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The respiratory DC/macrophage network at steady-state and upon influenza infection in the swine biomedical model

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Human and mouse respiratory tracts show anatomical and physiological differences, which will benefit from alternative experimental models for studying many respiratory diseases. Pig has been recognized as a valuable biomedical model, in particular for lung transplantation or pathologies such as cystic fibrosis and influenza infection. However, there is a lack of knowledge about the porcine respiratory immune system. Here we segregated and studied six populations of pig lung dendritic cells (DCs)/macrophages (M θ s) as follows: conventional DCs (cDC) 1 and cDC2, inflammatory monocyte-derived DCs (moDCs), monocyte-derived M θ s, and interstitial and alveolar M θ s. The three DC subsets present migratory and naive T-cell stimulation capacities. As observed in human and mice, porcine cDC1 and cDC2 were able to induce T-helper (Th)1 and Th2 responses, respectively. Interestingly, porcine moDCs increased in the lung upon influenza infection, as observed in the mouse model. Pig cDC2 shared some characteristics observed in human but not in mice, such as the expression of FC ϵ RI α and Langerin, and an intra-epithelial localization. This work, by unraveling the extended similarities of the porcine and human lung DC/M θ networks, highlights the relevance of pig, both as an exploratory model of DC/M θ functions and as a model for human inflammatory lung pathologies.

INTRODUCTION

Pig is being developed as a model for several respiratory pathologies leading to over-inflammation, such as cystic fibrosis,^{1,2} lung transplantation,³ and influenza A virus (IAV) infection.⁴ Indeed, pig and human respiratory systems present several anatomical, histological, physiological, and biochemical similarities.⁵ In addition, the pig immune system appears relatively close to the human one, as orthology preservation analysis on immune genes showed a greater similarity between human and pig than between human and mouse.⁶ For instance, and conversely to the murine ones, porcine alveolar macrophages (AMs) show no evidence of nitric oxide production after lipopolysaccharide and/or Concanavalin A stimulations⁷ exhibiting similar behavior to human AMs (for review, see ref. 8). Even if its immune system is one of the best

characterized after the human, primate, and murine ones,⁹ so far the few studies that have described pig's lung immune cells did not enter in dendritic cell (DC) and macrophage (M θ) subpopulation details,^{10,11} limiting its use as a preclinical model for respiratory pathologies.

One of the main components of the respiratory immune system is the DC/M θ network involved in sensing foreign antigens, controlling inflammation, and initiating the adaptive immune responses. For the sake of clarity, we will use an adaptation of a recently proposed nomenclature from Guilliams *et al.*,¹² distinguishing two levels of identification. The first level focuses on the origin of the cell-type progenitor (adult bone-marrow proDC for conventional DCs (cDCs), adult blood monocytes for monocyte-derived cells (moCells), or embryonic monocyte-derived precursors settled in peripheral tissues for

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M0s). The second level focuses on the cell functions (M0-like or DC-like).

In the mouse lung, four different DC/M0 subtypes have been described so far. Two types of cDCs arise from FLT3-dependent proDC. The cDC1 subset (CD103^{pos}/CD172a^{neg}/XCR1^{pos}) is mainly involved in the induction of primary CD8 and T-helper 1 (Th1) immune responses, whereas the cDC2 subset (CD11b^{pos}/CD172a^{pos}/XCR1^{neg}) is involved in the induction of Th2 lung immune responses. AMs derive from local precursors settled in the lungs before birth, which renew the AM pool in a bone marrow-independent manner.^{13,14} Finally, an inflammatory monocyte-derived DC (moDC) subset attracted to inflammatory tissues through the CCR2/CCL2 axis was recently defined as CD64^{pos}/FcεRIα^{pos}.¹⁵

In humans, CD141^{pos}/cDC1 and CD1c^{pos}/cDC2 have been well characterized in blood and skin,¹⁶ and their functions mainly correspond to the cDC1/cDC2 mouse paradigm. In the lung, DCs presenting both phenotypes have been described,^{16,17} although only one publication specifically addressed their functions, finding that both cDC1 and cDC2 presented antigens equally well to CD8⁺ memory T cells.¹⁸

Human AMs have been widely studied¹⁹ and behave similarly to the mouse ones, with the exception of nitric oxide production. However, no lung inflammatory moDCs have ever been described in the human lung.

We have previously shown that in the skin, porcine and human DC networks were very similar^{20,21} (for review, see ref. 22). Therefore, pig appears as a pertinent model to explore which murine lung DC/M0 functions can be extrapolated to other mammals, including humans, at steady state and upon pathological inflammation.

Unlike mice, pigs are natural hosts for IAV. They show identical symptoms and are infected by the same subtypes as humans (H1N1 and H3N2). Although H1N2 viruses have so far only been described in pigs, they might present some zoonotic potential,²³ like other pig IAV, as highlighted by the 2009 H1N1 pandemic of swine origin.²⁴ Similarly to humans, disease severity in IAV infection in pigs have been associated with increased local pro-inflammatory cytokines.^{10,25} In mice, inflammatory moDCs appear to have an important role in the IAV pathogenesis. Indeed, pharmacological or genetic downregulation of the trafficking of these cells moderates the inflammation without impacting the adaptive immune response and reduces morbidity and mortality.²⁶ Upon infection, AMs are thought to mainly downmodulate the pathogen-induced inflammation¹⁹ and to scavenge cell debris.²⁷ Thus, it seems relevant to confirm these results in other species than mouse, where AMs and DCs would then become potential therapeutic targets to reduce the pathological inflammation induced by some IAV. Therefore, pig appears as an ideal experimental animal to study different aspects of the interaction between IAV and the lung immune system.

In this study, we finely define for the first time the phenotypes and functions of DC/M0 populations in the different compartments of the swine respiratory tract at steady state and upon IAV infections.

RESULTS

Porcine lung MHCII^{high} cells can be divided into six populations of DCs and M0s

Based on our work on porcine skin,^{20,21} several putative DC populations were identified as MHCII^{high} in the porcine lung parenchyma (Figure 1a). Among the MHCII^{high} cells, five populations were distinguished using the CD163 and CD172a (Sirpα) surface markers. Population 1 was the less frequent (5.6 ± 5.1%) and the only negative one for CD172a expression (Figures 1a,e). Thus, it will hereafter be named CD172a^{neg}. The other four MHCII^{high} populations expressed CD172a and will be named based on their differential CD163 expression level: 2/CD163^{neg} (11.8 ± 12.3%), 3/CD163^{low} (17.4 ± 4.3%), 4/CD163^{int} (7.4 ± 4.3%), and 5/CD163^{high} (53.8 ± 28.1%). Finally, in the bronchoalveolar lavage (BAL), we also identified one homogeneous MHCII^{pos} population as AMs (Figure 1b). These cells were CD172a^{pos}/CD163^{high}.

In the parenchyma, the CD172a^{neg} population was CD11b-like^{neg}/CD1^{neg}/CadM1^{pos} and did not express the antigen recognized by an anti-human mannose receptor (MR-like) (Figure 1c). The CD163^{neg} population expressed low levels of CD1 and MR-like, and presented both CD11b-like^{pos} and CD11b-like^{neg} subpopulations. It also expressed CadM1 but at a lower level than the CD172a^{neg} population. The CD163^{low} population was CD11b-like^{pos}/CD1^{neg}/CadM1^{pos}/MR-like^{pos}. The CD163^{int} population was CD11b-like^{pos}/CD1^{neg}/CadM1^{low}/MR-like^{pos} and was the only one expressing CD14. The CD163^{high} population did not express CD11b-like, highly expressed MR-like and was autofluorescent, like AMs from BAL. AMs but not CD163^{high} cells expressed low levels of CadM1 in three out of four animals. To test the possibility that parenchymal CD163^{high} cells were AMs contaminating parenchymal samples, we differentially stained alveolar and parenchymal cells by injecting carboxyfluorescein succinimidyl ester (CFSE) in the bronchiole leading to the intermediate lobe alveoli. Next, the lobe was massaged for a good CFSE diffusion and cells from each compartment were collected separately. As expected, AMs showed strong CFSE staining due to the fact that they were directly in contact with injected CFSE. Conversely, none of the parenchymal cells, including CD163^{high} cells, were CFSE stained (Figure 1d), indicating that AMs and CD163^{high} parenchymal cells belonged to distinct compartments. In the upper respiratory track, the same five different populations of MHCII^{high} cells were detected in the tracheal mucosa, exhibiting a similar phenotype (Supplementary Data 1a online).

The different populations were sorted and stained on slides using May–Grünwald–Giemsa coloration, to analyze their morphology. As shown in Figure 1f, CD172a^{neg}, CD163^{neg}, and CD163^{low} cells showed DC morphology with dendrites and irregular serrated nuclei, whereas CD163^{int}, CD163^{high}, and AM cells clearly resembled M0s with a vacuolated cytoplasm and a round nucleus.

