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Digestion des aliments chez l'homme: Comment la microscopie aide à comprendre l'évolution des structures dans le tube digestif

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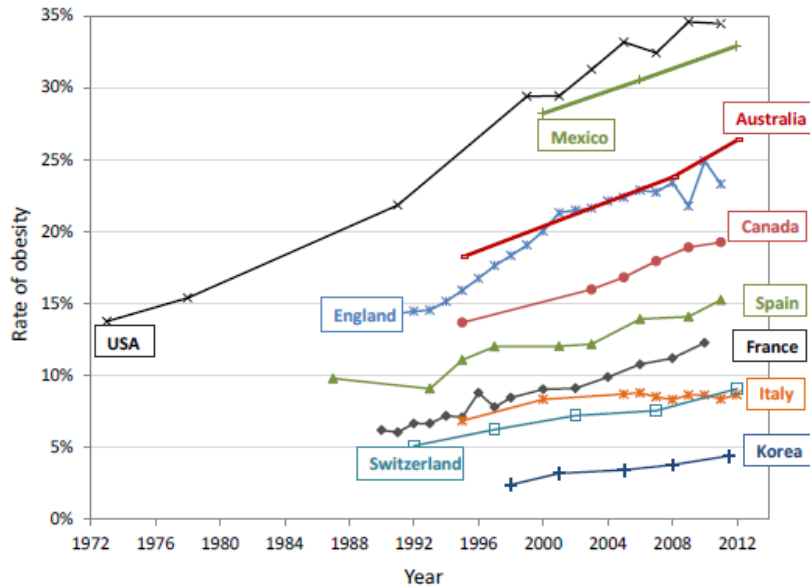


➤ Digestion des aliments chez l'homme:
Comment la microscopie aide à
comprendre l'évolution des structures
dans le tube digestif

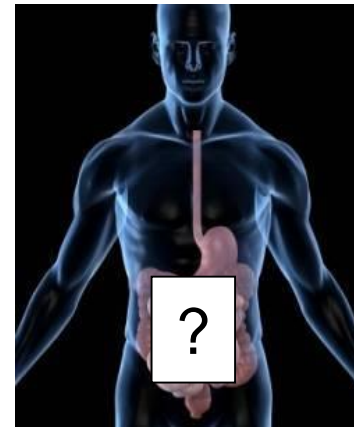
Steven LE FEUNTEUN et Didier DUPONT

INRAE – Institut Agro, STLO, Rennes, France

Food and human health: the key role of digestion



Diet-related diseases ↑
Prevent these pathologies rather than
cure them



Gut = interface between food and human body

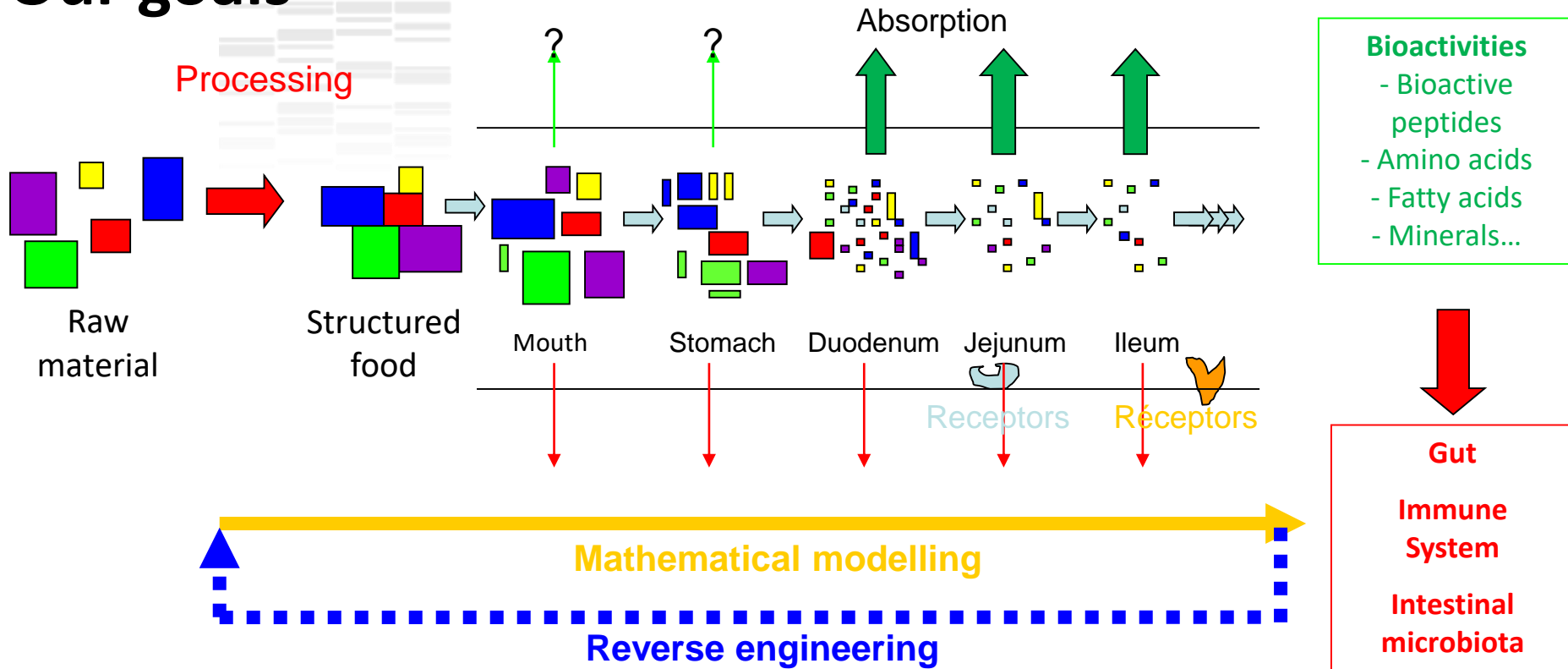
Digestion releases food components that can have a beneficial or a deleterious effect on human health

... but the mechanisms of food disintegration in the gastrointestinal tract remain unclear and the digestive process has been considered as a black box so far

By increasing our knowledge on food digestion, we will increase our knowledge on the effect of food on human health

