



HAL
open science

Slightly different metabolomic profiles are associated with high or low weight duck foie gras

Bara Lo, Nathalie Marty-Gasset, Helene Manse, Cécile Canlet, Renaud Domitile, Herve Remignon

► To cite this version:

Bara Lo, Nathalie Marty-Gasset, Helene Manse, Cécile Canlet, Renaud Domitile, et al.. Slightly different metabolomic profiles are associated with high or low weight duck foie gras. PLoS ONE, 2022, 17 (6), pp.e0255707. 10.1371/journal.pone.0255707 . hal-03726460

HAL Id: hal-03726460

<https://hal.inrae.fr/hal-03726460>

Submitted on 18 Jul 2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License

RESEARCH ARTICLE

Slightly different metabolomic profiles are associated with high or low weight duck foie gras

Bara Lo¹, Nathalie Marty-Gasset¹, Helene Manse¹, Cecile Canlet^{2,3}, Renaud Domitile⁴, Herve Remignon¹ ^{*}

1 GenPhySE, Université de Toulouse, INRAE, ENVT, Castanet Tolosan, France, **2** Toxalim, Université de Toulouse, INRAE, ENVT, INP-Purpan, UPS, Toulouse, France, **3** Axiom Platform, MetaToul-Me, National Infrastructure for Metabolomics and Fluxomics, Toulouse, France, **4** IDENA, Sautron, France

✉ Current address: Toxalim, Université de Toulouse, INRAE, ENVT, INP-Purpan, UPS, Toulouse, France
* herve.remignon@toulouse-inp.fr

 OPEN ACCESS

Citation: Lo B, Marty-Gasset N, Manse H, Canlet C, Domitile R, Remignon H (2022) Slightly different metabolomic profiles are associated with high or low weight duck foie gras. PLoS ONE 17(6): e0255707. <https://doi.org/10.1371/journal.pone.0255707>

Editor: Catherine Mounier, Université du Québec a Montreal, CANADA

Received: July 20, 2021

Accepted: June 14, 2022

Published: June 28, 2022

Copyright: © 2022 Lo et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper and its [Supporting information files](#).

Funding: For this specific study we received a grant from the ANR-T (grant number = ANR-T 2017/1414) to partly cover Bara LO's PhD. salary. An other financial support (number = ANR-10-INBS-08) was previously given to the technical platform in charge of our metabolomics analysis. The funders had no role in study design, data

Abstract

Understanding the evolution of fatty liver metabolism of ducks is a recurrent issue for researchers and industry. Indeed, the increase in weight during the overfeeding period leads to an important change in the liver metabolism. However, liver weight is highly variable at the end of overfeeding within a batch of animals reared, force-fed and slaughtered in the same way. For this study, we performed a proton nuclear magnetic resonance (¹H-NMR) analysis on two classes of fatty liver samples, called low-weight liver (weights between 550 and 599 g) and high-weight liver (weights above 700 g). The aim of this study was to identify the differences in metabolism between two classes of liver weight (low and high). Firstly, the results suggested that increased liver weight is associated with higher glucose uptake leading to greater lipid synthesis. Secondly, this increase is probably also due to a decline in the level of export of triglycerides from the liver by maintaining them at high hepatic concentration levels, but also of hepatic cholesterol. Finally, the increase in liver weight could lead to a significant decrease in the efficiency of aerobic energy metabolism associated with a significant increase in the level of oxidative stress. However, all these hypotheses will have to be confirmed in the future, by studies on plasma levels and specific assays to validate these results.

Introduction

France generates about 80% of the world production of "Foie gras". This is mainly composed of mule duck "foie gras", which are interspecific hybrids produced by crossing a common female duck (*Anas platyrhynchos*) with a Muscovy male duck (*Cairina moschata*). To obtain this product, male mule ducks are overfed for up to 12 days after a rearing period of about 12 weeks. During the overfeeding period, the ducks are fed ground corn mixed with water to form a flour that is more digestible. This diet is hypercaloric because corn is composed of 64% starch [1] and finally leads to a hepatic steatosis. Indeed, glucose absorbed by the hepatocytes

collection and analysis, decision to publish, or preparation of the manuscript.

Competing interests: RD was employed by company Idena. The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest. This does not alter our adherence to PLOS ONE policies on sharing data and materials.

is transformed into glycogen and when it reaches about 5% of the hepatic mass, the surplus is redirected to pathways leading to the synthesis of fatty acids which will be esterified into tri-glycerides to be exported to the adipose tissue in the form of very low-density lipoproteins [2, 3]. During the overfeeding period, unlike in mammals, in palmipeds, lipogenesis in the adipose tissue is very limited and mainly localized in the liver [4]. In the mulard duck, the export of *de novo* lipids from the liver to peripheral tissues is limited with low quantities of Very Low Density Lipoproteins or VLDL [5, 6]. Moreover, when the storage capacity of peripheral tissues reaches saturation, the reabsorption of circulating lipids allows the liver to also become the main fat storage site during this process [7]. Thus, at the end of the overfeeding period, a massive storage of lipids in the hepatocytes is observed and largely contribute to a huge increase in the weight of the liver (x10 between the beginning and the end of the overfeeding period). Livers issued from overfed ducks must have a weight greater than 300 g to be legally called « foie gras » [8] but their weights vary a lot above this low limit and often reach a value higher than 700 g at the end of this period [9]. To achieve this high level of synthesis and storage of lipids, several changes in various metabolic activities of the liver have also been observed such as various levels of hypoxia and of oxidative stress, modifications of cellular proteins metabolisms, cytoskeleton and extracellular matrix reorganizations and even cellular deaths by apoptosis [10–15]. However, a large intra-batch variability of final liver weights is generally observed by professionals despite similar intake amounts throughout the overfeeding process. Recent studies on the proteins fraction have been carried out on two classes of liver weights, light (weights between 550 and 600 g) and heavy (weights over 700 g) and showed marked differences in energy metabolism, oxidative stress, lipids and proteins metabolisms and in the extracellular membrane reorganization [12]. Recent metabolomics studies performed on duck livers overfed at 12 weeks of age for different durations (6 or 12 days) have shown that when the duration of overfeeding increases, the average liver weight increases and the hepatic metabolism is oriented towards a reduction of carbohydrate and amino acid metabolism, and the plasma membrane could be damaged [16].