Lung parenchymal MHCII^{high} cells can be divided in cDCs, moDCs, moM0s, and M0s

To assess DC/M0 canonical genes expression by reverse transcriptase-quantitative PCR (qPCR), all six populations

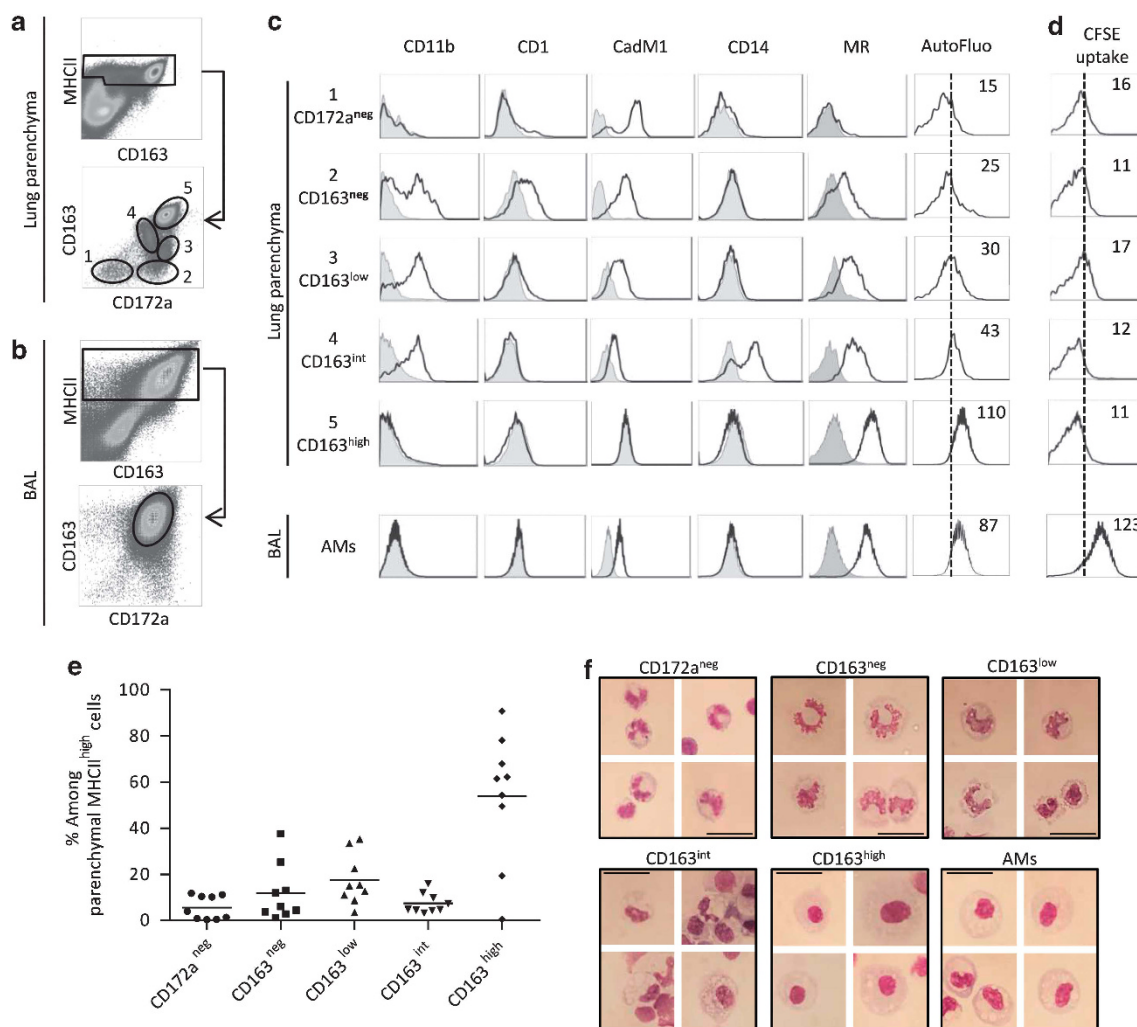


Figure 1 Phenotype of lung dendritic cells (DCs) and macrophages (Mφs). Bronchoalveolar lavages (BALs) and lung parenchymal cells were stained for multi-color flow cytometry. **(a)** In the parenchyma, DCs were identified as MHCII^{high} and divided into five subsets based on the expression of CD163 and CD172a. **(b)** In BAL, alveolar Mφ (AMs) were MHCII^{pos}. **(c)** Expression of the phenotypic markers CD11b-like, CD1, CadM1, CD14, and mannose receptor (MR)-like on DC subsets and AMs, along with their autofluorescence, were assessed by flow cytometry. The filled histogram represents the appropriate control isotype. **(d)** Cells present in the lumen of the intermediate lobe were carboxyfluorescein succinimidyl ester (CFSE) stained before being collected in the BAL. Next, parenchymal cells were extracted and CFSE expression was assessed by flow cytometry. Results are representative of at least three independent experiments. Numbers represent geometric fluorescence mean. **(e)** Percentages of each subset among total MHCII^{high} cells in the parenchyma. Each point represents one animal. **(f)** Cells were sorted, spun onto slides, and stained by a May–Grünwald–Giemsa coloration, to observe their morphology. Scale bar, 50 μm. All data are representative of at least three independent experiments.

were sorted. As shown in **Figure 2a**, both CD172a^{neg} and CD163^{neg} populations expressed high levels of FLT3, suggesting that they belong to the cDC lineage. CD172a^{neg} cells were the only ones expressing XCR1. These data, along with their CadM1^{pos}/CD11b-like^{neg}/MR-like^{neg} phenotype, indicate their adscription to the cDC1 subset. CD163^{neg} cDCs were FLT3^{high}/CD172a^{pos}/XCR1^{neg}, indicating that they correspond to the cDC2 subset. As observed in human and mouse cDC2,^{28,29} CD163^{neg} cDCs also expressed the myeloid genes *CSF1R*, *CCR2*, and *CX3CR1*, whose expressions overlapped with monocyte-derived cells. The CD163^{neg} cells, like human cDC2, expressed higher levels of Langerin³⁰ and FcεRIα³¹ compared with all other subtypes. As CD163^{neg} cells presented a bimodal expression of CD11b-like, we sought to

test CD11b-like^{neg} and CD11b-like^{pos} cells separately for cDC2 markers: both cell populations expressed FcεRIα (data not shown), comforting the belonging of the overall CD163^{neg} DC population to the cDC2 subset.

CD163^{low} cells expressed the myeloid genes *MerTK* and *CD64* at an intermediate level between cDCs and AMs, whereas they expressed less *CCR2* and *CX3CR1* than cDC2 but more than AMs. They also expressed the Mφ marker *CSF1R* but to a lesser extent than the CD163^{int} cells. Along with their DC morphology, these data suggest that they correspond to moDCs. Interestingly, and in parallel with their CadM1 expression, CD172a^{neg} but also CD163^{neg} and CD163^{low} cells expressed CD103, a cDC1 marker in mouse lung. The CD163^{int} population showed the same gene expression profile as

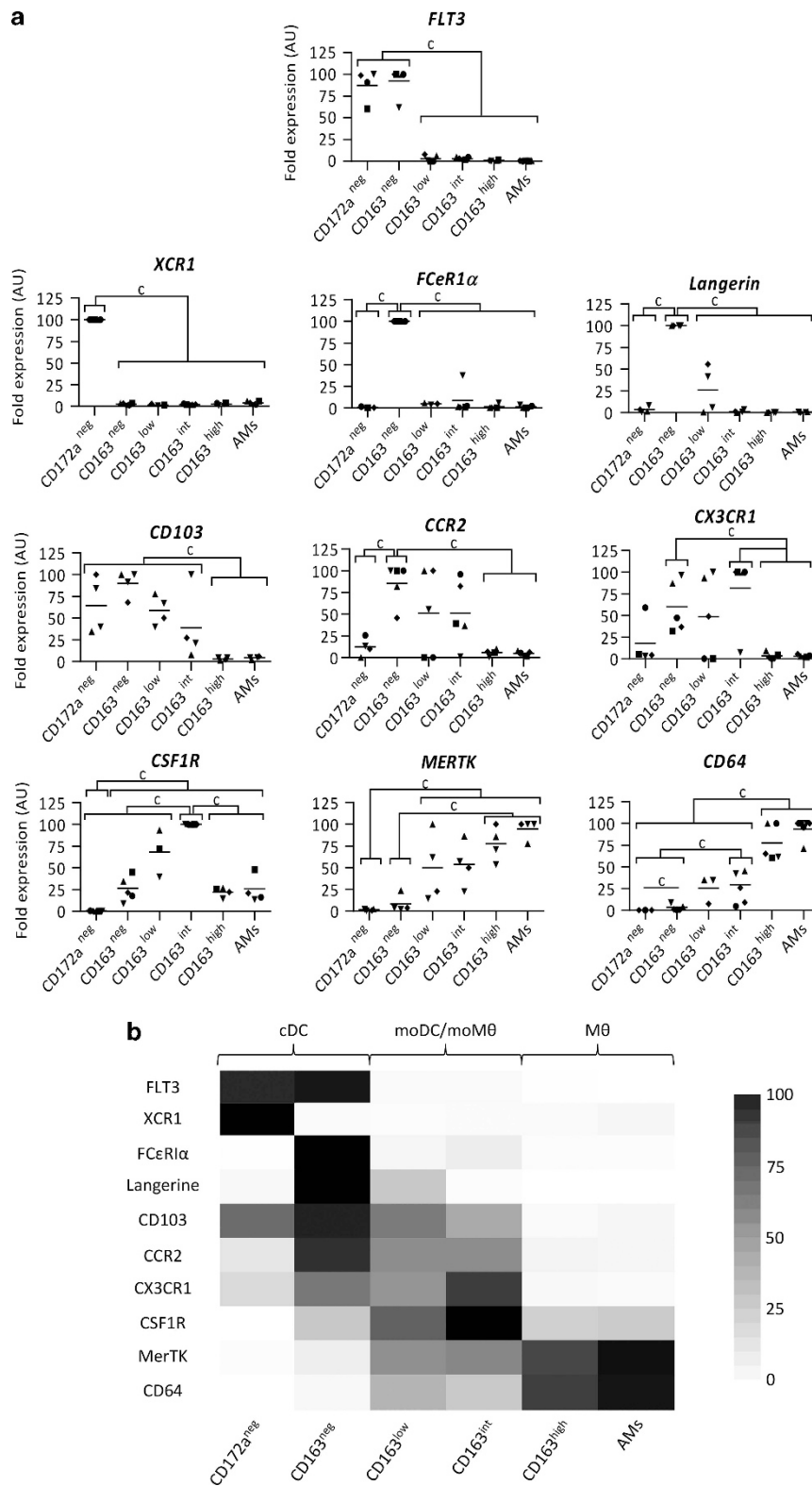


Figure 2 Lung parenchymal MHCII^{high} cells can be segregated in conventional dendritic cells (cDCs), monocyte-derived cells, and macrophages (M0s). The five DC subsets and alveolar M0s (AMs) were gated as in **Figure 1** and sorted by flow cytometry. **(a)** mRNA expression levels of FLT3, XCR1, FcεR1α, Langerin, CD103, CCR2, CX3CR1, CSF1R, merTK, and CD64 in DC subsets and AMs were assessed by quantitative real-time PCR. For each gene, data were normalized to the reference gene *RPS24* (ribosomal protein S24) expression and presented as relative expression (arbitrary units (AUs)): for each animal, the population with the highest expression for this gene was considered as 100 and the other populations were normalized to it. Each symbol represents one animal. **(b)** Heat map synthesizing the means (color scale on the right) of real-time PCR data from **(a)**. Statistic signs c: $P < 0.05$, b: $P < 0.01$, a: $P < 0.001$ with a Mann–Whitney test. moDC, monocyte-derived DC; moM0, monocyte-derived macrophages; M0, macrophages.

