

Damage to reproductive structures of broadleaf woody plants. Chapter 7

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Damage to reproductive structures of broadleaf woody plants

A. Roques, V. Talgø, J.-T. Fan and M.-A. Auger-Rozenberg

7.1. Flower (blossom, catkin, flower-head) galling

Description: Flower (catkin) distorted, swollen, or with tissue outgrowth(s) of any shape.

Possible damaging agents: Insects: Diptera (Cecidomyiidae midges: Figs. 7.1.5, 7.1.6), Hymenoptera (Cynipidae: Figs. 7.1.3., 7.1.4.), **Mites** (Acari, Eriophyiidae: Figs. 7.1.1., 7.1.2., 7.1.6.), **Fungi** (Ascomycetes, Taphrinales: Figs. 7.1.7., 7.1.8.), **Bacteria, Phytoplasma**.



Fig. 7.1.1. Newly-developed inflorescence **Fig. 7.1.2.** Cauliflower-like gall finally of ash (*Fraxinus excelsior*), galled by a mite resulting from mite damage shown in Fig. (Acari, Eriophyiidae: *Aceria fraxinivora*). 7.1.1. Hungary, GC. Marcillac, France, AR.



Fig. 7.1.3. Berry-like gall on a male catkin Fig. 7.1.4. Male catkin of Quercus of oak (Ouercus sp.) caused by a gall wasp myrtifoliae, deformed by a gall wasp (Hymenoptera, Cynipidae: quercusbaccarum). Hungary, GC.



Fig. 7.1.5. Inflorescence of birch (Betula sp.) Fig. 7.1.6. Symmetrically swollen catkin of deformed by a gall midge Cecidomyiidae: Semudobia betulae). Hungary, GC.



Neuroterus (Hymenoptera, Cynipidae: Callirhytis myrtifoliae). Florida, USA, GC.



(Diptera, hazelnut (Corylus sp.) caused by a gall midge (Diptera, Cecidomyiidae: Contarinia coryli) or a gall mite (Acari Eriophyiidae: Phyllocoptes corvli). The damaging agent can only be ascertained by catkin dissection. Germany, HJB.



Fig. 7.1.7. Catkin of poplar (Populus sp.) Fig. 7.1.8. Tongue-like gall on female deformed by a fungus (Ascomycota, catkin of alder (Alnus glutinosa) induced by Taphrinales: Taphrina johansonii). Hungary, a GC.



fungus (Ascomycota, Taphrinales: Taphrina alni). Germany, HJB.

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<u>Additional information</u>: Dissect the gall to check for the presence of insect larva(e) or minute mites. If insects and mites are absent without visible exit holes, check for fungi, bacteria or phytoplasmas. For insect sampling and preservation, see Chapter 3 and for fungal preservation, see Chapter 4.

7.2. Bacterial blight

Description: Tissue infected by bacteria will have a dark (blackish) appearance. Under humid conditions, droplets of bacterial exudates may occur. **Possible damaging agents: Bacteria** (Enterobacteriaceae: *Erwinia*).





Fig. 7.2.1. Flowers and fruits of *Cotoneaster* **Fig. 7.2.2.** Young fruits of *Malus* sp., *bullatus*, with fire blight due to a bacterium infected by fire blight due to a bacterium (Bacteria, Enterobacteriaceae: *Erwinia* (Bacteria, Enterobacteriaceae: *Erwinia amylovora*). Hordaland county, Norway, AS. *amylovora*). Slovakia, JM.

<u>Additional information</u>: Bacteria attacking reproductive structures are often transferred by pollinating insects and may lead to blossom blight, fruit rot and proceed to shoots.

7.3. Internal feeding by larva

Description: Flower remaining closed, with larval tunnels visible when the flower is opened.

Possible damaging agents: Insects: larvae of Coleoptera (Curculionidae (weevils): Figs. 7.3.1, 7.3.2, 7.3.8), Diptera (Anthomyiidae (maggots): Fig. 7.3.3, Cecidomyiidae (midges): Fig. 7.3.6), Hymenoptera (Tenthredinidae (sawflies): Fig. 7.3.4), Lepidoptera (Geometridae: Figs. 7.3.5, 7.3.6., Noctuidae, Tortricidae, Yponomeutidae).



Fig. 7.3.1. Salix catkin with brown frass on Fig. 7.3.2. The Salix catkin shown in 7.3.1 the top due to damage by a weevil larva sliced open to show the weevil larva (Coleoptera, Curculionidae: Dorvtomus taeniatus). Cesana Torinese, Italy, AR.



tunnelling the axis (Coleoptera, Curculionidae: Dorytomus taeniatus). Cesana Torinese, Italy, AR.



