

### The importance of myo-inositol and D-chiro-inositol to support fertility and reproduction

Fabio Facchinetti, Didier Dewailly, Christophe Soulage, Vittorio Unfer

### ► To cite this version:

Fabio Facchinetti, Didier Dewailly, Christophe Soulage, Vittorio Unfer. The importance of myoinositol and D-chiro-inositol to support fertility and reproduction. Médecine Thérapeutique. Médecine de la Reproduction, Gynécologie, Endocrinologie, 2021, 23 (3), pp.213-221. hal-03789151v2

### HAL Id: hal-03789151 https://hal.inrae.fr/hal-03789151v2

Submitted on 27 Sep 2022

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés. See discussions, stats, and author profiles for this publication at: https://www.researchgate.net/publication/355105046

The importance of myo-inositol and D-chiro-inositol to support fertility and reproduction L'importance du myo-inositol et du D-chiro-inositol pour soutenir la fertilité et la repro...

Article · October 2021				
DOI: 10.1684/mte.2021.0855				
TATIONS		READS		
)		345		
authors:				
F	Fabio Facchinetti		Didier Dewailly	
	Università degli Studi di Modena e Reggio Emilia		Centre Hospitalier Régional Universitaire de Lille	
7	729 PUBLICATIONS 15,311 CITATIONS		493 PUBLICATIONS 27,302 CITATIONS	
	SEE PROFILE		SEE PROFILE	
	Christophe O Soulage	100	Vittorio Unfer	
<b>1</b>	Institut National des Sciences Appliquées de Lyon		${\tt UniCamillus  International  University  of  {\tt Health  and  Medical  Sciences}}$	
1	129 PUBLICATIONS 3,206 CITATIONS		220 PUBLICATIONS 4,180 CITATIONS	
ſ	SEE PROFILE		SEE PROFILE	

Some of the authors of this publication are also working on these related projects:

Project AMH in PCOS View project

Ozone-induced insulin resistance View project

### **Review**

Médecine de la Reproduction 2021; 23 (3): 213-221

# The importance of myo-inositol and D-chiro-inositol to support fertility and reproduction

L'importance du myo-inositol et du D-chiro-inositol pour soutenir la fertilité et la reproduction

Fabio Facchinetti<sup>1,2</sup> Didier Dewailly<sup>2,3</sup> Christophe O. Soulage<sup>2,4</sup> Vittorio Unfer<sup>2,5</sup>

for EGOI\* (The Experts Group on Inositol in Basic, Clinical Research)<sup>a</sup>

- <sup>1</sup> Mother-Infant Department, University of Modena and Reggio Emilia, Modena, Italy <fabio.facchinetti@unimore.it>
- The Experts Group on Inositol in Basic and Clinical Research (EGOI)
- <fabio.facchinetti@unimore.it> <sup>3</sup> Faculty of Medicine, University of Lille, Lille, France

University of Lyon, Inserm U1060, Car-MeN, INSA de Lyon, Université Claude Bernard Lyon 1, Villeurbanne, France Systems Biology Group Lab, Rome, Italy

<sup>a</sup> The Experts Group on Inositol in Basic and Clinical Research (EGOI): Vittorio Unfer, Systems Biology Group Lab, Rome, Italy; Fabio Facchinetti, Mother-Infant Department, University of Modena and Reggio Emilia, Modena, Italy/Department of Obstetrics and Gynecology and Pediatrics, University of Modena and Reggio Emilia, Modena, Italy; Appetecchia Marialuisa, Oncological Endocrinology Unit, Regina Elena National Cancer Institute, IRCCS, Rome, Italy; Aragona Cesare, Systems Biology Group Lab, Rome, Italy; Barbaro Daniele, Director of U. O. Endocrinology in Livorno Hospital, USL Nordovest Toscana Italy. Professor at contract University of Pisa; Benvenga Salvatore, Department of Clinical and Experimental Medicine, University of Messina, Italy; Bevilacqua Arturo, Department of Dynamic and Clinical Psychology, Sapienza University, Rome, Italy; Bezerra Espinola Maria Abstract. This review details the physiologic roles of two insulin sensitizers, myo-inositol (MI) and D-chiro-inositol (DCI). In the human ovary, MI is a second messenger of follicle stimulating hormone (FSH) and DCI is an aromatase inhibitor. These activities allow a treatment for polycystic ovary syndrome (PCOS) to be defined based on the combined administration of MI and DCI, where the best MI:DCI ratio is 40:1. In addition, MI plays a pivotal role in the physiology of reproduction, and has beneficial effects on the development of oocytes, spermatozoa, and embryos. By contrast, DCI has little effect on spermatozoa, but high concentrations in the ovary can negatively affect the quality of oocytes and the blastocyst. Overall, the evidence in the literature supports the beneficial effects of MI in both female and male reproduction, warranting clinical use of MI in assisted reproductive treatment (ART).

Key words: myo-inositol, D-chiro-inositol, polycystic ovary syndrome, fertility, reproduction, 1V/F

Résumé. Cette revue détaille les rôles physiologiques de deux sensibilisateurs à l'insuline, le myo-inositol (MI) et le D-chiro-inositol (DCI). Dans l'ovaire humain, le MI est un second messager de l'hormone folliculostimulante (FSH) et le DCI est un inhibiteur de l'aromatase. Ces activités permettent de définir un traitement du syndrome des ovaires polykystiques (SOPK) basé sur l'administration combinée de MI et de DCI, où le meilleur rapport MI:DCI est de 40:1. En outre, le MI joue un rôle essentiel dans la physiologie de la reproduction et a des effets bénéfiques sur le développement des ovocytes, des spermatozoïdes et des embryons. En revanche, le DCI a peu d'effet sur les spermatozoïdes, mais des concentrations élevées dans l'ovaire peuvent avoir un effet négatif sur la qualité des ovocytes et du blastocyste. Dans l'ensemble, les données de la littérature confirment les effets bénéfiques du MI dans la reproduction féminine et masculine, ce qui justifie l'utilisation clinique du MI dans l'assistance médicale à la procréation.

