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1 **Short- and long-term responses of leaf day respiration to elevated**
2 **atmospheric CO₂**

3
4 **Running title:** CO₂ response of leaf respiration in the light

5
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24 (<https://academic.oup.com/plphys/pages/General-Instructions>) is Xiao Ying Gong.

25

1 AUTHOR CONTRIBUTIONS

2 X.Y.G. designed and planned the research; Y.R.S. and Y.N.X. performed the experiment, Y.R.S.
3 and W.T.M. analyzed the data and wrote the first draft, and all authors discussed the results and
4 implications and contributed to the revision.

6 ABSTRACT

7 Evaluating leaf day respiration rate (R_L), which is believed differ from that in the dark (R_{Dk}), is
8 essential for predicting global carbon cycles under climate change. Several studies have
9 suggested that atmospheric CO_2 impacts R_L . However, the magnitude of such an impact and
10 associated mechanisms remain uncertain. To explore the CO_2 effect on R_L , wheat (*Triticum*
11 *aestivum*) and sunflower (*Helianthus annuus*) plants were grown under ambient (410 ppm) and
12 elevated (820 ppm) CO_2 mole fraction ($[CO_2]$). R_L was estimated from combined gas exchange
13 and chlorophyll fluorescence measurements using the Kok method, the Kok-Phi method, and a
14 revised Kok method (Kok- C_c method). We found that elevated growth $[CO_2]$ led to an 8.4%
15 reduction in R_L and a 16.2% reduction in R_{Dk} in both species, in parallel to decreased leaf N and
16 chlorophyll contents at elevated growth $[CO_2]$. We also looked at short-term CO_2 effects during
17 gas exchange experiments. Increased R_L or R_L/R_{Dk} at elevated measurement $[CO_2]$ were found
18 using the Kok and Kok-Phi methods, but not with the Kok- C_c method. This discrepancy was
19 attributed to the unaccounted changes in C_c in the former methods. We found that the Kok and
20 Kok-Phi methods underestimate R_L and overestimate the inhibition of respiration under low
21 irradiance conditions of the Kok curve, and the inhibition of R_L was only 6%, representing 26%
22 of the apparent Kok effect. We found no significant long-term CO_2 effect on R_L/R_{Dk} , originating
23 from concurrent reduction in R_L and R_{Dk} at elevated growth $[CO_2]$, and likely mediated by
24 acclimation of nitrogen metabolism.

25
26 **Keywords:** leaf day respiration, Kok method, atmospheric CO_2 , photosynthesis, climate change,
27 carbon balance, mesophyll conductance, chlorophyll fluorescence

1 INTRODUCTION

2 Terrestrial vegetation assimilates ca. 120 pg carbon via photosynthesis but releases about half of
3 assimilated carbon via respiration (Gifford, 2003; Dusenge et al., 2019). The balance between
4 plant respiration and photosynthesis is therefore essential for plant productivity and global
5 carbon balance. Despite considerable variations depending on N fertilisation and climatic
6 conditions, the ongoing increase in atmospheric CO₂ mole fraction ([CO₂]) promotes leaf
7 photosynthesis and primary production, which is referred to as the “CO₂ fertilization effect”
8 (Drake et al., 1997; Cramer et al., 2001). Although the CO₂ fertilization effect on biomass (but
9 not necessarily yield) is evident from greenhouse and field experiments (Ainsworth and Long,
10 2005; Norby et al., 2005; Walker et al., 2021), the response of plant respiration to [CO₂] is rather
11 uncertain, limiting our ability to predict future climate change-driven modifications of plant
12 physiology.

13 The respiratory response is complicated by the fact that leaf respiration takes place not
14 only in darkness (the respiration rate is denoted as R_{Dk}), but also in the light. In illuminated
15 leaves, respiration is referred to as "respiration in the light" or "day respiration" (denoted as R_L ;
16 here we refer to CO₂ evolution rather than O₂ consumption). Leaf respiration has been shown to
17 be partially inhibited by the light although the magnitude of inhibition varies broadly, with
18 reported R_L/R_{Dk} values ranging from 0.2 to 1.3 (Ayub et al., 2011; Griffin and Turnbull, 2013;
19 Crous et al., 2017; Gong et al., 2018; Way et al., 2019). Given the longer light periods during the
20 growing season and higher temperature during the day than at night in most ecosystems, R_L is a
21 key component of plant- and community-scale carbon budgets (Atkin et al., 2007; Gong et al.,
22 2017). Experimental results revealed that the inhibition of respiration by light (i.e., $1-R_L/R_{Dk}$)
23 also occurs at the stand scale (Gong et al., 2017). Neglecting respiration inhibition might have
24 led to considerable errors in estimated gross primary production (Wehr et al., 2016; Gong et al.,
25 2017). Furthermore, the response of R_L to environmental cues are essential to predict carbon
26 balance, carbon use efficiency and improve land surface models (Wehr et al., 2016; Atkin et al.,
27 2017; Tcherkez et al., 2017b; Keenan et al., 2019).

28 So far, there is no consensus on the response of R_L to long-term [CO₂] increase. Some
29 studies have shown that R_L is stimulated by elevated growth [CO₂] (Wang et al., 2001; Shapiro et
30 al., 2004; Crous et al., 2012; Griffin and Turnbull, 2013), and this effect may be related to higher
31 carbohydrate concentrations in leaves (Rogers et al., 2004; Gong et al., 2017). Also, increased

1 leaf respiration at elevated $[\text{CO}_2]$ has been suggested to be associated with a larger mitochondrial
2 number per mesophyll cell (Griffin et al., 2001), indicating cellular and transcriptional (gene
3 regulation) mechanisms of respiratory control (Leakey et al., 2009). Other studies have reported
4 a decrease in R_L in plants grown under elevated $[\text{CO}_2]$ compared with that grown under ambient
5 $[\text{CO}_2]$ (Ayub et al., 2011; Ayub et al., 2014).

6 The decrease in R_L at elevated $[\text{CO}_2]$ has been suggested to be linked to either
7 photorespiration or nitrogen metabolism. Under elevated CO_2 , there is a reduction in
8 photorespiration rate (and the rate of oxygenation of RuBP, v_o), and this could cause an alteration
9 in R_L , as suggested by results obtained on short-term changes in respiratory metabolism under
10 varying CO_2 mole fraction. In effect, using ^{13}C -enriched substrates to trace decarboxylation
11 processes, Tcherkez *et al.* (2008) found that decarboxylation decreased when leaves were
12 exposed to elevated $[\text{CO}_2]$ for short periods. Likewise, results obtained using the Kok method
13 suggested there was a linear relationship between photorespiration rate and R_L (Griffin and
14 Turnbull, 2013). However, the mechanism behind this relationship is still unclear. In particular,
15 the Kok effect itself has been shown not to be fully caused by changes in respiration rate
16 (Gauthier et al., 2020) and thus, the relationships between photorespiration and Kok method-
17 based R_L are presently uncertain. In addition, R_L has been reported to either decrease (Pinelli and
18 Loreto, 2003; Tcherkez et al., 2008; Griffin and Turnbull, 2013), increase (Yin et al., 2020; Fang
19 et al., 2022) or remain unaffected (Sharp et al., 1984; Tcherkez et al., 2012), in the short-term
20 using gas-exchange experiments at elevated $[\text{CO}_2]$. Thus, conclusions drawn from short-term
21 changes in R_L caused by instantaneous elevation of $[\text{CO}_2]$ might not be relevant to long-term
22 changes in R_L .

23 The decrease of R_L at elevated $[\text{CO}_2]$ has also been suggested to be linked to nitrogen
24 metabolism. It has been observed in many free air CO_2 enrichment (FACE) experiments that
25 elevated $[\text{CO}_2]$ reduces leaf N content, which is accompanied by a down-regulation of
26 photosynthetic capacity (Long et al., 2004; Ainsworth and Long, 2005). It is believed that
27 elevated $[\text{CO}_2]$ inhibits N assimilation in leaves via the potential link between photorespiration
28 and nitrate assimilation (Bloom et al., 2010; Bloom et al., 2014; Busch et al., 2018). Given that N
29 assimilation in leaves is energy demanding and thus a driving factor for leaf respiration (Amthor,
30 2000; Reich et al., 2008), it would be important to know whether $[\text{CO}_2]$ affected R_L and R_{Dk} via

1 leaf N content. All in all, the response of R_L to elevated $[\text{CO}_2]$ appears to be highly variable and
2 mechanisms behind are unclear.

