



The fate of milk proteins and bioactive peptides in the gastrointestinal tract during digestion

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The fate of milk proteins and bioactive peptides in the gastrointestinal tract during digestion

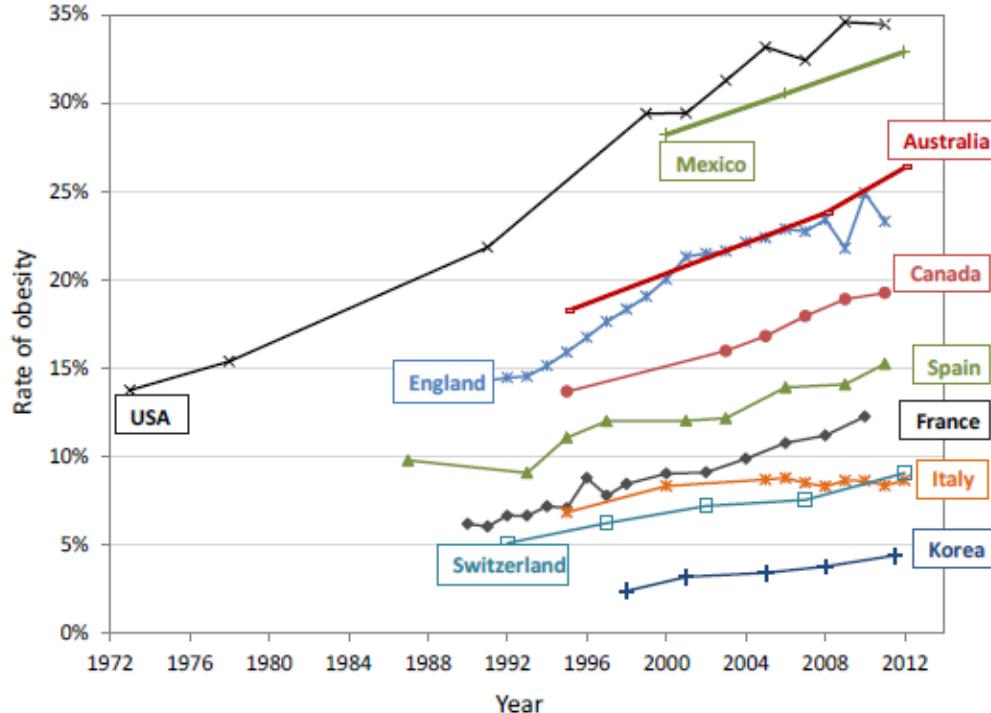
Didier Dupont

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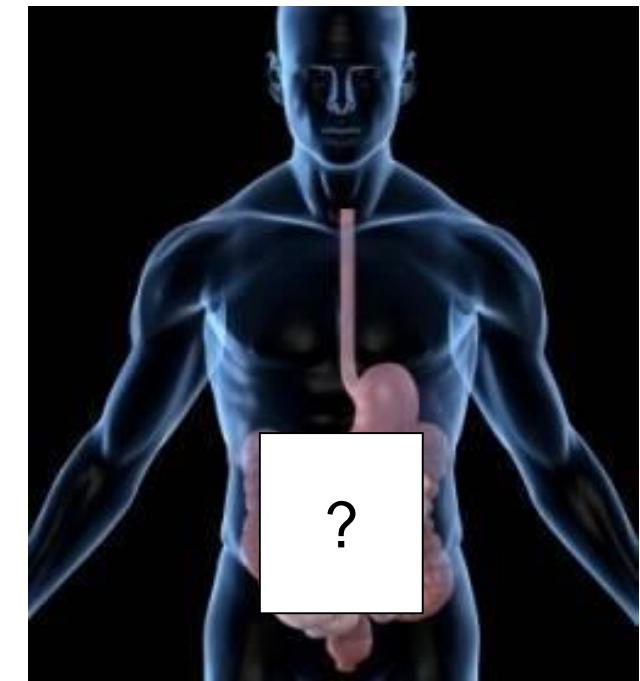


INRAE

Food and human health: the key role of the digestive process



Diet-related diseases ↑
→ Prevent these pathologies rather than cure them



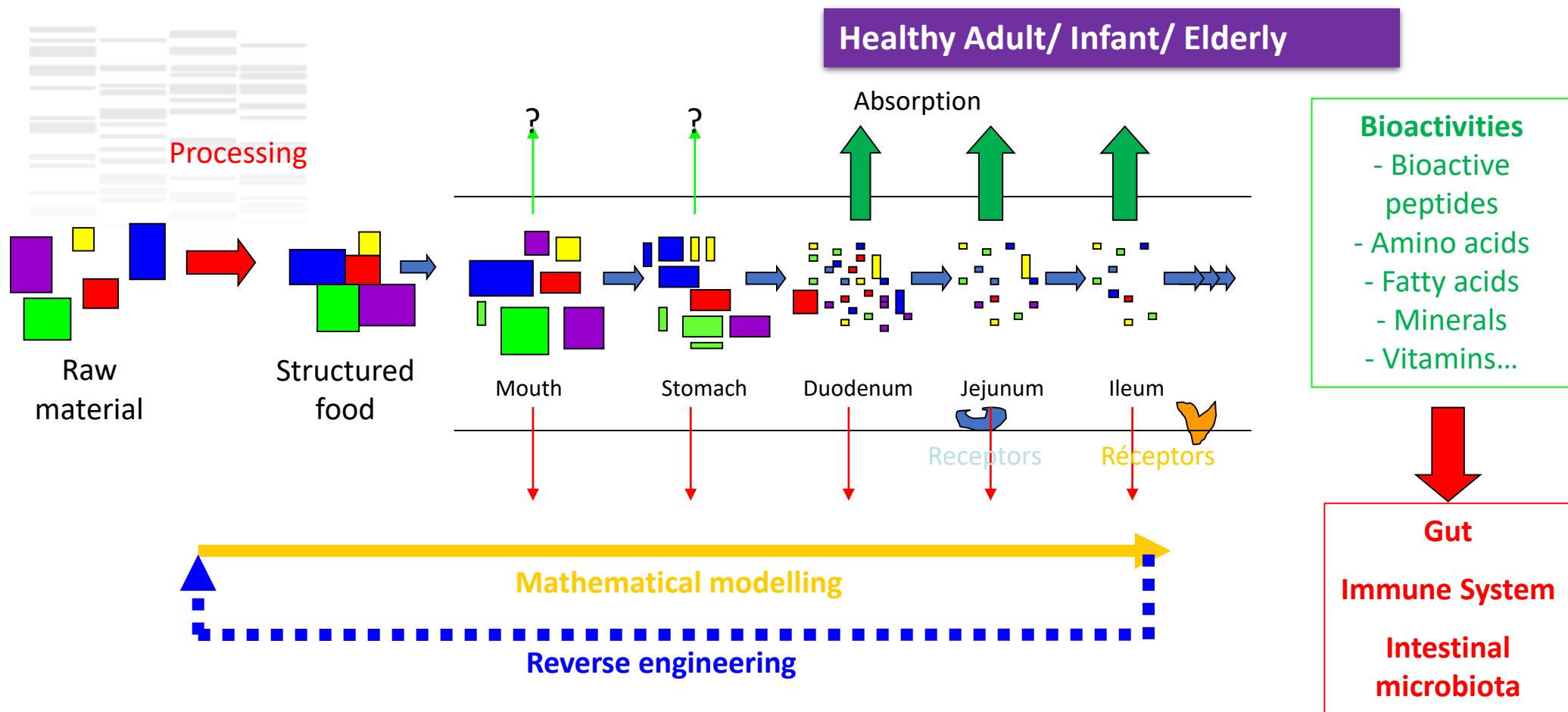
Gut = interface between food and human body

Digestion provides nutrients to the body and releases food components that can have a beneficial or a deleterious effect on human health

... but the mechanisms of food disintegration in the gastrointestinal tract remain unclear and the digestive process has been considered as a black box so far