Mots clés : myo-inositol , D-chiro-inositol , syndrome des ovaires polykystiques , fertilité , reproduction, fécondation in vitro

#### Synthesis and activities of MI and DCI

Inositols are cyclic polyols which are among the most ancient molecules on Earth. They can be found in 9 stereoisomers [1-3] and MI and DCI are the most prevalent.

In the human body, MI is actively synthesized in the kidneys, liver, testes, mammary gland, brain [4, 5].

Under insulin stimulation, a specific epimerase converts MI to DCI [6, 7].

Endogenously, the production of both stereoisomers depends on the specific tissue requirements [8].

As such, in healthy women the plasma MI:DCI ratio is 40:1 [9], whereas in ovarian follicular fluid is close to 100:1 [10].

#### MI and DCI activities

MI and DCI are deeply involved in insulin signaling, since insulin requires the presence of both stereoisomers to exert its functions. As inositolphosphoglycans, MI and DCI are second messengers in the insulin signaling, mediating different effects [3, 11-13]. Notably, being the two stereoisomers metabolically linked to

To cite this article: Facchinetti F, Dewailly D, Soulage CO, Unfer V. The importance of myo-inositol and D-chiro-inositol to support fertility and reproduction. Médecine de la Reproduction 2021; 23 (3): 213-221. doi: 10.1684/mte.2021.0855

each other, a drastic separation of their individual effects in vivo can be challenging. However, while MI mainly controls cellular glucose uptake, and its content is significantly higher in tissues with high-glucose utilization, like brain, heart, and ovaries [13-16], DCI is principally involved in glucose storage as glycogen.

Likely interfering with glucose intestinal uptake, MI seems to prevent glucose absorption at the duodenal level and decrease glucose rise in the blood [17]. Furthermore, MI improves insulin sensitivity in adipocytes by increasing lipid storage and glucose uptake, and by inhibiting lipolysis [18]. It also downregulates the inflammatory response, particularly in macrophages, probably through the inhibition of proinflammatory transcription factors [19].

#### Inositols in the ovary and in pregnancy

In the ovary MI (as InsP3) is one of the second messengers of FSH [20]. Its concentration in the mammalian female reproductive tract is higher than in

blood serum, suggesting that MI plays critical roles at the ovarian level, like ensuring correct oocyte maturation [21].

Instead, DCI down-modulates the activity of aromatase enzyme, found in fat tissue, ovaries, testicles, placenta, brain, bone [22]. Indeed, DCI decreases the expression of aromatase gene dose-dependently and consequently inhibits testosterone to estrogen conversion [23]. Furthermore, as insulin mediator, DCI stimulates testosterone biosynthesis by theca cells [24]. This effect is more marked in PCOS women compared to normal ones and contributes to explain the higher amounts of testosterone produced in PCOS patients, in comparison with healthy subjects [24].

Each organ balances the intracellular levels of inositols to achieve tissue-specific intracellular MI/DCI ratios that modulate metabolic processes [10]. In the ovaries, such ratio is about 100:1 [10].

The excess of DCI in ovarian follicles is potentially detrimental in some cases, as demonstrated by Ravanos *et al.* [25]. The authors observed that MI positively correlated with quality of blastocysts, while DCI concentrations above the MI/DCI limit ratio of 70:1 in follicular fluid decreased blastocyst quality [25].

Salomé, Systems Biology Group Lab, Rome, Italy; Bizzarri Mariano, Department of Experimental Medicine, Systems Biology Group, University La Sapienza, Rome, Italy; Systems Biology Group Lab, Rome, Italy; Cantelmi Tonino, Institute for Interpersonal Cognitive Therapy, Rome, Italy; Cavalli Pietro, Humanitas Research Hospital, Rozzano, Milan, Italy; Chan Shiao-Yng, Department of Obstetrics and Ginecology, Yong Loo Lin School of Medicine, National University of Singapore, Malaysia; Chiu Tony Tak Yu, IVF Centre, Hong Kong, China; Copp Andrew J., Newlife Birth Defects Research Centre and Developmental Biology and Cancer Programme, Institute of Child Health, University College London, London, UK; D'Anna Rosario, Department of Human Pathology, University of Messina, Messina, Italy; Dewailly Didier, Faculty of Medicine, University of Lille, Lille, France; Di Lorenzo Cherubino, Researcher at Department of Medico-Surgical Sciences and Biotechnologies, Sapienza University of Rome Polo Pontino, Latina, Italy; Diamanti-Kandarakis Evanthia, Department of Endocrinology and Diabetes, HYGEIA Hospital, Marousi, Athens, Greece; Dinicola Simona, Systems Biology Group Lab, Rome, Italy; Greene Nicholas D., Newlife Birth Defects Research Centre and Developmental Biology and Cancer Programme, Institute of Child Health, University College London, London, UK; Hernández Marín Imelda, Human Reproduction Department, Hospital Juárez de México, and Universidad Nacional Autónoma de México (UNAM), México City, México; Hod Moshe, Department of Obstetrics and Gynecology Sackler Faculty of Medicine, Tel-Aviv University, Israel; MOR Comprehensive Women's Health Center, Tel-Aviv, Israel; Kamenov Zdravko, Department of Internal Medicine, Medical University of Sofia, Sofia, Bulgaria; Kandaraki Eleni A., Department of Endocrinology and Diabetes, HYGEIA Hospital, Marousi, Athens, Greece; Laganà Antonio Simone, Department of Obstetrics and Gynecology, 'Filippo Del Ponte' Hospital, University of Insubria, Varese, Italy; Monastra Giovanni, Systems Biology Group Lab, Rome, Italy; Montanino Oliva Mario, Department of Obstetrics and Gynecology, Santo Spirito Hospital, Rome, Italy; Nestler John E., Division of Endocrinology, Diabetes and Metabolism, Department of Internal Medicine, Virginia Commonwealth University, Richmond, VA, USA; Nordio Maurizio, A.S.L. RMF, Civitavecchia (RM), Italy; Özay Ali Cenk, Near East University, Faculty of Medicine, Department of Obstetrics and Gynecology, and Research Center of Experimental Health Sciences, Nicosia, Cyprus; Papalou Olga, Department of Endocrinology and Diabetes, HYGEIA Hospital, Marousi, Athens, Greece; Pkhaladze Lali, Zhordania and Khomasuridze Institute of Reproductology, Tbilisi, Georgia; Porcaro Giuseppina, Women's Health Centre, USL UMBRIA 2, Terni, Italy; Prapas Nikos, Third Department of OB-GYNAE, Aristotle University of Thessaloniki, and IVF Laboratory, IAKENTRO Fertility Centre, Thessaloniki, Greece; Roseff Scott J., Reproductive Endocrinology and Infertility, South Florida Institute for Reproductive Medicine (IVFMD), Florida, USA; Soulage Christophe O., University of Lyon, INSERM U1060, CarMeN, INSA de Lyon, Université Claude Bernard Lyon 1, Villeurbanne, France; Stringaro Annarita, National Center for Drug Research and Evaluation, Italian National Institute of Health, Rome, Italy; Vazquez-Levin Mónica Hebe, National Council of Scientific and Technical Research, Instituto de Biología y Medicina Experimental (IBYME), Buenos Aires, Argentina; Vucenik Ivana, Department of Medical & Research Technology and Pathology, University of Maryland School of Medicine in Baltimore, MD, USA; Wdowiak Artur, Diagnostic Techniques Unit, Medical University of Lublin, Poland