CD163^{low} DCs but it did show M θ morphology as well, which suggest that they could belong to a monocyte-derived M θ (moM θ) population. Finally, AMs and parenchymal CD163^{high} cells presented a very similar expression pattern, with no expression of CCR2 and CX3CR1, and a higher expression of MerTK and CD64 compared with all the other cells.

This whole transcriptomic profile was similar in tracheal mucosa cells (**Supplementary Data 1b**).

CD172a^{neg}, CD163^{neg}, and CD163^{low} cells present DC functionalities, whereas CD163^{int}, CD163^{high}, and AMs behave like M θ s

Two of the main properties of mature DCs are their capacities to migrate toward the lymph node against a CCL21 gradient and to activate naive T cells. Thus, we tested the capacity of *in vitro*-matured lung populations to migrate toward CCL21 in a transwell assay. Strikingly, the three populations presenting DC morphology, i.e., the CD172a^{neg}, CD163^{neg}, and CD163^{low} cells migrated toward CCL21 (**Figure 3a**). To compare the migration of the various populations in spite of their different proportions in the upper chamber, we calculated their migration index³² as the percentage of cells having migrated for each population (**Figure 3b**). The results highlighted that both cDCs had a higher migration capacity compared with CD163^{low} cells, and that CD163^{int}, CD163^{high}, and AM subsets did not substantially migrate toward CCL21.

We next examined whether these subsets possessed the capacity to induce naive allogeneic T-cell proliferation (**Figure 3c–e** and **Supplementary Data 2**). As lungs were obtained from conventional, outbred animals, we experienced high variability in the percentages of responding T cells according to the degree of allogeneicity of each DC/T-lymphocyte match. For instance, CD4⁺ T-cell spontaneous proliferation ranged from 0.2 to 8.0%, and upon CD163^{neg} cells stimulation from 4.6 to 52.5%. Similarly, CD8⁺ T-cell spontaneous proliferation ranged from 1.0 to 15.6%, and upon CD163^{neg} cells stimulation from 7.3 to 55.5%. CD3⁺CD4⁻CD8⁻ $\gamma\delta$ T cells³³ presented a higher spontaneous proliferation ranging from 0.8 to 42.8%, and upon CD163^{neg} stimulation reached 4.1 to 66.7% of proliferation. We thus compared the proliferation index of each experiment as described in Methods. As shown in **Figure 3c**, CD163^{neg} cDCs were the best at inducing proliferation of CD4⁺ T cells, followed by CD172a^{neg} cells. The two cDC subsets were equally potent at inducing CD8⁺ T-cell proliferation (**Figure 3d**). CD163^{low} moDCs induced variable proliferation of CD8⁺, CD4⁺, and $\gamma\delta$ T cells, although without reaching significance. In agreement with their assignment to the M θ types, the CD163^{int}, CD163^{high}, and AM populations did not stimulate T-cell proliferation.

Subsequently, the capacity of each DC/M θ population to skew differentiation of naive allogeneic CD4⁺ T cells in different Th subsets was tested *in vitro*. CD172a^{neg} cDCs induced differentiation of interferon- γ -producing Th cells (**Figure 3f**). CD163^{neg} cDCs induced interleukin (IL)-13-producing Th cells (**Figure 3g**), whereas the three DC populations induced variable degrees of IL-17-producing Th cells (**Figure 3h**). Next, the capacity of each subset to produce inflammatory cytokines

after a lipopolysaccharide/polyinosinic:polycytidylic acid stimulation was studied. The CD163^{low} subset produced both IL-1 β (**Figure 3i**) and IL-8 (**Figure 3j**). Combining their phenotype, morphology, transcriptome, and functional capacities, we can assess that CD172a^{neg}/XCR1^{pos} cells and CD163^{neg}/CD172a^{pos} cells correspond to cDC1 and cDC2, whereas CD163^{low} cells may correspond to the inflammatory moDC type, as they present monocyte lineage features and DC capacities. The CD163^{int} population also presented monocyte-derived cell characteristics, but showed M θ features, leading us to name them moM θ s. Finally, CD163^{high} cells could be considered as ‘interstitial’ AMs.

DC/M θ localization in the respiratory tract

Both cDC subsets were more represented in the bronchus and trachea than in the lung parenchyma (**Figure 4**). The CD163^{low}/moDC population was equally represented all along the respiratory tract. Finally, the CD163^{int}/moM θ s and the CD163^{high} ‘interstitial’ AMs were overrepresented in the lower part of the lung.

DC/M θ sublocalization in respiratory tissues

Tissue sublocalization was assessed by confocal microscopy (**Figure 5**). CD172a^{neg}/cDC1 cells were interstitial, mainly localized around the alveoli in the lung parenchyma. CD163^{neg}/cDC2 cells had an intra-epithelial localization in tracheal and bronchial mucosa, and a sub-epithelial localization in the lung parenchyma bronchioles. The CD163^{low}/moDCs and CD163^{int}/moM θ populations cannot be distinguished from one another on these images. Parenchymal CD163^{high} cells could be distinguished from AMs in the lung interstitium and in the alveoli lumen, respectively.

IAV infection induces moDCs recruitment

Having characterized the equivalents of murine cDC1, cDC2, moDCs, moM θ s, and AMs in the porcine respiratory tract, we then assessed their recruitment upon IAV infection. Two field-isolated strains, swine (sw)H1N2 and swH3N2,³⁴ were tested in order to be able to draw robust, nonstrain-specific conclusions. Three groups of six conventional pigs were inoculated with phosphate-buffered saline (PBS), swH3N2, or swH1N2 (**Figures 6a–c**), and lungs were collected at 2 days post infection. Cells were extracted from the lobe presenting the higher macroscopic lesion score (cardiac lobe, arrow in **Figure 6a**) without performing BAL; thus, AMs and CD163^{high} cells could not be distinguished in this experiment. All populations were counted and analyzed by flow cytometry to measure their percentage among total cells (**Figures 6d–f**). The absolute number of each sub-population per cardiac lobe was then calculated. Interestingly, the CD163^{low}/moDCs population was the only one that significantly increased after both swH3N2 and swH1N2 infections (**Figure 6g**). Sorted lung DCs were tested for transcriptomic expressions of IL-12A, IL-4, IL-13, and IL-6, cytokines involved in the induction of respectively Th1, Th2, Th2, and Th17 cells.³⁵ CD172a^{neg}/cDC1 produced more IL-12A mRNA, the Th1 inducer cytokine, than CD163^{neg}/cDC1 and CD163^{low}/moDCs, both in mock