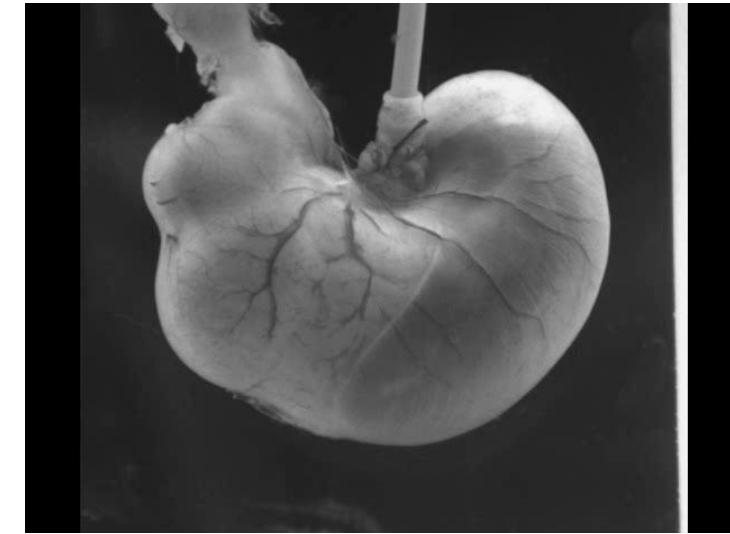
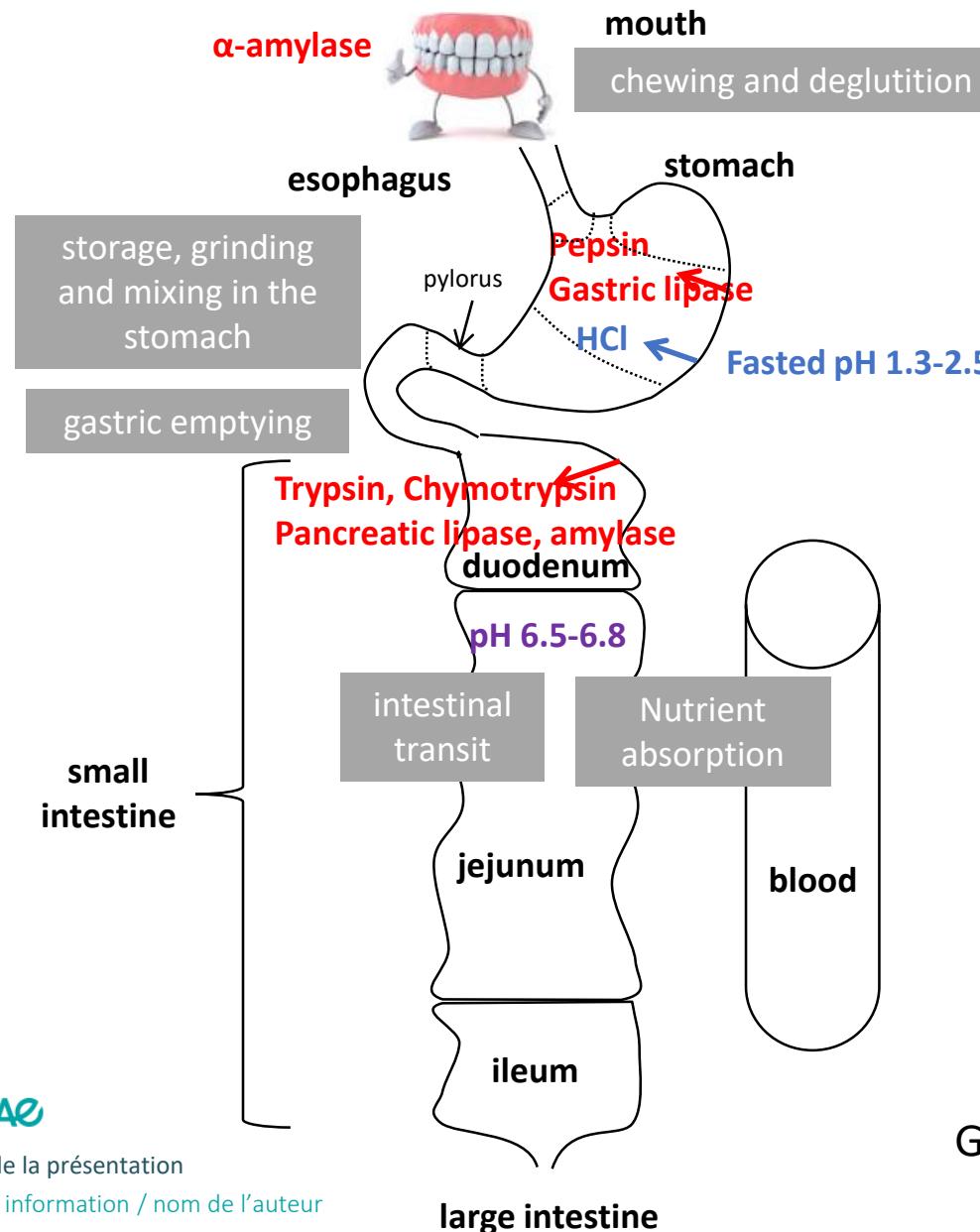
Increasing our knowledge on food digestion to increase our knowledge on the effect of food on human health

Our goals

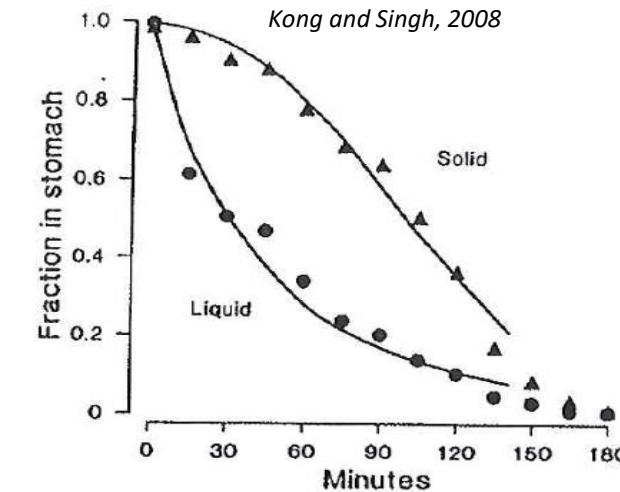


- >To understand the mechanisms of breakdown of food matrices and their constituents in the gut and identify the beneficial/deleterious food components released during digestion
- To determine the impact of the structure of food matrices on nutrient bioavailability
- To model these phenomena in order to develop a reverse engineering approach

The digestive process



From Roger Lentle, Massey Univ. NZ



Gastric phase = a very complex but crucial step for the whole digestion process

Models available at INRAE for simulating digestion

Brodkorb et al. 2019, Nat Protocols
Menard et al. 2018, Food Chem



Le Feunteun et al. 2014
Food Bioprocess Tech

$$\Phi_{12} = k_{12\text{whey}} \times (V_1 - m_{caswpd1} \times \alpha) + k_{12\text{aggr}} \times m_{caswpd1} \times \alpha$$

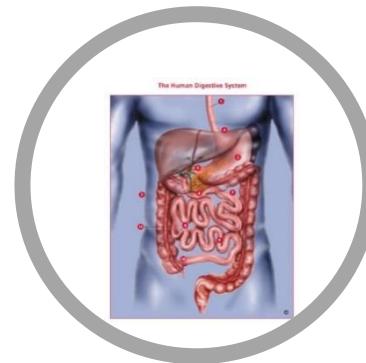
In silico models

In vitro static models
(infant, adult, elderly)

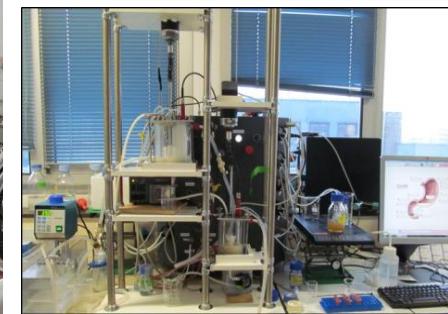
De Oliveira et al. 2016
Am J Clin Nutr
De Oliveira et al. 2017
Clin Nutr



Human models



Animal models



Menard et al. 2014, Food Chem
Sanchez et al. 2015 Food Res Int
Leroux et al. 2020, Foods
Reynaud et al. 2021 Food Chem

Barbé et al. 2013, 2014 Food Chem
Le Huerou-Luron et al. 2016 Eur J Nutr
Lorieau et al 2019 Food Chem
Nau et al. 2019, 2022 Food Chem

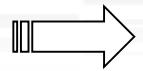


Does the food structure affect the release
of bioactive peptides during digestion?



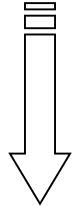
The food matrices

Ultra Low Heat powder



rehydration in water 14.5%

unheated milk (“raw” milk)



