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RESEARCH ARTICLE

Spatially balanced sampling methods are always more precise than random ones for estimating the size of aggregated populations

Jan Perret¹  | Anne Charpentier¹  | Roger Pradel¹  | Guillaume Papuga²  | Aurélien Besnard¹ 

¹CEFE, Univ Montpellier, CNRS, EPHE-PSL University, IRD, Montpellier, France

²Univ Montpellier, AMAP, IRD, CIRAD, CNRS, INRAE, Montpellier, France

Correspondence

Jan Perret

Email: jan.perret@gmail.com

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Abstract

1. Population size is a crucial parameter for both ecological research and conservation planning. When individuals are aggregated, estimating the size of a population through sampling raises methodological challenges, as the high variance between sampling units leads to imprecise estimates. Choosing the right sample design depending on the population aggregation level could improve the precision of estimates; however, this is difficult because studies comparing sample designs for aggregated populations have been limited to a few populations and sampling designs, so their results cannot be generalised.
2. To address this gap, we combined simulations of spatial point populations and field counts of three plant species to compare the relative precision of estimates between three sampling methods: simple random sampling (SRS), systematic sampling (SYS) and spatially balanced sampling (SBS). Comparisons were performed on density and aggregation gradients for a range of sample sizes.
3. Our simulations showed that SYS and SBS were always more precise than SRS when individuals were aggregated, reducing sampling variance up to 80% and 60%. The highest precision for estimating population size was always obtained when the average distance between sampling units equalled the diameter of the clusters (i.e. the groups of individuals). The difference in precision was similar for the natural populations, with sampling variance lowered by up to 75% (SYS) and 60% (SBS) compared to SRS.
4. These findings lead us to recommend using SYS or SBS rather than SRS to estimate population size when individuals are spatially aggregated, as these consistently provide more precise estimates. Assessing cluster diameters in the field enables a quick assessment of the potential gain in precision to expect, and thus the best choice of sampling method depending on the trade-off between precision and field constraints.

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KEYWORDS

autocorrelation, balanced acceptance sampling, clustered population, plant population, population monitoring, sampling error, spatial point process, survey design

1 | INTRODUCTION

Population size is a central parameter for all fields related to ecology and evolution. Evolutionary biologists use population size to predict the risk of genetic diversity loss due to inbreeding and genetic drift (Crow, 2010; Frankham, 1995). Ecologists study how population size varies over space and time to identify biotic interactions and abiotic factors that shape population dynamics (Bjørnstad & Grenfell, 2001; Quérroué et al., 2021) and their possible cascading consequences on ecosystems, as in biological invasions (Simberloff et al., 2013) or the reintroduction of keystone species (Ripple & Beschta, 2012; Watson & Estes, 2011). In conservation biology, population size is used to assess extinction risk (Dennis et al., 1991) and species conservation status (e.g. IUCN, 2019) and to evaluate the effectiveness of management actions (Beissinger & McCullough, 2002). Despite its apparent simplicity, estimating the size of a population raises several methodological challenges, as it is often impossible to count every individual. Ecologists thus rely on sampling: counting individuals in a subset of spatial units occupied by the studied population to infer the entire population size (Cochran, 1977).

Sampling is a prolific field of statistics, and various methods exist for selecting sampling units (Thompson, 2012). Whatever the sampling method, estimating the size of a population denoted Y is achieved through the same four steps (Cochran, 1977): (1) Defining a statistical population made up of N units, usually spatial units (e.g. all possible quadrats covering the study area); (2) Drawing from this a sample of n units on which to conduct counts of individuals, denoted y_1, y_2, \dots, y_n ; (3) Estimating the mean number of individuals per unit as the sample mean $\hat{Y} = \bar{y} = \frac{1}{n} \sum_{i=1}^n y_i$; finally (4) Inferring the entire population size by multiplying the sample mean by the number of sampling units in the statistical population $\hat{Y} = N\bar{y}$. The precision of the density estimate \hat{Y} depends on the sample size n , higher sample sizes reducing the sampling variance, and on two population parameters: the population mean \bar{Y} , and the dispersion of the number of individuals per sampling unit y_i around the mean. A higher population mean tends to increase the sampling variance. For a population with a given mean number of individuals per sampling unit, the more heterogeneous these numbers between units, the less precise the estimates of the mean number of individuals per sampling unit (Cochran, 1977). Therefore, the distribution of individuals in the studied population strongly affects the precision of the estimates.

In natural populations, individuals are usually not randomly distributed in space (Legendre, 1993; Levin, 1992). Distribution may indicate a process of repulsion, with individuals more distant from their nearest neighbours than expected in a random distribution. Repulsion can occur, for example, in territorial animals (e.g. Hinde, 1956; Maher & Lott, 2000) or in plant species due to intra-specific (Stoll & Bergius, 2005) or interspecific competition (Rayburn

& Schupp, 2013). When sampling such populations, the number of individuals per sampling unit is relatively homogeneous, so population density estimates have a high precision even with relatively small sample sizes (Cochran, 1977). Alternatively, individuals may aggregate, living closer to their nearest neighbours than expected in a random distribution (hereafter referred to as 'aggregated populations'). Aggregation of individuals is frequent in herbaceous plant species, leading many authors to state that most plant populations are aggregated (Damgaard & Irvine, 2019; Greig-Smith, 1983; Robinson, 1954). This arises due to limited dispersal capacity or a patchy habitat (Lara-Romero et al., 2016; Seabloom et al., 2005). Aggregation can also be observed in animals, such as colonial breeding vertebrates (Danchin & Wagner, 1997) and freshwater mussels (Morales et al., 2006; Smith et al., 2011). When sampling aggregated populations, samples are typically composed of many zeros and a few high counts, resulting in imprecise estimates (McGarvey et al., 2016). This makes improving the precision of population size estimates for aggregated populations by choosing an appropriate sampling design an important challenge (Thompson, 2004; Yoccoz et al., 2001).

Two types of sampling methods can be used for aggregated populations: one-step methods, in which all sampling units are selected prior to measurements in the field, and two-step methods, in which a sample of primary units is selected and counted in the field, and new units are added depending on the outcome of counts on primary units. Adaptive cluster sampling (Thompson, 1990) is a two-step method specifically developed for aggregated populations. Although it usually improves the precision of population size estimates (Turk & Borkowski, 2005), it is difficult to implement in the field (see, however, Philippi, 2005 and Morrison et al., 2008) and can lead to very uncertain estimates for small or very aggregated populations (Shackleton et al., 2020), so we did not cover it in this study. The most commonly used one-step sampling methods (Smith et al., 2017) are simple random sampling (SRS), in which the units are randomly selected, and systematic sampling (SYS), in which the location of the first unit is randomly selected and the others are arranged along a rectangular grid (Cochran, 1977). Spatially balanced sampling (SBS) is a more recent one-step sampling method in which selected units are evenly distributed over the study area but without imposing a strictly equal distance between units as in SYS. This can be done through various processes, such as dividing the study area into multiple spatial strata and selecting one unit from each (Stevens & Olsen, 2004) or using a random-start low-discrepancy sequence to select the location of the sampling units (Robertson et al., 2013). These sampling methods yield varyingly precise estimates depending on the spatial distribution of individuals in the studied population (McGarvey et al., 2016). However, no clear guidelines have yet been proposed on the best sampling method to choose depending on the observed aggregation.

To date, three types of studies have provided information about the relative precision of sampling methods for aggregated populations:

- (i) Some compare the precision of estimates between sampling methods in a purely analytical way: for example, they showed that SYS is more precise than SRS for populations with certain types of autocorrelation in the density of individuals (Cochran, 1946, 1977; Matérn, 1986; Quenouille, 1949). However, since the autocorrelation of density is generally unknown prior to fieldwork, these conclusions are difficult to translate into operational recommendations.
- (ii) Some map all individuals from a given population in the field and simulate multiple sampling designs on the raw data or simulated populations with similar characteristics. This approach is valuable in identifying the sampling methods that provide the most precise estimates for a given population but lacks generality, especially as the conclusions about the relative precision of the sampling methods vary between studies. Indeed, SYS or SBS is often found to be more precise than SRS, but the difference in precision varies considerably between studies (Kermorvant et al., 2020; Mier & Picquelle, 2008; Morrison et al., 2008), and a non-negligible fraction of the studies find all sampling methods to yield roughly the same precision (Khaemba et al., 2001; Smith et al., 2011). However, to our knowledge, no study has found SRS to be more precise than SYS or SBS.
- (iii) Some simulate virtual populations with various spatial distributions on which they then simulate multiple sampling designs. For example, McGarvey et al. (2016) simulated populations with multiple levels of aggregation and explored how the precision of estimates varied between SRS and SYS for a sample size of $n = 100$. They found that SYS was substantially more precise than SRS for all aggregated populations.