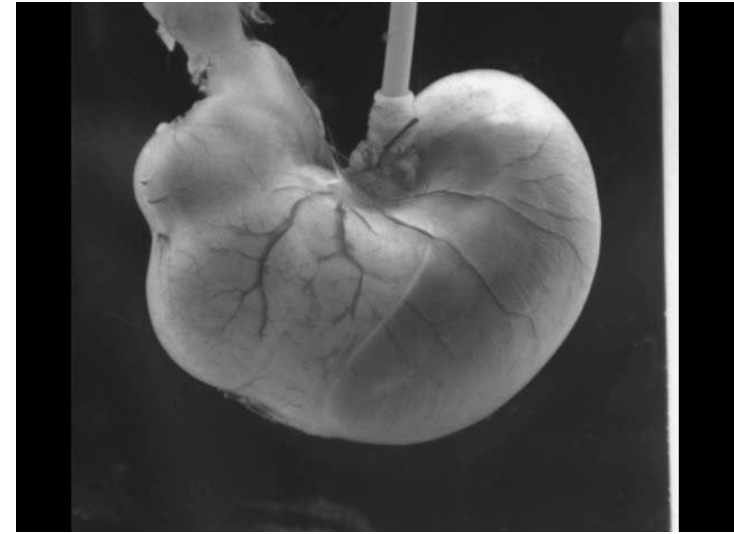
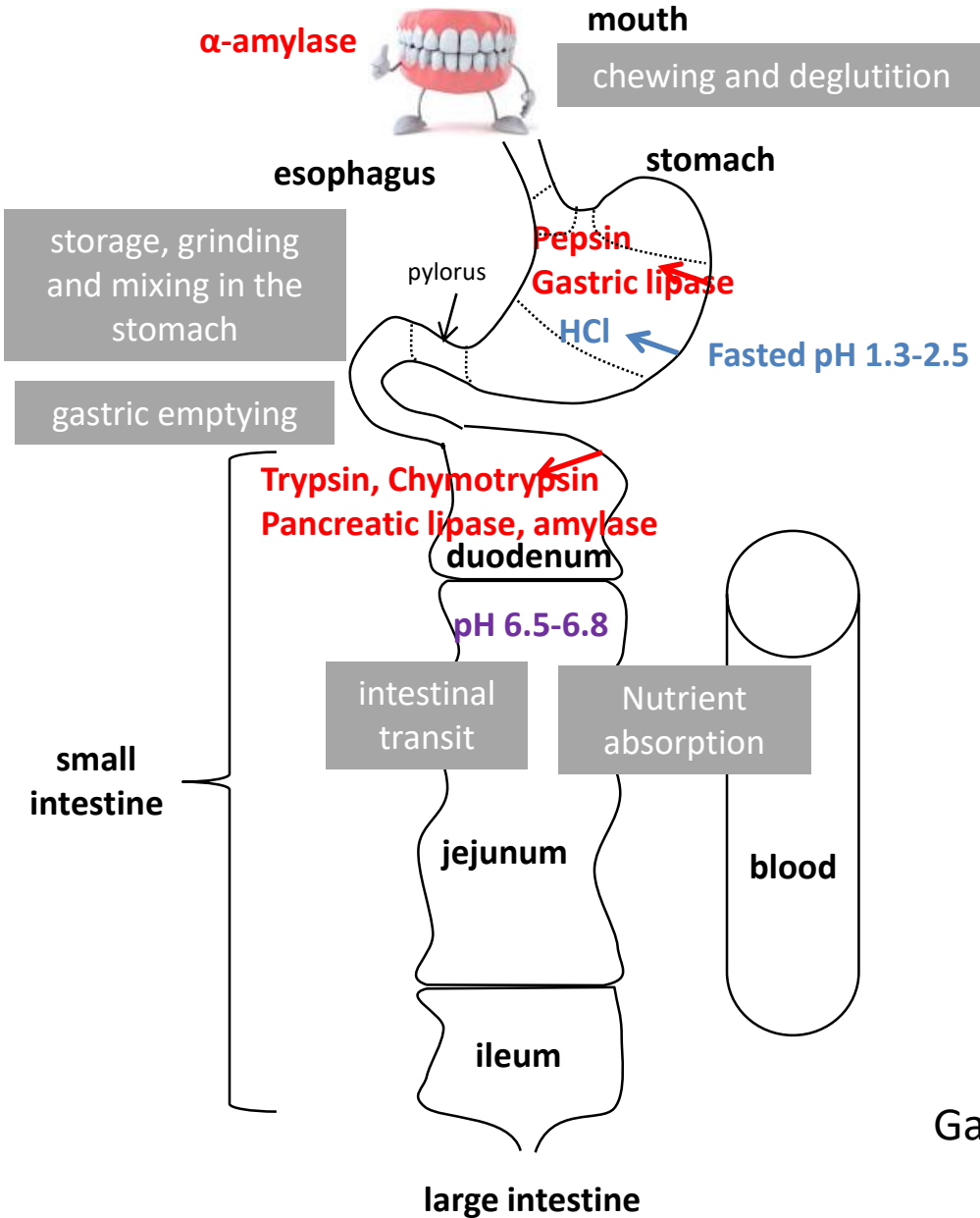
Our goals

Healthy Adult/ Infant/ Elderly

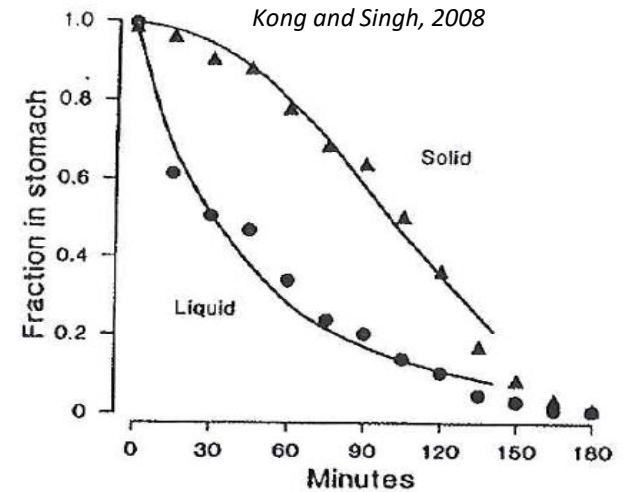


- ☞ To understand the mechanisms of breakdown of food matrices and their constituents in the gut and identify the beneficial/deleterious food components released during digestion
- ☞ To determine the impact of the structure of food matrices on nutrient bioavailability
- ☞ To model these phenomena in order to develop a reverse engineering approach

The digestive process



From Roger Lentle, Massey Univ. NZ



Gastric phase = a very complex but crucial step for the whole digestion process

Models available at INRAE for simulating digestion

Dupont et al.
2010ab,
Mol Nutr Food Res

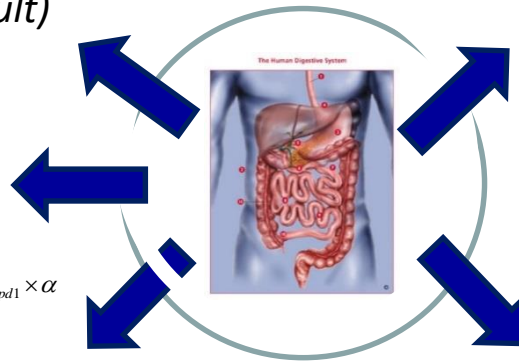


*In vitro static models
(infant, adult)*

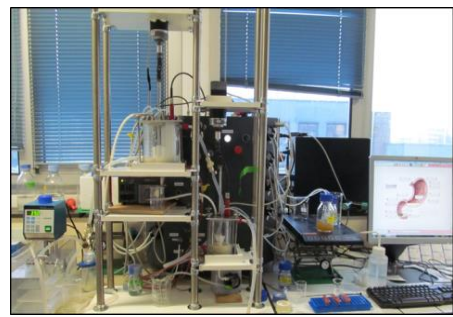
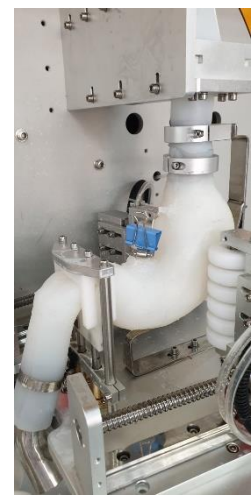
Le Feunteun et al.
2014
Food Bioprocess
Tech