Fig. 7.3.3. Salix catkin damaged by a Fig. 7.3.4. Dried tip of a Salix phylicifolia maggot (Diptera, Anthomyiidae: Egle sp.), catkin, indicating the presence of a catkinshowing woolly fluff expelled by the larva mining (top) and the emerging maggot larva Tenthredinidae: Pontopristia sp.) inside the (bottom). Marcillac, France, AR.



sawflv larva (Hymenoptera, catkin stem. Oulu, Finland, TN.



regia) infested internally by moth larvae regia) with emerging unidentified moth (Lepidoptera, Geometridae). Marcillac, France, AR.



to show damage by a midge larva (Diptera, closed following infestation by a weevil Cecidomyiidae: Dasineura oxyacanthae). Copenhagen, Denmark, SH.



Fig. 7.3.5. Male catkin of walnut (Juglans Fig. 7.3.6. Male catkin of walnut (Juglans larvae (Lepidoptera, Geometridae). Marcillac, France, AR.



Fig. 7.3.7. Flower of Crataegus sp. opened Fig. 7.3.8. Flower of Malus sp., remaining larva (Coleoptera, Curculionidae: Anthonomus pomorum), and manually opened to show the larva. Slovakia, JM.

Additional information: Dissect the flower to ascertain the presence of larva(e) or pupa(e). Note larva colour, if it is legless or not, and if it has a visible head or not. For insect preservation, see Chapter 3.

7.4. External feeding

Description: Flower gnawed or with cutout(s) (Figs. 7.4.1, 7.4.2, 7.4.3); fruit fed from exterior (Fig. 7.4.4.).

Possible damaging agents: Insects: Adults of Coleoptera (e.g., Cantharididae, Cetoniidae, Rutelidae, Scarabaeidae: Figs. 7.4.1, 7.4.2, 7.4.4) and Hymenoptera (Tenthredinidae), larvae of Lepidoptera (Fig. 7.4.3).



Fig. 7.4.1. Flower of quince (Cydonia Fig. 7.4.2. Flower of wild rose (Rosa sp.) oblonga) cut out by unidentified scarab with adult scarab beetles feeding on pollen beetles (Coleoptera, Rutelidae). Marcillac, (Coleoptera, France, AR.

Rutelidae: Hoplia sp.). Briançon, France, AR.



Fig. 7.4.3. Flower of quince (Cydonia Fig. 7.4.4. Fruit eaten by adult beetles oblonga) damaged by an early-instar moth (Coleoptera, Cetoniidae: Cetonia sp.). larva (Lepidoptera, Lasiocampidae: Slovakia, MZ. Malacosoma neusstria). Marcillac, France, AR.



Additional information: Observation by chance or by beating flowering branch over a Japanese umbrella. For insect collection and preservation, see Chapter 3.

7.5. Sucking arthropod damage

Description: Flower completely dried, often shed.

Possible damaging agents: Insects: Adults of Thysanoptera (Thrips: Fig. 7.5.1), nymphs and adults of Hemiptera (Aphididae: Fig. 7.5.2).





Fig. 7.5.1. Catkin of Betula sp. covered by a Fig. 7.5.2. Flower bud of wild rose (Rosa colony of bug nymphs (Hemiptera, sp.) covered by an aphid colony Aphididae: Acanthosomatidae: Elasmucha grisea). (Hemiptera, Macrosiphum Hungary, GC. rosae). Slovakia, MZ.

<u>Additional information</u>: Dissect the flower to look at the presence of minute insects (thrips); check the presence of aphid or bug colonies on the external part of the flower or on the pedicel.

7.6. Abiotic damage

Description: Flower completely dried, often shed; seed empty, without frass and without the remains of embryo.

Possible damaging agents: Late frosts, drought, genetic incompatibility.





Fig. 7.6.1. Flowers of Sorbus sp. killed by Fig. 7.6.2. Apple flower (Malus sp.) with frost and then colonized by aphids. Rodez, dark brown centre, indicating that it was France, AR.

killed by frost. Michigan, USA, ML.

Additional information: Dissect flowers to ascertain the absence of pests as well as of pest damage (e.g., tunnels and frass).

7.7. Fungal growth on catkins and fruits

Description: Catkins and fruits discoloured, with irregular chlorotic to necrotic spots with blighted margins or larger necrotic (yellowish) blotches. Possible damaging agents: Fungi: Ascomycota (Figs. 7.7.1 – 7.7.4).



Colletotrichum 7.7.1. Fig. (Ascomycota, Glomerellaceae) Rhododendron sp. Norway, VT.



Fig. 7.7.3. Berries of Mountain ash (Sorbus Fig. 7.7.4. Detail of a holly fruit (Ilex aucuparia) infested by Colletotrichum aquifolium) acutatum (Ascomycota, Glomerellaceae). acuminatum-like fungi. Rogaland county, Buskerud county, Norway, VT.



acutatum Fig. 7.7.2. Necrosis on a Salix catkin on caused by an unidentified fungus. Briançon, France, AR.



infested by Fusarium Norway, VT.