The three types of studies seem to indicate that SYS and SBS usually yield more or as precise estimates than SRS. However, it is unclear which method between SYS and SBS yields the most precise estimates, and no previous study has described how the relative precision of the sampling methods varies as a function of the three parameters affecting the precision of estimates: the mean density of individuals, their aggregation level and sample size. Furthermore, it has been shown analytically that SYS provides more precise estimates than SRS if the variance within the systematic samples is higher than the variance of the whole population (Cochran, 1977: 208). However, the mechanism determining the within-sample variance, and thus the precision of SYS relatively to SRS, has never been described. Understanding this mechanism would allow knowing with certainty which is the optimal sampling design for any studied population.

This study aimed to determine which sampling method provides the most precise estimates depending on the level of aggregation of the population. To this end, we combined computer-based simulations and field counts of plant populations to compare the precision

of three one-step sampling methods (SRS, SYS and SBS) over wide gradients of density, aggregation and sample size. We sought to answer four key questions: (i) Does aggregation have the same effect on estimate precision across the three sampling methods? (ii) Does the effect of aggregation on estimate precision change with population density? (iii) Does the effect of aggregation on estimate precision change with sample size? (iv) What is the mechanism generating the differences in precision between sampling methods for aggregated populations?

2 | MATERIALS AND METHODS

Our simulations followed a three-step procedure: (1) we generated virtual populations for a given combination of density and aggregation, (2) we drew samples from each virtual population with three sampling methods (SRS, SYS and SBS) and several sample sizes, and (3), we measured the precision of the density estimates obtained with each sampling method and sample size for each virtual population (Figure 1).

2.1 | Simulation of the virtual populations

We generated virtual populations by simulating spatial point patterns in which each point represented the location of an individual of the population of interest. We simulated the populations inside a study area of 100×100 spatial units (i.e. as the dimensions were virtual, they could be any surface area), and the sampling units were all 10,000 spatial units (hereafter called 'cells') with a dimension of 1×1 covering the study area. The distribution of individuals was generated using point process models (Baddeley et al., 2015). We investigated an aggregation gradient ranging from (i) populations in which individuals repel each other, (ii) to randomly located individuals and (iii) to populations in which individuals aggregate. We thus used three different point process models to generate the populations.

We simulated populations with randomly located individuals using a Poisson point process, in which point locations are determined by randomly drawing their x and y coordinates. We simulated populations with repulsion between individuals using a simple sequential inhibition (SSI) process. In this point process, randomly located points are added one by one to the study area, and if a point falls closer than a chosen distance r from an existing point, it is deleted, and a new random point is generated (Baddeley et al., 2015). Lastly, we simulated populations with individuals aggregating into clusters (i.e. circular groups of individuals) using a Matérn cluster process. This consists of four steps: (1) a set of 'parent' points is generated using a Poisson point process with a mean density denoted $kappa$; (2) a disc of a given radius, denoted $scale$, is centred on each parent point; (3) 'offspring' points are distributed across each disc using a Poisson point process with a mean density noted mu ; and (4) parent points are then suppressed from the simulated distribution. The clusters can overlap, leading

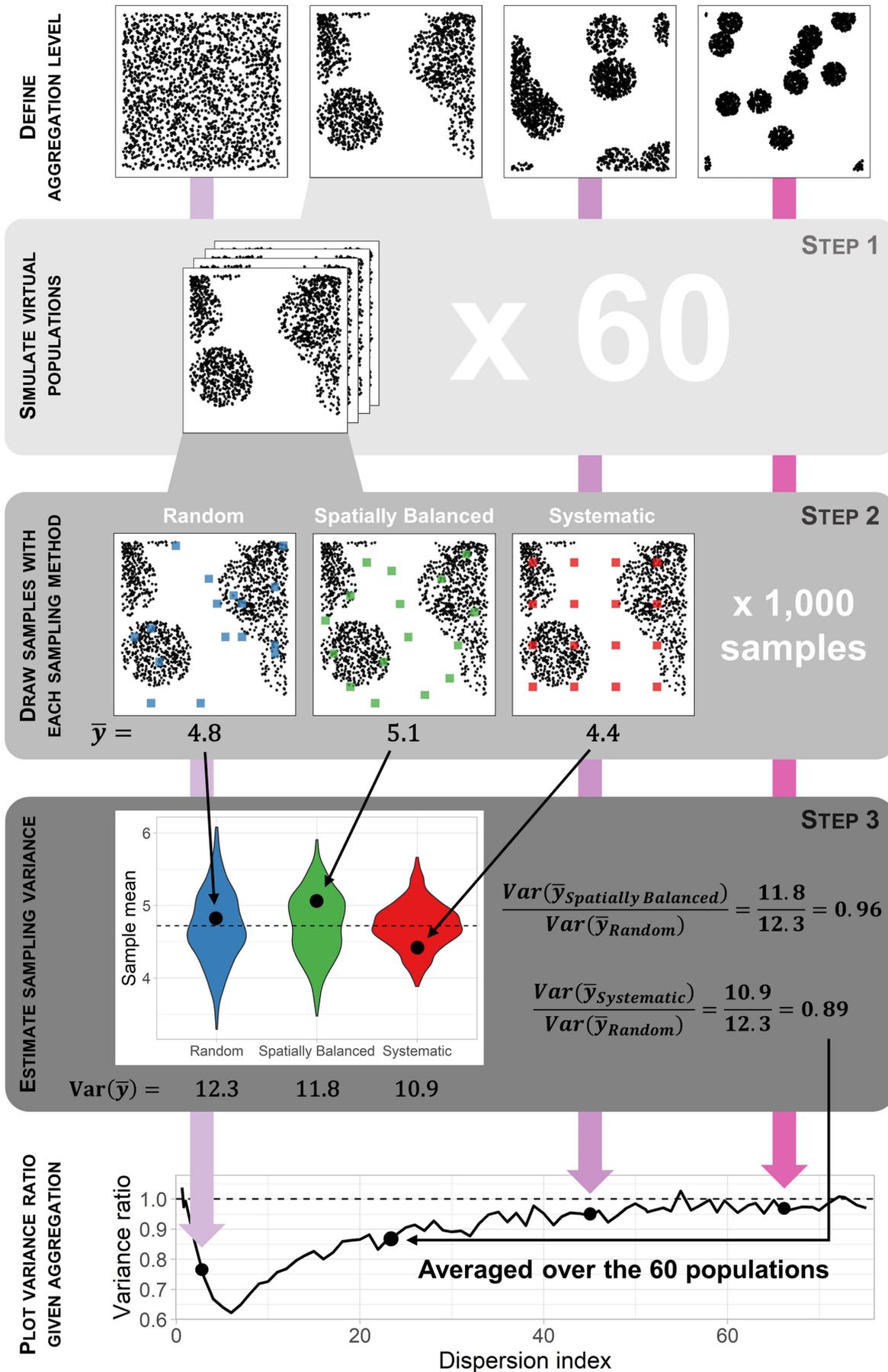


FIGURE 1 Workflow of the simulation process. One level of density (out of seven) and four levels of aggregation (out of 78) are represented. The simulation process is described for one aggregation level and one sample size (out of nine). For every level of aggregation (top row), we simulated 60 virtual populations (step 1). We drew 1000 samples of size n with each of the three sampling methods from every population and computed their means (step 2). We calculated the variance of the 1000 sample means to estimate the sampling variance and computed the ratios of the sampling variance of SYS and spatially balanced sampling (SBS) relative to simple random sampling (SRS; step 3). The horizontal dotted line on the violin plot represents the true mean density of the sampled population. The final result is the curve of the variance ratio as a function of the population aggregation level, each point of the curve being the average over 60 virtual populations. For clarity, only the curve for $\text{var}(\text{systematic})/\text{var}(\text{random})$ is shown.

to areas with a high point density (Matérn, 1986). The mean density of the individuals is $\lambda = \kappa \mu$, and scale only modulates the surface area of the clusters. Therefore, when simulating point patterns with a Matérn cluster process, the density of individuals can be increased by increasing only κ , only μ , or κ and μ simultaneously. These three ways to increase density result in populations with different spatial structures, that is, populations with more clusters, clusters with more individuals or both. As this could affect the precision of density estimates, we simulated all combinations of density and aggregation (as defined below) using the three approaches. For populations in which κ and μ increased simultaneously (modality 1), we kept the ratio of μ over κ at 10,000. For populations in which only κ was increased (modality 2), μ was kept constant at 100 individuals/cluster. For populations in which only μ was increased (modality 3), κ was kept at 0.01 clusters/cell.

