



HAL
open science

The seed germination spectrum of 528 plant species: a global meta-regression in relation to temperature and water potential

Keyvan Maleki, Elias Soltani, Charlotte Seal, Hugh W. Pritchard, Jay Ram Lamichhane

► To cite this version:

Keyvan Maleki, Elias Soltani, Charlotte Seal, Hugh W. Pritchard, Jay Ram Lamichhane. The seed germination spectrum of 528 plant species: a global meta-regression in relation to temperature and water potential. BioRxiv, In press, 25p. 10.1101/2022.08.24.504107 . hal-03764297

HAL Id: hal-03764297

<https://hal.inrae.fr/hal-03764297>

Submitted on 30 Aug 2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NoDerivatives 4.0 International License

1 **The seed germination spectrum of 528 plant species: a global meta-regression in** 2 **relation to temperature and water potential**

3 Keyvan Maleki¹, Elias Soltani², Charlotte E. Seal³, Hugh W. Pritchard⁴, Jay Ram Lamichhane^{5*}

4 ¹Department of Horticulture and Crop Science, Ohio State University, Columbus, OH, USA

5 ²Department of Agronomy and Plant Breeding Sciences, College of Aburaihan, University of Tehran, Tehran, Iran

6 ³Royal Botanic Gardens, Kew, Wakehurst, Ardingly, West Sussex, RH17 6TN, UK

7 ⁴Chinese Academy of Sciences, Kunming Institute of Botany, Kunming, Yunnan, P.R. China

8 ⁵INRAE, Université Fédérale de Toulouse, UMR AGIR, F-31326 Castanet-Tolosan Cedex, France

9

10 For correspondence: jay-ram.lamichhane@inrae.fr; Tel: 0033 561285250

11

12 **Abstract**

13 The germination niche of plant species depends primarily on the seeds' responsiveness to temperature and
14 water potential. However, to appreciate future climate risks to natural regeneration through germination, a
15 global level synthesis across species is needed. We performed a meta-regression of primary data from 377
16 studies on 528 species, including trees, grasses, crops and wild species, to determine patterns and co-correlants
17 in the cardinal values that define species' germination niche. A negative correlation was found between thermal
18 time and base temperature, and positive correlations between other cardinal temperatures and base
19 temperature. Mean values of thermal time indicate that annual crops germinate more rapidly compared to wild
20 species, potentially as a consequence of domestication, and tropical tree seeds the slowest. Dryland species
21 (Cactaceae and Agavaceae) have the widest upper thermal and lower moisture niche, indicative of an ability to
22 grow under harsh conditions, while forages have the narrowest thermal and moisture niche, suggesting higher
23 sensitivity to frost or drought. We propose a new conceptual framework for understanding germination niche as
24 shaped by thermal and moisture traits. Our database represents a unique source of information to further
25 determine the vegetation boundaries of wild or cultivated species, including within simulation studies on plant
26 species adaptations under changing land-use and climate.

27

28 **Keywords;** Adaptive traits, climate change, ecological niche, land-use change, seed functional traits, seed
29 germination niche, threshold-type models.

30

31

32

33 Introduction

34 Germination is a critical growth stage both for domesticated and wild plants and, as such, it plays a vital role in
35 their reproduction and regeneration. As the main environmental drivers, temperature and water potential
36 modulate the germination response of all species (Baskin & Baskin, 2014). Water potential is the most
37 important factor required for successful seed germination, influencing the vigour and final germination
38 level/percentage. The soil is the main source of water for seeds from the environment (Bewley *et al.*, 2012;
39 Zhang *et al.*, 2020). Temperature-regulated seed germination may act either by regulating dormancy status or
40 by controlling the capacity and rates of germination (Bewley *et al.*, 2012; Baskin & Baskin, 2014; Soltani *et al.*,
41 2017a,b). In the presence of adequate moisture, the germination process is limited to a permissible range of
42 temperatures that can be defined by 'cardinal' values; beyond these metabolic activities are impacted and
43 germination does not progress (Bewley *et al.*, 2012). A comprehensive understanding of seed germination
44 behavior and description of intra- and inter-variabilities between groups of species (perennials and annuals,
45 trees, grasses, crops and wild species etc.) to these two main environmental drivers of temperature and water
46 potential is thus informative for two key reasons. First, in defining where seed-producing species are currently
47 able to regenerate on the planet, and second, in predicting how these species will respond to unpredictable
48 environments in the future (Fenner & Thompson, 2005; Baskin & Baskin, 2014; Gremer *et al.*, 2020a,b).
49 Moreover, environmental cues, as an indication of climatic factors, shape the germination niche of plant species
50 that are responsive to specific germination requirements, including dormancy loss under warm and dry
51 conditions, contributing to the timing of germination (Carta *et al.*, 2022).

52 Identification of threshold-type responses to temperature and water potential values help characterize each
53 species' potential for regeneration via seed germination. Germination takes place between a minimum
54 (hereafter referred to as base temperature; T_b) and a maximum (T_{max}) threshold temperature (also known as the
55 ceiling temperature, T_c), with the highest germination speed used to define the optimal temperature (T_{opt})
56 (Bewley *et al.*, 2012; Baskin & Baskin, 2014). T_b is defined as the predicted minimum temperature to be
57 exceeded for germination to progress. It follows that germination will not progress at or below this temperature
58 (Garcia-Huidobro *et al.*, 1982a; Dahal & Bradford, 1994). Estimates of T_b are also valuable in calculating thermal
59 times for the completion of germination in the sub-optimal range between the T_{opt} and T_b . As thermal time
60 varies with sub-populations/percentiles, such estimates represent a powerful means for predicting germination
61 efficiency amongst a seed population under any, including changing, environmental conditions (Gummerson,
62 1986; Dahal & Bradford, 1994; Finch-Savage *et al.*, 2005; Maleki *et al.*, 2021). This concept of the thermal
63 parameters of germination can be expanded by incorporating different water potentials, leading to hydrotime
64 and hydrothermal time models describing the seed response above a base water potential for germination (b)
65 and its interaction with temperature, respectively (Gummerson, 1986; Dahal & Bradford, 1994; Finch-Savage *et al.*,
66 2005; Donohue *et al.*, 2010; Bewley *et al.*, 2012)

67 Estimates of thermal times, cardinal temperatures (i. e. T_b , T_{opt} and T_{max}) and b have been quantified for many
68 species, allowing an interpretation of seed germination within different environments (Trudgill *et al.*, 2005;
69 Orrù *et al.*, 2012; Dürr *et al.*, 2015; Seal *et al.*, 2017; Zhang *et al.*, 2020; Maleki *et al.*, 2021) as well as intra- and
70 inter-species comparisons facilitating prediction of their spatial distribution (Dürr *et al.*, 2015). How crop and
71 weed establishment occur under current and future climates (Forcella *et al.*, 2000; Dürr *et al.*, 2001; Gardarin
72 *et al.*, 2012; Lamichhane *et al.*, 2020, 2022) and how plants synchronize germination and subsequent seedling
73 growth with favorable conditions can be interpreted quantitatively (Dürr *et al.*, 2015; Gremer *et al.*, 2020a;

74 Maleki *et al.*, 2021). Application of these approaches for agricultural and ecological purposes has proven to be
75 useful for an increased understanding of crop diversification in space (intercropping, relay cropping) and time
76 (e.g. introduction of cover crops between two cash crops, double cropping). Designing innovative cropping
77 systems as well as improving environmental sustainability (e.g. restoration of forests, dunes, arid lowlands) also
78 benefit from thermal modelling (Waha *et al.*, 2020; Fernández-Pascual *et al.*, 2021; Beillouin *et al.*, 2021).

79 A comprehensive review of cardinal temperatures for germination indicated that species originating from
80 different geographical origins may show variable cardinal temperatures (Dürr *et al.*, 2015), and this response
81 may result from evolutionary adaptations (Donohue *et al.*, 2010; Baskin & Baskin, 2014). How trait-
82 environment interactions can shape the germination response through the thermal niche has been
83 comprehensively modelled in the laboratory (Thompson & Ceriani, 2003; Catelotti *et al.*, 2020) and *in situ*
84 (Porceddu *et al.*, 2013; Blandino *et al.*, 2022). Thermal time models in combination with relevant
85 environmental parameters have been widely used to investigate thermal niche and germination responses to
86 accumulated temperature in agricultural and more natural settings (Garcia-Huidobro *et al.*, 1982b; Covell *et al.*,
87 1986; Pritchard & KR., 1990; Hardegree, 2006; Porceddu *et al.*, 2013; Dantas *et al.*, 2020; Maleki *et al.*, 2021).
88 Previous studies have shown that variation in threshold-type responses to ongoing environmental conditions
89 among species may reflect the seeds' thermal memory of the maternal environment (Fernández-Pascual *et al.*,
90 2019), as evidenced by the cumulative thermal effects on seed development (Daws *et al.*, 2004; Baskin &
91 Baskin, 2014) and varying levels of seed dormancy (Pritchard *et al.*, 1999; Porceddu *et al.*, 2013).

92 The concept of ecological niche has been used to define the breadth of thermal and moisture ranges in which
93 seeds should be able to germinate under current or future climatic conditions (Porceddu *et al.*, 2013; Sultan,
94 2015; Catelotti *et al.*, 2020; Ordoñez-Salanueva *et al.*, 2021). While the thermal time approach has been used
95 to predict the consequences of climate change (Orrù *et al.*, 2012; Seal *et al.*, 2017; Dantas *et al.*, 2020), hydro
96 time and hydrothermal time quantification should better enable predictions of the impact of global
97 environmental change on species' emergence and habitat patterns. Such insight can then contribute to the
98 design of amelioration and adaptation strategies for environmental management based on seed fitness. A
99 previous study proposed a qualitative conceptual framework to define the temperature tolerance ranges of
100 plant species (Walck *et al.*, 2011) while more recent studies have shown that tropical plants will face the
101 greatest risk from climate warming as they experience temperatures closer to their upper germination limits
102 (Seal *et al.*, 2017; Sentinella *et al.*, 2020). However, no large scale study has yet quantified the germination
103 sensitivity to temperature and moisture that is useful to predict the potential impacts of shifts in environmental
104 conditions on germination.

105 Meta-analysis, meta-regression and systematic review have been widely used to analyze and synthesize data
106 from published and unpublished sources (Borenstein *et al.*, 2021). By employing a simple regression analysis
107 for synthesizing results from multiple studies, meta-regression seeks consensus on significant scientific issues
108 (Borenstein *et al.*, 2021). To this aim, we built on the framework used by Dürr *et al.* (2015) for a global analysis
109 of >200 species' seed germination response, but with two major differences. First, we updated their database,
110 which included 223 articles previously published on plant species with worldwide distribution, by including
111 data from an additional 154 studies published in the last decade on threshold-type responses to both
112 temperature and water potential conditions. Second, while Dürr *et al.* (2015) used a simple literature mining
113 method, we applied a meta-regression approach and employed a new statistical method to interpret changes in
114 threshold-type responses to temperature and water potential values and the relationships among the tested

115 traits. To this aim, we first took seven classes of variables into account, including the variation in ranges of
116 critical temperature and water potential values among species. We then assessed how each species has variable
117 threshold-type values, and what the relationship among traits would be. The information we collated provides
118 new insight into the relevant ecologically-meaningful traits required for plant recruitment and reproduction that
119 must be considered in studies within a quantitative framework for spatial distribution of habitats, and species
120 dispersal.

