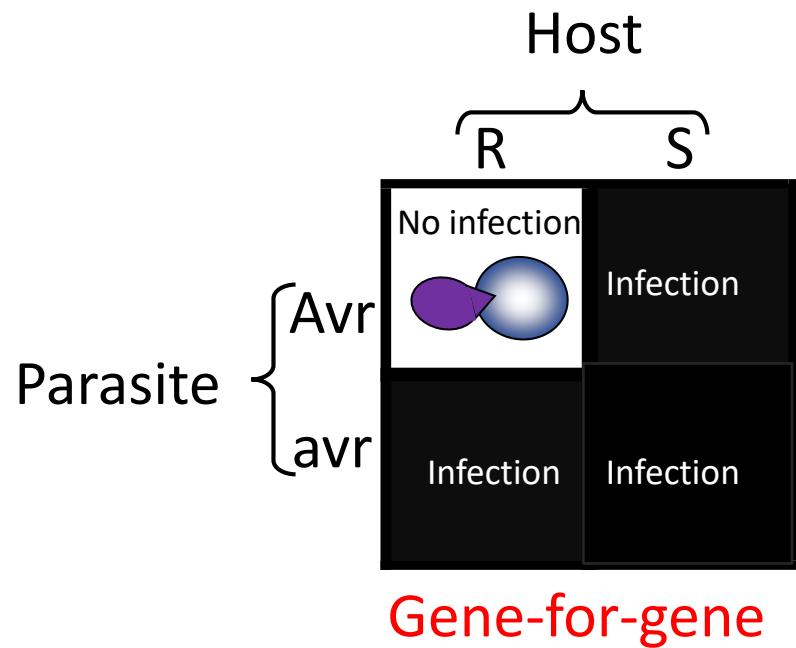


Many different recipes but one flavor

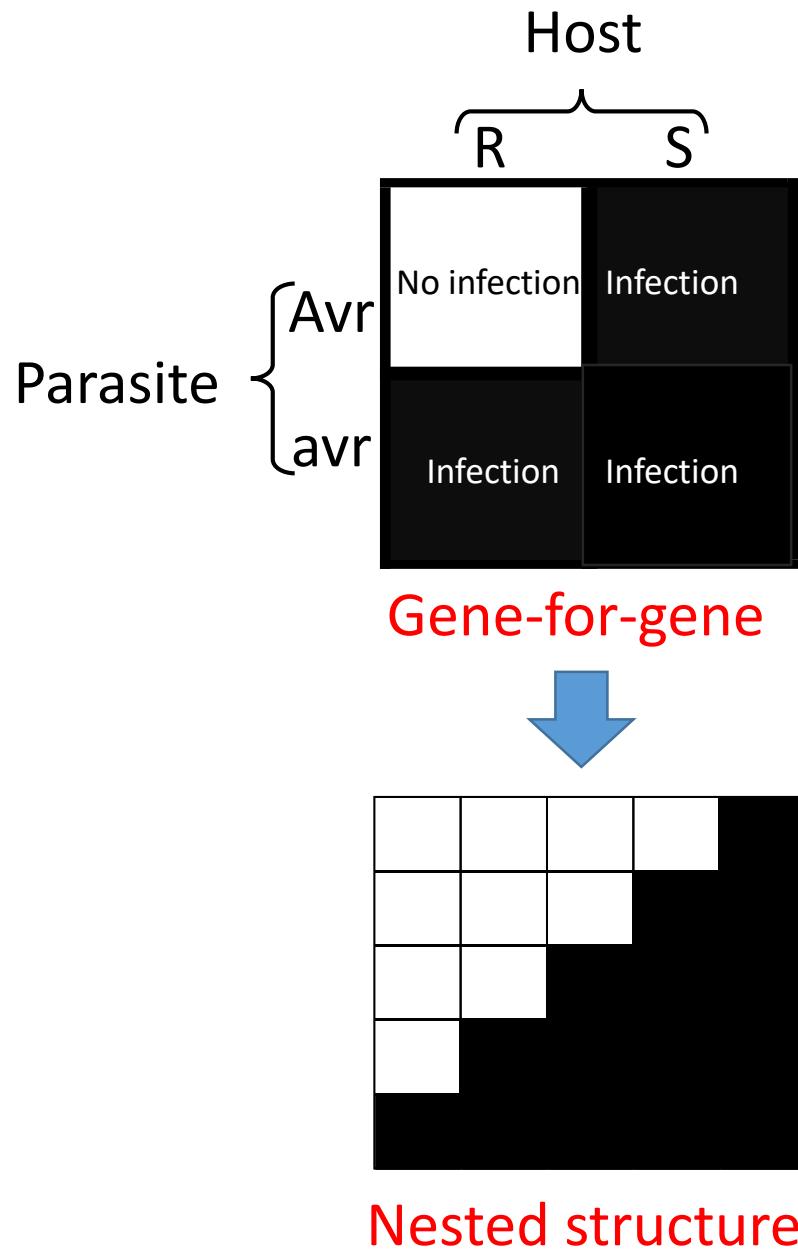
A universal nested structure of quantitative interactions
between plant genotypes and their parasites



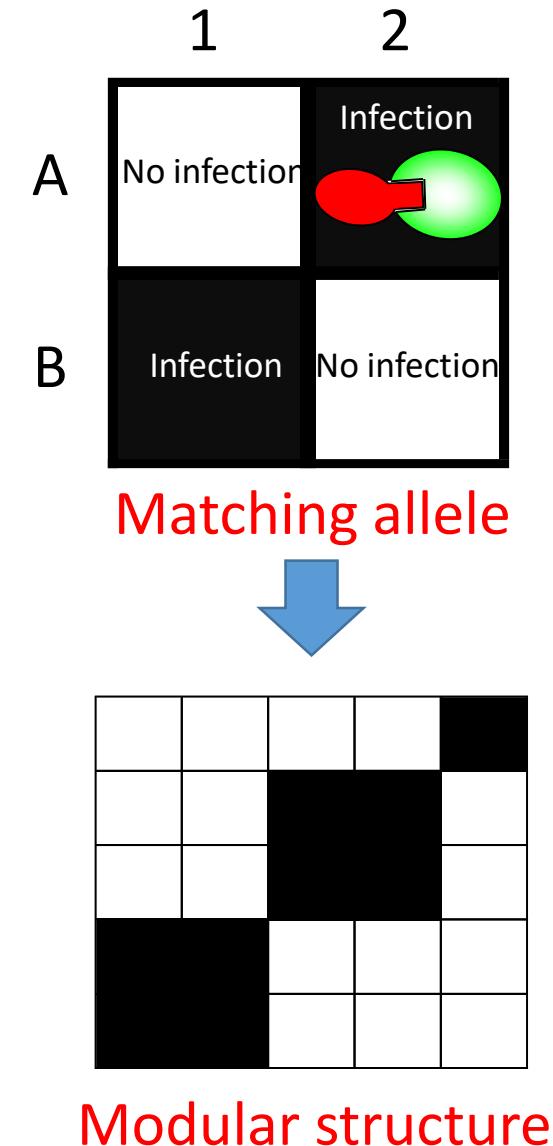
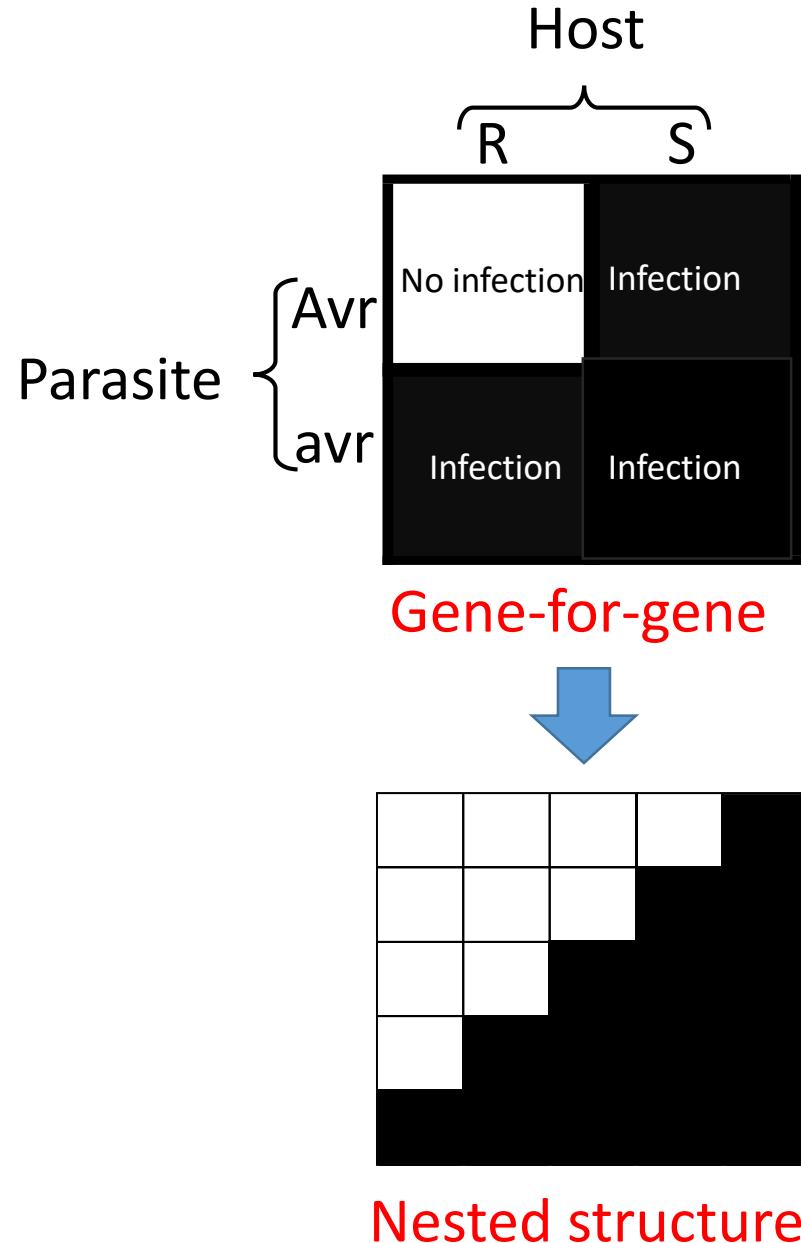
Qualitative plant immunity: two main genetic and evolutionary models