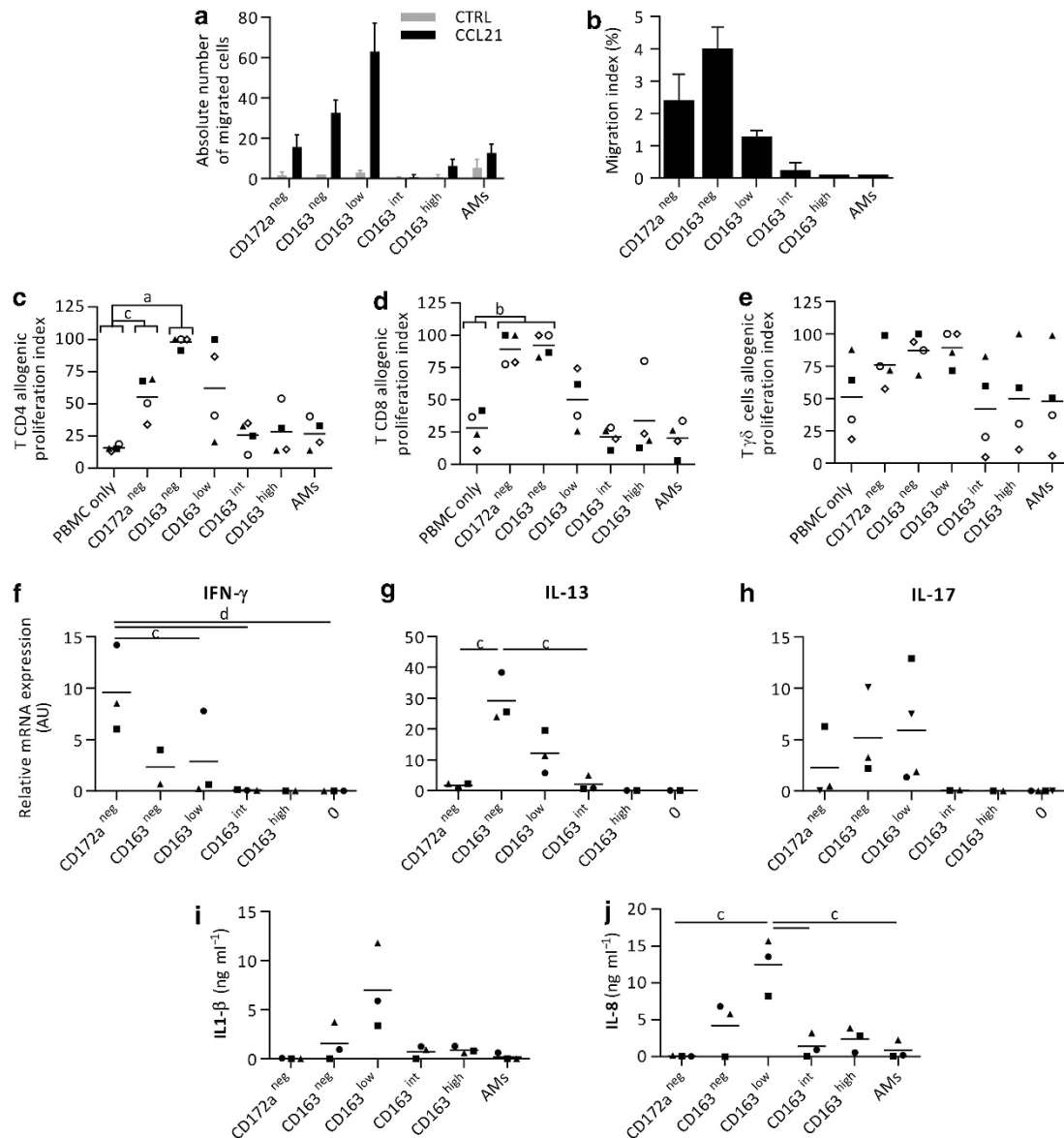


Figure 3 CD172a^{neg}, CD163^{neg}, and CD163^{low} cells present functionalities of respectively conventional dendritic cells (cDC)1, cDC2, and inflammatory (inf)DC. Bronchoalveolar lavage (BAL) cells and parenchymal cells were isolated and peripheral blood mononuclear cells (PBMCs) collected as described in the Methods section. DC-enriched parenchymal cells or BAL cells were laid on transwell inserts in contact with 400 μ l of CCL21 previously deposited in the lower chamber. Migrated DCs were collected after 2 h at 37 $^{\circ}$ C, stained, and counted by flow cytometry. Results are depicted either as the absolute number of migrated cells (**a**) or as the following percentage for each population: (number of migrated cells)/(initial number of cells deposited in the upper chamber) \times 100 (migration index) (**b**). Data are the means of triplicates and are representative of three different experiments. (**c,d,e**) DCs were sorted and plated out at a 1:8 ratio with 10^5 carboxyfluorescein succinimidyl ester (CFSE)-labeled allogeneic PBMCs from specific pathogen-free (SPF) pigs for 5 days at 37 $^{\circ}$ C. Proliferation of CD3⁺CD4⁺ (CD4⁺ T cells), CD3⁺CD4⁻CD8 α ⁺ (CD8⁺ T cells), and CD3⁺CD4⁻CD8 α ⁻ (γ δ T cells) was assessed by CFSE dilution (see **Supplementary Data 2** for gating strategy). For each animal, the highest proliferation value was considered as 100 and the other populations were normalized to it. (**f,g,h**) Lung DCs and macrophages (M ϕ) were sorted and plated out at a 1:10 ratio with previously enriched 2.5×10^5 allogeneic naive CD4⁺ T cells for 3 days. Total RNA was extracted and expressions of mRNA encoding interferon (IFN)- γ , interleukin (IL)-13, and IL-17 were assessed by quantitative reverse transcriptase-PCR (qRT-PCR). Data were normalized to the expression of the reference gene *RPS24* (ribosomal protein S24) and are representative of three distinct experiments. (**i,j**) The different DC subsets were sorted and cultured for one night with lipopolysaccharide (LPS) and polyinosinic:polycytidylic acid (poly:IC). Interleukins IL-8 and IL-1 β concentrations in the supernatants were assessed by cytokine beads analysis (CBA). Each symbol represents one animal. d: $P < 0.06$, c: $P < 0.05$, b: $P < 0.01$, a: $P < 0.001$ with a Student's *t*-test.

and IAV-infected animals (**Figure 6h,i**) in agreement with their Th1-inducing capacities in allogeneic reactions (**Figure 3h**). Neither IL-13 nor IL-4 transcripts, the Th2-inducing cytokines, were detected (data not shown). Finally, no

differences in IL-6 transcription (**Figure 3i**) were observed between the three DC subsets, both at steady state and upon infections, in agreement with the absence of a specific allogeneic Th17-inducing DC subset.

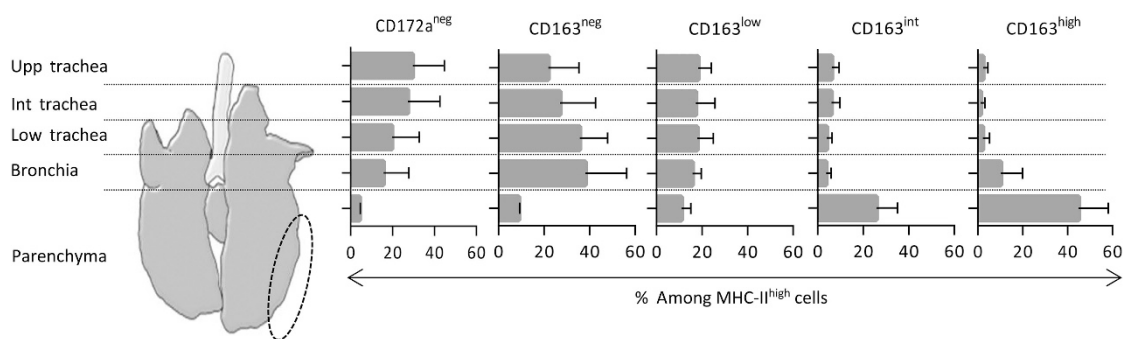


Figure 4 Dendritic cells (DCs) and macrophages (M0s) repartition in the respiratory tract. The five DC/M0 subsets were gated as in **Figure 1** and their percentage among all MHCII^{high} cells were compared in lung parenchyma, bronchial epithelium, and three different parts of the tracheal mucosa. Int, intermediate; Low, lower; Upp, upper.

CD163^{neg}/cDCs and CD163^{low}/moDCs can present IAV NP antigen to CD4⁺ effector/memory T cells

To further investigate the functions of lung DCs, we tested the capacity of each DC/M0 lung population to stimulate antigen-specific CD4⁺ and CD8⁺ effector/memory cells, using enriched spleen T cells from H1N1-vaccinated pigs and autologous cell-sorted lung DC/M0 loaded with purified IAV nucleoprotein (NP). Given the difficulty of such an experiment, it was only performed on three pigs, which obviously led to an important variability in the results. Both CD163^{neg}/cDC2 and CD163^{low}/moDCs appeared capable of presenting NP antigens to effector/memory CD4⁺ T cells (**Figure 7a** and **Supplementary Data 2**). Surprisingly, despite their strong capacity to activate naive allogeneic CD4⁺ and CD8⁺ T cells, to produce IL-12A and to induce Th1 cells, CD172a^{neg}/cDC1 appeared unable to present antigen to CD4⁺ or CD8⁺ effector/memory T cells (**Figure 7a,b**) under the experimental conditions tested here.

DISCUSSION

In order to develop valid and substantial alternative models to study respiratory pathologies, current knowledge on murine and human immune networks must be extended to other species. Here we show the potential of pig as a pertinent and intermediate model between mouse and human by ascribing six porcine respiratory DC/M0 cell types to their mouse and human counterparts, based on the recent unified nomenclature.¹²

Porcine AMs were CD163^{high}/CD11b-like^{neg} and expressed high levels of MerTK and CD64, as in mouse^{28,36} and in human.³⁷ Conversely to the other lung DCs/M0s, pig AMs expressed virtually no CCR2 or CX3CR1, in agreement with an origin that would be independent from blood monocytes, as recently demonstrated for mouse AMs.^{13,14} Interestingly, we observed an AM-like population, the CD163^{high} ‘interstitial’ AMs, unambiguously localized in the interstitium and representing >50% of the MHC-II^{high} parenchymal cells. It is noteworthy that this population has been observed in mouse lung,²⁷ without raising much interest. However, according to their frequency in pig parenchyma and to the essential role of AMs in the downmodulation of virus-induced inflammation in

mouse¹⁹ and in pig,³⁸ the presence of ‘interstitial’ AMs in human lungs and the respective roles of alveolar and ‘interstitial’ AMs deserve further investigations.

A third M0-like cell was described as MHC-II^{high} lung population: the CD163^{int} cells. According to their M0 features and their CCR2 and CX3CR1 expressions, they can be considered as moM0s. Interestingly, interstitial M0s, which are phenotypically distinct from AMs, have been described in mice. They express similar major histocompatibility complex II (MHC-II) levels as DCs and strongly downmodulate the allergic immune response.³⁹ The CD163^{int}/moM0s described here might be their pig counterpart (**Figure 8**). Again, to our knowledge, this population has never been reported in human.