Additional information: Anthracnose causes dark, slightly sunken lesions on flowers and fruits. Under humid conditions, the disease may develop and spread quickly.

7.8. Petal Blight

Description: Flower discoloured with tan, water-soaked spots or soft blighted tissue; petals stuck together and often shed prematurely. **Possible damaging agents: Fungi** (Figs. 7.8.1, 7.8.2).



Fig. 7.8.1. Flowers of *Prunus* sp. infected **Fig. 7.8.2.** Flowers of *Rhododendron* sp., during bloom by brown rot fungus infected by a petal blight fungus (Ascomycota, Helotiales: *Monilinia laxa*) to (Ascomycota, Helotiales: *Ovulinia* be compared to normally developed fruits on *azaleae*). Maryland, USA, UME. the same branch. Slovakia, JM.

<u>Additional information</u>: Petal blight is a commonly occurring problem on *Rhododendron* spp. and other woody plants and may destroy their economic and aesthetic value.

7.9. Damage on flowers by stone fruit scab

Description: Flower discoloured with black/blight necrosis, flecking, streaks on catkins (flowers) and/or with lesions. Stone fruit scab is caused by fungal infection (*Cladosporium carpophilum*), so there cannot be bacterial slime. **Possible damaging agents: Fungi** (Ascomycota, Pleosporales). See photos of final damage in 7.20.

7.10. External damage by sap feeders

Description: Fruit surface (partially) covered with insects, insect secretions or fungal structures, waxy or woolly covers and/or honeydew on surface.

Possible damaging agents: Insects: adults and nymphs of Hemiptera, especially scales (Coccoidae, Diaspididae: Figs. 7.10.1, 7.10.3), aphids (Aphididae: Fig. 7.10.2) and psyllids (Psyllidae: Fig. 7.10.4)





Fig. 7.10.1. Fruits of apple tree (Malus sp.) Fig. 7.10.2. Acorn of *Quercus ilex* with the infested by the San Jose scale (Hemiptera, basis covered by aphids (Hemiptera, Diaspididae: Diaspidiotus Slovakia, JM.

perniciosus). Aphididae: Thelaxes sp.). Toscana. Italy. GC.



Fig. 7.10.3. Twigs and fruits of *Euonymus* Fig. 7.10.4. Russeting damage to pear fruit sp. covered bv scales (Hemiptera, due to feeding by psyllids (Hemiptera, Diaspididae: Unaspis euonymi). Hungary, Psyllidae: Psylla pyricola). USA, EHB. GC.

Additional information: Note whether the organism present on the surface is protected by a soft or hard covering (scale), or if it is free living (aphid, woolly adelgid). Note the presence of honeydew. The damage symptoms caused by psyllids and mites are rather similar, but psyllid damage is typified by the presence of honeydew on the fruit. For insect preservation, see Chapter 3.

7.11. Rust diseases

Description: Yellow/orange/red spores/blisters and/or pustules on the fruit surface

Possible damaging agents: Fungi: Basidiomycota (Pucciniales: Figs. 7.11.1 -7.11.4).



Fig. 7.11.1. Rose hip (Rosa sp.) covered by a Fig. 7.11.2. Rose flower (Rosa sp.) infected rust fungus (Basidiomycota, Pucciniales: by Phragmidium sp.). Akershus Norway, VT.



Fig. 7.11.3. Fruit of pear (Pyrus communis) Fig. 7.11.4. Salix sp. attacked by a rust with medusa-like head due to a secondary fungus (Melampsora sp.). Sør-Trøndelag infestation by hawthorn rust (Basidiomycota, County, Norway, VT. Pucciniales: Gymnosporangium clavariiforme), the primary host being Juniperus. Slovakia, MZ.



а rust fungus (Basidiomycota, county, Pucciniales: Phragmidium sp.). Briancon, France, AR.



Additional information: Rust fungi are highly host specific, produce up to five spore stages and commonly need an alternate host to fulfil the life cycle.

7.12. Moulds

Description: Dark grey, green, blue or white velvet-like powdery mycelium on reproductive surfaces.

Possible damaging agents: Fungi: Heliothales (Sclerotinicaeae: Figs. 7.12.1 – 7.12.2).



Fig. 7.12.1. Flowers of *Rhododendron* **Fig. 7.12.2.** Detail of photo 7.12.1. *luteum* infected by grey mould (Heliothales, Sclerotinicaeae: *Botrytis cinerea*). Akershus county, Norway, VT.