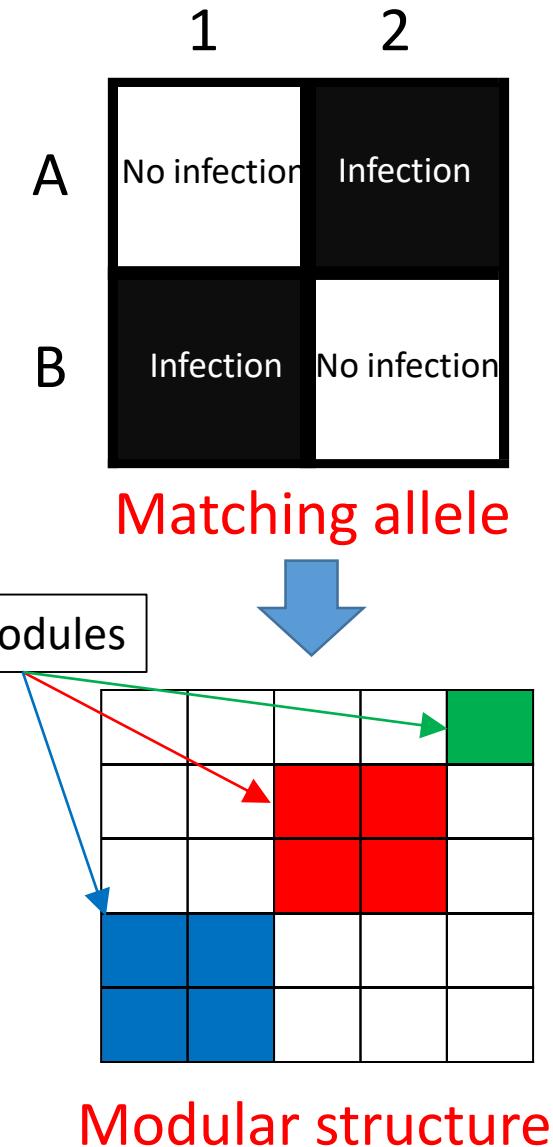
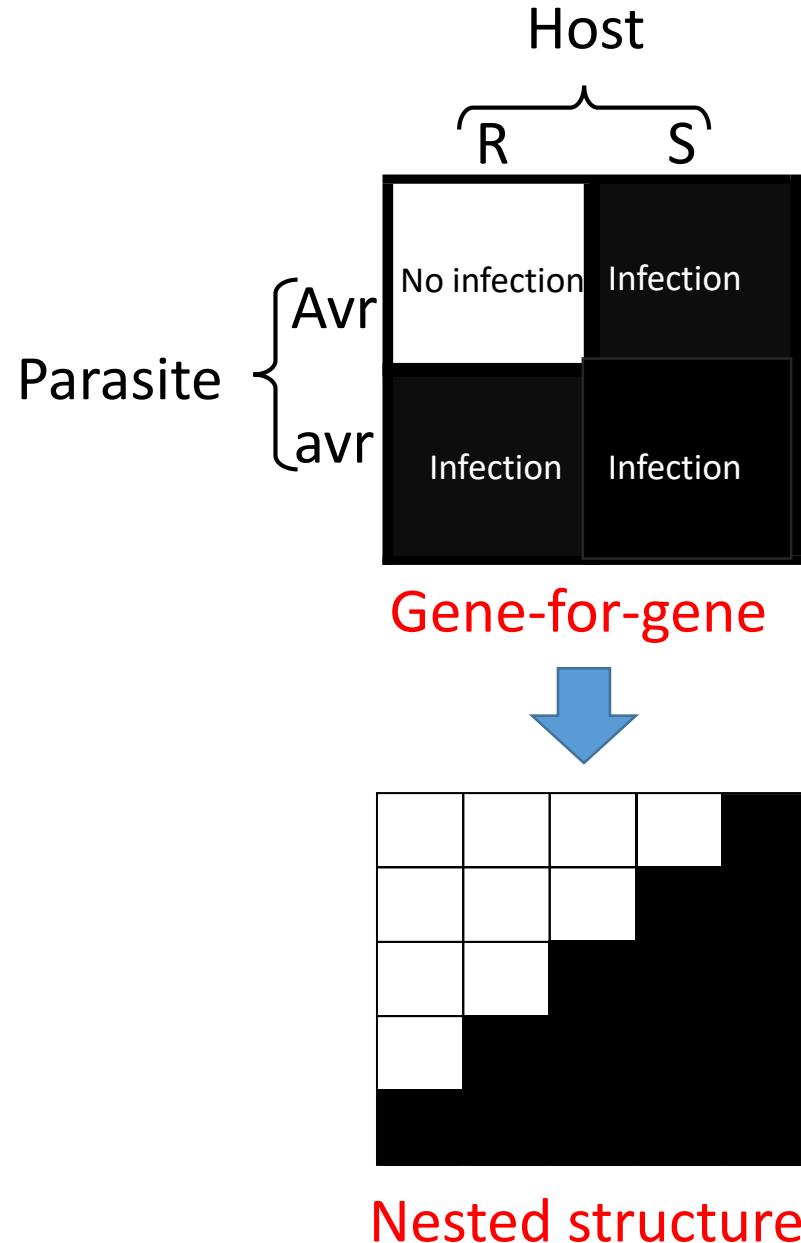
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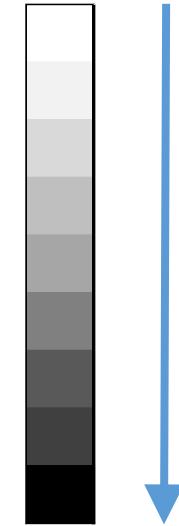
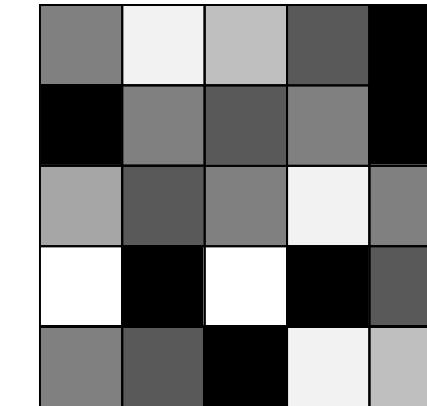
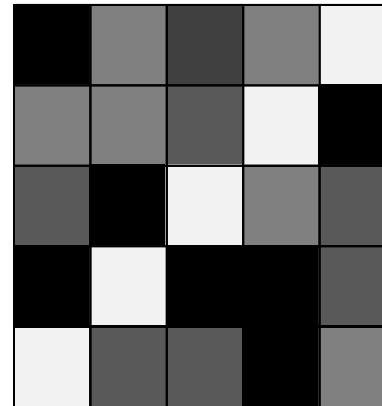
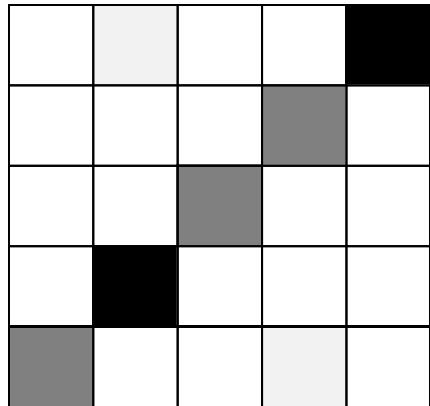
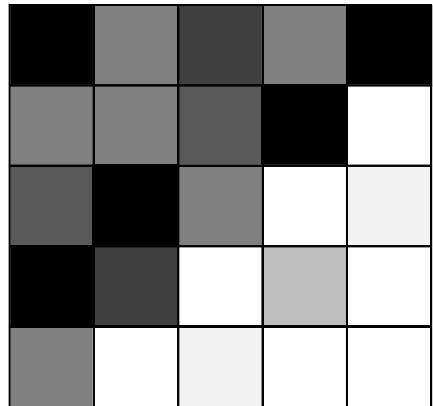
Qualitative plant immunity: two main genetic and evolutionary models