3 Another uncertainty associated with R_L and how it varies is technological. In fact, there are
4 several methods to estimate R_L , but none of them can measure R_L directly (for a review see
5 (Tcherkez et al., 2017b)). The Kok method (Kok, 1949) and the Laisk method (Laisk, 1977), the
6 two most commonly used methods, require manipulation of net CO_2 assimilation rates (A) at low
7 irradiances ($I_{\text{inc}} < 150 \mu\text{mol m}^{-2} \text{s}^{-1}$) (Kok) or low CO_2 (Laisk). Another method, the ^{13}C isotopic
8 disequilibrium method, uses two CO_2 sources with different $\delta^{13}\text{C}$ values to disentangle R_L and
9 photosynthesis under physiologically relevant environmental conditions without the need to
10 manipulate A (Gong et al., 2015; Gong et al., 2018). The ^{13}C disequilibrium method is valuable
11 since it does not require the use of low irradiance or low CO_2 and can be performed at any CO_2
12 mole fraction, and therefore, is suitable to study CO_2 effects on R_L . It is, however, technically
13 demanding (isotopic CO_2 sources, mass spectrometers). The Laisk method is, by definition, not
14 suitable for studying CO_2 effects because it manipulates $[\text{CO}_2]$ at sub-ambient levels. So far, the
15 response of R_L to $[\text{CO}_2]$ has mainly been estimated using the Kok method. However, as
16 mentioned above, the Kok method has been questioned since the Kok effect is not exclusively
17 caused by a decrease in respiration rates (Gauthier et al., 2020). Several studies showed that the
18 Kok method has conceptual uncertainties (Farquhar and Busch, 2017; Tcherkez et al., 2017a;
19 Tcherkez et al., 2017b; Yin et al., 2020). First, the Kok method assumes a constant
20 photochemical efficiency of PS II (Φ_2) along the A - I_{inc} curve (i.e. the Kok curve, see *Theory*). To
21 address this issue, Yin et al. (2009) suggested to use measured Φ_2 to improve the R_L estimation.
22 Second, the Kok method usually disregards variation in chloroplastic $[\text{CO}_2]$ (C_c) along the A - I_{inc}
23 curve, which could bias the estimates of R_L according to recent studies based on model analysis
24 (Farquhar and Busch, 2017; Yin et al., 2020). Estimating C_c along the A - I_{inc} curve requires
25 measurements of mesophyll conductance (g_m). Measuring g_m is challenging and this is
26 particularly true when measurements are performed at low irradiance (Pons et al., 2009; Gu and
27 Sun, 2014; Gong et al., 2015). So far, the uncertainty associated with C_c has not been fully
28 solved.

29 Taken as a whole, neither long-term nor short-term responses of R_L to CO_2 mole fraction
30 are well-known, and technologies used to measure R_L may be problematic. Here, we intend to
31 address the following questions: 1) how do short-, medium- and long-term CO_2 enrichment

1 affect R_L in C_3 leaves? 2) do the original- and revised Kok methods provide similar estimations
 2 of R_L ? To this end, we combine gas exchange and chlorophyll fluorescence (ChF) measurements
 3 to study the response of R_L of wheat (*Triticum aestivum* L.) and sunflower (*Helianthus annuus*
 4 L.) plants grown under ambient (410 ppm) and elevated $[CO_2]$ (820 ppm). We assessed the
 5 medium-to-long term CO_2 response (days to months) by comparing parameters of plants at
 6 different growth $[CO_2]$, and the short-term CO_2 response (minutes) by measuring the same leaves
 7 at 410 and 820 ppm of $[CO_2]$. We compared R_L estimated by the Kok method, the Yin method
 8 (i.e. the Kok-Phi method) and a revised Kok method (i.e. the Kok- C_c method) which takes the
 9 influence of Φ_2 and C_c into account.

11 RESULTS

12 *Effects of growth CO_2 on photosynthetic parameters and leaf traits*

13 Growth at elevated $[CO_2]$ led to a reduction in net CO_2 assimilation (A) for both species, when A
 14 values were compared at the same intercellular CO_2 concentration (C_i) levels (Fig. 1AB).
 15 Sunflower plants grown at elevated $[CO_2]$ exhibited lower E and g_{sw} compared with that grown
 16 at ambient CO_2 (Fig. 1DF). This effect on water vapour exchange was minor in wheat (Fig.
 17 1CE). In order to assess the long-term growth CO_2 effect on common grounds, gas exchange
 18 parameters of leaves were compared at their respective growth $[CO_2]$ (indicated by the subscript
 19 ‘growth’). Net CO_2 assimilation rate (A_{growth}), intrinsic water-use efficiency ($iWUE_{growth}$) and leaf
 20 carbon-use efficiency (CUE_L) of plants grown under elevated $[CO_2]$ were significantly higher
 21 than those of plants grown under ambient $[CO_2]$ in both species (Table 1). Averaged across
 22 species, growth at elevated $[CO_2]$ led to 5.6% reduction in A_{max} , 7.9% reduction in V_{cmax} , and
 23 8.0% in J , indicating a decline in photosynthesis capacity. The ratio of g_{sc} to g_m was not
 24 significantly affected by growth $[CO_2]$ or species. R_{Dk} of both species was lower at elevated
 25 $[CO_2]$ but this decrease differed between species (20% for wheat and 11% for sunflower).

26 Leaf chlorophyll content was significantly lower at elevated $[CO_2]$ compared with ambient
 27 $[CO_2]$. Similarly, elevated $[CO_2]$ led to 6.7% reduction (averaged across species) in nitrogen
 28 elemental content (N%) and 12% reduction in nitrogen content per surface area (N_{area}) on
 29 average, but the effect of CO_2 was not significant at a p -level of 0.05. SLA was significantly
 30 different between species but not affected by growth $[CO_2]$ (Table 1).

31

1 *CO₂ response of R_L estimated by different methods*

2 Φ_2 , C_c , and γ , the key parameters associated with assumptions in both original and revised Kok
3 methods, were found to decrease along the Kok curve in all species and treatments (Fig. 2). With
4 the increase of I_{inc} from 40 to 100 $\mu\text{mol m}^{-2} \text{s}^{-1}$, Φ_2 decreased by 3.1% for wheat and 2.4% for
5 sunflower and this trend was not substantially influenced by growth $[\text{CO}_2]$ (long term effect) and
6 measurement $[\text{CO}_2]$ (short-term effect). A short-term CO_2 effect on γ was detected, i.e., γ
7 decreased more strongly at measurement $[\text{CO}_2]$ of 410 ppm (by 5.2%) than that at measurement
8 $[\text{CO}_2]$ of 820 ppm (by 3.0%, averaged across species) with the increase in I_{inc} (Fig. S1). That is,
9 under our conditions, terms $(\gamma f_{act} \Phi_2 \rho_2 \alpha)$ in Eqn 3 and $(\gamma f_{act} \rho_2 \alpha)$ in Eqn 4 were not constant along a
10 Kok curve, causing errors in R_L estimated by the Kok and the Kok-Phi method, respectively.

11 Applying the Kok, the Kok-Phi and the Kok- C_c method, A was plotted against I_{inc} , $\Phi_2 I_{inc}$,
12 and $\gamma \Phi_2 I_{inc}$, respectively (Fig. 3). Both growth $[\text{CO}_2]$ and measurement $[\text{CO}_2]$ had impacts on A -
13 I_{inc} and $A-\Phi_2 I_{inc}$ response curves (Fig. 3A-D). As a result, growth at elevated $[\text{CO}_2]$ led to a
14 significant decrease in $R_{L \text{ Kok}}$ and $R_{L \text{ Kok-Phi}}$. The same was true for $R_{L \text{ Kok-}C_c}$ on average but it was
15 only significant with a p -value of 0.06 (Fig. 4, Table 2). There was a clear, although statistically
16 insignificant ($p > 0.05$), tendency for elevated measurement $[\text{CO}_2]$ to increase both $R_{L \text{ Kok}}$ and $R_{L \text{ Kok-Phi}}$
17 (Fig. 3 and Fig. 4) in both species. By contrast, $A-\gamma \Phi_2 I_{inc}$ curves obtained under different
18 measurement $[\text{CO}_2]$ seemed to coincide perfectly (Fig. 3EF), in agreement with the insignificant
19 effect of measurement $[\text{CO}_2]$ on $R_{L \text{ Kok-}C_c}$.

20
21 *CO₂ response of R_L/R_{DK} estimated by different methods*

22 We found no significant long-term CO_2 effect on R_L/R_{DK} estimated via all three methods (Table
23 3). There is a tendency that R_L/R_{DK} of wheat increased with the growth $[\text{CO}_2]$ for all three
24 methods (comparing a CO_2 -410 and e CO_2 -820), while that tendency was not found in sunflower.
25 That is, the long-term CO_2 effect on R_L/R_{DK} is not conclusive. Under elevated measurement
26 $[\text{CO}_2]$, significant increases in $R_{L \text{ Kok}}/R_{DK}$ and $R_{L \text{ Kok-Phi}}/R_{DK}$ were observed, but this short-term
27 response was not observed using the Kok- C_c method (Fig. 5). These results indicate that short-
28 term CO_2 effect on $R_{L \text{ Kok}}$ and $R_{L \text{ Kok-Phi}}$ could result from a technical bias simply due to
29 neglecting the change in C_c along the Kok curve.

30 When pooling all data across species and treatments together, R_{DK} was positively correlated
31 to $R_{L \text{ Kok}}$ ($r^2=0.82$, $p<0.05$), $R_{L \text{ Kok-Phi}}$ ($r^2=0.82$, $p<0.05$) and $R_{L \text{ Kok-}C_c}$ ($r^2=0.77$, $p<0.05$) (Fig. 6).

