

Supplementary Information

PeroxyHUB: a modular, cell-free biosensing platform using H₂O₂ as signal integrator.

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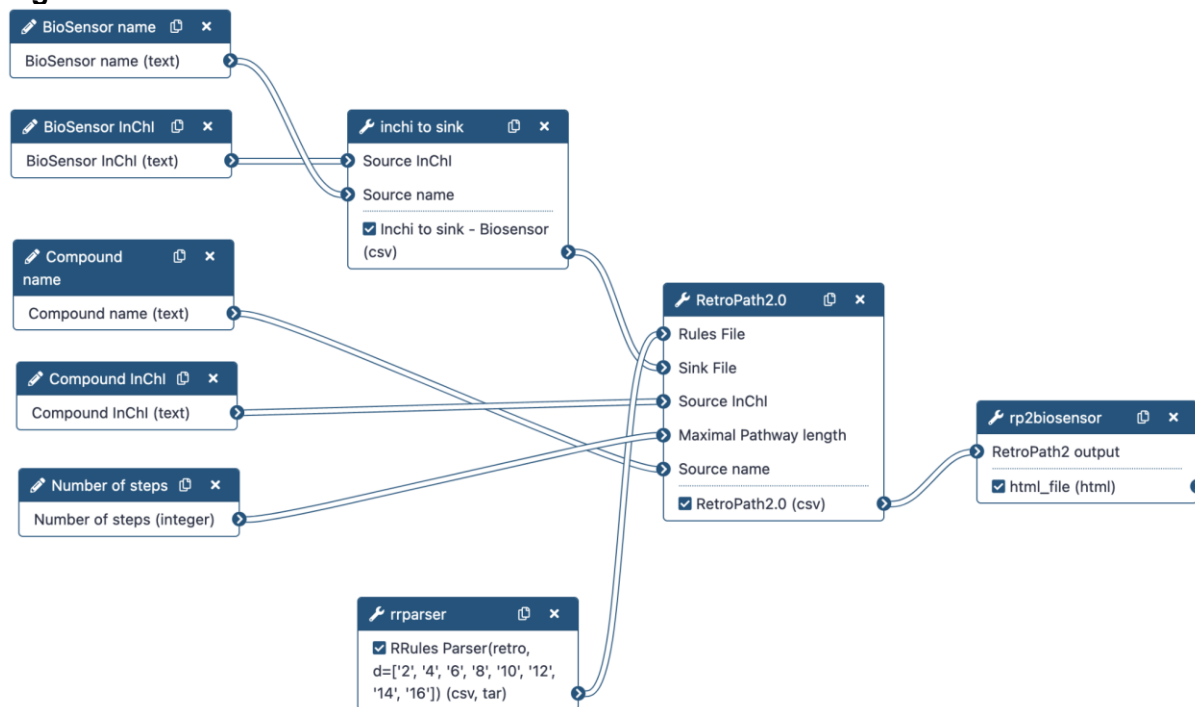
† These authors contributed equally.

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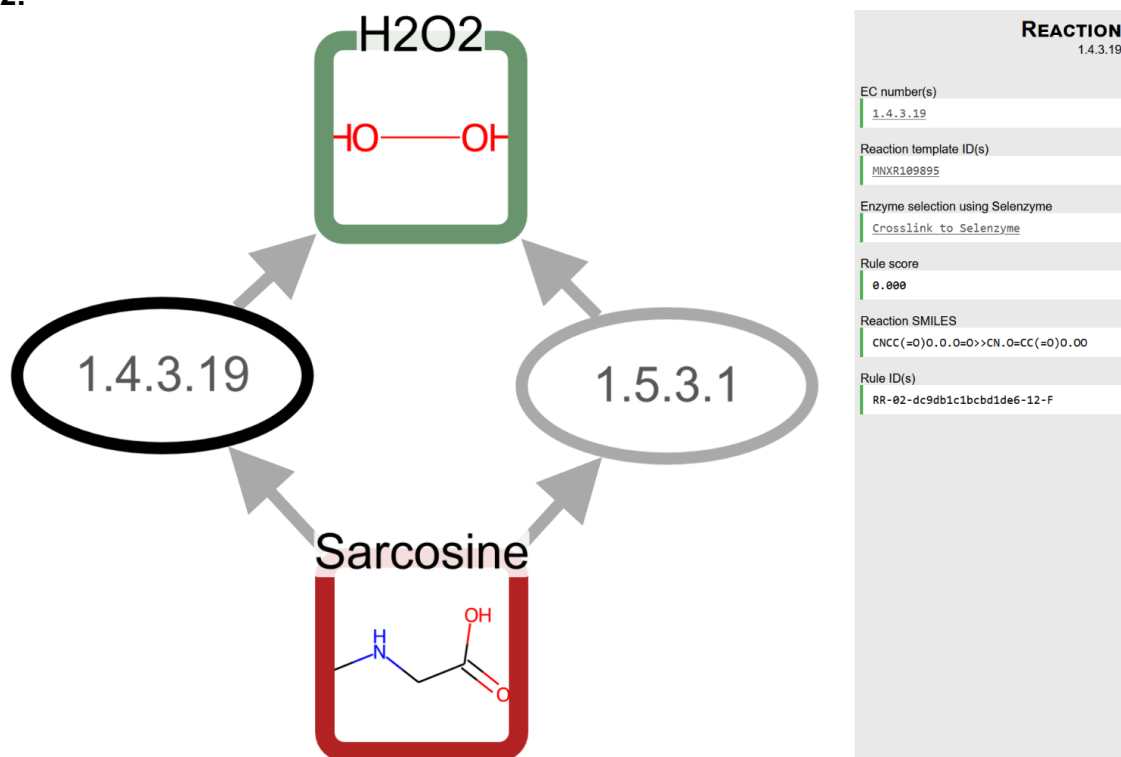
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Fig S1:



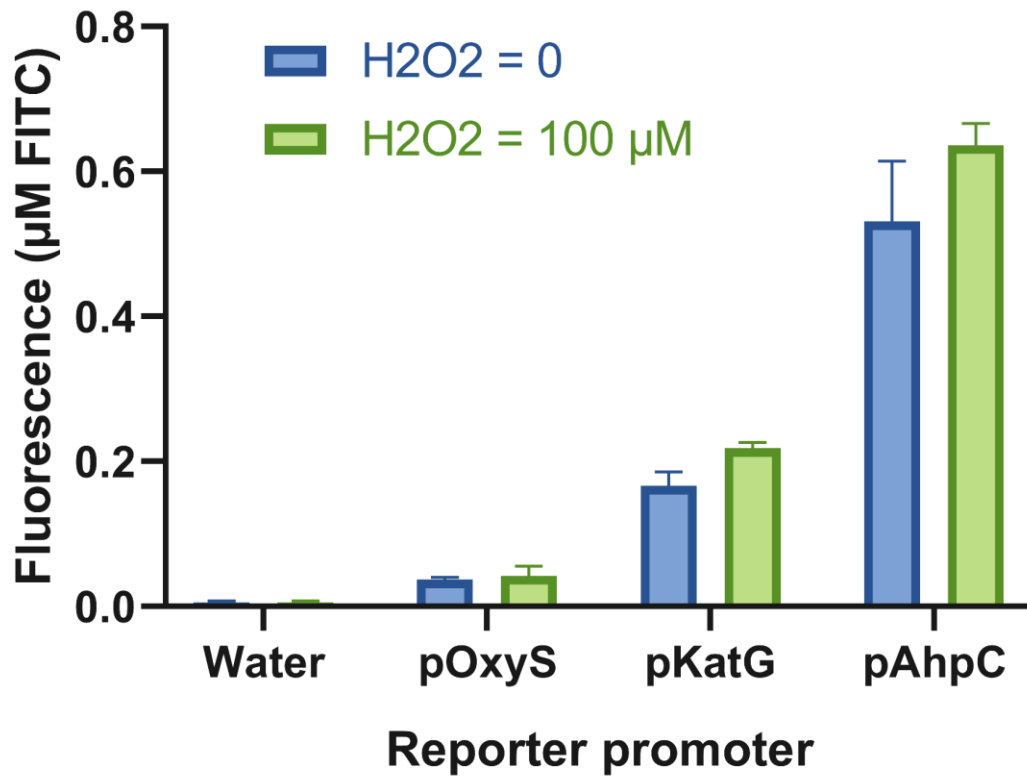
Supplementary Figure S1: Complete view of the BioSensor Galaxy workflow automatizing biosensor predictions. Characteristics of the main nodes are described in the method section.

Fig S2:



Supplementary Figure S2: Example of result web page output from the Biosensor workflow for sarcosine.

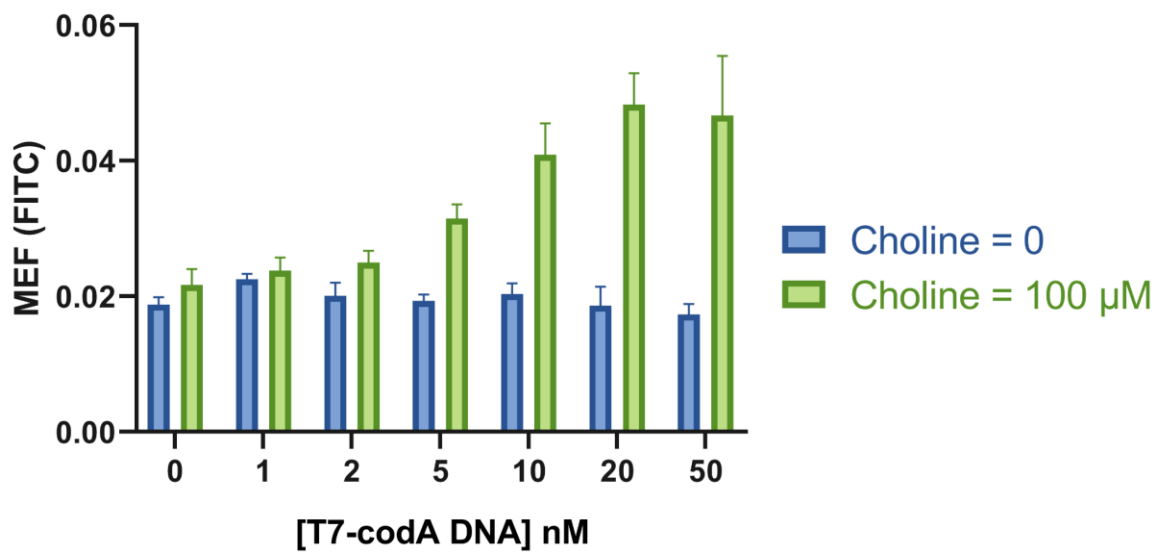
Fig S3:



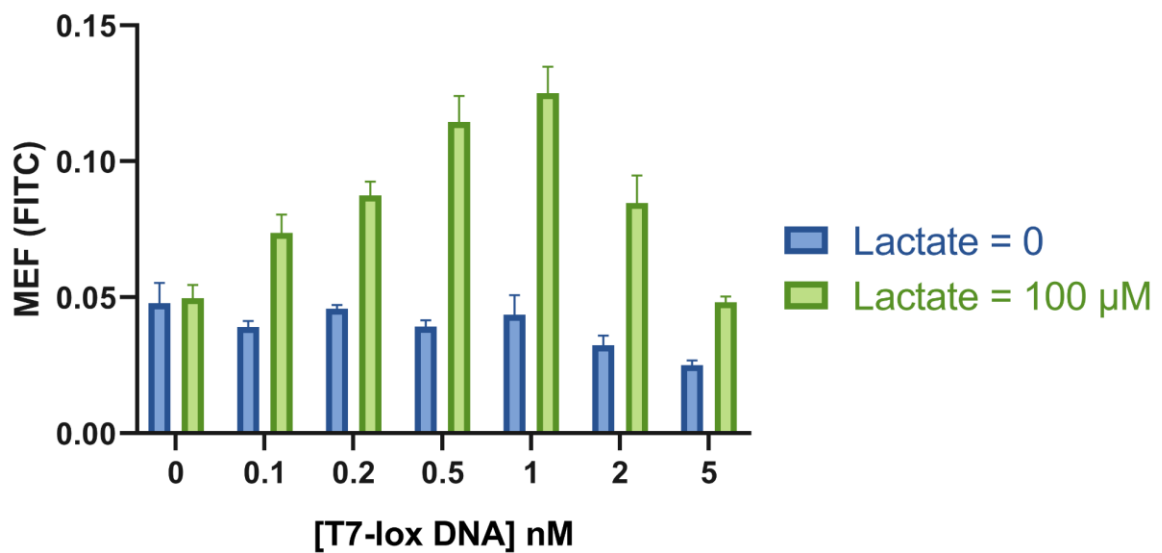
Supplementary Figure S3: Unoptimized H₂O₂ sensor response in cell-free system. Direct cell-free implementation of biosensor candidates derived from *in vivo* design did not show significant response to H₂O₂. 16.5µL of Cell-free mix was supplemented with 1.1µL of TF and Reporter plasmid each, 1.1µL water and 2.2µL of either water or H₂O₂. The final concentrations were of 10 nM for each plasmid added and 100µM for the H₂O₂ inducer. Fluorescence values are MEF measured at an endpoint of 8h

Fig S4:

A

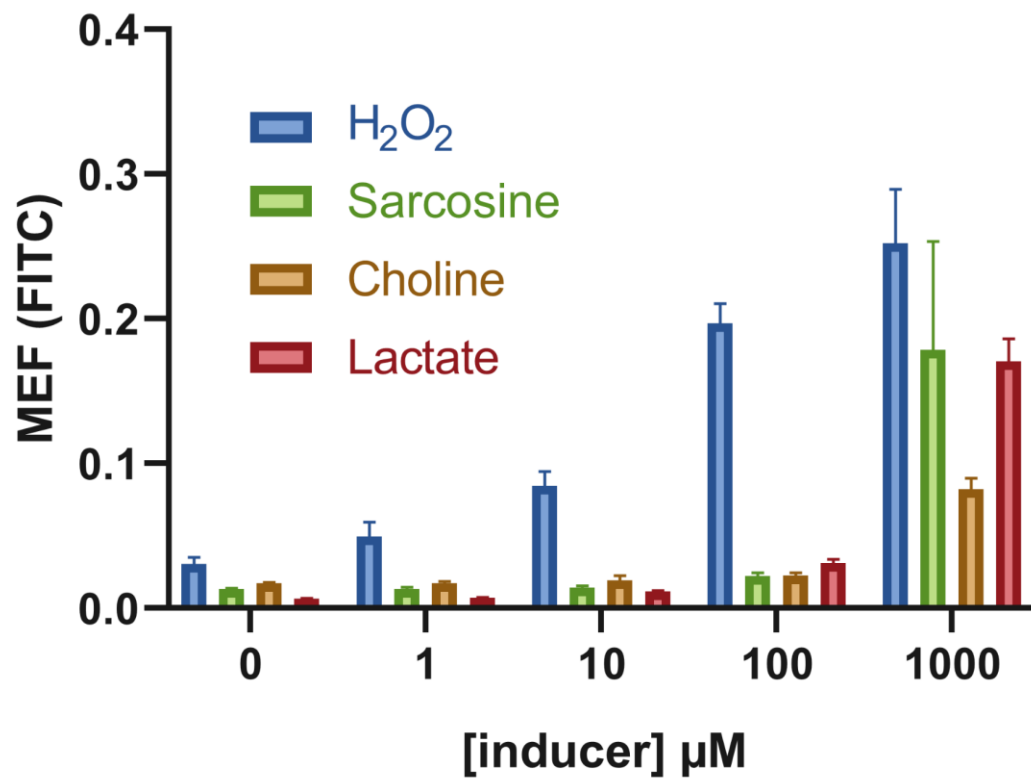


B



Supplementary Figure S4: Fine tuning of enzyme expression using DNA gradient (A) [T7-codA DNA] optimisation for choline sensing (B) [T7-lox DNA] optimisation for lactate sensing