<u>Additional information</u>: Moulds may grow on honeydew substances secreted by aphids and other insects, turn seed coats blackish, and form dark grey spots on cotyledon surfaces.

7.13. Powdery Mildew

Description: White velvet powdery mycelium on fruits and dark grey spots on cotyledon surfaces of the seeds.

Possible damaging agents: Fungi: Ascomycota (Erysiphales: Figs. 7.13.1 -7.13.4).



Fig. 7.13.1. Fruit of American gooseberry, Fig. 7.13.2. Achene of maple (Acer sp.) *Ribes uva-crispa*, infested by mildew with necrotized parts due to a fungus (Ascomycota, Erysiphales: Podosphaera (Ascomycota, mors-uvae). Slovakia, JM.



Ervsiphales: Sawadea bicornis). The necrotized sections were at first chlorotic, and covered with a whitish powdery coating. Zürich, Switzerland, OH.



Fig. 7.13.3. Fruit of peach, Prunus persica, Fig. infected by powdery mildew (Ascomycota, Rhododendron (Azalea molle X sinense), Erysiphales: Podosphaera pannosa var. infected by powdery mildew (Ascomycota, persicae). Slovakia, JM.



7.13.4. Fruits and leaves of Erysiphales: Erysiphe azaleae). Bergen, Norway, VT.

Additional information: Generally, powdery mildew is easy to recognize and most species are host specific.

7.14. Fruit galling

Description: Fruit clearly deformed, swollen, enlarged, or with uneven development. When the fruits are opened, larva(e), insect chamber(s), or galleries are visible.

Possible damaging agents: Insects: Larvae of Diptera (Cecidomyiidae midges: Figs. 7.14.4, 7.14.5) and Hymenoptera (Cynipidae: Figs. 7.14.1 – 7.14.3); Mites: Acari (Eriophiidae: Fig. 7.14.6).





Fig. 7.14.1. Acorn of oak (Quercus robur), Fig. 7.14.2. Acorn of oak (Quercus robur), completely deformed by a gall wasp with a spiny gall due to a wasp (Hymenoptera, Cynipidae: quercuscalicis; asexual Hungary, GC.

Andricus (Hymenoptera Cynipidae: Andricus generation). lucidus; asexual generation). Hungary, GC.



Fig. 7.14.3. Acorn of Ouercus cerris, sliced Fig. 7.14.4. Fruit of linden (Tilia cordata) open to show the larval chambers of a gall galled (reddish) by a midge larva (Diptera, (Hymenoptera, wasp Pseudoneuroterus saliens; generation). Hungary, GC.



Cynipidae: Cecidomyiidae: Contarinia tilliarum). sexual Hungary, GC.





7.14.5. Young fruit of pear (*Pyrus* **7.14.6.** Fruit of walnut (*Juglans regia*) with *communis*) galled in the calyx cavity by protruding galls on the surface, due to a larvae of pear gall midge (Diptera, mite (Acari, Eriophyidae: *Aceria tristriata*). Cecidomyiidae: *Contarinia pyrivora*). Slovakia, JM.

Additional information: Dissect the gall to check for the presence of insect larva(e) or minute mites. Note if there are distinct larval chambers. The insects and mites may have already left the gall, check also for the presence of exit holes. Such galls may also contain parasites, predators, or inquiline species. For insect preservation, see Chapter 3.

7.15. Fruit fungal deformation

Description: Fruit clearly deformed, swollen, enlarged, or with uneven development. An elongated, sac or pocket-like hollow fruit develops without an embryo, and without larva(e) or larva tunnel(s) inside.

Possible damaging agents: Fungi: Ascomycota (Taphrinomycetes: Figs. 7.15.1, 7.15.2).



Fig. 7.15.1. Plum tree (Prunus domestica) Fig. 7.15.2. Prunus fruits infected by plum with an elongated, slightly curved fruit due to infection by the plum pocket fungus (Ascomycota, Taphrinomycetes: Taphrina pruni). Sogn og Fjordane county, Norway, AS.



pocket fungus (Ascomycota, Taphrinomycetes: Taphrina pruni) at an advanced stage, showing a thick flour-like white coating. Later the deformed fruits will dry and rot, but may persist for a long time on twigs as mummies. Slovakia, JM.



Fig. 7.15.3. Fruits of wild cherry (Prunus Fig. 7.15.4. Detail of a deformed fruit padus) infected by plum pocket fungus shown in Fig. 7.15.3. (Ascomycota, Taphrinomycetes: Taphrina pruni). Akershus county, Norway, VT.

Additional information: Taphrina pruni commonly occurs on plum trees, but can also be found on other woody plants, e.g. Prunus padus (Figs. 7.15.3 -7.15.4).

7.16. Bacterial diseases and viruses

Description: Bacteria form black or brown lesions or blotches where bacterial exudate may ooze out under humid conditions. Virus attacks result in chlorotic ring structures or other patterns on the fruits.