1 These linear regressions yielded an average $R_L \text{ Kok}/R_{Dk}$ of $0.78 \pm 0.04(\text{SE})$, $R_L \text{ Kok-Phi}/R_{Dk}$ of
 2 0.85 ± 0.04 , and $R_L \text{ Kok-Cc}/R_{Dk}$ of 0.94 ± 0.04 . That is, the Kok-Cc method showed a small light-
 3 induced inhibition of respiration, of 6% only, thus much lower than inhibition values from the
 4 other two methods (22%, 15%).

6 **DISCUSSION**

7 **Growth at elevated CO₂ leads to reduction in R_L**

8 This study showed that R_L of plants grown at elevated [CO₂] was lower than that at ambient
 9 [CO₂], and this result was confirmed by all three methods: Kok, Kok-Phi and Kok-Cc. On
 10 average, elevated [CO₂] led to an 8.4% reduction in R_L , regardless of the method. This is in
 11 agreement with previous findings that leaf R_L of plants grown at elevated [CO₂] is lower (Ayub
 12 et al., 2014) despite opposite findings (Wang et al., 2001, Shapiro et al., 2004).

13 Interestingly, although a long-term CO₂ effect on R_L was evident, elevated [CO₂] had no
 14 influence on the R_L/R_{Dk} ratio, because R_{Dk} was also significantly lower at elevated [CO₂]. Similar
 15 to our study, there was no significant long-term CO₂ effect on R_L/R_{Dk} in Sydney blue gum
 16 (*Eucalyptus saligna*) (Ayub et al., 2011; Crous et al., 2012). However, Wang et al. (2001),
 17 Shapiro et al. (2004) and (Gong et al., 2017) found that non proportional changes in R_L and R_{Dk}
 18 led to higher R_L/R_{Dk} ratio in common cocklebur (*Xanthium strumarium*) leaves and sunflower
 19 stands grown at elevated [CO₂]. By contrast, R_L/R_{Dk} was reduced by elevated growth [CO₂] in
 20 wheat because R_L declined (Ayub et al., 2014) or R_{Dk} increased (Griffin and Turnbull, 2013).
 21 Presumably, variations in the response to growth CO₂ between species and conditions might be
 22 linked to differences in nutrient content, metabolism, protein content, etc. which are all related to
 23 respiration.

25 **The long-term response of R_L to CO₂ is associated with changes in leaf N status**

26 Leaf N has long been suggested to be a key parameter influencing respiration rate, and used to
 27 estimate leaf respiration in vegetation models (Atkin et al., 2017). In our study, the reduction in
 28 R_L and R_{Dk} was associated with a decrease in N_{area} and chlorophyll content, suggesting that leaf
 29 N effectively drives the respiration rate. Nitrate reduction and maintenance of proteins are energy
 30 consuming (Wullschlegel et al., 1997). Lower N content implies lower energy requirements and
 31 thus lower growth and maintenance respiration.

1 It has often been found in FACE or growth cabinet experiments that leaf N content was
 2 lower at elevated $[\text{CO}_2]$. This has been explained by different mechanisms. For example,
 3 elevated $[\text{CO}_2]$ was shown to cause a decrease in stomatal conductance of leaves, leading to
 4 decreasing transpiration rates (Ainsworth and Rogers, 2007) and thus, lower transpiration-driven
 5 mass flow of soil N to roots and stems (so-called transpiration mechanism (McGrath and Lobell,
 6 2013; Feng et al., 2015)). Another mechanism is associated with photorespiration. Generally N
 7 assimilation is believed to be lower due to lower photorespiration (Bloom et al., 2010), which is
 8 accompanied by the reduced reductant supplied via photorespiration at elevated $[\text{CO}_2]$ (Taub and
 9 Wang, 2008). Furthermore, a ‘dilution effect’ could occur whereby N uptake does not increase
 10 proportionally to the increase of biomass at elevated $[\text{CO}_2]$ (Feng et al., 2015).

11 The decreased leaf N content at elevated $[\text{CO}_2]$ has also consequences on photosynthetic
 12 capacity (i.e. V_{cmax}). It was reported that species grown under elevated $[\text{CO}_2]$ had lower
 13 maximum apparent carboxylation velocity (V_{cmax}) and carboxylation efficiency (Ainsworth and
 14 Long, 2005). Finally, elevated $[\text{CO}_2]$ significantly increased CUE_L by enhancing photosynthetic
 15 rate and reducing dark respiration. Gong et al. (2017) reported that CUE of sunflower stands was
 16 higher at 200 ppm growth $[\text{CO}_2]$ than that of 1000 ppm growth $[\text{CO}_2]$. This results thus could not
 17 be explained by the response of CUE_L itself since at the leaf level, CUE_L likely increased at
 18 elevated growth $[\text{CO}_2]$. We speculate that the reduction of whole plant CUE in their study was
 19 mainly due to enhanced respiration of heterotrophic organs or exudation.

21 **Changes in Φ_2 and C_c are involved in the Kok effect and impact on R_L estimates**

22 Our study found a short-term CO_2 effect on R_L estimated using the Kok and Kok-Phi method, but
 23 no effect using the Kok- C_c method. In fact, both Kok and Kok-Phi methods showed an increase
 24 in R_L when measured at elevated $[\text{CO}_2]$. This short-term response was in agreement with the
 25 finding of Yin et al. (2020) and Fang et al. (2021), but is not supported by the findings of other
 26 studies (Tcherkez et al., 2008; Griffin and Turnbull, 2013). We believe discrepancies in short-
 27 term CO_2 effect on R_L is mostly associated with methodological differences. As shown in the
 28 *Theory* section, the classical Kok method has conceptual uncertainty with the assumption that Φ_2
 29 remains constant across the Kok curve. This assumption must be rejected as Φ_2 decreases with
 30 increasing I_{inc} (Fig. 2). However, this short-term CO_2 effect on R_L cannot be explained by
 31 changes in Φ_2 because (i) the decrease in Φ_2 along Kok curve was similar at both measurement

1 [CO₂] and (ii) the effect persisted when the Kok-Phi method was used to account for variation in
2 Φ_2 .

3 Another assumption that has been made for both the Kok and Kok-Phi method is that γ
4 (determined by Γ^*/C_c) remains constant throughout the Kok curve. This assumption has also
5 been challenged in recent model analyses (Buckley et al., 2017; Farquhar and Busch, 2017), but
6 the question is how to quantify the change in C_c as this requires g_m estimates. Here, we used
7 species-specific g_{sc}/g_m ratios to calculate C_c , suggesting that C_c and γ decreased with increasing
8 I_{inc} . Importantly, measurement CO₂ influenced the trend of γ with increasing I_{inc} , which might be
9 the origin of this short-term CO₂ effect on $R_{L\text{ Kok}}$ and $R_{L\text{ Kok-Phi}}$. When changes in γ (or Γ^*/C_c) are
10 accounted for, the apparent short-term effect of CO₂ on R_L , as found with the Kok and Kok-Phi
11 methods, became insignificant (see also Fig. 3, 4).

12

13 **Kok- and Kok-Phi based estimates of R_L suppression are overestimates**

14 The inhibition of R_L by light is supported by biochemical evidence. Utilizing ¹³C labelling, flux
15 calculations suggest that decarboxylation rates associated to glucose catabolism and activation of
16 malic enzyme increase with decreasing irradiance in the irradiance region where the Kok effect
17 occurs (Gauthier et al., 2020). Recently, how much of the Kok effect is associated with
18 respiration has been under debate (Farquhar and Busch, 2017; Gauthier et al., 2020; Yin et al.,
19 2020). Indeed, the methods used in the present study show different levels of inhibition of
20 respiration by light. The average R_L/R_{Dk} was 0.74 for the Kok method, 0.81 for the Kok-Phi
21 method and 0.93 for the Kok- C_c method. That is, the change in Φ_2 , γ (or Γ^*/C_c), and real light
22 inhibition of R_L explained *c.* 27, 46 and 27% of the apparent Kok effect (i.e. the apparent 26%-
23 inhibition of R_L found with the classical Kok method), respectively. This is in agreement with the
24 results of previous model analyses which show that the Kok effect is not purely respiratory
25 (Farquhar and Busch, 2017; Yin et al., 2020), and both the Kok method and the Kok-Phi method
26 underestimated R_L and overestimated the inhibition of R_L (Yin et al., 2020).

27 The real light inhibition of R_L (as revealed by the Kok- C_c method) was only 6%, which is
28 close to the mean inhibition of 8% of several herbaceous species determined using the ¹³C
29 disequilibrium method (Gong et al., 2018) and the mean inhibition of 10% in wheat leaves
30 determined using a nonrectangular hyperbolic model to interactively solve g_m and R_L (Fang et al.,

2022). In line with these results, a break point in the linear section of the photosynthetic response curve could hardly be seen in the Kok- C_c plots (Fig. 3EF).