# Treatment with MI in impaired female fertility (PCOS and other conditions)

The first study (1992) on the role of MI in IVF found a positive correlation between serum concentrations of MI and successful IVF pregnancy. Serum samples with high MI content showed clear trophic properties when added to in vitro embryo cultures, and supported better post-implantation development of mouse embryos [26]. Subsequent studies showed that high MI levels in human follicular fluid correlate positively with satisfactory oocyte quality [27]. Moreover, addition of MI to the culture medium stimulated meiotic progression by mouse germinal vesicle oocytes, a process that requires intracellular calcium mobilization [28]. On the other hand, Goud and coworkers demonstrated that inositol-1,4,5-trisphosphate plays an important physiological role during in vitro maturation, fertilization, and early cleavages of human oocytes and embryos [29]. PCOS women undergoing ART are a particularly challenging target group and have been the subject of several recent studies. Wdowiak [30] investigated the activity of oral MI in PCOS women enrolled for intracytoplasmic sperm injection (ICSI). Over the 3 months before ICSI, 60 control PCOS patients received 200 µg of folic acid twice per day; the remaining 52 PCOS subjects were treated with 2 g of MI plus 200 µg folic acid, also twice per day. Further controls (no treatment) were 105 healthy women. A significant difference in the number of pregnancies was found: pregnancy was reported in 34.62% of the MI treated group but in only 20% of the PCOS controls.

In another IVF clinical trial [31], 133 PCOS and 137 non-PCOS women with preserved ovarian reserve were treated daily by the oral route for 3 months during the preconception period and ovarian stimulation. The first group (PCOS) received 1 g MI plus 400  $\mu$ g folic acid, whereas the second group (non-PCOS) received 400  $\mu$ g folic acid plus 2  $\mu$ g cyanocobalamin. The total number of mature oocytes in the MI-treated patients was significantly higher than in the second group. Pregnancy rates per embryo transfer, 'take home baby' index, and miscarriage rates were comparable. In conclusion, MI improves oocyte quality, thus potentially improving IVF outcome [31].

# MI reduces the amount of gonadotropin in IVF procedures

Emekçi Özay et al. [32] administered 4 g MI plus 400  $\mu$ g folic acid, before and during controlled ovarian hyperstimulation (COH) with recombinant (r)FSH and

intrauterine insemination (IUI), to 98 infertile PCOS women undergoing controlled ovulation induction and IUI. Controls (n = 98) received rFSH and 400 µg folic acid. Of the treated subjects, nine accomplished spontaneous pregnancy. During COH + IUI treatment three cycles were canceled in the study group and eight in the control group. In the treated patients, a significant decrease in total rFSH dose and cycle duration was reported; in addition, clinical pregnancy rate was higher (18.6%) in patients receiving MI compared with controls (12.2%) [32]. Systematic review and meta-analysis, including eight randomized controlled trials (RCTs) with a total of 812 participants [33], confirmed that oral inositol supplementation during controlled ovarian stimulation (COS) and ART can reduce the total amount of gonadotropins used and the length of COS in both PCOS and non-PCOS women undergoing IVF. According to the analysis, MI was effective in reducing gonadotropin administration in both PCOS and non-PCOS women. However, MI supplementation decreased the length of COH only in PCOS women. Although the data from the current literature do not take pharmacoeconomic aspects into account, based on the above results the authors assert that MI therapy may significantly reduce the overall cost of IVF procedures, with direct benefits for the patients [33]. Zheng and colleagues [34] performed systematic literature review and meta-analysis concerning the efficacy of MI administration to infertile (non-PCOS) patients undergoing ovulation induction for ICSI or IVF and embryo transfer (IVF-ET). Seven trials, with a total of 935 women, were taken in consideration. MI treatment was associated with a significantly higher clinical pregnancy rate and proportion of grade 1 embryos. The abortion rate in the MI group was significantly lower than in controls. Furthermore, the study group required significantly fewer total units of gonadotropins such as rFSH compared with controls. MI therefore increases both the clinical pregnancy rate in infertile women undergoing assisted motherhood and the quality of embryos, as well as reducing the number of unsuitable oocytes and the amount of stimulation drugs required [34]. Further evaluation of MI supplementation, in a larger cohort of patients, will be necessary to assess its economic advantages in IVF treatments and its effect on long-term fertility outcomes.