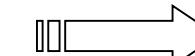
heat treatment 90°C-10 min

microstructure

heated milk

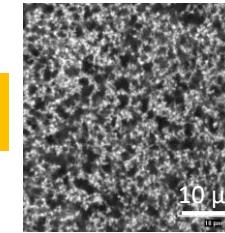


macrostructure

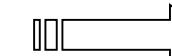


rennet gel

24h-20°C,
rennet 0.003 % v/w

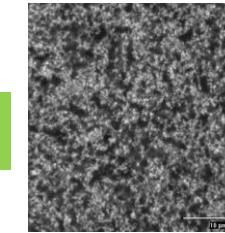


pH 6.6



rennet gel

24h-20°C,
rennet 0.3 % v/w

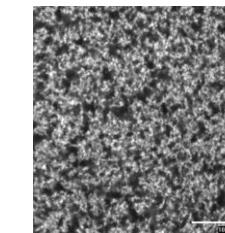


pH 6.6



acid gel

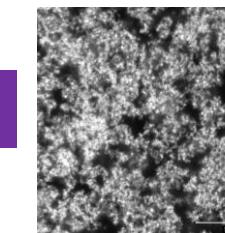
24h-20°C,
GDL 3 % w/w



pH 4

24h-20°C,
GDL 3 % w/w +
mixer 2 min

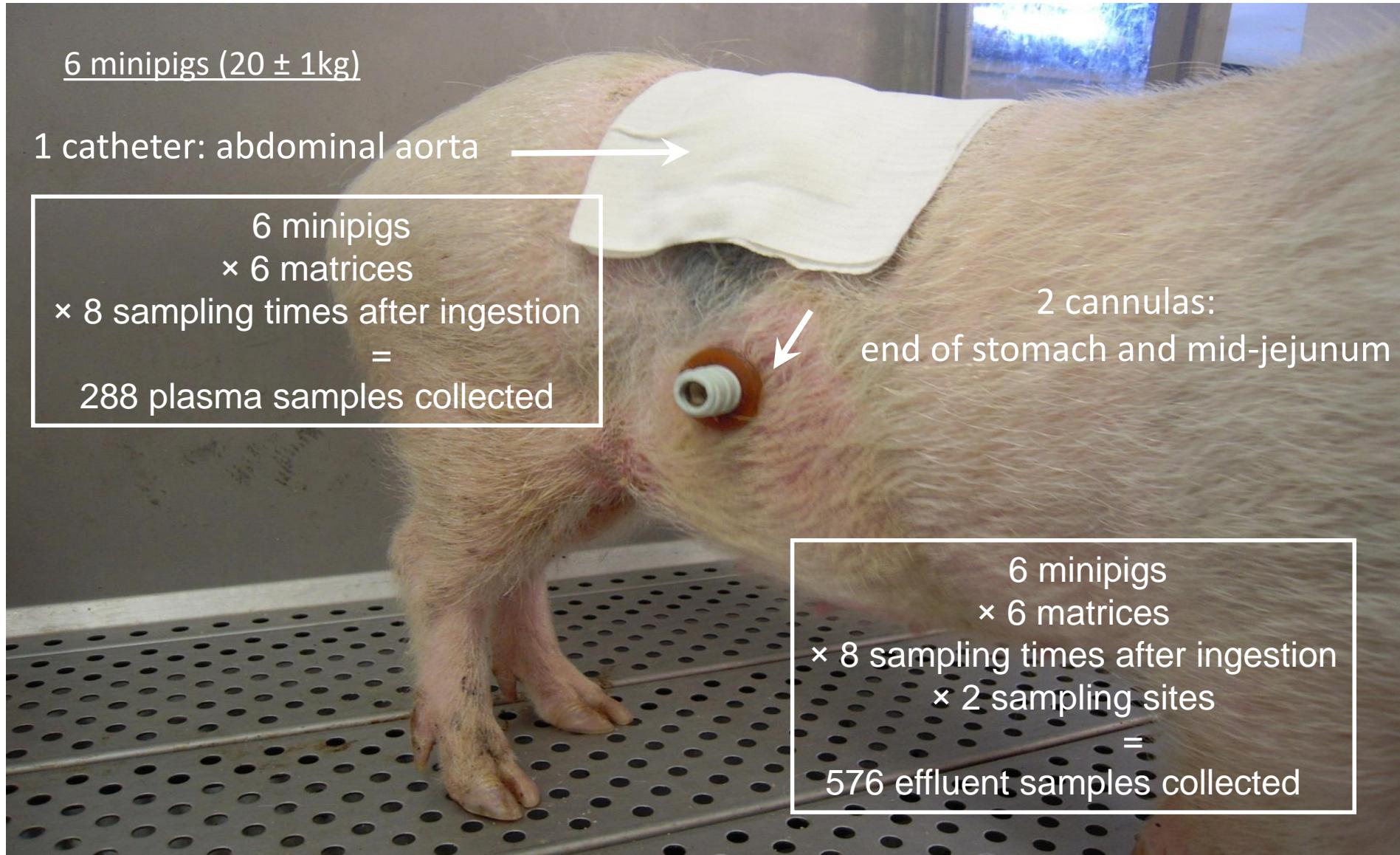
stirred acid gel



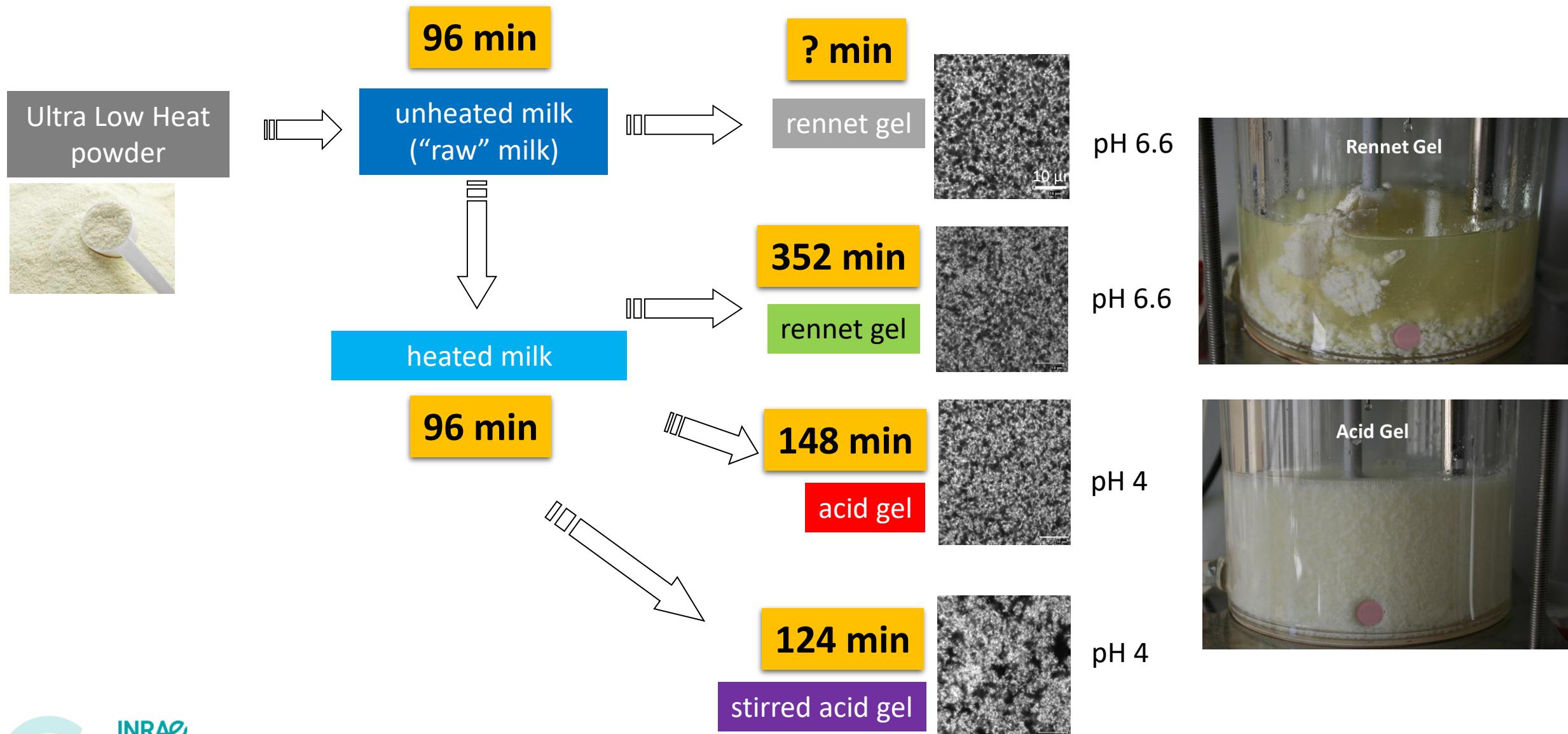
pH 4

Fat-free matrices: 40 g/L caseins, 10 g/L whey proteins, 95 g/L lactose and minerals
+ marker of the meal transit (Cr^{2+} -EDTA) → Gastric emptying half-time