To get a complementary overview of the involvement of main hepatic metabolites in this process, we performed a new comparison with a metabolomic approach using ¹H-NMR between ducks of the same breed, raised and overfed under the same conditions but presenting different liver weights at the end of the overfeeding period. The aim of this study is therefore to identify which essential metabolites significantly vary between low and weight livers and how they inform us about differences in metabolism activities between those two groups of weight livers.

Material and methods

The experiments described here fully comply with legislation on research involving animal subjects according to the European Communities Council directive of November 24, 1986 (86/609/EEC). All the experiment was conducted in a commercial partnership and no specific manipulations of the birds, neither in the farm nor at the slaughterhouse, were done to achieve the production of the samples of livers. Investigators were certificated by the French governmental authority (agreement no. R-31-ENVF-F1-12) and by the Federation of European Laboratory Animal Science Associations (agreement no. F011/05) for carrying out those experiments.

A flock of about 2000 male mule ducks (*Cairina moschata* x *Anas platyrhynchos*) was reared for 12 weeks according to standard commercial rules. Then, birds were overfed for 20 meals (twice a day during 10 days) according to a standard overfeeding program based on moistened corn flour. At the beginning of the overfeeding period, ducks received a quantity of 225 g/meal

which was progressively increased to a final value of 510 g for the last meal. As a whole, during the overfeeding period, ducks ingested an average quantity of 8,8 kg of feed. Ducks were slaughtered approximately 11 hours after the last meal in a commercial slaughterhouse. At 20 min *post-mortem*, 40 livers were randomly sampled according to their weights to create 2 experimental groups: 20 livers weighing between 550 g and 600 g (low weight livers or LWL group) and 20 livers with a weight over than 700 g (high weight livers or HWL group). These two liver weight ranges were chosen because the one corresponding to the LWL group is the considered as "the best one" by processing plant managers, while the one corresponding to the HWL is considered as "the worst one" from a sensory and technical point of view by these same professionals. From all those 20 livers, 50 g were collected from the median lobe and directly frozen in liquid nitrogen before storage at -80°C . The rest of the liver was cooled to 4°C in cooled air. Then, Near Infra-Red Spectra (NIRS) were performed in absorbance from 250 to 2500 nm with an interval of 1 nm with the Labspec[®] 5000 Pro spectrometer (ASD Inc., Boulder (CO), USA), on six independent points of the surface of each liver, to predict the raw biochemical characteristics of the samples.

1- Biochemical analyses

For the 20 harvested samples, the raw biochemical (i.e. dry matter, total lipids and total nitrogen) contents were determined from NIRS spectra according to the method described by Marie-Etancelin et al. (2014) [17]. Then, 16 samples (8 / group) were selected for being representative of the whole 40 samples and further analyzed for metabolomics. The total amount of proteins was determined by the formula: (% Proteins = % total nitrogen \times 6.25).

2- Metabolomic analysis

a- Metabolite's extraction and ¹H-NMR analysis. Polar metabolites were extracted from samples according to a modified protocol from Beckonert et al., (2007) [18]. Briefly, 0.25 g of crushed liver was added to methanol in a ratio 1:4 (w/v) and water in a ratio 1:0.85 (w/v) at 4°C and homogenized in Fastprep[®]-24 Instrument (MP Biomedicals, USA). Subsequently, dichloromethane in a ratio 1:2 (w/v) was added twice and mixed between each addition. Then, water was added in a ratio 1:2 (w/v). After 15 min at 4°C , the mixture was centrifuged at 1 000 g at 4°C for 15 min. The upper phase, composed of hydrophilic metabolites, was collected in new polypropylene tubes and evaporated with a vacuum concentrator (Concentrator Plus, Eppendorf, Hamburg, Germany). Then, each sample was diluted into 650 μl of NMR buffer phosphate pH = 7 in deuterated water (D_2O) with sodium trimethylsilylpropionate (17.2 g TMSP for 100 ml) and stored at -20°C until NMR (Nuclear Magnetic Resonance) analysis. Thereafter, the tube was thawed homogenized and centrifuged for 15 minutes at 5 350 g. Finally, 600 μl were transferred in a NMR tube.

The ¹H-NMR analysis of polar metabolites was done at MetaToul-AXIOM platform (<http://www.metatoul.fr/>) equipped with a Bruker Avance III HD NMR spectrometer at a proton resonance frequency of 600 Mhz. Topspin (V2.1, Bruker, Biospin, Munich, Germany) was used and the NOESYPR1D spin echo pulse sequence was applied to attenuate the signal in the water [19]. The spectra were acquired at 300 K and each spectrum collected with 32 K data points, 16 dummy scans and 512 scans. After Fourier's transformation, all spectra were manually phased, baseline corrected, and chemicals shifts were calibrated to TMSP at 0 ppm. To confirm the chemical structure of metabolites of interest, 2D ¹H - ¹H-COSY (COrrrelation SpectroscopY) and 2D ¹H - ¹³C HSQC (Heteronuclear Single Quantum Coherence spectroscopy) NMR experiment was performed on one sample.

b- Spectra pre-processing and statistical analysis. In a nutshell, the metabolite method involved the use of the ASICS R software package (R software package version 4.0.2 <https://bioconductor.org/packages/ASICS/>) to convert $^1\text{H-NMR}$ spectra into a metabolite relative concentration table. It contains an automatic identification method and quantifies the metabolites in the complex $^1\text{H-NMR}$ spectrum based on its unique peak shape [20, 21]. The metabolite database used consists of the spectra of 190 pure metabolites described by Lefort et al., (2019), therefore only metabolites included in this list will be identified and quantified [22]. The zones corresponding to water (5.0–4.7 ppm), dichloromethane (5.5–5.44 ppm) and methanol (3.38–3.34 ppm) were excluded. Finally, a table of metabolite relative concentration in column and sample number in rows was generated.