#### MI and embryo development in vitro

Preimplantation mouse embryos supplemented in vitro with MI exhibit (i) increased percentage of progression to the most advanced stage of development; (ii) overall increased percentage of development to the expanded blastocyst stage; (iii) increased average number of blastomeres forming the embryos at the blastocyst stage [35]. A plausible mechanism may include rapid incorporation of MI into phosphatidylinositides (PtdIns) and increased production of intracellular second messengers that enhance proliferation [36, 37]. In particular, MI supplementation of the culture medium of late preimplantation mouse embryos induced Akt phosphorylation at serine 473 [38]. This demonstrates that, in the early stages of development, new phosphorylation of Akt occurs in the mid-to-late preimplantation stages, and this process depends on the availability of MI. Previous evidence showed that the development of preimplantation mouse embryos (8-16 cell stage) requires the activity of PI3K, an enzyme that produces PIP3 from PtdIns [39, 40]. The increased synthesis of phosphoinositides [41], and the resulting increase in PIP3, may account for these observations. In conclusion, enhanced phosphorylation of Akt in the presence of MI may be responsible for the faster development rate of cultured embryos. Reasonably, MI supplementation to dividing blastomeres enhances the pathway leading to Akt phosphorylation and accelerates development. In conclusion, the direct effects of MI on oocyte health and subsequent development may improve ART outcome.

#### Inositols for the treatment of PCOS

#### **Pre-clinical evidence**

An important role of MI and DCI in PCOS patients is an insulin sensitizing action, mirrored by a decrease in the homeostatic model assessment (HOMA) index. Both isomers are useful in treating insulin resistance states [12, 42]. Bevilacqua et al. [43] recently carried out a preclinical study on an animal model of PCOS. Female mice received continuous light (L/L) for 10 weeks. At the end of this period, they developed a phenotype with several similarities to that of PCOS women. A group of mice kept under normal (12/12 h) light/dark cycle (L/D) served as control. The uteri of L/D mice had a proestrus/ estrus-like appearance, as normally found in sexually mature, cycling animals. Instead, the uteri of L/L mice exhibited immature/diestrus-like features, typical of noncycling animals. Ovaries from control mice presented a corpus albicans (from recent ovulations) and a corpus luteum and showed normal primary, secondary, and tertiary follicles upon histological analysis. On the contrary, ovaries from L/L mice were smaller, without corpus albicans and, upon histological analysis, revealed

paucity of primary and secondary follicles and cystic tertiary follicles that strongly resembled those in human polycystic ovaries. Such cystic follicles lacked the oocyte and presented variable amounts of granulosa cells. The early tertiary follicles with a living oocyte presented a hyperplastic theca cell layer and a thinner granulosa cell sheet. The ratio between the thickness of these two layers (TGR) allows reliable evaluation of the androgenic-like phenotype that typically occurs in PCOS [44]. In fact, a hypertrophic theca cell layer is a hallmark of polycystic ovaries and causes a greater production of androgens [45]. The study provided the first experimental evidence of the different efficacy exerted by various MI/DCI ratios (5:1; 20:1; 40:1; 80:1) in restoring a normal phenotype. Moreover, it supported the metabolic link between MI and DCI, specifically in PCOS. Mice treated daily with 420 mg/kg MI/DCI in a 40:1 molar ratio made a fast and almost full recovery from PCOS signs and symptoms. On the contrary, the other MI/DCI ratios were less effective or had even negative effects. In particular, the formulation with high content of DCI proved to worsen the PCOS pathological features.

#### **Clinical evidence**

Recently, a clinical trial confirmed these findings also in PCOS women [46]. Since the "ovarian paradox" [47] postulated that ovaries, unlike liver and muscles, never become insulin resistant [48-50], the hyperinsulinemia in PCOS women enhances ovarian MI to DCI epimerization, increasing DCI concentration at the expense of MI [7, 10, 47].

Indeed, while healthy women show an ovarian MI:DCI ratio around 100:1, in PCOS women it drops to 0.2:1 [10], likely affecting the FSH signaling. Therefore, restoring the physiological MI:DCI ratio in the follicular fluid may be crucial for proper ovarian function [10].

In a recent meta-analysis [51] 9 randomized controlled trials (RCTs) on PCOS patients were evaluated, with a total of 247 cases and 249 controls [32, 52-59].

The authors determined the efficacy of treatments (length: 12-24 weeks) with MI, alone, or in association with DCI in the 40:1 ratio, on fasting insulin, HOMA index and serum levels of testosterone, androstenedione, and sex hormone-binding globulin (SHBG). Inositol supplementation significantly reduced fasting insulin and HOMA index, slightly decreasing testosterone with respect to controls. After at least 24 weeks of administration, MI significantly increased SHBG levels.

Since high doses of DCI/day may be detrimental for ovaries and oocyte maturation, the authors recommend

avoiding exclusive DCI supplementation, also considering that DCI cannot be converted into MI and that deficiencies of MI correlate with insulin-resistance conditions.

On the contrary, the meta-analysis strongly supports MI supplementation for improving the metabolic profile of PCOS patients. Also, a systematic review and metaanalysis [60], including 10 RCTs and 573 patients further confirmed these results [52-55, 58, 59, 61-63].

Therefore, inositols can be recommended for managing PCOS with insulin resistance, as well as for improving symptoms caused by decreased estrogen in PCOS [60]. The best therapeutic regimen, clinically tested in women with PCOS, is the oral combination of MI and DCI in a molar ratio of 40:1.

The optimal daily amount of inositols is 4 g divided in two administrations, for at least 3 or 6 months.

