



HAL
open science

Distribution of *Achromobacter* species in 12 French cystic fibrosis centers in 2020 by a retrospective MALDI-TOF MS spectrum analysis

Thomas Garrigos, Manon Dollat, Arnaud Magallon, Anaïs Folletet, Julien Bador, Maryam Abid, Marlène Amara, Clémence Beauruelle, Olivier Belmonte, Pierre Boyer, et al.

► To cite this version:

Thomas Garrigos, Manon Dollat, Arnaud Magallon, Anaïs Folletet, Julien Bador, et al.. Distribution of *Achromobacter* species in 12 French cystic fibrosis centers in 2020 by a retrospective MALDI-TOF MS spectrum analysis. *Journal of Clinical Microbiology*, 2022, 60 (6), 7 p. 10.1128/jcm.02422-21 . hal-03685833

HAL Id: hal-03685833

<https://hal.inrae.fr/hal-03685833v1>

Submitted on 13 Jun 2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Copyright



Distribution of *Achromobacter* Species in 12 French Cystic Fibrosis Centers in 2020 by a Retrospective MALDI-TOF MS Spectrum Analysis

Thomas Garrigos,^{a,b} Manon Dollat,^a Arnaud Magallon,^{a,b} Anaïs Folletet,^a Julien Badot,^{a,b} Maryam Abid,^c Marlène Amara,^e Clémence Beuruelle,^{f,g} Olivier Belmonte,^h Pierre Boyer,^{ij} Emilie Cardot-Martin,^k Anne-Gaëlle Cauchie,^e Sylvie Colin de Verdière,^l Claire Daurel,^m Cécile Gaudru,^{n,o} Farida Hamdad,^q Geneviève Héry-Arnaud,^{f,g} Baptiste Hoellinger,ⁱ Claudie Lamoureux,^{f,g} Marie-Frédérique Lartigue,^{c,d} Damasie Malandain,^m Océane Marchand,^r Caroline Piau,^s Sandrine Picot,^t Hélène Revillet,^{n,o,p} Zeina Sabouni,^r Catherine Neuwirth,^{a,b} Lucie Amoureux^{a,b}

^aDepartment of Bacteriology, University Hospital of Dijon, Dijon, France

^bUMR/CNRS 6249 Chrono-environnement, University of Bourgogne-Franche-Comté, Besançon, France

^cDepartment of Bacteriology, Virology, Hospital Hygiene, Tours University Hospital, Tours, France

^dINRAE, ISP, Tours University, Tours, France

^eDepartment of Microbiology, Hospital of Versailles, Le Chesnay, France

^fINSERM, EFS, Univ Brest, UMR 1078, GGB, Brest, France

^gDepartment of Bacteriology, Virology, Hospital Hygiene, and Parasitology-Mycology, Brest University Hospital, Boulevard Tanguy Prigent, Brest, France

^hBacteriology Laboratory, Felix-Guyon University Hospital, Saint-Denis, France

ⁱBacteriology Department, Strasbourg University Hospital, Strasbourg, France

^jUR7290 Early Bacterial Virulence, Institute of Bacteriology, ITI InnoVec, Fédération de Médecine Translationnelle de Strasbourg, University of Strasbourg, Strasbourg, France

^kDepartment of Microbiology, Foch Hospital, Suresnes, France

^lRespiratory Diseases Department and Centre de compétence de la Mucoviscidose, Foch Hospital, Suresnes, France

^mMicrobiology Department, CHU de Caen, Caen, France

ⁿDepartment of Bacteriology, Purpan University Hospital, Toulouse, France

^oIRSD, Toulouse University, INSERM, INRA, ENVT, UPS, Toulouse, France

^pObservatoire Burkholderia cepacia, Toulouse University Hospital, Toulouse/Vaincre la Mucoviscidose, Paris, France

^qDepartment of Bacteriology, Amiens University Hospital, Amiens, France

^rDepartment of Microbiology, Toulon-La Seyne-sur-Mer Hospital, Toulon, France

^sDepartment of Clinical Microbiology, Rennes University Hospital, Rennes, France

^tDepartment of Bacteriology, Virology and Parasitology, University Hospital of La Réunion, Saint Pierre, Réunion

Thomas Garrigos and Manon Dollat contributed equally to this article. Author order was determined by seniority.