Possible damaging agents: Bacteria (Fig. 7.16.1), Viruses (Figs. 7.16.2 -7.16.4)



Fig. 7.16.1. Oozing canker on walnut Fig. 7.16.2. Fruit of peach (Prunus persica) (Juglans regia) due to a bacterium (Bacteria, with chlorotic ring structures on the fruit Proteobacteria, Xanthomonas arboricola py. juglandis). (Virus, Potyviridae). Slovakia, JM. Slovakia, JM.



Xanthomonadales: surface due to plum pox virus (sharka)

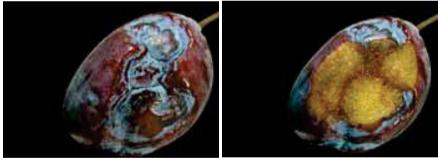


Fig. 7.16.3. Plum fruit (Prunus domestica) Fig. 7.16.4. Sharka blotches becoming with blotches due to plum pox virus (sharka) harder as the plum fruit shown in 7.16.3 is (Virus, Potyviridae). Slovakia, JM.

maturing. Slovakia, JM.

Additional information: Look for presence of aphids on the plant, which may transmit and inject bacteria into the fruit.

7.17. Nut rot

Description: Fruit entirely or partly discoloured; black or tan-brown discoloration on the inside or outside of the fruit, with presence of small raised, cream-coloured fruiting structures.

Possible damaging agents: Fungi: Ascomycota (Figs. 7.17.1 – 7.17.2).



Fig. 7.17.1. Mummified nuts of hazelnut Fig. 7.17.2. Nut of pecan tree (Carya (Corylus avellena) due to a brown rot fungus illinoinensis) showing shuck rot due to a (Ascomycota, Helotiales: Monilinia laxa). fungus (Ascomycota: Glomerella sp.). Slovakia, JM.

USA, WR.

Additional information: Infected nuts often die before they are fully grown. Diseased, mature nuts may have fungal growth on the seed inside the nutshell.

7.18. Hull Rot (mummification) of stone fruits

Description: Fruit entirely or partly discoloured. Shrivelled stone fruits or intermixed acorn tissues held together by fungal mycelium. Fruits turn chalky and sponge-like.

Possible damaging agents: Fungi: Ascomycota (Helotiales: Figs. 7.18.1 -7.18.4).



Fig. 7.18.1. Apple fruit (Malus sylvestris) Fig. 7.18.2. Mummified apple fruit (Malus with superficial, circular, brown spots sylvestris) as the final result of damage by a expanding on the surface due to a brown rot brown rot fungus (Ascomycota, Helotiales: fungus (Ascomycota, Helotiales: Monilinia Monilinia fructigena), Slovakia, MZ. fructigena), Slovakia, MZ.



Fig. 7.18.3. Mummified plum (Prunus Fig.7.18.4. Fruit of pear (Pyrus communis) domestica) due to a brown rot fungus with a circular brown spot expanding on (Ascomycota, Helotiales: Monilinia laxa), the surface due to a brown rot fungus Slovakia, JM.





(Ascomycota, Helotiales: Monilinia fructigena). Slovakia, MZ.

Additional information: Look for presence of mummified fruits on the ground or still attached to the tree.

7.19. Black rot (mummification) of acorns

Description: Fruit entirely or partly discoloured. Shrivelled stone fruits or mixed acorn tissues held together by fungal mycelia. Fruits turn chalky and sponge-like. Possible damaging agents: Fungi: Ascomycota (Helotiales: Figs. 7.19.1, 7.19.2).





Ciboria

Fig. 7.19.1. Acorns of oak (Quercus sp.) Fig. 7.19.2. Acorns of oak (Quercus sp.) partly infected by a ciboria fungus totally infected by a ciboria fungus (Ascomycota, Helotiales: Ciboria (Ascomycota, Helotiales: batschiana). Slovakia, AK. batschiana). Slovakia, AK.

Additional information: Infected acorns will normally fail to germinate.

7.20. Stone fruit scab

Description: Fruit entirely or partly discoloured. Extensive crusty corky spotting on the surface of fruit with cracks, lesions or necrosis, often clustered near the stem end of the fruit.

Possible damaging agents: Fungi: Ascomycota (Pleosporales: Figs. 7.20.1 -7.20.4).



Fig. 7.20.1. Pyrus communis fruit covered Fig. 7.20.2. Malus sylvestris fruit covered by brown-black spots due to pear scab by brown-black spots due to apple scab fungus (Ascomycota, Pleosporales: Venturia fungus pyrina). Slovakia, JM.