Statistical analyses were performed with the R software (version 4.0.2). First, a partial least squares discriminant analysis (PLS-DA) was performed with MixOmics R package (R package version 4.0.2. <https://CRAN.R-project.org/package=mixOmics>) as described by Lê Cao et al., (2011) and the ropls R package (R package version 4.0.2. <https://www.bioconductor.org/packages/release/bioc/html/ropls.html>) was used to highlight the metabolites useful for distinguishing the two (LWL and HWL) original groups of livers [23, 24]. The quality of fit of the models was estimated by the proportion of cumulative explained variance (R^2) for the variables X (X = metabolites) and the variable Y (Y = Liver weight) and by the predictive ability of the model (Q^2). First of all, Q^2 is calculated by a cross-validation, and predictive data are then compared with the original ones and the sum of squared errors calculated for the whole dataset. Then, the Root Mean Square Error of Estimation (RMSEE) was computed and indicated the fit of the observations to the model. When its value is zero, it means that the correlation coefficient is 1 and implies that all points lie on the regression line. So the lower its value, the better the quality of the model [25]. A latent variable is only included in the PLS model if its Q^2 value is greater than or equal to 0.0975 [26]. Finally, the model parameters of the original data (R^2Y and Q^2) and those of all models for 500 datasets when Y (liver weight) is randomly permuted with an unchanged X (metabolite) matrix (pR^2Y and pQ^2) were compared. The model was validated only if all parameter values of the permuted model were much lower than those of the original model and the intercept of the regression line passing through the scatterplot was less than 0. For the final model, only variables with Variable Importance in Projection (VIP) values greater than 1 were considered to be discriminatory. Then, the Student–Wilcoxon’s mean comparison analysis was performed and only metabolites with a p-value < 0.05 were considered as significantly different with multiple testing correction false-discovery rate (FDR).

Results

1- Biochemicals analysis

Liver weights and total protein contents from the two experimental groups were largely different (p-value < 0.001, Table 1). On the contrary, no significant differences (p-value > 0.05)

Table 1. Liver weights (LW), gross chemical composition and color parameters of livers from the two studied groups (n = 8 samples / group).

	LWL	HWL	p-value
LW (g)	581 ± 18	745 ± 24	***
Lipids (% of dry matter)	61.4 ± 0.5	61.6 ± 0.7	NS
Dry Matter (% of raw weight)	71.3 ± 1.8	71.4 ± 1.5	NS
Proteins (% of dry matter)	6.64 ± 0.41	5.76 ± 0.38	***

Values are means ± SD. Within a line, p-values indicate the level of significance (*P ≤ 0.05, **P ≤ 0.01, ***P ≤ 0.001, and NS: not significant).

<https://doi.org/10.1371/journal.pone.0255707.t001>

Table 2. Standard scaling of predictors and response.

Component	R ² X	R ² X(cum)	R ² Y	R ² Y(cum)	Q ²	Q ² (cum)	RMSEE	pR ² Y	pQ ²
1	0.21	0.21	0.792	0.792	0.587	0.587	0.244	0.002	0.004
2	0.19	0.4	0.074	0.866	0.01	0.597	0.203	0.022	0.002

<https://doi.org/10.1371/journal.pone.0255707.t002>

were observed for dry matter and total lipids contents between the two studied groups. All these values are within the ranges of the ones observed in previous studies [9, 12, 27].

2- Metabolomic analysis

As a result, ASICS R software was used to identify a table of 30 metabolites with their intensities from the 16 spectra composed mainly of sugars and amino acids and their derivatives. With the PLS-DA, using one latent variable model, the cumulative parameters of the model were R²X = 0.21 and R²Y = 0.792 and the prediction of the model was Q² = 0.587 (Table 2). Adding a second latent variable only allowed to gain 0.01 for Q², so less than the threshold value of 0.0975 [28]. Consequently, the model with one latent variable was kept as the final model. The model values for R²Y(cum) and Q²(cum) were higher than those obtained after permutation (pR²Y(cum) = 0.002 and pQ²(cum) = 0.004) (Table 2). The final RMSEE value was considered as being low (= 0.244).

Thanks to the PLS-DA, the distinction of the two groups of livers, based on their respective metabolomics profiles, was effective along the horizontal axis which corresponds to the first latent variable (Fig 1).

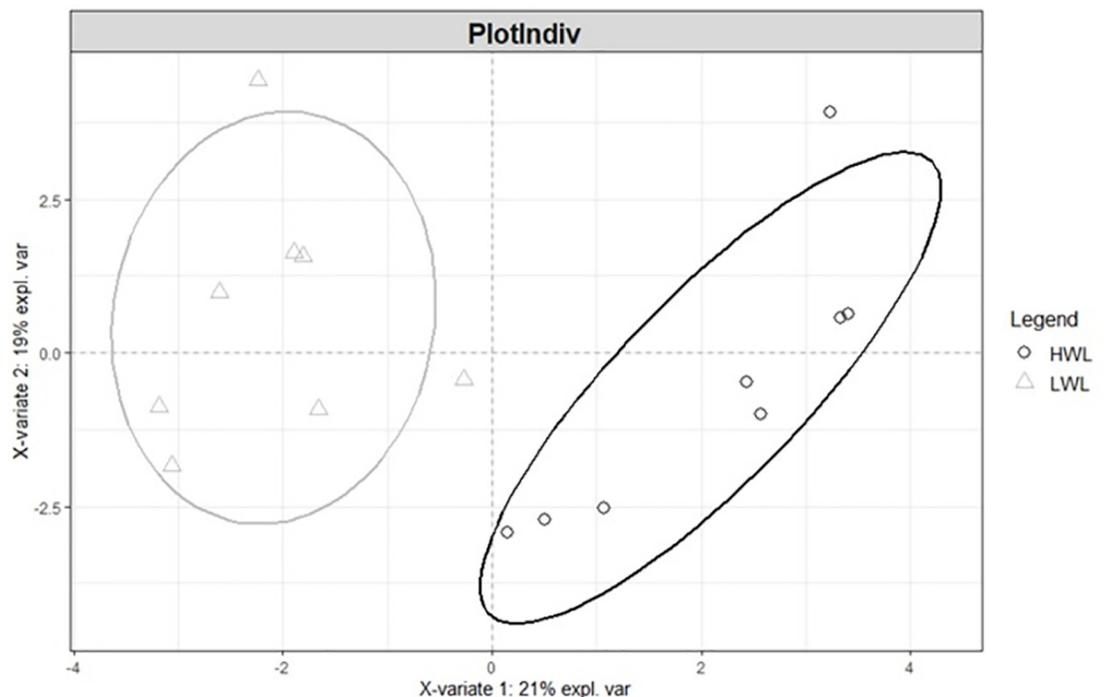


Fig 1. PLS-DA score plots according to the two first latent variables for the two liver weight groups and their 30 variant metabolites (n = 8 livers / group).