The Kok- C_c method developed here requires g_{sc}/g_m to estimate C_c along a Kok curve since C_c cannot be directly measured. Estimating g_m under low light remains technically very challenging. We used species specific g_{sc}/g_m values measured under the growth condition to estimate g_m at each step of Kok curves. Similar approach has been applied to estimate C_c to improve the Laisk method (Gong et al., 2018; Way et al., 2019). These calculations assume that g_{sc}/g_m was the same under the measurement condition of the Kok method and the growth condition. In another word, g_{sc} and g_m should decrease similarly with the decrease of PPFD. This assumption is supported by experimental results (Flexas et al., 2008; Douthe et al., 2011; Xiong et al., 2015). Estimating g_m from species specific g_{sc}/g_m ratio is supported by the robust relationship between g_{sc} and g_m observed in different species under manipulated CO_2 , irradiance, and drought stress (Flexas et al., 2008; Ma et al., 2021; Gong et al., 2022). Although the g_{sc}/g_m ratio estimated here could have a certain level of uncertainty due to methodological issues associated with g_m estimation (Pons et al., 2009; Gu and Sun, 2014; Gong et al., 2015), R_L Kok- C_c was not very sensitive to g_{sc}/g_m . Importantly, the factor that directly influences R_L Kok- C_c estimation is the decreasing rate of γ with the increase of I_{inc} ($d\gamma/dI_{inc}$) but not absolute values of g_m or C_c . Varying g_{sc}/g_m by ± 0.4 or assuming a constant g_m has little effect on $d\gamma/dI_{inc}$ and a negative $d\gamma/dI_{inc}$ was evident in all cases (Fig. S1, S2). In effect, our sensitivity tests showed that varying g_{sc}/g_m by ± 0.4 has a minor influence on both R_L estimates and the CO_2 effect (Fig. S3). However, R_L/R_{Dk} is sensitive to small variations in R_L and thus is affected by g_{sc}/g_m (Fig. S4). Adjusting g_{sc}/g_m (± 0.4 units) leads to changes of mean light inhibition from 4% to 10%. These results highlighted that accounting for $d\gamma/dI_{inc}$ is essential for estimating R_L (Farquhar and Busch, 2017), and the uncertainty associated with the accuracy of $d\gamma/dI_{inc}$ is much less than assuming a constant γ along a Kok curve. The Kok- C_c based estimates of R_L suppression could be further improved if a new method is developed to precisely estimate g_m at very low light. Taken as a whole, neither the Kok nor Kok-Phi method seem suitable to quantify the inhibition of respiration by light (as also suggested by Yin et al. 2020 and Tcherkez et al. 2017a, 2017b), and the inhibition of R_L at the operating PPFDs of this study should be lower than 10%.

30
31

1 **Conclusions and Perspectives**

2 This study showed that elevated growth [CO₂] reduced R_L and R_{DK} likely as a result of decreasing
 3 leaf N status and chlorophyll content. We found no significant long-term CO₂ effect on R_L/R_{DK} ,
 4 indicating a concurrent response of R_L and R_{DK} to elevated growth [CO₂], mediated by the
 5 adjustment of nitrogen metabolism in leaves. These results shed light into the incorporation of R_L
 6 into the carbon cycling models. We revisited the theoretical basis of the Kok method, revised
 7 Kok methods and discussed their respective limitations. Using Kok and Kok-Phi methods, we
 8 found that R_L were stimulated by short-term CO₂ enrichment, while the effect was not supported
 9 by the data of the Kok- C_c method. We attributed this short-term CO₂ effect to methodological
 10 uncertainty associated with unaccounted changes in γ (or Γ^*/C_c) along a Kok curve. Accounting
 11 for those effects, we found that the Kok and Kok-Phi method underestimate R_L and overestimate
 12 the inhibition of respiration under low irradiance conditions of the Kok method, and the
 13 inhibition of R_L is only $6\pm 4\%$, which represents 26% of the Kok effect (i.e. of the apparent
 14 inhibition of R_L found using the classical Kok method). Although the Kok- C_c method has less
 15 theoretical uncertainty and is thus in principle more reliable, we are aware that all three methods
 16 have operating PPFD much lower than usual, ambient irradiance encountered by plants.
 17 However, estimated R_L could vary with irradiance. Earlier studies have showed a decrease of R_L
 18 with the increase of operating PPFD (Brooks and Farquhar, 1985; Atkin et al., 1998; Atkin et al.,
 19 2000) by using the Laisk method which also has the uncertainty associated with the unaccounted
 20 changes in C_c (Farquhar and Busch 2017). To date, the effect of irradiance on R_L is poorly
 21 known and this should be addressed in subsequent studies.

22 **MATERIALS AND METHODS**

23 **Theory**

24 When estimating R_L with the Kok method, A should be measured at low irradiance, where A is
 25 limited by the light-dependent electron transport rate. According to the equation of the electron
 26 transport-limited photosynthesis (Farquhar et al., 1980), A at low light is described as:

$$27 \quad A = J \frac{1 - \Gamma^*/C_c}{4 + 8\Gamma^*/C_c} - R_L \quad \text{Eqn 1}$$

28 where J is the electron transport rate that is used for CO₂ fixation and photorespiration, Γ^* is the
 29 C_c -based CO₂ compensation point in the absence of mitochondrial respiration ($37.4 \mu\text{mol mol}^{-1}$
 30 at 25°C, (Silva-Perez et al., 2017)). According to the theoretical evaluations of Yin et al. (2011,
 31

2020), Eqn 1 forms the theoretical basis of the Kok method, and is useful for evaluating methodological uncertainties.

In this equation, J can be replaced by $f_{\text{aet}}\Phi_2\rho_2\alpha I_{\text{inc}}$, where f_{aet} is the fraction of electron transport for photosynthesis, ρ_2 is the fraction of absorbed irradiance partitioned to PS II, α is the absorptance by leaf photosynthetic pigments and I_{inc} is incident irradiance (Yin et al., 2011). Here, we define the term $\frac{1-\Gamma^*/C_c}{4+8\Gamma^*/C_c}$ as γ , so that Equation (1) becomes:

$$A = \gamma f_{\text{aet}} \Phi_2 \rho_2 \alpha I_{\text{inc}} - R_L \quad \text{Eqn 2}$$

With the Kok method, net CO₂ assimilation rates are plotted against I_{inc} and datapoints that fall above the breakpoint are used to extrapolate A up the y -axis and thereby estimate R_L . In fact, if the term $\gamma f_{\text{aet}} \Phi_2 \rho_2 \alpha$ is assumed to be constant, thus the intercept of this linear relation provides the estimate of $R_{L \text{ Kok}}$. In terms of equation, this can be written as:

$$A = (\gamma f_{\text{aet}} \Phi_2 \rho_2 \alpha) I_{\text{inc}} - R_{L \text{ Kok}} \quad \text{Eqn 3}$$

However, it has been shown that Φ_2 could decrease with increasing I_{inc} even within the range of low irradiance (Genty and Harbinson, 1996; Yin et al., 2020). Alternatively, Φ_2 can be obtained from chlorophyll fluorescence measurements. Yin et al. (2009) thus suggested to plot A against $\Phi_2 I_{\text{inc}}$ as:

$$A = (\gamma f_{\text{aet}} \rho_2 \alpha) \Phi_2 I_{\text{inc}} - R_{L \text{ Kok-Phi}} \quad \text{Eqn 4}$$

The Yin et al. (2011) method can be considered as a revised Kok method with variation in Φ_2 accounted for, and thus it is renamed as the 'Kok-Phi' method here to highlight the modification. This method assumes that γ is constant across Kok curve, which is obviously not true under photorespiratory conditions, i.e. under ambient conditions where O₂ mole fraction is about 21% (Yin et al., 2014). Theoretically, the Kok-Phi method is applicable for measuring C₃ leaves at nonphotorespiratory conditions or C₄ leaves (Yin et al., 2011; 2020; Fang et al., 2021).

On the basis of these two methods, we propose a revised Kok method, named 'Kok-C_c' method, accounting for variations in γ caused by the decrease in C_c along the Kok curve. In the Kok-C_c method, A should be plotted against $\gamma \Phi_2 I_{\text{inc}}$, the intercept of the linear relation yields the estimation of R_L ($R_{L \text{ Kok-Cc}}$):

$$A = (f_{\text{aet}} \rho_2 \alpha) \gamma \Phi_2 I_{\text{inc}} - R_{L \text{ Kok-Cc}} \quad \text{Eqn 5}$$

This method requires estimates of C_c at each step of the A - I_{inc} curve (see below the section dedicated to C_c estimation). It is worth noting that in practice all 'Kok type' methods, assume

1 that R_L is not sensitive to changes in C_c along the Kok curve, as they rely on linear
2 extrapolations. To our knowledge, this assumption has not been verified (see *Introduction*).

3

4 **Plant material and growth conditions**

5 Sunflower (*Helianthus annuus* L.) and wheat (*Triticum aestivum* L.) plants were grown from
6 seed in plastic pots with garden soil and thinned to one plant per pot. Initial nutrient composition
7 of the garden soil (Scotts Miracle-Gro, USA) was 0.68% N, 0.27% P_2O_5 , and 0.36% K_2O . Plants
8 were randomly placed in two growth chambers, where CO_2 mole fraction was 410 ppm
9 (ambient) and 820 ppm (elevated [CO_2]) respectively. In both chambers, air temperature was
10 maintained at 25°C and the relative humidity of the air was 70% for both light and dark periods.
11 The photosynthetic photon flux density (PPFD) was $700 \mu mol m^{-2} s^{-1}$ for 16-h photoperiod. All
12 plants were watered every 2-3 days to prevent water stress. This experiment had six replicates
13 per treatment, and in total 24 plants were used for measurements.