To compare the populations simulated with the three types of point processes (Poisson, SSI and Matérn cluster), we used the index of dispersion I as a common metric of aggregation. This index is defined as $\frac{\sigma^2}{\bar{Y}}$, where \bar{Y} is the mean density of individuals in every possible cell of the study area, and σ^2 is the associated variance. I is equal to one for populations with randomly distributed individuals, decreases when individuals show repulsion (its minimum possible value being 0) and increases when individuals aggregate (with no upper bound; Baddeley et al., 2015: 201). For the populations simulated with the Matérn cluster process, we had to increase the cluster radius when increasing population density to maintain the same value of dispersion index in modalities 1 and 3. Thus, populations with the same dispersion index had larger clusters as density increased, except for modality 2, in which clusters had the same radius for the same level of dispersion index at all densities (Appendix S1).

We chose the extent of the density and aggregation gradients based on our knowledge of plant ecology and published literature (see, e.g. Greig-Smith, 1983; Morrison et al., 2008; Reisch et al., 2018). Thus, we simulated populations with seven densities (1, 5, 10, 15, 20, 25 and 30 individuals/cell) and 78 different aggregation levels ($I = 0.4, 0.6$ and 0.8 with the SSI process; $I = 1$ with the Poisson process; $I =$ from 2 to 75 with the Matérn cluster process). We simulated all aggregation levels for all densities, except those generated with the SSI process, which we simulated only for densities of 1, 10 and 20 individuals/cell to save computation time. Altogether, we simulated populations for 534 combinations of density and aggregation, and for each combination we simulated 60 populations. Examples of these populations are found in Appendix S2.

2.2 | Sampling process

We selected nine sample sizes: $n = 9, 15, 25, 49, 100, 150, 196, 300$ and 400 (out of 10,000 possible sample units) because for SYS they allow the sample units to be arranged in a square or quasi-square grid, which limits major spatial coverage differences between sample sizes. Many ways of performing SBS have been proposed, and we chose one of the most commonly used, called balanced acceptance sampling (BAS), in which the sampling units are selected by drawing their coordinates from a low discrepancy sequence, in our case the Halton sequence (Robertson et al., 2013). BAS is one of the methods yielding the best spatial balance when all units are accepted, which was always the case in our simulations (Robertson et al., 2018). For each virtual population, we simulated 1000 sampling surveys for each of the nine sample sizes. A survey consisted of two steps: drawing a sample of size n from a population with each of the three sampling methods and storing the mean densities calculated from the three samples. For SYS and SRS, all sampling units have the same inclusion probability; thus, the sample mean is an unbiased estimator of the population mean density (Cochran, 1977). For SBS as we simulated it, inclusion probabilities are equal to the 3rd or 4th decimal (Robertson et al., 2013), so we treated them as equiprobable, as weighting the sample mean by the inclusion probability of the units would have had no discernible effect on our results. Altogether, we simulated 60,000 sampling surveys by combination of density, aggregation and sample size, that is, 1000 sampling surveys on 60 virtual populations.

2.3 | Comparison of estimate precision between sampling methods

Before comparing the precision of the density estimates obtained with each sampling method, we first checked whether they were unbiased. Then, for each combination of density, aggregation and sample size, we computed the mean of the 1000 mean densities calculated from the samples for each of the 60 virtual populations. We then averaged the resulting 60 mean densities and compared the result to the mean of the true densities of the 60 virtual populations that were sampled. The estimated density was always very close to the true density (see Appendix S3), indicating that the estimates obtained with the three sampling methods were unbiased, as expected from sampling theory (Thompson, 2012).

To measure the precision of the density estimates, we estimated the sampling variance for every combination of sampling method and sample size by computing the variance of the means of the 1000 corresponding sampling surveys for each virtual population. This method of estimating the sampling variance avoids the need for variance estimators, but requires knowing the mean of all possible samples (although in our case we did not draw all possible samples but a large proportion of them) and can therefore only be used in simulation studies (Magnussen et al., 2020; Magnussen & Fehrmann, 2019; McGarvey et al., 2016). This method allows to estimate the conditional sampling variance, that is, conditional on a realised population covering a finite area as encountered in the field, within the context of design-based inference (see Brus, 2021, for a synthesis on design-based and model-based inference). To compare the estimate precision between the three sampling methods for a given sample size, we calculated the ratio of the sampling variance of SYS and SBS, relative to the sampling variance of SRS. This indicator represents the rate of reduction in variance obtained on average when changing from SRS to another sampling method (Kish, 1965). A variance ratio below one means that a method (here SYS or SBS) is more precise than SRS. The variance ratio can also be interpreted as the effective sample size (Kish, 1965). Its inverse then represents the rate of increase in sample size required to achieve the same precision as the alternative method using SRS. For example, for a sample size of $n = 100$, a ratio of 0.5 between the sampling variance of SYS and SRS means that SRS would need, on average, a sample size of $n = 100 \times (1/0.5) = 200$ to achieve the same precision as SYS with $n = 100$. To obtain a generalizable result, for each sample size, we averaged the variance ratio values over the 60 virtual populations that were sampled for each combination of density and aggregation. Hence, the values of variance ratio we present are the expected values for each combination of density, aggregation and sample size (Magnussen et al., 2020).

2.4 | Field study

To compare our results from simulated populations with natural populations, we conducted a field survey to map the distribution of all individuals in one population for three herbaceous plant species exhibiting different levels of density and aggregation. We chose a population of *Bellis sylvestris* Cirillo, a common Mediterranean Asteraceae, to illustrate a population with approximately randomly distributed individuals; a population of *Sanguisorba minor* L., a Rosaceae common in the Mediterranean basin, to illustrate a population with high density and slightly aggregated individuals; and a population of *Limonium girardianum* Guss. to illustrate a population in which individuals are highly aggregated and form well-delimited clusters—this Plumbaginaceae is endemic in the north-west Mediterranean basin and grows in saltmarshes on small sand mounds a few metres wide.

To survey these populations, we placed a 20×20m square on each and mapped all individuals in each 10×10cm cell using 1×1m

quadrats subdivided with a 100-cell grid. To analyse the data from the three natural populations, we grouped the cells four by four to obtain the total number of individuals per 20×20cm cell to have a study area of 100×100 cells as for the virtual populations. We simulated the same sampling surveys on the counts of the three natural populations as on the virtual populations and computed the variance ratio of SYS and SBS relative to SRS. To test whether cluster diameter can be quickly assessed in the field, an observer surveyed the study area by splitting the 20×20m square into 20 columns 1 m in width. He then walked through every second column and measured the largest width of five randomly selected clusters intersected by the column.

All the simulations were performed in R version 3.6.1 (R Core Team, 2019). The spatial point patterns were generated with the package SPATSTAT (Baddeley et al., 2015). The functions used to draw the samples were either programmed by the authors or adapted from the package SDRAW (McDonald & McDonald, 2020).

3 | RESULTS

3.1 | Comparison of estimate precision between the three sampling methods for the virtual populations

The relative precision of the three sampling methods varied considerably with the aggregation level (Figure 2). For randomly distributed populations ($I = 1$) and populations exhibiting repulsion ($I < 1$), the values of the variance ratio were close to one: that is, SRS, SBS and SYS had equivalent precision. This result was consistent for all densities and sample sizes tested. For all populations with aggregated individuals ($I > 1$), whatever the sample size, SYS and SBS were always as precise or more precise than SRS, and SYS was always more precise than SBS. For both SYS and SBS, the variance ratio decreased rapidly below one as aggregation increased, and after reaching a minimum, it slowly increased towards 1. This means that as the aggregation of individuals increased, the estimates obtained with SYS and SBS quickly became more precise than those obtained with SRS until they reached a maximum of relative precision. After this maximum, their relative precision slowly decreased until it became equivalent to SRS for high aggregation levels. When the sample size was increased, the minimum of the variance ratio had a lower value and shifted towards higher levels of aggregation. In other words, as sample size increased, SYS and SBS became increasingly more precise than SRS, and maximum relative precision was reached at higher aggregation levels.