121 **Materials and method**

122 Data collection

123 We updated a previous database from Dürr et al. (2015) that was based on 223 articles published before
124 December 2011. We integrated additional data that were extracted from 154 articles published between
125 December 2011 and April 2021. We used ISI-Web of Science database to retrieve these articles by using the
126 keywords "thermal requirements + germination" (199 publications), "cardinal temperature + germination"
127 (194 publications), "germination + hydrotime" (94 publications), "germination + hydrothermal time" (275
128 publications), and "thermal ranges + germination" (429 publications) for a total of 377 publications. The
129 original dataset can be accessed at <https://doi.org/10.15454/XP3XHW> (see Data Availability statement). We
130 used Microsoft Excel (version 2016) to collate data on traits related to hydro-thermal times and germination
131 thresholds from the published papers (Garcia-Huidobro *et al.*, 1982b; Gummerson, 1986; Dahal & Bradford,
132 1994). The traits considered in this study included cardinal temperatures (T_b , T_{opt} and T_{max} , which are estimated
133 through regression procedure), thermal time required for germination (θ_{50} , which is calculated via $\theta_{sub}(g) = (T -$
134 $T_b) t_g$ and $\theta_{sup}(g) = (T_c - T) t_g$), and base water potential (ψ which is quantified by $\theta_H = (\psi - \psi_0) t_g$ and $probit(g) = [$
135 $\psi(\theta_H/t_g) - \psi_0] / \sigma_{\psi}$). For further information, see Garcia-Huidobro *et al.*, (1982), Gummerson (1986), Dahal &
136 Bradford (1994) and Maleki et al. (2021), and references therein. For literature mining, seven categories of
137 plant species were defined as previously (Dürr *et al.*, 2015): crops, horticultural species (vegetables, leafy
138 vegetables, ornamentals and medicinal plants), forage and rangeland species, Cactaceae and Agavaceae, wild
139 species (invasive, endangered, wild potential medicinal plants), tropical trees and other trees. All perennial
140 plants (trees and shrubs) were put into the trees category. To collect data on forage species, crops, and
141 vegetables, only species that are important for agricultural purposes, were taken into account. Our study was
142 premeditatedly limited to species with non-dormant seeds, or to seed lots where a pretreatment was applied to
143 release dormancy to avoid the influence of differing patterns of dormancy on germination and on their
144 interrelationships. Furthermore, weeds were excluded because of considerable variations in dormancy level as
145 dormancy status can change cardinal temperatures (Baskin & Baskin, 2014).

146 Determination of germination niche

147 We proposed a new conceptual framework based on the assumption that species construct their germination
148 niche in response to environmental conditions they have experienced (Sultan, 2015; Fernández-Pascual *et al.*,
149 2019). In considering germination niche, we took into account both thermal and moisture dependency. Then,
150 we collected all available data on threshold-type responses to μ and cardinal temperatures. We then used a well-
151 established framework for explaining the seed thermal niche. Based on the framework, thermal niche falls into
152 two distinct categories, namely, the sub-optimal temperature range and the supra-optimal temperature range.

153 The sub-optimal temperature range, from T_b to T_{opt} , in which the germination rate increases as the prevailing
154 temperature rises up to T_{opt} . The supra-optimal temperature range, from T_{opt} to T_{max} , in which the germination
155 rate becomes progressively slower with temperature increase. To determine differences in moisture niche
156 shaped by b values among plant categories, we set a range varying from 0 to negative values assigned to each
157 plant category, suggesting a range in which seeds are able to germinate rapidly compared to the niche range
158 occupied. To calculate both dimensions of niche, we computed the average potential and cardinal temperature
159 values for each plant category, and then plotted the computed values on separate graphs to illustrate how the
160 ecological niche might be distinct among species and plant category.

161 Meta-regression analysis

162 Meta-regression is conceptually similar to simple linear regression, in which explanatory variables predict
163 response variables. Here, the slopes of meta-regression are the effect size predicted by a regression line. Meta-
164 regression coefficients explain how the response variable changes with an increase in the explanatory variable.
165 The effect estimate denotes log risk ratio. Explanatory variables define aspects of studies that have considerable
166 impact on the effect size (also called co-variates). A meta-regression method differs from linear regression in two
167 ways: in a meta-regression, larger studies are more influential than smaller ones; and the residual
168 heterogeneity can be modelled by explanatory variables, giving rise to random-effects meta-regression. The
169 correlation between the tested traits is indicated by positive regression slopes with significant P-values.

170 To perform meta-regression analysis, the following standard regression model was used;

$$Y = \theta + X +$$

171
172 Where θ is intercept showing the overall effect size, Y denotes outcome variable that estimates the changes in
173 traits of interest. X is matrix of explanatory variable. and indicate the vector of coefficients and the random error
174 that refers to the classical regression model, respectively.

175 Weighted least-squares estimators of θ and θ were computed as follows (Hartung *et al.*, 2008);

$$\begin{aligned} \theta &= \frac{\sum (x_i - \bar{x})(y_i - \hat{y})}{\sum (x_i - \bar{x})^2} \\ \hat{y} &= \hat{\theta} \cdot X \\ \hat{y} &= \frac{W_i y_i}{\sum W_i} \end{aligned}$$

176 \hat{y} denotes the estimates of the population effect size, W_i is weight of each study.

177 Residual sum of squares was calculated using following equation:

$$Q_{residual} = \sum W_i (\theta + X + y_i)^2$$

178

179 W_i is weight of each study.

180 We employed log risk ratio as dependent variable and threshold-type traits were used as co-variate variable;

$$\begin{aligned}\log \text{ risk ratio} &= \log (RR_i) \\ \ln(R) &= \ln (X_1) - \ln(X_2)\end{aligned}$$

181 X_1 is T_b values regressed on X_2 representing other traits, including T_{max} , T_{opt} , b and 50

182 The variance of log risk ratio ($v\ln R$) is calculated as follows;

$$v\ln R = \left(\frac{1}{n_1(X_1)^2} \right) + \left(\frac{1}{n_2(X_2)^2} \right)$$

183

184 Where n_1 and n_2 denotes number of studies incorporated into analysis; X_1 and X_2 are T_b values regressed on X_2
185 representing other traits, including T_{max} , T_{opt} , b and 50

186 The approximate standard error (SE) was computed as follows:

187
$$SE \ln R = \sqrt{v\ln R}$$

188 We performed meta-regression on only one covariate (e.g. T_b vs. T_{max}), and, therefore, we suggested the
189 possibility of the Z-test to examine its relationship with effect size. This meta-regression is thus based on the Z-
190 distribution, which is a statistical approach to test the significance of the regression slopes. Therefore, we
191 reported the Z-value with a corresponding p-value to indicate significant correlations, and, we also computed
192 the magnitude of the relationship. The relationship of traits to effect size (defined as log risk ratio) is calculated
193 as follows:

194
$$\ln(RR) = \text{intercept} - \text{the slope of each covariate}(X)$$

195 Where X is the absolute value of traits.

196 The 95% confidence interval for each covariate is estimated as follows:

197
$$LL = X - 1.96 \times SE$$

198
$$UL = X + 1.96 \times SE$$

199 LL and UL refer to lower limit and upper limit, respectively. In the above-mentioned equations, 1.96 shows the
200 Z-value corresponding to confidence limits of 95% (making space for 2.5% error at both end of the distribution).

201 **Results**

202 Intraspecific variation in traits among plant categories

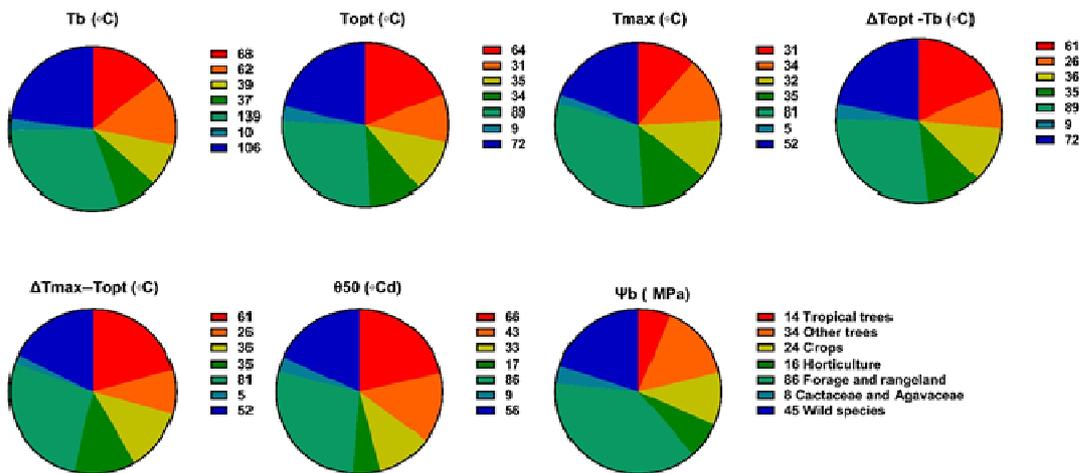
203 Values related to intraspecific variation in traits among plant categories are reported in **Table 1** while the
204 number of species included in intraspecific variation in the analyzed traits among the plant categories is
205 presented in **Figure 1**.

206 **Table 1.** Intraspecific variation in the analyzed traits among the plant categories. The estimates were separately computed using the
 207 standard deviation of observed values for each category. T_b , T_{opt} and T_{max} , $\Delta T_{opt}-T_b$, $\Delta T_{max}-T_{opt}$ indicate the base, optimum, maximum
 208 sub- and supra optimal temperature values, respectively; θ_{50} indicates thermal time required to attain 50% germination while ψ_b
 209 indicates the base water potential values.

Plant category	T_b (C)	T_{opt} (C)	T_{max} (C)	$\Delta T_{opt}-T_b$ (C)	$\Delta T_{max}-T_{opt}$ (C)	θ_{50} ($^{\circ}$ Cd)	ψ_b (MPa)
Tropical trees	4.034	3.404	4.852	5.352	6.006	134.113	0.392
Other trees	4.993	9.732	8.199	9.012	11.794	44.567	1.018
Crops	4.589	5.160	5.722	5.071	7.595	12.791	0.578
Horticulture	4.106	5.679	5.616	3.809	4.185	23.200	0.266
Forage and rangeland	2.322	4.599	4.735	3.456	5.037	12.495	0.344
Cactaceae and Agavaceae	3.477	4.504	5.535	4.527	5.366	19.434	0.569
Wild species	4.657	7.438	6.046	6.386	4.200	43.880	0.448
Average	4.025	5.788	5.815	5.373	6.343	41.497	0.516

210

211 Much information was available on forage and rangeland (198 species) and wild species (107 species) followed
 212 by tropical trees (85 species) and crops (45 species). In contrast, we found much less data on Cactaceae and
 213 Agavaceae (10 species). The range of variation in T_b was similar among plant categories with the ranges varying
 214 from 3 $^{\circ}$ C for forage and rangeland to nearly 5 $^{\circ}$ C for other trees. There was a considerable variation in θ_{50} with a
 215 window ranging from 12 $^{\circ}$ Cd in forage and rangeland to 134 $^{\circ}$ Cd in tropical trees. Variation in T_{opt} , T_{max} and $\Delta T_{opt}-$
 216 T_b was not important among plant categories. ψ_b values showed the lowest variation among the plant categories.
 217 Among all traits considered, forage and rangeland species included higher number of species followed by wild
 218 species and tropical trees (**Figure 1**).