14

15 **Gas exchange and chlorophyll fluorescence measurements**

16 Photosynthetic gas exchange and ChF parameters were measured when there were 4 fully
17 expanded leaves in each plant (sunflower) or tiller (wheat). Using a portable gas exchange
18 system (LI-6800; Li-Cor Inc., Lincoln, NE, USA), measurements were undertaken on the second
19 youngest fully developed leaves. Light response curves and ChF parameters were measured to
20 estimate R_L . When stable gas exchange rates were achieved, we measured A starting at $120 \mu mol$
21 $m^{-2} s^{-1}$, and the PPFD was sequentially reduced to 100, 80, 60, 40, 20 and 0 (i.e., with light
22 source switched off) $\mu mol m^{-2} s^{-1}$. ChF measurements were done at PPFD of 120, 100, 80, 60 and
23 $40 \mu mol m^{-2} s^{-1}$ using the multi-phase flash method. Φ_2 was calculated as

$$24 \quad \Phi_2 = (F_m' - F_s) / F_m' \quad \text{Eqn 6}$$

25 where F_s is the steady-state fluorescence in the light conditions and F_m' is maximal
26 fluorescence during short saturating pulses of light. For each leaf, the irradiance response of
27 photosynthesis rates was determined at two atmospheric [CO_2] (410 and 820 ppm) to assess
28 short-term CO_2 response of R_L .

29 All gas exchange parameters have been corrected for the leak effect (i.e. CO_2 diffusion
30 across gaskets of leaf chamber) using the measured leak coefficients of intact leaves (Gong et al.,
31 2015; Gong et al., 2018). R_{Dk} measured at 410 and 820 ppm [CO_2] was used to calculate the

1 cuvette leak coefficient for CO₂ (K_{CO_2}) with the leaf present in the leaf chamber using the
 2 equations in (Gong et al., 2015). K_{CO_2} was not significantly different between species and
 3 growth [CO₂], with a mean K_{CO_2} of 0.21 for wheat and 0.30 for sunflower (Fig. S5). Thereafter,
 4 the response of A to [CO₂] (i.e. $A-C_i$ curve) was determined under an irradiance of 700 $\mu\text{mol m}^{-2}$
 5 s^{-1} and varying CO₂, using a [CO₂] sequence of 410, 200, 150, 100, 50, 410, 800 and 1600 μmol
 6 mol^{-1} . ChF parameters were acquired at 200, 410, 800 and 1600 $\mu\text{mol mol}^{-1}$ CO₂. Leaf
 7 temperature was maintained at 25°C for all gas exchange measurements, there is thus no
 8 temperature correction needed to compare R_L and R_{Dk} .

10 Estimation of day respiration and C_c

11 For the Kok method, the data of the linear range of the $A: I_{inc}$ curve at PPFD levels above the
 12 Kok breakpoint (kink) were used to estimate R_L according to Eqn 3. Each $A: I_{inc}$ curve was
 13 visually inspected to identify the irradiance at the Kok breakpoint, which was 40 $\mu\text{mol m}^{-2} \text{s}^{-1}$
 14 (Fig. S6). The data measured at PPFD of 120 $\mu\text{mol m}^{-2} \text{s}^{-1}$ deviated from the linear relation (i.e.
 15 the linear domain of assimilation response curve to light between 40-100 $\mu\text{mol m}^{-2} \text{s}^{-1}$), thus they
 16 were excluded from the dataset used for the estimation of R_L via all methods. Linear regressions
 17 were performed using data of the PPFD levels of 40, 60, 80, and 100 $\mu\text{mol m}^{-2} \text{s}^{-1}$ for all three
 18 methods, with the exception of 5 out of 45 curves in which a point that deviated from the linear
 19 relation was excluded for the estimation of R_L . For the Kok-Phi method, the data from the same
 20 PPFD range were used to estimate R_L by plotting A against $\Phi_2 I_{inc}$ according to Eqn 4. We have
 21 not intensively measured A at very low PPFD levels to accurately identify the breakpoint.
 22 However, the data at 40 $\mu\text{mol m}^{-2} \text{s}^{-1}$ PPFD seem to be above the Kok breakpoint and in the
 23 linear domain of $A: I_{inc}$ curves. Our approach is similar to recent studies which compared the Kok
 24 and the Kok-Phi method (Yin et al., 2011; Fang et al., 2021).

25 Estimating R_L from the Kok- C_c method requires estimates of mesophyll conductance (g_m).
 26 According to the variable J method of Harley et al. (1992), g_m could be calculated as:

$$27 \quad g_m = \frac{A}{C_i \frac{\Gamma^*[J+8(A+R_L)]}{J-4(A+R_L)}} \quad \text{Eqn 7}$$

28 Here, we used R_L estimated using the Kok-Phi method to calculate g_m , given that this method
 29 addresses the issue of decreasing Φ_2 and provides a more reliable estimation of R_L , compared to
 30 the Kok method (Yin et al., 2011). Furthermore, using $R_{L \text{ Kok}}$ or $R_{L \text{ Kok-Phi}}$ has minor influence on

1 g_{sc}/g_m , thus should have no influence on our conclusions (see the discussion on the uncertainty
 2 associated with g_{sc}/g_m). We chose data in a reliable range of dC_c/dA between 10 and 50 for
 3 estimating g_m as suggested by Harley et al. (1992). dC_c/dA was calculated as:

$$4 \quad dC_c/dA = 12\Gamma^*J/(J - 4(A + R_L))^2 \quad \text{Eqn 8}$$

5 Most of the data obtained with sunflower met this empirical criterion of dC_c/dA , while dC_c/dA of
 6 wheat exceeded this range ($dC_c/dA > 100$) in most cases. Therefore, the $A-C_i$ curve-fitting method
 7 was used to estimate the g_m value of each leaf in wheat. Based on the FvCB photosynthesis
 8 model (Farquhar et al., 1980), the $A-C_i$ curve fitting tool developed by Sharkey et al. (2007) was
 9 used to estimate g_m by minimizing the sum of squared deviations between the observed and
 10 modelled data.

11 Recently, it has been found that g_m and stomatal conductance to CO_2 (g_{sc}) are strongly
 12 related (Flexas et al., 2012; Ma et al., 2021). A nearly fixed g_{sc}/g_m ratio across different
 13 environments and plant functional groups was shown by Ma et al. (2021), offering a useful
 14 solution to estimate g_m . We first obtained species- and treatment-specific g_{sc}/g_m using Equation 7
 15 (sunflower) or curve fitting (wheat), and then g_m along the Kok curve was estimated from
 16 measured g_{sc} and previously estimated g_{sc}/g_m . C_c was calculated from g_m as:

$$17 \quad C_c = C_i - A/g_m \quad \text{Eqn 9}$$

18 With C_c , γ could be calculated and thus $R_{L \text{ Kok-Cc}}$ could be estimated by plotting A against $\gamma\Phi_2I_{inc}$
 19 using data of the PPFD range of 40-100 $\mu\text{mol m}^{-2} \text{s}^{-1}$ according to Eqn 5. We also tested the
 20 sensitivity of $R_{L \text{ Kok-Cc}}$ to g_{sc}/g_m by adjusting obtained species- and treatment-specific g_{sc}/g_m (\pm
 21 0.4).

22 The daily carbon-use efficiency of leaves, the ratio of net carbon gain to assimilated carbon
 23 (integrated photosynthesis) was calculated as:

$$24 \quad \text{CUE}_L = (\int A - \int R_{Dk}) / (\int A + \int R_{L \text{ Kok-Cc}}) \quad \text{Eqn 10}$$

25 Since plants were grown in controlled environments, the daily carbon fluxes were calculated as
 26 $\int A = A \times \text{light hours}$, $\int R_{L \text{ Kok-Cc}} = R_{L \text{ Kok-Cc}} \times \text{light hours}$, and $\int R_{Dk} = R_{Dk} \times \text{dark hours}$.

27

28 **Plant sampling and leaf trait parameters**

29 After gas exchange and ChF measurements, the measured leaves were harvested. We measured
 30 leaf area and fresh weight, and the chlorophyll content (Chl) was determined by a chlorophyll
 31 meter (SPAD-502 Plus; Konica Minolta Inc., Tokyo, Japan). The chlorophyll content was

1 calculated from the observed SPAD values as $\text{Chl} = (99 \text{ SPAD}) / (144 - \text{SPAD})$ (Cerovic et al.,
2 2012). All leaves were dried at 70 °C to constant mass after drying to stop enzymatic activity at
3 105 °C for 1 hour. We measured dry mass of individual leaves, and then the leaves were ground
4 with a ball mill (Tissuelyser-24, Jingxin Ltd., Shanghai, China). Leaf N content was measured
5 using an elemental analyzer (VARIO ELIII, Elementar Analysensysteme GmbH, Hanau,
6 Germany).