The effect of the mean density depended on the modality we used to simulate increasing density. For modality 1 (shown in Figure 2), when density increased, the variance ratio had lower values, and its minimum shifted towards higher aggregation levels. In other words, as density increased, SYS and SBS became more precise relative to SRS, and their maximum relative precision shifted towards more aggregated populations. For modality 3, the same pattern occurred in a more pronounced way. In contrast, for modality 2, the variance

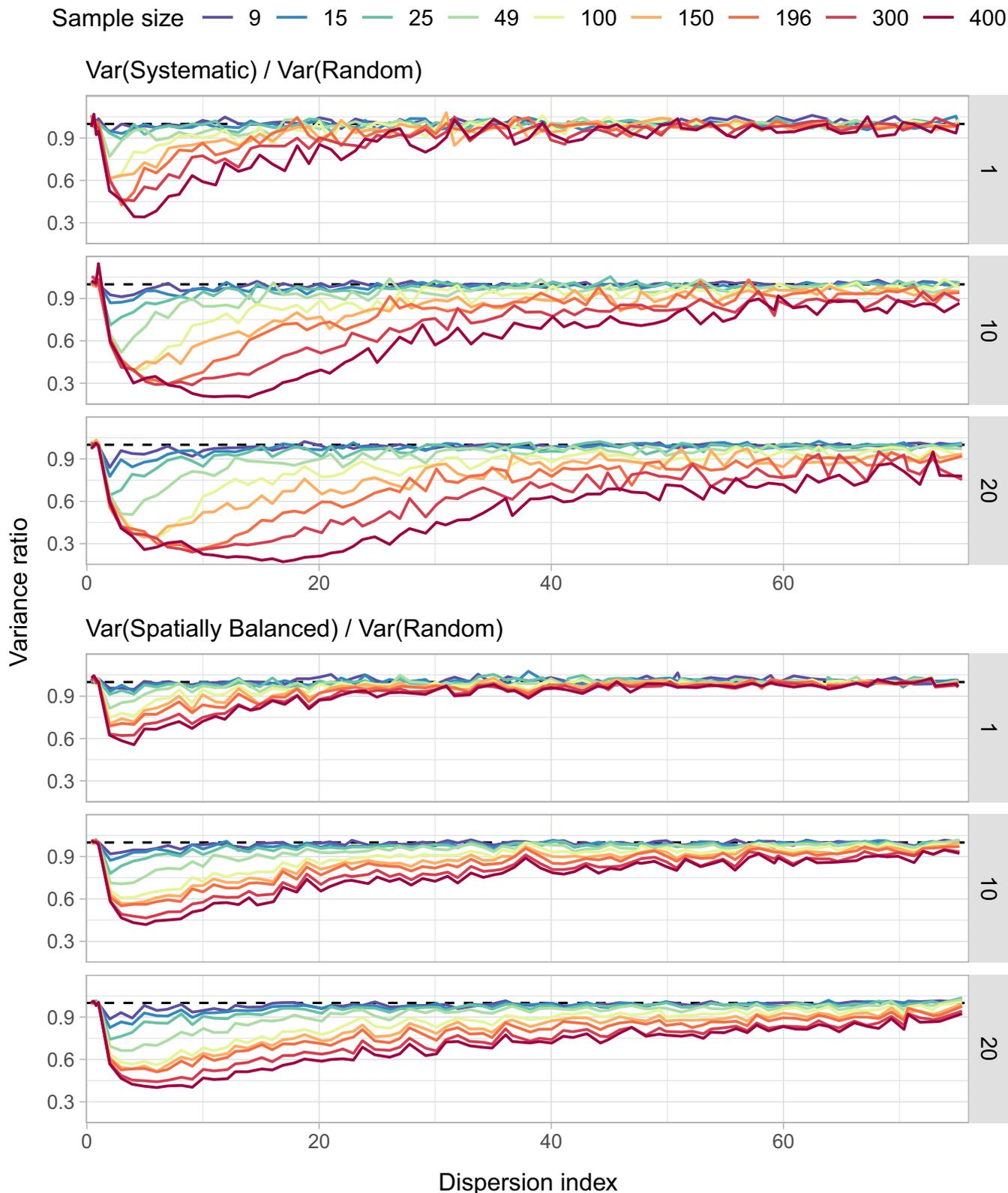


FIGURE 2 Ratio of sampling variance of systematic sampling (SYS) and spatially balanced sampling (SBS) over sampling variance of simple random sampling (SRS), as a function of the dispersion index (I) of the sampled populations. Panels show three mean densities (1, 10 and 20 individuals/cell). For the aggregated populations, density was increased by simultaneously increasing the number of clusters and the number of individuals per cluster (modality 1). Each point of the curves is the ratio of two sampling variances, each estimated from 60,000 sampling survey simulations (1000 for 60 populations).

ratio curves were identical for all densities (Appendix S1). These modalities differed in that the diameter of the clusters changed with density for modalities 1 and 3, but not for modality 2. Therefore, cluster diameter appeared central to explain the relative precision of the sampling methods.

To investigate this result, we represented the variance ratio as a function of the diameter of clusters instead of the dispersion index. Figure 3 shows that the relative precision of SYS compared to SRS for a given sample size was the same for all densities and depended only on cluster diameter. Moreover, the variance ratio only reached one when the cluster diameter was so small that clusters could fit in one sampling unit. We obtained the same result for SBS (Appendix S1) and for all populations, regardless of the modality used to simulate increasing density. Furthermore, the minimum variance ratio of SYS and SBS was always reached when the mean distance between the sampling units was equal to the cluster diameter.

3.2 | Relative precision of estimates for three plant populations

The *B. sylvestris* population had a mean density of 0.079 individual per 20 × 20 cm cell and a dispersion index of 2.87. Individuals

were thus slightly aggregated, although it was difficult to clearly distinguish clusters (Figure 4). The *S. minor* population had a mean density of 1.47 individuals/cell and a dispersion index of 7.63. Individuals formed clusters of various diameters with indistinct boundaries. The population of *L. girardianum* had a mean density of 1.66 individuals/cell and a dispersion index of 8.42. Individuals were grouped into clusters with clear boundaries, corresponding to the sand mounds present in the study area. For *B. sylvestris*, the cluster diameters measured in the field were within the range (20 cm; 110 cm) with a median of 45 cm; for *S. minor*, they were within the range (60 cm; 290 cm) with a median of 145 cm; and for *L. girardianum* they were within the range (20 cm; 520 cm) with a median of 90 cm.

For the three populations, the variance ratio was below 1 for both SYS and SBS for almost all sample sizes, indicating that density estimates obtained with SYS and SBS were generally more precise than those obtained with SRS (Figure 4). The variance ratios decreased overall as the sample size increased. The minimum values reached by the variance ratio (0.25 for SYS and 0.40 for SBS) imply that the sample size would have to be increased by 400% and 250% to achieve the same precision as SYS or SBS using SRS. The SYS and SBS methods did not reach their highest relative precision at the point where the mean distance between the sampling units was equal to the median cluster

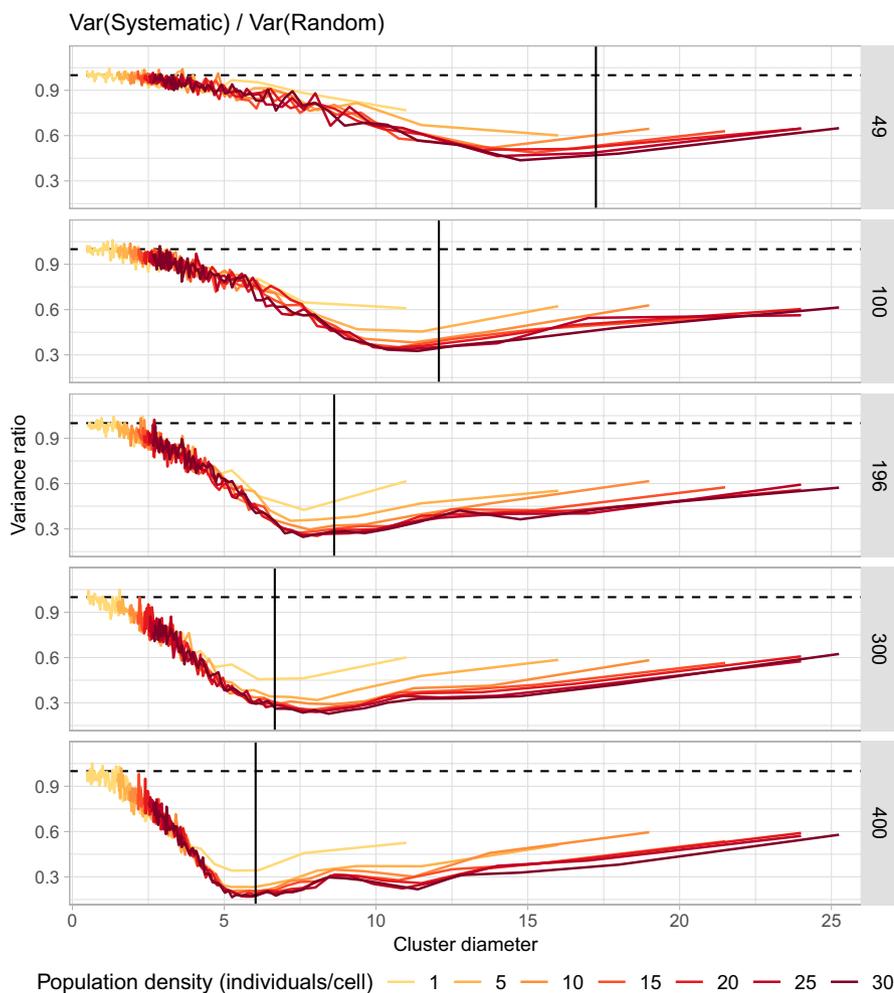


FIGURE 3 Ratio of sampling variance of systematic sampling (SYS) over sampling variance of simple random sampling (SRS) for the aggregated populations. The x-axis is the cluster diameter, and the panels show five different sample sizes. Each curve shows the variance ratio obtained for a given mean density of individuals (from 1 to 30 individuals/cell). The black vertical lines indicate where the distance between the sampling units for SYS is equal to the cluster diameter.