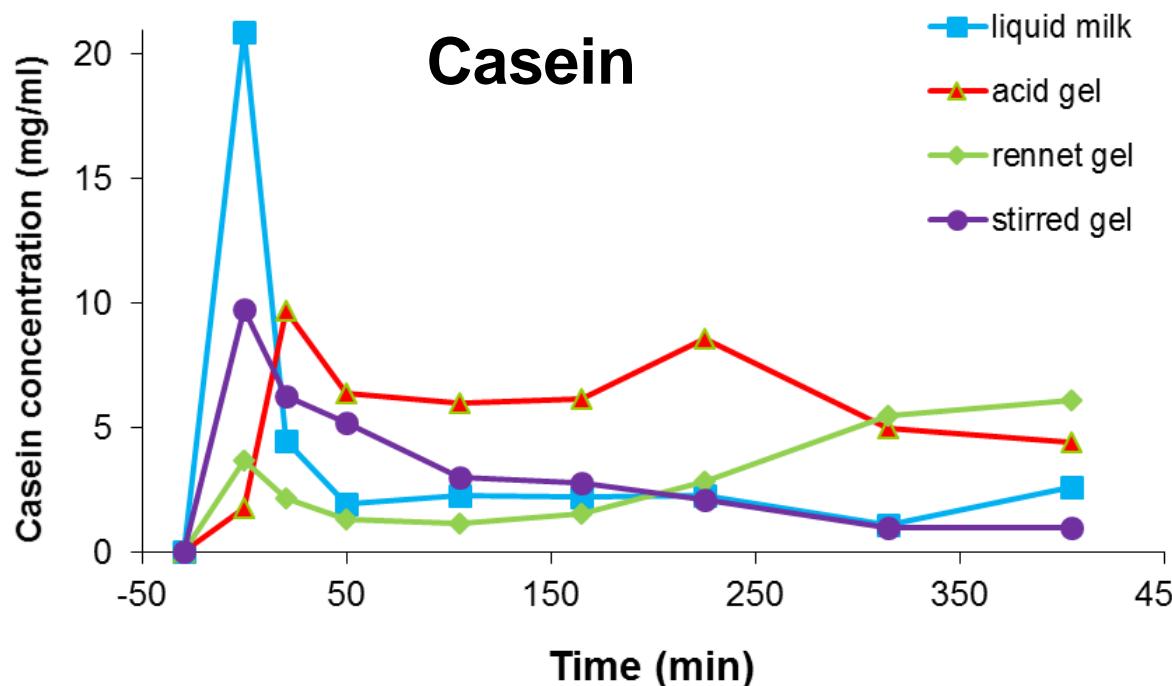
The multi-canulated mini-pigs



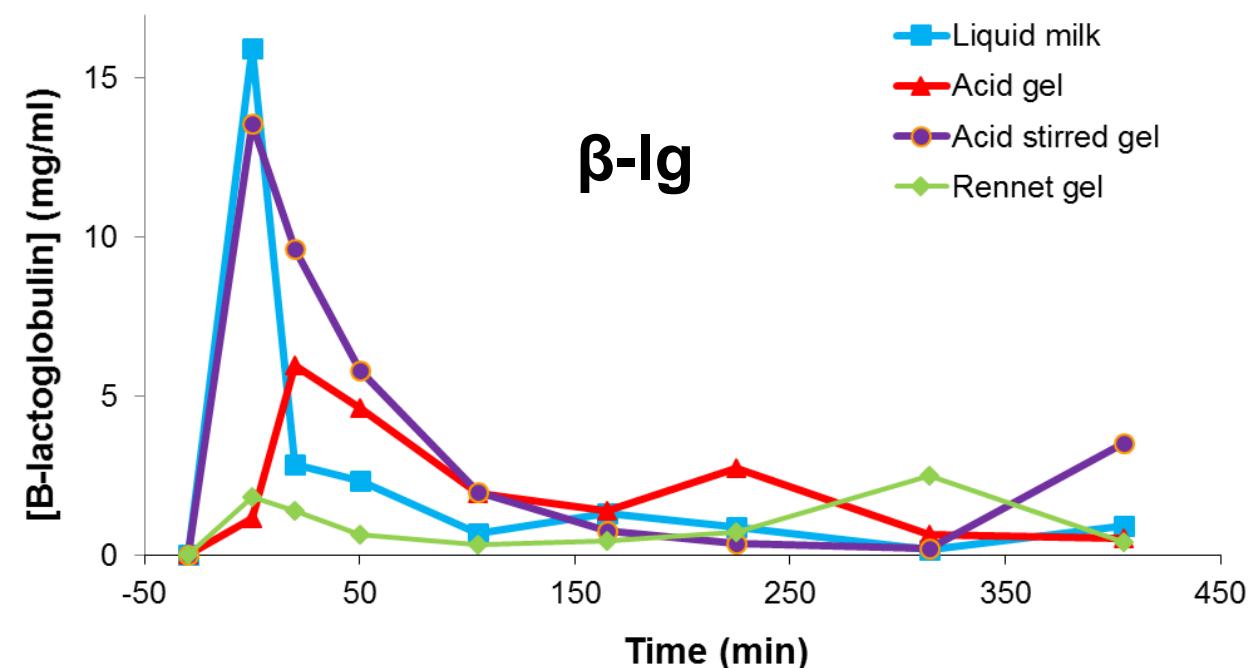
Impact of food structure on gastric emptying half-time



Concentration of Milk proteins in the duodenum (ELISA)

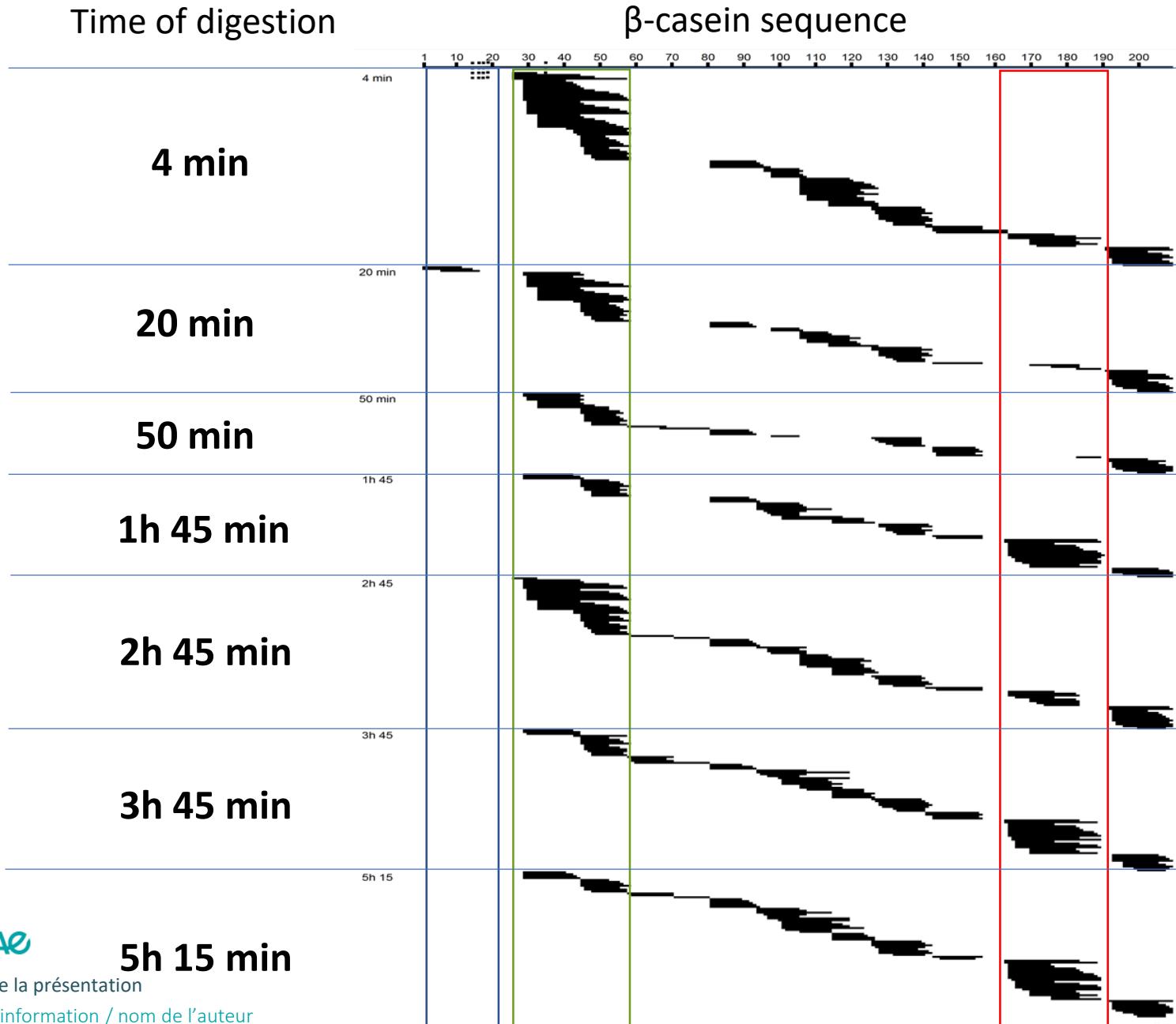


- Intense and early peak with milk/ lower and delayed with gels
- Intermediate behaviour with stirred gel
- Low concentrations with rennet gel but casein release tends to increase over time



- Only traces of milk proteins found in the jejunum
- Dairy products remain highly digestible

β -casein peptides released in the duodenum during digestion of acid gels



More than 16,000 milk peptides identified in the gastrointestinal tract of pigs

Bioactive peptides released during digestion differ from one matrix to another

Protein	Sequence	Activity	Reference	4	20	50	105	165	225	315
αs1	1-23	EMUL	Shimizu et al. (1984)							
αs1	23-34	HYP	Maruyama & Suzuki (1982)							
αs1	30-45	MB	Meisel et al. (1991)							
αs1	40-52	MB	Adamson & Reynolds (1996)							
αs1	43-58	MB	Meisel et al. (1991)							
αs1	91-100	STRE	Miclo et al. (2001)							
αs1	99-109	MIC	McCann et al. (2006)							
αs1	167-180	MIC	Hayes et al. (2006)							
αs1	180-193	MIC	Hayes et al. (2006)							
αs2	1-24	MB	Miquel et al. (2005)							
αs2	124-146	MB	Miquel et al. (2005)							
αs2	183-206	TRAN	Kizawa et al. (1996)							
αs2	183-207	MIC	Recio & Visser (1999)							
αs2	189-197	HYP	Maeno et al. (1996)							
αs2	190-197	HYP	Maeno et al. (1996)							
β	1-24	MB	Bouhallab et al. (1999)							
β	33-52	MB	Miquel et al. (2005)							
β	60-80	OPI	Jinsmaa & Yoshikawa (1999)							
β	98-105	OXI	Rival et al. (2001)							
β	114-119	OPI	Jinsmaa & Yoshikawa (1999)							
β	132-140	HYP	Robert et al. (2004)							
β	192-209	IMM	Coste et al. (1992)							
β	193-202	IMM	Kayser & Meisel (1996)							
β	193-209	IMM	Coste et al. (1992)							
κ	18-24	HYP	Lopez-Exposito et al. (2007)							
κ	106-116	THR	Jolles et al. (1986)							
β-Ig	32-40	HYP	Pihlanto-Leppala et al. (2000)							
β-Ig	92-100	MIC	Pellegrini et al. (2001)							
β-Ig	142-148	HYP	Mullally et al. (1997)							

Acid Gel

Protein	Sequence	Activity	Reference	4	20	50	105	165	225	315
αs1	40-52	MB	Adamson & Reynolds (1996)							
αs1	43-58	MB	Meisel et al. (1991)							
αs1	99-109	MIC	McCann et al. (2006)							
αs1	167-180	MIC	Hayes et al. (2006)							
αs1	180-193	MIC	Hayes et al. (2006)							
αs2	1-24	MB	Miquel et al. (2005)							
αs2	189-197	HYP	Maeno et al. (1996)							
β	33-52	MB	Miquel et al. (2005)							
β	166-175	HYP	Hayes et al. (2007)							
β	193-202	IMM	Kayser & Meisel (1996) (8))							
β-Ig	92-100	MIC	Kayser & Meisel (1996) (9))							
β-Ig	142-148	HYP								