<https://doi.org/10.1371/journal.pone.0255707.g001>

Table 3. List of the 30 metabolites identified in the samples from the LWL and/or the HLW groups.

Metabolites	LWL/HWL	VIP	FDR-corrected p-value
D-Glucose	1.26	1.91	7.72E-04
Lactate	0.81	1.87	1.11E-03
L-Glutathione-oxidized	0.83	1.66	5.96E-03
CholineChloride	2.98	1.55	0.01
Creatine	0.75	1.45	0.02
Ethanolamine	1.16	1.44	0.03
Uridine	1.14	1.18	0.07
Glycerol	1.20	0.57	0.07
AceticAcid	1.18	1.13	0.09
Glycerophosphocholine	1.18	1.11	0.09
L-Glutamine	0.76	1.07	0.11
D-Sorbitol	0.57	1.26	0.12
L-Alanine	0.89	1.10	0.12
D-Maltose	0.68	1.19	0.14
L-Serine	0.89	0.86	0.27
MalicAcid	1.56	1.04	0.29
Succinate	1.10	0.72	0.36
L-GlutamicAcid	0.89	0.65	0.39
L-Carnitine	1.06	0.64	0.40
L-Leucine	1.07	0.70	0.45
L-Proline	1.07	0.64	0.48
Myo-Inositol	0.94	0.43	0.54
L-Valine	1.06	0.59	0.61
Phosphocholine	0.92	0.35	0.69
L-Glycine	0.97	0.42	0.72
ArgininosuccinicAcid	0.97	0.41	0.72
Betaine	1.09	0.18	0.81
L-Aspartate	1.03	0.24	0.84
Taurine	1.02	0.14	0.90
2-Oxoglutarate	0.86	0.68	0.96

VIP values > 1 and FDR-corrected p-value < 0.05 allowed to identify the significant LWL / HWL ratios which were reported in bold (n = 8 samples / group).

<https://doi.org/10.1371/journal.pone.0255707.t003>

Fourteen metabolites presented a VIP > 1 and then represented about 50% of the initial 30 quantified metabolites (Table 3). Among this shortened list, 6 metabolites were identified as being the significant ones because they presented an FDR-corrected p-value < 0.05 (Table 2) corresponding to 20% of the initial quantified metabolites. Finally, calculations of the ratios between LWL and HWL for each significant metabolite indicate that 3 of them (creatine, lactate and L-glutathione.oxidized) were in higher amounts in the HWL than in the LWL group. On the contrary, 3 metabolites (choline chloride, D-glucose and ethanolamine) were more present in samples from the LWL group than in those from the HWL group (Table 2).

For the 6 metabolites with a significantly different amount between the two groups, a correlation matrix between metabolite amounts and liver weight was computed (Fig 2).

Discussion

During the overfeeding period, the high-caloric diet provided to the animals leads to a large amount of lipids synthesis in the liver and a large part of these lipids remains stored in

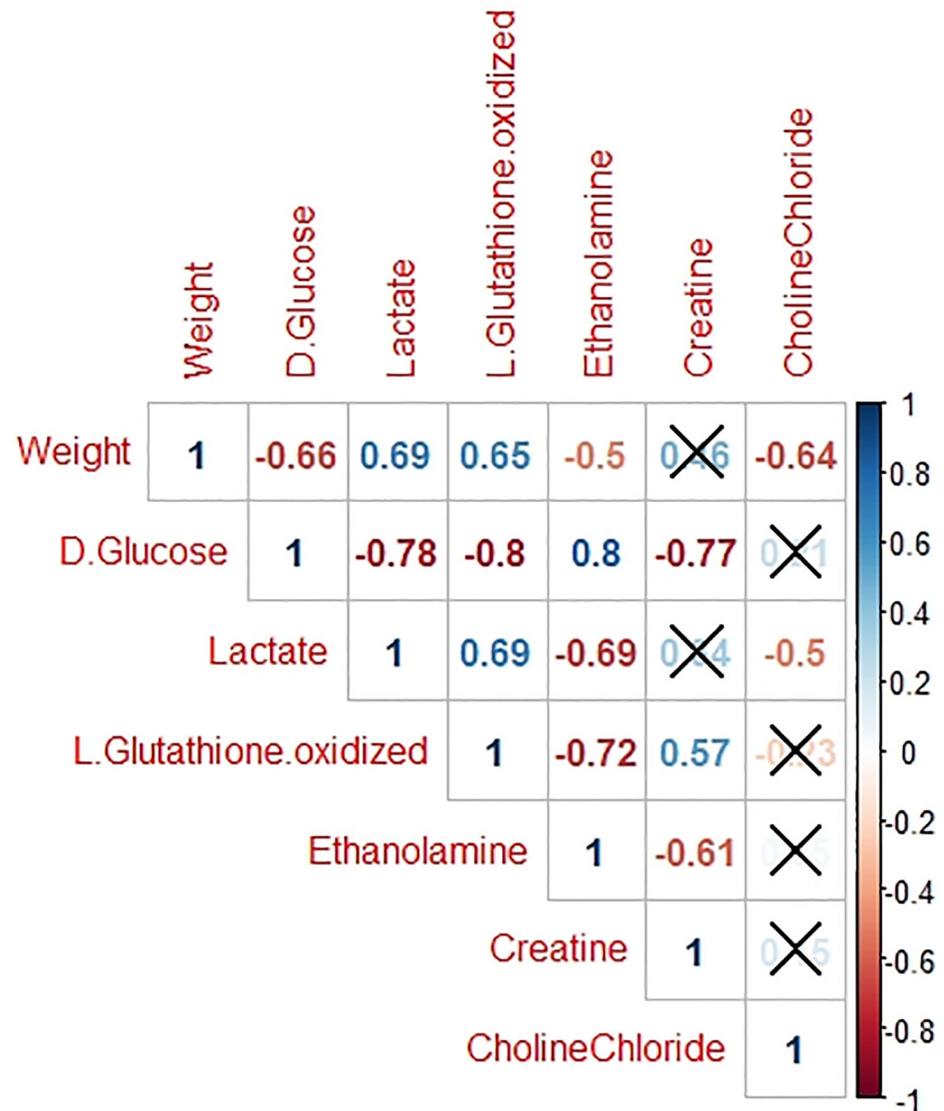


Fig 2. Correlation matrix of liver weight and metabolites identified as most significant in distinguishing the 2 liver weight group. Non-significant correlations (p-value > 0.05) are indicated by a cross.