ABSTRACT *Achromobacter* spp. are nonfermenting Gram-negative bacilli mainly studied among cystic fibrosis (CF) patients. The identification of the 19 species within the genus is time-consuming (*nrdA*-sequencing), thus data concerning the distribution of the species are limited to specific studies. Recently, we built a database using MALDI-TOF mass spectrometry (MS) (Bruker) that allows rapid and accurate species identification and detection of the multiresistant epidemic clones: *A. xylosoxidans* ST137 spreading among CF patients in various French and Belgium centers, and *A. ruhlandii* DES in Denmark. Here, we first assessed whether species identification could be achieved with our database solely by analysis of MS spectra without availability of isolates. Then, we conducted a multicentric study describing the distribution of *Achromobacter* species and of the clone ST137 among French CF centers. We collected and analyzed with our local database the spectra of *Achromobacter* isolates from 193 patients (528 samples) from 12 centers during 2020. In total, our approach enabled to conclude for 502/528 samples (95.1%), corresponding to 181 patients. Eleven species were detected, only five being involved in chronic colonization, *A. xylosoxidans* (86.4%), *A. insuavis* (9.1%), *A. mucicolens* (2.3%), *A. marplatensis*

Editor Nathan A. Ledebor, Medical College of Wisconsin

Copyright © 2022 American Society for Microbiology. All Rights Reserved.

Address correspondence to Lucie Amoureux, lucie.amoureux@chu-dijon.fr.

The authors declare no conflict of interest.

Received 10 December 2021

Returned for modification 11 January 2022

Accepted 18 March 2022

(1.1%) and *A. genogroup 3* (1.1%). This study confirmed the high prevalence of *A. xylosoxidans* in chronic colonizations and the circulation of the clone *A. xylosoxidans* ST137 in France: four patients in two centers. The present study is the first to report the distribution of *Achromobacter* species from CF patients samples using retrospective MALDI-TOF/MS data. This easy approach could enable future large-scale epidemiological studies.

KEYWORDS *Achromobacter*, cystic fibrosis, epidemiology, Maldi-TOF, mass spectrometry, identification, ST137, DES

A *chromobacter* spp. are Gram-negative nonfermenting bacilli frequently reported in the respiratory samples of cystic fibrosis (CF) patients. To date, 19 officially validated species can be identified on the pubMLST database (<https://pubmlst.org/achromobacter/>) within the genus (1). The distribution of the species is variable among CF centers and countries (2–14), *A. xylosoxidans* being the most prevalent one. The pathogenicity of these bacteria remains controversial in CF patients, but chronic colonization has been associated with higher rates of mortality and transplantation among these patients (15). Particular attention should be paid to the *A. xylosoxidans* ST137 clone detected so far in five French centers and three Belgian centers and the *A. ruhlandii* Danish epidemic strain (DES) responsible for epidemics in Denmark. These clones are multi-drug resistant and also responsible for chronic colonizations (7, 12, 16).

However, because of time-consuming methods of identification, epidemiological data remain scarce, and more studies are needed to help determine which species or which strains might be of clinical importance. Therefore, it is important to describe the distribution of the species and of these clones among CF patients (2–14, 17, 18). In France, the prevalence of *Achromobacter* spp. in CF patients raised from 2.7% in 2001 to 6.9% in 2019 (Registre Français de la Mucoviscidose, www.vaincrelamuco.org) and the only study reporting the different species detected was based on isolates collected in 2014 (7).

The current methods of reference for *Achromobacter* species identification are based on housekeeping gene sequencing and therefore not performed by routine diagnosis laboratories (*nrdA*-sequencing or multilocus sequence typing [MLST]) (2, 19, 20). Indeed, the databases of commercially available mass spectrometry (MS) systems do not include all the species described to date in the genus and sometimes misidentify species within the genus (21). We recently developed at Dijon center a database for accurate *Achromobacter* species identification by matrix-assisted laser desorption/ionization-time of flight mass spectrometry (MALDI-TOF/MS, Bruker Daltonics). This database allows the identification of 19 species and also the detection of the multiresistant epidemic clones *A. xylosoxidans* ST137 and *A. ruhlandii* DES (21, 22). The spectra obtained during MS analysis are automatically recorded in the instrument and the files can be exported to other laboratories using the same technology for further analysis.

