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# Ovine fetal testis stage-specific sensitivity to environmental chemical mixtures

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## Abstract

Exposure of the fetal testis to numerous individual environmental chemicals (ECs) is frequently associated with dysregulated development, leading to impaired adult reproductive competence. However, 'real-life' exposure involves complex mixtures of ECs. Here we test the consequences, for the male fetus, of exposing pregnant ewes to EC mixtures derived from pastures treated with biosolids fertiliser (processed human sewage). Fetal testes from continuously exposed ewes were either unaffected at day 80 or exhibited a reduced area of testis immunostained for CYP17A1 protein at day 140. Fetal testes from day 140 pregnant ewes that were exposed transiently for 80-day periods during early (0–80 days), mid (30–110 days), or late (60–140 days) pregnancy had fewer Sertoli cells and reduced testicular area stained for CYP17A1. Male fetuses from ewes exposed during late pregnancy also exhibited reduced fetal body, adrenal and testis mass, anogenital distance, and lowered testosterone; collectively indicative of an anti-androgenic effect. Exposure limited to early gestation induced more testis transcriptome changes than observed for continuously exposed day 140 fetuses. These data suggest that a short period of EC exposure does not allow sufficient time for the testis to adapt. Consequently, testicular transcriptomic changes induced during the first 80 days of gestation may equate with phenotypic effects observed at day 140. In contrast, relatively fewer changes in the testis transcriptome in fetuses exposed continuously to ECs throughout gestation are associated with less severe consequences. Unless corrected by or during puberty, these differential effects would predictably have adverse outcomes for adult testicular function and fertility.

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## Introduction

Exposure to environmental pollutants during fetal, neonatal, and/or adult life is associated with adverse effects on male reproductive development and function (Sumner *et al.* 2020). Chemical release into the environment has increased markedly since the 1950s (>140,000 chemicals) and this is associated with adverse temporal trends in human male reproductive health (Landrigan 2017). Meta-analyses now confirm that human sperm counts have declined by 50% over 70 years (Carlsen *et al.* 1992, Swan *et al.* 2000, Levine *et al.* 2017). Concomitant with these temporal changes, incidences of testicular cancer in young adults and malformations in male newborns (cryptorchidism and hypospadias) have also increased (Skakkebaek *et al.* 2016, Park *et al.* 2018). In support of an environmental cause, the occupational exposures of mothers to chemical pollutants have been associated with reduced semen quality in their offspring (Istvan *et al.* 2021). Exposure studies in sheep,

dogs, and rodents are similarly indicative of pollutant effects on male reproductive health (Paul *et al.* 2005, Salian *et al.* 2011, Sumner *et al.* 2020). In addition, animal model studies indicate that male reproductive disorders are linked and originate following exposure during fetal life (Skakkebaek *et al.* 2001, Wohlfahrt-Weje *et al.* 2009). The majority of studies have focussed on single chemicals and, in some cases, at doses much higher than environmental exposure levels. Since environmental pollutants constitute a complex mixture, extrapolating from single chemical effects to real-life exposure is problematic.

To investigate real-world exposures, pregnant ewes are grazed on pastures treated with biosolids fertiliser generated from processed human sewage sludge. This is recognised agricultural practice worldwide and the fertiliser contains chemical pollutants which, in combination, represent exposure in humans. A wide range of chemical types have been detected in biosolids and in maternal and fetal livers collected from ewes



that had grazed-treated pastures (Lea *et al.* 2016, Viguie *et al.* 2020). Although directly equating changes in liver chemical load with concentrations in the soil are problematic, exposure has been linked to an increased chemical burden in maternal livers and developmental effects in the mid-gestation male fetal testis. These include reduced numbers of gonocytes, Sertoli and Leydig cells, and a parallel reduction in fetal inhibin A and testosterone (Paul *et al.* 2005). Extending gestational exposure to day 1 neonates and to 7 months in offspring reduced germ cell numbers and induced Sertoli cell-only tubules in cohorts of neonatal and adult animals. In adults, reduced Sertoli cell numbers were also reported (Bellingham *et al.* 2012, Elcombe *et al.* 2021). Biosolids exposure is also associated with adverse effects on the fetal ovary, thyroid, and hypothalamo-pituitary axis as well as in the adult liver (Fowler *et al.* 2008, Bellingham *et al.* 2009, 2010, Hombach-Klonisch *et al.* 2013, Lea *et al.* 2016, Filis *et al.* 2019). Taken together, a conclusion that exposure to chemicals not only impacts the developing fetus but also has longer-term consequences for adult wellbeing is inescapable.

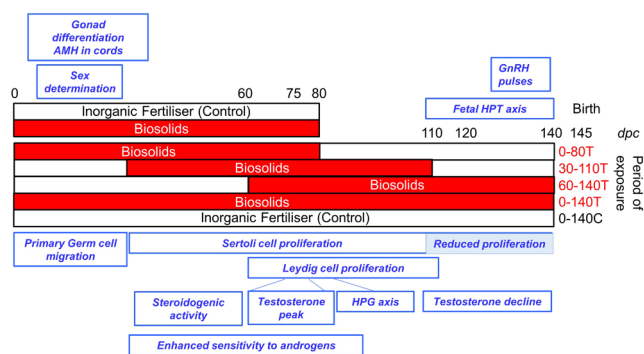
We previously reported that exposing ewes for overlapping 80-day periods that encompass early (0–80 days), mid (30–110 days), and late (60–140 days) gestation induce differential and adverse changes in the fetal ovary (Lea *et al.* 2016). Since these 80-day periods also encompass critical developmental stages in the developing fetal testis (Fig. 1), the current study was designed to determine the effects of transient and consistent exposures on the developing male. Given the continuum of testis development, days 0, 30, and 60 fetal testes will have different cellular compositions largely reflecting periods of Sertoli and Leydig cell proliferation. Since this depends on gestational age, we postulated that the 80-day exposure periods will differentially impact developmental processes including Sertoli and Leydig cell proliferation. Our primary morphological studies therefore focussed on these cell types. Here, we report that exposure during the first third of gestation induced transcriptomic changes measurable in the late gestation fetal testis and exposure in the last third of gestation induced broader phenotypic changes in the male fetus.

## Methods

### Animals, treatment groups, and tissue collection

All animal experimental protocols were approved by the James Hutton Institute's Local Ethical Committee and licensed by the United Kingdom's Animals Scientific Procedures Act 1986 (Project license: 60/3356). All procedures were performed in accordance with relevant guidelines and regulations.

The treatment of pastures with biosolids and grazing of sheep were carried out as previously described (Lea *et al.* 2016). In brief, pastures were fertilised with a single treatment of thermally dried sewage sludge (2.25 metric tons of dry matter/ha; treated: T) or with inorganic fertiliser balanced for



**Figure 1** Experimental design to investigate the effects of biosolids exposure on the fetal testis. Pregnant ewes were sacrificed at day 80 (2 cohorts) or day 140 (5 cohorts) of gestation. In the day 80 group, ewes were exposed to biosolids-treated pastures or pastures fertilised with inorganic fertiliser. In pregnant ewes maintained to day 140, exposure to biosolids-treated pastures occurred from: days 0 to 140 (0-140T, continuous exposure), days 0 to 80 (0-80T, early exposure), days 30 to 110 (30-110T, mid-exposure), and days 60 to 140 (60-140T, late exposure). Control group pregnant ewes (C) were maintained on pastures treated with inorganic fertiliser from days 0 to 140 of gestation (0-140C). For array transcriptomics, 0-80T, 0-140T, and 0-140C groups were used. The main events of differentiation in the fetal testis are described in the blue rectangles. Key references: Sertoli cell proliferation: (Hochereau-de Reviers *et al.* 1995), steroidogenic activity/testosterone: (Pomerantz & Nalbandov 1975, Quirke *et al.* 2001), fetal HPT function: (Mesiano *et al.* 1991, Roselli *et al.* 2016).

nitrogen (225 kg/ha/year; control: C). Experimental animals were maintained on either C or T pastures. Mature Texel ewes were allocated to one of seven treatment groups (Fig. 1) randomly keeping body condition score as consistent as possible. Although initial group size comprised approximately 14 pregnant ewes, the results reported concern only those carrying at least 1 male fetus.

Two groups were either exposed to biosolid treated or control pastures from mating to sacrifice at mid-gestation (day 80). The remaining five groups were all maintained to sacrifice at late gestation (day 140) and comprised one group grazed on C pastures throughout (0-140C), one group exposed to T pastures throughout (0-140T), and three transiently exposed groups where exposure was limited to 80-day periods during early (0-80 days), mid (30-110 days), or late (60-140 days) gestation (Fig. 1). Pregnancies were established by mating ewes with Texel rams at the second synchronized estrus following the withdrawal of progestagen sponges (chronolone, 30 mg; Intervet, Cambridge, UK). Pregnant ewes were either maintained on T pastures or transferred between C and T pastures (and back again), as appropriate. All ewes were habituated to being moved between pastures, thus minimising stress. When pregnant ewes were moved, they were maintained on a separate pasture for a few days to minimise C pasture contamination by faeces and urine.