<https://doi.org/10.1371/journal.pone.0255707.g002>

macrovacuoles located in the cytoplasm of hepatocytes [29]. This process, physiologically defined as liver steatosis, results in an increase in the liver weight from about 80 g to over 700 g [9, 11]. In parallel, the percentage of lipids stored in hepatocytes rose from 5% to more than 50% of the total dry matter (DM) [7, 9].

The glucose, issued from the digestion of the starch which is present in large amounts in the corn, is at the origin of the huge increase in the synthesis of lipids because it is the main precursor of triacylglycerols synthesis in the liver of ducks [30]. Indeed, according to Evans (1972), in birds, about 75% of the glucose entering into the hepatocytes is converted into lipids in the form of fatty acids [31]. Although starch intake increased with increasing amounts of feed at each meal, Mozduri et al., (2021) showed that the plasmatic glucose content was negatively correlated to liver weight ($r = -0.94$) in force-fed ducks [32]. Therefore, a decrease in the

amount of glucose present in the liver may indicate an increase of its utilization for an in-situ synthesis of lipids and consequently the increase in the content of lipids in hepatocytes. Therefore, in the present experiment, the higher amount of glucose recorded in LWL than in HWL induces that lipids synthesis might be more active in HWL than in LWL.

Choline chloride is a salt of choline and is known to be an essential nutrient for poultry [33]. In fact, choline plays important roles in the synthesis of the phospholipids of the membranes and in the methyl-group metabolism [34]. According to Wen et al., (2014), as choline increases in the diet, weight gain and food consumption rose linearly or quadratically [35]. However, maize is a choline poor diet [1], therefore a variation in the amount of choline between LWL and HWL livers is not related to the feeding but to an increase in its biosynthesis in the liver. Several studies have shown that choline prevented excessive lipids accumulation and the incidence of fatty liver in animals [36–38]. This is largely due to the fact that, phosphatidylcholine, a metabolite derived from the choline, is required for the packaging and the export of triglycerides into very low density lipoproteins (VLDL) [39]. Then, if choline is present at a low level in the liver, less VLDL can be synthesized and consequently less lipids will be exported. This favors the development of the hepatic steatosis. In the present study, a greater amount of choline was reported in LWL samples. This probably indicates that LWL, at the end of the overfeeding period, were still able to synthesize large amounts of VLDL and thus to export a part of the lipids neo-synthesized by the hepatocytes. In consequence, the weight of LWL is lower than HWL but this also signifies those hepatocytes from LWL are probably in a more active metabolism than those from HWL which are finally forced to store the lipids because of their incapacity to export them.

Ethanolamine is known to be a precursor of the phosphatidylethanolamine (PE) which is an important component of the cellular membranes. Shimada et al., (2003) compared rats fed an ethanolamine-enriched diet with control rats and observed that the supplementation with ethanolamine induced, in the liver, a significant reduction in cholesterol and triglyceride amounts and an increase in phospholipids [40]. According to Leonardi et al. (2009) and Pavlovic and Bakovic (2013) the reduction of phosphatidylethanolamine synthesis in the liver triggered an increase in fatty acid synthesis to be incorporated in triglycerides and thus favored the development of a hepatic steatosis [41, 42]. In the present study, the amount of ethanolamine was higher in LWL than in HWL samples. This indicates that HWL had less capacities to elaborate PE and then are more oriented in the synthesis of triglycerides for in situ storage rather than in phospholipids.

Sahoo et al., (2017) reported that the reduced glutathione (GSH) is the most important intracellular antioxidant [43]. Thanks to the activity of the glutathione peroxidase, GSH reacts with various oxidizing radicals (including hydrogen peroxide or reactive oxygen species ROS) to produce oxidized glutathione (GSSG) and reduced products. The system GSH-GSSG is one of the most powerful radical scavenger present in various cellular types and it is largely admitted that the increase of the ratio GSH/GSSG is a good indicator of the level of the oxidative status of the cell. According to Chen et al. (2013), the level of oxidized glutathione could be used as a marker of steatosis and oxidative stress [44]. In the present study, the amount of oxidized glutathione is higher in HWL, indicating that high weight livers have a more important level of oxidative stress than low weight livers. These results are in good accordance with previous observations focused on different proteic markers associated with the oxidative status of hepatocytes at the end of the overfeeding period [12]. They indicate that, mainly in HWL samples, the cellular homeostasis of hepatocytes is dysregulated by a metabolic stress.

Lactate is the product of the reduction of pyruvate, issued from glucose during glycolysis, in anaerobic conditions. In humans, Kalhan et al. (2011) showed that plasmatic lactate levels were higher when hepatic steatosis developed [45]. Metabolomic studies performed on duck

livers fed for increasing lengths of time (between 6 and 12 days) showed a positive correlation between liver weight and lactate in plasma ($r = 0.62$) and in liver ($r = 0.98$) [16, 32]. According to Murphy et al. (2001), hyperlactatemia could be the result of cellular hypoxia or hypoperfusion with resulting anaerobic metabolism [46]. Furthermore, McGarry et al. (2018) reported that hypoxia induces an increase in glycolytic activity through an increase in intermediate metabolites such as lactate [47]. Thus, the observed increase in lactate content in HWL (Fig 2) at the end of the overfeeding period could reflect an orientation of the energetic metabolism of hepatocytes towards more anaerobism. This confirms previous reports indicating that high weight livers may suffer from hypoxia when steatosis develops [27].