## Effects of inositols in the ovary and impact on pregnancy in PCOS women

As mentioned, the polycystic ovaries exhibit specific MI depletion and DCI overload [10], with impaired FSH signaling and poor-quality oocytes [64]. Therefore, a probable treatment may be to restore the physiological levels of the two isomers in the FF and reestablish proper ovarian functioning [10]. In a clinical study [58], 46 obese PCOS women (BMI N 30) received combined MI and DCI (40:1 ratio) for 6 months. The authors observed improved insulin sensitivity and ovulatory function, along with decreased luteinizing hormone (LH) and free testosterone levels. The lower LH/FSH ratio subsequently reduces the observed hyperandrogenism. The authors also reported significantly reduced HOMA index and fasting insulin and significantly increased E2 and SHBG. The overall improved hormonal status restored the ovulation, without observed side effects. On the contrary, the placebo group reported no relevant changes in the levels of sex hormones [58, 65]. The content of MI in human FF seems to play a role in follicular maturity and high concentrations represent a potential marker of good oocyte quality. As previously reported, studies have demonstrated that increased MI content improves the quality of blastocysts, while excess DCI has deleterious effects [25]. Indeed, DCI increases testosterone levels through two different pathways: in the theca cells from PCOS women as insulin mediator (inositolglycan mediator) [24] and in the granulosa cells as aromatase inhibitor [23]. Such evidence could provide a plausible explanation for the higher amounts of testosterone in women suffering PCOS, as compared with healthy individuals. In summary, rebalancing the hormonal status and the metabolic parameters is beneficial to reproductive outcomes in humans, enhancing oocyte health and ovulatory function. The importance of preserving the balance between MI and DCI concentration in FF is also highlighted in the trials that used combined treatment of MI and DCI. Indeed, the physiologic ratio appeared to optimize the improvement of fertility [13]. In addition, literature data indicate that MI signaling may regulate the AMH production induced by FSH in the granulosa cells [66]. AMH decreases oocyte sensitivity to FSH and participates in regulating follicle maturation. Treatment with MI in in vitro fertilization (IVF) allows a decrease in the amount of recombinant FSH administered and in the duration of the ovulation induction for follicular development [33] and an increase in the clinical pregnancy rate [32].

#### Association with CC

Clomiphene citrate (CC) is an antiestrogenic compound, used as first-choice drug in the therapy for oligo-anovulatory infertility. Although some patients are resistant, CC was widely used in PCOS women to induce ovulation because it increases the pituitary production of FSH and LH [67]. Researchers investigated inositol treatment, combined with CC, to assess possible fertility improvements in PCOS women seeking pregnancy [68]. In the study, 50 anovulatory PCOS patients received MI for three spontaneous cycles. If they remained anovulatory and/or failed to achieve pregnancy, they received a combination of MI and CC in the following three cycles. MI improved ovarian activity in PCOS women, as spontaneous ovulation occurred in 61.7% of patients, while 72.2% of MI-resistant subjects eventually ovulated after clomiphene treatment. A recent pilot study [69] further demonstrated that the combination of MI and CC significantly increases the ovulation rate, decreases the rate of resistance to CC, and improves the pregnancy rate. The study shows a potential benefit for MI supplementation during CC ovulation induction PCOS patients, even though the results failed to reach statistical significance for most outcomes, probably because of the small number of patients. However, further studies are required to draw more definite conclusions [70]. A double-blinded, randomized, and controlled trial is currently recruiting patients with PCOS seeking pregnancy and eligible to simple ovulation induction by CC. Half of them will receive MI + folic acid in addition to CC, whereas the other half will receive a placebo containing only folic acid, in addition to CC.

#### Overcoming issues in inositol therapy

#### **Reduced** absorption of inositol

Competition with DCI or interference of other molecules (e.g., glucose) on the transport mechanism may cause a reduced absorption of MI. Competition for the

#### Review

same transporter may cause insufficient MI passage across the intestinal barrier or inside the cells. This condition occurs when a competitor has affinity for the transporter greater than MI, or when a competitor has lower affinity but large concentration to displace MI. Two groups of inositol transporters exist, with different tissue distribution: sodium/ myo-inositol cotransporter 1 and 2 (SMIT1 and SMIT2), coupled with sodium ions; proton/myo-inositol cotransporter (HMIT), coupled with protons. Both of them are expressed in several tissues and organs (kidneys, brain, placenta, pancreas, heart, skeletal muscles, lungs, liver, intestine, adipose tissue, and oocytes) [71]. To date, SMIT2 is the only known transporter of MI located in the intestine (duodenum and jejunum). In vitro experiments identified a Km (DCI) lower than Km (MI), hence DCI transport is slightly favored. To avoid enhanced intestinal absorption of DCI at the expense of MI, determining the proper MI: DCI ratio to administer to patients is essential. The abovementioned 40:1 ratio proved to be optimal. MI and DCI have a much greater affinity (more than 100 times) than glucose for the transporter [72, 73].

#### **Inositol resistance**

Some patients exhibit a reduced intestinal absorption of inositol and are defined "inositol-resistant". Researchers tried to overcome this problem combining MI with the whey protein alpha-lactalbumin ( $\alpha$ -LA), already known for its propriety as carrier for metal ions and vitamin D [74, 75].

To test this association, Monastra *et al.* [76] supplemented 18 healthy volunteers with a single dose of MI (6 g) and, one week later, with 6 g of MI + 150 mg of  $\alpha$ -LA in a single dose.

After the combined administration, the authors observed that maximum MI plasma concentration (Cmax) and area under the time course curve of MI plasma concentration (AUC) were significantly higher (+32.4% and +27.5%, respectively) in respect to when MI alone was administrated alone.

Subsequently, the authors investigated the possible mechanism underlying this effect. In particular, they evaluated the transport of MI, alone and combined with  $\alpha$ -LA across the human intestinal Caco-2 cell monolayer [76], currently used as in vitro model of gut mucosa [77, 78].

In the presence of  $\alpha$ -LA the passage of MI across the monolayer was increased, due to the transient opening of the tight junctions between the cells [76] and the subsequent 'passive' transport of MI.

The clinical study by Montanino Oliva *et al.* on 37 anovulatory PCOS women confirmed the beneficial effects of the combination of MI and  $\alpha$ -LA [79].

After an orally supplementation with 2g of MI twice a day for 3 months, 23 women (62%) ovulated, whereas 14

(38%) remained anovulatory, showing hallmarks of inositol resistance. These 14 women were further supplemented with the association of 2 g MI and 50 mg  $\alpha$ -LA, twice a day for an additional 3 months. As a result, 12 (86%) patients ovulated, featuring significantly higher serum levels of MI and better hormone and lipid profiles with respect to the baseline.

This study supports Monastra's findings [76], providing a promising option to overcome some limitations in the clinical approaches using inositol.

Future research is needed in order to confirm these results on larger cohorts of patients as well as on different PCOS phenotypes and to accurately tailor the proper administration and the best combinations of MI, DCI, and  $\alpha$ -LA.

#### **Conflicts of interest**

The authors declare no conflict of interest concerning this article.

#### References

**1.** Parthasarathy R, , Eisenberg Jr F. . The inositol phospholipids: a stereochemical view of biological activity. *Biochem J* 1986; 235 (2):313-22.