All pregnant ewes were euthanised according to Schedule 1 protocols (UK Animals Scientific Procedures Act, 1986). Samples and data (organ weights, etc.) were collected and processed according to our standard protocols (Bellingham *et al.* 2013,

Lea et al. 2016). In brief, fetal anogenital distance (AGD) and selected organ weights were measured and recorded. One testis per fetus was analysed for histology and the second for transcriptomic studies. Testes were Bouin's fixed for histological analysis or snap-frozen in liquid nitrogen and stored at  $-80^{\circ}\text{C}$  for RNA extraction. Maternal and fetal liver sample processing and chemical measurements were reported by (Lea et al. 2016).

### Testis immunohistochemistry and histomorphology

Bouin's-fixed testes were processed for immunohistochemistry: Sertoli cells (anti-Mullerian hormone; AMH), Leydig cells (steroidogenic enzymes: CYP11A1, CYP17A1), and proliferation (Ki67). Established immunohistochemistry protocols were applied (Paul et al. 2005, Andrade et al. 2013) including epitope retrieval (microwaving:  $3 \times 5$  min in 0.01 M citrate buffer pH 6.0) and application of the Vectastain ABC universal Elite kit (2B Scientific Ltd, Stonesfield, UK) protocol. Tissue sections were incubated with primary antibodies for 1 h at room temperature: (a) polyclonal goat AMH (Santa Cruz:  $0.125 \mu\text{g}/\text{mL}$ ); (b) monoclonal mouse anti-human Ki67 (Clone MIB-1: Dakocytomation:  $0.8 \mu\text{g}/\text{mL}$ ); (c) polyclonal rabbit anti-CYP11A1 (Merck Millipore:  $4 \mu\text{g}/\text{mL}$ ); (d) polyclonal rabbit anti-CYP17A1 (gift: Professor Ian Mason, Edinburgh:  $10 \mu\text{g}/\text{mL}$ ). Negative controls comprised incubation with non-specific mouse, rabbit, or goat IgG. Sections were visualised by incubation with DAB chromagen according to kit instructions (2B Scientific Ltd). The area of cells positively stained for CYP17A1 or CYP11A1 was measured as a percentage of total interstitial area (Image-Pro Plus: Media Cybernetics, Maryland, USA). Twenty randomly selected digital images ( $\times 400$ ) were analysed by a single observer (BE) while blinded to the group being examined.

Sertoli cell numbers were determined stereologically (Paul et al. 2005, Andrade et al. 2013). In brief,  $5\text{-}\mu\text{m}$  sections were subjected to immunohistochemistry for AMH. Forty images at  $\times 630$  magnification were captured from one testis cross-section using Leica software, ten per pole. Images were overlaid with a 432 point grid (Image-Pro Plus: Media Cybernetics). Sertoli cells across an intersection were counted and this total point value was expressed as a percentage of the maximum count across all 40 images ( $40 \times 432 = 17,280$ ). Testis weight occupied by Sertoli cells was calculated (e.g. 1 g testis with an average point count of  $10\% = 0.1$  g of Sertoli cells). A weight-to-volume conversion was applied ( $1 \text{ g} = 1 \text{ cm}^3$ ) to generate the absolute volume (AV) of the testis occupied by Sertoli cells. The average Sertoli cell diameter was measured (Image-pro plus) and the mean nuclear volume (MNV) of each Sertoli cell was determined from the formula  $(4/3\pi)r^3$ . The total number of Sertoli cells per testis was calculated  $(AV (\mu\text{m}^3)/\text{MNV}(\mu\text{m}^3))$  and adjusted to account for testis weight.

### Measurement of testosterone

Fetal serum concentrations of total testosterone (bound and unbound) were measured using the automated ADVIA Centaur XP competitive immunoassay system (Siemens Healthcare Diagnostics), as previously reported (Lea et al. 2016). Total serum testosterone assay sensitivity was

$0.35 \text{ nmol}/\text{L}$  and the mean intra- and inter-assay CV values were 4.4 and 6.2%, respectively.

### Fetal testicular RNA extraction

RNA, DNA, and protein were extracted from fetal testis using an AllPrep DNA/RNA/Protein mini kit (Qiagen Ltd.). Samples were homogenised and processed as previously described (Lea et al. 2016). The manufacturer's instructions were followed with the optional on-column DNase digestion included. Samples were homogenised in  $600 \mu\text{L}$  RNeasy lysis buffer for tissues (RLT) buffer for 2 min at 30 Hz, centrifuged after which the lysate was added to an ALLprep DNA column. After centrifugation, the column flow-through was processed by passing through an RNeasy spin column according to kit instructions. In brief, RNA wash buffer (RW1) buffer was centrifuged through the column after which a DNase I incubation mix was added followed by a 15-min incubation at room temperature. The column was then washed/centrifuged with RW1 buffer ( $350 \mu\text{L}$ ) and  $2 \times$  RNA purification and elution buffer (RPE) buffer ( $500 \mu\text{L}$ ) after which the addition of  $30 \mu\text{L}$  of RNase-free water followed by centrifugation yielded RNA containing flow through for cDNA synthesis.

### Customised ovine microarray and pathway analyses

Transcriptome analysis was conducted using a custom 15K Agilent oligo sheep microarray generated as previously described (Lea et al. 2016). In brief, a catalogue Sheep Gene Expression Microarray  $8 \times 15\text{K}$  (G4813A-019921) was modified by adding new target sequences and removing redundancies. The latter was identified from sheep ESTs assembly and oligo annotations performed by Sigenae (<http://www.sigenae.org/>). Several different oligonucleotides were used to target sheep contigs, and the best annotated of these at the 3' end were conserved (2 per contig max). Other oligos specific for the same transcript were removed from the array. Agilent 'GE Probe Design' eArray workflow and tools (<https://earray.chem.agilent.com/earray/>) were used for oligo design. All 1500 new oligos, and those remaining on the original Agilent array, were annotated using Sigenae SigReannot tool49. The array was enriched with genes initially identified in the fetal sheep gonad and completed with control genes known to be expressed in developing fetal gonads from both sexes. The final version of the array comprised 7500 different genes and labelling and hybridisation were performed at the 'Plate-forme Biopuces et Sequencage' (<http://www-microarrays.u-strasbg.fr/>). Following one-colour labelling and hybridisation using the Quick Amp Labelling kit (Agilent, 5190-0442) and One-colour RNA Spike-in Kit (Agilent, 5188-5282), arrays were scanned with the Agilent DNA Microarray Scanner Model G2565B. Image analysis was performed with Agilent Feature Extraction software v9.5.3.1.

### Gene ontology enrichment

Identified sheep differentially expressed probes were analysed with Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway membership with Database was performed using the DAVID Bioinformatic Database 6.8 (<https://david.ncifcrf.gov/>). These analyses and pathways were

considered significant for a Benjamini corrected enrichment score of less than 0.05. Further analysis of gene pathways utilised the Ingenuity Pathway Analysis software (<http://www.ingenuity.com/>). This pathway analysis system uses the same hypergeometric test as DAVID, and topology-based output was used to assess the biological processes impacted by biosolids exposure (Nguyen *et al.* 2019). We used the eXploring Genomic Relations web tool (<http://galahad.well.ox.ac.uk:3040/>) under default parameters and with lists of differentially expressed genes tested for enrichment of annotations (Fang *et al.* 2016).

### Pathway and functional analysis of differentially expressed genes (DEGs)

To further understand biological functions and pathways, sheep differentially expressed transcripts (false discovery rate (FDR)  $\leq$  5%; LogFC threshold  $\pm$  0.2) were functionally annotated based on GO terms and KEGG pathway through the DAVID ontology database. Uncharacterized, putative genes and redundant probes were removed so that 1596 (0–80T) and 1567 (0–140T) official gene name symbols were subjected to DAVID analyses. To increase the depth of genes with GO annotations, *Homo Sapiens* genome annotation was used as background and statistically enriched biological processes and molecular functions were obtained in which the proteins are involved (Ha *et al.* 2015). The transcripts were classified as: biological process, cellular component, and molecular function. An individual transcript may be represented in several categories.

### Validation of transcriptomic data by qPCR

Using the same samples used for microarray, transcript expression was confirmed by real-time PCR (Lea *et al.* 2016). DNase-treated RNA was used to synthesise cDNA (Transcriptor First Strand cDNA Synthesis Kit: Roche). Standard curves were generated from pooled cDNA (1:5 serial dilutions), and all cDNA samples were diluted at 1:50 in dH<sub>2</sub>O. Primers and probes were designed using Primer3 software, and DNA probes were synthesised with a 5'-FAM fluorophore and a 3'-TAMRA quencher (Eurofins, Ebersberg, Germany). Genbank accession numbers and probe and primer sequences are listed in Supplementary Table 1 (see section on [supplementary materials](#) given at the end of this article). Samples were run in triplicate using the LightCycler® 480 Probes Master (Roche), and 'no template controls' were included as standard. Reaction mixtures and cycling conditions were as previously reported (Lea *et al.* 2016). All qPCR data were analysed using Roche LightCycler480 software and normalised using the geNorm method. Three housekeeping genes (*GAPDH*, *HPRT*, *YWHAZ*) were tested for stability using geNorm, Normfinder, and ANOVA analysis.