This prompted us to evaluate the feasibility and reliability of a retrospective spectrum analysis in order to conduct a multicentric epidemiological study describing the distribution of the various species of *Achromobacter* in sporadic and chronic colonizations and detecting the eventual presence of the *A. xylosoxidans* ST137 and *A. ruhlandii* DES clones in various CF centers in France.

MATERIALS AND METHODS

Evaluation of the feasibility and reliability of retrospective spectrum analysis. For this purpose, we collected the spectra from all isolates (recovered from various anatomic sites, from non-CF patients and sputum from CF patients) ($n = 164$ isolates from 83 patients) identified in our clinical laboratory (Dijon) as *Achromobacter* spp. by MALDI-TOF MS (Bruker Daltonics) using Bruker database (MBT-IVD-DB-7712) during 2019. The corresponding strains had all been identified to the species level by a reference method (MLST; $n = 36$), *nrdA* gene sequencing ($n = 29$), or MS using our database from a sheep-blood agar spot ($n = 99$) (2, 19–21). The spectra were retrospectively analyzed by MALDI-TOF/MS using our database as already described (21). We deliberately chose an operator who was not involved in the construction of the database and did not have access to samples and patients information. During this first step, three situations were observed: i) the spectrum was directly “interpretable” if the first two best matches were obtained with a log score ≥ 2 with the same species, ii) directly “uninterpretable” if the log score was < 2 , and iii) interpretation was considered “uncertain” if the first two best matches were obtained with a log score ≥ 2 corresponding to two different species or to *A. ruhlandii* (because of the risk of misidentification, as already described) (21).

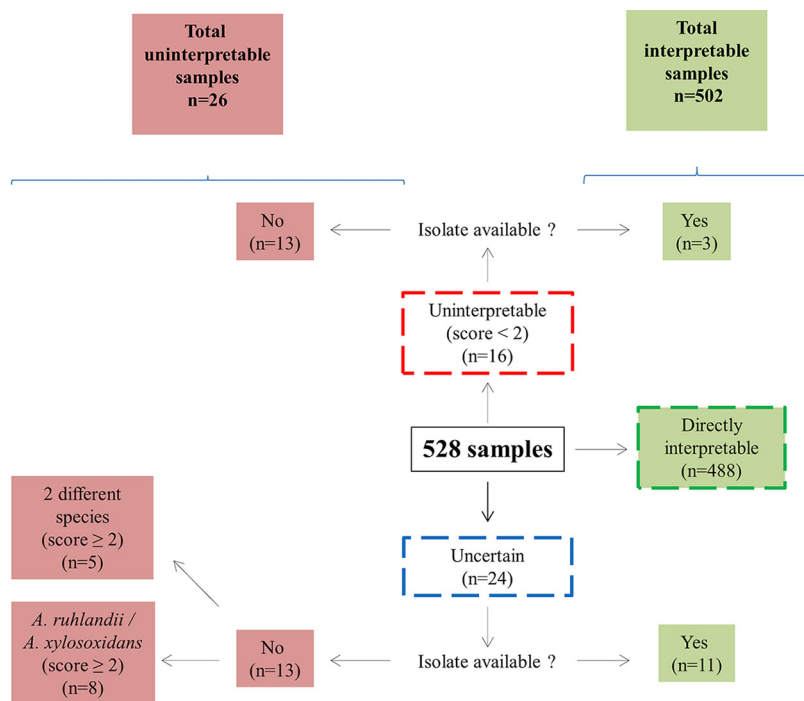


FIG 1 Analysis process of the isolates.

We were able to conclude to a reliable retrospective species identification (i.e., “directly interpretable” spectra) for 92.7% of the isolates, which allowed us to initiate the retrospective study, by collecting spectra from collaborating centers.