The CD163^{low} cells presented full DC functionalities. Their frequency increases in the lung on IAV infection and they secrete the pro-inflammatory cytokines IL-1 β and IL-8/CXCL8 when stimulated. They also appeared potent in antigen-specific restimulation of the CD4 recall response. Thus, they were ascribed to the inflammatory moDC population. In mouse, lung inflammatory DCs¹⁵ have been defined as Fc ϵ RI α ^{pos}/CD64^{pos}. In pig, they expressed intermediate levels of CD64 compared with AMs and did not express Fc ϵ RI α conversely to CD163^{neg}/cDC2, which strongly expressed Fc ϵ RI α . In human, inflammatory DCs (from arthritic synovial fluid and ascites), cDC1, and cDC2 (from blood and lung) express Fc ϵ RI α .^{31,40,41} These data suggest that Fc ϵ RI α might not be the best *trans*-species marker of inflammatory moDCs.

Characterization of these inflammatory moDCs is very relevant, considering not only their pathological role in mice during an IAV infection^{26,42} but also their involvement in asthma.¹⁵ For example, during the last decade, CCR2 pharmacological antagonists have been developed, patented, and tested in different human inflammatory pathologies.⁴³ The definition of inflammatory moDCs in pig might allow to better describe, understand, and refine the different modes of action of these and other future drugs to be used for treatment in human.

Finally, two populations presented the hallmarks of cDCs; they expressed FLT3 along with strong migration and antigen presentation capacities. The CD172a^{neg} cDCs expressed XCR1, activated CD8⁺ better than CD4⁺ naive allogeneic T cells, and

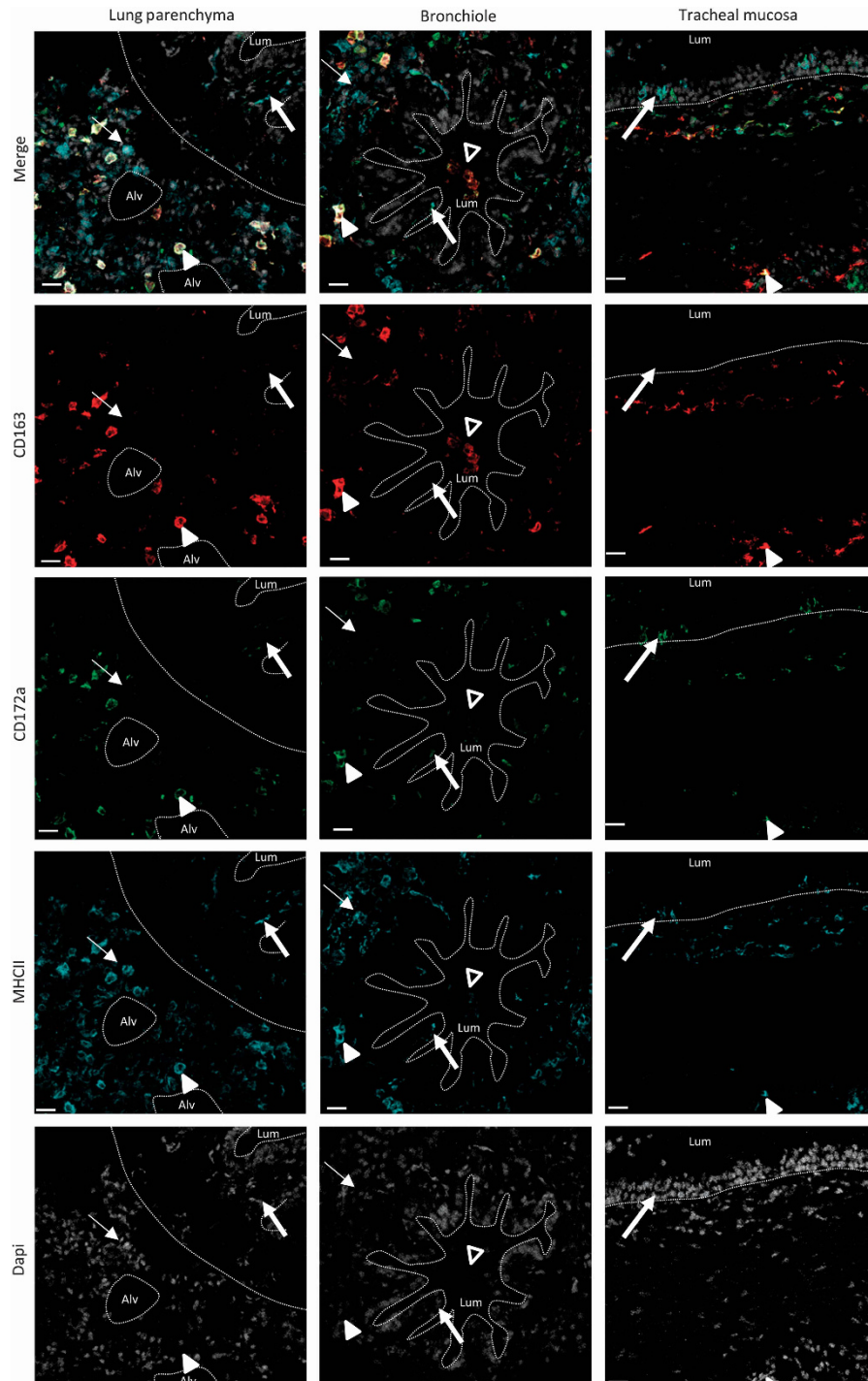


Figure 5 Dendritic cells (DCs) and macrophages (M0s) sublocalization. Four DC/M0 subsets were localized by confocal microscopy on parenchymal and tracheal mucosa cryosections fixed with acetone/methanol and stained for the indicated specificity. Images are representative of at least three pictures from different regions of biopsies from at least two different animals, independently stained. Objective used, $\times 40$ oil immersion. Scale bars, $20\mu\text{m}$. Thin arrow: $\text{CD172a}^{\text{neg}}$; large arrow: $\text{CD163}^{\text{neg}}$; filled arrowhead: $\text{CD163}^{\text{high}}$; empty arrowhead: AM.

induced a Th1 response. All in all, these properties clearly defined them as cDC1. However, they were unable to induce antigen-specific proliferation of effector/memory CD8^+ T cells. In humanized mice,¹⁸ both cDC subsets from the lungs or lymph nodes were able to reactivate memory CD8^+ T cells.

However, it has been shown in mouse⁴⁴ that although cDC1 from the lymph node and lung were able to activate naive CD8^+ T cells, only cDC1 from the lymph node could efficiently activate memory CD8^+ T cells. The authors hypothesized that the inability of lung cDC1 to activate memory CD8^+ T cells

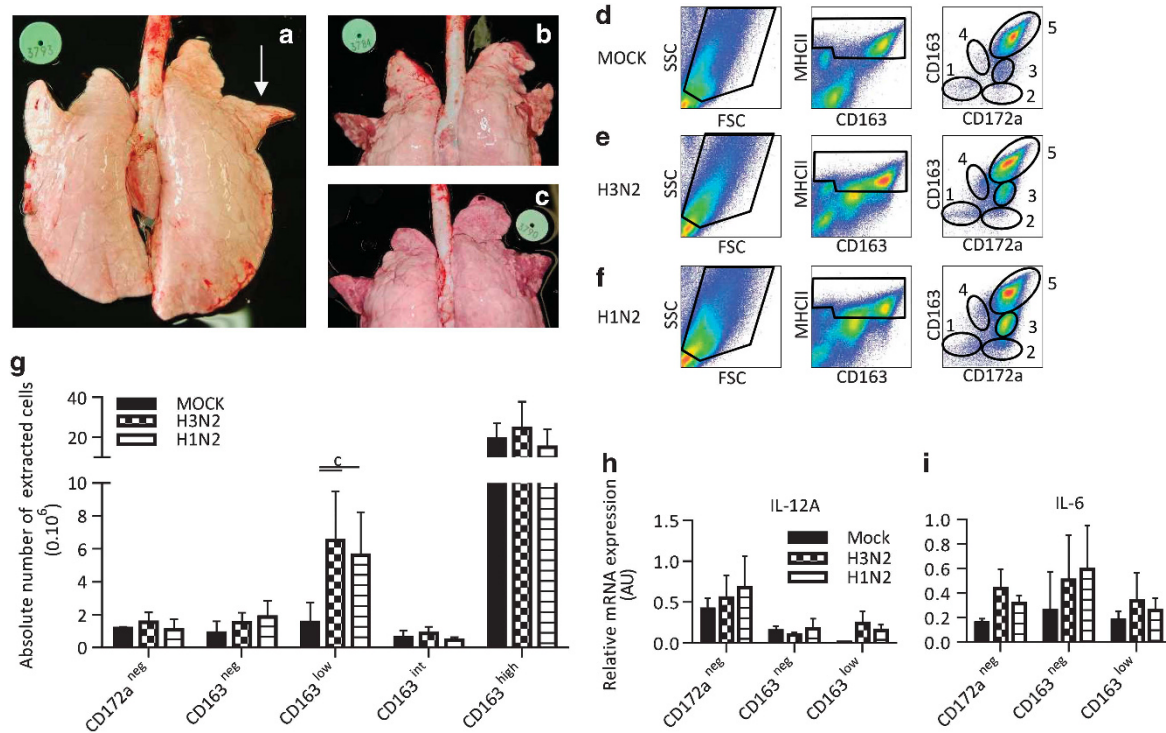


Figure 6 Influenza A virus (IAV) infection induces a recruitment of monocyte-derived dendritic cells (moDCs). Three groups of six pigs were inoculated intratracheally with swH3N2, swH1N2, or PBS and euthanized at 2 days post infection (dpi). Lungs from MOCK- (a), H3N2- (b), and H1N2-infected pigs (c) were sampled. Cells from the cardiac lobe (pointed by the white arrow) were extracted, stained, and analyzed by flow cytometry (d,e,f). Percentages of each population among total extracted cells were represented as means of six pigs for each group (g). c: $P < 0.05$ with a Student's *t*-test. Lung-extracted DCs were stained and sorted by flow cytometry. Total RNA was extracted and expressions of mRNA encoding interleukins (IL)-12A (h) and IL-6 (i) were assessed by quantitative real-time PCR. Data were normalized to the expression of the reference gene *RPS24* (ribosomal protein S24).

would allow a better priming of naive T cells, even in conditions in which robust memory predominates, allowing the emergence of CD8⁺ T cells presenting new antigen specificities. Although further *in-vivo* experiments are required, porcine lung cDC1 might be closer to murine lung cDC1 in their low capacity, to reactivate effector/memory CD8⁺ T cells than their human counterparts.