### Statistical analysis

Morphometric data comprising fetal mass, organ weights (testis, thyroid, adrenal, liver), and AGD were analysed using Generalized Linear Regression (GLR: Genstat statistical package version 20; <https://www.vsnl.co.uk/>). Since males were from singleton and twin pregnancies, data were adjusted

for litter size (fixed effects: number of fetuses and treatment, random effect: ewe). The Bonferroni multiple comparisons adjustment was applied, and between-group differences were identified. Sertoli cell numbers and Leydig cell staining were also analysed by GLR. Sertoli and Leydig cell data are presented as scatter plots with means (GraphPad Prism Prism, version 8). Fold changes were presented as positive or negative values relative to controls.

For gene array, data processing and analysis were conducted using Bioconductor packages suite (<http://www.bioconductor.org/index.html>) and LIMMA package50 with the R statistical programme. Raw median signal from Feature Extraction array files was used as non-processed signal and log<sub>2</sub>-transformed. Background was then subtracted locally, and intra-array normalisation was performed by subtracting the array median signal from each spot signal on the same array. Multiple testing corrections were applied, and differentially expressed transcripts were considered under a FDR of 5% (Benjamini & Hochberg 1995).

## Results

### Effects of biosolids exposure on the fetal urogenital tract and testosterone

At day 80, male fetal mass was not altered by continuous exposure to biosolids and there were no treatment effects on AGD or organ weights (i.e. testis, thyroid, adrenal, and liver: [Table 1](#)). In contrast, day 140 male fetuses from ewes exposed from 60 to 140 days of gestation (60–140T group) had a lower fetal mass than non-exposed controls (0–140C), continuously exposed fetuses (0–140T) and fetuses exposed mid-gestation (30–110T) ( $P < 0.001$ ). AGD was shorter in the 60–140T group than controls ( $P < 0.05$ ) ([Table 1](#)). The late exposure group (60–140T) also had smaller adrenals ( $P < 0.05$ ), smaller testes ( $P < 0.001$ ), and lower levels of testosterone ( $P < 0.01$ ) compared to control non-exposed fetuses ([Table 1](#)).

### Effects of biosolids exposure on testis development

*Day 80:* continuous maternal exposure to biosolids had no effect on AMH-positive Sertoli cell numbers per gram of testis ([Fig. 2A](#) and Supplementary Fig. 1A). There was no treatment effect on per cent nucleated area stained for the steroidogenic enzyme CYP11A1 ([Fig. 2C](#) and Supplementary Fig. 1C) or CYP17A1 ([Fig. 2E](#) and Supplementary Fig. 1D).

*Day 140:* Sertoli cell numbers per gram of testis were reduced by transient exposure of the mother to biosolids, regardless of when exposure occurred. In contrast, continuous exposure from 0 to 140 days had no effect on Sertoli cell numbers ([Fig. 2B](#)) ( $P < 0.001$ ). Positive staining for Ki67 was indicative of Sertoli cell proliferation as expected at this developmental stage (Supplementary Fig. 1B). The per cent testicular area stained for CYP11A1 was reduced in the 0–140T ( $P < 0.001$ ) and 0–80T ( $P < 0.01$ ) exposure groups

**Table 1** Morphology (days 80 and 140) and endocrinology (day 140) of male fetuses following exposure to biosolids-treated pastures. Values represent predicted means  $\pm$  S.E.M. Experimental groups: C=control, T=exposed. Paired organs were combined into a single weight. Data were fitted to a general linear model, adjusting for litter size, and analysed by general linear regression (Genstat). Between-group differences were analysed by Bonferroni (day 140).

	Exposure groups								
	Day 80 fetuses			Day 140 fetuses					
	0–80C	0–80T	<i>P</i>	0–140C	0–140T	0–80T	30–110T	60–140T	<i>P</i>
Morphology									
No. ewes > 1 male fetus (D80 <sup>A</sup> , D140 <sup>B</sup> )	8	9		11	9	7	8	11	
No. fetuses <sup>C</sup>	13	11		15	12	9	12	15	
Fetal mass (g, D80; kg D140)	346.8 $\pm$ 11.66	318.2 $\pm$ 12.69	NS	5.39 $\pm$ 0.22 <sup>a</sup>	5.24 $\pm$ 0.26 <sup>a</sup>	5.01 $\pm$ 0.31 <sup>ab</sup>	5.32 $\pm$ 0.24 <sup>a</sup>	<b>4.05 <math>\pm</math> 0.24<sup>b</sup></b>	<0.001
AGD (mm)	47.07 $\pm$ 0.66	47.77 $\pm$ 0.71	NS	151.6 $\pm$ 3.38 <sup>a</sup>	144.6 $\pm$ 4.06 <sup>ab</sup>	150.7 $\pm$ 4.67 <sup>ab</sup>	142.4 $\pm$ 3.61 <sup>ab</sup>	<b>134.9 <math>\pm</math> 3.56<sup>b</sup></b>	<0.05
Testis (mg)	155.7 $\pm$ 9.4	141.4 $\pm$ 10.2	NS	2590 $\pm$ 112.3 <sup>a</sup>	2534 $\pm$ 128.3 <sup>a</sup>	2521 $\pm$ 155.5 <sup>a</sup>	2192 $\pm$ 120.3 <sup>a</sup>	<b>1677 <math>\pm</math> 118.3<sup>b</sup></b>	<0.001
Thyroid (mg)	141.5 $\pm$ 12.84	128.7 $\pm$ 14.0	NS	1370 $\pm$ 107.2	1354 $\pm$ 128.8	1117 $\pm$ 148.0	1249 $\pm$ 119.8	1328 $\pm$ 113.0	NS
Adrenals (mg)	126.6 $\pm$ 8.5	132.4 $\pm$ 9.2	NS	647.8 $\pm$ 28.47 <sup>a</sup>	572.9 $\pm$ 32.53 <sup>ab</sup>	549.0 $\pm$ 39.44 <sup>ab</sup>	591.5 $\pm$ 30.50 <sup>ab</sup>	<b>505.1 <math>\pm</math> 29.99<sup>b</sup></b>	<0.05
Liver (g)	21.21 $\pm$ 0.6	19.6 $\pm$ 0.7	NS	132.0 $\pm$ 7.97	130.9 $\pm$ 9.11	134.7 $\pm$ 11.04	149.8 $\pm$ 8.54	117.3 $\pm$ 8.40	NS
Fetal endocrinology									
Fetal testosterone				3.60 $\pm$ 0.21 <sup>b</sup>	3.75 $\pm$ 0.28 <sup>b</sup>	3.45 $\pm$ 0.30 <sup>ab</sup>	3.52 $\pm$ 0.23 <sup>b</sup>	<b>2.54 <math>\pm</math> 0.23<sup>a</sup></b>	<0.01

<sup>A</sup>Ewes at day 80: 82% (14/17) carried twin pregnancies and the remaining pregnancies were triplets ( $n = 3$ ); <sup>B</sup>Ewes at day 140: 69% (32/46) carried twin pregnancies and remaining pregnancies were singles ( $n = 7$ ) and triplets ( $n = 7$ ); <sup>C</sup>Fetuses were used from all pregnant ewes. Differing superscripts indicate differences between groups and values in bold indicate differences relative to 0–80C or 0–140C. AGD, anogenital distance.

(Fig. 2D), whereas staining for CYP17A1 was reduced in continuous and all transient exposure groups (Fig. 2F).

### Effects of biosolids exposure on the testis transcriptome

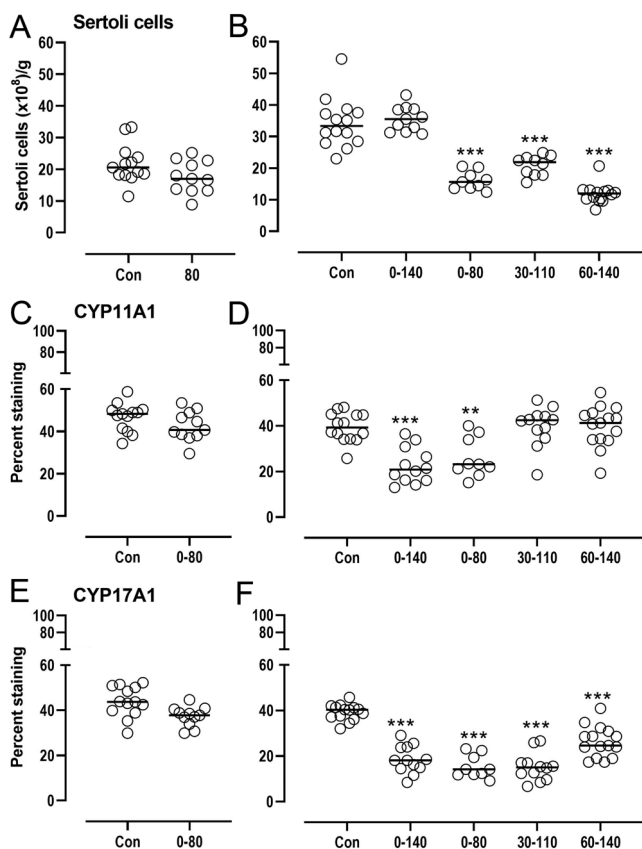
Gene array analysis was carried out on three groups of fetuses at 140 days: 0–140C, 0–140T, and 0–80T. Analyses revealed that 249 transcripts (3.3% of 7500) were differentially expressed ( $P < 0.05$  after Benjamini–Hochberg multiple testing correction (FDR at 5%) and absolute fold change > 1.5) between controls and exposed fetal testes (Fig. 3A). Of these transcripts, 47 were differentially expressed in testes from continuously exposed mothers (0–140T). In contrast, 202 transcripts were differentially expressed in the 0–80T group (Fig. 3A). A total of 22 differentially expressed genes were common to both the 0–80 and 0–140 exposure groups (Fig. 3B). In the exposure groups, the majority of genes were downregulated, particularly in the 0–80T group (0–80T, 174/202; 86% vs 0–140T, 27/47, 57%) (Fig. 3A and Supplementary Table 2). The complete raw data transformed by the platform (processed signal) are provided as Supplementary Table 3.