Rennet Gel

- More bioactive peptides identified during digestion of acid gel than rennet gel
- Nature of peptides is identical (clearly defined by the digestive enzyme specificity)
 - Kinetics of release are different

Barbé et al. 2014
Food Res Int

INRAe

Titre de la présentation
Date / information / nom de l'auteur

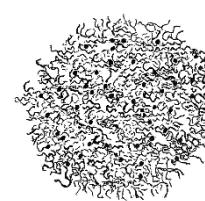
The intestinal peptidome highly depends on food structure



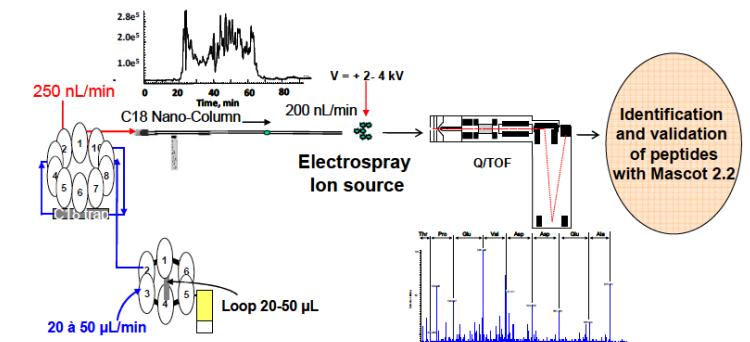
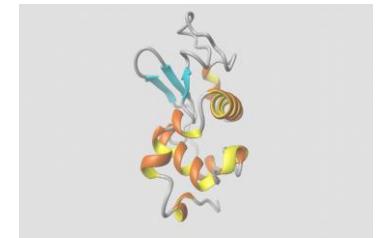
Characterization of bioactive peptides released in the small intestine of human volunteers

> *In vivo* digestion of milk proteins

- 16 subjects were fed with 30g of either:
 - Caseins
 - Whey proteins
- Intestinal contents were collected through a double lumen nasogastric tube placed in their jejunum during 6 h after ingestion (location of the sampling site was controlled by radiography)
- Effluents were then freeze-dried until further use. Before analysis, effluents were rehydrated in 50 mM Tris-HCl buffer pH 8.0 and 2M urea, centrifuged for 10 min at 2000 g
- Samples were characterized by ESI-MS-MS. Only peptides larger than 5 amino acids are considered



/



Example of data collected

prot hit num	prot desc	prot cover	pep exp mr	start	peptide	end	score	res before	pep seq	res after
1	18KD Betalg D bovine	26,5	724,3362	94	94-99	99	25,15	L	VLDTDY	K
1	18KD Betalg D bovine	26,5	771,3746	50	50-56	56	39,24	T	PEGDLEI	L
1	18KD Betalg D bovine	26,5	837,42	93	93-99	99	26,65	V	LVLDTDY	K
1	18KD Betalg D bovine	26,5	1116,4984	125	125-134	134	73,04	R	TPEVVDDEALE	K
1	18KD Betalg D bovine	26,5	1203,6312	42	42-51	51	42,99	V	YVEQLKPTPE	G
1	18KD Betalg D bovine	26,5	1212,6165	43	43-53	53	55,16	Y	VEQLKPTPEGD	L
1	18KD Betalg D bovine	26,5	1226,6485	45	45-55	55	45,06	E	QLKPTPEGDLE	I
1	18KD Betalg D bovine	26,5	1244,6017	125	125-135	135	82,11	R	TPEVVDDEALEK	F
1	18KD Betalg D bovine	26,5	1272,6174	124	124-134	134	55,85	V	RTPEVVDDEALE	K
1	18KD Betalg D bovine	26,5	1325,706	43	43-54	54	60	Y	VEQLKPTPEGDL	E
1	18KD Betalg D bovine	26,5	1355,6724	44	44-55	55	62,52	V	EQLKPTPEGDLE	I
1	18KD Betalg D bovine	26,5	1375,6718	42	42-53	53	46,74	V	YVEQLKPTPEGD	L
1	18KD Betalg D bovine	26,5	1454,7508	43	43-55	55	56,53	Y	VEQLKPTPEGDLE	I
1	18KD Betalg D bovine	26,5	1567,8385	43	43-56	56	64,32	Y	VEQLKPTPEGDLEI	L
1	18KD Betalg D bovine	26,5	1617,8032	42	42-55	55	36,18	V	YVEQLKPTPEGDLE	I
2	24KD cas BetaB bovine	19,6	587,2972	115	115-119	119	28,04	Y	PVEPF	T
2	24KD cas BetaB bovine	19,6	753,4528	81	81-87	87	43,75	T	PVVVPPF	L
2	24KD cas BetaB bovine	19,6	882,4862	73	73-80	80	39,25	Q	NIPPLTQT	P
2	24KD cas BetaB bovine	19,6	888,4869	59	59-66	66	49,43	L	VYPFPGPPI	H
2	24KD cas BetaB bovine	19,6	1220,7164	81	81-91	91	48,36	T	PVVVPPFLQPE	V

➡ 4704 dietary peptides unambiguously identified

➤ A large proportion of the sequence covered

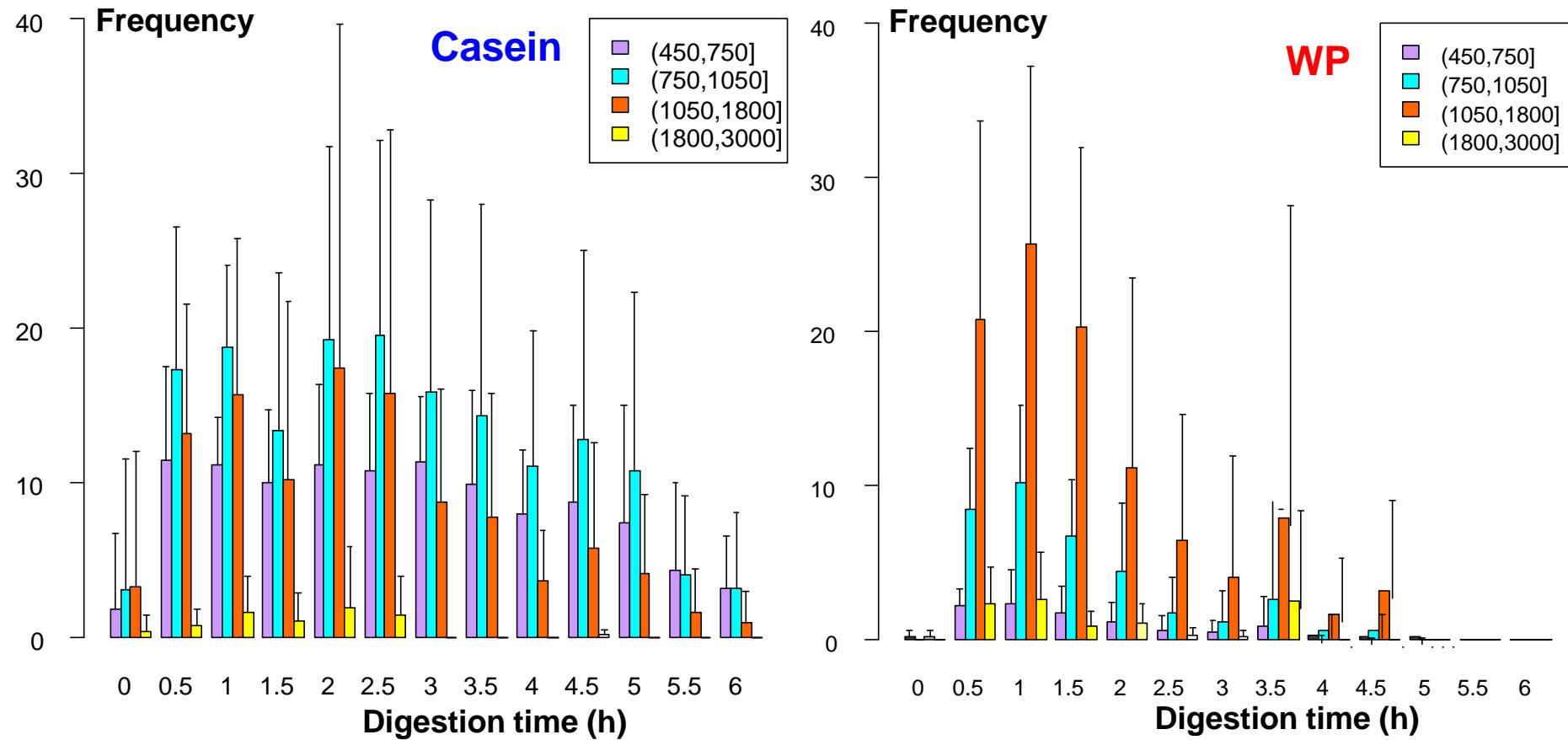
β-lactoglobulin : 70 % of the sequence

1 **LIVTQTMKGL DIQKVAGTWY SLAMAASD**I**S LLDAQSAPLR VYVEELKPTF**
51 **EGDLEILL**Q**K WENGECAQKK **IIAEKT**KIP**A VF**KIDALNEN KV**LVLDTDYK******
101 **KYLLFCMENS AEPEQSLACQ CL**VRTPEVDD EALEKF**D**KAL KALPMH**I**R**L**S**********
151 **F**MPTQ**L EEQC HI**************