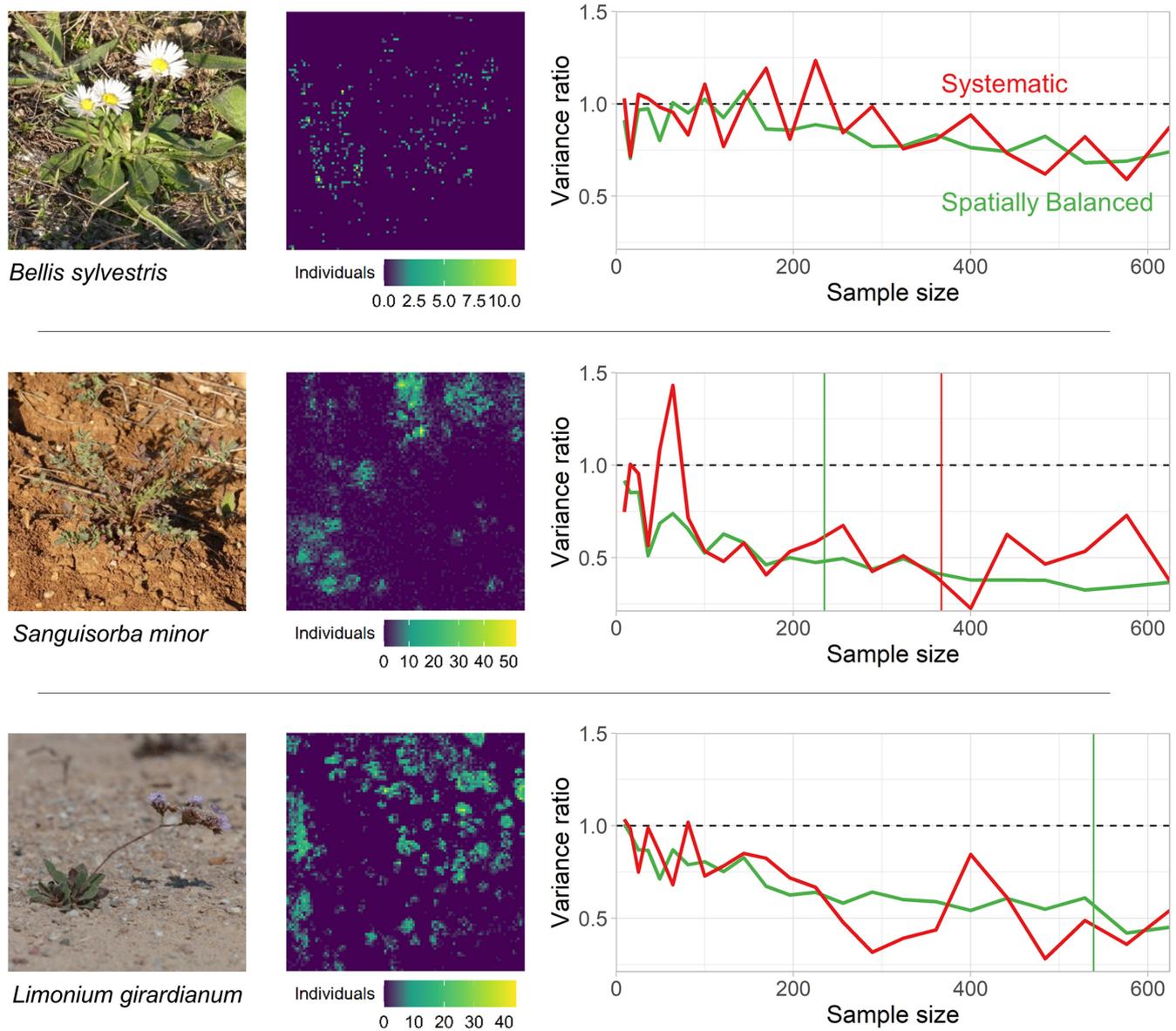


FIGURE 4 Pictures of the three plant species in their natural habitat (left panel), maps of the number of individuals in every 20×20 cm cell covering the study area (middle panel) and curves of the ratios $\text{var}(\text{systematic})/\text{var}(\text{random})$ in red and $\text{var}(\text{spatially balanced})/\text{var}(\text{random})$ in green, as a function of sample size (right panel). The vertical lines represent where the mean distance between the sampling units equals the median cluster diameter we measured in the field. For *B. sylvestris*, this exceeds the extent of the sample size gradient shown here. The maximum simulated sample size for the virtual populations was $n = 400$, but here we simulated sample sizes up to $n = 625$ for illustrative purposes.

diameter, as was the case for the virtual populations. However, at this point, SYS and SBS were considerably more precise than SRS.

4 | DISCUSSION

Our findings provide evidence that different one-step sampling methods do not yield estimates of the same precision when estimating the size of aggregated populations. Our simulations showed that this difference in precision depends on the aggregation level of individuals and sample size. For populations of individuals aggregated in clusters of equal diameter, SYS is on average always as or more

precise than SBS, which is always as or more precise than SRS for a given sample size. For the virtual populations, sampling variance obtained with SYS and SBS was, respectively, up to 80% and 60% lower than that obtained with SRS. The difference in precision was similar for the natural populations, with sampling variance up to 75% lower for SYS and 60% lower for SBS compared to SRS. In other words, the fieldwork effort required to estimate population size could be divided by four or five without altering the precision of the final estimates if the sampling design is well calibrated to the aggregation level of the population.

The overall result that SYS is more precise than SRS for aggregated populations is in line with theoretical work (Matérn, 1986;

Quenouille, 1949) and previous simulation studies (McGarvey et al., 2016). Our results show that this holds true for a wide range of population densities, aggregations and sample sizes. The magnitude of difference in precision we found in our simulations is consistent with several previous studies (Ambrosio et al., 2004; Mier & Picquelle, 2008; Morrison et al., 2008), especially regarding the largest differences in precision that were found. For example, McGarvey et al. (2016) found a reduction in sampling variance of up to 82% for SYS compared to SRS, and Cochran (1977: 223) reported studies finding up to 83% lower variance for SYS compared to stratified random sampling. SBS has been less investigated, but the existing simulation studies have also shown that it is usually more precise than SRS (see Kermorvant et al., 2019, for a summary). Our findings demonstrate that this is true for a large variety of aggregated populations and sample sizes.

4.1 | How the distribution of individuals drives sampling variance in the virtual populations

Simulating sampling surveys over gradients of population density, aggregation and sample size showed that the relative precision of the sampling methods always followed the same pattern as aggregation increased. Changing the population density and sample size modulated this general pattern, shifting the minimum variance ratio to a higher level of aggregation and lowering its value. Nevertheless, the highest relative precision for SYS and SBS was always achieved when the mean distance between the sampling units was equal to the cluster diameter, whatever the density of individuals (Figure 3).

This is because the aggregated populations we simulated were constituted of clusters with the same diameter and number of individuals. In this setup, sampling units located in clusters have values close to each other, and all sampling units located outside clusters are equal to zero. Thus, the sampling variance mainly depends on the between-sample variability in the proportion of sampling units located within clusters. The closer this proportion is between samples, the lower the sampling variance. With SYS and SBS, the spacing between units (fixed for SYS and slightly variable for SBS) leads the proportion of units located in clusters to be less variable between samples than with SRS (Figure 5). Therefore, SYS and SBS will always lead to more precise estimates than SRS, except if the clusters are smaller than the sampling units, in which case all sampling methods will achieve the same precision.