7

8 **Statistical analysis**

9 Statistical analysis was performed using SPSS (v. 25.0, SPSS, Chicago, IL, USA). Leaf traits and
10 photosynthetic parameters were analyzed with two-way ANOVAs to determine the influence of
11 growth [CO_2], species and their interaction. Besides, ANOVAs were carried out to clarify the
12 effect of growth [CO_2], measurement [CO_2], their interaction and species on R_L and R_L/R_{DK} . A p -
13 value lower than 0.05 is considered statistically significant.

14

15 **FUNDING**

16 This work was supported by the National Natural Science Foundation of China (NSFC
17 31870377, 32120103005).

18

19 **DATA AVAILABILITY**

20 All data that support the findings of this study are included in the published article and its
21 Supplementary Information.

22

1 **TABLES**

2 Table 1. Leaf traits and photosynthetic parameters of wheat (*T. aestivum*) and sunflower (*H. annuus*)
3 grown under ambient or elevated CO₂ (aCO₂ or eCO₂). Leaf trait parameters include: specific leaf area
4 (SLA, cm² mg⁻¹), leaf nitrogen content per dry mass (N%), leaf nitrogen content per area (N_{area}, g m⁻²),
5 chlorophyll content (Chl, g m⁻²). Photosynthetic parameters include net CO₂ assimilation rate at the
6 growth CO₂ (A_{growth} , $\mu\text{mol m}^{-2} \text{s}^{-1}$), maximum CO₂ assimilation rate (A_{max} , $\mu\text{mol m}^{-2} \text{s}^{-1}$), respiration rate in
7 the dark (R_{Dk} , $\mu\text{mol m}^{-2} \text{s}^{-1}$), intrinsic water-use efficiency ($i\text{WUE}_{\text{growth}}$, $\mu\text{mol mol}^{-1}$), maximum
8 carboxylation rates by Rubisco (V_{cmax} , $\mu\text{mol m}^{-2} \text{s}^{-1}$), electron transport rate (J , $\mu\text{mol m}^{-2} \text{s}^{-1}$), ratio of
9 stomatal conductance for CO₂ to mesophyll conductance ($g_{\text{sc}}/g_{\text{m}}$), leaf carbon-use- efficiency (CUE_L).
10 Data are mean \pm SE ($n=6$); significant treatment effects ($P<0.05$) tested with two-way ANOVAs are
11 shown in bold.

	<i>T. aestivum</i>		<i>H. annuus</i>		Significance		
	aCO ₂	eCO ₂	aCO ₂	eCO ₂	<i>spe</i>	CO ₂	<i>spe</i> ×CO ₂
A_{growth}	28.42±0.73	32.79±2.01	27.27±2.57	31.43±2.06	0.529	0.042	0.956
A_{max}	37.68±1.29	34.04±2.22	32.32±2.40	31.83±1.73	0.068	0.304	0.430
R_{Dk}	2.26±0.14	1.81±0.17	1.48±0.11	1.32±0.14	<0.001	0.046	0.327
$i\text{WUE}_{\text{growth}}$	52.64±1.30	62.25±3.94	37.30±10.25	102.14±24.99	0.380	0.013	0.057
SLA	0.24±0.01	0.27±0.02	0.22±0.02	0.23±0.02	0.038	0.252	0.511
N%	6.61±0.17	6.39±0.11	3.51±0.67	3.16±0.56	<0.001	0.527	0.882
N _{area}	2.70±0.06	2.37±0.10	1.53±0.18	1.35±0.16	<0.001	0.062	0.586
Chl	0.69±0.02	0.56±0.03	0.49±0.05	0.48±0.02	<0.001	0.04	0.083
V_{cmax}	160.9±3.2	142.7±13.0	123.0±11.7	117.4±3.3	0.002	0.203	0.493
J	183.4±3.3	160.1±9.3	164.2±10.4	158.9±8.6	0.234	0.101	0.293
$g_{\text{sc}}/g_{\text{m}}$	0.95±0.03	1.16±0.07	1.74±0.55	1.97±0.42	0.087	0.222	0.859
CUE _L	0.89±0.01	0.92±0.01	0.92±0.02	0.94±0.01	0.011	0.004	0.547

12

13

1 Table 2. ANOVA tests for R_L estimated by the Kok, Kok-Phi and Kok- C_c methods. Significant
2 treatment effects ($P < 0.05$) are shown in bold.

Source	df	$R_{L\text{ Kok}}$		$R_{L\text{ Kok-Phi}}$		$R_{L\text{ Kok-Cc}}$	
		F	P	F	P	F	P
Species	1	28.36	<0.001	29.74	<0.001	20.06	<0.001
Growth CO ₂	1	6.681	0.013	5.777	0.021	3.921	0.055
Measurement CO ₂	1	1.227	0.275	1.714	0.198	0.198	0.658
Growth CO ₂ *	1	0.092	0.763	0.107	0.745	0.072	0.790
Measurement CO ₂	1						

3
4 Table 3. ANOVA tests for R_L/R_{Dk} estimated by the Kok, Kok-Phi and Kok- C_c methods.
5 Significant treatment effects ($P < 0.05$) are shown in bold.

Source	df	$R_{L\text{ Kok}}/R_{Dk}$		$R_{L\text{ Kok-Phi}}/R_{Dk}$		$R_{L\text{ Kok-Cc}}/R_{Dk}$	
		F	P	F	P	F	P
Species	1	2.824	0.101	1.638	0.208	0.095	0.759
Growth CO ₂	1	2.216	0.144	0.799	0.377	0.389	0.536
Measurement CO ₂	1	7.480	0.009	10.410	0.003	1.140	0.292
Growth CO ₂ *	1	0.328	0.570	0.441	0.511	0.243	0.625
Measurement CO ₂	1						

6
7 **FIGURE LEGENDS**

8 **Figure 1.** Net CO₂ assimilation rate (A), transpiration rate (E) and stomatal conductance for
9 water vapor (g_{sw}) in response to short-term variation of intercellular CO₂ concentration (C_i) for
10 wheat (*T. aestivum*) and sunflower (*H. annuus*). Blue circles refer to ambient (410 $\mu\text{mol mol}^{-1}$)
11 growth CO₂, and red squares refer to elevated (820 $\mu\text{mol mol}^{-1}$) growth CO₂. Data are shown as
12 mean \pm SE ($n=6$).

13 **Figure 2.** Photochemical efficiency of photosystem II (Φ_2), chloroplastic CO₂ concentration
14 (C_c) and γ (the lumped parameter in Eqn 2) in response to incident irradiance (I_{inc}) for wheat (*T.*
15 *aestivum*) and sunflower (*H. annuus*). Plants grown under ambient CO₂ (aCO₂, blue circles) or
16 elevated CO₂ (eCO₂, red squares) were measured at gaseous conditions of 410 $\mu\text{mol mol}^{-1}$ (open
17 symbols) or 820 $\mu\text{mol mol}^{-1}$ (closed symbols) CO₂ in the leaf chamber. Data are shown as mean
18 \pm SE ($n=6$).

19 **Figure 3.** Net CO₂ assimilation rate (A) in response to I_{inc} (incident irradiance), $\Phi_2 I_{inc}$ (Φ_2 ,
20 photochemical efficiency of photosystem II) or $\gamma \Phi_2 I_{inc}$ (γ , the lumped parameter in Eqn 2) for
21 wheat (*T. aestivum*) and sunflower (*H. annuus*). Data are mean \pm SE ($n=6$). Meaning of symbols
22 of different CO₂ treatments and measurement conditions are shown in Fig. 2.

23 **Figure 4.** Effects of growth CO₂ treatments (aCO₂ and eCO₂) and measurement conditions (410
24 and 820 ppm CO₂) on respiration rates in the light (R_L) estimated by three methods for wheat (*T.*

1 *aestivum*) and sunflower (*H. annuus*). R_L was measured by the Kok (A and D), Kok-Phi (B and
2 E) and Kok- C_c (C and F) methods. Data are mean \pm SE ($n=5-6$). The results of ANOVA tests are
3 shown in Table 2.

4 **Figure 5.** Effects of growth CO_2 treatments (a CO_2 and e CO_2) and measurement conditions (410
5 and 820 ppm) on ratio of respiration in the light to respiration in the dark (R_L/R_{Dk}) for wheat (*T.*
6 *aestivum*) and sunflower (*H. annuus*). R_L/R_{Dk} was estimated by the Kok (A and D), Kok-Phi (B
7 and E) and Kok- C_c (C and F) methods. Data are mean \pm SE ($n=5-6$). The results of ANOVA
8 tests are shown in Table 3.

9 **Figure 6.** Correlation between respiration in the dark (R_{Dk}) and respiration in the light (R_L). R_L
10 was measured by the Kok (A), Kok-Phi (B) and Kok- C_c (C) methods. The average R_L/R_{Dk} (\pm SE,
11 $n=45$) was calculated by pooling over the data of species (wheat and sunflower) and CO_2
12 treatments. Grey dashed lines give the 1:1 relationship.