β-casein : 67 % of the sequence

1 **RELEELNVPG E**I**VESLSSSE ESITRINKKI EKFQSEKQQQ TEDELQDKIH**
51 PFAQTQ**SLVY PFPGP**I**HNSL PQMIPPLTQT PVVVPPFLQP EVMGVSKVKE******
101 **AMAPKHK**KEMP FPKY**PVEPFT ESQ**SL TL TDV ENL**HLPLPL**L QS**W**M**H**QPH**QP************
151 **LPPTVMFPPQ SVLSLSQS**KV LPVPQ**KAVPY PQRD**MPI**IQAF LLY**QEPVLGP********
201 **VRGPFPI**IV******************

➤ Kinetics of peptides release



- ☞ Release of casein peptides throughout digestion
- ☞ Peptides generated from whey proteins digestion are larger
- ☞ Generation of a unique database of dietary peptides bioaccessible

> High variability in patterns but some peptides present in all subjects

31 peptides present throughout digestion

14 peptides found in all the subjects

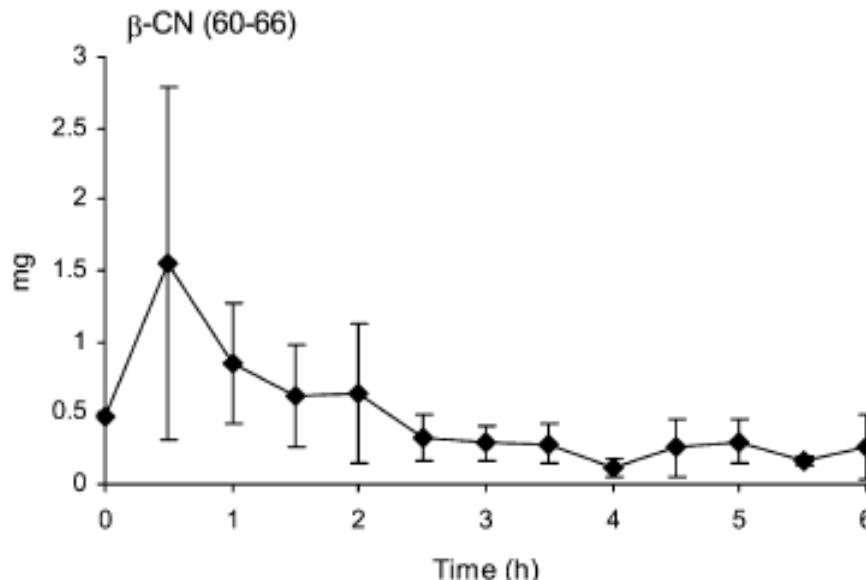
1-6	RELEEL	
57-66	SLVYFPFGPI	57-66 SLVYFPFGPI
58-66	LVYFPFGPI	
58-68	LVYFPFGPIP	
59-66	VYPFGPPI	59-66 VYPFGPPI
59-67	VYPFGPIP	
59-68	VYPFGPIP	59-68 VYPFGPIP
60-66	YPFGPPI	60-66 YPFGPPI
61-66	PFPGPI	60-67 YPFGPPIP
73-79	NIPPLTQ	
73-80	NIPPLTQT	
73-82	NIPPLTQTPV	
80-91	TPVVVPPFLQPE	
81-87	PVVVPPF	
81-91	PVVVPPFLQPE	81-91 PVVVPPFLQPE
81-92	PVVVPPFLQPEV	81-92 PVVVPPFLQPEV
83-91	VVPPFLQPE	
84-91	VPPFLQPE	
85-091	PPFLQPE	
107-113	KEMPEPK	
108-113	EMPFPK	108-113 EMPFPK
114-119	YPVEPF	114-119 YPVEPF
134-139	HPLPL	
135-139	LPLPL	
156-160	MFPPQ	
168-173	VAPFPQ	168-173 VAPFPQ
170-175	VLPVPQ	170-175 VLPVPQ
171-175	LPVPQ	
194-201	QEPVLGPV	177-182 AVPYQP
195-201	EPVLGPV	194-201 QEPVLGPV
196-201	PVLGPV	195-201 EPVLGPV

β -CN
Among those 5 bioactive peptides

Boutrou et al. 2013
Am J Clin Nutr

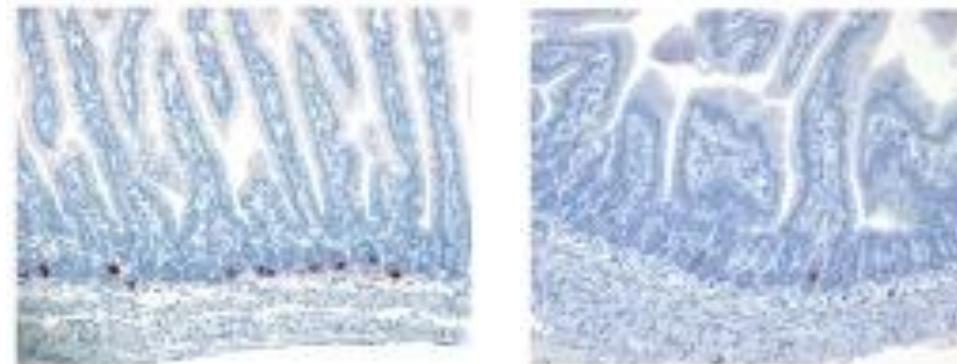
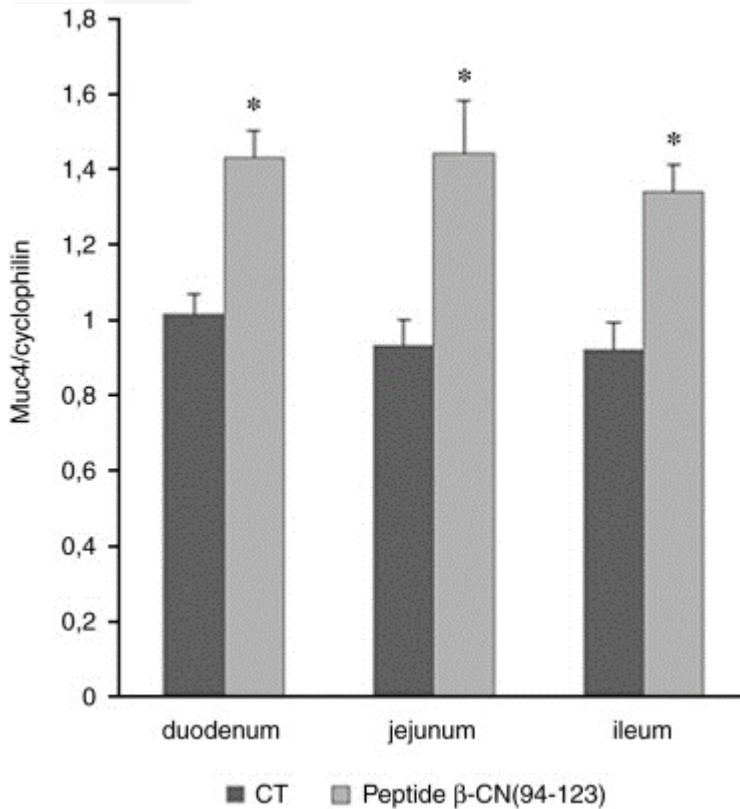
59-66 VYPFGPPI
59-68 VYPFGPIP
60-66 YPFGPPI
108-113 EMPFPK
114-119 YPVEPF

Anti-hypertensive Opioïd



Bioactive peptides are released in the gut lumen at concentrations compatible with a biological action

Bioactive peptides released during digestion can reinforce the defense of the intestinal epithelium



Before After

Sections of duodenum after 10 d of administration of peptide β -CN (94-123)

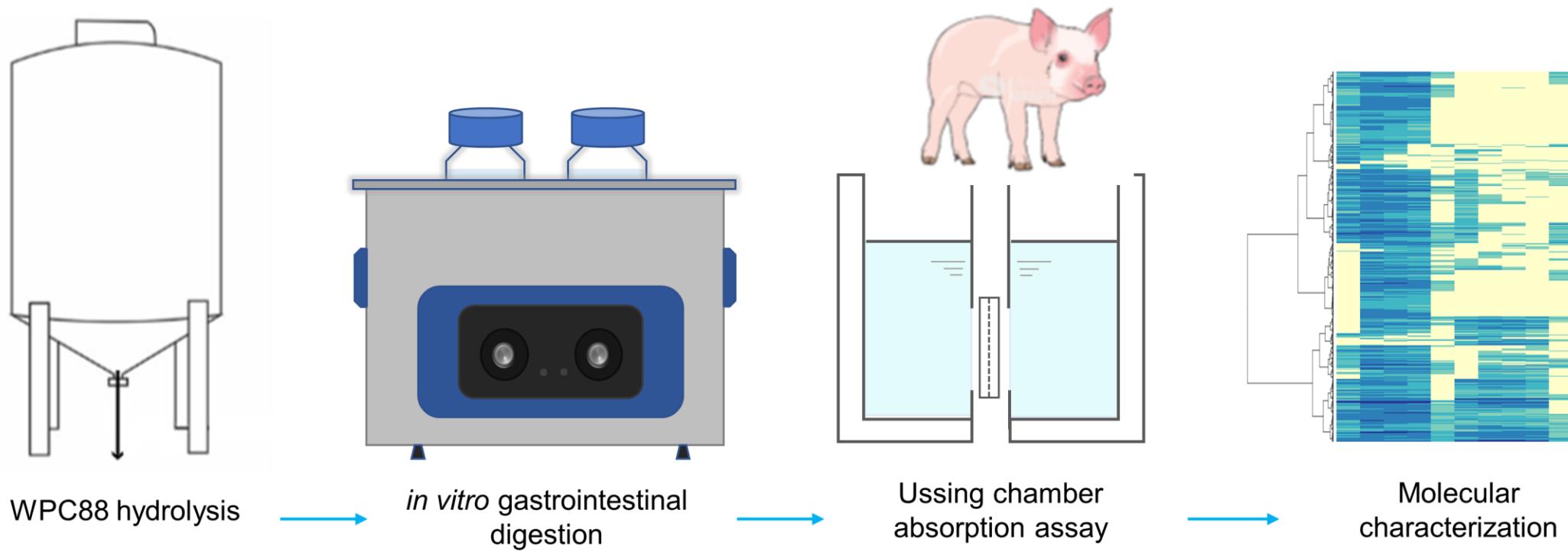
β -CN(94-123) stimulates the mucus production *in vitro* (HT-29 MTX) and *in vivo* (rat)

Plaisancie et al. 2013, J Nutr Biochem
Plaisancie et al. 2015, J Dairy Res
Bruno et al. 2017, J Dairy Sci



Are these bioactive peptides
absorbed?