*In silico
models*

$$\Phi_{12} = k_{12whey} \times (V_1 - m_{caswpd1} \times \alpha) + k_{12aggr} \times m_{caswpd1} \times \alpha$$



Menard et al. 2014,
Food Chem
Sanchez et al. 2015
Food Res Int



*In vitro dynamic models
(infant, adult, elderly)*



*Human
models*



Animal models



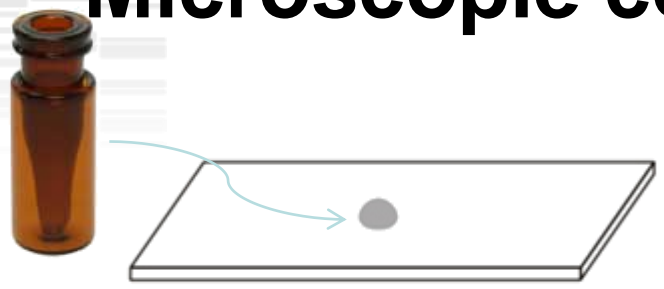
De Oliveira et al. 2016
Am J Clin Nutr
De Oliveira et al. 2017
Clin Nutr

Barbé et al. 2013, 2014
Food Chem
Le Huerou-Luron et al.
2016 Eur J Nutr

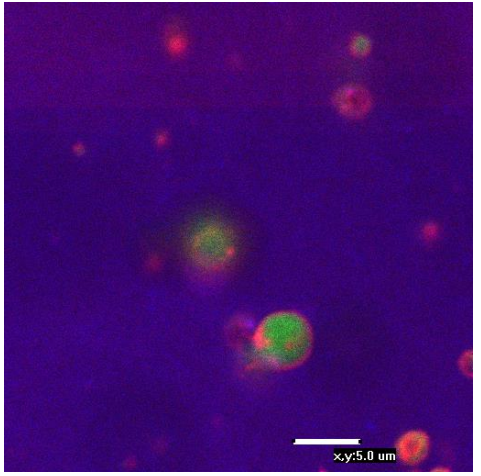


Microscopie confocale

Digestas
+ 3 sondes
fluorescentes

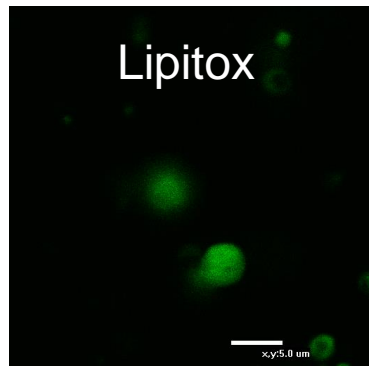


OBS X60

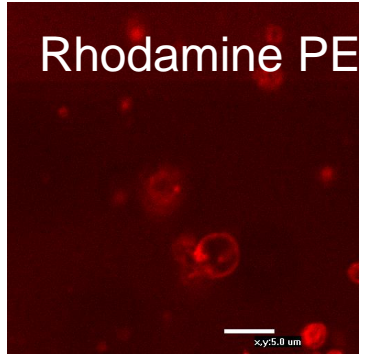


3 sondes

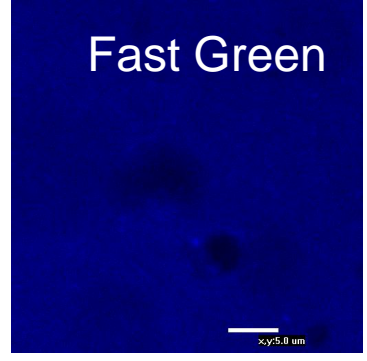
Lipides
Apolaires



Amphiphiles



Protéines

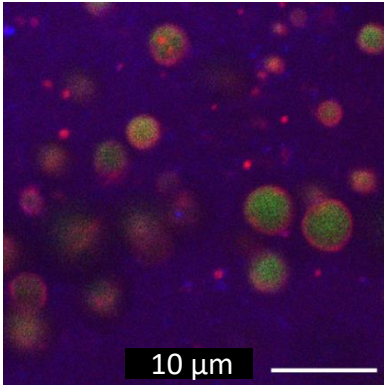


Pasteurization affected the initial structure and the emulsion disintegration of HM

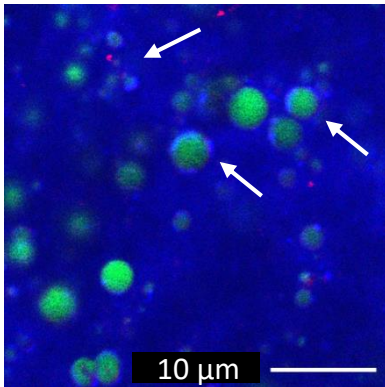
(n = 6 infants)

Initial structure

Raw HM



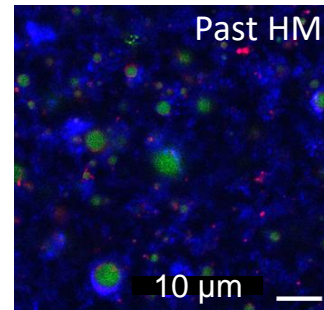
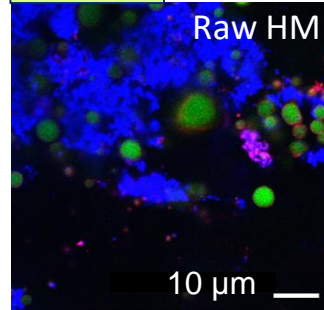
Past HM



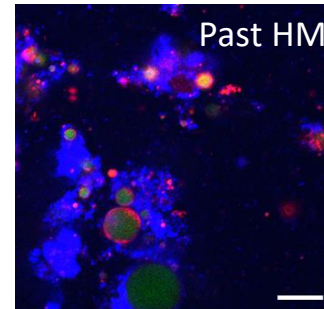
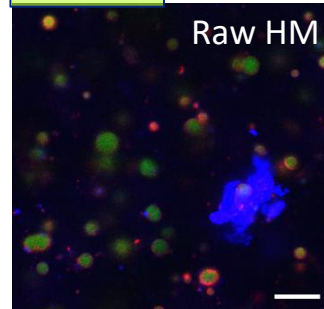
→ Protein heat-induced aggregation

