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First report of barley virus G infecting winter barley (*Hordeum vulgare* L.) in France

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1 As part of a cereals virome project high throughput sequencing (HTS)-based viral indexing was 2 performed on plants of various cereals with symptoms of barley vellow dwarf disease collected in June (2017-2020) in the main French cereals production areas. Total RNAs from 32 3 4 individual plants, including 9 winter barley plants were purified (RNeasy Plant Mini Kit, 5 Qiagen, Courtaboeuf, France) and Illumina sequenced (2x150 nt) following ribodepletion 6 (Genewiz-Azenta, Leipzig, Germany). Following quality trimming, reads for each sample were 7 de novo assembled (CLC Genomics Workbench 21, Qiagen) [1] and contigs annotated by 8 BlastX analysis. In four winter barley samples collected in 2018 (18-58, 18-325 and 18-326) 9 and 2019 (19-30A), besides contigs representing diverse viruses such as barley yellow dwarf 10 viruses-PAV and PAS, Hordeum vulgare endornavirus, cereal yellow dwarf virus-RPV (18-11 326), wheat dwarf virus (18-325 and 18-326) and a novel Polerovirus (18-58 and 18-326), 12 large contigs with high identity to barley virus G (BVG) were identified. BVG, a tentative 13 Polerovirus, was initially reported in barley in South Korea in 2016 [2] and has so far been 14 identified in a few other hosts including wheat, oat, maize, proso and foxtail millets as well as 15 switchgrass. It has been reported from the USA, Australia [3] and, in Europe, from the 16 Netherlands, Germany, Hungary and Greece [4]. Large BVG scaffolds representing near 17 complete genomes could be reconstructed for each sample, integrating a total of 128.339, 18 7.188, 8.078 and 20.073 reads, for samples 19-30A, 18-58, 18325 and 18-326 respectively. 19 Given that between 17.2 and 20.5 million reads had been obtained per sample, these values 20 translate into between 0.04% (18-58 and 18-325) and 0.6% (19-30A) of total reads, and to 21 average coverages of between 158x (18-58) and 2866x (19-30A) for the genomic scaffolds. 22 The four assembled sequences (5584-5610 nt) have been deposited in GenBank (ON419453-ON419456). They are nearly identical (98.4 to 99.5% nt identity) and share between 97.7% and 23 24 98.5% nt identity with a barley reference isolate from the South Korea (NC 029906). To 25 confirm the presence of BVG, a primer pair was designed based on available BVG sequences.

Primers BVG-F(5'-CTAGCCCAACGAGTTGCGGG-3') 26 and BVG-R(5'-27 GGTACAGAAGCTCTACGGTTC-3') amplifying a 394 nt product were used in a two-step RT-PCR on new RNA extracts obtained from the 18-325 and 18-326 infected plants. The 28 29 amplicons were directly sequenced and showed respectively 99.2% (ON419457, 18-325) and 30 100% (18-326) nt identity with the corresponding de novo scaffolds. The four analyzed samples 31 have been collected respectively in 2018 (18-58, 18-325, 18-326) and 2019 (19-30A) in three 32 different regions of France (Auvergne-Rhône-Alpes, Occitanie and Centre-Val de Loire), 33 indicating a wide distribution and a persistence over time of BVG in France. To our knowledge, this represents the first report of a natural infection of BVG in cultivated winter 34 35 barley in France. Presence of BVG may have been overlooked in a range of situations, as indicated by its retrospective discovery in a 34 years old Australian sample [3], possibly 36 37 explaining its broad distribution in France. While the mixed infection status of the analyzed 38 plants precludes any conclusion on its pathogenicity in French cereals, BVG has been reported 39 to be associated with a range of symptoms in various hosts so that further studies to evaluate its 40 prevalence and impact in France and to begin to understand its epidemiology are clearly 41 warranted by the present results.

42 References

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