Qualitative plant immunity: two main genetic and evolutionary models

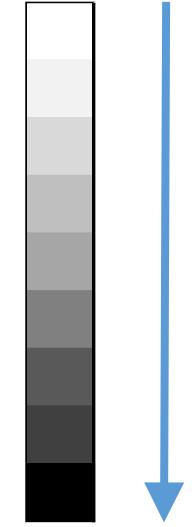
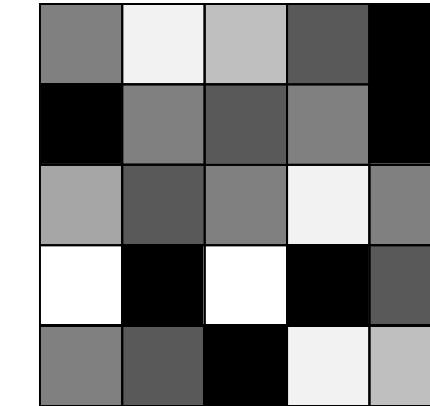
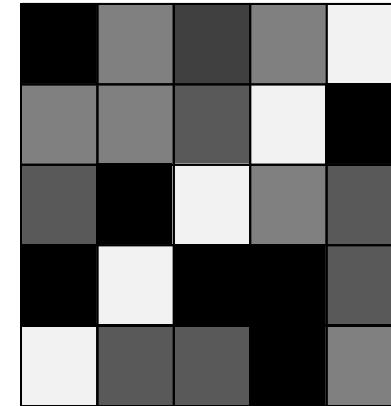
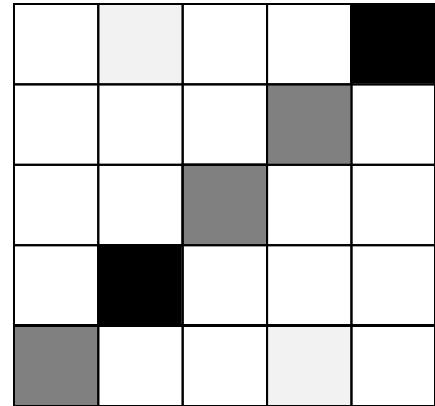
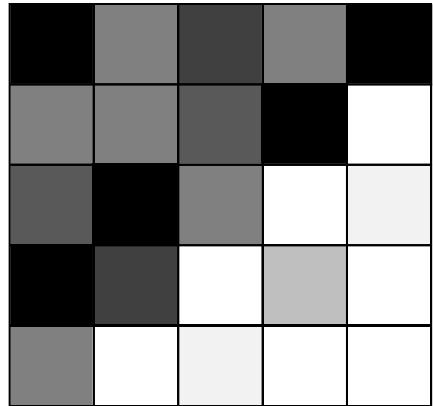


Is there analogous structures (and genetic models) in the case of quantitative infection matrices?



Increasing
infection

**Is there analogous structures (and genetic models)
in the case of quantitative infection matrices?**



**Increasing
infection**

**What can these structures tell us about evolution and genetic bases
of quantitative plant – parasite interactions?**

Analysis of 32 quantitative matrices

Plant	Parasite	
Apricot	<i>Pseudomonas syringae</i>	Bacteria
Barley	<i>Puccinia hordei</i>	
Apple tree	<i>Venturia inaequalis</i>	
Tomato	<i>Botrytis cinerea</i>	
Melon	<i>Podosphaera xanthii</i>	
Wheat	<i>Zymoseptoria tritici</i>	
Pepper	<i>Phytophthora capsici</i>	
Tomato and other <i>Solanum</i> spp.	<i>Phytophthora infestans</i>	
Fabaceae	<i>Aphanomyces euteiches</i>	
Pea	<i>Aphanomyces euteiches</i>	
Grapevine	<i>Plasmopara viticola</i>	
Melon	<i>Aphis gossypii</i>	Insects
Potato	<i>Globodera pallida</i>	
Potato and other <i>Solanum</i> spp.	<i>Globodera pallida</i>	Nematodes
Pepper	<i>Potato virus Y</i>	Viruses

Analysis of 32 quantitative matrices

- Resulting from cross-inoculations under controlled conditions
- Dimensions: at least 6×6
- Without missing data
- Quantitative traits: parasite load, parasite-induced damages, latency period, dissemination capacity

Estimating the degrees of nestedness/modularity and testing their significance

0,27	0,69	0,36	0,47	0,15
0,32	0,45	0,36	1,02	0,04
0,1	3,01	0,89	1,34	0
0	1,26	2,51	0,84	0,09
0,78	1	1,77	0,99	1,03

Infection trait values

Estimating the degrees of nestedness/modularity and testing their significance

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Infection trait values



Transformation into integer values [0-9]

0	2	0	1	0
1	1	0	3	0
0	9	2	4	0
0	4	8	2	0
2	1	5	3	3

Estimating the degrees of nestedness/modularity and testing their significance

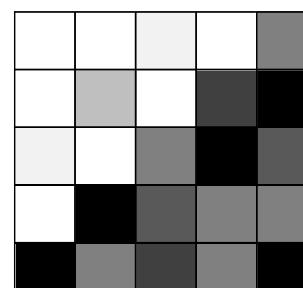
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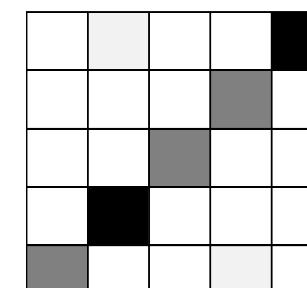
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Transformation into integer values [0-9]