The functional analysis of differentially expressed transcripts common to both exposure groups highlighted differentially expressed genes associated with VEGF and signalling pathways: SMAD2/3, IGF1, ERK1/ERK2 MAPK, ErbB1,2,3 (Fig. 4). Differentially expressed transcripts specific to the 0–80T exposure group were associated with a range of cell specific functions: androgen signalling, angiogenesis, and cell signalling (Fig. 5A). In the lesser perturbed group (0–140T), differentially expressed transcripts were associated

with cell signalling, metabolism (e.g. insulin receptor pathway), and angiogenesis (Fig. 5B).

Topological analyses by ingenuity pathway analysis revealed differentially expressed gene networks associated with drug metabolism, disorders of haematological neurological, cardiovascular systems, and metabolic disease (Table 2). High scoring gene networks in the 0–140T group were primarily linked to cell function and these networks were also identified in the 0–80T group (Table 2). Biological functions primarily highlighted in the 0–80T group were linked with cancer and reproductive system disease, dermatological disease, and genetic disorders (Table 3). The latter two functions were also highlighted in the 0–140T group along with inflammatory disease and skeletal/muscular disorders. Molecular, cellular, and physiological functions in the 0–140T group were low scoring. However, in the 0–80T group, organismal and tissue development along with cell morphology, assembly, and organisation were high-scoring networks (Table 3).

Of the 22 differentially expressed transcripts common to both exposure groups (Fig. 3B), the four most upregulated genes were genes encoding the human leukocyte antigens A and B (Supplementary Table 2: day 80 and day 140 testes, respectively: *HLA-A*: +1.75- and +1.67-fold change, *HLA-B*: +1.3- and +1.6-fold change). The two most downregulated genes in both groups (0–80T, 0–140T) were *POSTN* (periostin osteoblast specific factor) and ovin *ZFNLOC101104520* (zinc finger protein) (Supplementary Table 2: day 80 and day 140 testes, respectively: *POSTN*: –1.4- and –2.0-fold change, associated with tumour progression and *LOC101104520*: –1.3- and –1.2-fold change, respectively).



**Figure 2** Disturbance in Sertoli cells numbers and area positive for steroidogenic cells in fetal testes following continuous or transient exposure of pregnant ewes to biosolids. Fetal testes were collected at days 80 or day 140 of gestation. Day 80 testes were derived from control (con) non-exposed and continuously exposed mothers. Day 140 testes were derived from control (con) non-exposed mothers, from mothers exposed continuously (0–140) and from mothers exposed for a period of 80 days limited to early (0–80 days), mid (30–110 days), and late gestation (60–140 days). Sertoli cells (A: day 80, B: day 140) were identified by AMH staining, quantified stereologically, and expressed as numbers per gram of testis. Steroidogenic interstitial cells were identified by immunohistochemistry quantified by image analysis as per cent nucleated interstitial area stained for CYP11A1 (C: day 80, D: day 140) and CYP17A1 (E: day 80, F: day 140). Each symbol corresponds to an individual control or exposed animal. The horizontal line indicates the mean for each group. Day 140 data were fitted to a generalized linear regression model, adjusting for litter size (fixed effects: number of fetuses and treatment, random effect: ewe) (Genstat). Treatment effect:  $P < 0.001$ . Between-group differences were analysed by Bonferroni; asterisks indicate significant differences with control non-exposed group.

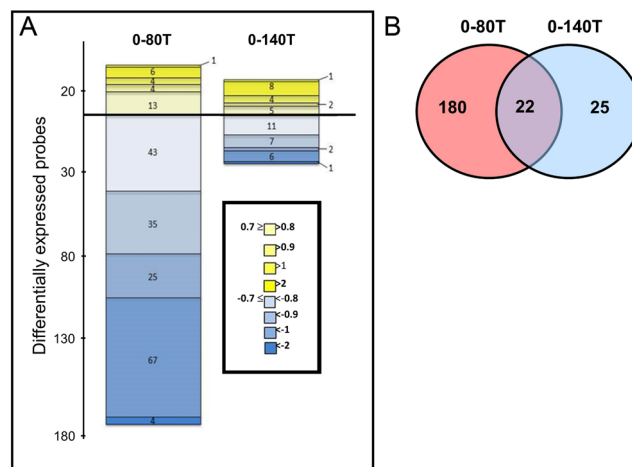
Confirmation of microarray changes was carried out by qPCR and comparable exposure-linked changes were observed for *POSTN* (decreased), *MHC class 1 HLA-B* (increased), and *PTGER3* (marginal decrease) (Supplementary Fig. 2 and Supplementary Table 2).

## Discussion

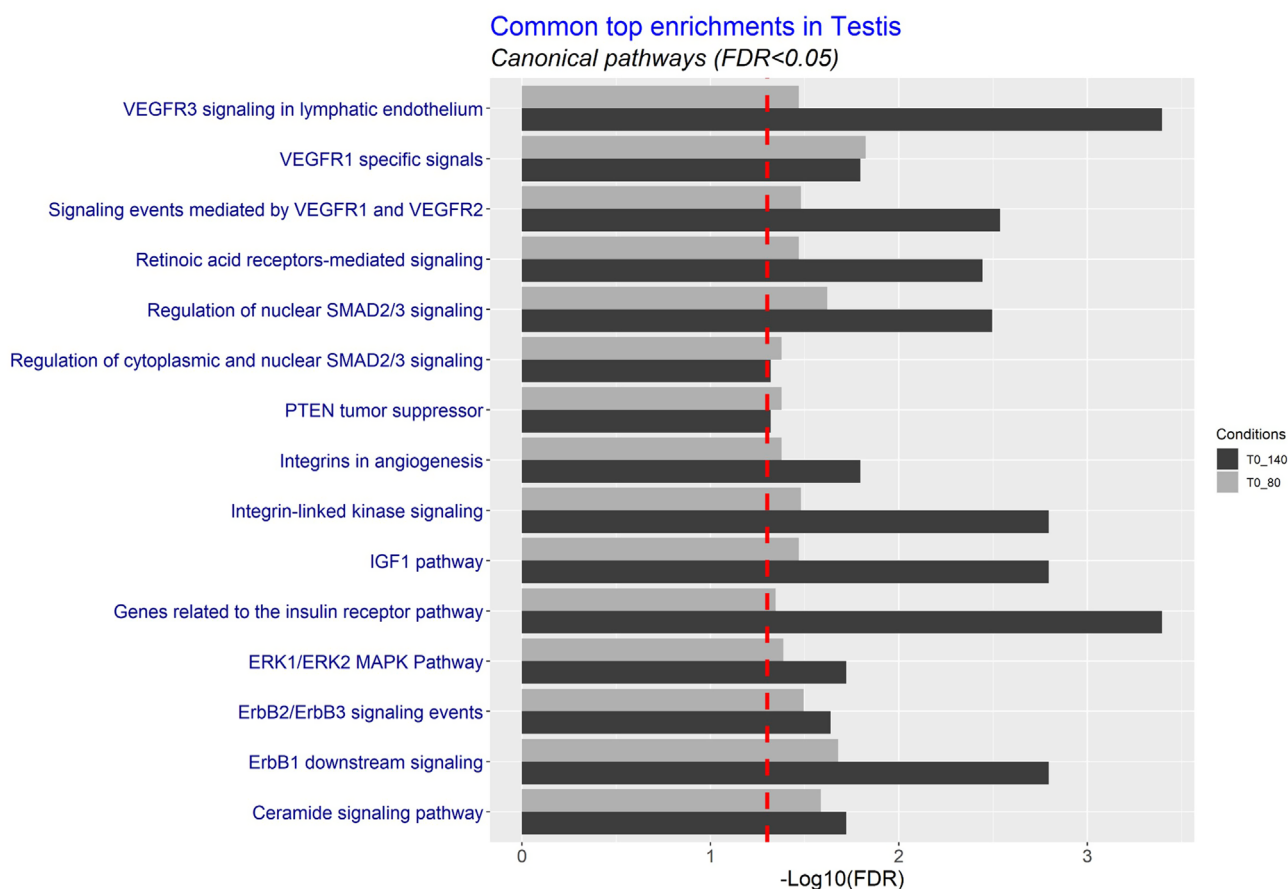
The current study identifies stages of male fetal gonad development in sheep that are developmentally

sensitive to an EC mixture relevant to human exposure. In the current paradigm, exposure occurred when pregnant ewes grazed pastures fertilised with biosolids generated from processed human sewage sludge. Sertoli cell number in day 140 fetuses exposed throughout gestation (0–140 days) remained unchanged but Leydig cell CYP17A1 and CYP11A1 staining was reduced. In contrast, exposure for periods of 80 days limited to early, mid, or late gestation reduced both Sertoli cell numbers and Leydig cell CYP17A1 immuno-reactivity in day 140 male fetal testes, regardless of developmental stage. Of note is that the reduced proportion of interstitial cells positive for CYP17A1 in the 30–110 and 60–140T exposure groups is not reciprocated with CYP11A1. Since fetal testosterone concentrations were reduced only in the 60–140 exposure group, this likely reflects fetal Leydig cell activity and raises the possibility of an exposure-induced reduction in the proportion of CYP17A1-positive testosterone-secreting Leydig cells. Intriguingly, exposure limited to 30–110 days was not sufficient to reduce testosterone but still reduced the proportion of CYP17A1. The lack of an effect on CYP11A1 may reflect differential sensitivity of the steroidogenic enzymes and awaits further characterisation of these cells at this late stage of gestation.