Fig. 7.20.3. Prunus persica fruit covered by Fig. 7.20.4. Nuts of pecan tree (Carva brown-black spots due to peach scab fungus *illinoinensis*) showing increasing degrees of (Ascomycota, Pleosporales: carpophila). Slovakia, JM.



(Ascomycota, Pleosporales: Venturia inaequalis). Slovakia, JM.



Venturia pecan scab (Ascomycota, Pleosporales: Fusicladium effusum). USA, WR.

Additional information: Scab greatly reduces the fruit quality for consumption. Thus, proper management is required in the fruit producing industry.

7.21. Fruit spot

Description: Fruit entirely or partly discoloured. Circular or irregular, elongated, bright or dark brown, red, nearly black, distinct and gradually coalescing or sharply outlined spots of various dimensions, but no larva or tunnel present when the fruit is opened.

Possible damaging agents: Fungi: Ascomycota (Capnodiales: Figs.7.21.3 -7.21.4; Diaporthales: Figs. 7. 21.1 – 7.21.2).

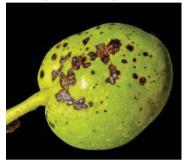


Fig. 7.21.1. Nut of walnut (Juglans regia) Fig. 7.21.2. The same damage as 7.21.1., showing brown patches due to walnut-leaf but at the end of the growing season: the blotch fungus (Ascomycota, Diaporthales: blotches having merged. Slovakia, AK. Gnomonia leptostyla). Slovakia, AK.



Fig. 7.21.3. Apricot fruit, covered by spots Fig. 7.21.4. Peach fruit (Prunus persica) shot-hole disease due to а Capnodiales: (Ascomvcota. carpophilum Stigmina = Slovakia, JM.





fungus with spots due to a shot-hole disease fungus Thyrostroma (Ascomycota, Capnodiales: Thyrostroma *carpophila*). *carpophilum* Stigmina carpophila), = Slovakia, JM.

Additional information: Be aware that fruit spots may also be caused by bacterial infections

7.22. Internal fruit feeding by insect larva

Description: Fruit entirely or partly discoloured or with scattered deposits of gum (thick sap: Fig. 7.22.1) or frass (larval excrements: Figs. 7.22.3, 7.22.4) on the surface. When the fruit is cut open, larva(e) or tunnel(s) are visible (Figs 7.22.2, 7.22.6, 7.22.7).

Possible damaging agents: Insects: larvae of Coleoptera (Curculionidae and Rhynchitidae weevils: Figs. 7.22.5, 7.22.6), Diptera (Cecidomyiidae, Phoridae, Sciaridae and Tephritidae fruit flies: Fig. 7.22.7), Hymenoptera (Tenthredinidae sawflies: Fig. 7.22.8) and Lepidoptera (Gelechiidae, Geometridae, Noctuidae, Tortricidae, Yponomeutidae: Figs. 7.22.1 – 7.22.4).



Fig. 7.22.1. Plum fruit (Prunus domestica) Fig. 7.22.2. The plum fruit shown in Fig. with gum deposit on the surface, resulting 7.22.1., sliced to show the moth larva and from internal feeding by a moth larva its damage. Slovakia, JM. (Lepidoptera, Tortricidae: Cydia funebrana). Slovakia, JM.





Fig. 7.22.3. Apple fruit (Malus sylvestris) Fig. 7.22.4. Hip of wild rose (Rosa sp.) with protruding frass resulting from internal with protruding frass due to a moth larva damage by a larva of codling moth (Lepidoptera, (Lepidoptera, Tortricidae: Cydia pomonella). scirrhosella). Hungary, GC. Slovakia, MZ.

Tortricidae: Carposina



attacked by larvae of apple fruit weevil sativa) sliced to show damage by weevil (Coleoptera, Rhynchitidae: Tatianaerhynchites aequatus). Slovakia, MZ.



Fig. 7.22.7. Cherry fruit (Prunus cerasus) Fig. 7.22.8. Apple fruit (Malus sylvestris) sliced to show the damage caused by a larva with a gallery along the underside of the of a fruit fly (Diptera, Tephritidae: epidermis made by a sawfly larva Rhagoletis cerasi). Slovakia, JM.



Fig. 7.22.5. Apple fruit (Malus sylvestris) Fig. 7.22.6. Chestnut fruit (Castanea larvae (Coleoptera, Curculionidae: Curculio sp.). Marcillac, France, AR.



(Hymenoptera, Tenthredinidae: Hoplocampa testudinea). Slovakia, JM.

Additional information: Note whether the extruded frass is coarse or light, and combined or not with gum. If the fruit is only partly discoloured and without exit holes, the damaging larvae are still present. Carefully cut the fruit longitudinally, like in Figs. 7.22.2, 7.22.6 and 7.22.7, and check the discoloured parts for the presence of larvae. Note the position of the larvae and the larval tunnels. Then, extract larvae, and note the shape of the tunnel. Note that infested fruits can be secondarily attacked by fungal pathogens. For insect preservation, see Chapter 3.