Nestedness/modularity value (X)
of the experimental matrix

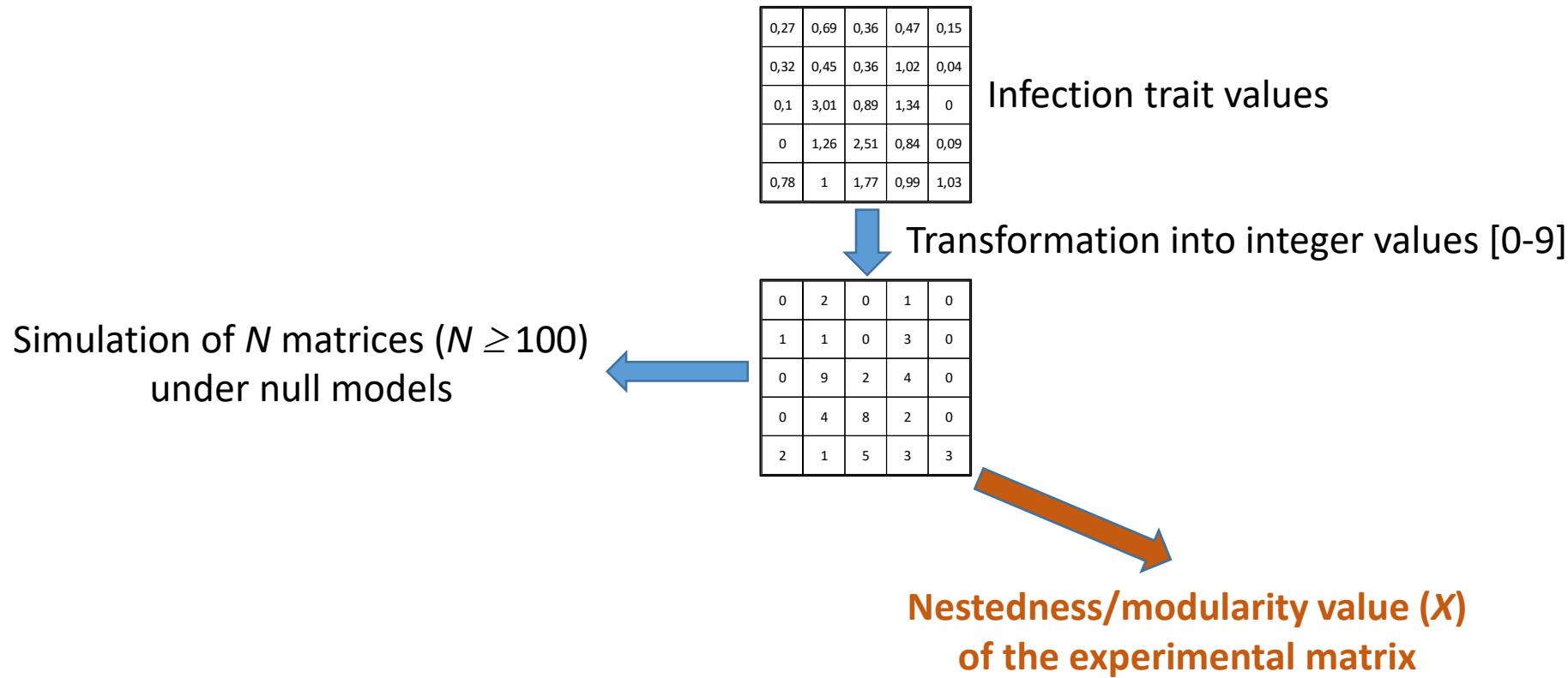


High nestedness

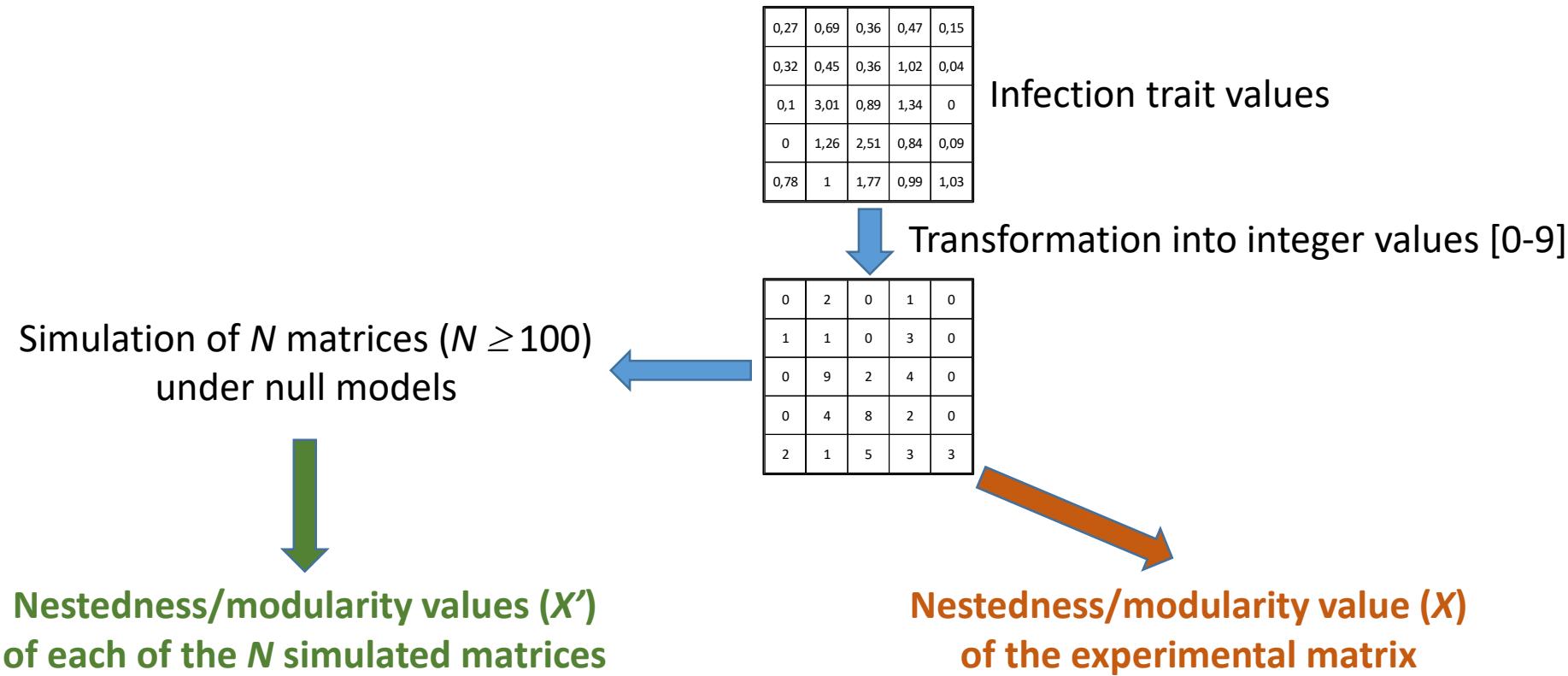


High modularity

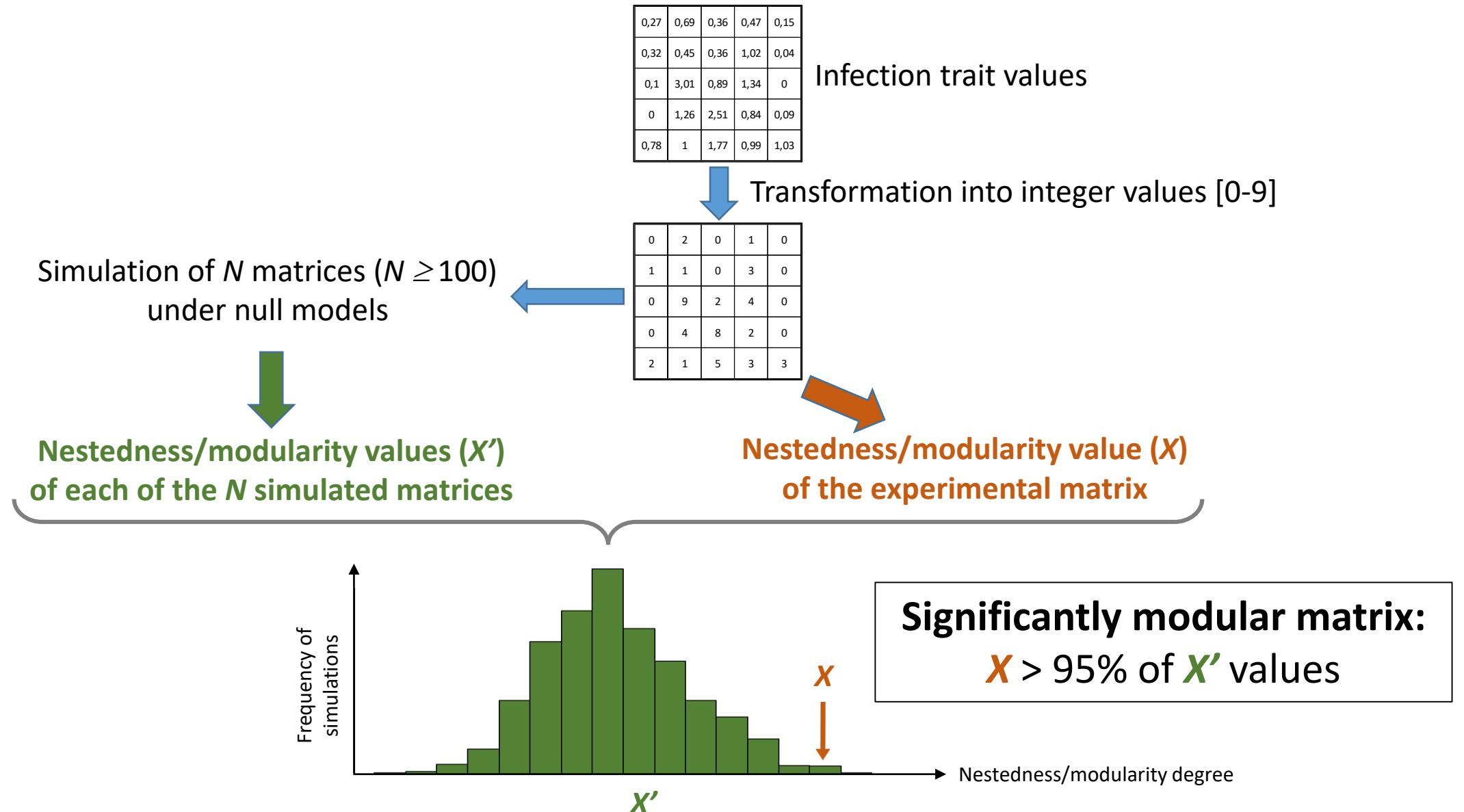