Ex vivo absorption of milk peptides



A whey protein hydrolysate was submitted to *in vitro* digestion

Peptide absorption was investigated by Ussing Chambers

Absorbed peptides were identified by mass spectrometry

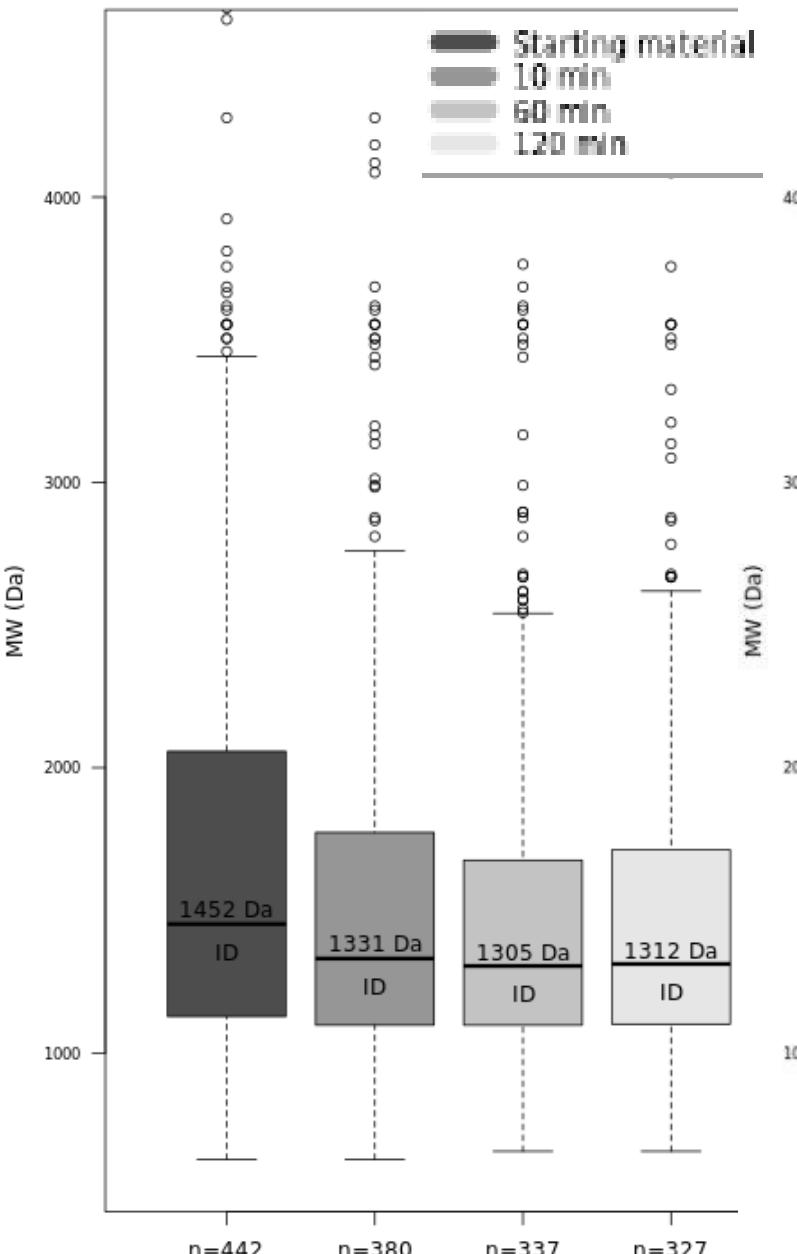
Ozorio et al. 2021
Foods

Mw of the peptides identified in the apical and basolateral chambers

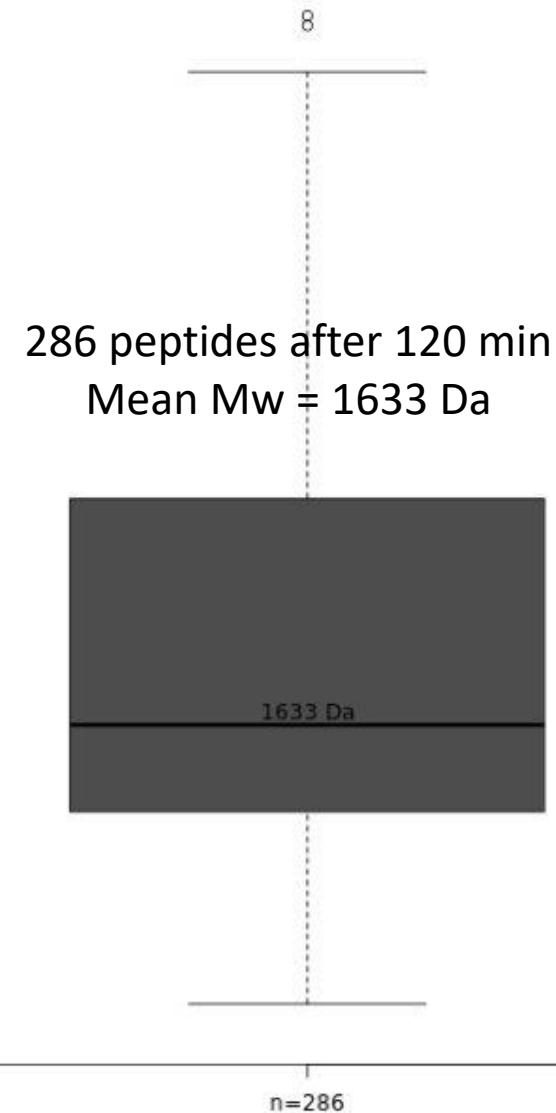
Even large peptides ($Mw > 4kDa$) can go through the epithelial barrier

This confirms our preliminary work on the absorption of a 17 AA peptide (Regazzo et al. 2010)