The process of creatine synthesis is divided into two steps, catalysed by the L-arginine:glycine amidinotransferase (AGAT) and the guanidinoacetate N-methyltransferase (GAMT) which takes place mainly in kidneys and livers, respectively [48]. Recently, Aljobaily et al. (2020) showed that a creatine supplementation in the feed of rats resulted in a decrease in the oxidative stress and damages in high weight livers [49]. This mainly reflects that creatine can play a direct role as an antioxidant [50]. According to Persky and Brazeau (2001), the benefits of creatine are generally attributed to the creatine-induced buffering of cellular ATP levels and its significant depletion will lead to accumulation of intracellular Ca^{2+} , formation of ROS and oxidative tissue damages [51]. In the present study, the level of creatine content was higher in HWL than in LWL. Probably because the oxidative stress occurred more in HWL than in LWL, HWL metabolism was more oriented towards a higher production of creatine to act as a supplementary antioxidant.

Conclusion

The results of this study showed that the more or less important increase in the liver weight during an overfeeding period is associated with differences in the amounts of some hepatic metabolites. Livers with a relative low weight increase (final weight 550 g and 600 g) could be characterized by a moderate utilization of the available quantity of glucose and therefore a lower lipid synthesis, more lipid export thanks to the higher quantity of choline available in the liver associated with a possible decrease in the hepatic concentrations of triglycerides and cholesterol shown by a more marked presence of ethanolamine in those LWL. On the contrary, livers with a weight higher than 700 g could more characterized by a less efficient aerobic energetic metabolism (high amounts of lactate in HWL) associated with a higher level of oxidative stress reported by higher levels of glutathione oxidized and creatine.

Because it is known that different birds will differently support the metabolic shift imposed by the overfeeding, it is not totally surprising to finally see a more or less developed fatty liver. It is associated with a metabolomic profile illustrating a more or less active synthesis and exportation of lipids or the beginning of their excessive accumulation associated with the beginning of cellular oxidation. However, to confirm these hypotheses, other experiments would be complementary such as metabolomic assays of plasma on these two classes of liver weights or even specific assays (measurements of plasmatic glucose or VLDL contents, . . .). In addition, further studies will try to demonstrate if those metabolic states are transitory or definitive and therefore characteristics of the achievement of two different states of hepatic steatosis.

Supporting information

S1 Data.
(CSV)

Author Contributions

Conceptualization: Bara Lo, Renaud Domitile, Herve Remignon.

Data curation: Bara Lo.

Formal analysis: Bara Lo.

Funding acquisition: Renaud Domitile.

Investigation: Bara Lo, Helene Manse.

Methodology: Bara Lo, Nathalie Marty-Gasset, Helene Manse, Cecile Canlet.

Resources: Cecile Canlet.

Supervision: Herve Remignon.

Validation: Nathalie Marty-Gasset.

Visualization: Bara Lo.

Writing – original draft: Bara Lo.

Writing – review & editing: Nathalie Marty-Gasset, Cecile Canlet, Renaud Domitile, Herve Remignon.

References

1. Souci SW, Fachmann W, Kraut H, Scherz H, Senger F. Food composition and nutrition tables. 6th ed. Stuttgart: Medpharm Scientific Publishers; 2000.
2. Postic C, Girard J. Contribution of de novo fatty acid synthesis to hepatic steatosis and insulin resistance: lessons from genetically engineered mice. *J Clin Invest*. 2008; 118: 829–838. <https://doi.org/10.1172/JCI34275> PMID: 18317565
3. Rui L. Energy Metabolism in the Liver. *Comprehensive Physiology*. American Cancer Society; 2014. pp. 177–197. <https://doi.org/10.1002/cphy.c130024> PMID: 24692138
4. Leveille GA, Romsos DR, Yeh Y, O'Hea EK. Lipid biosynthesis in the chick. A consideration of site of synthesis, influence of diet and possible regulatory mechanisms. *Poult Sci*. 1975; 54: 1075–1093. <https://doi.org/10.3382/ps.0541075> PMID: 240159
5. Saez G. Relation entre l'engraissement intramusculaire chez le canard, la lipogénèse hépatique, la sécrétion hépatique des lipides et la capacité de captage des lipides par les muscles. Thèse de doctorat, Université de Pau et des Pays de l'Adour. thesis, Pau. 2009. <http://www.theses.fr/2009PAUU3007>
6. Baeza E, Marie-Etancelin C, Davail S, Diot C. La stéatose hépatique chez les palmipèdes. *INRA Prod. Anim*. 2013; 26: 403–414.
7. Hermier D, Salichon MR, Guy G, Peresson R. Differential channelling of liver lipids in relation to susceptibility to hepatic steatosis in the goose. *Poult Sci*. 1999; 78: 1398–1406. <https://doi.org/10.1093/ps/78.10.1398> PMID: 10536788
8. UE. RÈGLEMENT (CE) N o 543/2008 DE LA COMMISSION du 16 juin 2008. Journal officiel de l'Union européenne. 16 Jun 2008. <https://eur-lex.europa.eu/legal-content/FR/TXT/HTML/?uri=CELEX:32008R0543&from=FR>. Accessed 15 Jul 2019.
9. Bonnefont CMD, Molette C, Lavigne F, Manse H, Bravo C, Lo B, et al. Evolution of liver fattening and foie gras technological yield during the overfeeding period in mule duck. *Poult Sci*. 2019. <https://doi.org/10.3382/ps/pez359> PMID: 31347676
10. Theron L, Fernandez X, Marty-Gasset N, Pichereaux C, Rossignol M, Chambon C, et al. Identification by proteomic analysis of early post-mortem markers involved in the variability in fat loss during cooking of mule duck “foie gras”. *J Agric Food Chem*. 2011; 59: 12617–12628. <https://doi.org/10.1021/jf203058x> PMID: 21999348
11. Rémignon H, Ben Haj Yahia R, Marty-Gasset N, Wilkesman J. Apoptosis during the development of the hepatic steatosis in force-fed ducks and cooking yield implications. *Worlds Poult Sci J*. 2018; 97: 2211–2217. <https://doi.org/10.3382/ps/pey054> PMID: 29509948