Notably, exposure for the first 80 days of gestation resulted in a dramatic change in the day 140 testis



**Figure 3** Analysis of differentially expressed (relative to control) fetal testis genes following maternal exposure to biosolids. At day 140, more genes were altered relative to control following biosolids exposure in the 0–80T than in the 0–140T exposure group. (A) Numbers and fold change of differentially expressed genes. Only probes that met an FDR of 5% and a threshold of  $\pm 0.7$  on the  $\log_2$ -transformed fold change (LogFC) are displayed. Probes are represented according to their positive (yellow) or negative (blue) fold change. (B) Venn diagram showing the number of differentially expressed transcripts unique to and common to both exposure groups. Day 140 testes from 0 to 80T exposed fetuses had  $>8\times$  the number of differentially expressed transcripts compared to testes from the 0 to 140T group.



**Figure 4** Common canonical testicular pathways following maternal exposure to biosolids in 0–80T and 0–140T groups. Common enrichment probes coding for canonical pathways were analysed using the eXploring Genomic Relations (XGR) web tool.

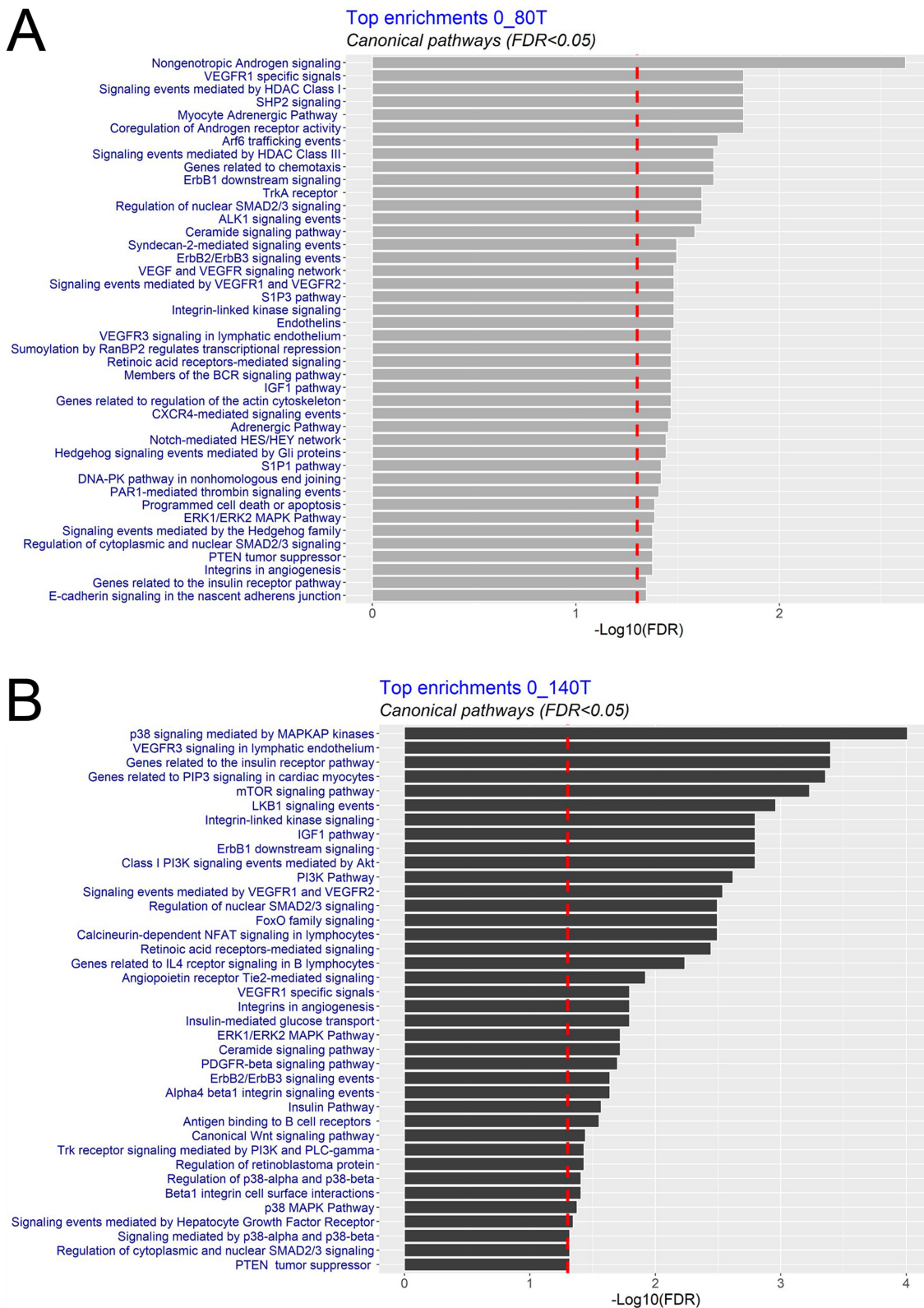
transcriptome, whereas exposure for the last 80 days induced phenotypic differences manifest by male fetuses with reduced body mass, reduced AGD, smaller testes and adrenals, and reduced fetal testosterone. Indeed, these phenotypic observations are consistent with an anti-androgenic effect on the male fetus (Recabarren *et al.* 2008, Scully *et al.* 2018). Taken together, these data indicate that fetal testes transiently exposed to a cocktail of environmental contaminants either do not have time to adapt and/or that exposure alters the cellular composition of the testis between days 80 and 140. This raises the possibility that the transcriptomic changes observed may reflect some of the cellular changes in transient exposure groups.

#### **Transient vs continuous fetal testis exposure**

Fetuses exposed to ECs for the first 80 days of gestation exhibited a greater change in testis transcriptome than those continuously exposed. Biological consistency can be drawn from similar transcriptomic changes in fetal ovaries examined from the same experimental paradigm. That is, a greater degree of ovarian

transcriptomic change was observed in transient vs continuously exposed female fetuses (Lea *et al.* 2016). Furthermore, relative to animals exposed from mating to 110 days, exposure limited to the pre-conception period reduced the proportion of mid-gestation type 1a ovarian follicles, reduced small blood vessel numbers in the fetal thyroid and specific to the males, increased thyroid organ weights. (Bellingham *et al.* 2013, Hombach-Klonisch *et al.* 2013). It is plausible that in the current study, the lesser degree of testicular transcriptomic change reflects adaptation to a longer and continuous period of exposure with less severe consequences for the fetus. In contrast, altering the environment by moving pregnant ewes from biosolids to control pastures or vice versa requires the fetal testis to adapt at more advanced developmental stages with different longer-term consequences. This applies to all three transient exposure groups (0–80T, 30–110T, 60–140T), and the shorter adaptation period of the 0–80T group may account for the greater phenotypic changes observed at day 140. Although transcriptomics was not carried out on day 140 fetal testes transiently exposed during mid or late gestation, our observations on Sertoli and Leydig





**Figure 5** Functional analysis of differentially expressed testis transcripts following maternal exposure to biosolids. Top canonical pathways are shown in (A) 0–80T and (B) 0–140T exposure groups. Analysis was carried out using the eXploring Genomic Relations (XGR) web tool.

**Table 2** Functional analysis of differentially expressed transcripts representative of specific biological pathways.

Treatment group	Associated network functions <sup>a</sup>	Score
0–80T	1. Cellular movement, cell morphology, drug metabolism	44
	2. Cell death, hematological system development and function, cell-mediated immune response	42 33
	3. Cell cycle, hair and skin development and function, connective tissue development and function.	27 25
	4. Developmental disorder, neurological disease, cardiovascular system development and function	
	5. Genetic disorder, metabolic disease, cellular assembly and organisation	
0–140T	1. Cell signalling, small molecule biochemistry, inflammatory response	34
	2. RNA post-transcriptional modification, cell to cell signalling and interaction, cellular assembly and organisation	30 3
	3. Embryonic development, organ development, organismal development	3
	4. Endocrine system disorders, gastrointestinal disease, genetic disorder	3
	5. Lipid metabolism, small molecule biochemistry, connective tissue disorders	

<sup>a</sup>Biological pathways identified by ingenuity pathway analysis (IPA) software (FDR < 7%).

cell markers support this theory. This raises concerns that a transient environmental change during fetal development may adversely affect male reproduction during adulthood, a critically important area not yet explored within the current paradigm.