The second cDC subset, CD163^{neg}/CD172a^{pos}/XCRI^{neg}, activated CD8⁺ and CD4⁺ naive allogeneic T cells and induced a Th2 response. Although the association of CD8⁺ T-cell activation with a Th2 response seems contradictory, it is important to point out that our results were obtained in the context of an allogeneic reaction, which does not involve cross-priming of CD8⁺ T cells, and induces a strong helper response. CD163^{neg} cDC induced an antigen-specific proliferation of CD4⁺ effector/memory T cells and, in a less clear-cut way, of CD8⁺ T cells, as previously observed in mouse cDC2.⁴⁵ According to these properties and in agreement with mouse data,¹⁵ CD163^{neg} cells can be ascribed to the cDC2 lineage.

Pig lung cDC2 expressed strong levels of Langerin. We have previously described its expression in some skin cDC2.^{20,21} In addition, it has been recently shown in human that Langerin is expressed by some cDC2 from the dermis, lung, tonsil, and liver, and it is rapidly induced in blood cDC2 on tumor growth factor- β exposure.³⁰ Interestingly, similar to human cDC2 (ref. 18) porcine cDC2 are localized in close proximity with

epithelial cells, a potential source of tumor growth factor- β , whereas in mouse cDC1 have been described at this location.⁴⁶ In line with this, CD103 and CadM1, integrins involved in homophilic or heterophilic interactions with epithelial cells,^{47,48} are expressed by pig cDC1 and cDC2. CD103 is a marker of cDC1 in mouse peripheral tissues,^{46,49} whereas CadM1 is a marker of cDC1 in different tissues of mice, humans, pigs, sheep,^{21,32,50} and bats (Ginhoux and Dutertre, personal communication). In swine lung, these two markers are expressed on the two cDC subsets and on moDCs. In our previous work on porcine skin, we observed that CadM1 was expressed on dermal cDC1 and on intra-epithelial Langerhans cells.²⁰ We can thus hypothesize that in pig, and potentially in other species, CadM1 and CD103 may be expressed on cDC1 regardless of their location, but also on any DC in an epithelial environment.

It is important to highlight that the most striking similarities between swine and human are related to the cDC2 population: conversely to the mouse one, swine and human cDC2 are localized in or next to the tracheal and bronchial epithelia and express FcER1 α (Figure 8). These two properties, associated with the Th2-inducing capacity of cDC2, might imply similar allergen responses of these two species as opposed to mouse. Indeed, intraepithelial localization of cDC2 must facilitate their sampling of noninvasive antigens such as allergens, whereas FcER1 α expression might allow self-amplification⁵¹ or

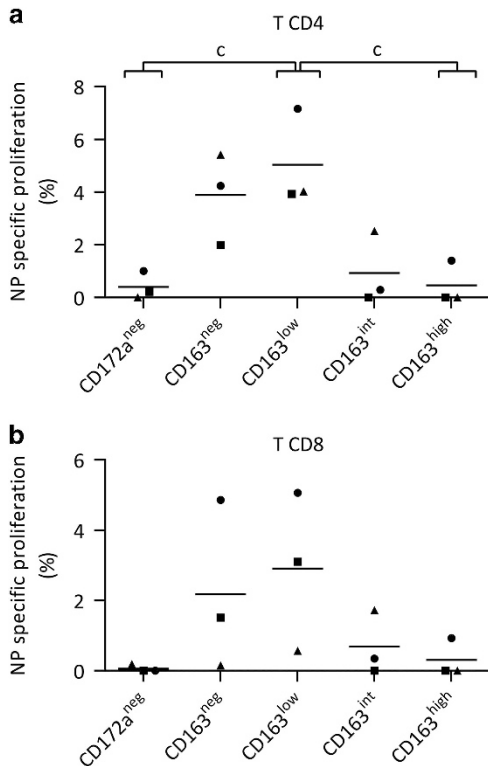


Figure 7 CD163^{neg} conventional dendritic cells (cDCs) and CD163^{low} monocyte-derived DCs (moDCs) induce antigen-specific proliferation of CD4⁺ memory T cells. Three pigs were intramuscularly vaccinated twice with inactivated H1N1 virus supplemented with a CD8-inducing adjuvant (ISA201-SEPPIC), and lung and spleen cells were collected at day 60. Lung DCs and macrophages (M θ s) were sorted and either incubated alone or charged with recombinant influenza nucleoprotein (NP), then plated with carboxyfluorescein succinimidyl ester (CFSE)-stained, enriched T cells, depleted of myeloid cells, B cells, and $\gamma\delta$ T cells, at a 1:20 ratio. CD4⁺ (CD3⁺CD4⁺) (**a**) or CD8⁺ (CD3⁺CD4⁻CD8 α ⁺) (**b**) proliferation was assessed by flow cytometry after 5 days. For each subset of each pig, the percentage of proliferation induced by unloaded DCs/M θ s was subtracted from the percentage of proliferation induced by NP-loaded DCs/M θ s, in order to represent NP-specific proliferation only. c: $P < 0.05$ with a Student's *t*-test.

autoregulation⁵² of the pathological inflammation, differently from the murine cDC2. Thus, pig respiratory system might be considered a better model than mouse for human allergic syndromes.

On the contrary, during pathologies targeting epithelial cells such as IAV infection, the deep localization of cDC1 in porcine and in human¹⁸ pulmonary tissues might undermine their access to antigens and thus lessen their role in the induction of the immune response compared with that in mouse. Different cDC1 and cDC2 localizations between mice and humans might explain why human cDC2 are shown to be more involved in the induction of anti-IAV CD8 response¹⁸ than their mouse counterparts.⁵³ It would be of great interest to test this hypothesis by monitoring the role of cDC1 and cDC2 in the initiation of anti-IAV CD8 immune response in pig.

Indeed, as stated before, the porcine respiratory and immune systems are similar to the human one at many levels. In addition,

unlike mice, pigs are IAV natural hosts that are able to be infected with the same strains as human. We showed here that it was possible to use pig as a pertinent model for respiratory immune parameters and to study each population of DC/M θ as precisely as in mouse. It is noteworthy to emphasize the importance of working on different complementary species such as mouse, nonhuman primate and nonconventional models such as pig, ferret, or horse, in order to draw more robust conclusions, which allow an easier translation to human clinic.

METHODS

Animals, *in vivo* infections, and tissue collection. For phenotypic and functional assays, tissue samples were obtained from 5- to 7-month-old Large White conventionally bred sows from UEPAO, Tours, France.

IAV infection experiments were conducted at CReSA (Barcelona, Spain), in compliance with the Ethical Committee for Animal Experimentation of the institution (Universitat Autònoma de Barcelona). The treatment, housing, and husbandry conditions conformed to the European Union Guidelines (Directive 2010/63/EU on the protection of animals used for scientific purposes). Animal care and procedures were in accordance with the guidelines of the Good Laboratory Practices under the supervision of the Ethical and Animal Welfare Committee of the Universitat Autònoma de Barcelona (number 1189) and under the supervision of the Ethical and Animal Welfare Committee of the Government of Catalonia (number 5796). Eighteen pigs (7–8 weeks old, LandraceXPietrain) were housed in separate isolation rooms and were randomly assigned to three experimental groups of six pigs. The animals were seronegative to IAV (ID Screen Influenza A Antibody Competition ELISA, ID-Vet, Grabels, France) at the time of the experiment. On day 0, one group of pigs was intratracheally inoculated with $3 \times 10^{6.5}$ TCID₅₀ of a swH3N2 virus (A/Swine/Spain/SF32071/2007) in 2 ml inoculum produced on Madin–Darby canine kidney (MDCK) cells. Another group was similarly inoculated with $3 \times 10^{7.2}$ TCID₅₀ per 2 ml of a swH1N2 virus (A/Swine/Spain/SF12091/2007). The third group was mock infected, receiving 2 ml of PBS. Each of the three groups was euthanized at 2 days post infection and cardiac lobes were collected.

For the antigen-specific presentation assay, experiments were conducted at PFIE (Tours, France), in accordance with the animal welfare experimentation recommendations under the responsibility of I Schwartz (authorization 00783.02). The animal experiment protocol was approved by the French national ethics committee CEEA Val de Loire. At days 1 and 30, three 2- to 4-month-old Large White pigs were injected intramuscularly with 2 ml of H1N1 vaccine composed of PBS, $18.5 \mu\text{g ml}^{-1}$ inactivated A/Swine/Paris/2590/2009 and ISA201VG adjuvant (SEPPIC, Paris, France). Pigs were euthanized at day 60. The lung and spleen were collected.

