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➤ Nanopore Adaptive sampling: A powerful approach to decipher the genetic diversity of the NLRome/resistome in melon

Javier BELINCHON-MORENO

Nanopore Day, Paris, 21st March 2023

➤ EPGV – INRAE: plant genomic platform located at Genoscope, Evry - France



support-EPGV@inrae.fr



-Experiment reporting
-Data archiving

Bioanalyses

High quality
data
LIMS

WetLab
expertise

Expertise and
consulting in
Genomics

- Genome assembly & annotations
- Variants detection



- Variants calling (SNPs/SVs)

Genotypes matrix
Bam / vcf / fasta
Genome sequence



EPGV
Etude du Polymorphisme
des Génomes Végétaux

INRAE

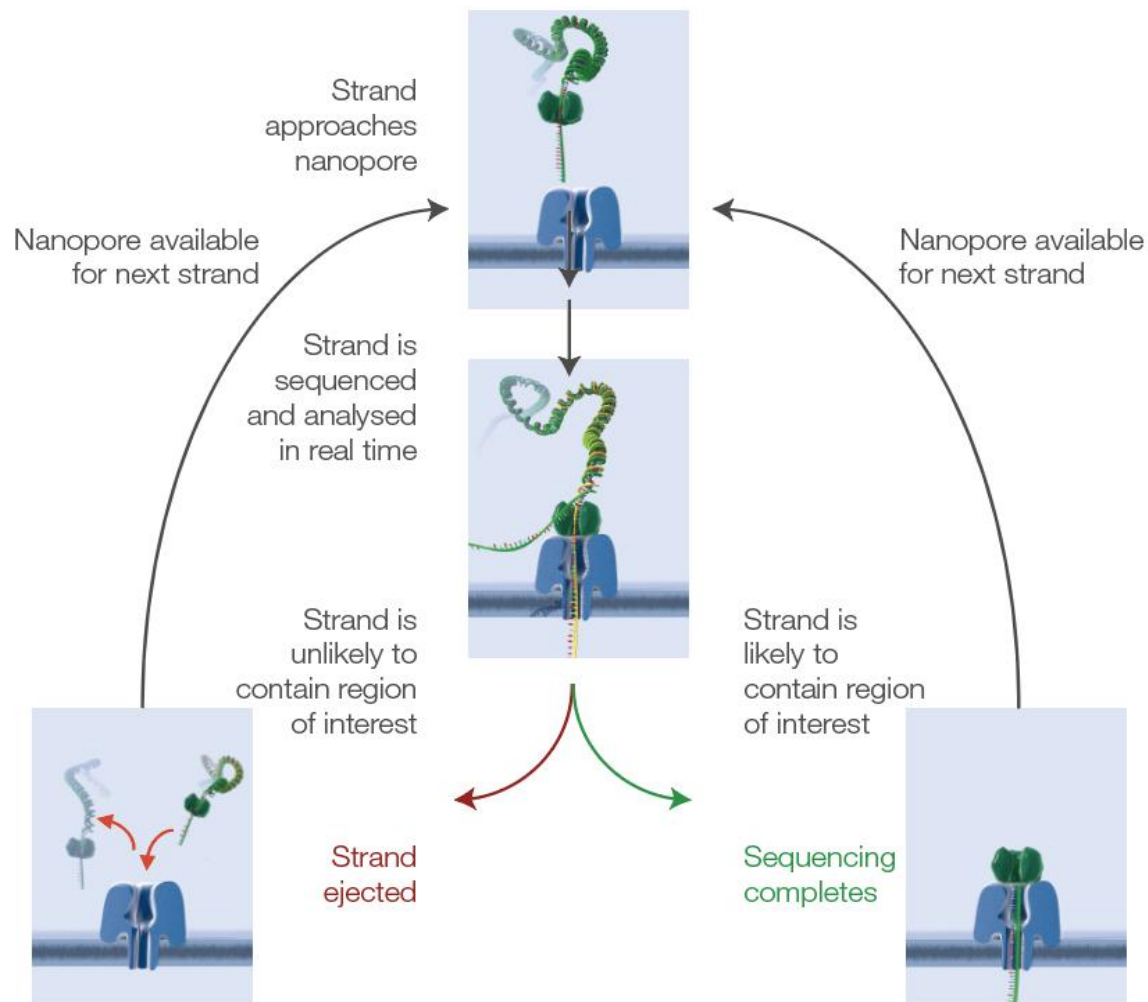
**INRAE
GENOMICS**

INRAE

21 March 2023 / Javier BELINCHON-MORENO

➤ What is Nanopore adaptive sampling?