These data are consistent with three previous studies demonstrating developmental exposure effects on testes from mid-gestation male fetuses, neonatal day 1 males, and post-weaning ram lambs (Paul *et al.* 2005, Bellingham *et al.* 2012, Elcombe *et al.* 2021). Mid-gestation male fetuses continuously exposed to day

110 exhibited reduced Sertoli and Leydig cell numbers and this was reflected by reduced inhibin A and testosterone. In the current study, the lack of an effect at day 80 likely reflects the earlier developmental stage and/or the shorter period of continuous exposure. In neonatal lambs exposed *in utero* via the mother, and in adult males exposed via the mother and for a period of 7 months following parturition, testicular abnormalities were observed in a cohort of male offspring, for example fewer germ cells and Sertoli cell-only tubules (Bellingham *et al.* 2012, Elcombe *et al.* 2021).

**Table 3** Biological functions associated with testicular differentially expressed genes following biosolids exposure.

Biological functions	Treatment groups			
	0–80T		0–140T	
	<i>P</i>	No. mols	<i>P</i>	No. mols
Diseases and disorders				
Dermatological disease/conditions	9.47E–04 – 5.96E–03	23	3.83E–05 – 4.58E–02	8
Genetic disorder	9.47E–04 – 4.86E–02	25	9.47E–04 – 5.96E–03	23
Cancer	1.06E–04 – 3.66E–02	45		
Reproductive system disease	3.83E–05 – 4.67E–02	27		
Gastrointestinal disease	9.47E–04 – 4.95E–02	11		
Connective tissue disorders			3.83E–05 – 4.67E–02	3
Inflammatory disease			9.47E–04 – 4.86E–02	13
Skeletal and muscular disorders			9.47E–04 – 4.95E–02	12
Molecular and cell functions				
Cell morphology	1.75E–04 – 3.01E–02	20	1.16E–03 – 4.48E–02	9
Cell assembly and organisation	3.24E–04 – 3.66E–02	29		
Cellular compromise	3.24E–04 – 3.66E–02	9		
Lipid metabolism	5.38E–04 – 3.38E–02	8		
Molecular transport	5.38E–04 – 3.66E–02	12		
Gene expression			4.53E–04 – 4.10E–02	3
Cell function and maintenance			8.66E–04 – 4.29E–02	8
Cell death			1.12E–03 – 4.67E–02	8
Cellular movement			1.41E–03 – 4.48E–02	6
Physiological system development and function				
Organismal development	2.62E–04 – 3.66E–02	37	1.12E–03 – 3.72E–02	8
Cardiovascular system	1.30E–03 – 2.93E–02	14	1.84E–03 – 3.72E–02	5
Tissue development	8.03E–04 – 3.66E–02	34		
Hair and skin	1.54E–03 – 3.10E–02	10		
Renal and urological	1.81E–03 – 3.66E–02	13		
Reproductive system			3.97E–04 – 2.56E–02	6
Organ development			1.84E–03 – 1.19E–02	4
Visual system			1.84E–03 – 1.99E–03	3

### Anti-androgenic effects

Our observations of reduced fetal AGD and impaired testis development support studies linking these observations with chemical exposures during fetal developmental (Gray *et al.* 2006). Furthermore, a common masculinisation programming period has been identified in rats that occurs just after testis differentiation (Welsh *et al.* 2008). During this period, exposure to an androgenic chemical reduces AGD concomitantly with the induction of hypospadias and cryptorchidism. In developing male fetal sheep, prenatal androgen exposure mid-gestation also alters gonadal development although an increase in AGD has been reported (Manikkam *et al.* 2004, Padmanabhan *et al.* 2010). Notably, we have previously reported that female fetuses exposed to biosolids via the mother have an increased AGD and postulated that this may be androgenic (Lea *et al.* 2016, Dorman *et al.* 2018). In the current study, the reduction in AGD was specific to smaller male fetuses and this effect characterised only the late gestation exposure group. However, the fact that CYP17A1 and testosterone were significantly reduced and other organ weights (e.g. thyroid) were not affected in this group argues for a dysregulation of androgen production and target tissue response, probably exacerbated by the concomitant reduced growth. An increased risk of urogenital defects, like hypospadias, is associated with small for gestational age in humans and this is associated with altered placental androgen production (O'Shaughnessy *et al.* 2019). This raises the possibility that biosolids chemicals may disturb placental function/androgen production, leading to an increased risk of impaired masculinisation.

Importantly, during fetal life and after the proliferation of germ cells, the stages of development between the testis and ovary are fundamentally different. In the ovine (and human) ovary, two essential steps take place: meiosis initiation and blockage of the oocyte at the end of prophase I and follicle formation (Mandon-Pepin *et al.* 2003). In the testis, germ cells remain quiescent until puberty. Despite this observation, female fetuses had greater sensitivity to exposure during mid and late gestation rather than in early gestation as described for the males. This may reflect the earlier rate of development of the testis compared to the ovary (Smith *et al.* 2014, Wilhelm *et al.* 2007) and/or the differential sensitivity of the testis and ovary to specific chemical types. Of note is that maternal liver phthalate was increased specifically in the 0–80T group while specific changes in liver PCBs and PBDEs characterised the mid and late gestation groups (Lea *et al.* 2016).

### Transcriptomic analyses

Although direct causality cannot be established, the combined chemical measurements and transcriptomic studies suggest that the developing gonad is less

well-equipped to respond to a transient change in environment, than to a consistent but abnormal change. In the current study, transcriptomic analyses highlighted drug metabolism as a key function of the differentially expressed genes in the 0–80T group but not the 0–140T group. Drug-metabolising enzymes such as the differentially expressed gonadal P450 (CYP) family genes reported in the current study are also reported to show altered expression during development (Hines 2008). We therefore propose that this may account for some of the differences between exposure groups. Also unique to this group were gene networks associated with development or disorders in a range of key body systems including haematological, connective tissue, neurological, cardiovascular, and metabolism. In contrast, high scoring networks in the 0–140T group were primarily linked to cell functions: signalling, interaction, assembly, and organisation, and these were also highlighted in the 0–80T group. Notably, our previous transcriptomic studies of biosolids-exposed fetal ovaries indicated that genes linked to drug metabolism were altered in the most perturbed 60–140T group (Lea *et al.* 2016). We postulated that activation of these genes following intermittent exposure of the fetus may impact later development and that this may account for the greater effect of transient exposures to biosolids. This raises the possibility that this may also apply to the developing male.

In the 0–80T group, the transcriptional regulator *SOHLH1*, associated with early spermatogonia differentiation and subsequent male fertility, was a top upregulated gene (Toyoda *et al.* 2014). In mice, *SOHLH1* is expressed in gonadal germ cells (fetal and post-natal) and is crucial for germ cell survival and spermatogenesis (Barrios *et al.* 2012). The dysregulation of *SOHLH1* may be associated with the altered androgen regulation of Sertoli cells and thus germ cell development. Furthermore, altered *SOHLH1* may also be indirectly linked to exposure-induced changes in testicular drug metabolisms pathways. These areas constitute lines of further investigation.

The upregulation of HLA antigens in both exposure groups and the testicular cell-mediated/inflammatory gene networks identified may reflect exposure-induced changes in immune function (Bansal *et al.* 2018). Since immunological competence in fetal lambs develops progressively throughout gestation (Nalubamba *et al.* 2008), markers such as pattern recognition receptors (TLR10) and chemokine receptors (CXCR5) provide potential follow-up genes for further investigation. For both exposure groups, RXR function and PPAR $\alpha$  activation constituted the top toxicological network. The PPAR receptors, including PPAR $\alpha$  (NR1C1), are expressed in mammalian fetal testis (Froment *et al.* 2006) and the alpha form is implicated in Sertoli cell metabolism (Rouiller-Fabre *et al.* 2015). Since maternal liver phthalate levels were increased in the 0–80T group

and rodent studies indicate phthalate toxicity effects on the testis are partly mediated by PPAR and RXR, phthalates may contribute to the effects of biosolids on the male ovine fetus (Ward *et al.* 1998, Ryu *et al.* 2008, Lea *et al.* 2016).

Androgen signalling was perturbed in the 0–80T group (DAVID) and this is consistent with the phenotypic effects observed in exposed male (reduced AGD, lowered testosterone, reduced fetal body, adrenal and testis mass) and female fetuses (e.g. increased AGD, increased testosterone, reduced fetal and uterus mass) (Lea *et al.* 2016). Angiogenic factors were also differentially affected across the two treated male groups along with classical cell signalling systems: retinoic acid, SMAD2/3, IGF1, ERK1/2/MAPK, and ErB1. The trend for downregulation of differentially expressed fetal testicular transcripts in the 0–80T group parallels the downregulation of fetal ovarian transcripts in fetuses from the same experiment (Lea *et al.* 2016). This may be reflective of an underlying epigenetic mechanism in both sexes. Interestingly, histone deacetylase signalling was altered in the 0–80T males, and in our previous study of the females, histone methylation genes were altered

## Conclusions

In conclusion, the data presented in this study demonstrate that the ovine fetal testis exhibits differential and temporal sensitivity to chemical mixtures present in pastures treated with biosolids fertiliser. We have demonstrated, for the first time, that a short period of environmental exposure applied for the first 80 days of gestation dramatically alters the fetal testis transcriptome at day 140. Furthermore, restricting the period of environmental change to the final 80 days of gestation induces phenotypic changes indicative of an anti-androgenic effect on the male fetus. Relative to the shorter periods of biosolids exposure, continuous exposure was associated with fewer differentially regulated genes and a lesser effect on the male phenotype (fetal, adrenal, testis mass, AGD, and fetal testosterone unaltered), we propose that a short period of exposure either does not allow the developing fetus to adapt and/or alters the cellular composition of the testis. Collectively, these data indicate that exposure of grazing ruminants to mixtures of chemicals contained in biosolids fertiliser may be linked to perturbations in testicular development. Given the relevance of the biosolids model to human exposures and the consumption of ruminant-derived food products, these data should be viewed as a concern for animal and human male reproductive health.