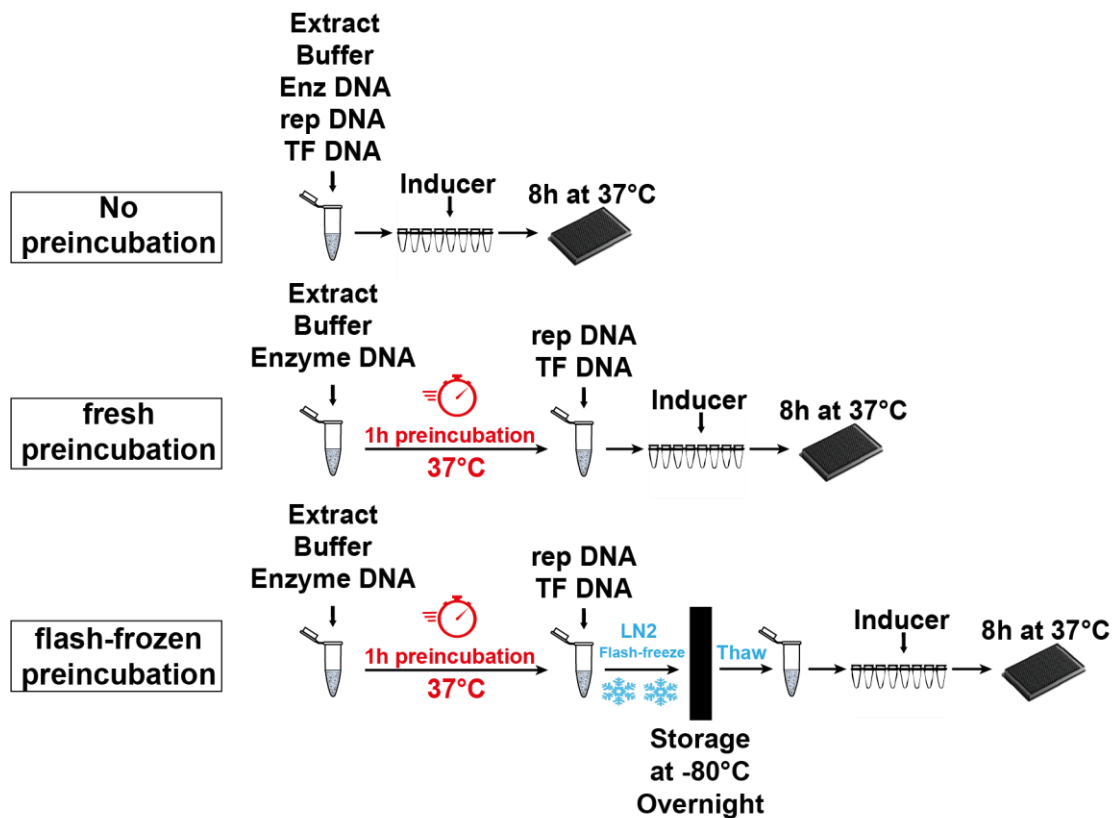
Fig S5:



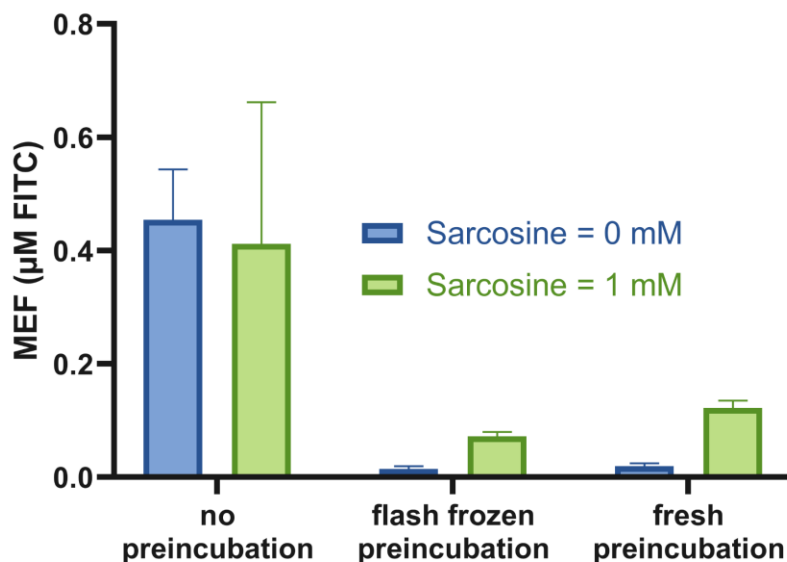
Supplementary Figure S5: Final sensors fluorescent dose response

Fig S6:

A



B

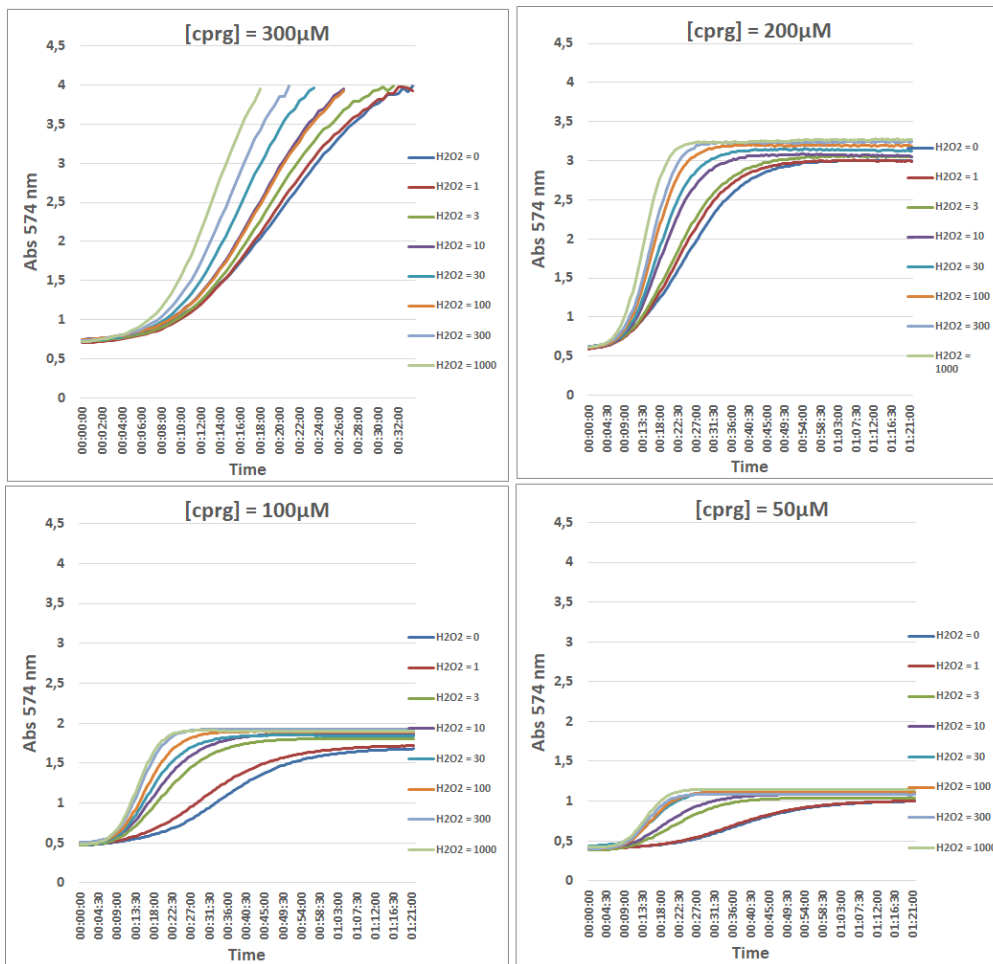


Supplementary Figure S6: Liquid Nitrogen Flash-Freezing of preincubated mix (A)

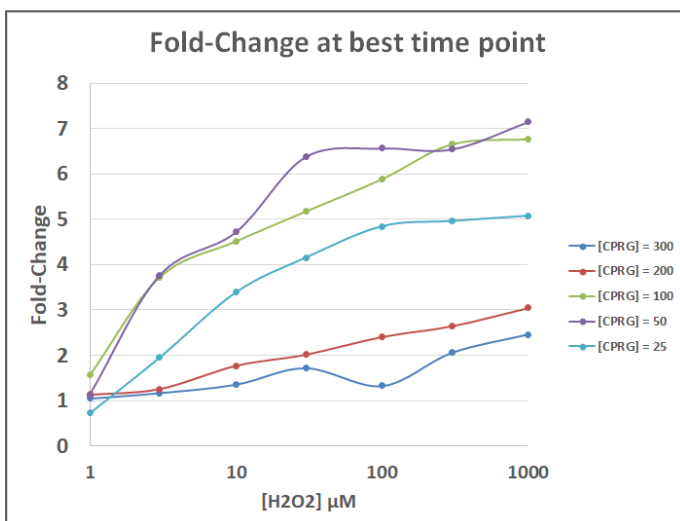
Experimental workflows detailing the no preincubation, the fresh preincubation and the flash frozen preincubation pipelines used for biosensors evaluation (B) 2h30 endpoint fluorescence taken in presence and absence of inducer for each pipeline tested reveals few differences of responses between the fresh and the flash frozen preincubation workflows.

Fig S7:

A

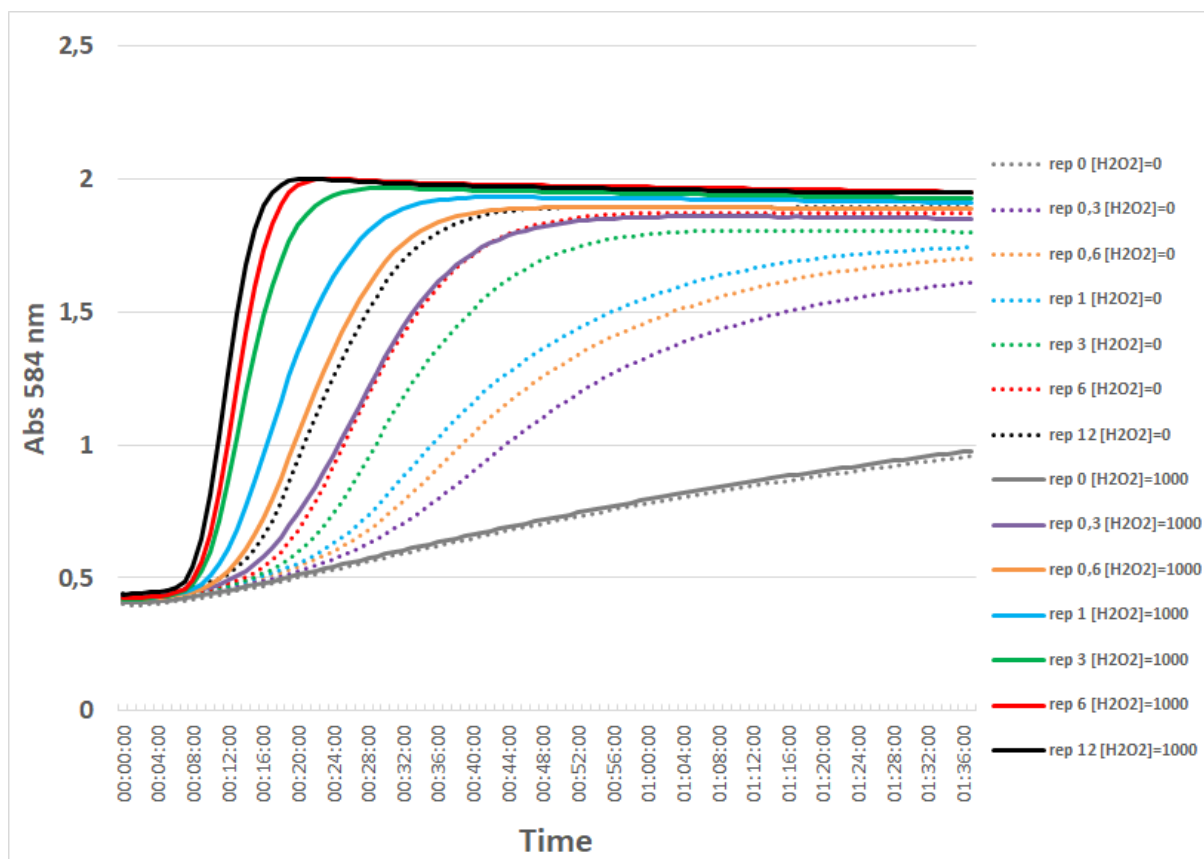


B



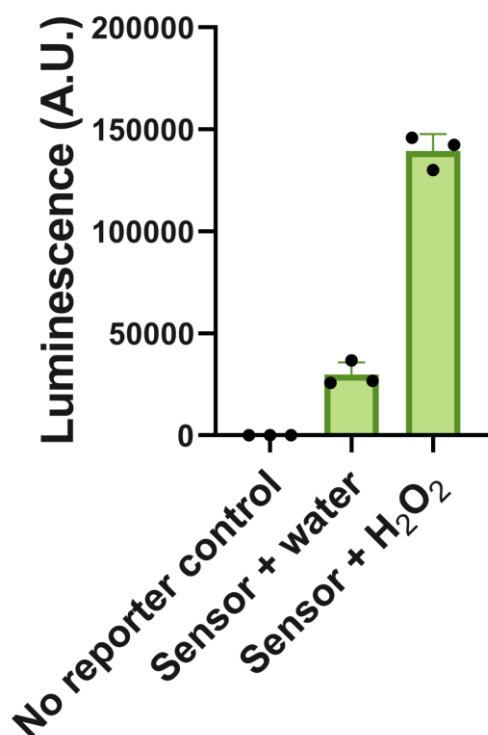
Supplementary Figure S7: CPRG concentration optimization for colorimetric H₂O₂ biosensor (A) 574 nm absorbance time course kinetic of colorimetric H₂O₂ biosensor with various initial concentration of CPRG (B) Blank subtracted H₂O₂ dose response curve of colorimetric biosensor with various initial concentration of CPRG

Fig S8:



Supplementary Figure S8: [pAhpC-LacZ DNA] concentration optimization for colorimetric H₂O₂ biosensor

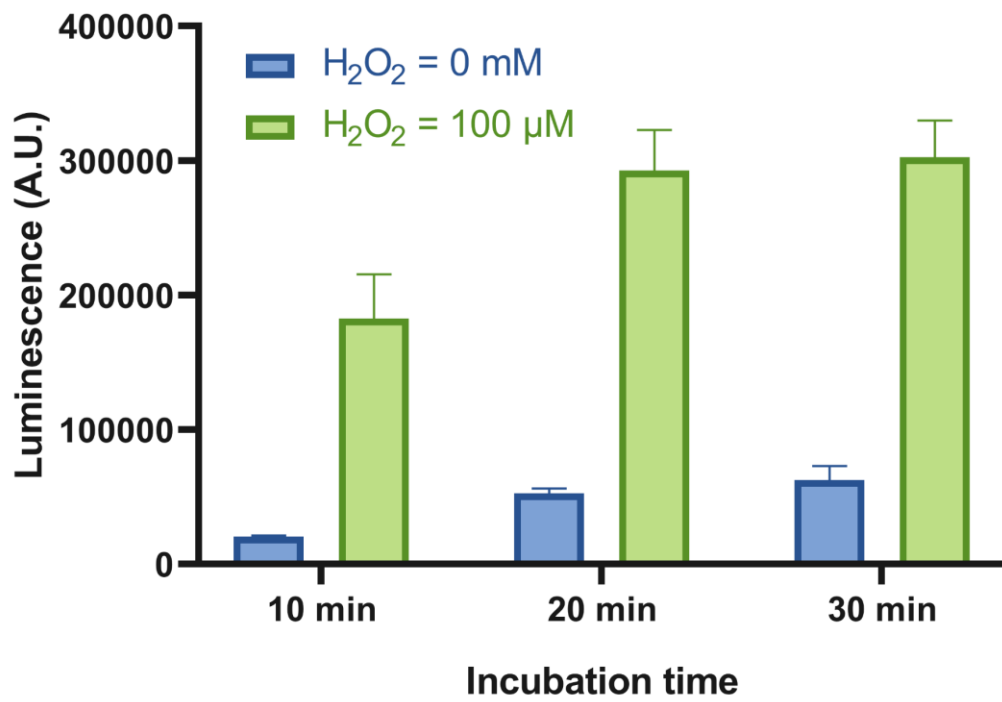
Fig S9:



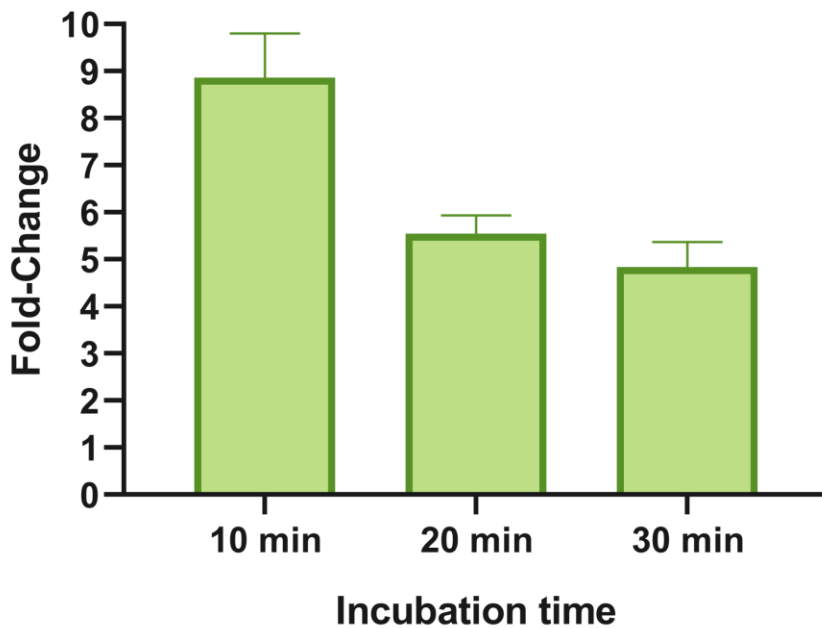
Supplementary Figure S9: Luminescent sensor early evaluation. [pAhpC-Luc DNA] was added at 0 or 6nM and [j23101-OxyR DNA] at 24 nM to 1h preincubated cell-free mix together with either water or 1mM H₂O₂. The reaction was incubated for 30 min at 37°C and 20µL of final reaction was added to 50 µL of Luciferin reagent mix before taking a luminescence point measurement. Dots represent individual values obtained. Error bars represent the standard deviation calculated from the 3 replicates.

Fig S10:

A



B



Supplementary Figure S10: Luminescent sensor incubation time optimisation (A)

Luminescence absolute values measured in arbitrary units **(B)** Luminescence Fold-Change between induced and non induced condition

Supplementary Table S1

Chemicals identifiers used in the study

Compound	InChI
S-lactate	InChI=1S/C3H6O3/c1-2(4)3(5)6/h2,4H,1H3,(H,5,6)/p-1/t2-/m0/s1
choline	InChI=1S/C5H14NO/c1-6(2,3)4-5-7/h7H,4-5H2,1-3H3/q+1
sarcosine	InChI=1S/C3H7NO2/c1-4-2-3(5)6/h4H,2H2,1H3,(H,5,6)

Supplementary Table S2

Characteristics of enzymes used in the study

substrate	name	uniprot ID	organism	kinetic		catalytic efficiency
				Km	Kcat	Kcat/Km
sarcosine	SoxA	P40859	<i>Bacillus subtilis</i>	141,6 mM source	0.204 s ⁻¹ source	1.441 s ⁻¹ .M ⁻¹ source
choline	CodA	Q7X2H8	<i>Arthrobacter globiformis</i>	0,6 mM source	13.4 s ⁻¹ source	22000 s ⁻¹ .M ⁻¹ source
lactate	lox	D4YFM2	<i>Aerococcus viridans</i>	0,5 mM source	88 s ⁻¹ source	176000 s ⁻¹ .M ⁻¹ source

Supplementary Table S3

Plasmids used in this study:

Plasmid	Antibiotic Selection	Accession/ Catalog reference	Source
pBeast-J23101-RBS-OxyR	Amp ^R		This work
pBeast-pAhpC-RBS-sfGFP	Amp ^R		This work
pBeast-pOxyS-RBS-sfGFP	Amp ^R		This work
pBeast-pKatG-RBS-sfGFP	Amp ^R		This work
pBeast-pYjjZ-RBS-sfGFP	Amp ^R		This work
pBeast-pZint-sfGFP	Amp ^R		This work
pBeast-J23101-RBS-soxA	Amp ^R		This work
pBeast-pT7-soxA	Amp ^R		This work
pBeast-pT7-codA	Amp ^R		This work
pBeast-pT7-lox	Amp ^R		This work
pBeast-pAhpC-RBS-LacZ	Amp ^R		This work
pBeast-pAhpC-RBS-Luc	Amp ^R		This work

Supplementary Table S4

DNA sequences for constructs used in this study:

Plasmid/ DNA construct	Sequence (5'->3')
pBeast-J23101-RBS-OxyR	<p>ATACTAGAGGATGACCCCATCTGTTTACAGCTAGCTCAGTCCTAG GTATTATGCTAGCTAGTAGAGTCACACAGGAAAGTAGTAGATGAAT ATTCGTGATCTTGAGTACCTGGTGGCATTGGCTGAACACCGCCAT TTTCGGCGTGCGGCAGATTCCCTGCCACGTTAGCCAGCCGACGCTT AGCGGGCAAATTCGTAAGCTGGAAGATGAGCTGGGCGTGATGTT GCTGGAGCGGACCAGCCGTAAGTGTGTTTACCCAGGCGGGAA TGCTGCTGGTGGATCAGGCGCGTACCGTGCTGCGTGAGGTGAAA GTCCTTAAAGAGATGGCAAGCCAGCAGGGCGAAACCATGTCCGG ACCGCTGCACATTGGTTTGATTCCACAGTTGGACCGTACCTGCT ACCGCATATTATCCCTATGCTGCACCAGACCTTCCAAAGCTGGAA ATGTATCTGCATGAAGCACAGACCCACCAGTTACTGGCGCAACTG GACAGCGGCAAACCTCGATTGCGTGATCCTCGCGCTGGTGAAAGA GAGCGAAGCATTGATTGAAGTGCCGTTGTTTATGAGCCAATGTT GCTGGCTATCTATGAAGATCACCCGTGGGCGAACCAGCAATGCGT ACCGATGGCCGATCTGGCAGGGGAAAAACTGCTGATGCTGGAAG ATGGTCACTGTTTGC GCGATCAGGCAATGGGTTTCTGTTTTGAAG CCGGGGCGGATGAAGATACACACTTCCGCGCGACCAGCCTGGAA ACTCTTCGCAACATGGTGGCGGCAGGTAGCGGGATCACTTTACTG CCAGCGCTGGCTGTGCCGCCGAGCGCAAACGCGATGGGGTTGT TTATCTGCCGTGCATTAAGCCGGAACCACGCCGCACTATTGGCCT GGTTTATCGTCTGGCTCACCGCTGCGCAGCCGCTATGAGCAGCT GGCGGAAGCCATCCGCGCAAGAATGGATGGCCATTCGATAAAGT TTAAAACAGGCGGTTTAAactttatctgagaatagtcaatcttcggaatcccaggtg gcatgctaaaagtctcgtaaagcgttctatcaataaccggttggtgccaggcatcaataaaaacga aaggctcagtcgaaagactgggcctttcgtttatctggtggtggtcggtgaacgctctactagagtc acactggctcacctcgggtggcctttctgctgttataccgtctcagaatcgcccggaacaataaa atagttcgggtattattgaccactccgagtagaatcgtgcttcagtaagagtcgaccgatgccctga gagcctcaaccagtcagctcctccggtgggcgcggggcaTGACTATCGTCGCCG CACTTATGACTGTCTTCTTTATCATGCAACTCGTAGGACAGGTGCC GGCAGCGCTCTTCCGCTTCCCTCGCTCACTGACTCGCTGCGCTCG GTCGTTCCGGCTGCGGCGAGCGGTATCAGCTCACTCAAAGGCGGT AATACGGTTATCCACAGAATCAGGGGATAACGCAGGAAAGAACAT GTGAGCAAAAGGCCAGCAAAAGGCCAGGAACCGTAAAAAGGCCG CGTTGCTGGCGTTTTTCCATAGGCTCCGCCCCCTGACGAGCATC ACAAAATCGACGCTCAAGTCAGAGGTGGCGAAACCCGACAGGA CTATAAGATAACCAGGCGTTTCCCCTGGAAGCTCCCTCGTGCGC TCTCCTGTTCCGACCCTGCCGCTTACCGGATACCTGTCCGCCTTT CTCCCTTCGGGAAGCGTGGCGCTTTCTCAATGCTCACGCTGTAGG TATCTCAGTTCGGTGTAGGTCGTTCCGCTCCAAGCTGGGCTGTGTG CACGAACCCCGTTTCCAGCCCGACCGCTGCGCCTTATCCGGTAA CTATCGTCTTGAGTCCAACCCGGTAAGACACGACTTATCGCCACT GGCAGCAGCCACTGGTAACAGGATTAGCAGAGCGAGGTATGTAG GCGGTGCTACAGAGTTCTTGAAGTGGTGGCCTAACTACGGCTACA CTAGAAGGACAGTATTTGGTATCTGCGCTCTGCTGAAGCCAGTTA CCTTCGGAAAAAGAGTTGGTAGCTCTTGATCCGGCAAACAAACCA CCGCTGGTAGCGGTGGTTTTTTTGTGTTGCAAGCAGCAGATTACGC</p>

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<p>pBeast-pKatG- RBS-sfGFP</p>	<p>TGTGGCTTTTATGAAAATCACACAGTGATCACAAATTTAAACAGA GCACAAAATGCTGCCTCGAAATGAGGGCGGGAAAATAAGGTTATC AGCCTTGTTTTCTCCCTCATTACTTGAAGGATATGAAGCTAAAACC CTTTTTATAAAGCATTGTCCGAATTCGGACATAATCAAAAAGCT TAATTAAGATCAATTTGATCTACATCTCTTAAACCAACAATATGTAA GATCTCAACTATCGCATCCGTGGATTAATTCAATTATAACTTCTCTC TAACGCTGTGTATCGTAACGGTAACACTGTAGAGGGGAGCACATT GATGCTAGAGAAAGAGGAGAAATACTAGatgctgtaaaggcgaagagctgtt cactggtgctgcccctattctggtggaactggatggatgtcaacggctataagtttccgctgctggtg cgaggggtgaaggtagcgaactaatggtaaactgacgctgaagttcatctgtactactggtaaact gccggtaccttgccgactctggtaacgacgctgactatggtgttcagtgtttgctcgttatccgga ccatatgaagcagcatgactcttcaagtcgccatgccggaaggctatgtgcaggaacgcacg attcctttaaggatgacggcacgtacaaaacgctgctggaagtgaattgaaggcgataccct ggtaaaccgcatgagctgaaaggcattgactttaaagaagacggcaataatcctgggccataag ctggaatacaatttaacagccacaatgttcatcaccgccgataaacaataaaatggcattaa gcgaatttaaaatccgccacaacgtggaggatggcagcgtgcagctggctgatcactaccagca aaacactccaatcggatggtctgttctgctgcccagacaatcactatctgagcagcgaagcgtt ctgtctaaagatccgaacgagaaaacgcatcatatggttctgctggagtctgaaccgcagcggg catcacgcatggtatggatgaactgtacaaatgatgaactttatctgagaatagtcaatcttcggaa atcccagtggtcatgctaaaagtctcgtaaagcgttctatcaataaccggttggtgccagcatca aataaaacgaaaggctcagtcgaaagactggcctttcgtttatctgtgtttgctggtgaaacgctct ctactagagtcacactggctcacctcgggtgggctttctgctgttataccgtctcagaatcggccgt gaacaataaaatagttcggattattgaccactccgagtagaatcgtgcttcagtaagagtcgac cgatgcccttgagagcctcaaccagtcagctcctccgggtggcgcggggcaTGACTATC GTCGCCGCACTTATGACTGTCTTCTTTATCATGCAACTCGTAGGAC</p>

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