Application to the analysis of spectra from 12 CF centers. For this study, only laboratories using MALDI-TOF/MS (Bruker Daltonics) for routine identification according to the manufacturer’s instructions for *in vitro* diagnostics were retained: Toulouse, Suresnes-Foch, Giens, Caen, Roscoff, Versailles, La Réunion-Saint-Denis, La Réunion-Saint Pierre, Rennes, Strasbourg, Tours, and Dijon.

Each laboratory provided all the spectra matching with the genus *Achromobacter* using Bruker database from 1 January 2020 to 31 December 2020 for CF patients sputum isolates. Each center sent the spectra by email to Dijon center with the following informations: CF patient number, sample number, type of colonization (chronicity being defined according to the criteria of Pereira: “when at least three positive cultures in 1 year were obtained, with a minimum 1-month interval between them, for at least 2 years”) (17).

A total of 988 spectra corresponding to 528 samples from 193 patients were included. All the spectra were analyzed in Dijon center by MALDI-TOF/MS using our database for species level identification as already described (21). In case of uninterpretable or uncertain interpretation of the spectra, the sending laboratory was asked for the availability of the isolate. The isolates were subcultured on sheep-blood agar and re-analyzed by MALDI-TOF/MS with our database and, if not conclusive, by *nrdA* gene sequencing as already described (2). If the isolate was not available, the sample was classified as uninterpretable.

Each spectrum identified as *A. xylosoxidans* was visually analyzed in flexAnalysis software (Bruker Daltonics) as already described (22) to search for the specific absence of any peaks between *m/z* 6,640 – 6,700 for *A. xylosoxidans* ST137.

RESULTS AND DISCUSSION

The present study is the first retrospective multicentric study to describe the distribution of *Achromobacter* species directly from MALDI-TOF/MS data obtained in various laboratories. This approach has the advantage of being easier, faster, and less costly than *nrdA*-sequencing or MLST currently performed for species identification in the available epidemiological studies. It allowed the quick analysis of 988 spectra corresponding to 528 samples from 193 patients attending 12 CF centers in 2020 (Fig. 1). For 488/528 (92.6%) samples, the spectrum was directly interpretable for species identification. For 16/528 and 24/528 samples, the spectra were respectively uninterpretable or uncertain, probably mostly because of the poor quality of the original spot, and in 8 cases because of the *A. ruhlandii/A. xylosoxidans* discrimination issue (21). However, we cannot exclude that some isolates belonged to novel species not characterized yet and not included in our database. For example, in one case, the *nrdA*-