**2.** Michell RH. Inositol and its derivatives: their evolution and functions. *Adv Enzyme Regul* 2011; 51(1):84-90.

**3.** Michell RH. Do inositol supplements enhance phosphatidylinositol supply and thus support endoplasmic reticulum function? *Br J Nutr* 2018; 120(3):1-16.

**4.** Beemster P, Groenen P, Steegers-Theunissen R. Involvement of inositol in reproduction. *Nutr Rev* 2002; 60(3):80-7.

5. Clements Jr RS, Diethelm AG. The metabolism of myo-inositol by the human kidney. *J Lab Clin Med* 1979; 93(2):210-9.

**6.** Sun TH, Heimark DB, Nguygen T, *et al*. Both myo-inositol to chiroinositol epimerase activities and chiro-inositol to myo-inositol ratios are decreased in tissues of GK type 2 diabetic rats compared to Wistar controls. *Biochem Biophys Res Commun* 2002; 293(3):1092-8.

7. Heimark D, McAllister J, Larner J. Decreased myo-inositol to chiroinositol (M/C) ratios and increased M/C epimerase activity in PCOS theca cells demonstrate increased insulin sensitivity compared to controls. *Endocr J* 2014; 61(2):111-7.

**8.** Monastra G, Unfer V, Harrath AH, *et al*. Combining treatment with myo-inositol and D-chiro-inositol (40:1) is effective in restoring ovary function and metabolic balance in PCOS patients. *Gynecol Endocrinol* 2017; 33(1):1-9.

**9.** Facchinetti F, Dante G, Neri I. The ratio of MI to DCI and its impact in the treatment of polycystic ovary syndrome: experimental and literature evidences. In : Genazzani AR, Tarlatzis BC, (eds). *Frontiers in gynecological endocrinology: volume 3: ovarian function and reproduction – from needs to possibilities*. Cham: Springer International Publishing. 103-9. **10.** Unfer V, Carlomagno G, Papaleo E, *et al*. Hyperinsulinemia alters myoinositol to d-chiroinositol ratio in the follicular fluid of patients with PCOS. *Reprod Sci* 2014; 21(7):854-8.

**11.** Bizzarri M, Fuso A, Dinicola S, *et al.* Pharmacodynamics and pharmacokinetics of inositol(s) in health and disease. *Expert Opin Drug Metab Toxicol* 2016; 12(10):1181-96.

**12.** Laganà AS, Garzon S, Casarin J, *et al.* Inositol in polycystic ovary syndrome: restoring fertility through a pathophysiology-based approach. *Trends Endocrinol Metab* 2018; 29(11):768-80.

**13.** Nestler JE, Unfer V. Reflections on inositol(s) for PCOS therapy: steps toward success. *Gynecol Endocrinol* 2015; 31(7):501-5.

**14.** Larner J, Huang LC, Tang G, *et al*. Insulin mediators: structure and formation. *Cold Spring Harb Symp Quant Biol* 1988; 53(Pt 2):965-71.

**15.** Huang LC, Fonteles MC, Houston DB, *et al.* Chiroinositol deficiency and insulin resistance. III. Acute glycogenic and hypoglycemic effects of two inositol phosphoglycan insulin mediators in normal and streptozotocin-diabetic rats *in vivo. Endocrinology* 1993; 132(2):652-7.

**16.** Bevilacqua A, Bizzarri M. Inositols in insulin signaling and glucose metabolism. *Int J Endocrinol* 2018; 2018 : 1968450.

**17.** Chukwuma CI, Ibrahim MA, Islam MS. Myo-inositol inhibits intestinal glucose absorption and promotes muscle glucose uptake: a dual approach study. *J Physiol Biochem* 2016; 72(4):791-801.

**18.** Kim JN, Han SN, Kim HK. Phytic acid and myo-inositol support adipocyte differentiation and improve insulin sensitivity in 3T3-L1 cells. *Nutr Res* 2014; 34(8):723-31.

**19.** Ricote M, Li AC, Willson TM, *et al*. The peroxisome proliferatoractivated receptor-gamma is a negative regulator of macrophage activation. *Nature* 1998; 391(6662):79-82.

**20.** Milewska EM, Czyzyk A, Meczekalski B, *et al.* Inositol and human reproduction. From cellular metabolism to clinical use. *Gynecol Endocrinol* 2016; 32(9):690-5.

**21.** Lewin LM, Yannai Y, Melmed S, *et al.* myo-inositol in the reproductive tract of the female rat. *Int J Biochem* 1982; 14(2):147-50.

**22.** Stocco C. Tissue physiology and pathology of aromatase. *Steroids* 2012; 77(1-2):27-35.

**23.** Sacchi S, Marinaro F, Tondelli D, *et al.* Modulation of gonadotrophin induced steroidogenic enzymes in granulosa cells by d-chiroinositol. *Reprod Biol Endocrinol* 2016; 14(1):52.

**24.** Nestler JE, Jakubowicz DJ, de Vargas AF, *et al.* Insulin stimulates testosterone biosynthesis by human thecal cells from women with polycystic ovary syndrome by activating its own receptor and using inositolglycan mediators as the signal transduction system. *J Clin Endocrinol Metab* 1998; 83(6):2001-5.

**25.** Ravanos K, Monastra G, Pavlidou T, *et al.* Can high levels of D-chiro-inositol in follicular fluid exert detrimental effects on blastocyst quality? *Eur Rev Med Pharmacol Sci* 2017; 21(23):5491-8.

**26.** Chiu TT, Tam PP. A correlation of the outcome of clinical *in vitro* fertilization with the inositol content and embryotrophic properties of human serum. *J Assist Reprod Genet* 1992; 9(6):524-30.

**27.** Chiu TT, Rogers MS, Law EL, *et al.* Follicular fluid and serum concentrations of myo-inositol in patients undergoing IVF: relationship with oocyte quality. *Hum Reprod* 2002; 17(6):1591-6.

**28.** Chiu TT, Rogers MS, Briton-Jones C, *et al*. Effects of myo-inositol on the *in-vitro* maturation and subsequent development of mouse oocytes. *Hum Reprod* 2003; 18(2):408-16.