With SYS, when the distance between the sampling units is equal to the cluster diameter (Figure 5, 4th column), all clusters are intersected by a single sampling unit so that the proportion of units located in clusters is strictly identical for all samples. Consequently, the means are very close for all samples, and SYS reaches its optimal precision relative to SRS. When the distance between units is smaller or greater than the cluster diameter, the proportion of units located in clusters is not strictly identical between samples, and the relative precision deviates from the optimum. Using the formulation of Cochran (1977: 208), for any aggregated population, the within-sample variance is the highest for SYS when the distance between the sampling units is equal to the cluster diameter, and thereby SYS achieves its highest precision relative to SRS. The same mechanism operates for SBS, but as the distance between sampling units is not strictly constant, the proportion of sampling units located in clusters

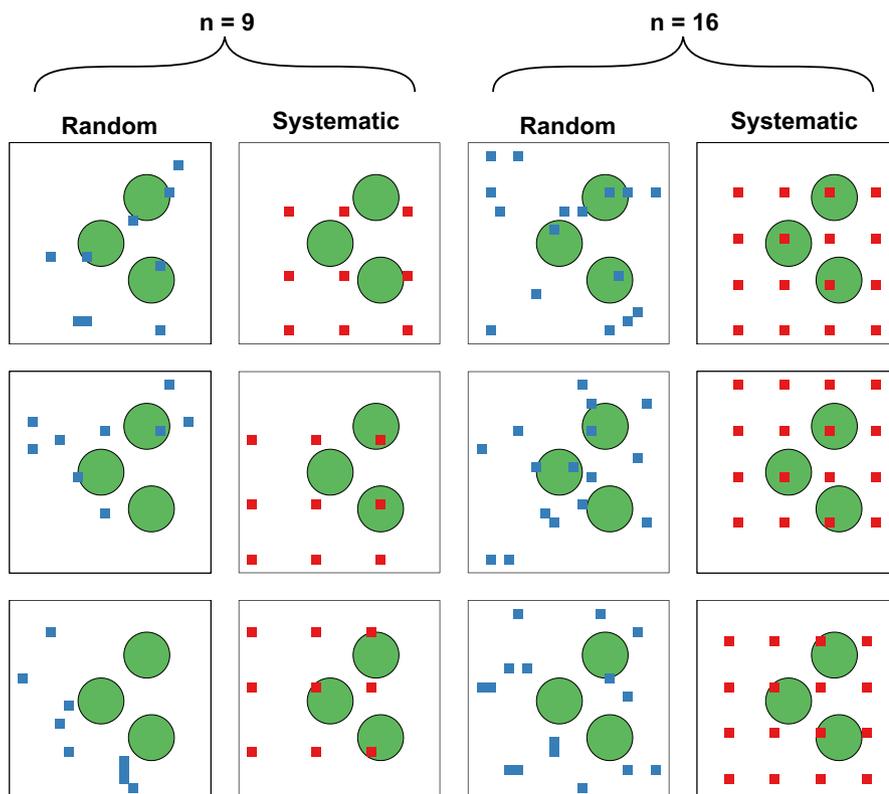


FIGURE 5 Illustration of the mechanism underlying the simulation results regarding the impact of the distance between sampling units on the relative precision of the sampling methods. Three samples drawn from the same virtual population are presented for random (SRS) and systematic sampling (SYS), with two sample sizes ($n = 9$ and $n = 16$). The green circles represent clusters of individuals. For SYS, with $n = 9$ (2nd column), the cluster diameter is inferior to the inter-unit distance, so some clusters can be missed, but there is never more than one unit in each cluster. With $n = 16$ (4th column), the cluster diameter equals the inter-unit distance; all clusters present in the study area are sampled at least once in every sample. This leads to a lower sampling variance for SYS than random sampling, in which the proportion of units located in clusters varies more between samples (1st and 3rd columns).

varies more between samples than in SYS. The SBS method is consequently less efficient than SYS for the virtual populations we simulated. Although it has often been stated that sampling methods with better spatial coverage are usually more precise for spatially structured populations (Kermorvant et al., 2019; Robertson et al., 2013; Stevens & Olsen, 2004), the mechanism driving this expectation has, to our knowledge, never been reported before. This mechanism certainly determines the relative precision of all one-step sampling methods when sampling aggregated populations, not only that of the versions of SYS and SBS that we investigated in this study (systematic square-grid sampling and BAS, respectively).

4.2 | From computer simulations to field studies

The aggregated populations we simulated had a simplistic spatial structure, as clusters were discs of equal diameter with clear boundaries and containing the same number of individuals. The three plant populations we exhaustively mapped illustrate how natural populations are far less simplistic (Figure 4). In the population of *B. sylvestris*, individuals appeared to be slightly aggregated, but no clear cluster could be identified. The two other populations showed clusters of varying size, with indistinct boundaries, and the distribution of individuals within clusters was heterogeneous. Despite these differences between the virtual and natural populations, the general result that SYS and SBS are more precise than SRS on aggregated populations held true for the three natural populations, and the variance ratio reached similar values than for the virtual populations. Thus, sampling methods with a balanced spatial coverage seem to yield more precise estimates than SRS, even for populations with more complex spatial structures than our virtual populations. However, two differences emerged: (1) the variance ratio did not exhibit a localised minimum at the point where the mean distance between sampling units equalled the diameter of the clusters and instead appeared to be stable for sample sizes above $n = 200$; and (2) SYS and SBS seemed to have roughly similar precision. We expected that the variance ratio curves would not have a minimum as clearly localised for natural populations as for virtual populations. Indeed, for populations consisting of clusters with various sizes and densities of individuals, it is likely that multiple sample sizes may yield a similar proportion of units located in the clusters and, therefore, a similar relative precision. The fact that SYS and SBS achieved a similar precision could stem from the same phenomenon, but this requires further investigation.

Further studies need to be conducted to identify the other characteristics of the distribution of individuals than the diameter of the clusters involved for natural populations. This will first require to better understand how individuals are distributed within natural populations. Currently, datasets containing the location of all individuals in a population, or more synthetic aggregation metrics, remain scarce (but see Morrison et al., 2008, and Law et al., 2009), making it difficult to build simulations with realistic distributions of individuals. Once this barrier is removed, simulation studies will

have to be carried out to identify the characteristics other than cluster diameter that drive the relative precision of sampling methods. For example, the non-random location of clusters, heterogeneity in cluster size and shape, or heterogeneity in density between clusters and within each cluster might be good candidates. Finally, metrics that can be measured easily in the field will have to be identified so that the sampling design can be adapted to the spatial structure of the population. This last step will be critical for the results of future methodological work to be implemented in the field.

4.3 | Recommendations for field studies

Given our results on both virtual and natural populations, we recommend using SYS or SBS when studying populations with signs of spatial aggregation. These sampling methods will, on average, provide more precise estimates than SRS unless the clusters are of similar size to the sampling units, in which case all sampling methods will achieve equivalent precision. However, as the results between simulations and natural populations differed, we do not recommend SYS over SBS, as both methods might provide roughly equivalent precision estimates for natural populations.

To check whether it is worthwhile to choose SYS or SBS instead of SRS, cluster diameter measurements can be used to verify if the planned sample size allows a large increase in precision or not. If clusters are identifiable in the study area, one can randomly select clusters and measure their greatest width. If the median diameter is close to the mean inter-unit distance obtained for the planned sample size, the increase in precision will be large. For the plant populations we mapped, the median cluster diameter we measured identified sample sizes for which SYS and SBS were much more precise than SRS. Another solution proposed by Kermorvant et al. (2020) is to use the available information on the population (i.e. results from a pilot study, expert knowledge, etc.) to simulate a virtual population with a distribution of individuals as close as possible to the studied population, and to perform sampling simulations to identify the optimal sampling design. Furthermore, the consequence of the better spatial coverage of SYS and SBS compared to SRS is that the distance to visit all sample units is generally greater. If travel time is a strong constraint and the expected gain in precision is low, SRS might be favoured over the other methods.

Field ecologists should be aware that SYS has a statistical drawback. For SYS, the sample mean is an unbiased estimator of the population mean, but there is no universally unbiased estimator for the variance of this estimate, that is, the sampling variance (Cochran, 1977: 224). In other words, all existing estimators can sometimes give biased estimates of the sampling variance so that although the population mean will be estimated without bias, the precision of this estimate can be under- or overestimated (Magnussen & Fehrmann, 2019). Nevertheless, we argue that this problem should not prevent using SYS, given the substantial potential increase in precision using this sampling method. The search for better estimators is an ongoing research topic, and the best estimators currently

known usually allow to see a substantial gain in precision by shifting from SRS to SYS, although variances tend to be slightly overestimated. For example, the two best estimators found by McGarvey et al. (2016) overestimated the sampling variance by less than 20% for most populations, although cases of overestimation above 60% were found for a few populations. Simulating even more populations, Magnussen et al. (2020) found two other estimators to overestimate sampling variance on average by less than 10%. We tested these four estimators on our virtual and natural populations (Appendix S4). For the virtual populations, the four estimators overestimated the sampling variance, which was expected given that they are all based on measurements of the correlation among neighbouring units. This is not adapted for populations following a Matérn cluster process as in our simulations. However, we propose a new estimator that is more adapted for this type of populations, and it provided nearly unbiased estimates of the sampling variance. For the natural populations, we found similar levels of performance to previous studies (Magnussen et al., 2020; McGarvey et al., 2016), with several estimators based on correlation between neighbouring units, as well as our new estimator, providing almost unbiased estimates of the sampling variance. Estimating sampling variance is less problematic for SBS, although some cases of biased estimates have also been reported (Robertson et al., 2013; Stevens & Olsen, 2003). We tested the most commonly used estimator, and it provided nearly unbiased estimates for both the virtual and natural populations (Appendix S4). These results confirm that given a variance estimator appropriate for the studied population is used, a substantial gain in precision will be seen when using SYS or SBS instead of SRS. To help choose the variance estimator, future simulation studies need to be conducted to screen the performance of estimators across many distributions of individuals. This will be particularly useful if realistic distributions of individuals can be simulated based on a better understanding of how individuals are distributed in natural populations. Other advantages of SBS over the other two methods include the ability to incorporate legacy sites where data have already been accumulated (Foster et al., 2017) and to draw oversamples to replace units that could not be observed in the field (e.g. inaccessible sites) while maintaining spatially balanced samples (Kermorvant et al., 2019).