Lung and spleen cell isolation. A BAL procedure was performed twice on the isolated left lung with 250 ml of PBS + 2 mM EDTA (PBS/EDTA), to collect AMs. Next, a 1-cm slice of external lung parenchyma was dissected from the same lung. For *in vivo*-infected tissues, the whole cardiac lobe was dissected without performing any BAL, resulting in the extraction of 60–170 million live cells with no significant differences between conditions. Tracheal mucosa was separated from the cartilage with pliers. Tissues were minced and incubated in nonculture-treated Petri dishes, to avoid differential plastic adherence of M θ s and DCs, for 2 h at 37 °C in complete RPMI, consisting of RPMI 1640 supplemented with 100 IU ml⁻¹ penicillin, 100 mg ml⁻¹ streptomycin, 2 mM L-glutamine, and 10% inactivated fetal calf serum (FCS) (all from Invitrogen, Paisley, UK), containing 2 mg ml⁻¹ collagenase D (Roche, Meylan, France), 1 mg ml⁻¹ dispase (Invitrogen), and 0.1 mg ml⁻¹ Dnase I (Roche). Cells were passed through 40 μm cell strainers and red blood cells lysed with erythrocytes

	Mouse	Pig	Human
DC	Migrate toward CCL21		
cDC	FLT3 ^{pos}		
cDC1	Activate allogeneic naïve T cells and induce a Th1 response		
	XCR1 ^{pos} , CD172a ^{neg}		
	No effector/memory CD8 restimulation	Memory CD8 restimulation	
	Induce mucosal effector CD8 differentiation Proximity with epithelial cells	Proximity with alveoli	
cDC2	Activate allogeneic naïve T cells and induce a Th2 response		
	CSF1R ^{pos} /CCR2 ^{pos} /CX3CR1 ^{pos}		
	Proximity with epithelial cells		
	Langerin ^{pos} /FcεR1α ^{pos}		
	CadM1 ^{pos} /CD103 ^{pos}		
	Induce central memory CD8 differentiation	Induce mucosal effector CD8 differentiation	
Monocyte-derived Cells	Induce antigen-specific proliferation of effector/memory CD4 ⁺ T cells		
	CCR2 ^{pos} /CX3CR1 ^{pos} /CSF1R ^{pos}		
Inflammatory moDC	CD64 ^{pos} /FcεR1α ^{pos}	CD64 ^{int} /FcεR1α ^{neg}	Population Not described
	Recruited upon influenza infection		
	Induce pathologic inflammation upon influenza infection		
	Induce antigen-specific proliferation of effector/memory CD4 ⁺ T cells		
moMacro	MHCII ^{high} /MerTK ^{pos} /CD64 ^{pos}		Population Not described
Interstitial AMs	Autofluorescent/CD11b-like ^{neg}		
	CD163 ^{high} /MerTK ^{high} CCR2 ^{neg} /CX3CR1 ^{neg} /CD64 ^{pos}		
AMs	Autofluorescent/CD11b-like ^{neg} /CD163 ^{high} /CD64 ^{pos} /MerTK ^{high} /CCR2 ^{neg} /CX3CR1 ^{neg}		

(Bibliographic references can be found in the main text)

Figure 8 Comparison of mouse, pig and human lung DCs/Mφs. DCs, dendritic cells; Mφs, macrophages.

lysis buffer (10 mM NaHCO₃, 155 mM NH₄Cl, and 10 mM EDTA). Next, cells were washed with PBS–EDTA, counted, and step-frozen in FCS + 10% dimethyl sulfoxide (Sigma-Aldrich, St Louis, MO).

Spleen cells were extracted by scraping with a scalpel blade and collected in PBS 1.3 mM citrate, then filtered on a 100-μm strainer in complete RPMI, and enriched on Ficoll-Hypaque density gradient (Amersham Biosciences, Uppsala, Sweden). Red blood cells were lysed as described above.

Ex-vivo CFSE staining assay. In order to stain the alveolar cells *ex vivo*, the intermediate lobe was sampled and injected with 20 ml of 50 μM CFSE (Invitrogen) via the connecting bronchiole, while being rubbed for a better diffusion in the alveoli. Next, it was clamped and after 30 min incubation at 37 °C, alveolar cells were collected by one lavage with 10 ml FCS and two lavages with 15 ml PBS–EDTA. Two additional lavages were performed to eliminate remaining alveolar cells. The lobe was then minced and enzymatically digested as described above, to retrieve interstitial lung cells.

Flow cytometry analysis. Cell surface stainings were performed in PBS–EDTA supplemented with 5% horse serum and 5% swine serum for 30 min on ice. Primary antibodies and their working dilutions are listed in **Table 1**. The anti-CD11b antibody used (M1/70) recognizes murine and human CD11b but had never been described in pig. It must bind to one out of the three potential orthologs of CD11c and CD11b in pig, namely CD11R1, CD11R2, or CD11R3.⁵⁴ Thus, for clarity we referred to the antigen recognized by this antibody as ‘CD11b-like’. The anti-MR also being an anti-human antibody, it will be referred to as ‘MR-like’.

Matched isotype controls for mouse IgG1, IgG2b, and IgG2a were purchased from Invitrogen and were used at the same concentration as the corresponding antibody. Secondary antibodies, anti-mouse IgG1, IgG2b, IgG2a coupled to Alexa-488, Phycoerythrin-PE, Tricolor, or Alexa-647 were from Invitrogen and were used at a 1/200 dilution. Between labelings, cells were washed twice with PBS–EDTA. Cells from IAV-infected pigs were then fixed in 4% paraformaldehyde (EMS, Hatfield, PA) before flow cytometry analysis. Samples were acquired on

Table 1 Antibodies used for flow cytometry, microscopy, and CBA assays (working dilutions are specified for flow cytometry assays. A 1/40 dilution was used for each antibody for depletions)

Antibody	Clone	Isotype	Species	Working dilution	Supplier
Anti-CD1	76-7-4	IgG2a	Mouse	1/250	Monoclonal
Anti-CD172a	74-22-15, 74-22-15a	IgG1, IgG2b	Mouse	1/500	Antibody
Anti-MHCII	Th21a, MSA3	IgG2b, IgG2a	Mouse	1/500	Center
Anti-CD4	PT90A, 74-12-4	IgG2a, IgG2b	Mouse	1/400	Washington
Anti-CD3	8E6	IgG1	Mouse	1/100	State
Anti- $\gamma\delta$	86D, PGBL22A	IgG1, IgG1	Mouse		University
Anti-CD8a	76-2-11	IgG2a	Mouse	1/50	(Pullmann,
Anti-CD8b	PG164A	IgG2a	Mouse		WA)
Anti-IgM	PG145A	IgM	Mouse		
Anti-humCD14	TUK4	IgG2a	Mouse	1/100	
Anti-CD25	K231.3B2	IgG1	Mouse		
Anti-CD16	G7	IgG1	Mouse	1/100	AbDSerotec
Anti-IgL	K139-3E1	IgG2a	Mouse		(Oxford, UK)
Anti-CD163 unlabeled or PE-conjugated	2A10/11	IgG1	Mouse	1/400 1/20	
Anti-humMR-like	122D2.08	IgG1	Mouse	1/100	Dendritics (Lyon, France)
Anti-humCadM1	3E1	IgY	Chicken	1/250	MBL (Woburn, MA)
Anti-humCD11b (CD11b-like)	M1/70	IgG2b	Rat	1/100	BD Pharmingen
Anti-humCD21 (CD21-like)	B-ly4	IgG1	Mouse		(Le Pont de Claix, France)
Cytokines	Recombinant cytokines	Biotinylated antibody			
IL1- β	R&D Systems 681-PI	R&D Systems BAF6811			
CXCL8 /IL-8	R&D Systems 535-IN	R&D Systems BAF535			

Abbreviations: CBA, cytometric beads assay; IL1- β , interleukin-1 β ; MR, mannose receptor.

a FACS-Calibur (BD-Biosciences, Oxford, UK) or sorted on a MoFlo ASTRIOS (Beckman-Coulter, Paris, France). For sorting, dead cells were excluded by Dapi staining (Sigma-Aldrich). Acquired data were analyzed using FlowJo software (version X.0.6; Tree Star, Ashland, OR).

Microscopy. Lung parenchyma biopsies and tracheal mucosa pieces were snap-frozen in OCT (Sakura, Paris, France) and conserved at -80°C . Cryosections ($7\mu\text{m}$) were obtained using a cryostat (Leica CM3050S, Nanterre, France). Sections were fixed in methanol/acetone (1:1) at -20°C for 20 min and stained using previously described anti-swine antibodies, matched isotype controls, and isotype-specific secondary antibodies. Sections were mounted in SlowFade mounting medium (Invitrogen). Slides were examined on a LSM510 confocal microscope (Zeiss, LePecq, France), using a $40\times$, oil-immersion objective (MIMA2 Platform, INRA, Jouy en Josas, France), and analyzed with Zen 2012 Software (Zeiss, Jena, Germany).

May-Grünwald-Giemsa staining and histology. Sorted cells were deposited on microscope slides (Superfrost, Thermo, Villebon sur Yvette, France) by cytocentrifugation and stained with May-Grünwald-Giemsa stain. Images were acquired with a Leica Leitz DMRB microscope equipped with a $\times 63$ oil-immersion objective (numerical aperture 1.3) and a DP50 imaging camera coupled to the CellF software (Olympus, Tokyo, Japan).

DC enrichment and stimulation. Cells were cultured in complete RPMI. Preparations were enriched in DC by gradient²⁰ (Optiprep;

Nycomed Pharma, Oslo, Norway). Enriched preparations routinely contained 5–25% of DC/M θ as checked by flow cytometry. For transwell and cytokine production assays, enriched DCs were matured by a 24-h *in vitro* culture in complete RPMI supplemented with $10\mu\text{g ml}^{-1}$ lipopolysaccharide and polyinosinic:polycytidylic acid (Sigma-Aldrich).