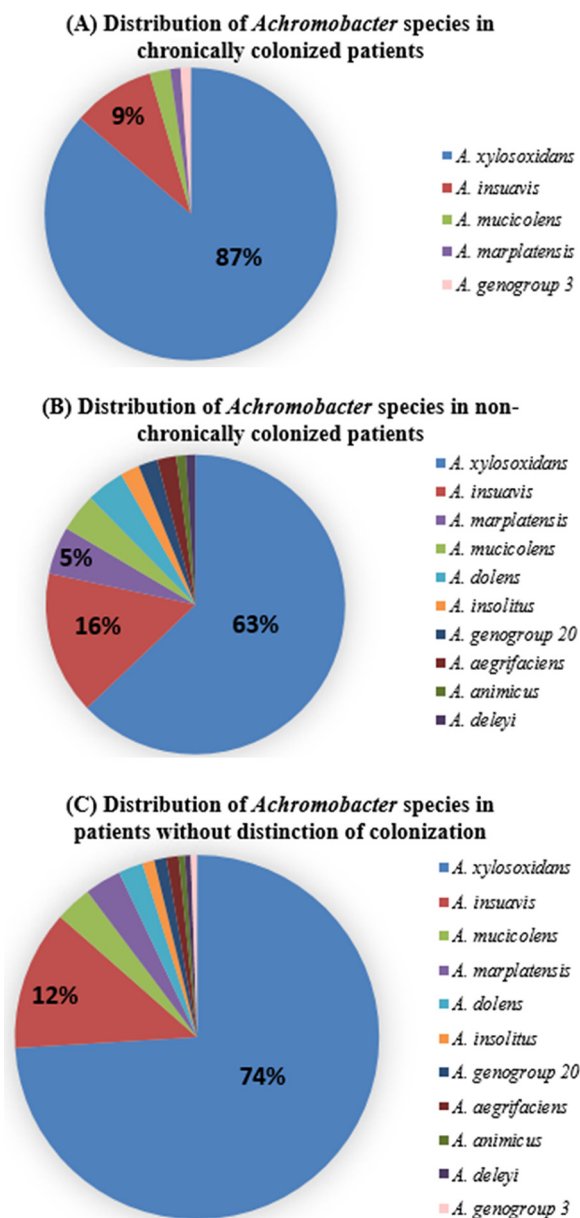


FIG 2 Distribution of *Achromobacter* species among the 181 CF patients (% of patients): (A) chronic colonized patients, (B) nonchronic colonized patients and (C) without distinction of colonization. Only values > 5% are shown.

analysis of the available isolate enabled us to identify a putative novel species belonging to genogroup 3. In total, our approach enabled to conclude for 502/528 samples (95.1%), corresponding to 181 patients (Fig. 1). The number of patients may have been slightly overestimated due to anonymized data, in the case of patients attending different centers. We always ensured the coherence of the species between the different spectra of the same sample. For example, we were able to detect, for two patients, the presence of two different species on the same sample. Overall, the prevalence of *Achromobacter* was of 7.9% and varied from 3.1 to 18.8% in the 12 CF centers.

A total of 11 species were identified, *A. xylooxidans* being the most prevalent species in each CF center (74.6% patients), as already reported (2–11, 13, 14, 23), followed by *A. insuavis* (12.1%), *A. mucicolens* (3.2%), *A. marplatensis* (3.2%), *A. dolens* (2.1%), *A. aegrifaciens* (1.1%), *A. insolitus* (1.1%), *A. genogroup 20* (1.1%) (identified by *nrdA* gene sequencing), *A. genogroup 3* (0.5%) (identified by *nrdA* gene sequencing), *A. animicus* (0.5%), and *A. deleyi* (0.5%). As

TABLE 1 Distribution of *Achromobacter* species among the patients of the 12 French centers in 2020^a

Center	<i>A. xylosoxidans</i>		<i>A. insuavis</i>		<i>A. mucicolens</i>		<i>A. maripalensis</i>		<i>A. insolitus</i>		<i>A. aegyri-faciens</i>		<i>A. geno-group 20</i>		<i>A. geno-group 3</i>		<i>A. animicus</i>		<i>A. deleyi</i>		<i>A. dolens</i>		Total patients		Total patients in each center	Achromobacter prevalence in each center (%)			
	C	NC	C	NC	C	NC	C	NC	C	NC	C	NC	C	NC	C	NC	C	NC	C	NC	T	TT							
Roscoff	8	1	0	1 ^b	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	5 ^b	13	185	7		
Caen	3	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	4	128	3.1		
Dijon	4	4	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	8	13	129	10.1		
Suresnes-Foch	13	11	2	3	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	16	14	32	508	6.3	
La Réunion	4	9	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	4	12	16	85	18.8	
St Pierre	0	7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	9	9	11	64	17.2	
St Denis	6	8	1	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	7	12	19	228	8.3	
Strasbourg	6	3	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	4	10	14	318	4.4	
Glens	10	2	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	11	2	13	17	191	8.9
Toulouse	11 ^c	3	1	7	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	14 ^{c,d}	19 ^{c,d}	31	294	10.5
Tours	10	3	1	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	11	5	16	16	223	7.2
Versailles	1	3	1	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	2	6 ^e	7	70	10	
Total	76	61	8	15	2	4	1	5	0	2	0	2	0	2	1	0	0	0	0	0	0	0	0	88	97	181	193	2423	7.9

^aC: chronic; NC: nonchronic; T: total patients of the center with *Achromobacter* identified at the species rank; TT: total patients of the center with *Achromobacter* (includes patients with isolates unidentified to the species rank).
^bOne patient chronically colonized with *A. mucicolens* had a positive sample with *A. insuavis*.
^cOne patient chronically colonized with *A. xylosoxidans* had a positive sample with *A. deleyi*.
^dOne patient chronically colonized with *A. insuavis* had a positive sample with *A. xylosoxidans*.
^eOne patient had a positive sample with *A. insolitus* and *A. aegyri-faciens*.

already noticed in our former studies in France, we did not detect any isolate belonging to *A. ruhlandii* in this study although this species is the second most frequent species reported in United States, Brazil, Argentina and Denmark. (2, 9, 11, 14) (Fig. 2A and Table 1).