**29.** Goud PT, Goud AP, Van Oostveldt P, *et al*. Presence and dynamic redistribution of type I inositol 1,4,5-trisphosphate receptors in human oocytes and embryos during *in-vitro* maturation, fertilization and early cleavage divisions. *Mol Hum Reprod* 1999; 5(5):441-51.

**30.** Wdowiak A. Myoinositol improves embryo development in PCOS patients undergoing ICSI. *Int J Endocrinol* 2016; 2016 : 6273298.

**31.** Vartanyan EV, Tsaturova KA, Devyatova EA, *et al.* Improvement in quality of oocytes in polycystic ovarian syndrome in programs of in vitro fertilization. *Gynecol Endocrinol* 2017; 33(S1):8-11.

**32.** Emekçi Özay Ö, Özay AC, Çagaliyan E, et al. Myo-inositol administration positively effects ovulation induction and intrauterine insemination in patients with polycystic ovary syndrome: a prospective, controlled, randomized trial. *Gynecol Endocrinol* 2017; 33(7):524-8.

**33.** Laganà AS, Vitagliano A, Noventa M, *et al.* Myo-inositol supplementation reduces the amount of gonadotropins and length of ovarian stimulation in women undergoing IVF: a systematic review and meta-analysis of randomized controlled trials. *Arch Gynecol Obstet* 2018; 298(4):675-84.

**34.** Zheng X, Lin D, Zhang Y, *et al.* Inositol supplement improves clinical pregnancy rate in infertile women undergoing ovulation induction for ICSI or IVF-ET. *Medicine (Baltimore)* 2017; 96(49): e8842.

**35.** Colazingari S, Fiorenza MT, Carlomagno G, *et al.* Improvement of mouse embryo quality by myo-inositol supplementation of IVF media. *J Assist Reprod Genet* 2014; 31(4):463-9.

**36.** Fujiwara T, Nakada K, Shirakawa H, *et al.* Development of inositol trisphosphate-induced calcium release mechanism during maturation of hamster oocytes. *Dev Biol* 1993; 156(1):69-79.

**37.** Mehlmann LM, Kline D. Regulation of intracellular calcium in the mouse egg: calcium release in response to sperm or inositol trisphosphate is enhanced after meiotic maturation. *Biol Reprod* 1994; 51(6):1088-98.

**38.** Kuşcu N, Bizzarri M, Bevilacqua A. Myo-inositol safety in pregnancy: from preimplantation development to newborn animals. *Int J Endocrinol* 2016; 2016 : 2413857.

**39.** Fiorenza MT, Torcia S, Canterini S, *et al.* TCL1 promotes blastomere proliferation through nuclear transfer, but not direct phosphorylation, of AKT/PKB in early mouse embryos. *Cell Death Differ* 2008; 15(2):420-2.

**40.** Luconi M, Torcia S, Grillo D, *et al*. Enhancement of mouse sperm motility by the PI3-kinase inhibitor LY294002 does not result in toxic effects on preimplantation embryo development. *Hum Reprod* 2005; 20(12):3500-4.

**41.** Kane MT, Norris M, Harrison RA. Uptake and incorporation of inositol by preimplantation mouse embryos. *J Reprod Fertil* 1992; 96 (2):617-25.

**42.** Muscogiuri G, Palomba S, Laganà AS, *et al.* Inositols in the treatment of insulin-mediated diseases. *Int J Endocrinol* 2016; 2016 : 3058393.

**43.** Bevilacqua A, Dragotto J, Giuliani A, et al. Myo-inositol and D-chiro-inositol (40:1) reverse histological and functional features of

polycystic ovary syndrome in a mouse model. *J Cell Physiol* 2019; 234 (6):9387-98.

**44.** Caldwell AS, Middleton LJ, Jimenez M, *et al.* Characterization of reproductive, metabolic, and endocrine features of polycystic ovary syndrome in female hyperandrogenic mouse models. *Endocrinology* 2014; 155(8):3146-59.

**45.** Gilling-Smith C, Willis DS, Beard RW, *et al.* Hypersecretion of androstenedione by isolated thecal cells from polycystic ovaries. *J Clin Endocrinol Metab* 1994; 79(4):1158-65.

**46.** Nordio M, Basciani S, Camajani E. The 40:1 myo-inositol/Dchiro-inositol plasma ratio is able to restore ovulation in PCOS patients: comparison with other ratios. *Eur Rev Med Pharmacol Sci* 2019; 23(12):5512-21.

**47.** Carlomagno G, Unfer V, Roseff S. The D-chiro-inositol paradox in the ovary. *Fertil Steril* 2011; 95(8):2515-6.

**48.** Harwood K, Vuguin P, DiMartino-Nardi J. Current approaches to the diagnosis and treatment of polycystic ovarian syndrome in youth. *Horm Res* 2007; 68(5):209-17.

**49.** Matalliotakis I, Kourtis A, Koukoura O, *et al.* Polycystic ovary syndrome: etiology and pathogenesis. *Arch Gynecol Obstet* 2006; 274(4):187-97.

**50.** Rice S, Christoforidis N, Gadd C, *et al.* Impaired insulindependent glucose metabolism in granulosa-lutein cells from anovulatory women with polycystic ovaries. *Hum Reprod* 2005; 20(2):373-81.

**51.** Unfer V, Facchinetti F, Orrù B, *et al.* Myo-inositol effects in women with PCOS: a meta-analysis of randomized controlled trials. *Endocr Connect* 2017; 6(8):647-58.

**52.** Artini PG, Di Berardino OM, Papini F, *et al.* Endocrine and clinical effects of myo-inositol administration in polycystic ovary syndrome. A randomized study. *Gynecol Endocrinol* 2013; 29(4):375-9.

**53.** Costantino D, Minozzi G, Minozzi E, *et al*. Metabolic and hormonal effects of myo-inositol in women with polycystic ovary syndrome: a double-blind trial. *Eur Rev Med Pharmacol Sci* 2009; 13 (2):105-10.