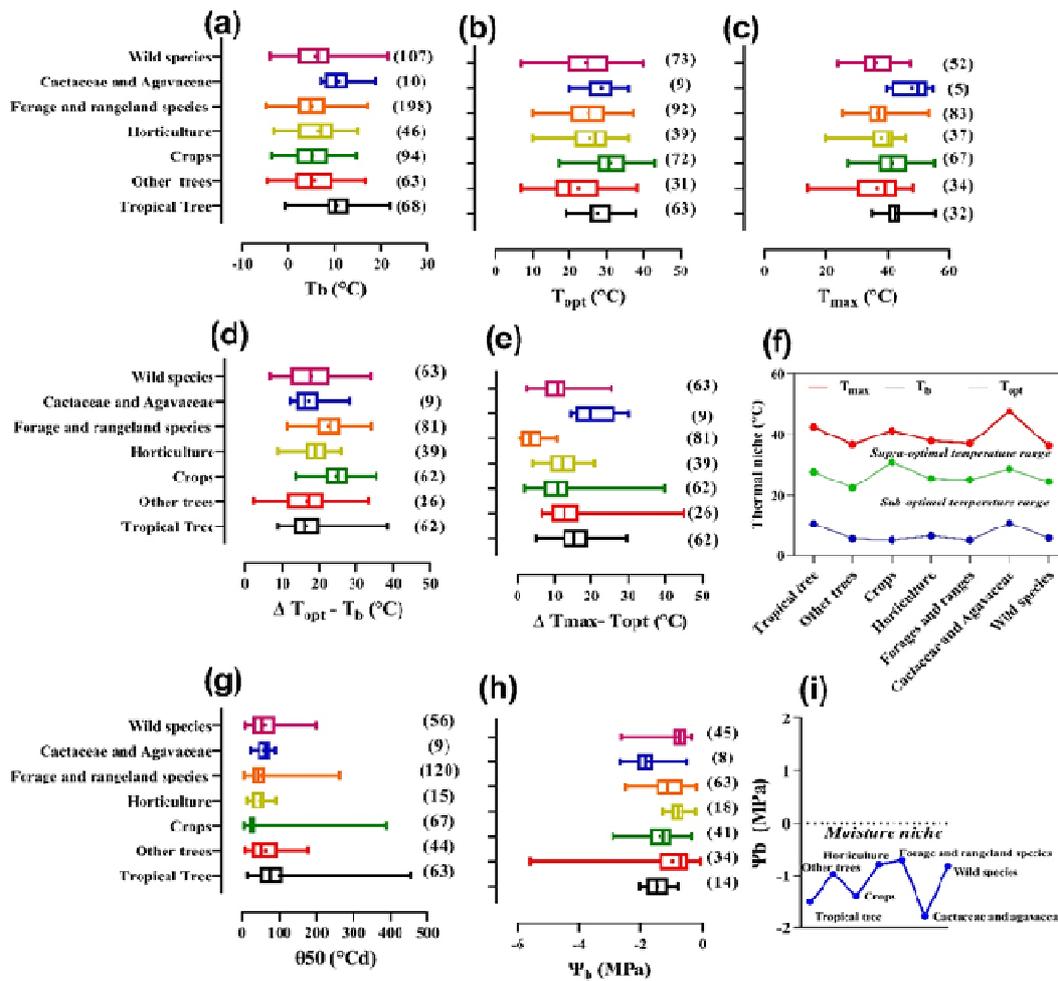
219

220 **Figure 1.** Number of species included in intraspecific variation in the analyzed traits among the plant categories. The numbers
 221 reported in the legend show the number of species in this set of analysis (the number of studies is provided in parenthesis of Figure
 222 1). The number of species exceeds the number of studies as a given study may have reported data on different species. T_b , T_{opt} and T_{max} ,
 223 $\Delta T_{opt}-T_b$, $\Delta T_{max}-T_{opt}$ indicate the base, optimum, maximum sub- and supra optimal temperature values, respectively; θ_{50} indicates
 224 thermal time required to attain 50% germination while ψ_b indicates the base water potential values.

225

226 Cardinal temperatures

227 While Ψ_b values were reported only for 226 species, temperature-based traits were the most studied with T_b and
 228 T_{opt} values being reported for 461 and 334 species, respectively. T_b values varied greatly among plant groups,
 229 ranging from -4.8°C for *Koeleria varilochensis* (i.e. forage and rangeland category) to 21.9°C for *Terminalia*
 230 *brassii* (Tropical trees category; **Figure 2a**). Succulent species (Cactaceae and Agavaceae families) showed less
 231 extreme values with the same mean of 10 C while other species groupings in the analysis had wider variabilities
 232 in T_b values.



233

234 **Figure 2.** Ranges of base (T_b ; **a**), optimum (T_{opt} ; **b**), maximum (T_{max} ; **c**) temperatures, sub-optimal ($\Delta T_{opt} - T_b$; **d**), supra-optimal ($\Delta T_{max} - T_{opt}$; **e**), thermal niche (upper and lower; **f**), thermal time required to attain 50% germination (t_{50} ; **g**), base water potential (Ψ_b ; **h**), and moisture niche (**i**) values of different plant categories considered in this study. The numbers in the parenthesis represent the number of studies found in the literature for each plant group. A frequency distribution was used to show the overview of all traits in some plant categories. In this case, the approach used represents how frequencies are distributed over the observed values. Two-dimensional visualization of the ecological niche of plant species indicating the mean thermal (**f**) and moisture niche (**i**) occupied by each plant category. The breadth of sub-optimal temperature range characterizing the lower thermal niche was estimated by the difference between minimum and optimum temperature while the of supra-optimal temperature range characterizing the upper thermal niche was estimated by the differences between maximum and optimum temperature. Moisture niche was a dimension estimated by taking the mean Ψ_b values of each plant category from the database.

243

244 T_{opt} for germination of crops was limited to a window ranging from 17°C to 44°C, the latter being the highest
245 observed values (**Figure 2b**). The widest range of T_{opt} for germination was found for wild species, from 7 C to 40
246 C, followed by the category "other trees" from 6°C to 38°C. In contrast, inter-species variation for T_{opt} for
247 Cactaceae and Agavaceae and tropical trees had the narrowest range, with values of 20 C to 39 C and 20°C to
248 40°C, respectively.

249 T_{max} also varied considerably among plant categories. The lowest T_{max} values were observed for non-tropical trees
250 (14C for *Acer saccharum*) and horticultural plants (20°C for *Muscari comosum*). In contrast, the highest T_{max}
251 values were around 55°C, in tropical trees (55.4°C for *Cenostigma microphyllum*), Cactaceae and Agavaceae
252 (54.5°C for *Polaskia chende*), forage and rangeland species (53.3°C for *Urochloa brizantha*) and crops (55°C for
253 *Cicer arietinum* L.) (**Figure 2c**). Amongst species groupings, T_{max} varied the most for non-tropical trees (14°C for
254 *Acer saccharum* to 48°C for *Anadenanthera colubrine*; **Figure 2c**) and the least for Cactaceae and Agavaceae
255 (from 39.5°C to 54.5°C for *Manfreda brachystachya* and *Polaskia chende*, respectively).

256 Thermal niche for germination and τ_{50}

257 Values of sub- and supra-optimal temperature range varied markedly among plant species (**Figure 2d,e,f**).
258 Forage and rangeland species and crops, showed the widest sub-optimal temperature range, with the extreme
259 values of 20°C and 26°C, respectively. In contrast, non-tropical and tropical trees had the narrowest sub-optimal
260 temperature range, with an average value of 17C for both plant categories. In the supra-optimal temperature
261 range all species had a thermal niche spanning mean values of 12°C to 19°C, e.g. Cactaceae and Agavaceae
262 (19°C), tropical trees (15°C), non-tropical trees (14°C), crops (11°C) and wild species and horticulture species
263 (both 12°C).

264 τ_{50} values varied nearly two orders of magnitude among plant groups ranging from 6 °Cd to 500Cd (**Figure 2g**).
265 Tropical tree species had the highest range of τ_{50} values, ranging from 15.1°Cd for *Anadenanthera colubrina* to
266 477°Cd for *Araucaria angustifolia*. In crops, τ_{50} ranged only about 10-fold, from 6.3°Cd for *Sesamum indicum* L.
267 to 59.4°Cd for *Vicia variabilis*. Wild species, forage and rangeland species, and non-tropical trees showed a
268 similar range in terms of thermal time required for germination, varying from 6°Cd to 263°Cd. Cactaceae and
269 Agavaceae all had relatively short τ_{50} varying only two-fold from 39.3°Cd to 87.6°Cd.

270 Ψ_b and moisture niche for germination

271 Ranges of Ψ_b values are presented **Figure 2h**. Compared to the other traits mentioned above, we found fewer
272 data on Ψ_b values of tropical trees (14 papers) and horticultural plants (18 papers). In contrast, much more
273 information was available on Ψ_b values of forage and rangeland species (63 papers) and wild species (45
274 papers). Overall, Ψ_b values differed considerably among plant species, ranging from a lowest value of -6 MPa for
275 *Atriplex halimus* (a non-tropical tree species) to nearly -0.2 MPa observed for *Stipa grandis* (a forage and
276 rangeland species). Wild species, Cactaceae and Agavaceae, and crops showed similar Ψ_b values, with a range
277 varying from -3 MPa to nearly 0 MPa. In contrast, the narrowest range of Ψ_b values was for horticultural crops
278 (from -1.27 MPa for *Cucurbita pepo* to -0.21 MPa for *Trachyspermum ammi*) followed by that for tropical trees
279 (from -2.02 MPa for *Apeiba tiborbou* to -0.81 MPa for *Peltophorum dubium*) and wild species (from -2.62 for
280 *Pectocarya heterocarpa* to -0.34 MPa for *Erodium cicutarium*). Forage and rangeland species showed the highest
281 Ψ_b values (near to zero as for example -0.21 MPa for *Stipa grandis*).