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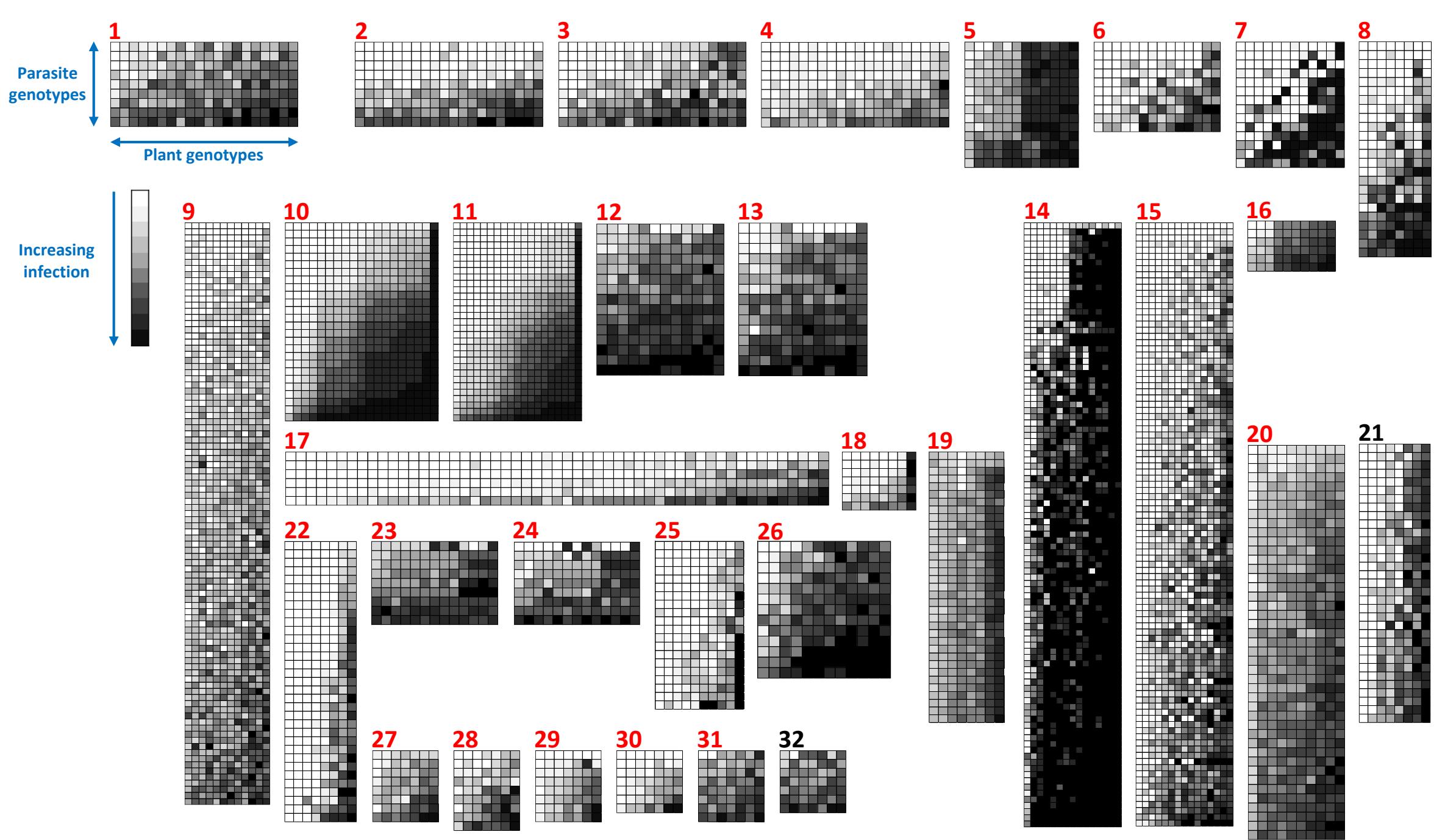
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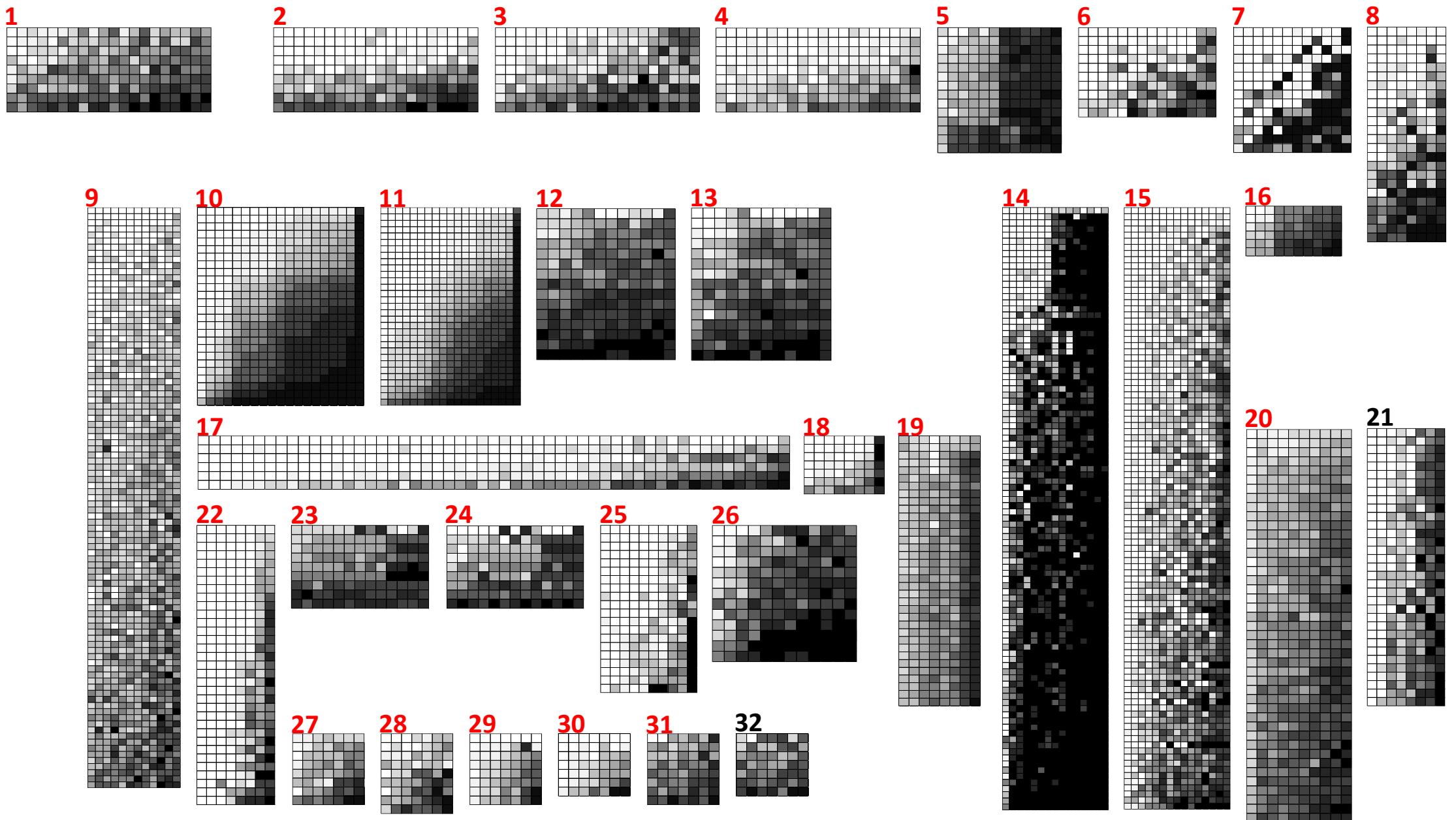