Transwell assay. A Chinese Hamster Ovary (CHO) clone expressing porcine CCL21-GFP has been derived as previously described.²¹ Four hundred microliters of supernatants from this clone or from the parental CHO cells were deposited in the lower wells of a 24-well plate. DCs or AMs were enriched and stimulated as described above and re-suspended in complete RPMI at 1×10^6 cells per ml. Next, 100 μl were deposited on the inserts (Costar, 5 μm pore filter, Corning, NY). After 2 h at 37°C , cells having migrated in the lower compartment were stained and counted by flow cytometry.

Mixed lymphocytes reaction assay. Mixed lymphocytes reaction assays were performed as previously described.²¹ Briefly, peripheral blood mononuclear cells from large, white, specific pathogen-free pigs (Anses, Ploufragan, France) were CFSE stained and re-suspended in X-vivo medium (Ozyme, Saint-Quentin-en-Yvelines, France) supplemented with 2% FCS and penicillin/streptomycin at 3×10^6 cells per ml (or 1×10^6 cells per ml for enriched naive CD4⁺ T cells) and 100 μl per well were plated in a 96-well U-bottom plate. The sorted DCs were mixed with the allogeneic CFSE-labeled T cells at a 1:8 ratio. Cells were cultured for 5 days in X-vivo medium + 2% FCS + penicillin/streptomycin before staining with anti-CD3, anti-CD4, and

Table 2 Primers used for qPCR

Primers	Sequences (5'-3')	Gene ID	Size	Efficiency
RPS24	F: AAGGAACGCAAGAACAGAATGAA R: TTTGCCAGCACCAACGTTG	100155012	62 bp	0.98
FLT3	F: TGTTACGCTGAATATAAGAAGGAA R: GGAGCAGGAAGCCTGACTTG	100515445	70 bp	0.92
XCR1	F: CGATGCCGTCTTCCACAAG R: GGAACCACTGGCGTTCTGA	414375	61 bp	1.01
CSF1R	F: TGAACGACTCCAACACTACATTGTCA R: TGTAGACGCAGTCGAAGATGCT	100517086	86 bp	0.86
CD64 (FCGR1A)	F: TCCAACCTGGTGGGCAAATG R: GGAGGCTGCAAGGTGATCAC	613130	53 bp	0.98
FCeR1 α	F: AATTTACAGACCCACAGCCTAGCT R: TGCTATCGCAGATGTTTCTTGAG	100152827	74 bp	0.76
CCR2	F: ACACGCTTTCCCGGTTCA R: CCCTTGATATTCATTGTAAGCAGAGA	414372	69 bp	0.88
CX3CR1	F: CGTGGCCCTGGGAACTG R: CGAGGCCAAAGGCAAAA	100622126	60 bp	1.08
CD103 (ITGAE)	F: GATGCGGAACATCTATGAGAAGTG R: CTGGATGACGCTCCCGTATT	100519919	69 bp	1.09
Langerin	F: GGTCTGGTTCGCTCCAT R: GACATCTGACATTGTGCCATAA	100037935	69 bp	0.99
IFN- γ	F: TGGTAGCTCTGGGAACTGAATG R: TGGCTTTGCGCTGGATCT	396991	79 bp	0.97
IL-13	F: CTGACCACCAGCATGCAGTACT R: CGCTGGCAGTCGGAGATGTT	396721	59 bp	0.93
IL-17	F: CCAGAOCGGCCCTCAGATTAC R: GGTCTCGTTGCGTTGGA	449530	65 bp	0.86

Abbreviations: IFN- γ , interferon- γ ; IL, interleukin; qPCR, quantitative PCR.

anti-CD8 α antibodies, and flow cytometry analysis. For each of the four distinct experiments, the cell subtype inducing the highest percentage of T-cell proliferation was considered as 100 and the percentage of T-cell proliferation induced by the other cell subtypes of the same experiment were normalized to it.

For naive CD4⁺ T-cell functional polarization, allogeneic naive CD4⁺ T cells were enriched by depletion of myeloid cells, B cells, natural killer cells, $\gamma\delta$ T, CD8⁺ T cells, and memory CD4⁺ T cells. Fresh peripheral blood mononuclear cells from Melanoma Libechov Minipig pigs⁵⁵ were stained with anti-CD172a, anti-CD21-like, anti-IgM, anti-IgL, anti-CD16, anti-CD8 α , anti-MHC-II, and anti- $\gamma\delta$ T cell receptor, all at a 1/40 dilution. They were then incubated with goat anti-mouse IgG microbeads (Miltenyi-Biotech, Bergisch Gladbach, Germany) at a 1/5 dilution, washed, and passed on an LD column (Miltenyi-Biotech) for depletion. Naive T cells represented >70% of the collected cells. They were then mixed with the sorted DCs/M θ s at a 1:10 ratio. Seventy-two hours later, cells were lysed and RNA extracted. Reverse transcriptase-quantitative PCR were then performed as stated below using interferon- γ -, IL-13-, and IL-17-specific primers.

Cytokine detection. Sorted DCs were re-suspended in complete RPMI at 1×10^6 cells per ml and plated in 200 μ l per well of a 96-well U-bottom plate for one night at 37 °C with 10 μ g ml⁻¹ lipopolysaccharide and polyinosinic:polycytidylic acid. The supernatants were frozen at -20 °C until cytokine detection. Concentrations of IL-1 β and IL-8 were assessed by cytometric beads assay for simultaneous detection of 10 swine cytokines. Antibodies and recombinant cytokines

used are listed in **Table 1**. Capture antibodies were covalently coupled to the surface of fluorescent Cyto-Plex carboxylated Microspheres (Thermo Fisher Scientific, Courtabouef, France) according to the manufacturer's instructions. Each capture antibody was applied to a given microsphere category. All assays were made in duplicates. Fifty microliters of samples or standards, diluted in diluent buffer (PBS 0.5 \times and Tris buffer 0.5 \times), supplemented with 2.5% bovine serum albumin, 0.5% polyvinylalcohol (Sigma-Aldrich), and 0.8% polyvinylpyrrolidone (Sigma-Aldrich) were incubated in membrane filter-bottomed microplates (MultiScreen HTS BV 1.2 μ m Opaque Non-sterile, Millipore, Darmstadt, Germany) with 2,000 beads per well of each antibody-coated beads. Beads were washed with washing buffer (PBS 10 \times (Thermo Fisher Scientific), Tris 0.5 M, pH 7.2, 1:1) by aspiration. Twenty-five microliters of biotinylated detection antibodies (1 μ g ml⁻¹ diluted in assay buffer) were added and incubated for 90 min. Fifty microliters of PE-labelled streptavidine (5 μ g ml⁻¹ diluted in assay buffer, Molecular Probes) were added in each well after three washes, followed by 30 min incubation. Beads were then washed and re-suspended in PBS before acquisition on a Guava easyCyte 6HT-2L cytometer (Millipore). The results were analyzed using FloCytomix Pro software (eBioscience, Paris, France). A five-parameter logistic regression model was used to fit the curve.

Antigen-specific presentation assay. For antigen-specific presentation assay, spleen cells were collected from pigs vaccinated as described above. Dead cells were eliminated on an Optiprep gradient. Cells were then enriched in T lymphocytes by depletion of myeloid

cells, B cells, and $\gamma\delta$ T cells. They were stained with anti-CD172a, anti-CD21-like, anti-IgM, anti-IgG, and anti- $\gamma\delta$ TCR, all at a 1/40 dilution, and then with goat anti-mouse IgG microbeads at a 1/5 dilution. Cells were subsequently washed and passed on an LS column (Miltenyi-Biotec) for depletion and they were CFSE stained. The sorted lung DCs/M Φ s were re-suspended in X-vivo medium + 2% FCS + penicillin/streptomycin, and 20,000 cells were plated in 200 μ l per well of a 96-well U-bottom plate. For each subset of each pig, one well was then incubated at 37 °C for 90 min with medium alone and another well with 2 μ g ml⁻¹ recombinant IAV NP, produced as previously described.⁵⁶ After centrifugation, the NP-containing medium was discarded and 400,000 CFSE-stained T cells from the same pig were added to each well. Cells were cultured for 5 days at 37 °C before staining with anti-CD3, anti-CD4, and anti-CD8 α antibodies, and flow cytometry analysis. In order to represent the NP-specific proliferation only, the percentage of proliferation induced by unstimulated DCs/M Φ s was subtracted from the percentage of proliferation induced by NP-loaded DCs/M Φ s.

RNA extraction. Total RNA from sorted DCs/M Φ s or cultured allogeneic naive T cells were extracted using the Arcturus PicoPure RNA Isolation kit (ThermoFisher Scientific, St Aubin, France) according to the manufacturer's instructions. Contaminating genomic DNA was removed using a Qiagen RNase free DNase set (Courtaboeuf, France).

Real-time qPCR. RNA was reverse transcribed using random hexamers and the Multiscribe reverse transcriptase (ThermoFisher Scientific). All cDNA were examined for the frequency of different transcripts using qPCR. All qPCR reactions were performed in 25 μ l volumes using iTaq Universal SYBR Green Supermix (Biorad, Hercules, CA). Primers are detailed in Table 2. Relative quantification was determined using the Δ Ct method and normalized to expression of the reference gene *RPS24* (ribosomal protein S24). This gene has been chosen as the reference gene because of its highly stable expression in the different DC and M Φ lung subsets, and upon IAV infection (data not shown), as compared with other genes previously shown as stable in pig lungs: *GAPDH*, *RPL19*, and *HPRT*.⁵⁷

Statistical analysis. All data were analyzed using the GraphPad Prism v5.0 statistical software package (GraphPad Software, La Jolla, CA). Statistical tests applied to each data set are indicated in the relevant figure legend.

SUPPLEMENTARY MATERIAL is linked to the version of the paper at <http://www.nature.com/mi>

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DISCLOSURE

The authors declare no conflict of interest.

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