Among the 181 patients, 88 (48.6%) were chronically colonized. This result is in accord with previous studies (Table S1 in the supplemental material) (5, 6, 8, 10, 12). The number of chronic patients might have been underestimated since some consultations did not take place in the year 2020 because of COVID-19. Despite the predominance of *A. xylosoxidans*, we found a greater diversity of species during nonchronic colonization than in chronic colonization (total of 10 versus 5 species) (Fig. 2B, Table 1, and Table S1 in the supplemental material), as previously reported in France, Canada, and Denmark (5, 12, 23).

Among chronically colonized patients, as expected, *A. xylosoxidans* was the most predominant species. Noteworthy, 3 species never reported to date in chronic colonization were also detected (Table S1 in the supplemental material) (4–6, 8–10, 12, 14, 23, 24): *A. genogroup 3*, *A. mucicolens*, and *A. marplatensis*.

Among the 181 patients, the epidemic clone *A. xylosoxidans* ST137 was detected in four patients from two centers: three in Foch CF center and one in Giens CF center. The presence of ST137 had not been documented yet in these patients and no patient carrying the ST137 clone was previously known in Giens center. Each time the strain was responsible for chronic colonization, and multiresistant as in previous descriptions (7, 12, 16). These data show that the clone with epidemic potential continues to spread in new centers. It should be noted that our approach allowed us to detect this clone easily and quickly, and that this method will help in patients monitoring and management of segregation measures.

In conclusion, this study showed that retrospective analyses by our MALDI-TOF/MS database of spectra collected from samples from various centers was possible and led to excellent *Achromobacter* species identification. It allowed the detection of the multiresistant epidemic clone *A. xylosoxidans* ST137. Our database is currently only available in our center (Dijon) or by contacting the corresponding author (21). These easy-to-use MALDI-TOF/MS retrospective studies could be used on a large scale for enrichment of epidemiological data concerning the distribution of *Achromobacter* species and the survey of the circulation of epidemic clones.

SUPPLEMENTAL MATERIAL

Supplemental material is available online only.

SUPPLEMENTAL FILE 1, PDF file, 0.2 MB.