282 Values of moisture niche are presented in **Figure 2i**. Similar to the higher thermal niche, tropical trees (14
 283 species and 13 papers), and Cactaceae and Agavaceae (12 species and 8 papers) were revealed to be the most
 284 tolerant groups to drought, with a range varying from -2.64 MPa to -0.5 MPa . In contrast, non-tropical trees,
 285 and forage and rangeland species showed higher (less negative) Ψ_b values, of -0.07 and -0.19 MPa,
 286 respectively. Crops had the widest moisture niche, with a range varying from 0 MPa to -1.39 MPa. Forage and
 287 rangeland, and horticulture species have the narrowest moisture niche, with the occupancy range varying from
 288 0 to -0.71 MPa and from 0 to -0.79 MPa, respectively.

289 Correlations between traits

290 A positive correlation was found between T_b and T_{opt} values for all plant categories (**Figure 3a**). The steepness of
 291 regression slopes was the highest for forage and rangeland species (0.07; $P=0.02$; **Table 2**) and lowest for
 292 Cactaceae and Agavaceae and tropical tree species (0.02 and $P=0.00$ for both species; **Table 2**).

293 **Table 2.** Meta-Regression reporting T_{opt} , T_{max} , $\Delta T_{opt}-T_b$, $\Delta T_{max}-T_{opt}$, θ_{50} and b_b values for different categories of plant species as a function
 294 of T_b for germination. The coefficients were obtained using linear regression. Residual heterogeneity indicates variation around the
 295 true regression line. Tau-squared shows the variance of the effect size across studies included in the analysis. T_b , T_{opt} and T_{max} , $\Delta T_{opt}-T_b$,
 296 $\Delta T_{max}-T_{opt}$ indicate the base, optimum, maximum sub- and supra optimal temperature values, respectively; θ_{50} indicates thermal time
 297 required to attain 50% germination while b_b indicates the base water potential values.

298

Plant group	Coefficient		SE	P-value	Residual heterogeneity	Tau-squared
	Intercept	Slope				
T_{opt}						
Tropical trees	-0.71	0.02	0.00	0.00	86.44	0.00
Other trees	-0.87	0.04	0.01	0.03	22.41	0.00
Crops	-1.05	0.05	0.00	0.00	89.24	0.00
Horticulture	-0.84	0.03	0.00	0.03	52.40	0.00
Forage and rangeland	-1.17	0.07	0.00	0.02	74.14	0.00
Cactaceae and Agavaceae	-0.71	0.02	0.0	0.00	6.38	0.00
Wild species	-0.95	0.05	0.00	0.01	10.02	0.01
Average	-1.08	0.05	0.00	0.014	53.04	0.01
T_{max}						
Tropical trees	-1.20	0.06	0.00	0.01	14.30	0.00
Other trees	-1.07	0.04	0.00	0.01	18.68	0.00
Crops	-1.16	0.05	0.00	0.00	74.76	0.00
Horticulture	-1.06	0.04	0.00	0.04	45.38	0.00
Forage and rangeland	-1.26	0.07	0.00	0.02	83.64	0.00
Cactaceae and Agavaceae	-1.03	0.03	0.01	0.03	4.69	0.00
Wild species	-1.15	0.06	0.00	0.02	53.17	0.00
Average	-1.23	0.06	0.00	0.00	55.31	0.01
$\Delta T_{opt}-T_b$						
Tropical trees	-0.84	0.06	0.00	0.04	51.12	0.00
Other trees	-0.69	0.04	0.00	0.00	17.58	0.00
Crops	-1.02	0.06	0.00	0.03	12.46	0.01
Horticulture	-0.87	0.05	0.00	0.01	52.93	0.00
Forage and rangeland	-1.19	0.09	0.00	0.00	62.39	0.00
Cactaceae and Agavaceae	-0.74	0.04	0.00	0.02	8.11	0.00

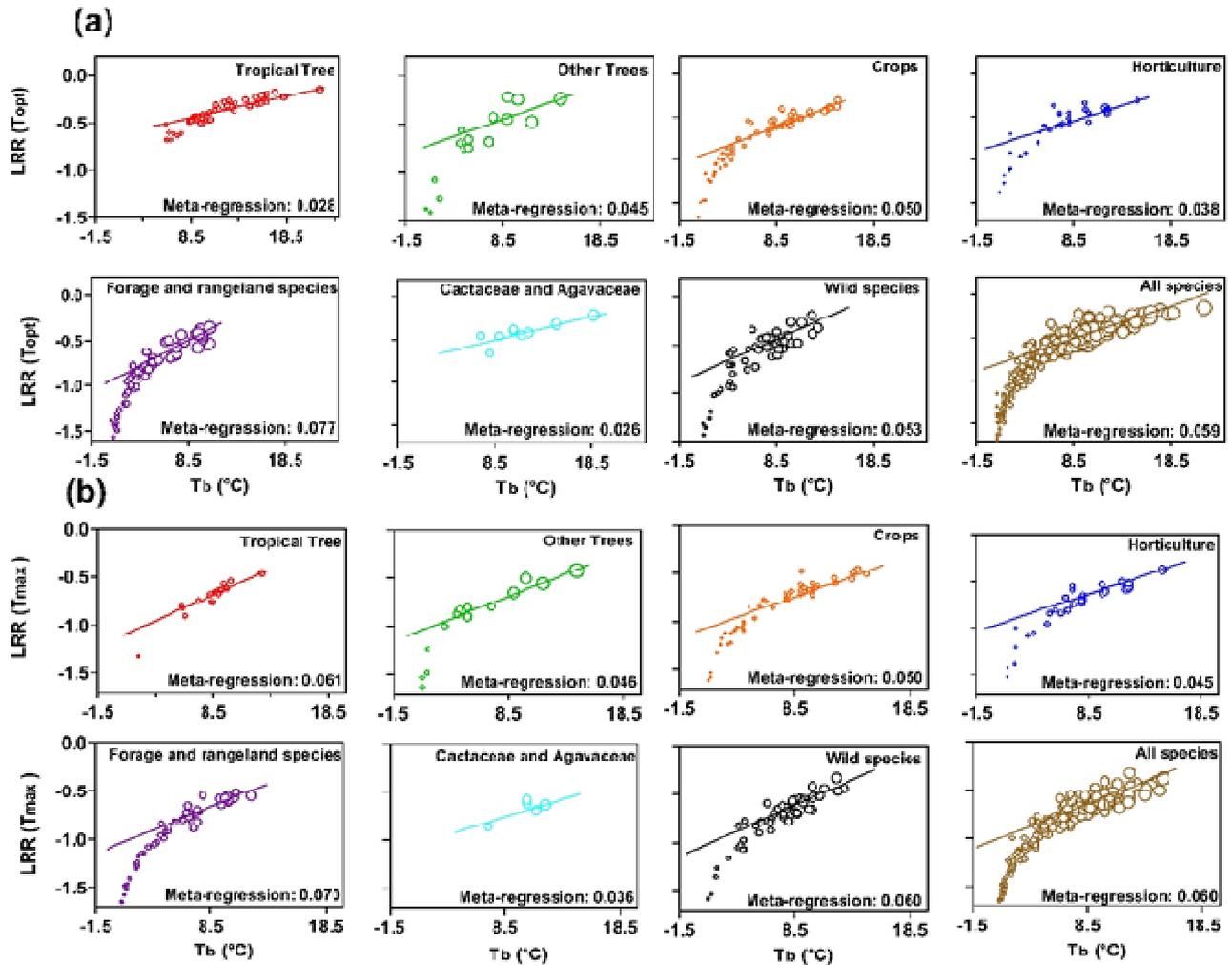
Wild species	-0.97	0.07	0.00	0.02	82.97	0.01
Average	-1.01	0.07	0.00	0.00	63.75	0.01
$\Delta T_{\max} - T_{\text{opt}}$						
Tropical trees	-0.938	0.07	0.00	0.04	31.16	0.00
Other trees	-1.15	0.09	0.00	0.00	27.32	0.01
Crops	-0.79	0.06	0.00	0.03	17.46	0.01
Horticulture	-0.71	0.06	0.00	0.01	22.53	0.00
Forage and rangeland	-0.91	0.07	0.00	0.00	12.25	0.00
Cactaceae and Agavaceae	-1.03	0.07	0.00	0.02	18.56	0.01
Wild species	-0.79	0.08	0.00	0.02	62.67	0.01
Average	-0.65	0.05	0.00	0.00	52.25	0.00
50						
Tropical trees	1.25	-0.03	0.01	0.02	60.32	0.06
Other trees	1.02	-0.03	0.01	0.00	63.74	0.09
Crops	0.52	-0.04	0.03	0.01	30.18	1.18
Horticulture	1.61	-0.11	0.02	0.03	11.79	0.07
Forage and rangeland	0.82	-0.02	0.05	0.02	19.38	0.00
Cactaceae and Agavaceae	0.93	-0.02	0.02	0.00	8.48	0.00
Wild species	1.40	-0.07	0.00	0.00	63.02	0.01
Average	0.90	-0.02	0.00	0.01	21.50	0.12
b						
Tropical trees	0.08	0.40	0.41	0.03	0.33	0.00
Other trees	-1.38	-0.44	0.22	0.02	8.67	0.00
Crops	-1.30	-0.60	0.13	0.01	18.20	0.08
Horticulture	-1.12	-0.38	0.37	0.01	19.61	0.01
Forage and rangeland	-0.50	-0.09	0.04	0.01	55.63	0.03
Cactaceae and Agavaceae	-1.70	-0.54	0.30	0.02	1.60	0.00
Wild species	-1.27	-0.73	0.14	0.01	40.08	0.01
Average	-0.003	-0.26	0.05	0.00	30.29	0.05

299

300 Values of T_b positively correlated with T_{\max} , with a high proportion of variance corresponding to high T_b values
 301 (**Figure 3b**). The steepness of the regression slopes differed markedly among plant categories, from 0.07 for
 302 Forage and Rangeland species ($P=0.02$ and 0.01 , respectively; **Table 2**), then wild species and tropical trees,
 303 (0.06 for both, $P=0.02$ and 0.00 , respectively; **Table 2**). In contrast, a weaker correlation was observed between
 304 T_b and T_{\max} in Cactaceae and Agavaceae, and horticulture species as shown by the regression slope values of 0.03
 305 and 0.04 , respectively ($P=0.03$ and 0.04 , respectively; **Table 2**).

306 Different data inputs and variance are shown in **Figure 3** by varying size of bubbles such that inflated bubbles
 307 refer to higher variance. Despite differing levels of data coverage that may influence the slopes of meta-
 308 regression and subsequently the correlations obtained, the general trend indicates that there is a positive
 309 correlation between T_b and T_{\max} (the slope of 0.060 ; **Table 2**; **Fig 3b**). Tropical trees, and forage and rangeland
 310 species showed steeper slopes than the general trend while all other plant categories were characterized by
 311 values lower than the general trend (**Table 2**; **Fig 3b**). Overall, variance inflated with increasing T_b , indicating
 312 that there is considerable difference between species groups and data shortage (**Figure 3**). This trend may be
 313 reflected in other parameters as well.