The results of this paper refer to design-based inference, in which the estimates of the population mean and sampling variance are generally unbiased (except for SYS as discussed above), and no assumptions about the studied population need to be made. Model-based inference is an alternative approach in which the estimates are obtained by fitting a spatial variation model to the sample. This approach usually provides more precise estimates than design-based inference when there is spatial dependence in the density of individuals. Yet, the accuracy of this approach strongly relies on the realism of model assumptions, and unrealistic assumptions can lead to spurious estimates. Moreover, hybrid methods, called model-assisted inference, which combine the advantages of both approaches have been developed (Brus, 2021). Using model-based or preferably model-assisted inference seems a promising way to further increase the precision of estimates for aggregated populations. However, the

difference in estimate precision between design-based and model-based approaches seems limited when the sample is selected with a SBS method, while it is large with SRS (Dumelle et al., 2022). Finding ways to combine the strengths of model-assisted inference and SBS methods tailored to the aggregation of the studied population would be the next step to further improve the precision of aggregated population size estimates.

AUTHOR CONTRIBUTIONS

Jan Perret, Anne Charpentier, Guillaume Papuga and Aurélien Besnard conceived the study and designed the methodology; Jan Perret and Roger Pradel analysed the data; Jan Perret led the writing of the manuscript. All authors contributed critically to the drafts and gave final approval for publication.

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CONFLICT OF INTEREST

All authors declare no conflicts of interest.

PEER REVIEW

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DATA AVAILABILITY STATEMENT

All data and R codes used for this study are available from the GitHub repository: https://github.com/JanPerret/Sampling_aggregated_populations. The repository is also archived on Zenodo (Perret, 2022).

ORCID

Jan Perret  <https://orcid.org/0000-0002-1912-6995>

Anne Charpentier  <https://orcid.org/0000-0002-8588-6001>

Roger Pradel  <https://orcid.org/0000-0002-2684-9251>

Guillaume Papuga  <https://orcid.org/0000-0002-7803-2219>

Aurélien Besnard  <https://orcid.org/0000-0002-2289-9761>

REFERENCES

- Ambrosio, L., Iglesias, L., Marín, C., & Del Monte, J. P. (2004). Evaluation of sampling methods and assessment of the sample size to estimate the weed seedbank in soil, taking into account spatial variability. *Weed Research*, 44(3), 224–236. <https://doi.org/10.1111/j.1365-3180.2004.00394.x>
- Baddeley, A., Rubak, E., & Turner, R. (2015). *Spatial point patterns: Methodology and applications with R*. Chapman and Hall/CRC Press. <https://doi.org/10.1201/b19708>

- Beissinger, S. R., & McCullough, D. R. (2002). *Population Viability Analysis*. University of Chicago Press.
- Bjørnstad, O. N., & Grenfell, B. T. (2001). Noisy clockwork: Time series analysis of population fluctuations in animals. *Science*, 293(5530), 638–643. <https://doi.org/10.1126/science.1062226>
- Brus, D. J. (2021). Statistical approaches for spatial sample survey: Persistent misconceptions and new developments. *European Journal of Soil Science*, 72(2), 686–703. <https://doi.org/10.1111/ejss.12988>
- Cochran, W. G. (1946). Relative accuracy of systematic and stratified random samples for a certain class of populations. *The Annals of Mathematical Statistics*, 17(2), 164–177. <https://doi.org/10.1214/aoms/1177730978>
- Cochran, W. G. (1977). *Sampling techniques* (3rd ed.). John Wiley & Sons.
- Crow, J. F. (2010). Wright and Fisher on inbreeding and random drift. *Genetics*, 184(3), 609–611. <https://doi.org/10.1534/genetics.109.110023>
- Damgaard, C. F., & Irvine, K. M. (2019). Using the beta distribution to analyse plant cover data. *Journal of Ecology*, 107, 2747–2759. <https://doi.org/10.1111/1365-2745.13200>
- Danchin, E., & Wagner, R. H. (1997). The evolution of coloniality: The emergence of new perspectives. *Trends in Ecology & Evolution*, 12(9), 342–347. [https://doi.org/10.1016/S0169-5347\(97\)01124-5](https://doi.org/10.1016/S0169-5347(97)01124-5)
- Dennis, B., Munholland, P. L., & Scott, J. M. (1991). Estimation of growth and extinction parameters for endangered species. *Ecological Monographs*, 61(2), 115–143. <https://doi.org/10.2307/1943004>
- Dumelle, M., Higham, M., Ver Hoef, J. M., Olsen, A. R., & Madsen, L. (2022). A comparison of design-based and model-based approaches for finite population spatial sampling and inference. *Methods in Ecology and Evolution*, 13(9), 2018–2029. <https://doi.org/10.1111/2041-210X.13919>
- Foster, S. D., Hosack, G. R., Lawrence, E., Przeslawski, R., Hedge, P., Caley, M. J., Barrett, N. S., Williams, A., Li, J., Lynch, T., Dambacher, J. M., Sweatman, H. P. A., & Hayes, K. R. (2017). Spatially balanced designs that incorporate legacy sites. *Methods in Ecology and Evolution*, 8(11), 1433–1442. <https://doi.org/10.1111/2041-210X.12782>
- Frankham, R. (1995). Effective population size/adult population size ratios in wildlife: A review. *Genetics Research*, 66(2), 95–107. <https://doi.org/10.1017/S0016672300034455>
- Greig-Smith, P. (1983). *Quantitative plant ecology*. University of California Press.
- Hinde, A. (1956). The biological significance of the territories of birds. *Ibis*, 98(3), 340–369. <https://doi.org/10.1111/j.1474-919X.1956.tb01419.x>
- IUCN Standards and Petitions Committee. (2019). *Guidelines for using the IUCN red list categories and criteria—version 14*.
- Kermorvant, C., Coube, S., D'amico, F., Bru, N., & Caill-Milly, N. (2020). Sequential process to choose efficient sampling design based on partial prior information data and simulations. *Spatial Statistics*, 38, 100439. <https://doi.org/10.1016/j.spasta.2020.100439>
- Kermorvant, C., D'Amico, F., Bru, N., Caill-Milly, N., & Robertson, B. (2019). Spatially balanced sampling designs for environmental surveys. *Environmental Monitoring and Assessment*, 191(8), 524. <https://doi.org/10.1007/s10661-019-7666-y>
- Khaemba, W. M., Stein, A., Rasch, D., De Leeuw, J., & Georgiadis, N. (2001). Empirically simulated study to compare and validate sampling methods used in aerial surveys of wildlife populations. *African Journal of Ecology*, 39(4), 374–382. <https://doi.org/10.1046/j.0141-6707.2001.00329.x>
- Kish, L. (1965). *Survey sampling*. John Wiley & Sons.
- Lara-Romero, C., de la Cruz, M., Escribano-Ávila, G., García-Fernández, A., & Iriondo, J. M. (2016). What causes conspecific plant aggregation? Disentangling the role of dispersal, habitat heterogeneity and plant–plant interactions. *Oikos*, 125(9), 1304–1313. <https://doi.org/10.1111/oik.03099>
- Law, R., Illian, J., Burslem, D. F. R. P., Gratzner, G., Gunatilleke, C. V. S., & Gunatilleke, I. a. U. N. (2009). Ecological information from spatial patterns of plants: Insights from point process theory. *Journal of Ecology*, 97(4), 616–628. <https://doi.org/10.1111/j.1365-2745.2009.01510.x>
- Legendre, P. (1993). Spatial autocorrelation: Trouble or new paradigm? *Ecology*, 74(6), 1659–1673. <https://doi.org/10.2307/1939924>
- Levin, S. (1992). The problem of pattern and scale in ecology. *Ecology*, 73(6), 1943–1967. <https://doi.org/10.2307/1941447>
- Magnussen, S., & Fehrmann, L. (2019). In search of a variance estimator for systematic sampling. *Scandinavian Journal of Forest Research*, 34(4), 300–312. <https://doi.org/10.1080/02827581.2019.1599063>
- Magnussen, S., McRoberts, R. E., Breidenbach, J., Nord-Larsen, T., Ståhl, G., Fehrmann, L., & Schnell, S. (2020). Comparison of estimators of variance for forest inventories with systematic sampling—Results from artificial populations. *Forest Ecosystems*, 7(1), 17. <https://doi.org/10.1186/s40663-020-00223-6>
- Maher, C. R., & Lott, D. F. (2000). A review of ecological determinants of territoriality within vertebrate species. *The American Midland Naturalist*, 143(1), 1–29. [https://doi.org/10.1674/0003-0031\(2000\)143\[0001:AROEDO\]2.0.CO;2](https://doi.org/10.1674/0003-0031(2000)143[0001:AROEDO]2.0.CO;2)
- Matérn, B. (1986). *Spatial variation* (2nd ed.). Springer. <https://doi.org/10.1007/978-1-4615-7892-5>
- McDonald, T., & McDonald, A. (2020). *SDraw: Spatially balanced samples of spatial objects* (2.1.13). [computer software]. <https://CRAN.R-project.org/package=SDraw>
- McGarvey, R., Burch, P., & Matthews, J. M. (2016). Precision of systematic and random sampling in clustered populations: Habitat patches and aggregating organisms. *Ecological Applications*, 26(1), 233–248. <https://doi.org/10.1890/14-1973>
- Mier, K. L., & Picquelle, S. J. (2008). Estimating abundance of spatially aggregated populations: Comparing adaptive sampling with survey designs. *Canadian Journal of Fisheries and Aquatic Sciences*, 65(2), 176–197. <https://doi.org/10.1139/F07-138>
- Morales, Y., Weber, L. J., Mynett, A. E., & Newton, T. J. (2006). Effects of substrate and hydrodynamic conditions on the formation of mussel beds in a large river. *Journal of the North American Benthological Society*, 25(3), 664–676. [https://doi.org/10.1899/0887-3593\(2006\)25\[664:EOSAHC\]2.0.CO;2](https://doi.org/10.1899/0887-3593(2006)25[664:EOSAHC]2.0.CO;2)
- Morrison, L. W., Smith, D. R., Young, C., & Nichols, D. W. (2008). Evaluating sampling designs by computer simulation: A case study with the Missouri bladderpod. *Population Ecology*, 50(4), 9. <https://doi.org/10.1007/s10144-008-0100-x>
- Perret, J. (2022). JanPerret/Sampling_aggregated_populations. *Zenodo*. <https://doi.org/10.5281/zenodo.6954499>
- Philippi, T. (2005). Adaptive cluster sampling for estimation of abundances within local populations of low-abundance plants. *Ecology*, 86(5), 1091–1100. <https://doi.org/10.1890/04-0621>
- Quenouille, M. H. (1949). Problems in plane sampling. *The Annals of Mathematical Statistics*, 20(3), 355–375. <https://doi.org/10.1214/AOMS/1177729989>
- Quéroué, M., Barbraud, C., Barraquand, F., Turek, D., Delord, K., Pacoureaux, N., & Gimenez, O. (2021). Multispecies integrated population model reveals bottom-up dynamics in a seabird predator–prey system. *Ecological Monographs*, 91(3), e01459. <https://doi.org/10.1002/ecm.1459>
- R Core Team. (2019). *R: A language and environment for statistical computing*. R Foundation for Statistical Computing. <https://www.R-project.org/>
- Rayburn, A. P., & Schupp, E. W. (2013). Effects of community- and neighborhood-scale spatial patterns on semi-arid perennial grassland community dynamics. *Oecologia*, 172(4), 1137–1145. <https://doi.org/10.1007/s00442-012-2567-6>
- Reisch, C., Schmid, C., & Hartig, F. (2018). A comparison of methods for estimating plant population size. *Biodiversity and Conservation*, 27(8), 2021–2028. <https://doi.org/10.1007/s10531-018-1522-1>
- Ripple, W. J., & Beschta, R. L. (2012). Trophic cascades in Yellowstone: The first 15 years after wolf reintroduction. *Biological Conservation*, 145(1), 205–213. <https://doi.org/10.1016/j.biocon.2011.11.005>