**54.** Genazzani AD, Lanzoni C, Ricchieri F, *et al.* Myo-inositol administration positively affects hyperinsulinemia and hormonal parameters in overweight patients with polycystic ovary syndrome. *Gynecol Endocrinol* 2008; 24(3):139-44.

**55.** Gerli S, Papaleo E, Ferrari A, *et al.* Randomized, double blind placebo-controlled trial: effects of myo-inositol on ovarian function and metabolic factors in women with PCOS. *Eur Rev Med Pharmacol Sci* 2007; 11(5):347-54.

**56.** Pizzo A, Laganà AS, Barbaro L. Comparison between effects of myo-inositol and D-chiro-inositol on ovarian function and metabolic factors in women with PCOS. *Gynecol Endocrinol* 2014; 30(3):205-8.

**57.** Nordio M, Proietti E. The combined therapy with myo-inositol and D-chiro-inositol reduces the risk of metabolic disease in PCOS overweight patients compared to myo-inositol supplementation alone. *Eur Rev Med Pharmacol Sci* 2012; 16(5):575-81.

**58.** Benelli E, Del Ghianda S, Di Cosmo C, *et al*. A combined therapy with myo-inositol and D-chiro-inositol improves endocrine param-

eters and insulin resistance in PCOS young overweight women. *Int J Endocrinol* 2016; 2016 : 3204083.

**59.** Pkhaladze L, Barbakadze L, Kvashilava N. Myo-inositol in the treatment of teenagers affected by PCOS. *Int J Endocrinol* 2016; 2016 : 1473612.

**60.** Zeng L, Yang K. Effectiveness of myoinositol for polycystic ovary syndrome: a systematic review and meta-analysis. *Endocrine* 2018; 59 (1):30-8.

**61.** Fruzzetti F, Perini D, Russo M, *et al.* Comparison of two insulin sensitizers, metformin and myo-inositol, in women with polycystic ovary syndrome (PCOS). *Gynecol Endocrinol* 2017; 33 (1):39-42.

**62.** Donà G, Sabbadin C, Fiore C, *et al.* Inositol administration reduces oxidative stress in erythrocytes of patients with polycystic ovary syndrome. *Eur J Endocrinol* 2012; 166(4):703-10.

**63.** Gerli S, Mignosa M, Di Renzo GC. Effects of inositol on ovarian function and metabolic factors in women with PCOS: a randomized double blind placebo-controlled trial. *Eur Rev Med Pharmacol Sci* 2003; 7(6):151-9.

**64.** Arya BK, Haq AU, Chaudhury K. Oocyte quality reflected by follicular fluid analysis in poly cystic ovary syndrome (PCOS): a hypothesis based on intermediates of energy metabolism. *Med Hypotheses* 2012; 78(4):475-8.

**65.** Gateva A, Unfer V, Kamenov Z. The use of inositol(s) isomers in the management of polycystic ovary syndrome: a comprehensive review. *Gynecol Endocrinol* 2018; 34(7):545-50.

**66.** Dinicola S, Chiu TT, Unfer V, *et al.* The rationale of the myoinositol and D-chiro-inositol combined treatment for polycystic ovary syndrome. *J Clin Pharmacol* 2014; 54(10):1079-92.

**67.** Thessaloniki ESHRE/ASRM-Sponsored PCOS Consensus Workshop Group. . Consensus on infertility treatment related to polycystic ovary syndrome. *Fertil Steril* 2008; 89(3):505-22.

**68.** Kamenov Z, Kolarov G, Gateva A, *et al.* Ovulation induction with myo-inositol alone and in combination with clomiphene citrate in polycystic ovarian syndrome patients with insulin resistance. *Gynecol Endocrinol* 2015; 31(2):131-5.

**69.** Rolland AL, Peigné M, Plouvier P, et al. Could myo-inositol soft gel capsules outperform clomiphene in inducing ovulation? Results of a pilot study. *Eur Rev Med Pharmacol Sci* 2017; 21(2 Suppl):10-4.

**70.** Showell MG, Mackenzie-Proctor R, Jordan V, *et al.* Inositol for subfertile women with polycystic ovary syndrome. *Cochrane Database Syst Rev* 2018; 12(12):Cd012378.

**71.** Schneider S. Inositol transport proteins. *FEBS Lett* 2015; 589 (10):1049-58.

**72.** Lin X, Ma L, Fitzgerald RL, *et al*. Human sodium/inositol cotransporter 2 (SMIT2) transports inositols but not glucose in L6 cells. *Arch Biochem Biophys* 2009; 481(2):197-201.

**73.** Aouameur R, Da Cal S, Bissonnette P, *et al.* SMIT2 mediates all myo-inositol uptake in apical membranes of rat small intestine. *Am J Physiol Gastrointest Liver Physiol* 2007; 293(6):G1300-1307.

**74.** Wang X, Ai T, Meng XL, *et al.* In vitro iron absorption of  $\alpha$ -lactalbumin hydrolysate-iron and  $\beta$ -lactoglobulin hydrolysate-iron complexes. J Dairy Sci 2014; 97(5):2559-66.

**75.** Delavari B, Ali AM-M, Ali Akbar S, *et al*. Alpha-lactalbumin: a new carrier for vitamin D3 food enrichment. *Food Hydrocolloids* 2015; 45 : 124-31.

**76.** Monastra G, Sambuy Y, Ferruzza S, *et al.* Alpha-lactalbumin Effect on Myo-inositol Intestinal Absorption: In vivo and In vitro. *Curr Drug Deliv* 2018; 15(9):1305-11.

77. Sambuy Y, De Angelis I, Ranaldi G, et al. The Caco-2 cell line as a model of the intestinal barrier: influence of cell and culture-related

factors on Caco-2 cell functional characteristics. *Cell Biol Toxicol* 2005; 21(1):1-26.

**78.** Lemmer HJ, Hamman JH. Paracellular drug absorption enhancement through tight junction modulation. *Expert Opin Drug Deliv* 2013; 10(1):103-14.

**79.** Montanino Oliva M, Buonomo G, Calcagno M, *et al*. Effects of myo-inositol plus alpha-lactalbumin in myo-inositol-resistant PCOS women. *J Ovarian Res* 2018; 11(1):38.