7.23 Mite-induced discoloration

Description: Fruit noticeably speckled or russeted. **Possible damaging agents: Mites:** Acari (Eriophyiidae: Figs. 7.23.1 – 7.23.2).



Fig. 7.23.1. Fruit of plum (*Prunus* Fig. 7.23.2. Russeting of pear fruit by pear *domestica*) attacked by silver mites (Acari, rust mite (Acari, Eriophyiidae: *Epitrimerus* Eriophyiidae: *Aculus fockeui*). Slovakia, JM. *pyri*). California, USA, JKC.

<u>Additional information</u>: Look for the presence of minute mites on the fruit. For collection and preservation, refer to Chapter 3.

7.24. Larval emergence hole

Description: Presence of irregular exit hole(s) on the surface.

Possible damaging agents: Insects: mature larvae, falling down to pupate on the ground, of Coleoptera (Curculionidae and Rhynchitidae weevils: Figs. 7.24.1 -7.24.3), Diptera (Cecidomyiidae, Phoridae, Sciaridae and Tephritidae fruit flies: Fig. 7.24.6), Hymenoptera (Tenthredinidae sawflies: Fig. 7.24.4), and Lepidoptera (Gelechiidae, Geometridae, Noctuidae, Tortricidae, Yponomeutidae: Fig. 7.24.5); immature larvae of Lepidoptera may move from one fruit to another.



with exit holes of larvae of apple fruit weevil sativa) with exit holes of weevil larvae Rhynchitidae: (Coleoptera, (Coleoptera, Tatianaerhynchites aequatus). Slovakia, elephas). Gorduno, Switzerland, BW. MZ.

Fig. 7.24.1. Apple fruit (Malus sylvestris) Fig. 7.24.2. Fruit of chestnut (Castanea Curculionidae: Curculio

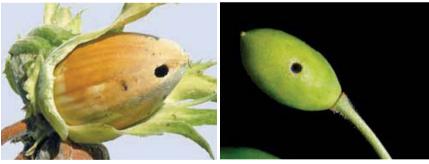


Fig. 7.24.3. Hazelnut (Corylus avellana) Fig. 7.24.4. Plum (Prunus sylvestris) with with an exit hole of a weevil larva an exit hole of a sawfly (Coleoptera, Curculionidae: nucum). Slovakia, JM.

larva Curculio (Hymenoptera, Tenthredinidae: Hoplocampa minuta). Slovakia, JM.



spinosissima) with an exit hole of a moth (central arrow), exit hole for larva (left larva (Lepidoptera, Tortricidae: Carposina arrow) and the larva (right arrow) of a fruit scirrhosella). Briançon, France, AR.

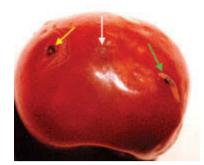


Fig. 7.24.5. Hip of wild rose (Rosa Fig. 7.24.6. Cherry fruit oviposition sting fly (Diptera, Tephritidae: Drosophila suzukii). Howard County, Maryland, USA, GB.

Additional information: Usually, no more larvae are present, but check for the presence of dead specimens which could not emerge from the fruit. Note the shape and size of the exit hole, which can help the identification of the insect.

Description: Presence of exit hole(s) of regular shape on fruit surface. Possible damaging agents: Insects: adults of Coleoptera seed beetles (Bruchidae, Curculionidae: Fig. 5.25.3) and Hymenoptera seed chalcids (Eurytomidae: Fig. 5.25.4, Torymidae: Figs. 5.25.1 - 5.25.2).



Fig. 5.25.1. Hip of wild rose (Rosa sp.) with Fig. 5.25.2. Fruit of Rhus natalensis an emergence hole of an adult seed chalcid showing an adult exit hole of a seed chalcid (Hymenoptera, Torymidae: Megastigmus (Hymenoptera, Torymidae: Megastigmus sp.). Lijiang, China, AR.



transvaalensis). Ronge Nyika, Kenya, RC.



insect exit holes (among others, seed beetles with exit holes of adult seed chalcids belonging to Coleoptera, Bruchidae). Moshi (Hymenoptera, Eurytomidae: Eurytoma District. Tanzania. RE.



Fig. 5.25.3. Pods of Prosopis juliflora with Fig. 5.25.4. Almond fruits (Prunus dulcis) amygdali). Sallèles, France, AR.

Additional information: Usually, no more insects are present, but check for the presence of dead specimens, which could not emerge from the fruit. Also, check the presence of exit holes on seeds. Note the diameter of the exit hole. Emerging parasites of the phytophagous pests may also bore similar circular holes to emerge from the fruit.