Gastric disintegration

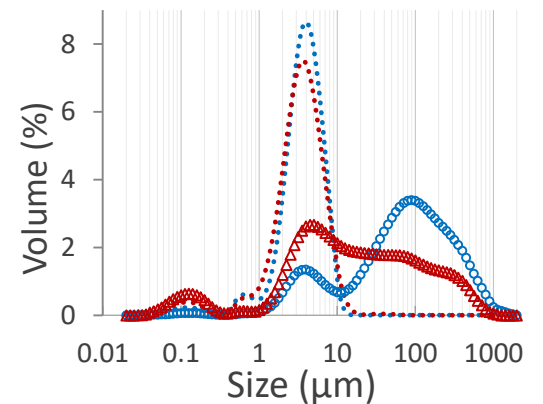
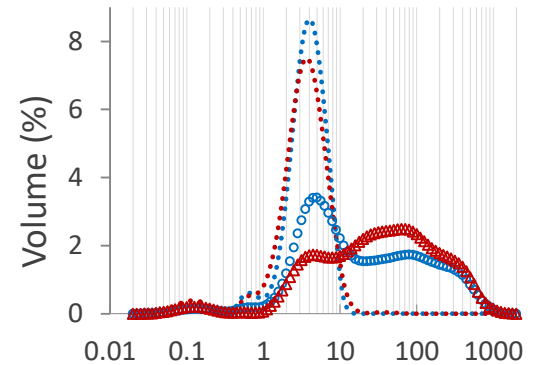
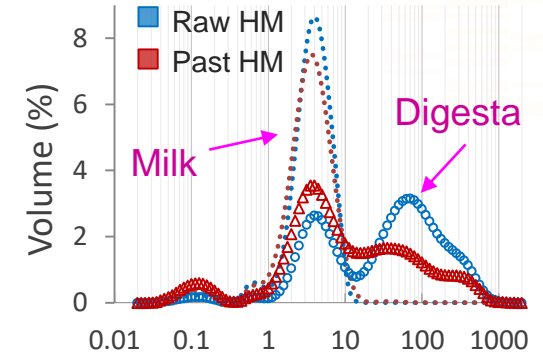
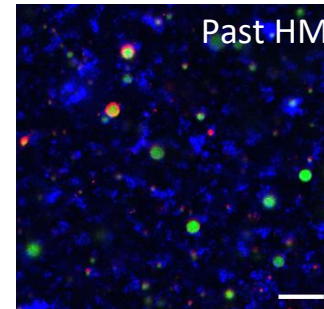
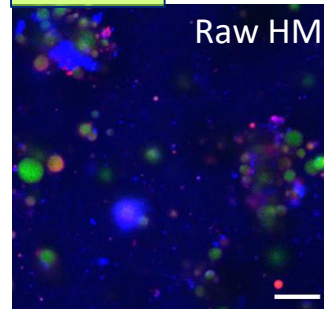
35 min



60 min



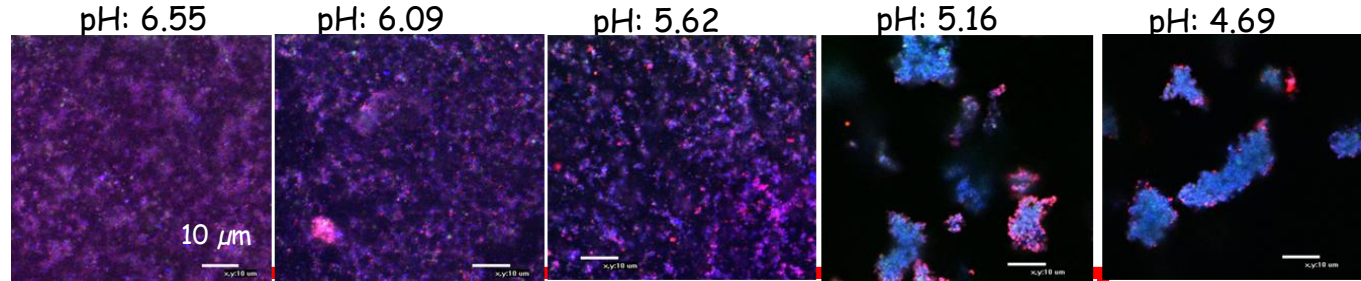
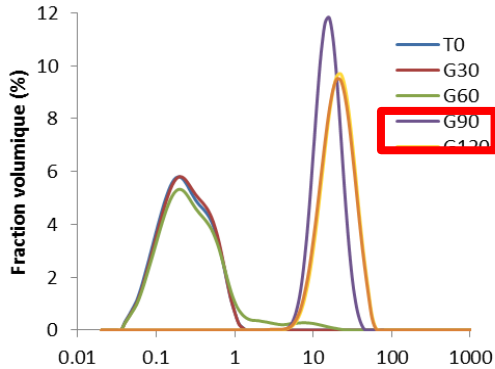
90 min



■ Apolar lipids ■ Amphiphiles ■ Proteins

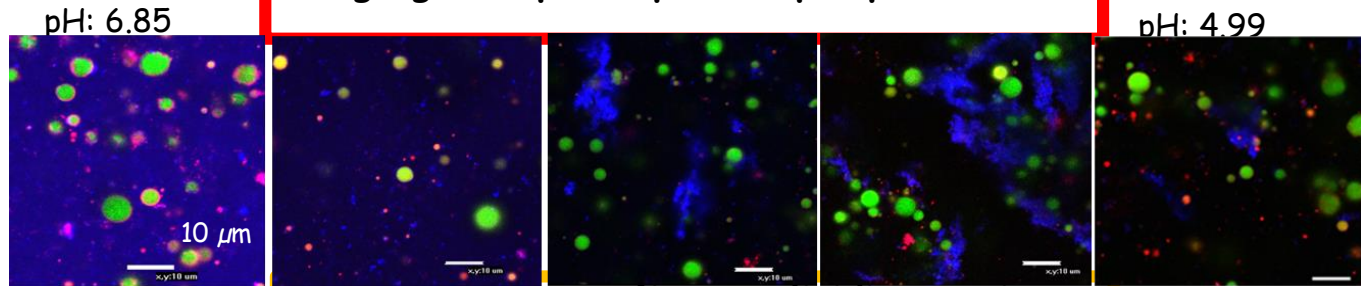
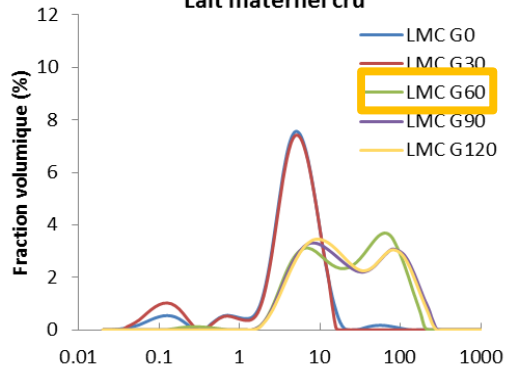
STRUCTURE - PHASE GASTRIQUE