314

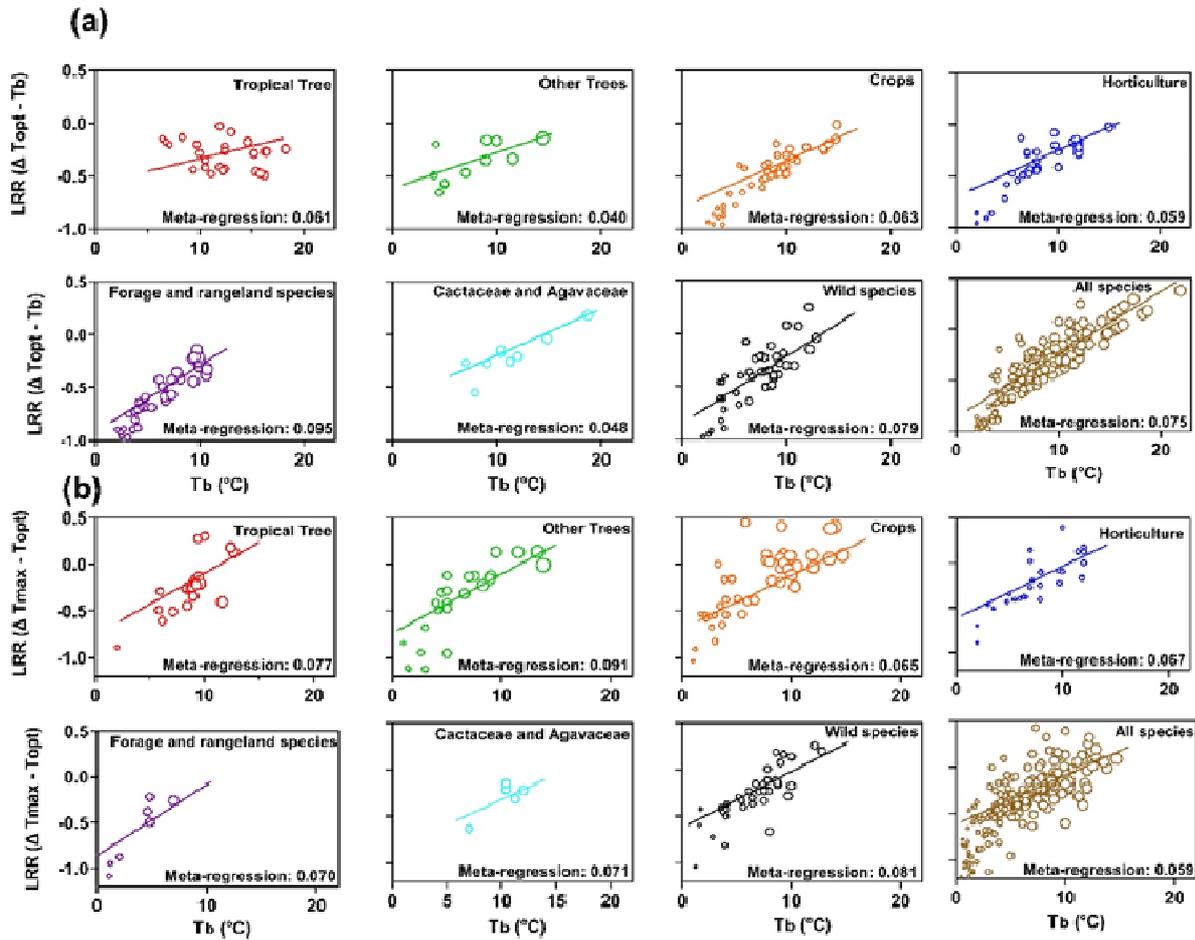


315

316 **Figure 3.** Regression of log risk ratio (LRR) of optimum (a) and T_{max} (b) temperature on base temperature (T_b). The size of each bubble is
 317 inversely correlated with the variance of the log relative risk estimate with larger bubbles showing more inflated variance. LRR
 318 represents the probability of changes in the range of T_{opt} and T_{max} as a function of differing T_b ranges. Fitted lines were obtained
 319 through linear regression approach ($Y = \theta + X +$). Each plant category is indicated as a separate panel.

320 Correlation results between T_b and thermal niche are presented in **Figure 4**. We found a positive correlation
 321 between T_b and $\Delta T_{opt} - T_b$ values for all plant categories (**Fig 4a**). The steepness of regression slopes was the
 322 highest for forage and rangeland species (0.095; $P=0.00$; **Table 2**), followed by that of wild (0.079; $P=0.02$;
 323 **Table 2**), crops (0.063; $P=0.03$; **Table 2**), tropical tree (0.061; $P=0.04$ Table 1), horticulture (0.0596; $P=0.01$
 324 Table 1), Cactaceae and Agavaceae (0.048; $P=0.02$), and non-tropical tree (0.04; $P=0.00$; **Table 2**).

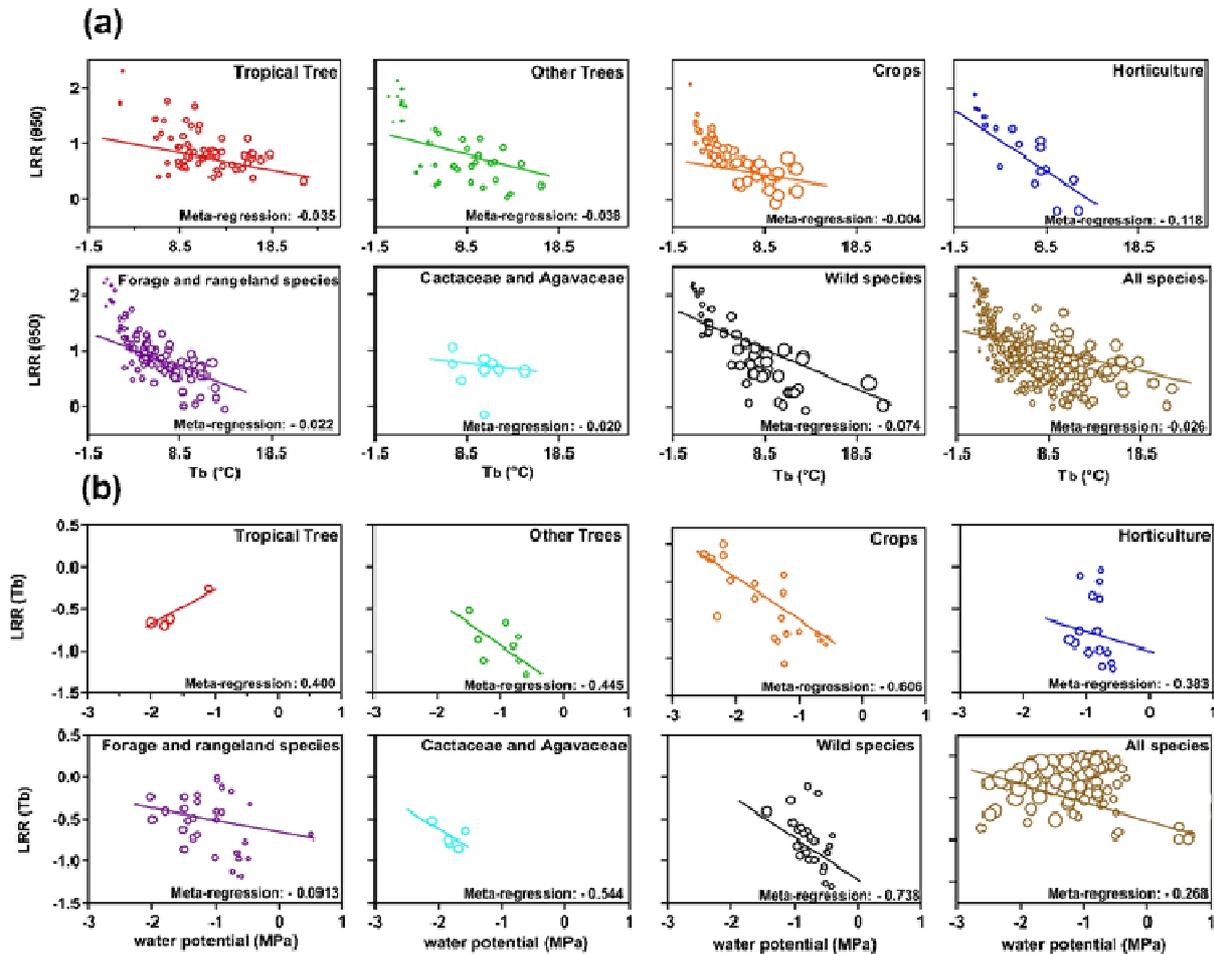
325



326

327 **Figure 4.** Regression of log risk ratio (LRR) of sub-optimal ($\Delta T_{opt}-T_b$; **a**), supra-optimal ($\Delta T_{max}-T_{opt}$; **b**), on base temperature (T_b). The size
 328 of each bubble is inversely correlated with the variance of the log relative risk estimate with larger bubbles showing more inflated
 329 variance. LRR represents the probability of changes in the range of Δ values of sub- and supra-optimal temperature as a function of
 330 changes in T_b . Fitted lines were obtained through linear regression approach ($Y=\sigma+X +$). Each plant category is indicated as a
 331 separate panel.

332 There was also a positive correlation between T_b and $\Delta T_{max}-T_{opt}$ values for all plant categories (**Fig 4b**). Other
 333 trees and wild species showed the highest steepness of regression slopes, with the slope of 0.091 and
 334 0.081($P=0.00$; **Table 2**). Followed by other trees (0.091) and wild species (0.081), the highest slope was for
 335 tropical trees (0.077) and forage and rangeland species (0.070). In contrast, the steepness of regression slopes
 336 was the lowest for crops (0.065).



337

338 **Figure 5.** Regression of log risk ratio (LRR) of thermal time required to attain 50% germination (θ_{50}) on base temperature (T_b ; **a**)
 339 and LRR of base temperature (T_b) on base water potential (Ψ_b) values (**b**). The size of each bubble is inversely correlated with the variance
 340 of the log relative risk estimate with larger bubbles showing more inflated variance. LRR represents the probability of changes in
 341 θ_{50} as a function of changes in T_b (**a**) values and in the range of T_b as a function of differing Ψ_b values (**b**). Fitted lines were obtained
 342 through linear regression approach ($Y = \theta + X + \dots$). Each plant category is indicated as a separate panel.

343 Correlation results between T_b and θ_{50} are presented in **Figure 5a**. We observed a negative relationship between
 344 T_b and θ_{50} with different meta-regression values among plant categories. The steepness of slope was the highest
 345 in horticultural species (i.e. -0.11; **Table 2**) followed by wild species (-0.07) while the lowest slope was
 346 observed for crops (-0.04). Horticultural species were followed by Cactaceae and Agavaceae, and forage and
 347 rangeland species showed a slight increase in θ_{50} as a function of T_b , with the slope steepness of -0.020 and -
 348 0.022, respectively (**Table 2**).