## Supplementary materials

This is linked to the online version of the paper at <https://doi.org/10.1530/REP-21-0235>.

<https://rep.bioscientifica.com>

## Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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## Author contribution statement

R G L, C C, P A F, and K D S conceived of and designed the study. B F E and A B carried out the histological studies and testicular cell counts. C C, B M P, B L, E P, and L J designed and analysed the microarray results. L P, B M P, and R G L carried out all qPCR studies and analyses. Z Z carried out all chemical analyses and academic input in interpreting chemical effects. R G L and K D S wrote the paper. P A F co-ordinated the REEF EU FP7 project which funded the work and R G L, K D S, and C C were work-package leaders.

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## References

- Andrade LP, Rhind SM, Rae MT, Kyle CE, Jowett J & Lea RG 2013 Maternal undernutrition does not alter Sertoli cell numbers or the expression of key developmental markers in the mid-gestation ovine fetal testis. *Journal of Negative Results in Biomedicine* **12** 2. (<https://doi.org/10.1186/1477-5751-12-2>)
- Bansal A, Filippini D, Campolo F, Gori M, Bramucci F, Pellegrini M, Ottolenghi S, Rossi P, Jannini EA & Dolci S 2012 SOHLH1 and SOHLH2 control kit expression during postnatal male germ cell development. *Journal of Cell Science* **125** 1455–1464. (<https://doi.org/10.1242/jcs.092593>)
- Bellingham M, Fowler PA, Amezaga MR, Rhind SM, Cotinot C, Mandon-Pepin B, Sharpe RM & Evans NP 2009 Exposure to a complex cocktail of environmental endocrine-disrupting compounds disturbs the kisspeptin/GPR54 system in ovine hypothalamus and pituitary gland. *Environmental Health Perspectives* **117** 1556–1562. (<https://doi.org/10.1289/ehp.0900699>)
- Bellingham M, Fowler PA, Amezaga MR, Whitelaw CM, Rhind SM, Cotinot C, Mandon-Pepin B, Sharpe RM & Evans NP 2010 Foetal hypothalamic and pituitary expression of gonadotrophin-releasing hormone and galanin systems is disturbed by exposure to sewage sludge chemicals via maternal ingestion. *Journal of Neuroendocrinology* **22** 527–533. (<https://doi.org/10.1111/j.1365-2826.2010.01974.x>)
- Bellingham M, McKinnell C, Fowler PA, Amezaga MR, Zhang Z, Rhind SM, Cotinot C, Mandon-Pepin B, Evans NP & Sharpe RM 2012 Foetal and post-natal exposure of sheep to sewage sludge chemicals disrupts sperm production in adulthood in a subset of animals. *International Journal of Andrology* **35** 317–329. (<https://doi.org/10.1111/j.1365-2605.2011.01234.x>)

- Bellingham M, Amezaga MR, Mandon-Pepin B, Speers CJ, Kyle CE, Evans NP, Sharpe RM, Cotinot C, Rhind SM & Fowler PA 2013 Exposure to chemical cocktails before or after conception – the effect of timing on ovarian development. *Molecular and Cellular Endocrinology* **376** 156–172. (<https://doi.org/10.1016/j.mce.2013.06.016>)
- Benjamini Y & Hochberg Y 1995 Controlling the false discovery rate: a practical and powerful approach to multiple testing. *Journal of the Royal Statistical Society: Series B* **57** 289–300. (<https://doi.org/10.1111/j.2517-6161.1995.tb02031.x>)
- Carlsen E, Giwercman A, Keiding N & Skakkebaek NE 1992 Evidence for decreasing quality of semen during past 50 years. *BMJ* **305** 609–613. (<https://doi.org/10.1136/bmj.305.6854.609>)
- Dorman DC, Chiu W, Hales BF, Hauser R, Johnson KJ, Mantus E, Martel S, Robinson KA, Rooney AA, Rudel R *et al.* 2018 Systematic reviews and meta-analyses of human and animal evidence of prenatal diethylhexyl phthalate exposure and changes in male anogenital distance. *Journal of Toxicology and Environmental Health: Part B, Critical Reviews* **21** 207–226. (<https://doi.org/10.1080/10937404.2018.1505354>)
- Elcombe CS, Monteiro A, Ghasemzadeh-Hasankolaei M, Evans NP & Bellingham M 2021 Morphological and transcriptomic alterations in neonatal lamb testes following developmental exposure to low-level environmental chemical mixture. *Environmental Toxicology and Pharmacology* **86** 103670. (<https://doi.org/10.1016/j.etap.2021.103670>)
- Fang H, Knezevic B, Burnham KL & Knight JC 2016 XGR software for enhanced interpretation of genomic summary data, illustrated by application to immunological traits. *Genome Medicine* **8** 129. (<https://doi.org/10.1186/s13073-016-0384-y>)
- Filis P, Walker N, Robertson L, Eaton-Turner E, Ramona L, Bellingham M, Amezaga MR, Zhang Z, Mandon-Pepin B, Evans NP *et al.* 2019 Long-term exposure to chemicals in sewage sludge fertilizer alters liver lipid content in females and cancer marker expression in males. *Environment International* **124** 98–108. (<https://doi.org/10.1016/j.envint.2019.01.003>)
- Fowler PA, Dora NJ, McFerran H, Amezaga MR, Miller DW, Lea RG, Cash P, McNeilly AS, Evans NP, Cotinot C *et al.* 2008 In utero exposure to low doses of environmental pollutants disrupts fetal ovarian development in sheep. *Molecular Human Reproduction* **14** 269–280. (<https://doi.org/10.1093/molehr/gan020>)
- Fromont P, Gizard F, Defever D, Staels B, Dupont J & Monget P 2006 Peroxisome proliferator-activated receptors in reproductive tissues: from gametogenesis to parturition. *Journal of Endocrinology* **189** 199–209. (<https://doi.org/10.1677/joe.1.06667>)
- Gray Jr LE, Wilson VS, Stoker T, Lambright C, Furr J, Noriega N, Howdeshell K, Ankley GT & Guillette L 2006 Adverse effects of environmental antiandrogens and androgens on reproductive development in mammals. *International Journal of Andrology* **29** 96–104; discussion 105–108. (<https://doi.org/10.1111/j.1365-2605.2005.00636.x>)
- Ha M, Sabherwal M, Duncan E, Stevens S, Stockwell P, McConnell M, Bekhit Ael Ael-D & Carne A 2015 In-depth characterization of sheep (*Ovis aries*) milk whey proteome and comparison with cow (*Bos taurus*). *PLoS ONE* **10** e0139774. (<https://doi.org/10.1371/journal.pone.0139774>)
- Hines RN 2008 The ontogeny of drug metabolism enzymes and implications for adverse drug events. *Pharmacology and Therapeutics* **118** 250–267. (<https://doi.org/10.1016/j.pharmthera.2008.02.005>)
- Hochereau-de Reviers MT, Perreau C, Pisselet C, Locatelli A & Bosc M 1995 Ontogenesis of somatic and germ cells in sheep fetal testis. *Journal of Reproduction and Fertility* **103** 41–46. (<https://doi.org/10.1530/jrf.0.1030041>)
- Hombach-Klonisch S, Danescu A, Begum F, Amezaga MR, Rhind SM, Sharpe RM, Evans NP, Bellingham M, Cotinot C, Mandon-Pepin B *et al.* 2013 Peri-conceptual changes in maternal exposure to sewage sludge chemicals disturbs fetal thyroid gland development in sheep. *Molecular and Cellular Endocrinology* **367** 98–108. (<https://doi.org/10.1016/j.mce.2012.12.022>)
- Istvan M, Rahban R, Dananche B, Senn A, Stettler E, Multigner L, Nef S & Garlandezec R 2021 Maternal occupational exposure to endocrine-disrupting chemicals during pregnancy and semen parameters in adulthood: results of a nationwide cross-sectional study among Swiss conscripts. *Human Reproduction* **36** 1948–1958. (<https://doi.org/10.1093/humrep/deab034>)
- Landrigan PJ 2017 Air pollution and health. *Lancet Public Health* **2** e4–e5. ([https://doi.org/10.1016/S2468-2667\(16\)30023-8](https://doi.org/10.1016/S2468-2667(16)30023-8))
- Lea RG, Amezaga MR, Loup B, Mandon-Pepin B, Stefansdottir A, Filis P, Kyle C, Zhang Z, Allen C, Purdie L *et al.* 2016 The fetal ovary exhibits temporal sensitivity to a ‘real-life’ mixture of environmental chemicals. *Scientific Reports* **6** 22279. (<https://doi.org/10.1038/srep22279>)
- Levine H, Jorgensen N, Martino-Andrade A, Mendiola J, Weksler-Derri D, Mindlis I, Pinotti R & Swan SH 2017 Temporal trends in sperm count: a systematic review and meta-regression analysis. *Human Reproduction Update* **23** 646–659. (<https://doi.org/10.1093/humupd/dmx022>)
- Mandon-Pepin B, Oustry-Vaiman A, Vigier B, Piumi F, Cribiu E & Cotinot C 2003 Expression profiles and chromosomal localization of genes controlling meiosis and follicular development in the sheep ovary. *Biology of Reproduction* **68** 985–995. (<https://doi.org/10.1095/biolreprod.102.008557>)
- Manikkam M, Crespi EJ, Doop DD, Herkimer C, Lee JS, Yu S, Brown MB, Foster DL & Padmanabhan V 2004 Fetal programming: prenatal testosterone excess leads to fetal growth retardation and postnatal catch-up growth in sheep. *Endocrinology* **145** 790–798. (<https://doi.org/10.1210/en.2003-0478>)
- Mesiano S, Hart CS, Heyer BW, Kaplan SL & Grumbach MM 1991 Hormone ontogeny in the ovine fetus. XXVI. A sex difference in the effect of castration on the hypothalamic-pituitary gonadotropin unit in the ovine fetus. *Endocrinology* **129** 3073–3079. (<https://doi.org/10.1210/endo-129-6-3073>)
- Nalubamba KS, Gossner AG, Dalziel RG & Hopkins J 2008 Differential expression of pattern recognition receptors during the development of foetal sheep. *Developmental and Comparative Immunology* **32** 869–874. (<https://doi.org/10.1016/j.dci.2007.12.007>)
- Nguyen TM, Shafi A, Nguyen T & Draghici S 2019 Identifying significantly impacted pathways: a comprehensive review and assessment. *Genome Biology* **20** 203. (<https://doi.org/10.1186/s13059-019-1790-4>)
- O’Shaughnessy PJ, Antignac JP, Le Bizet B, Morvan ML, Svechnikov K, Soder O, Savchuk I, Monteiro A, Soffientini U, Johnston ZC *et al.* 2019 Alternative (backdoor) androgen production and masculinization in the human fetus. *PLoS Biology* **17** e3000002. (<https://doi.org/10.1371/journal.pbio.3000002>)
- Padmanabhan V, Sarma HN, Savabieasfahani M, Steckler TL & Veiga-Lopez A 2010 Developmental reprogramming of reproductive and metabolic dysfunction in sheep: native steroids vs. environmental steroid receptor modulators. *International Journal of Andrology* **33** 394–404. (<https://doi.org/10.1111/j.1365-2605.2009.01024.x>)
- Park JS, Kim J, Elghiaty A & Ham WS 2018 Recent global trends in testicular cancer incidence and mortality. *Medicine* **97** e12390. (<https://doi.org/10.1097/MD.00000000000012390>)
- Paul C, Rhind SM, Kyle CE, Scott H, McKinnell C & Sharpe RM 2005 Cellular and hormonal disruption of fetal testis development in sheep reared on pasture treated with sewage sludge. *Environmental Health Perspectives* **113** 1580–1587. (<https://doi.org/10.1289/ehp.8028>)
- Pomerantz DK & Nalbandov AV 1975 Androgen level in the sheep fetus during gestation. *Proceedings of the Society for Experimental Biology and Medicine* **149** 413–416. (<https://doi.org/10.3181/00379727-149-38818>)
- Quirke LD, Juengel JL, Tisdall DJ, Lun S, Heath DA & McNatty KP 2001 Ontogeny of steroidogenesis in the fetal sheep gonad. *Biology of Reproduction* **65** 216–228. (<https://doi.org/10.1095/biolreprod65.1.216>)
- Recabarren SE, Rojas-García PP, Recabarren MP, Alfaro VH, Smith R, Padmanabhan V & Sir-Petermann T 2008 Prenatal testosterone excess reduces sperm count and motility. *Endocrinology* **149** 6444–6448. (<https://doi.org/10.1210/en.2008-0785>)
- Roselli CE, Amodei R, Gribbin KP, Corder K, Stormshak F & Estill CT 2016 Excess testosterone exposure alters hypothalamic-pituitary-testicular axis dynamics and gene expression in sheep fetuses. *Endocrinology* **157** 4234–4245. (<https://doi.org/10.1210/en.2016-1411>)
- Rouiller-Fabre V, Guerin MJ, N’Tumba-Byn T, Muczynski V, Moison D, Tourpin S, Messiaen S, Habert R & Livera G 2015 Nuclear receptors and endocrine disruptors in fetal and neonatal testes: a gapped landscape. *Frontiers in Endocrinology* **6** 58. (<https://doi.org/10.3389/fendo.2015.00058>)
- Ryu JY, Lee E, Kim TH, Lee YJ, Lee J, Lee BM, Kwack SJ, Jung KK, Han SY, Kim SH *et al.* 2008 Time-response effects of testicular gene expression profiles in Sprague-Dawley male rats treated with di(n-butyl) phthalate. *Journal of Toxicology and Environmental Health: Part A* **71** 1542–1549. (<https://doi.org/10.1080/15287390802391992>)