Estimating the degrees of nestedness/modularity and testing their significance



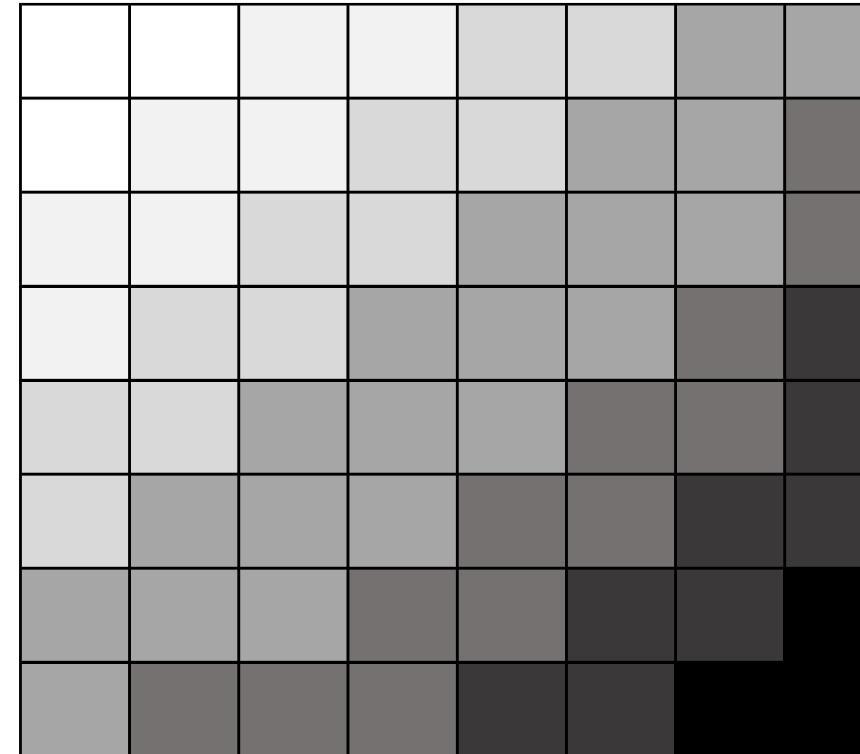
Nestedness results





All matrices but two (n°21 and 32) are strongly and significantly nested

Biological interpretation of nestedness: genetics

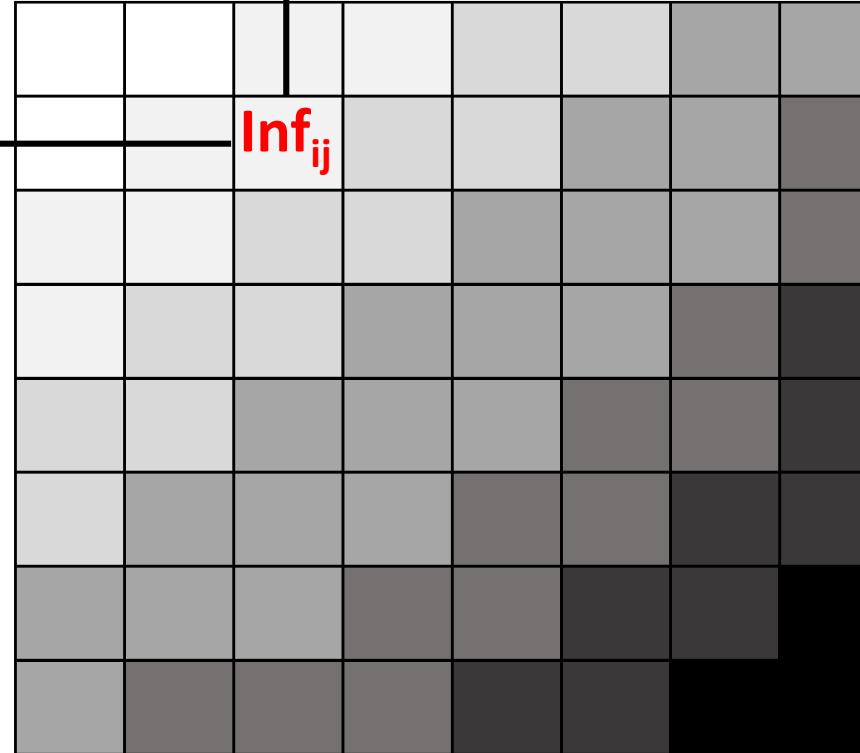


Biological interpretation of nestedness: genetics

Susceptibility of plant genotype j:

Pathogenicity of parasite genotype i: P_i

S_j



→ Suitability of an « additive » model: Inf_{ij} (infection level) = $P_i \times S_j$

Biological interpretation of nestedness: genetics

Suitability of the additive model

- Model 1 (27 matrices) : Infection ~ parasite + plant + parasite * plant
 - Interaction not significant for 5 matrices
 - Part of variance explained by interaction $\omega^2 = 0$ to 0.28 (mean 0.11)
- Model 2 (all 32 matrices) : Infection ~ parasite + plant
 - part of variance explained by model $\omega^2 = 0.40$ to 0.98 (mean 0.69)

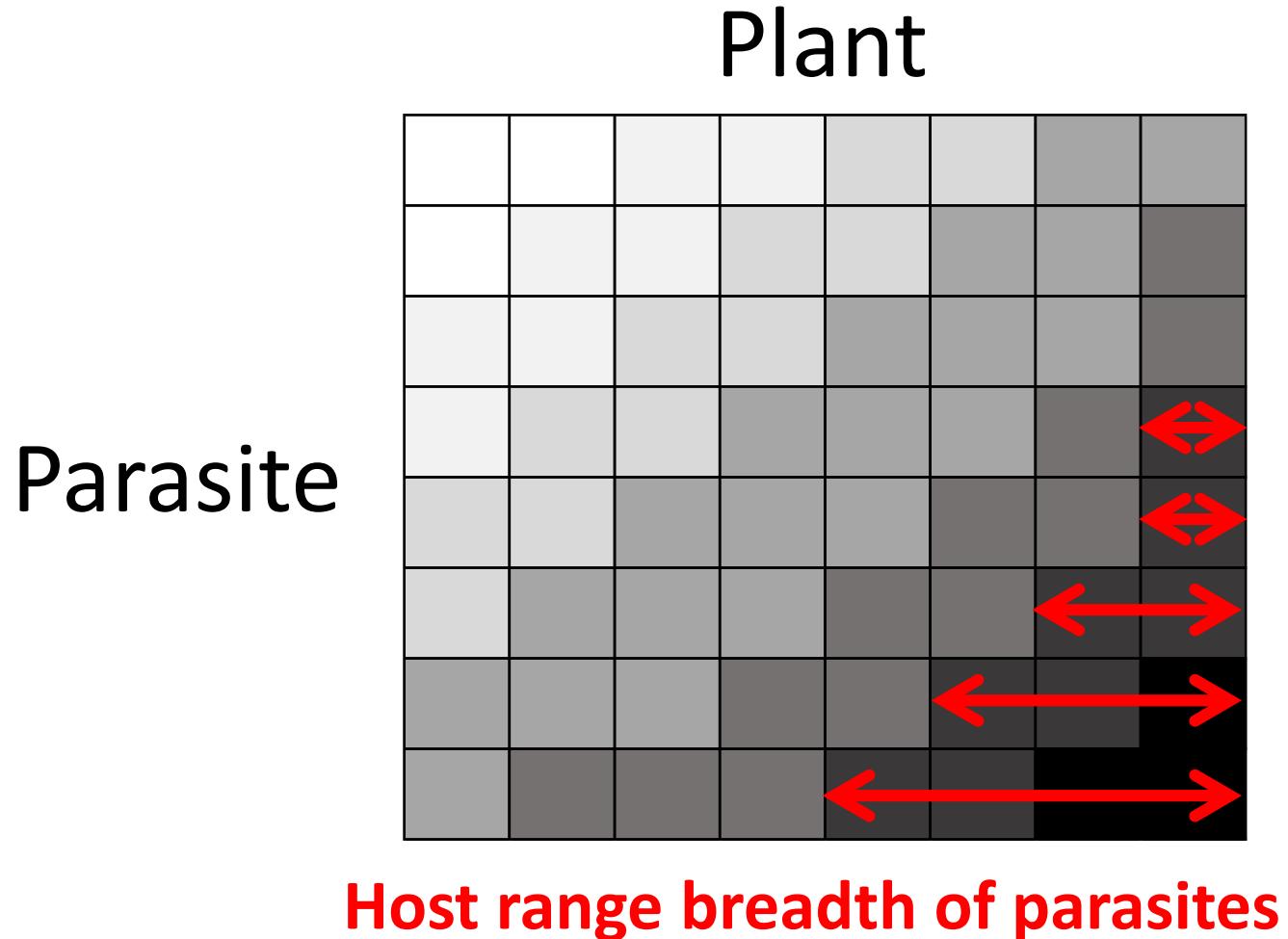
Biological interpretation of nestedness: trade-offs

One could have imagined **trade-offs** between:

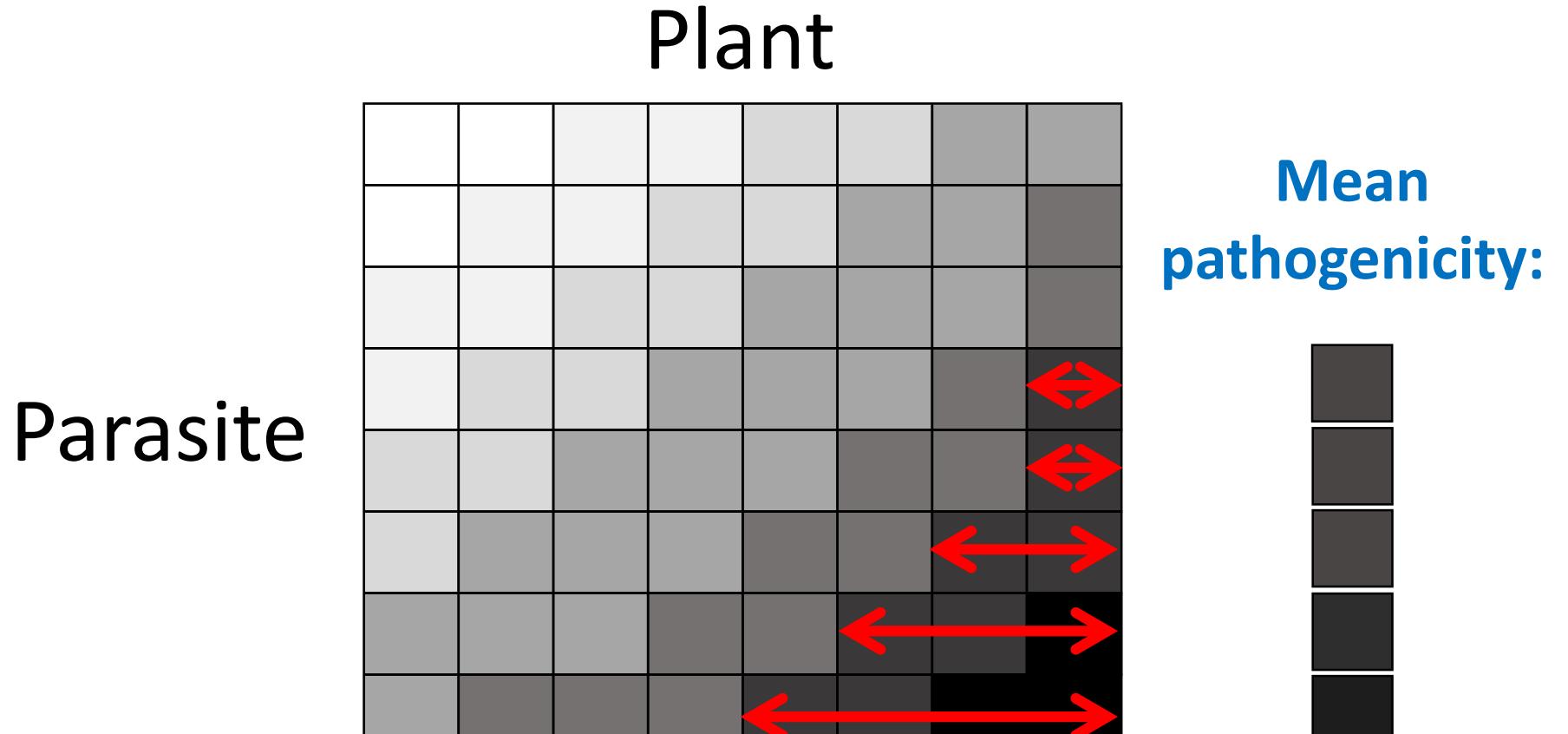
- spectrum and efficiency of resistance among plant genotypes
- host range breadth and pathogenicity among parasite genotypes

... but nestedness suggests the opposite

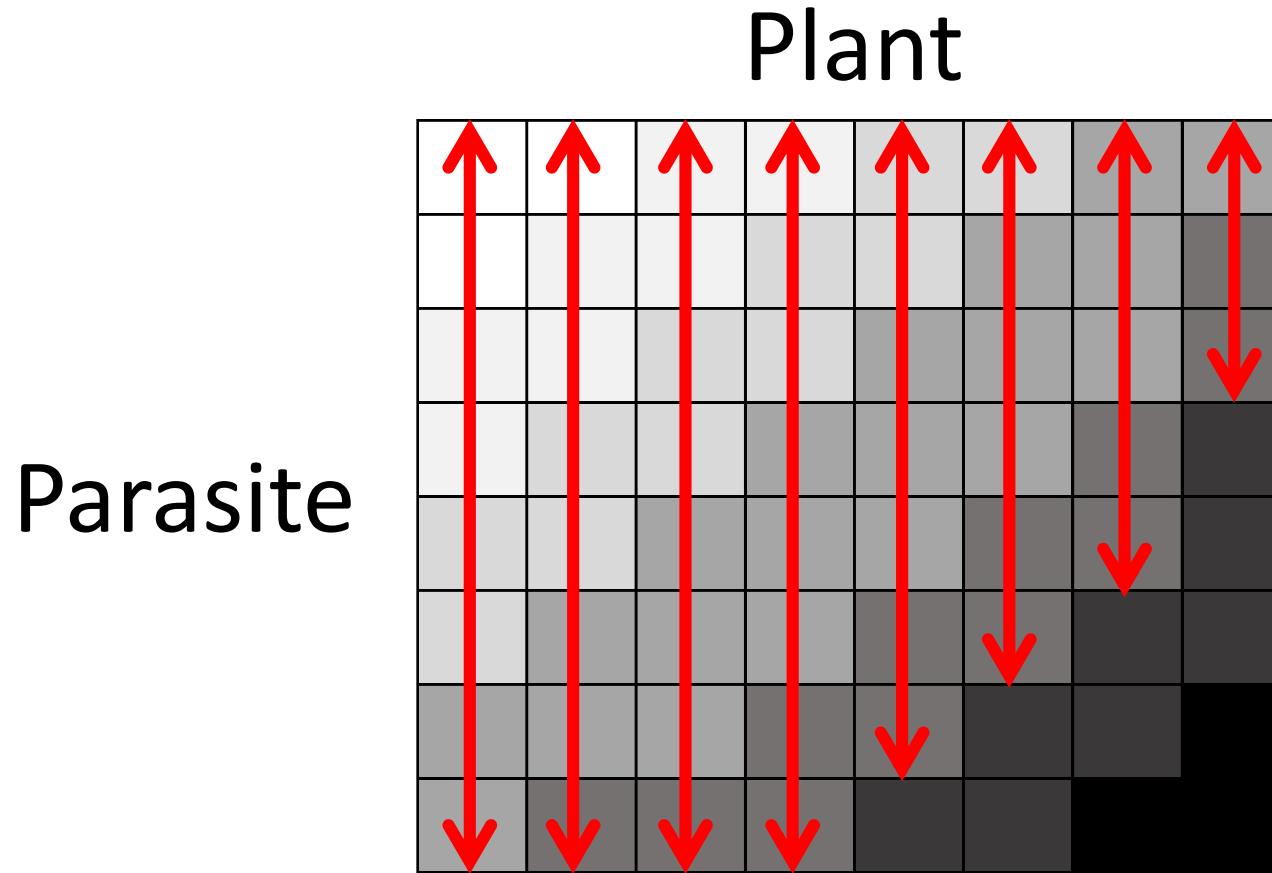
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Biological interpretation of nestedness: trade-offs



Biological interpretation of nestedness: trade-offs

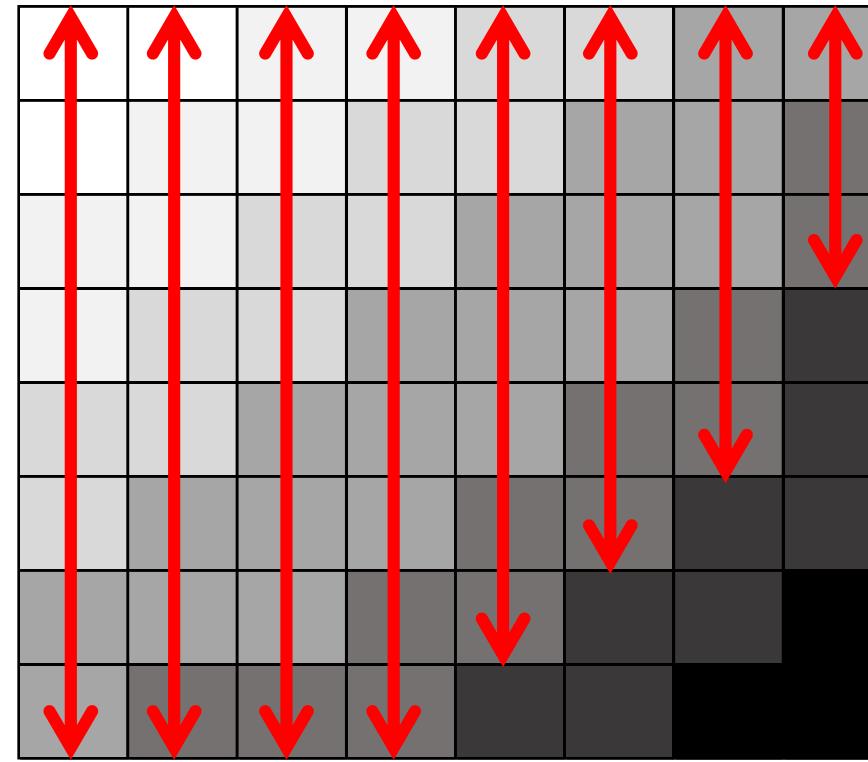


Spectrum of action of plant resistance

Biological interpretation of nestedness: trade-offs

Parasite

Plant



Mean resistance efficiency:



**Spectrum of action of plant resistance
is positively correlated to the mean resistance efficiency**

Occurrence of trade-offs in plant or parasite genotypes?