349 Overall, a negative relationship was found between T_b and Ψ_b values in all species included in this study, with
 350 the regression slope of -0.26 (-0.26 $P=0.00$; **Table 2**). A strong negative correlation between T_b and Ψ_b values was
 351 observed for all but tropical tree species (**Figure 5b**), with considerable difference in the steepness of regression
 352 slopes among plant categories ($P=0.03$; **Table 2**). Wild species had the lowest values of the regression slopes (-
 353 0.73; $P=0.01$; **Table 1**), followed by those of crops (-0.60; $P=0.01$), Cactaceae and Agavaceae (-0.54; $P=0.02$;
 354 **Table 1**), non-tropical trees (-0.44; $P=0.02$; **Table 2**), and horticulture species (-0.38; $P=0.01$; **Table 2**). In

355 contrast, a positive correlation between the T_b and b values was observed for tropical tree species with a slope of
356 0.40, although this plant category had few data (only 4 species).

357 4. Discussion

358 Sensitivity to environmental factors is a critical ecological process for plants occurring in unpredictable habitats
359 where successful germination is highly dependent on the spatio-temporal variability of temperature and rainfall
360 (Batlla & Benech-Arnold, 2015; Soltani *et al.*, 2017a,b; Maleki *et al.*, 2021). For 569 species of wild and
361 cultivated origin, we analysed the thermal and moisture germination niches through temperature and water
362 potential thresholds and thermal timing, and considered how these varied among different species and plant
363 categories.

364 Thermal germination niche was classified into two distinct ranges viz. sub- and supra- optimal ranges. Within
365 sub-optimal range, crops showed the widest thermal germination niche that could be a result of the selection
366 and domestication process. A wide sub-optimal temperature range increases the possibility of crop adaptations
367 across contrasted thermal environments, including fluctuating temperatures, which may allow the relative
368 synchronization of germination under known environmental and management conditions. The availability of fit-
369 for-purpose seed lots greatly reduces risks of germination failure, particularly at sub-optimal temperatures.
370 Predictable performance at low temperature means that spring crops can be sown early to escape late season
371 summer drought and winter crops sown to avoid early/late season frosts. In contrast to crops with the widest
372 thermal germination niche, species originating from warmer environments, such as tropical trees, had the
373 narrowest thermal niche within the sub-optimal temperature range. This could be due to a strong natural
374 selection and environmental drivers such that thermal niche is narrow when environmental filters are stronger
375 (Fernández-Pascual *et al.*, 2017). Consequently, when considering germination niche, habitat-specific strategies
376 should be taken into account as habitat-generalist and habitat-specialist plants can have broader and narrower
377 germination niches, respectively (Marques *et al.*, 2014). Although we did not focus on habitat-specific
378 strategies, our results highlight the need for future research on the environmental and abiotic conditions (i.e.,
379 environmental and physico-chemical) which may limit the emergence of plant species (Tudela-Isanta *et al.*,
380 2018). We also show that crops relative to tropical trees have the steepest regression slope when considering
381 the relationship between T_b and T_{opt} and Δ values, indicating that crops have higher sensitivity to sub-optimal
382 temperatures compared to tropical trees (**Figure 4a; 5a**). Such enhanced sensitivity effectively means that crop
383 seed germination is a more efficient over a range of conditions.

384 In contrast to sub-optimal temperature, the sensitivity pattern of supra-optimal temperature range was
385 considerably different, with Cactaceae and Agavaceae having the widest and crops showing the narrowest
386 supra-optimal temperature range (**Figure 2f**). This suggests that exposure of Cactaceae and Agavaceae to
387 extreme tropical dryland conditions may have facilitated population divergence through an adjustment in
388 germination behaviour as a consequence of parental environmental (Lampe *et al.*, 2017). Based on the concept
389 of parental environmental effects, offspring can take advantage of information about parental environment to
390 drive specific phenotypes for optimizing the match with the offspring environment (Lacey, 1988). This
391 ecological process requires a correlation between parental and offspring environments (Ezard *et al.*, 2014;
392 Burgess & Marshall, 2014; Leimar & McNamara, 2015). In contrast to Cactaceae and Agavaceae, crops, which
393 are often grown under optimal management conditions, have a lower capacity to tolerate warmer conditions,
394 and thus, could be subjected to higher mortality risks under climate change. An example is the reduced

395 capability of sunflower (*Helianthus annuus*) compared to its wild relatives to convert to normal seedlings at high
396 temperatures and low water potentials (Castillo-Lorenzo *et al.*, 2019a,b)

397 Our meta-regression results highlight a sensitivity pattern whereby, within both sub- and supra-optimal
398 temperature ranges, species (or groups) with lower T_b might be the most sensitive to climate change because of
399 their smaller delta values (i.e., $T_o - T_b$; $T_{max} - T_o$). This means that increases in global average temperature could
400 impact such species germination performance as the supra-optimal temperature range would be relatively
401 easier to reach. Most likely, and if genotypic limits are not exceeded, natural selection for and evolution towards
402 higher T_b and wider delta values could enable continuing emergence. Such adaptation to a potentially new (but
403 still narrower) germination niche (Fernández-Pascual *et al.* (2017) might be more likely for annual species than
404 perennials, such as trees, Cactaceae and Agavaceae.

405 The trend of supra-optimal temperature range also reflects the moisture niche dependency, with Cactaceae and
406 Agavaceae occupying broader moisture niche. Again, this may suggest that divergence in parental
407 environmental effects could have driven Cactaceae and Agavaceae toward wider moisture niche to be able to
408 capitalize on short periods of water availability under unpredictable conditions (Baskin & Baskin, 2014; Lampei
409 *et al.*, 2017) and the rapid initiation of the germination process as soil moisture increases. Overall, we show that
410 species with narrower moisture niche have a stronger relationship between T_b and Ψ_b explaining that species
411 with wider supra-optimal temperature range may have broader moisture niche, which results from a correlation
412 between parental and offspring environments (Ezard *et al.*, 2014; Burgess & Marshall, 2014; Leimar &
413 McNamara, 2015).

414 The sensitivity pattern reflected in sub- and supra-optimal temperature range is also evidence in the Ψ_b values.
415 Species with higher T_b are potentially more tolerant to drought as shown by lower Ψ_b values. For instance,
416 tropical trees show a positive correlation between T_b and Ψ_b that is probably due to a high frequency of rainfall
417 in tropical areas. However, the ongoing variability in rainfall frequency and intensity due to climate change may
418 affect tropical trees through the germination phase of the life cycle (Pritchard *et al.*, 2022).

419 Variability in T_b values among plant categories

420 Cactaceae and Agavaceae are characterized by the lowest variabilities in terms of T_b extreme values, suggesting
421 their sensitivity to lower temperature range. A negative correlation was observed between T_b and T_{50} values.
422 Annual plant species of tropical origin (e.g. cotton and mungbean), have the highest T_b values, while tree
423 species from cooler regions (e.g. such as oak, betula and ash) show the lowest T_b values. Moreover, some crops
424 (e.g. winter pea) are projected to be able to germinate at sub-zero temperatures ($T_b = -1.10$ °C), which could be
425 due to selection through breeding (Stupnikova *et al.*, 2006) Overall, we found positive correlations between T_b
426 and other traits related to cardinal temperatures suggesting that plant species alter T_b values in harmony with
427 other thermal traits as a highly efficient adaptation strategy to coping with harsh conditions.

428 Based on the results of this study, T_b for some plant categories are below zero except for Cactaceae and
429 Agavaceae and tropical trees. Germination at subzero temperatures is likely because proteins and sugars in
430 plant cell that protect cell environment from damages caused by ice formation. A recent study (Jaganathan *et al.*,
431 2020) proposed three main mechanisms for sustaining survival at subzero temperatures, including the
432 existence of water impermeable seed coats, the super-cooling of seed tissue and freezing tolerance triggered by

433 extracellular-freezing. Species included in our study might take advantage of these mechanisms to ensure their
434 survival at subzero temperatures.

435 Amplitude of T_{opt} and T_{max} values among plant categories

436 The range of T_{opt} was narrow compared to T_{max} range. The range of T_{max} ranged from 8C in forages and rangeland
437 species to 44C in crops, suggesting that crops seem to be a category that has higher T_{opt} (from 17.2C for
438 *Onobrychis subnitens* to 43C for *Sorghum bicolor*) due to domestication and selection events targeting rapid
439 germination across fluctuating environmental conditions.

440 Crops and horticultural species are likely to have higher T_{opt} and T_{max} values compared with other categories. This
441 could be driven by their selection aimed at adapting to wider environmental conditions, including hotter
442 regions of the world. For instance, horticultural species are high-value crops that are most often grown under
443 irrigated conditions that make it possible to grow them even across extremely hot conditions. Crops with
444 tropical origin, such as cotton and mungbean, have the highest T_{opt} and T_{max} values. For many species, the range
445 of T_{max} was 40-45 C, and it seems that seeds of these species are not able to progress towards completion of
446 germination above T_{max} . However, some crops are able to germinate at temperatures > 45°C (e.g. *Phaseolus*
447 *vugaris* L., *Glycine max* L., *Cicer arietinum* L. and *Sesamum indicum* L.). These values suggest that there may be
448 an upper temperature limit (T_{max}) to the germination process around 50-55°C. This limit could be imposed by
449 the onset of enzyme denaturation and activities at molecular level leading to cell death and failure of cell
450 growth. For example, lentil and pea seed amylase required for starch degradation in the early stages of
451 germination becomes inactive within minutes of exposure to 70°C and 80 °C, respectively (Tárrago & Nicolás,
452 1976; Adegbanke *et al.*, 2021).

453 This suggests that in addition to adaptation to environmental conditions within genetic limits, there might be
454 cell functional and signaling constraints that control the biological range within which seeds are able to
455 germinate. In contrast, the threshold-type response of wild species (other than Cactaceae and Agavaceae) to T_{max}
456 was weaker than that of crops and was restricted to 20-45°C. For instance, non-tropical trees such as sugar
457 maple (*Acer saccharum* M.) have the lowest T_{max} values (i.e. 14°C). Such a low T_{max} value may be confounded by
458 the presence of dormancy in this species. Species with such non-deep physiological dormancy have limited
459 cardinal temperatures at maturity, e.g. *Aesculus hippocastanum* (Pritchard *et al.*, 1999), and seeds with this
460 'conditional dormancy' progressively gain the ability to germination over a broader temperature range (Baskin
461 & Baskin, 2014; Maleki *et al.*, 2021; Soltani *et al.*, 2022). As for T_{opt} values, they increased as a function of T_b , as
462 shown by the regression slope, which may suggest coevolution of threshold-type responses to temperature that
463 need to be explored by future research.