13

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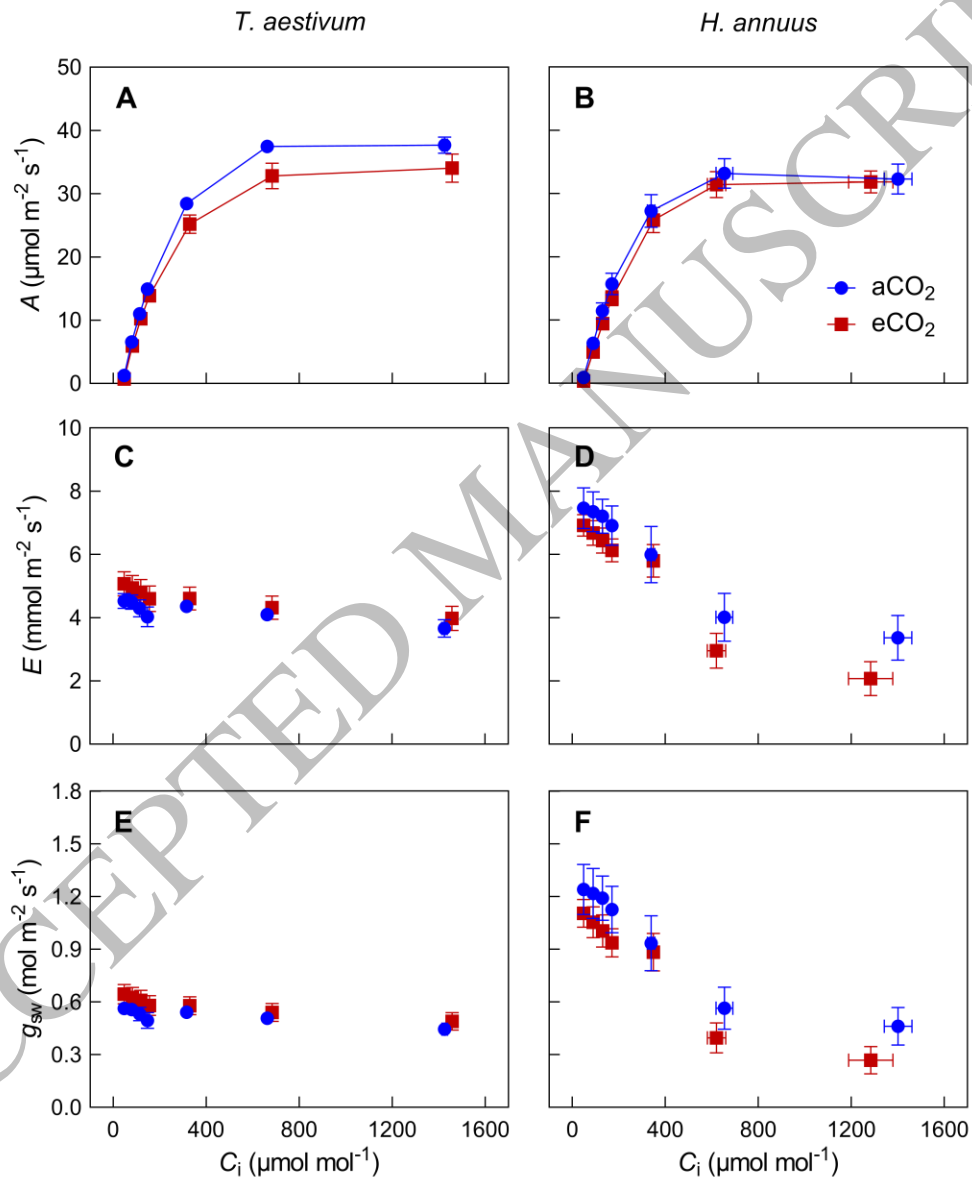
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7
 8 **Figure 1**
 9 **159x182 mm (9.1 x DPI)**
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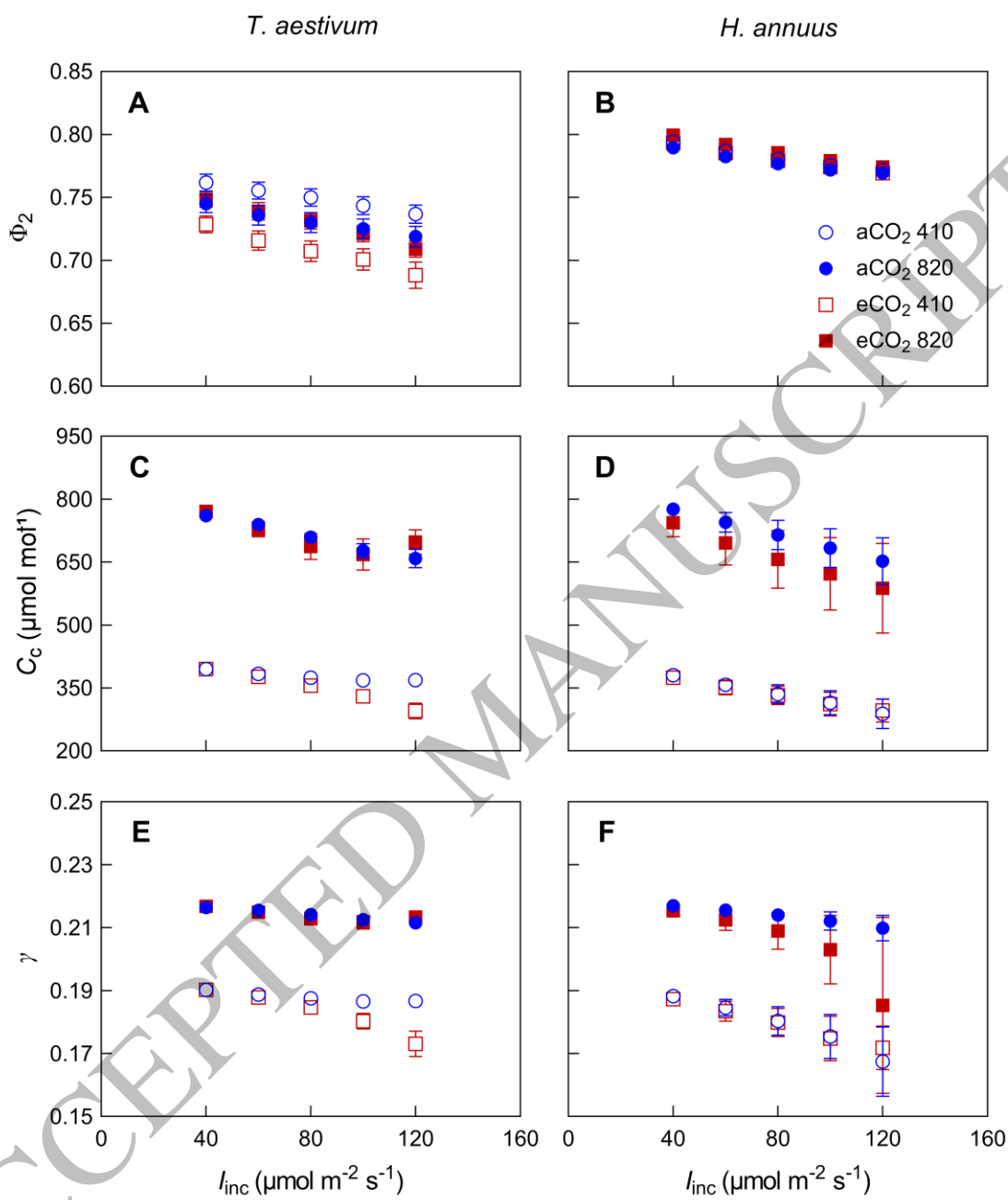


Figure 2
159x181 mm (9.1 x DPI)