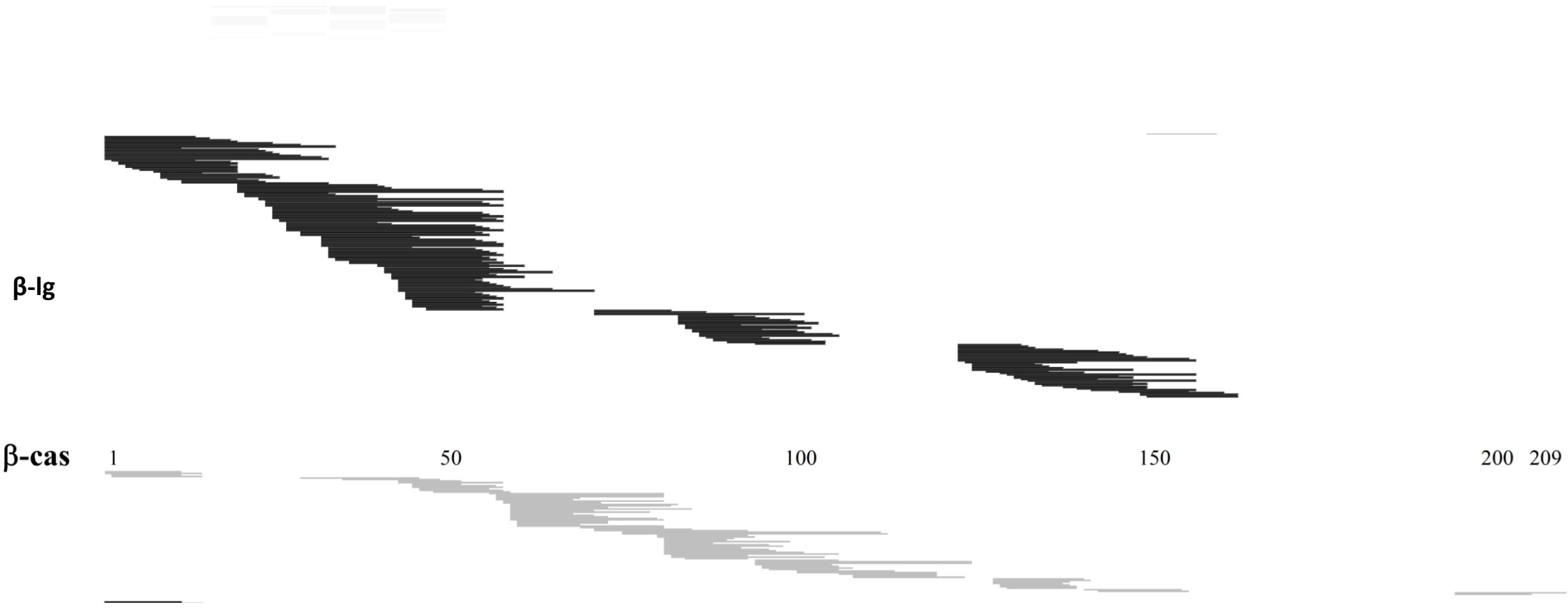
Apical



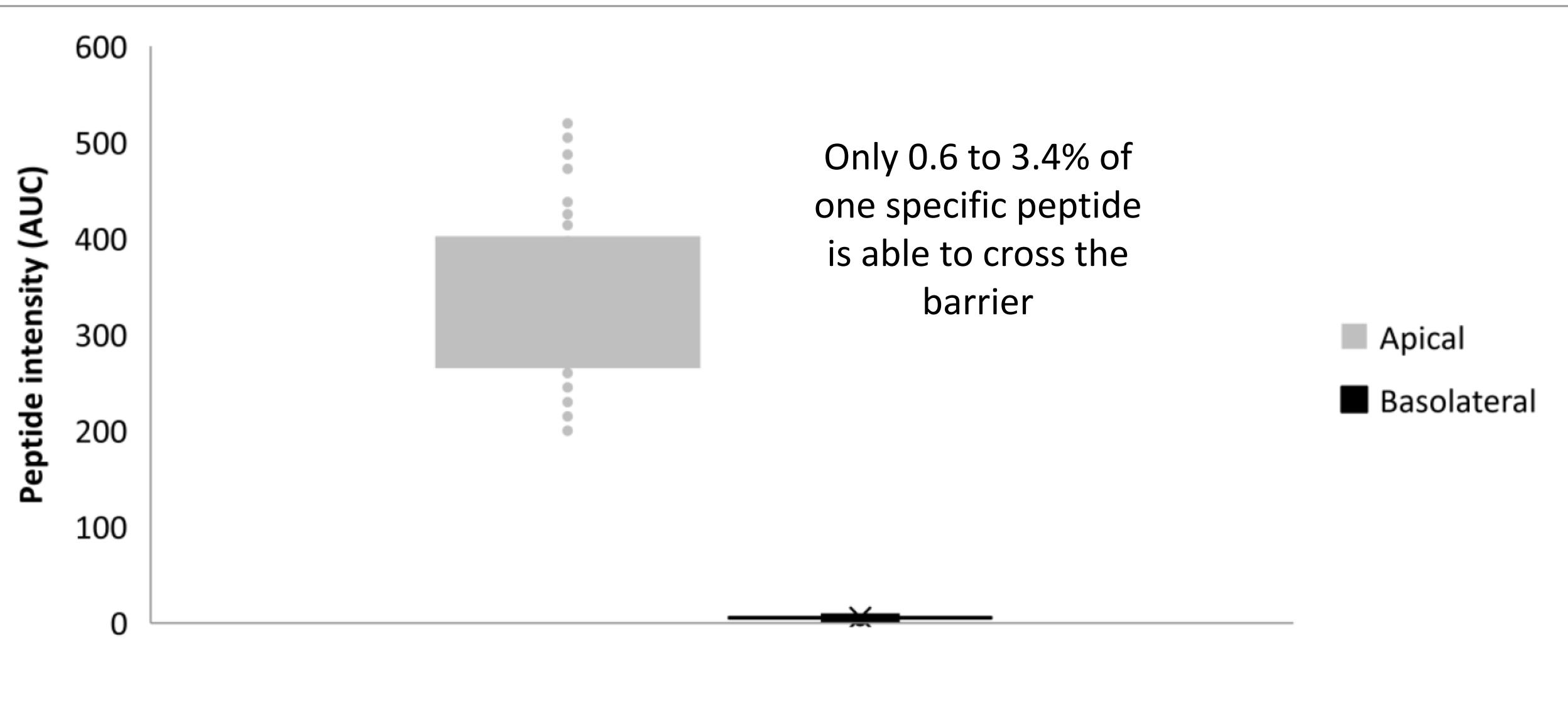
Basolateral



➤ **β -lactoglobulin (β -Ig) and β -casein (β -cas) peptides identified by NanoLC-MS/MS in the basolateral compartment of the Ussing chambers**



Median intensity (AUC) of the 147 peptides identified in both the apical and basolateral compartments of the Ussing chambers



Some of the peptides able to cross the epithelial barrier are bioactive

Protein	Peptide	WPH	ID	Bioactivity	Reference
	KGLDIQKVAGTW	X	X		[42]
	GLDIQK	X	X		[43]
	VLDTDYK	X			
	IVTQTMKG	X			
	VLDTDYKK	X		ACE-Inhibitory	
	KTKIPAV	X	X		[44]
	VEELKPTPEGDLE	X			
	LEKFDFK	X			
	LDIQKVAGTW	X			[45]
	RELKDLKGYGG	X	X		
	LIVTQTMKGLD	X			
	IVTQTMKGLD	X		Anti-diabetic	[46]
β -Ig	IVTQTMKGLDIQ	X			
	LKPTPEGDL	X	X		
	TPEVDDEALEK	X	X	DPP-IV inhibitory	[45]
	VLVLDTDYK	X	X		
	LVLDTDYK	X			[47]
	ALKALPMHI	X			[48]
	GLDIQKVAGT	X	X		
	TPEVDDEALEK	X	X	Antimicrobial	
	VLVLDTDYK	X	X		[45]
	IDALNENK	X	X		
	SLAMAASDISLL	X	X		
	KPTPEGDLEI	X		Memory function	[49]
	KTKIPAVF	X			[50]
	NENKVLVLDTDYKKY	X		Antioxidant	[51]

α -la	GGVSLPEW	X		[52]
	DKVGINYW	X		[45]
	LKGYGGVSLPEW	X		
	LVYPFPGPPIP	X	X	ACE-Inhibitory
	VYPFPGPPIP	X	X	[9]
	YPFPGPPIP	X	X	[45]
	YPFPGPPIHNSLPQ	X		
	VENLHLPLPL	X		Anticancer
	VYPFPGPPIP	X	X	Antioxidant
	YPFPGPPIP	X	X	DPP-IV inhibitory
	VYPFPGPPIP	X	X	Prolyl endopeptidase-inhibitory
	PVVVPPFLQPE	X	X	[45]
κ -cas	VQVTSTAV	X		
α -cas1	SDIPNPIGSENSEK	X	X	Antimicrobial

Take home messages

Dietary proteins are hydrolyzed in the gastrointestinal tract into a myriad of bioactive peptides

Concentrations of these bioactive peptides are sufficient to exert a biological activity in the gut lumen (antimicrobial, increase in mucus and defensins production)

Peptides up to 4 kDa are able to cross the epithelial barrier

But only 0.6-3.4% of a peptide will pass through the epithelium

Since the lifetime of a peptide in the bloodstream is a question of minutes, one can wonder whether bioactive peptides targeting a peripheric organ can really exert a biological activity?

> The Bioactivity & Nutrition team at INRAE Rennes

Head

Didier DUPONT - Senior Scientist

Scientists

Amélie DEGLAIRE – Lecturer
Juliane FLOURY – Lecturer
Catherine GUERIN - Lecturer
Steven LE FEUNTEUN – Senior Scientist
Martine MORZEL – Senior Scientist
Françoise NAU – Professor
Frédérique PEDRONO – Lecturer

Imen JEBALIA – Post-Doc

Stefano NEBBIA – Post-doc

Ines GRECO – Post-doc



PhD students

Jun WANG (2018-2021)
Lea SALLELES (2018-2021)
Elise CHARTON (2019-2022)
Lucile CHAUVENT (2019-2022)
Ousmane SUWAREH (2019-2022)
Rozenn LE FOLL (2020-2023)
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Technicians

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Engineers

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Julien JARDIN
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Jordane OSSEMOND

Masters students

Improving health properties of food by sharing our knowledge on the digestive process

International Network

Dr. Didier DUPONT, Senior Scientist, INRAE, France



Main objective: understanding the mechanisms of food digestion

- Develop new *in vitro*, *in vivo* and *in silico* digestion models including some for specific populations (infant, elderly)
- Harmonize the methodologies and propose guidelines for performing experiments
- Validate *in vitro* models towards *in vivo* data (animal and/or human)
- Identify the beneficial/deleterious components that are released in the gut during food digestion
- Determine the effect of the food matrix on the bioavailability of food nutrients and bioactive molecules

Industry involvement

👉 More than 60 companies are following INFOGEST





INFOGEST



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Vice-chair
Alan Mackie - UK

www.cost-infogest.eu

In vitro
models of
digestion
WG1

Food
interaction –
meal digestion
WG2

Absorption
models
WG3

Digestive
lipases and
lipid digestion
WG4

Digestive
amylases and
starch
digestion
WG5

In silico
models of
digestion
WG6



Isidra Recio



Pasquale Ferranti



Linda Giblin



Myriam Grundy



Nadja Siegert



Choi-Hong Lai



Titre de la présentation Lotti Egger
Date / Brodkorb / nom de l'auteur



Uri Lesmes



Brigitte Graf



Frederic
Carriere



Anabel
Mulet-Cabero



Caroline Orfila



Steven Le Feunteun



INFOGEST

International Conference

Porto, Portugal
2024

> We are pleased to announce the next
8th International Conference on Food Digestion



Ae

INRAE

Titre de la présentation

Date / information / nom de l'auteur

in Porto, Portugal, 9-11 April 2024

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