7.26. Seed galling

Description: Seed noticeably deformed.

Possible damaging agents: Insects: Larvae of Diptera (Cecidomyiidae gall midges: Fig. 7.26.1) and Hymenoptera (Cynipidae gall wasps: Fig. 7.26.2), Mites: Acari (Eriophyiidae).



Fig. 7.26.1. Seed of birch (Betula sp.), galled Fig. 7.26.2. Oak acorn (Quercus sp.) sliced by a midge larva (Diptera, Cecidomyiidae: to show the gall chambers (note the larvae) Semudobia *betulae*). North Denmark, LKT.

Zeeland, of a gall wasp (Hymenoptera, Cynipidae: Callirhytis glandium). Hungary, GC.

Additional information: Collect mature seeds/acorns. The seed must be opened or, better, X-rayed if possible to ascertain the presence of larvae. Alternatively, for a more easy identification of the damaging species, infested seeds can be reared until adult emergence. For insect preservation see Chapter 3.

7.27. Seed rot

Description: Seed partly/completely discoloured with dark grey, green, blue, white velvet-powdery mycelium on the seed coat.

Possible damaging agents: Fungi: Ascomycota (Penicillium, Fusarium, Phoma and others).



Fig. 7.27.1. Seed of European beech (Fagus Fig. 7.27.2. Seed of European beech sylvatica) infected by Penicillium sp. (Fagus sylvatica) infected by fungal rot (Ascomycota, Eurotiales). Vestfold county, (Ascomycota, Pleosporales: Phoma sp.). Norway, VT.

Vestfold county, Norway, VT.

7.28. Seed insect damage

Description: Presence of circular hole(s) in external seed coat or presence of larva(e) or pupa(e) in the seed when opened (or larvae/pupae visible using Xrays).

Possible damaging agents: Insects: Larvae of Coleoptera seed beetles (Bruchidae, Curculionidae: Figs. 7.28.4 left - 7.28.5), Diptera seed midges (Cecidomviidae, Phoridae: Fig. 7.28.7), Hymenoptera seed chalcids (Eurytomidae: Fig. 7.28.6, Torymidae: Fig. 7.28.8) and Lepidoptera (Nepticulidae, Yponomeutidae, Tortricidae: Figs. 7.28.1, 7.28.2, 7.28.3, 7.28.4 right).



7.28.1. Seed of maple (Acer Fig. 7.28.2. Fig. pseudoplatanus) with an exit hole of a pseudoplatanus) with a lepidopteran seed moth Tortricidae: Pammene regiana). Hungary, Tortricidae: Pammene regiana). Hungary, GC.



Seed of maple (Acer larva of a (Lepidoptera, lepidopteran seed moth (Lepidoptera, GC.





Fig. 7.28.3. sylvatica) with emergence holes of a seed sliced to show damage by a weevil larva moth (Lepidoptera, Tortricidae: Cvdia sp.). (Coleoptera, Curculionidae: Curculio sp.) Hungary, GC.

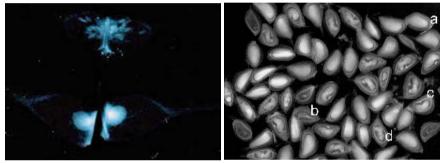
Acorn of beech (Fagus Fig. 7.28.4. Acorns of oak (Ouercus sp.), (left) and a moth larva (Lepidoptera, Tortricidae: Cydia sp.) (right). Hungary, GC.



Fig. 7.28.5 Seed of maple pseudoplatanus) dissected to show the pupa lentiscus) with an exit hole of a seed of a seed weevil (Coleoptera, Curculionidae: chalcid Bradybatus kellneri). Hungary, GC.



(Acer Fig. 7.28.6. Seed of pistachio (Pistacia (Hymenoptera, Eurytomidae: Eurytoma plotnikovi). Tabarka, Tunisia, AR.



(Acer pseudoplatanus), to show seed rose (Rosa odorata), infested by chalcid infested by dipteran larva (top) (Diptera, larva (c) and pupae (d) (Hymenoptera, Phoridae: Megaselia sp.) and sound seed Torymidae: Megastigmus sp.) and sound (bottom). Grenoble, France, AR.

Fig. 7.28.7. X-ray picture of seed of maple Fig. 7.28.8. X-ray picture of seed of wild (a) and empty (b) seed. Lijiang, China, AR.

Additional information: Collect mature seeds. Usually, an infested seed does not differ from a healthy one in shape, colour or weight, although exceptions exist. The seed must be opened or, better, X-rayed if possible to ascertain the presence of larvae. Alternatively, for a more easy identification of the damaging species, infested seeds can be reared until adult emergence. For insect preservation see Chapter 3.