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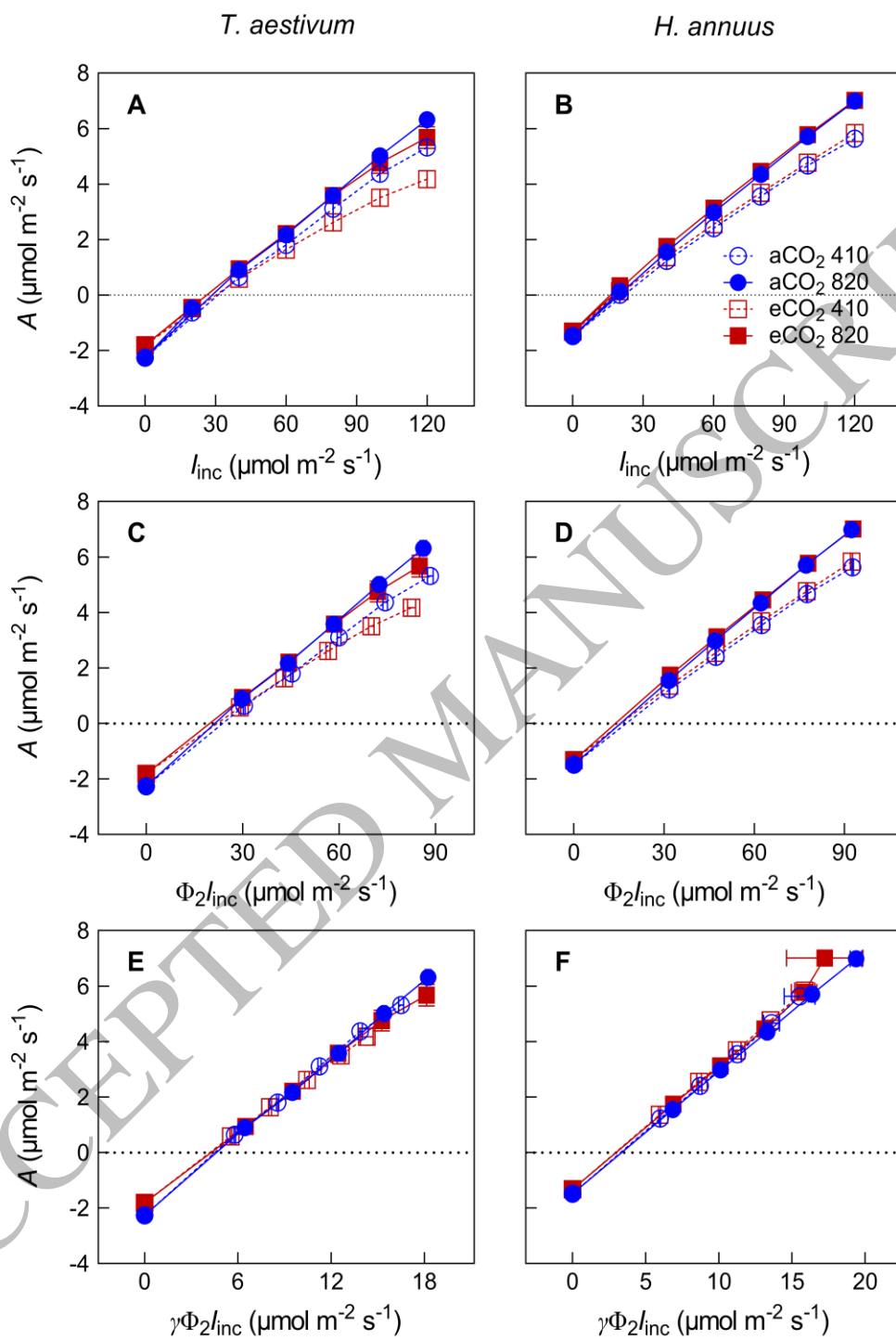


Figure 3
159x212 mm (9.1 x DPI)

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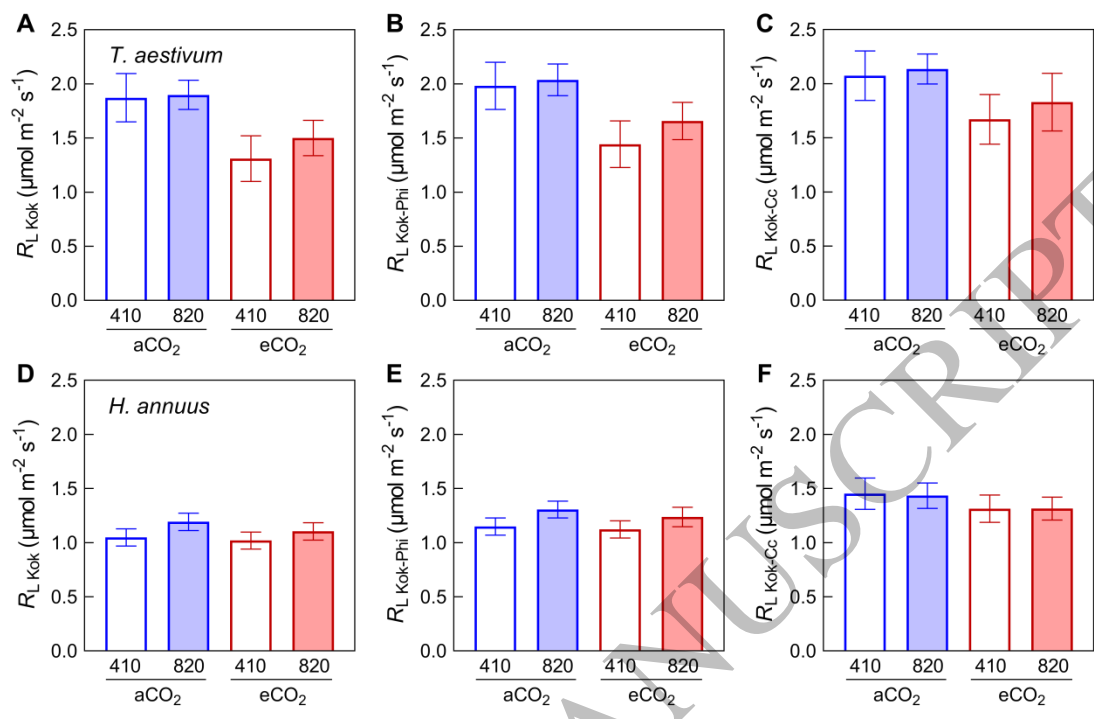


Figure 4
153x100 mm (9.1 x DPI)

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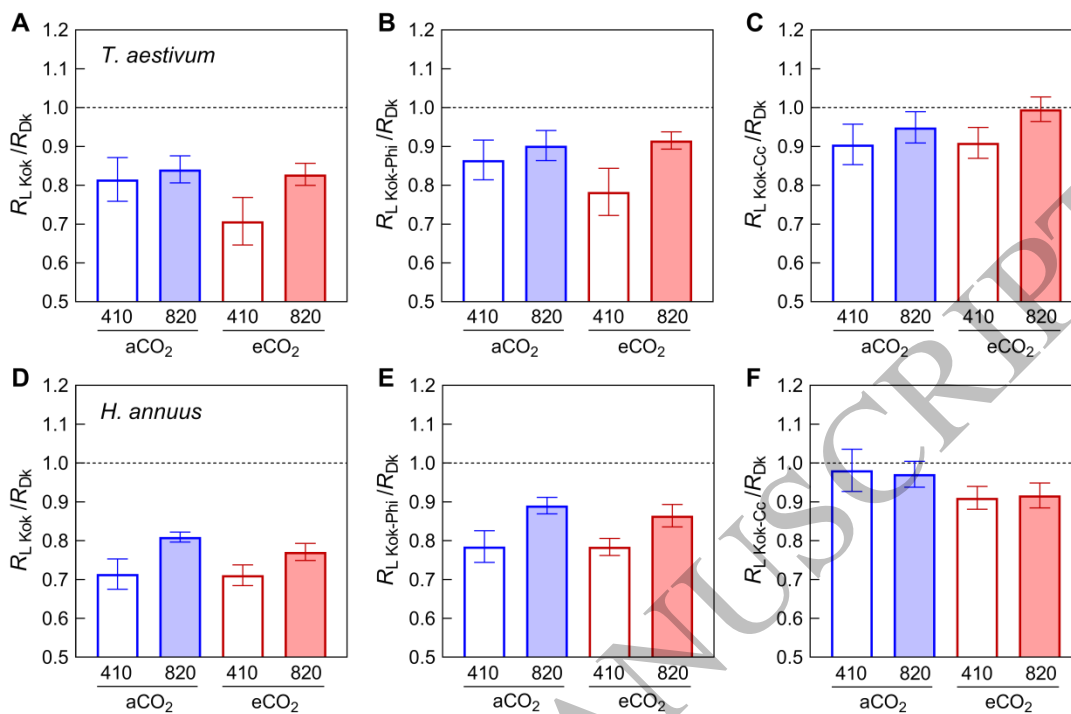


Figure 5
152x101 mm (9.1 x DPI)

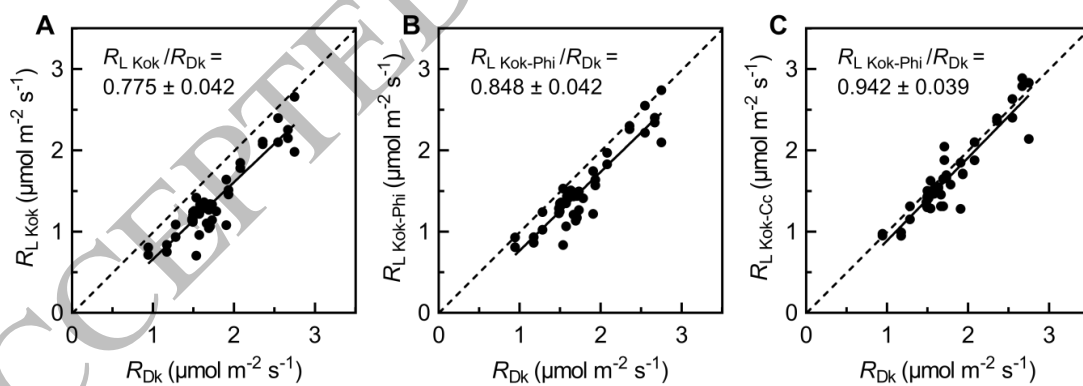


Figure 6
159x59 mm (9.1 x DPI)

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