12. Lo B, Marty-Gasset N, Pichereaux C, Bravo C, Manse H, Domitile R, et al. Proteomic Analysis of Two Weight Classes of Mule Duck “foie gras” at the End of an Overfeeding Period. *Front Physiol.* 2020; 11: 569329. <https://doi.org/10.3389/fphys.2020.569329> PMID: 33041868
13. François Y, Marie-Etancelin C, Vignal A, Viala D, Davail S, Molette C. Mule Duck “Foie Gras” Shows Different Metabolic States According to Its Quality Phenotype by Using a Proteomic Approach. *J Agric Food Chem.* 2014; 62: 7140–7150. <https://doi.org/10.1021/jf5006963> PMID: 24976256
14. Héroult F, Houée-Bigot M, Baéza E, Bouchez O, Esquerré D, Klopp C, et al. RNA-seq analysis of hepatic gene expression of common Pekin, Muscovy, mule and hinny ducks fed ad libitum or overfed. *BMC Genom.* 2019; 20: 13. <https://doi.org/10.1186/s12864-018-5415-1> PMID: 30616512
15. Pioche T, Skiba F, Bernadet M-D, Seilliez I, Massimino W, Houssier M, et al. Kinetic study of the expression of genes related to hepatic steatosis, glucose and lipid metabolism, and cellular stress during overfeeding in mule ducks. *Am J Physiol Regul Integr Comp Physiol.* 2020; 318: R453–R467. <https://doi.org/10.1152/ajpregu.00198.2019> PMID: 31913683
16. Mozduri Z, Lo B, Marty-Gasset N, Masoudi AA, Arroyo J, Morisson M, et al. Application of Metabolomics to Identify Hepatic Biomarkers of Foie Gras Qualities in Duck. *Front Physiol.* 2021; 12: 966. <https://doi.org/10.3389/fphys.2021.694809> PMID: 34305649
17. Marie-Etancelin C, Vitezica ZG, Bonnal L, Fernandez X, Bastianelli D. Selecting the quality of mule duck fatty liver based on near-infrared spectroscopy. *Genet Sel Evol.* 2014; 46: 38. <https://doi.org/10.1186/1297-9686-46-38> PMID: 24917150
18. Beckonert O, Keun HC, Ebbels TMD, Bundy J, Holmes E, Lindon JC, et al. Metabolic profiling, metabolomic and metabolomic procedures for NMR spectroscopy of urine, plasma, serum and tissue extracts. *Nature Protocols.* 2007; 2: 2692–2703. <https://doi.org/10.1038/nprot.2007.376> PMID: 18007604
19. Parella T. Pulse program catalogue: 1d & 2d nmr experiments. Bruker Biospin. 2006; 2: 373.
20. Tardivel P, Canlet C, Lefort G, Tremblay-Franco M, Debrauwer L, Concordet D, et al. ASICS: an automatic method for identification and quantification of metabolites in complex 1D 1H NMR spectra. *Metabolomics.* 2017; 13: Non Paginé. <https://doi.org/10.1007/s11306-017-1244-5>
21. Lefort G, Liaubet L, Marty-Gasset N, Canlet C, Vialaneix N, Servien R. Joint Automatic Metabolite Identification and Quantification of a Set of 1H NMR Spectra. *Anal Chem.* 2021; 93: 2861–2870. <https://doi.org/10.1021/acs.analchem.0c04232> PMID: 33497193
22. Lefort G, Liaubet L, Canlet C, Tardivel P, Pèrè M-C, Quesnel H, et al. ASICS: an R package for a whole analysis workflow of 1D 1H NMR spectra. *Bioinformatics.* 2019; 35: 4356–4363. <https://doi.org/10.1093/bioinformatics/btz248> PMID: 30977816
23. Lê Cao K-A, Boitard S, Besse P. Sparse PLS discriminant analysis: biologically relevant feature selection and graphical displays for multiclass problems. *BMC Bioinformatics.* 2011; 12: 253. <https://doi.org/10.1186/1471-2105-12-253> PMID: 21693065
24. Thevenot E. ropls: PCA, PLS(-DA) and OPLS(-DA) for multivariate analysis and feature selection of omics data. Bioconductor version: Release (3.12); 2021.
25. Glen S. RMSE: Root Mean Square Error. In: “RMSE: Root Mean Square Error” From StatisticsHowTo.com: Elementary Statistics for the rest of us! [Internet]. 2021 [cited 16 Apr 2021]. <https://www.statisticshowto.com/probability-and-statistics/regression-analysis/rmse-root-mean-square-error/>
26. Tenenhaus M. La régression PLS: théorie et pratique. Editions TECHNIP; 1998.
27. Lo B, Marty-Gasset N, Manse H, Bannelier C, Bravo C, Domitile R, et al. Cellular markers of mule duck livers after force-feeding. *Poult Sci.* 2020; 99: 3567–3573. <https://doi.org/10.1016/j.psj.2020.03.048> PMID: 32616253
28. Kerkri A, Allal J, Zarrouk Z. The L-Curve Criterion as a Model Selection Tool in PLS Regression. *J Appl Probab Stat.* 2019; 2019: e3129769. <https://doi.org/10.1155/2019/3129769>
29. Locsmándi L, Hegedüs G, Andrásy-Baka G, Bogenfürst F, Romvári R. Following the goose liver development by means of cross-sectional digital imaging, liver histology and blood biochemical parameters. *Acta Biol Hung.* 2007; 58: 35–48. <https://doi.org/10.1556/ABiol.58.2007.1.4> PMID: 17385542
30. Saez G, Baéza E, Davail S, Durand D, Bauchart D, Gruffat D. Is the hepatic metabolism of glucose and linoleic acid influenced by species in overfed ducks? *Comp Biochem Physiol Part A Mol Integr Physiol.* 2008; 151: 576–581. <https://doi.org/10.1016/j.cbpa.2008.07.015> PMID: 18687407
31. Evans AJ. In vitro lipogenesis in the liver and adipose tissues of the female Aylesbury duck at different ages. *Br Poult Sci.* 1972; 13: 595–602. <https://doi.org/10.1080/00071667208415986> PMID: 4638819
32. Mozduri Z, Marty-Gasset N, Lo B, Masoudi AA, Morisson M, Canlet C, et al. Identification of Plasmatic Biomarkers of Foie Gras Qualities in Duck by Metabolomics. *Front Physiol.* 2021; 12. <https://doi.org/10.3389/fphys.2021.628264> PMID: 33643071
33. Hegsted DM, Mills RC, Elvehjem CA, Hart EB. Choline in the nutrition of chicks. *J Biol Chem.* 1941; 138: 459–466.