Threshold = 30% of
maximal infection
value

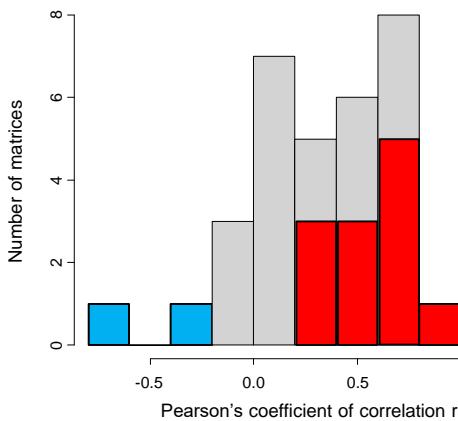


Occurrence of trade-offs in plant or parasite genotypes?

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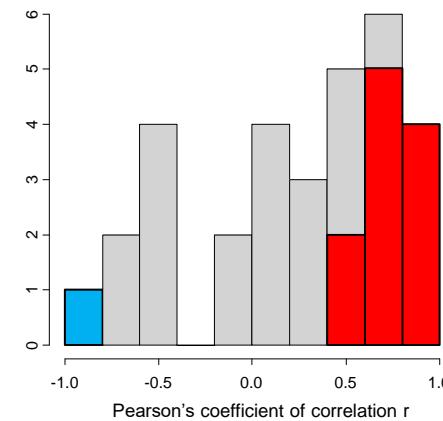
Among parasites

Host range breadth and pathogenicity



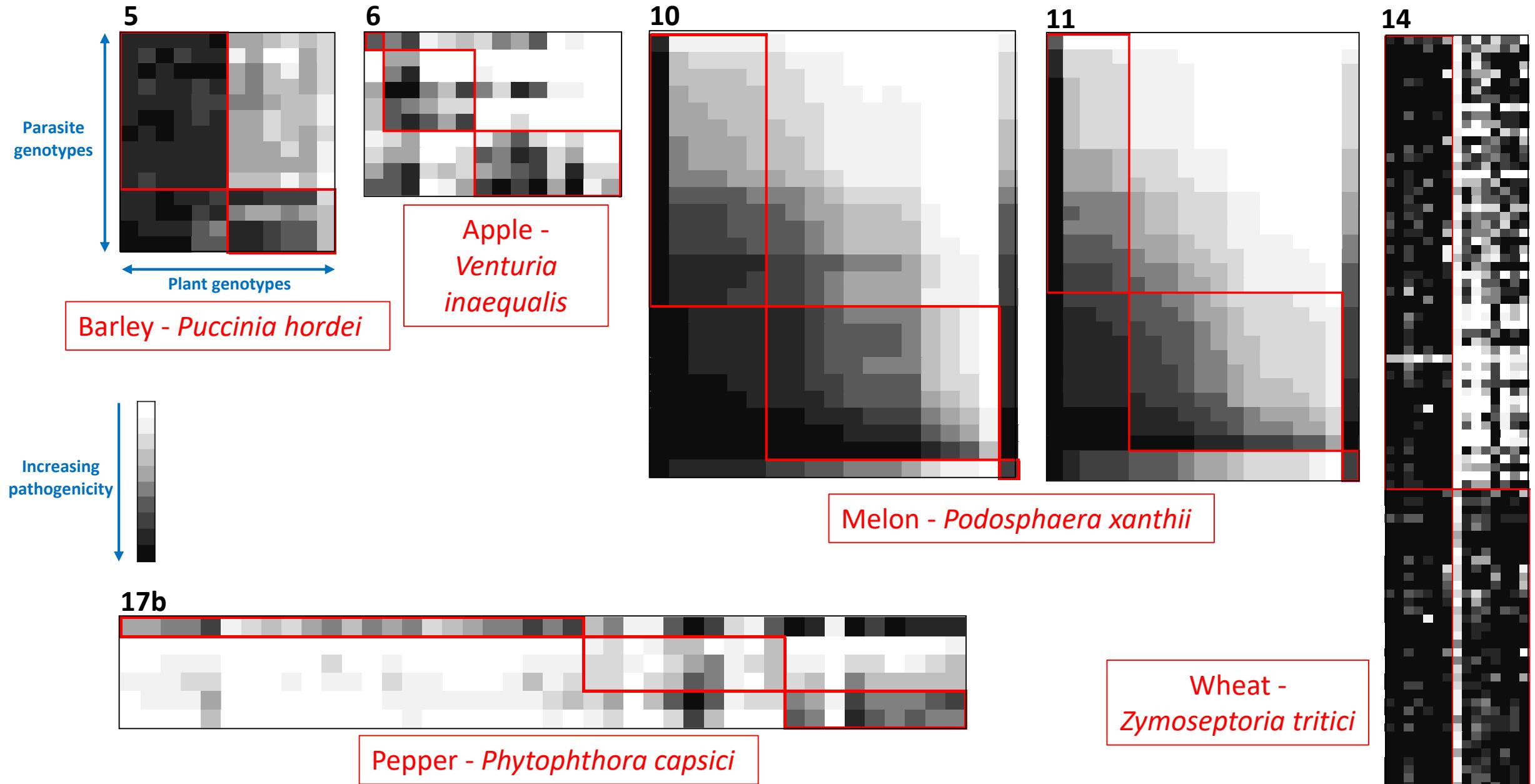
Among plants

Spectrum and efficiency of resistance

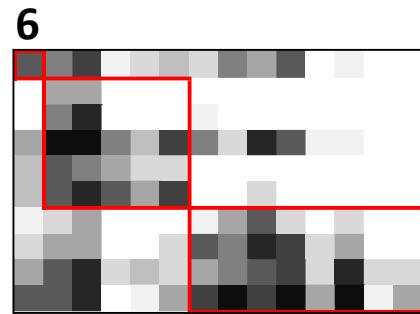
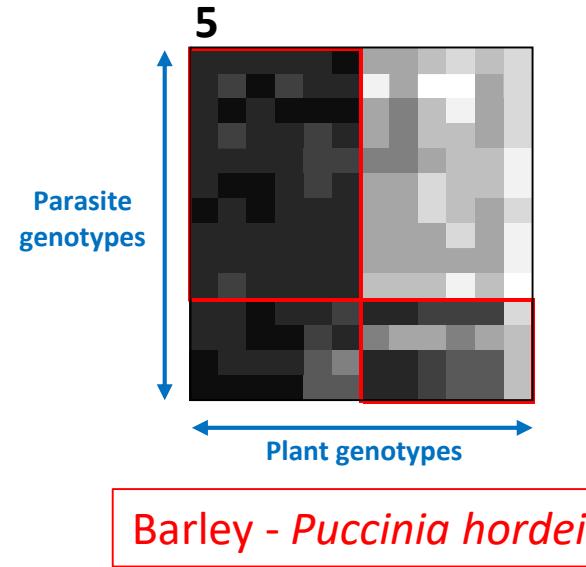


■ Significantly positive r
■ Significantly negative r

Modularity results



- Six matrices show weak but significant modularity (2 or 3 modules)



- Modules linked to the presence of resistance genes / QTLs
(3 of 6 matrices)

Messages à rapporter à la maison

- Strong nestedness for almost all matrices (30/32) / weak and rare modularity (6/32)
 - Reject the "matching allele" model
 - Possible models: additive model; variations on the "gene for gene" model

Messages à rapporter à la maison

- Strong nestedness for almost all matrices (30/32) / weak and rare modularity (6/32)
 - Reject the "matching allele"
 - Possible models: additive model; variations on the "gene for gene" model
- Rare trade-off between the level of resistance and its spectrum of action (plant)
- Rare trade-off between the level of pathogenicity and host range breadth (parasite)
 - Consequences in terms of management of quantitative resistance

Contributors

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