464 Range of τ_{50} values among plant categories

465 Values of τ_{50} provide a context for adaptive strategies optimizing the efficiency of germination in relation to
466 temperature and impacting timing (Donohue *et al.*, 2010; Maleki *et al.*, 2021). Higher thermal requirement is
467 mainly linked to dormancy status (Baskin & Baskin, 2014; Maleki *et al.*, 2021), although dormancy-breaking
468 treatment may be applied to overcome seed dormancy. Tropical trees, non-tropical trees and wild species are
469 shown to have longer τ_{50} (i.e. larger thermal inputs for the same germination proportion). This could be a
470 valuable adaptation to cope with changing environments, such that germination is spread and the likelihood of
471 all seeds in a cohort dying due to unfavorable conditions is reduced. In contrast, crops showed the smallest τ_{50}

472 values, suggesting domestication has resulted in selection for fast germination and high vigour. This is also true
473 for horticultural species, particularly ornamental or medicinal species, that have been selected for a specific use
474 (provisioning, nutrients) and cropping cycle. A similar narrow range of short thermal requirements (i. e. lowest
475 Ψ_b values) for germination found in horticultural species and crops is evident in Cactaceae and Agavaceae. Rather
476 than domestication and selection being important, for the latter groupings an adaptive germination strategy in
477 response to harsh environments is likely. In this way, Cactaceae and Agavaceae can take advantage of temporary
478 suitable conditions and rapidly complete this critical step (germination) in their life cycle. Such considerations
479 support the hypothesis of a bet-hedging strategy for germination that depends on specific environmental
480 conditions (Gremer *et al.*, 2016).

481 Extent of Ψ_b values among plant categories

482 Unlike for traits related to T_b , T_{50} and cardinal temperatures, we found less literature on Ψ_b with only for 226
483 species compared to T_b and T_{opt} reported for 461 and 334 species, respectively. Nonetheless, studies published
484 in the last decade have focused more on seed Ψ_b compared with those published before 2011 (database of Dürr
485 *et al.*, 2015). Undoubtedly, with concerns growing about the wider impacts of climate change on drought, a
486 wider dataset on species' germination responses to water potential would be valuable.

487 Overall, the lowest values of Ψ_b were found for wild woody species, in particular -5.58 MPa for *Atriplex halimus*.
488 This shrub species is adapted to extreme environments, being highly drought resistant and tolerant of saline
489 conditions, and highlights one of the most important aspects of our new data: to identify model species at the
490 limits of environmental adaptability for further study.

491 Although we found 65 and 45 studies reporting Ψ_b values of forage and rangeland species, and wild species,
492 respectively, the dearth of information on tropical trees and horticultural plants (14 and papers, respectively)
493 calls for further research as germination response to water availability is one of the most important
494 environmental driver for adaptation of plant species.

495 Contrasted germination niches among plant categories

496 We found that sensitivity to environmental factors is a critical ecological process for plants occurring in
497 unpredictable habitats where successful germination is highly dependent on the spatio-temporal variability of
498 temperature. Therefore, germination niche, as a fundamental ecological indicator, should be included more in
499 future risk assessments to species survival. Environmental factors may determine germination niche through
500 regulating both seed dormancy and germination behaviour (Batlla & Benech-Arnold, 2015; Soltani *et al.*,
501 2017a,b; Maleki *et al.*, 2021). We found distinct thermal and moisture germination niche, which is defined as
502 limits characterizing species distribution, for various groups of species. Moreover, differential thermal times for
503 germination describes changes in the sensitivity of germination responses to randomly changing
504 environments, leading to various strategies adopted by plants in term of niche construction and regulation of
505 germination process. Maternal thermal environments may play a regulatory role in niche construction.
506 Cactaceae and Agavaceae, for example, have the widest supra-optimal temperature range, suggesting that
507 species experiencing higher temperatures during their lifetime, or more precisely species having experience of
508 higher temperatures, should have wider thermal niche, which is consistent with the ecological concept of
509 germination niche, in which species construct their niche in response to environmental conditions they have
510 experienced (Sultan, 2015; Fernández-Pascual *et al.*, 2019). Although Cactaceae and Agavaceae have wider

511 upper thermal niche, a recent study showed that these plants are sensitive to increasing global temperatures as
512 the thermal environments in which they inhabit are close to their upper thermal limit (Sentinella *et al.*, 2020).
513 Similarly, species facing dry environments could be generally more tolerant to drought condition by having
514 wider moisture niche as seen in *Cactaceae* and *Agavaceae*. However, in some cases, species from wetter
515 environmental conditions may be better tolerant to drought as observed in *Brassica* sp. (Castillo-Lorenzo *et al.*,
516 2019a). Interestingly, plant groups manipulated to adapt to changing environmental conditions, such as crops
517 and horticultural species, are better suited to lower temperatures by showing wider lower thermal niche.
518 However, these plants are highly sensitive to higher temperatures as shown by a narrow upper thermal niche,
519 suggesting that genetic manipulation and agronomic practices applied to these plant species (e.g. irrigation)
520 may have led to these changes.

521 **5. Conclusion**

522 In this study, we extracted data from 377 studies conducted worldwide on 569 plant species including trees,
523 grasses, crops and wild species to determine ranges of threshold-type responses to temperature and water
524 potential, as two main environmental filters, to determine relationship among these traits. Despite all the
525 complexities in collecting and interpreting data, our results show a strong relationship among ecologically
526 meaningful traits quantifying ranges of threshold-type responses to temperature and water potential and the
527 results of our meta-regression provide insight into the adaptation of various categories of species distributed
528 worldwide to the environments in which they inhabit, factors constraining adaptation and the role of
529 domestication under current and future climate. We found that, under environmental disturbances, distinct
530 germination traits expressed among and within species can allow ecosystems to persist since such adaptive
531 traits are likely to determine threshold-type values for life-history transitions, particularly germination as an
532 important determinant of plant recruitment. Because climate change is expected to become increasingly severe
533 in the future, any changes in threshold-type responses to temperature and water potential may considerably
534 influence plant function and its performance. A better understanding of functional traits and precise field-based
535 validation underlying adaptation would be useful in providing detailed information on plant regeneration.

536 **Author contributions:** JRL conceived the original idea. ES collected and extracted the data from the literature.
537 KM assembled and managed the database, analyzed the data and drafted the manuscript. JRL and ES
538 supervised the work. KM, JRL, ES, CES and HWP commented on the data analyses and revised the manuscript.
539 All authors approved the final version.

541 **Acknowledgments**

542 The authors thank Carolyne Dürr for her earlier feedback on this work. JRL received financial support from the
543 AgroEcoSystem division of INRAE.

545 **Conflicts of Interest:** The authors declare no conflict of interest

546 **Data availability**

547 The dataset has been deposited to a data repository of INRAE (Data identification number: XP3XHW_2022). A
548 version of record of the repository can be found at <https://doi.org/10.15454/XP3XHW>.

550
551

552 **References**

- 553 **Adegbanke OR, Adepomola AF, Adeniran EA, Bamidele OS. 2021.** Effect of pH and Temperature on the
554 Isolation, Purification and Characterization of α -Amylase from *Aspergillus niger* Produced from Pigeon Pea. *Acta*
555 *Scientific Nutritional Health* **5**: 62–67.
- 556 **Baskin CC, Baskin JM. 2014.** *Seeds: Ecology, Biogeography, and Evolution of Dormancy and Germination, 2nd*
557 *edn.* San Diego, CA: Elsevier/Academic Press.
- 558 **Batlla D, Benech-Arnold R. 2015.** A framework for the interpretation of temperature effects on dormancy and
559 germination in seed populations showing dormancy. *Seed Science Research* **10**: 147–158.
- 560 **Beillouin D, Ben-Ari T, Malézieux E, Seufert V, Makowski D. 2021.** Positive but variable effects of crop
561 diversification on biodiversity and ecosystem services. *Global Change Biology* **27**: 4697–4710.
- 562 **Bewley JD, Bradford K, Hilhorst H. 2012.** *Seeds: physiology of development, germination and dormancy.*
563 Springer Science & Business Media.
- 564 **Blandino C, Fernández-Pascual E, Newton RJ, Pritchard HW. 2022.** Regeneration from seed in herbaceous
565 understorey of ancient woodlands of temperate Europe. *Annals of Botany*: mcac003.
- 566 **Borenstein M, Hedges L V, Higgins JPT, R RH. 2021.** *Introduction to meta-analysis.* John Wiley & Sons.
- 567 **Burgess SC, Marshall DJ. 2014.** Adaptive parental effects: the importance of estimating environmental
568 predictability and offspring fitness appropriately. *Oikos* **123**: 769–776.
- 569 **Carta A, Fernández-Pascual E, Gioria M, Müller J V, Rivière S, Rosbakh S, Saatkamp A, Vandeloock F,**
570 **Mattana E. 2022.** Climate shapes the seed germination niche of temperate flowering plants: a meta-analysis of
571 European seed conservation data. *Annals of Botany*: mcac037.
- 572 **Castillo-Lorenzo E, Finch-Savage WE, Seal CE, Pritchard HW. 2019a.** Adaptive significance of functional
573 germination traits in crop wild relatives of Brassica. *Agricultural and Forest Meteorology* **264**: 343–350.
- 574 **Castillo-Lorenzo E, Pritchard HW, Finch-Savage WE, Seal CE. 2019b.** Comparison of seed and seedling
575 functional traits in native *Helianthus* species and the crop *H. annuus* (sunflower). *Plant Biology* **21**: 533–543.
- 576 **Catelotti K, Bino G, Offord CA. 2020.** Thermal germination niches of *Persoonia* species and projected
577 spatiotemporal shifts under a changing climate. *Diversity and Distributions* **26**: 589–609.
- 578 **Covell S, Ellis RH, Roberts EH, Summerfield RJ. 1986.** The influence of temperature on seed germination
579 rate in grain legumes: I. A comparison of chickpea, lentil, soyabean and cowpea at constant temperatures.
580 *Journal of Experimental Botany* **37**: 705–715.
- 581 **Dahal P, Bradford KJ. 1994.** Hydrothermal time analysis of tomato seed germination at suboptimal
582 temperature and reduced water potential. *Seed Science Research*.