34. Corbin KD, Zeisel SH. Choline Metabolism Provides Novel Insights into Non-alcoholic Fatty Liver Disease and its Progression. *Curr Opin Gastroenterol*. 2012; 28: 159–165. <https://doi.org/10.1097/MOG.0b013e32834e7b4b> PMID: 22134222
35. Wen ZG, Tang J, Hou SS, Guo YM, Huang W, Xie M. Choline requirements of White Pekin ducks from hatch to 21 days of age. *Poult Sci*. 2014; 93: 3091–3096. <https://doi.org/10.3382/ps.2014-03994> PMID: 25260528
36. Gabarrou JF, Salichon MR, Guy G, Blum JC. Hybrid ducks overfed with boiled corn develop an acute hepatic steatosis with decreased choline and polyunsaturated fatty acid level in phospholipids. *Reprod Nutr Dev*. 1996; 36: 473–484. <https://doi.org/10.1051/rnd:19960503> PMID: 8987099
37. Cooke RF, Silva Del Río N, Caraviello DZ, Bertics SJ, Ramos MH, Grummer RR. Supplemental choline for prevention and alleviation of fatty liver in dairy cattle. *J Dairy Sci*. 2007; 90: 2413–2418. <https://doi.org/10.3168/jds.2006-028> PMID: 17430945
38. Pickens MK, Yan JS, Ng RK, Ogata H, Grenert JP, Beysen C, et al. Dietary sucrose is essential to the development of liver injury in the methionine-choline-deficient model of steatohepatitis. *J Lipid Res*. 2009; 50: 2072–2082. <https://doi.org/10.1194/jlr.M900022-JLR200> PMID: 19295183
39. Noga AA, Vance DE. A gender-specific role for phosphatidylethanolamine N-methyltransferase-derived phosphatidylcholine in the regulation of plasma high density and very low density lipoproteins in mice. *J Biol Chem*. 2003; 278: 21851–21859. <https://doi.org/10.1074/jbc.M301982200> PMID: 12668679
40. Shimada Y, Morita T, Sugiyama K. Dietary Eritadenine and Ethanolamine Depress Fatty Acid Desaturase Activities by Increasing Liver Microsomal Phosphatidylethanolamine in Rats. *J Nutr*. 2003; 133: 758–765. <https://doi.org/10.1093/jn/133.3.758> PMID: 12612149
41. Leonardi R, Frank MW, Jackson PD, Rock CO, Jackowski S. Elimination of the CDP-ethanolamine Pathway Disrupts Hepatic Lipid Homeostasis*. *J Biol Chem*. 2009; 284: 27077–27089. <https://doi.org/10.1074/jbc.M109.031336> PMID: 19666474
42. Pavlovic Z, Bakovic M. Regulation of Phosphatidylethanolamine Homeostasis—The Critical Role of CTP:Phosphoethanolamine Cytidylyltransferase (Pcyt2). *Int J Mol Sci*. 2013; 14: 2529–2550. <https://doi.org/10.3390/ijms14022529> PMID: 23354482
43. Sahoo S, Awasthi JP, Sunkar R, Panda SK. Determining Glutathione Levels in Plants. In: Sunkar R, editor. *Plant Stress Tolerance*. New York, NY: Springer New York; 2017. pp. 273–277. https://doi.org/10.1007/978-1-4939-7136-7_16 PMID: 28735403
44. Chen Y, Dong H, Thompson D, Shertzer H, Nebert D, Vasiliov V. Glutathione defense mechanism in liver injury: insights from animal models. *Food Chem Toxicol*. 2013; 60: 38–44. <https://doi.org/10.1016/j.fct.2013.07.008> PMID: 23856494
45. Kalhan SC, Guo L, Edmison J, Dasarathy S, McCullough AJ, Hanson RW, et al. Plasma metabolomic profile in nonalcoholic fatty liver disease. *Metabolism*. 2011; 60: 404–413. <https://doi.org/10.1016/j.metabol.2010.03.006> PMID: 20423748
46. Murphy ND, Kodakat SK, Wendon JA, Jooste CA, Muiesan P, Rela M, et al. Liver and intestinal lactate metabolism in patients with acute hepatic failure undergoing liver transplantation. *Crit Care Med*. 2001; 29: 2111–2118. <https://doi.org/10.1097/00003246-200111000-00011> PMID: 11700405
47. McGarry T, Biniiecka M, Veale DJ, Fearon U. Hypoxia, oxidative stress and inflammation. *Free Radic Biol Med*. 2018; 125: 15–24. <https://doi.org/10.1016/j.freeradbiomed.2018.03.042> PMID: 29601945
48. Barcelos RP, Stefanello ST, Mauriz JL, Gonzalez-Gallego J, Soares FAA. Creatine and the Liver: Metabolism and Possible Interactions. *Mini-Rev Med Chem*. 2016; 16: 12–18. <https://doi.org/10.2174/1389557515666150722102613> PMID: 26202197
49. Aljobaily N, Viereckl MJ, Hydock DS, Aljobaily H, Wu T-Y, Busekrus R, et al. Creatine Alleviates Doxorubicin-Induced Liver Damage by Inhibiting Liver Fibrosis, Inflammation, Oxidative Stress, and Cellular Senescence. *Nutrients*. 2020; 13. <https://doi.org/10.3390/nu13010041> PMID: 33374297
50. Sestili P, Martinelli C, Colombo E, Barbieri E, Potenza L, Sartini S, et al. Creatine as an antioxidant. *Amino Acids*. 2011; 40: 1385–1396. <https://doi.org/10.1007/s00726-011-0875-5> PMID: 21404063
51. Persky AM, Brazeau GA. Clinical pharmacology of the dietary supplement creatine monohydrate. *Pharmacol Rev*. 2001; 53: 161–176. PMID: 11356982