Formule Infantile



Agrégation protéique et lipidique à G90

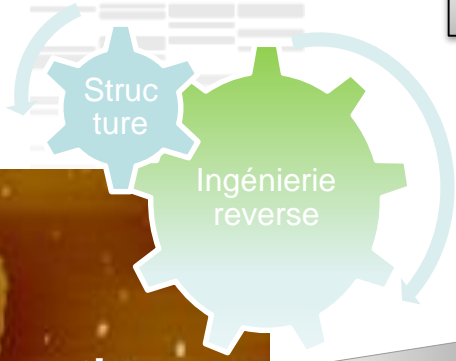
Lait maternel cru



**Agrégation protéique et lipidique moins prononcée à G60 - distribution bimodale
Reste quelques GG natifs**

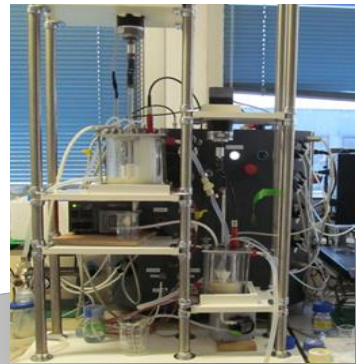
■ Protéines ■ Apolaires ■ Amphiphiles





Etude interactions lipases / substrat laitier en interface plane et en émulsion

in vitro digestion



Samira de Oliveira (FRI)

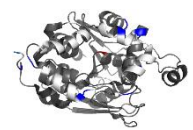
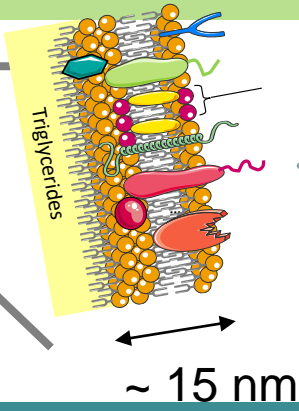
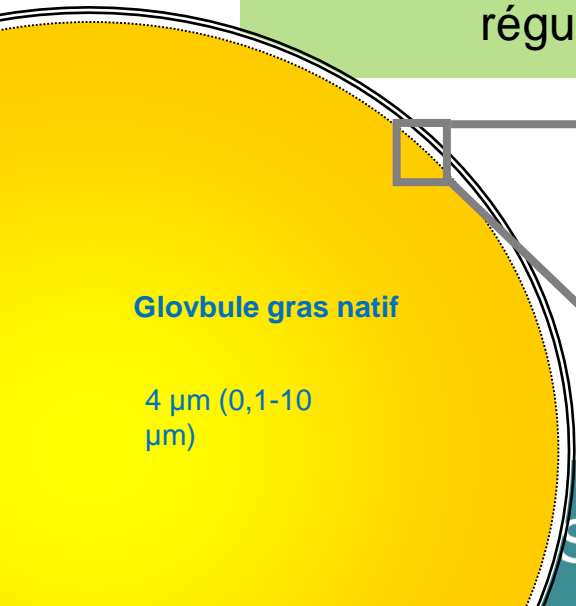
in vivo digestion



ARCHILACT clinical trial (2014-2015)

Systèmes de complexité croissante

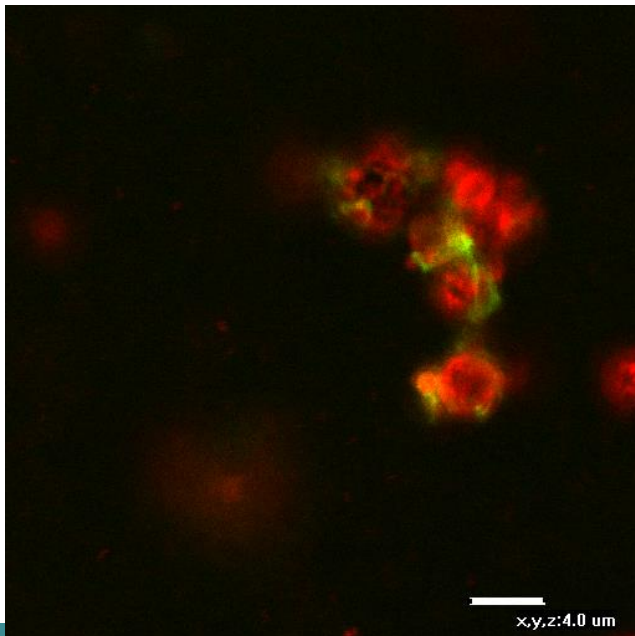
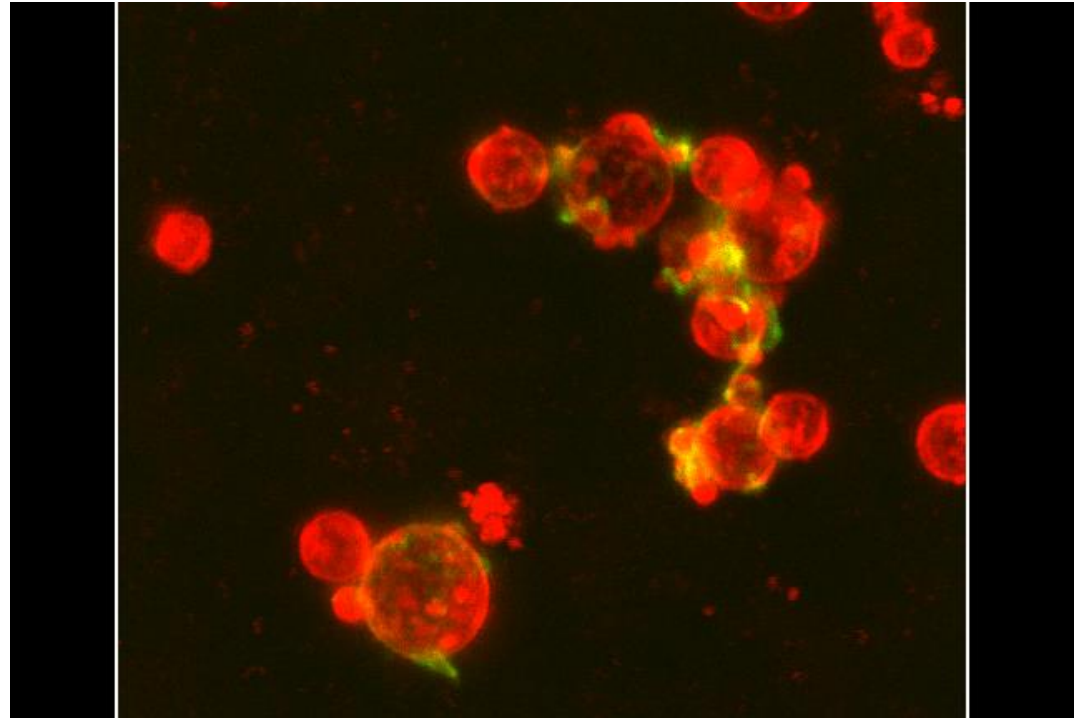
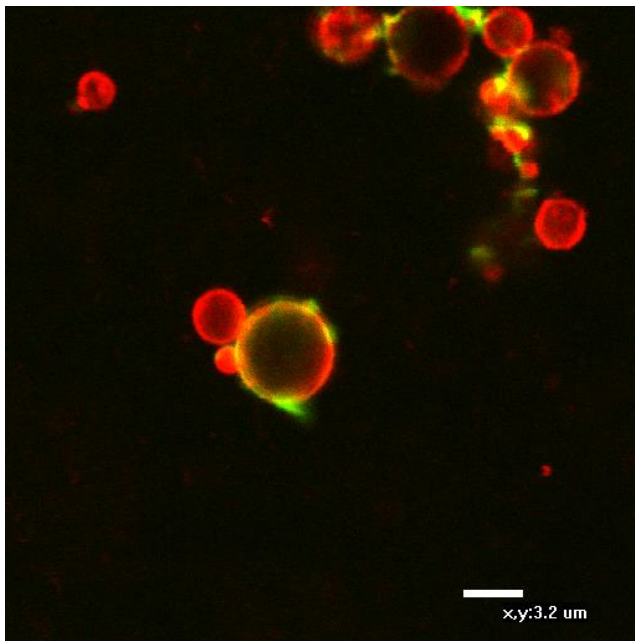
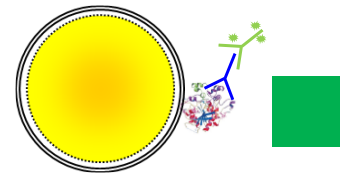
En quoi la composition et l'organisation latérale de la membrane de globule gras de lait (bovin ou humain) régulent l'activité des lipases digestives ?