- Robertson, B., McDonald, T., Price, C., & Brown, J. (2018). Halton iterative partitioning: Spatially balanced sampling via partitioning. *Environmental and Ecological Statistics*, 25(3), 305–323. <https://doi.org/10.1007/s10651-018-0406-6>
- Robertson, B. L., Brown, J. A., McDonald, T., & Jaksons, P. (2013). BAS: Balanced acceptance sampling of natural resources. *Biometrics*, 69(3), 776–784. <https://doi.org/10.1111/biom.12059>
- Robinson, P. (1954). The distribution of plant populations. *Annals of Botany*, 18(69), 35–45. <https://doi.org/10.1093/oxfordjournals.aob.a083380>
- Seabloom, E. W., Bjornstad, O. N., Bolker, B. M., & Reichman, O. J. (2005). Spatial signature of environmental heterogeneity, dispersal, and competition in successional grasslands. *Ecological Monographs*, 75(2), 199–214. <https://doi.org/10.1890/03-0841>
- Shackleton, R. T., Petitpierre, B., Pajkovic, M., Dessimoz, F., Brönnimann, O., Cattin, L., Čejková, Š., Kull, C. A., Pergl, J., Pyšek, P., Yoccoz, N., & Guisan, A. (2020). Integrated methods for monitoring the invasive potential and Management of *Heracleum mantegazzianum* (giant hogweed) in Switzerland. *Environmental Management*, 65(6), 829–842. <https://doi.org/10.1007/s00267-020-01282-9>
- Simberloff, D., Martin, J.-L., Genovesi, P., Maris, V., Wardle, D. A., Aronson, J., Courchamp, F., Galil, B., Garcia-Berthou, E., Pascal, M., Pyšek, P., Sousa, R., Tabacchi, E., & Vila, M. (2013). Impacts of biological invasions: What's what and the way forward. *Trends in Ecology & Evolution*, 28(1), 58–66. <https://doi.org/10.1016/j.tree.2012.07.013>
- Smith, A. N. H., Anderson, M. J., & Pawley, M. D. M. (2017). Could ecologists be more random? Straightforward alternatives to haphazard spatial sampling. *Ecography*, 40(11), 1251–1255. <https://doi.org/10.1111/ecog.02821>
- Smith, D. R., Rogala, J. T., Gray, B. R., Zigler, S. J., & Newton, T. J. (2011). Evaluation of single and two-stage adaptive sampling designs for estimation of density and abundance of freshwater mussels in a large river. *River Research and Applications*, 27(1), 122–133. <https://doi.org/10.1002/rra.1334>
- Stevens, D. L., & Olsen, A. R. (2003). Variance estimation for spatially balanced samples of environmental resources. *Environmetrics*, 14(6), 593–610. <https://doi.org/10.1002/env.606>
- Stevens, D. L., & Olsen, A. R. (2004). Spatially balanced sampling of natural resources. *Journal of the American Statistical Association*, 99(465), 262–278. <https://doi.org/10.1198/016214504000000250>
- Stoll, P., & Bergius, E. (2005). Pattern and process: Competition causes regular spacing of individuals within plant populations. *Journal of Ecology*, 93(2), 395–403. <https://doi.org/10.1111/j.0022-0477.2005.00989.x>
- Thompson, S. K. (1990). Adaptive cluster sampling. *Journal of the American Statistical Association*, 85(412), 1050–1059. <https://doi.org/10.2307/2289601>
- Thompson, S. K. (2012). *Sampling* (3rd ed.). Wiley.
- Thompson, W. (2004). *Sampling rare or elusive species: Concepts, designs, and techniques for estimating population parameters* (2nd ed.). Island Press.
- Turk, P., & Borkowski, J. J. (2005). A review of adaptive cluster sampling: 1990–2003. *Environmental and Ecological Statistics*, 12(1), 55–94. <https://doi.org/10.1007/s10651-005-6818-0>
- Watson, J., & Estes, J. A. (2011). Stability, resilience, and phase shifts in rocky subtidal communities along the west coast of Vancouver Island, Canada. *Ecological Monographs*, 81(2), 215–239. <https://doi.org/10.1890/10-0262.1>
- Yoccoz, N. G., Nichols, J. D., & Boulinier, T. (2001). Monitoring of biological diversity in space and time. *Trends in Ecology & Evolution*, 16(8), 446–453. [https://doi.org/10.1016/S0169-5347\(01\)02205-4](https://doi.org/10.1016/S0169-5347(01)02205-4)

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