- 583 **Dantas BF, Moura MSB, Pelacani CR, Angelotti F, Taura TA, Oliveira GM, Bispo JS, Matias JR, Silva FFS,**
584 **Pritchard HW, et al. 2020.** Rainfall, not soil temperature, will limit the seed germination of dry forest species
585 with climate change. *Oecologia* **192**: 529–541.
- 586 **Daws MI, Lydall E, Chmielarz P, others. 2004.** Developmental heat sum influences recalcitrant seed traits in
587 *Aesculus hippocastanum* across Europe. *New Phytologist* **162**: 157–166.
- 588 **Donohue K, Rubio de Casas R, Burghardt L, Kovach K, Willis CG. 2010.** Germination, Postgermination
589 Adaptation, and Species Ecological Ranges. *Annual Review of Ecology, Evolution, and Systematics* **41**: 293–319.
- 590 **Dürr C, Aubertot JN, Richard G, Dubrulle P, Duval Y, Boiffin J. 2001.** SIMPLE: a model for SIMulation of
591 PLant Emergence predicting the effects of soil tillage and sowing operations. *Soil Science Society of America*
592 *Journal* **65**: 414–442.
- 593 **Dürr C, Dickie JB, Yang XY, Pritchard HW. 2015.** Ranges of critical temperature and water potential values for
594 the germination of species worldwide: contribution to a seed trait database. *Agricultural and forest meteorology*
595 **200**: 222–232.
- 596 **Ezard THG, Prizak R, Hoyle RB. 2014.** The fitness costs of adaptation via phenotypic plasticity and maternal
597 effects. *Functional Ecology* **28**: 693–701.
- 598 **Fenner M, Thompson K. 2005.** *The Ecology of Seeds*. Cambridge: Cambridge University Press.
- 599 **Fernández-Pascual E, Carta A, Mondoni A, Cavieres LA, Rosbakh S, Venn S, Satyanti A, Guja L, Briceño**
600 **VF, Vandeloos F, et al. 2021.** The seed germination spectrum of alpine plants: a global meta-analysis. *New*
601 *Phytologist* **229**: 3573–3586.
- 602 **Fernández-Pascual E, Mattana E, Pritchard HW. 2019.** Seeds of future past: climate change and the thermal
603 memory of plant reproductive traits. *Biological Reviews* **94**: 439–456.
- 604 **Fernández-Pascual E, Pérez-Arcoiza A, Prieto JA, Díaz TE. 2017.** Environmental filtering drives the shape and
605 breadth of the seed germination niche in coastal plant communities. *Annals of Botany* **119**: 1169–1177.
- 606 **Finch-Savage WE, Rowse HR, Dent KC. 2005.** Development of combined imbibition and hydrothermal
607 threshold models to simulate maize (*Zea mays*) and chickpea (*Cicer arietinum*) seed germination in variable
608 environments. *New Phytologist* **165**: 825–838.
- 609 **Forcella F, Arnold RLB, Sanchez R, Ghersa CM. 2000.** Modeling seedling emergence. *Field Crops Research*
610 **67**: 123–139.
- 611 **Garcia-Huidobro J, Monteith JL, GR. S. 1982a.** Time, temperature and germination of pearl millet
612 (*Pennisetum typhoides* S. & H.). *Journal of Experimental Botany* **33**: 288–296.
- 613 **Garcia-Huidobro J, Monteith JL, Squire GR. 1982b.** Time, Temperature and Germination of Pearl Millet
614 (*Pennisetum typhoides* S. & H.). CONSTANT TEMPERATURE. *Journal of Experimental Botany* **33**: 288–296.
- 615 **Gardarin A, Dürr C, Colbach N. 2012.** Modeling the dynamics and emergence of a multispecies weed seed
616 bank with species traits. *Ecological Modelling* **240**: 123–138.

- 617 **Gremer JR, Chiono A, Suglia E, Bontrager M, Okafor L, Schmitt J. 2020a.** Variation in the seasonal
618 germination niche across an elevational gradient: the role of germination cueing in current and future climates.
619 *American Journal of Botany* **107**: 350–363.
- 620 **Gremer JR, Kimball S, Venable DL. 2016.** Within-and among-year germination in Sonoran Desert winter
621 annuals: bet hedging and predictive germination in a variable environment. *Ecology Letters* **19**: 1209–1218.
- 622 **Gremer JR, Wilcox CJ, Chiono A, Suglia E, Schmitt J. 2020b.** Germination timing and chilling exposure
623 create contingency in life history and influence fitness in the native wildflower *Streptanthus tortuosus*. *Journal*
624 *of Ecology* **108**: 239–255.
- 625 **Gummerson RJ. 1986.** The effect of constant temperatures and osmotic potentials on the germination of sugar
626 beet. *Journal of Experimental Botany* **37**: 729–741.
- 627 **Hardegee HP. 2006.** Predicting germination response to temperature. I. Cardinal temperature models and
628 subpopulation-specific regression. *Annals of Botany* **97**: 1115–1125.
- 629 **Hartung J, Knapp G, Sinha BK. 2008.** Statistical Meta-Analysis with Applications. In: John Willey & Sons, p.
630 127–137.
- 631 **Jaganathan GK, Dalrymple SE, Pritchard HW. 2020.** Seed Survival at Low Temperatures: A Potential
632 Selecting Factor Influencing Community Level Changes in High Altitudes under Climate Change. *Critical*
633 *Reviews in Plant Sciences* **39**: 479–492.
- 634 **Lacey EP. 1988.** What is an adaptive environmentally induced parental effect. In: Mousseau TA, Fox CW, eds.
635 *Maternal Effects as Adaptations*. New York: Oxford University Press, 54–66.
- 636 **Lamichhane JR, Constantin J, Schoving C, Maury P, Debaeke P, Aubertot J-N, Dürr C. 2020.** Analysis of
637 soybean germination, emergence, and prediction of a possible northward establishment of the crop under
638 climate change. *European Journal of Agronomy* **113**: 125972.
- 639 **Lamichhane JR, Wojciechowski A, Bourgeois C, Debaeke P. 2022.** Genetic variability for early growth traits
640 in second season sunflower. *Frontiers in Agronomy* **4**: 822456.
- 641 **Lampe C, Metz J, Tielbörger K. 2017.** Clinal population divergence in an adaptive parental environmental
642 effect that adjusts seed banking. *New Phytologist* **214**: 1230–1244.
- 643 **Leimar O, McNamara JM. 2015.** The Evolution of Transgenerational Integration of Information in
644 Heterogeneous Environments. *The American Naturalist* **185**: E55–E69.
- 645 **Maleki K, Soltani E, Arabhoseini A, Lakeh AM. 2021.** A quantitative analysis of primary dormancy and
646 changes during burial in seeds of *Brassica napus*. *Nordic Journal of Botany* **39**.
- 647 **Marques AR, Atman APF, Silveira FAO, de Lemos-Filho JP. 2014.** Are seed germination and ecological
648 breadth associated? Testing the regeneration niche hypothesis with bromeliads in a heterogeneous neotropical
649 montane vegetation. *Plant Ecology* **215**: 517–529.

- 650 **Ordoñez-Salanueva CA, Orozco-Segovia A, Mattana E, Castillo-Lorenzo E, Davila-Aranda P, Pritchard HW,**
651 **Ulian T, Flores-Ortiz CM. 2021.** Thermal niche for germination and early seedling establishment at the leading
652 edge of two pine species, under a changing climate. *Environmental and Experimental Botany* **181**: 104288.
- 653 **Orrù M, Mattana E, Pritchard HW, Bacchetta G. 2012.** Thermal thresholds as predictors of seed dormancy
654 release and germination timing: altitude-related risks from climate warming for the wild grapevine *Vitis vinifera*
655 subsp. *sylvestris*. *Annals of Botany* **110**: 1651–1660.
- 656 **Porceddu M, Mattana E, Pritchard HW, Bacchetta G. 2013.** Thermal niche for in situ seed germination by
657 Mediterranean mountain streams: model prediction and validation for *Rhamnus persicifolia* seeds. *Annals of*
658 *Botany* **112**: 1887–1897.
- 659 **Pritchard HW, KR. M. 1990.** Quantal response of fruit and seed germination rate in *Quercus robur* L. and
660 *Castanea sativa* Mill., to constant temperatures and photon dose. *Journal of Experimental Botany* **41**: 1549–
661 1557.
- 662 **Pritchard HW, Ser-shen, Tsan FY, Wen B, Jaganathan GK, Calvi G, Pence VC, Mattana E, Ferraz IDK, Seal**
663 **CE. 2022.** Chapter 19 - Regeneration in recalcitrant-seeded species and risks from climate change. In: Baskin
664 CC, Baskin JM, eds. *Plant Regeneration from Seeds*. Academic Press, 259–273.
- 665 **Pritchard HW, Steadman KJ, Nash J V, Jones C. 1999.** Kinetics of dormancy release and the high
666 temperature germination response in *Aesculus hippocastanum* seeds. *Journal of Experimental Botany* **50**:
667 1507–1514.
- 668 **Seal CE, Daws MI, Flores J, Ortega-Baes P, Galíndez G, León-Lobos P, Sandoval A, Ceroni Stuva A,**
669 **Ramírez Bullón N, Dávila-Aranda P, et al. 2017.** Thermal buffering capacity of the germination phenotype
670 across the environmental envelope of the Cactaceae. *Global Change Biology* **23**: 5309–5317.
- 671 **Sentinella AT, Warton DI, Sherwin WB, Offord CA, Moles AT. 2020.** Tropical plants do not have narrower
672 temperature tolerances, but are more at risk from warming because they are close to their upper thermal limits.
673 *Global Ecology and Biogeography* **29**: 1387–1398.
- 674 **Soltani E, Baskin CC, Baskin JM. 2017a.** A graphical method for identifying the six types of non-deep
675 physiological dormancy in seeds. *Plant Biology* **10**: 673–682.
- 676 **Soltani E, Gruber S, Oveisi M, Salehi N, Alahdadi I, Javid MG. 2017b.** Water stress, temperature regimes
677 and light control induction, and loss of secondary dormancy in *Brassica napus* L. seeds. *Seed Science Research*
678 **10**: 217–230.
- 679 **Soltani E, Maleki K, Heshmati S. 2022.** Application of a process-based model to quantifying dormancy loss in
680 seeds of *Parrotia persica* C.A. Meyer. *South African Journal of Botany* **144**: 97–104.
- 681 **Stupnikova I, Benamar A, Tolleter D, Grelet J, Borovskii G, Dorne A-J, Macherel D. 2006.** Pea seed
682 mitochondria are endowed with a remarkable tolerance to extreme physiological temperatures. *Plant Physiol.*
683 **140**: 326–335.

- 684 **Sultan SE. 2015.** *Organism and environment: Ecological development, niche construction, and adaption.* USA:
685 Oxford University Press.
- 686 **Tárrago JF, Nicolás G. 1976.** Starch Degradation in the Cotyledons of Germinating Lentils. *Plant Physiology*
687 **58:** 618–621.
- 688 **Thompson K, Ceriani RM. 2003.** No relationship between range size and germination niche width in the UK
689 herbaceous flora. *Functional Ecology* **17:** 1365–2435.
- 690 **Trudgill DL, Honek A, Li D, van Straalen NM. 2005.** Thermal time – concepts and utility. *Annals of Applied*
691 *Biology* **146:** 1–14.
- 692 **Tudela-Isanta M, Ladouceur E, Wijayasinghe M, Pritchard HW, Mondoni A. 2018.** The seed germination
693 niche limits the distribution of some plant species in calcareous or siliceous alpine bedrocks. *Alpine Botany*
694 **128:** 83–95.
- 695 **Waha K, Dietrich JP, Portmann FT, Siebert S, Thornton PK, Bondeau A, Herrero M. 2020.** Multiple
696 cropping systems of the world and the potential for increasing cropping intensity. *Global Environmental Change*
697 **64:** 102131.
- 698 **Walck JL, Hidayati SN, Dixon KW, Tthompson KEN, Poschlod P. 2011.** Climate change and plant
699 regeneration from seed. *Global Change Biology* **17:** 2145–2161.
- 700 **Zhang R, Luo K, Chen D, Baskin J, Baskin C, Wang Y, Hu X. 2020.** Comparison of Thermal and Hydrotime
701 Requirements for Seed Germination of Seven Stipa Species From Cool and Warm Habitats. *Frontiers in Plant*
702 *Science* **11.**