Lipase gastrique humaine

$S=19.6 \text{ nm}^2$
 $\varnothing=5 \text{ nm}$

Immunolocalization lipase gastrique(pH 6)



Observation des micro-domaines **membranaires**
Lipase adsorbée à l'interface (colocalisation zones
rouge et verte) – mais limite de résolution en
microscopie confocale

→ COMPLEMENT synchrotron SOLEIL – ligne UV

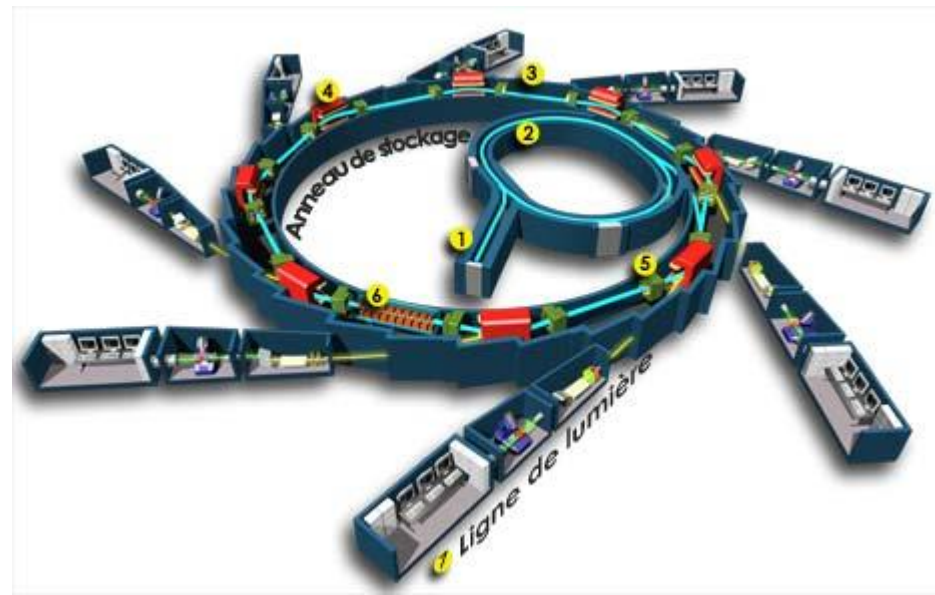


Soleil is a particle (electron) accelerator that produces the synchrotron radiation, an extremely powerful source of light that permits exploration of inert or living matter

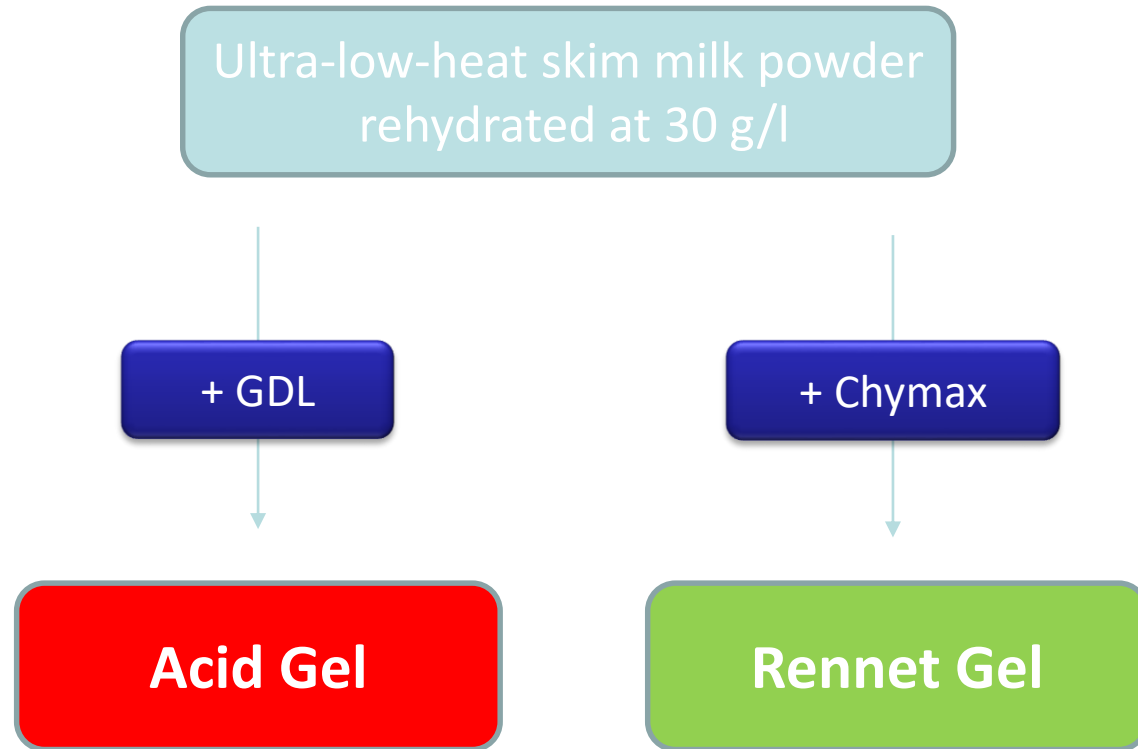


DISCO is a VUV to visible beamline dedicated to biochemistry, chemistry and cell biology. The spectral region is optimized between 60 and 700 nm with conservation of the natural polarization of the light