REFERENCES

- Dumolin C, Peeters C, Ehsani E, Tahon G, De Canck E, Cnockaert M, Boon N, Vandamme P. 2020. *Achromobacter veterisilvae* sp. nov., from a mixed hydrogen-oxidizing bacteria enrichment reactor for microbial protein production. *Int J Syst Evol Microbiol* 70:530–536. <https://doi.org/10.1099/ijsem.0.003786>.
- Spilker T, Vandamme P, LiPuma JJ. 2013. Identification and distribution of *Achromobacter* species in cystic fibrosis. *J Cyst Fibros* 12:298–301. <https://doi.org/10.1016/j.jcf.2012.10.002>.
- Coward A, Kenna DTD, Perry C, Martin K, Doumith M, Turton JF. 2016. Use of *nrdA* gene sequence clustering to estimate the prevalence of different *Achromobacter* species among cystic fibrosis patients in the UK. *J Cyst Fibros* 15:479–485. <https://doi.org/10.1016/j.jcf.2015.09.005>.
- Coward A, Kenna DTD, Woodford N, Turton JF, Armstrong M, Auckland C, Bowler I, Burns P, Cargill J, Carroll M, Flight W, Graver M, Green H, Horner C, Jones A, Jones AM, Jones G, Mayell S, Orendi J, Perry A, Robb A, Tucker N, Waine D, Winstanley T, Withers N, and members of the UK CF Surveillance Working Group., The UK CF Surveillance Working Group comprised. 2020. Structured surveillance of *Achromobacter*, *Pandoraea* and *Ralstonia* species from patients in England with cystic fibrosis. *J Cyst Fibros* 19: 388–393. <https://doi.org/10.1016/j.jcf.2019.11.005>.
- Edwards BD, Greyson-Wong J, Somayaji R, Waddell B, Whelan FJ, Storey DG, Rabin HR, Surette MG, Parkins MD. 2017. Prevalence and outcomes of *Achromobacter* species infections in adults with cystic fibrosis: a North American cohort study. *J Clin Microbiol* 55:2074–2085. <https://doi.org/10.1128/JCM.02556-16>.
- Veschetti L, Sandri A, Patuzzo C, Melotti P, Malerba G, Lleo MMY. 2021. Genomic characterization of *Achromobacter* species isolates from chronic and occasional lung infection in cystic fibrosis patients. *Microb Genom* 7:000606.
- Amoureux L, Sauge J, Sarret B, Lhoumeau M, Bajard A, Tetu J, Bador J, Neuwirth C, Caillon J, Cardot-Martin E, Cattoir V, Doléans-Jordheim A, Ferroni A, Guet-Revillet H, Héry-Arnaud G, Segonds C, Thomas E, Plésiat P, Vu-Thien H, MucoMicrobes group. 2019. Study of 109 *Achromobacter* spp. isolates from 9 French CF centres reveals the circulation of a multiresistant clone of *A. xylosoxidans* belonging to ST 137. *J Cyst Fibros* 18:804–807. <https://doi.org/10.1016/j.jcf.2019.04.005>.
- Barrado L, Brañas P, Orellana MÁ, Martínez MT, García G, Otero JR, Chaves F. 2013. Molecular characterization of *Achromobacter* isolates from cystic fibrosis and non-cystic fibrosis patients in Madrid, Spain. *J Clin Microbiol* 51:1927–1930. <https://doi.org/10.1128/JCM.00494-13>.
- Pereira RHV, Leão RS, Carvalho-Assef AP, Albano RM, Rodrigues ERA, Firmida MC, Folescu TW, Plotkowski MC, Bernardo VG, Marques EA. 2017. Patterns of virulence factor expression and antimicrobial resistance in *Achromobacter xylosoxidans* and *Achromobacter ruhlandii* isolates from patients with cystic fibrosis. *Epidemiol Infect* 145:600–606. <https://doi.org/10.1017/S0950268816002624>.
- Filipic B, Malešević M, Vasiljević Z, Lukic J, Novovic K, Kojic M, Jovcic B. 2017. Uncovering differences in virulence markers associated with *Achromobacter* species of CF and non-CF origin. *Front Cell Infect Microbiol* 7: 224. <https://doi.org/10.3389/fcimb.2017.00224>.
- Papalia M, Steffanowski C, Traglia G, Almuzara M, Martina P, Galanternik L, Vay C, Gutkind G, Ramírez MS, Radice M. 2020. Diversity of *Achromobacter*