- Salian S, Doshi T & Vanage G** 2011 Perinatal exposure of rats to bisphenol A affects fertility of male offspring – an overview. *Reproductive Toxicology* **31** 359–362. (<https://doi.org/10.1016/j.reprotox.2010.10.008>)
- Scully CM, Estill CT, Amodei R, McKune A, Gribbin KP, Meaker M, Stormshak F & Roselli CE** 2018 Early prenatal androgen exposure reduces testes size and sperm concentration in sheep without altering neuroendocrine differentiation and masculine sexual behavior. *Domestic Animal Endocrinology* **62** 1–9. (<https://doi.org/10.1016/j.domaniend.2017.07.001>)
- Skakkebaek NE, Rajpert-De Meyts E & Main KM** 2001 Testicular dysgenesis syndrome: an increasingly common developmental disorder with environmental aspects. *Human Reproduction* **16** 972–978. (<https://doi.org/10.1093/humrep/16.5.972>)
- Skakkebaek NE, Rajpert-De Meyts E, Buck Louis GM, Toppari J, Andersson AM, Eisenberg ML, Jensen TK, Jorgensen N, Swan SH, Sapra KJ et al.** 2016 Male reproductive disorders and fertility trends: influences of environment and genetic susceptibility. *Physiological Reviews* **96** 55–97. (<https://doi.org/10.1152/physrev.00017.2015>)
- Smith P, Wilhelm D & Rodgers RJ** 2014 Development of mammalian ovary. *Journal of Endocrinology* **221** R145–R161. (<https://doi.org/10.1530/JOE-14-0062>)
- Sumner RN, Harris IT, Van der Mescht M, Byers A, England GCW & Lea RG** 2020 The dog as a sentinel species for environmental effects on human fertility. *Reproduction* **159** R265–R276. (<https://doi.org/10.1530/REP-20-0042>)
- Swan SH, Elkin EP & Fenster L** 2000 The question of declining sperm density revisited: an analysis of 101 studies published 1934–1996. *Environmental Health Perspectives* **108** 961–966. (<https://doi.org/10.1289/ehp.00108961>)
- Toyoda S, Yoshimura T, Mizuta J & Miyazaki J** 2014 Auto-regulation of the *Sohlh1* gene by the SOHLH2/SOHLH1/SP1 complex: implications for early spermatogenesis and oogenesis. *PLoS ONE* **9** e101681. (<https://doi.org/10.1371/journal.pone.0101681>)
- Viguie C, Chaillou E, Gayraud V, Picard-Hagen N & Fowler PA** 2020 Toward a better understanding of the effects of endocrine disrupting compounds on health: human-relevant case studies from sheep models. *Molecular and Cellular Endocrinology* **505** 110711. (<https://doi.org/10.1016/j.mce.2020.110711>)
- Ward JM, Peters JM, Perella CM & Gonzalez FJ** 1998 Receptor and nonreceptor-mediated organ-specific toxicity of di(2-ethylhexyl) phthalate (DEHP) in peroxisome proliferator-activated receptor alpha-null mice. *Toxicologic Pathology* **26** 240–246. (<https://doi.org/10.1177/019262339802600208>)
- Welsh M, Saunders PT, Fiskin M, Scott HM, Hutchison GR, Smith LB & Sharpe RM** 2008 Identification in rats of a programming window for reproductive tract masculinization, disruption of which leads to hypospadias and cryptorchidism. *Journal of Clinical Investigation* **118** 1479–1490. (<https://doi.org/10.1172/JCI34241>)
- Wilhelm D, Palmer S & Koopman P** 2007 Sex determination and gonadal development in mammals. *Physiological Reviews* **87** 1–28. (<https://doi.org/10.1152/physrev.00009.2006>)
- Wohlfahrt-Veje C, Main KM & Skakkebaek NE** 2009 Testicular dysgenesis syndrome: foetal origin of adult reproductive problems. *Clinical Endocrinology* **71** 459–465. (<https://doi.org/10.1111/j.1365-2265.2009.03545.x>)

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