☞ Allow the imaging of protein intrinsic fluorescence with a UV microscope



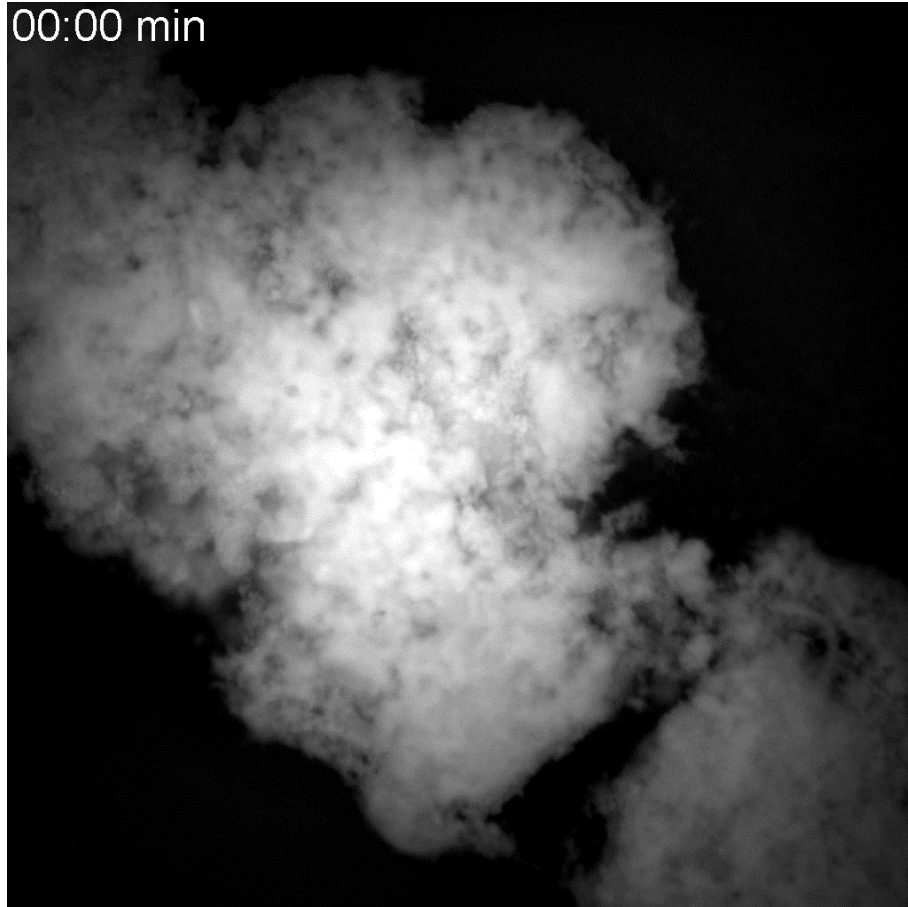
Protocol



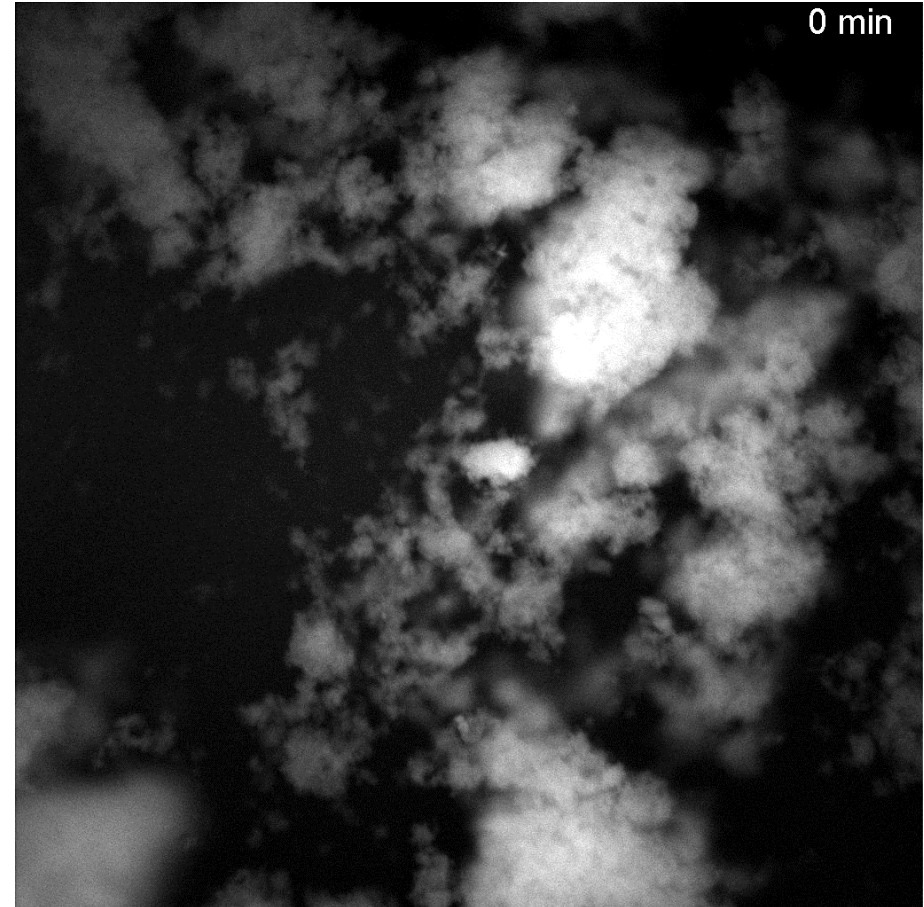
Gels soaked in SGF at pH 2.0 with or without pepsin (2000 U/mL)

Observation 2h UV microscope (x10 and x100)