- species recovered from patients with cystic fibrosis, in Argentina. *Rev Argent Microbiol* 52:13–18. <https://doi.org/10.1016/j.ram.2019.03.004>.
12. Gade SS, Nørskov-Lauritsen N, Ridderberg W. 2017. Prevalence and species distribution of *Achromobacter* sp. cultured from cystic fibrosis patients attending the Aarhus centre in Denmark. *J Med Microbiol* 66: 686–689. <https://doi.org/10.1099/jmm.0.000499>.
 13. Rodrigues ERA, Ferreira AG, Leão RS, Leite CCF, Carvalho-Assef AP, Albano RM, Marques EA. 2015. Characterization of *Achromobacter* species in cystic fibrosis patients: comparison of *bla*OXA-114 PCR amplification, multilocus sequence typing, and matrix-assisted laser desorption ionization–time of flight mass spectrometry. *J Clin Microbiol* 53:3894–3896. <https://doi.org/10.1128/JCM.02197-15>.
 14. Gabrielaite M, Bartell JA, Nørskov-Lauritsen N, Pressler T, Nielsen FC, Johansen HK, Marvig RL. 2021. Transmission and antibiotic resistance of *Achromobacter* in cystic fibrosis. *J Clin Microbiol* 59:e02911–20. <https://doi.org/10.1128/JCM.02911-20>.
 15. Somayaji R, Stanojevic S, Tullis DE, Stephenson AL, Ratjen F, Waters V. 2017. Clinical outcomes associated with *Achromobacter* species infection in patients with cystic fibrosis. *Ann Am Thorac Soc* 14:1412–1418. <https://doi.org/10.1513/AnnalsATS.201701-071OC>.
 16. Cools P, Ho E, Vranckx K, Schelstraete P, Wurth B, Franckx H, Ieven G, Van Simaey L, Van Daele S, Verhulst S, De Baets F, Vaneechoutte M. 2016. Epidemic *Achromobacter xylosoxidans* strain among Belgian cystic fibrosis patients and review of literature. *BMC Microbiol* 16:122. <https://doi.org/10.1186/s12866-016-0736-1>.
 17. Pereira RHV, Carvalho-Assef AP, Albano RM, Folescu TW, Jones MCMF, Leão RS, Marques EA. 2011. *Achromobacter xylosoxidans*: characterization of strains in Brazilian cystic fibrosis patients. *J Clin Microbiol* 49:3649–3651. <https://doi.org/10.1128/JCM.05283-11>.
 18. Voronina OL, Kunda MS, Ryzhova NN, Aksenova EI, Sharapova NE, Semenov AN, Amelina EL, Chuchalin AG, Gintsburg AL. 2018. On Burkholderiales order microorganisms and cystic fibrosis in Russia. *BMC Genomics* 19:74. <https://doi.org/10.1186/s12864-018-4472-9>.
 19. Ridderberg W, Wang M, Nørskov-Lauritsen N. 2012. Multilocus sequence analysis of isolates of *Achromobacter* from patients with cystic fibrosis reveals infecting species other than *Achromobacter xylosoxidans*. *J Clin Microbiol* 50:2688–2694. <https://doi.org/10.1128/JCM.00728-12>.
 20. Spilker T, Vandamme P, LiPuma JJ. 2012. A multilocus sequence typing scheme implies population structure and reveals several putative novel *Achromobacter* species. *J Clin Microbiol* 50:3010–3015. <https://doi.org/10.1128/JCM.00814-12>.
 21. Garrigos T, Neuwirth C, Chapuis A, Bador J, Amoureux L, Andre E, Barbier E, Caillon J, Cardot ME, Cattoir V, Doléans JA, Echahidi F, Ferroni A, Guet RH, Héry AG, Lipuma J, Nørskov LN, Peeters C, Pierard D, Segonds C, Thomas E, Plésiat P, Vandamme P, Verroken A, Vu TH, Collaborators. 2021. Development of a database for the rapid and accurate routine identification of *Achromobacter* species by matrix-assisted laser desorption/ionization–time-of-flight mass spectrometry (MALDI-TOF MS). *Clin Microbiol Infect* 27:126.e1–126.e5. <https://doi.org/10.1016/j.cmi.2020.03.031>.
 22. Garrigos T, Dollat M, Magallon A, Chapuis A, Varin V, Bador J, Makki N, Cremet L, Persyn E, Cardot-Martin E, Echahidi F, Peeters C, Pierard D, Vandamme P, Verroken A, Neuwirth C, Amoureux L. 2021. Matrix-assisted laser desorption ionization-time of flight mass spectrometry for rapid detection of isolates belonging to the epidemic clones *Achromobacter xylosoxidans* ST137 and *Achromobacter ruhlandii* DES from cystic fibrosis patients. *J Clin Microbiol* 59:e0094621. <https://doi.org/10.1128/JCM.00946-21>.
 23. Amoureux L, Bador J, Bounoua Zouak F, Chapuis A, de Curraize C, Neuwirth C. 2016. Distribution of the species of *Achromobacter* in a French cystic fibrosis centre and multilocus sequence typing analysis reveal the predominance of *A. xylosoxidans* and clonal relationships between some clinical and environmental isolates. *J Cyst Fibros* 15:486–494. <https://doi.org/10.1016/j.jcf.2015.12.009>.
 24. Dupont C, Michon A-L, Jumas-Bilak E, Nørskov-Lauritsen N, Chiron R, Marchand H. 2015. Inpatient diversity of *Achromobacter* spp. involved in chronic colonization of Cystic Fibrosis airways. *Infect Genet Evol* 32:214–223. <https://doi.org/10.1016/j.meegid.2015.03.012>.