

Fermentation of Chlorella vulgaris, a microalga used in human food, by 89 strains of lactic bacteria

Victoria Chuat, Valérie Gagnaire, Piot Carmine, Maeva Subileau, Claire Bourlieu-Lacanal, Florence Valence

▶ To cite this version:

Victoria Chuat, Valérie Gagnaire, Piot Carmine, Maeva Subileau, Claire Bourlieu-Lacanal, et al.. Fermentation of Chlorella vulgaris, a microalga used in human food, by 89 strains of lactic bacteria. 23ème édition du colloque du Club des Bactéries Lactiques, Jun 2022, Rennes, France. , 2022. hal-03695703

HAL Id: hal-03695703 https://hal.inrae.fr/hal-03695703

Submitted on 15 Jun2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial - NoDerivatives 4.0 International License

RÉPUBLIQUE RÉPUBLIQUE FRANÇAISE Liberté Égalité Fraternité



Fermentation of *Chlorella vulgaris*, a microalga used in human food, by 89 strains of lactic bacteria.

Victoria Chuat¹, Valérie Gagnaire¹, Carmine Piot¹, Maeva Subileau², Claire Bourlieu², Florence Valence¹

- ¹ UMR STLO, INRAE, Institut Agro, F35000, Rennes, France.
- ² UMR IATE, Univ Montpellier, INRAE, Institut Agro, F34000, Montpellier, France.

CONTEXT & AIM

- Microalgua are photosynthetic unicellular organisms rich in nutriments, constituting a biomass with high potential for food applications.
- Their production is energy consuming with a high risk of alteration by spoilage flora during the process. Fermentation, an ancestral preservation process, offers an alternative to current processes.
- ✓ By degrading the sugars naturally present in the matrix, lactic acid bacteria (LAB) produce organic acids, thus lowering the pH, which helps to inhibit pathogenic and spoilage flora when the pH reaches values around 4.6.
- ✓ In this study, we screened the ability of 89 LAB strains (10 species) to ferment *Chlorella vulgaris*, microalgae approved for human consumption.

STRATEGY Table 1 : Number of tested LAB strains by species

Levilactobacillus brevis2Pediococcus pentosaceus2Lactiplantibacillus pentosus5Lactiplantibacillus paraplantarum5Lactiplantibacillus plantarum1Lacticaseibacillus paracasei5Loigolactobacillus coryniformis2	 -
Pediococcus pentosaceus4Lactiplantibacillus pentosus5Lactiplantibacillus paraplantarum6Lactiplantibacillus plantarum1Lacticaseibacillus paracasei5Loigolactobacillus coryniformis2	-
Lactiplantibacillus pentosusSLactiplantibacillus paraplantarumSLactiplantibacillus plantarum1Lacticaseibacillus paracaseiSLoigolactobacillus coryniformisS)
Lactiplantibacillus paraplantarum3Lactiplantibacillus plantarum1Lacticaseibacillus paracasei3Loigolactobacillus coryniformis2	CONTRACTOR CONTRACTOR CONTRACTOR CONTRACTOR
Lactiplantibacillus plantarum1Lacticaseibacillus paracasei5Loigolactobacillus coryniformis2)
Lacticaseibacillus paracasei	4
Loigolactobacillus coryniformis	,)
) -
Leuconostoc mesenterolaes cremoris)
Fructilactobacillus fructivorans	-
Lactococcus lactis	,
TOTAL 8	•









ESULTS	ABILITY OF 89 LAB STRAINS TO FERMENT CHLORELLA VULGARIS
7 T0 pH = 6.57	
~	
6	
$5 \xrightarrow{7} 7 \xrightarrow{7} \overrightarrow{7} \overrightarrow{7} \xrightarrow{7} \overrightarrow{7} \overrightarrow{7} \overrightarrow{7} \overrightarrow{7} \xrightarrow{7} \overrightarrow{7} $	
4 6 6 7 7 7 7 7 7 8 8 8 8 8 8 8 6 7 8 8 8 8	8 7 7 7 7 7 8 8 8 7 9 8 8 8 6 7 9 7 8 8 7 8 9 1 7 8 8 7 8 9 1 7 8 8 8 8 8 8 7 8 9 1 7 8 8 8 8 9 7 8 7 8 7 8 7 8 8 8 8 7 8 9 7 8 8 8 8
L. lactis 24 L. lactis 24 L. lactis 24 L. lactis 24 L. lactis 24 L. lactis 24 L. lactis 25 L. lactis 21 antarum 22 antarum 21 pentosus 21 pertosus 21 ventosus 21 antarum 23 antarum 23 antarum 23 antarum 23 antarum 23 antarum 23 antarum 23 antarum 23 antarum 23	 porocase i 2. L. brevis 15. L. brevis 15. L. brevis 15. L. brevis 22. antarum 14. vorans 214. voranum 215. voranum 216. voranum 216.
שליביש לאינישי רובישרי בישריביים רובישרי בישריביים אונים אונים אונים אונים אוניים אוניים אוניים אוניים אוניים אוניים אוניים אוני	F. P.

- > A pH decrease was observed for the majority of the strains (72%) at 12h. After 12h of fermentation, the lowest pH reached 4.7 with the strain Lactococcus lactis CIRM-BIA2469.
- > The fermentation profiles of the 89 strains were divided into three « acidification groups »:
 - group of strains which finished acidifying at 12h (60% of the strains);
 - group of strains which finished acidifying at 36h (12% of the strains);
 - group of non-acidifying strains with no acidification at 36h (28% of the strains).
- > Lactococcus lactis, Lactiplantibacillus plantarum and Lactiplantibacillus pentosus were the most acidifying species.
- > A pH decrease was observed for the unfermented microalgal biomass used as the control, 5.78±0.37 (red arrow) suggesting the development of a native spoilage flora.



- A native flora was present in the suspension of Chlorella vulgaris around 3 log₁₀ CFU/ml at TO, enumerated on BHI-YE.
- > The spoilage flora was identified by sequencing of DNAr16S as **Bacillus sp.**
- The optimization was performed on the three best strains from the screening belonging to three different species (*L. lactis, L. plantarum* and *L. pentosus*).
- Addition of glucose enhanced the acidification rate for the three strains used. The effect was dependent on the concentration of glucose. Addition of 2g of glucose allowed to reach the targeted pH of 4.5 for all strains.
- After 24h of incubation at 30°C, the concentration of the native flora reached 8 log₁₀ CFU/ml in the non-inoculated suspension.
- When the algal suspension was inoculated with LAB, the spoilage flora was not enumerated on BHI-YE (data not shown).
- The addition of yeast extract combined with glucose did not increase the acidification rate compared to glucose alone, whatever the strains used.

CONCLUSION & PERSPECTIVES

- We confirmed the ability of LAB to ferment a freeze-dried Chlorella vulgaris biomass. To our knowledge, this is the first study of microalgae fermentation by such a large number of LAB strains.
 The LAB were able to inhibit the native spoilage flora, identified as the genus Bacillus sp..
- ✓ The addition of glucose allowed to optimize the acidification rate with a targeted pH around 4.5 after 24h of fermentation.
- Work is in progress to extend the screening to other freeze-dried microalgae species as well as to fresh microalgae supensions.

Centre Bretagne - Normandie



65 rue de Saint Brieuc 35 042 Rennes Cedex cirm-bia@inrae.fr