Kinetics of gel particles disintegration



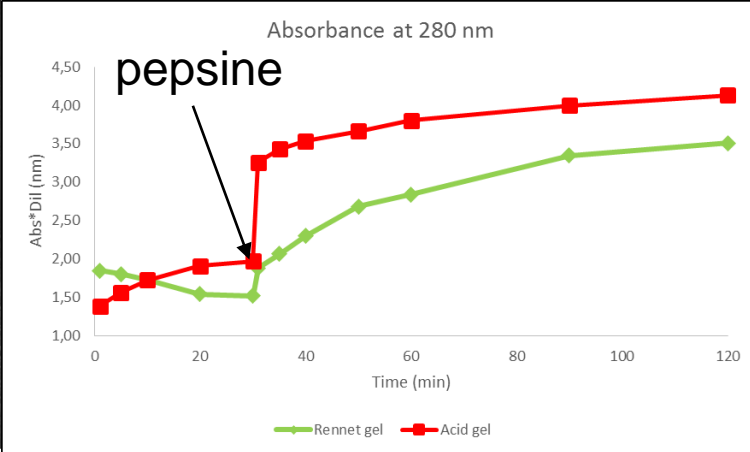
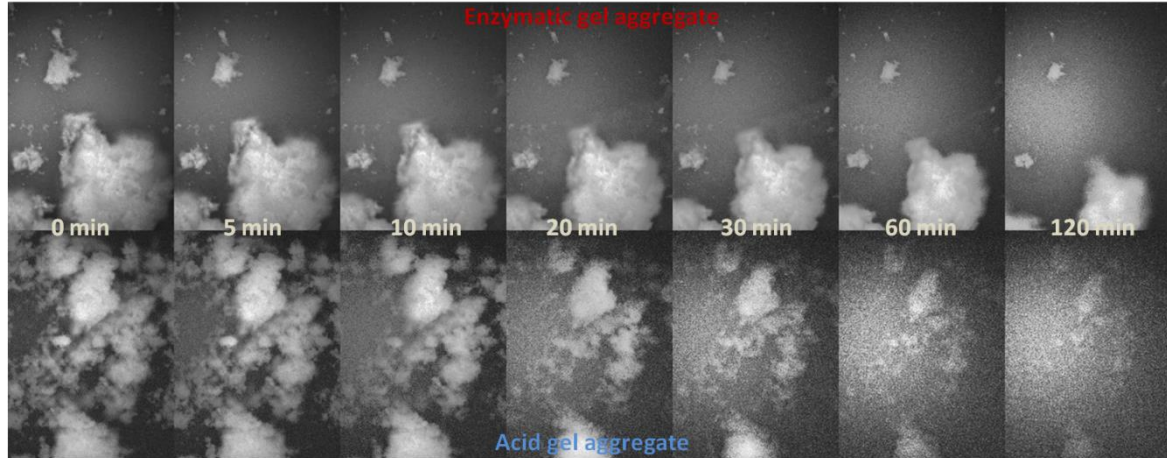
Rennet Gel



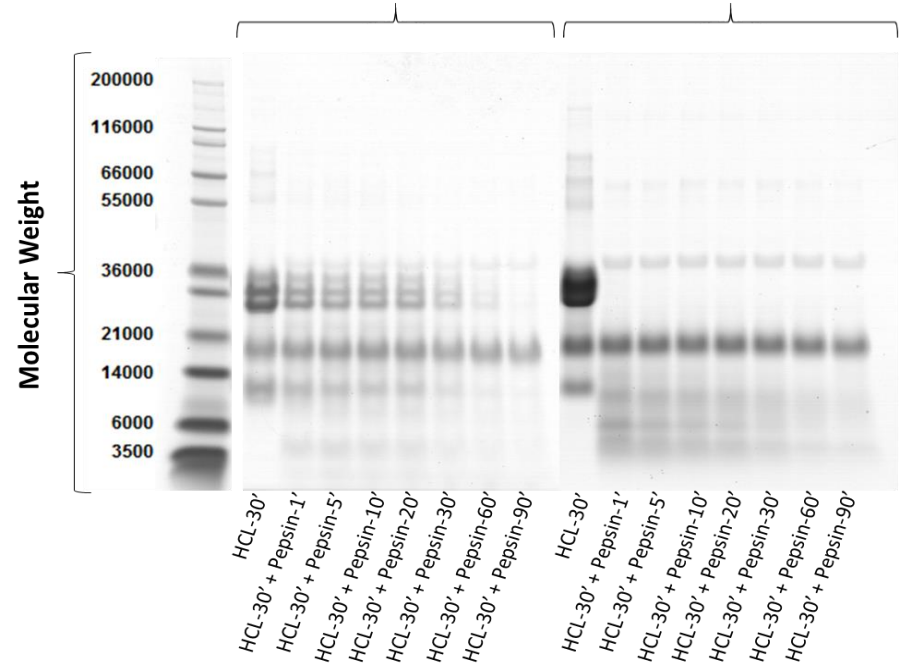
Acid Gel

Disintegration of an acid gel particle is faster than that of a rennet gel

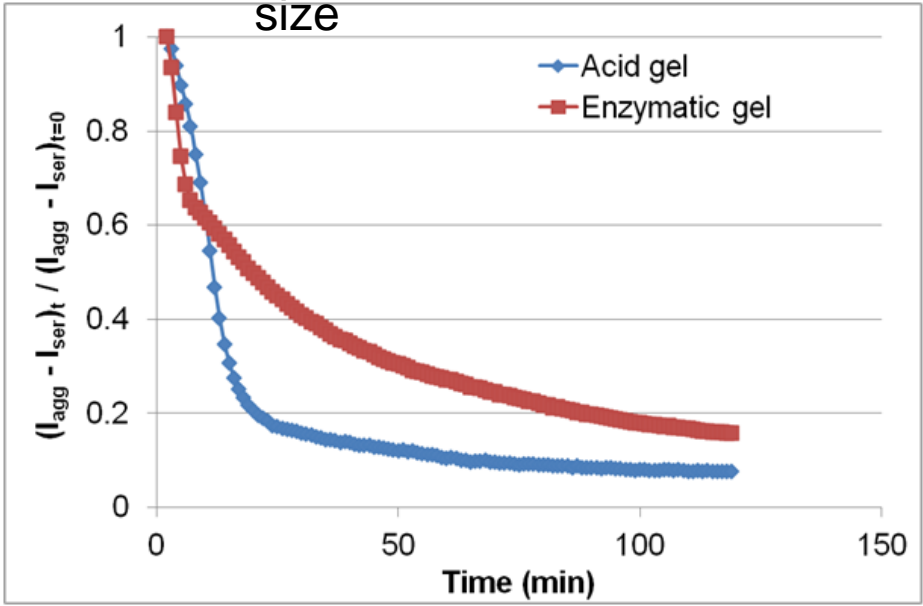
Evolution of soluble proteins



Rennet gel Acid gel



Evolution of particle size



The Bioactivity & Nutrition Team

20-25 people, 15 permanent staff



2015-2021 : 12 PhD, 7 post-docs, 5 international visiting scientists

Head

Didier DUPONT – DR

Scientists

Françoise NAU – PR
Amélie DEGLAIRE – MC
Juliane FLOURY – MC
Catherine GUERIN – MC
Frédérique PEDRONO – MC
Steven LE FEUNTEUN – CR
Martine MORZEL – CR



PhDs

Elise CHARTON (2019-2022)
Lucile CHAUVET (2020-2023)
Ousmane SUWAREH (2019-2022)
Jiajun FENG (2020-2023)
Rozenn LE FOLL (2020-2023)

Post-docs

Ines GRECO (2021-2023)
Stefano NEBBIA (2020-2022)
Imen JEBALIA (2021-2023)

Engineers & Technicians

Yann LE GOUAR – TR
Gwénaële HENRY – AI
Julien JARDIN – IE
Olivia MENARD – IE
Jordane OSSEMOND – IE
Marie-Françoise COCHET – IE



We are pleased to announce the next
8th International Conference on Food Digestion



in Porto, Portugal, April 2024