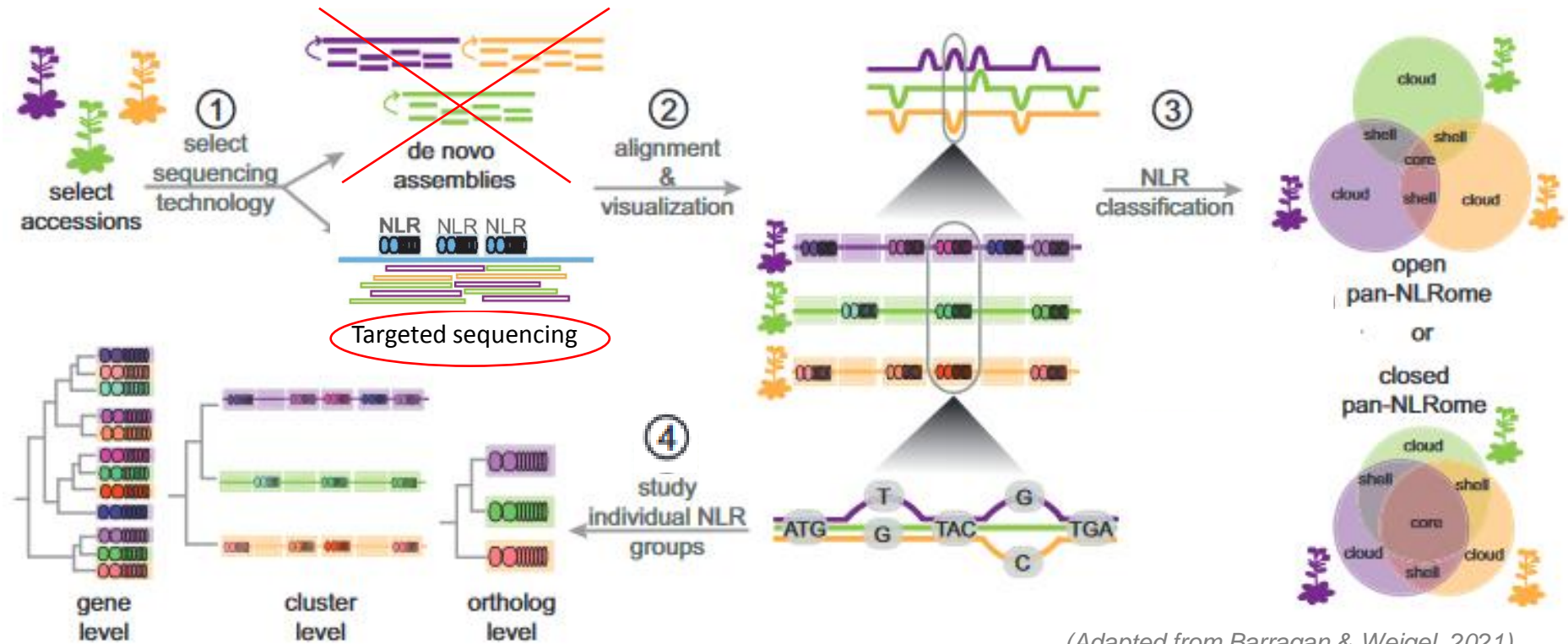
- Accept/reject molecules based on **small initial part** of sequence (~500 bp)
- **Enrich/deplete** specific regions of interest.
- Increase on-target data, reduce time-to-answer.
- **Real time** target selection: **easy** library preparation.



➤ Our objective: Nanopore adaptive sampling to decipher the genetic diversity of the NLRome in melon

Reference genome not enough → Construction of **pan-NLRome**

High number of varieties (~150 varieties) → Need for **selective sequencing**



(Adapted from Barragan & Weigel, 2021)

➤ NLR resistance genes: a conserved structure

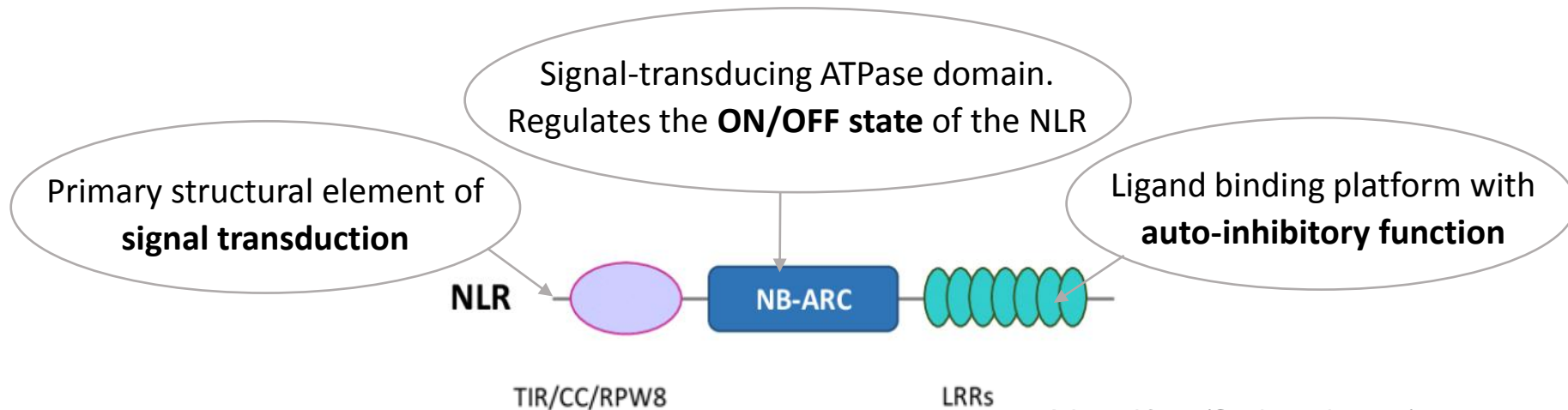
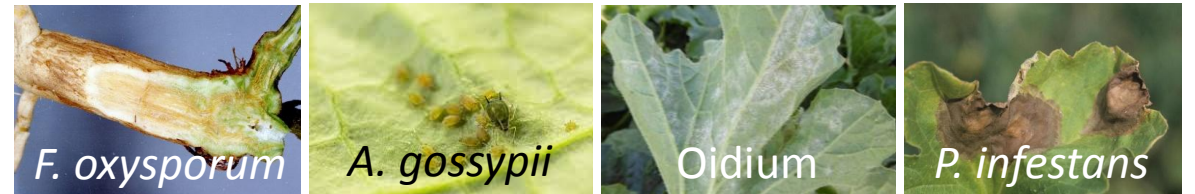
Nucleotide binding site (NB) -leucine rich repeat (LRR)



Intracellular immune receptor proteins

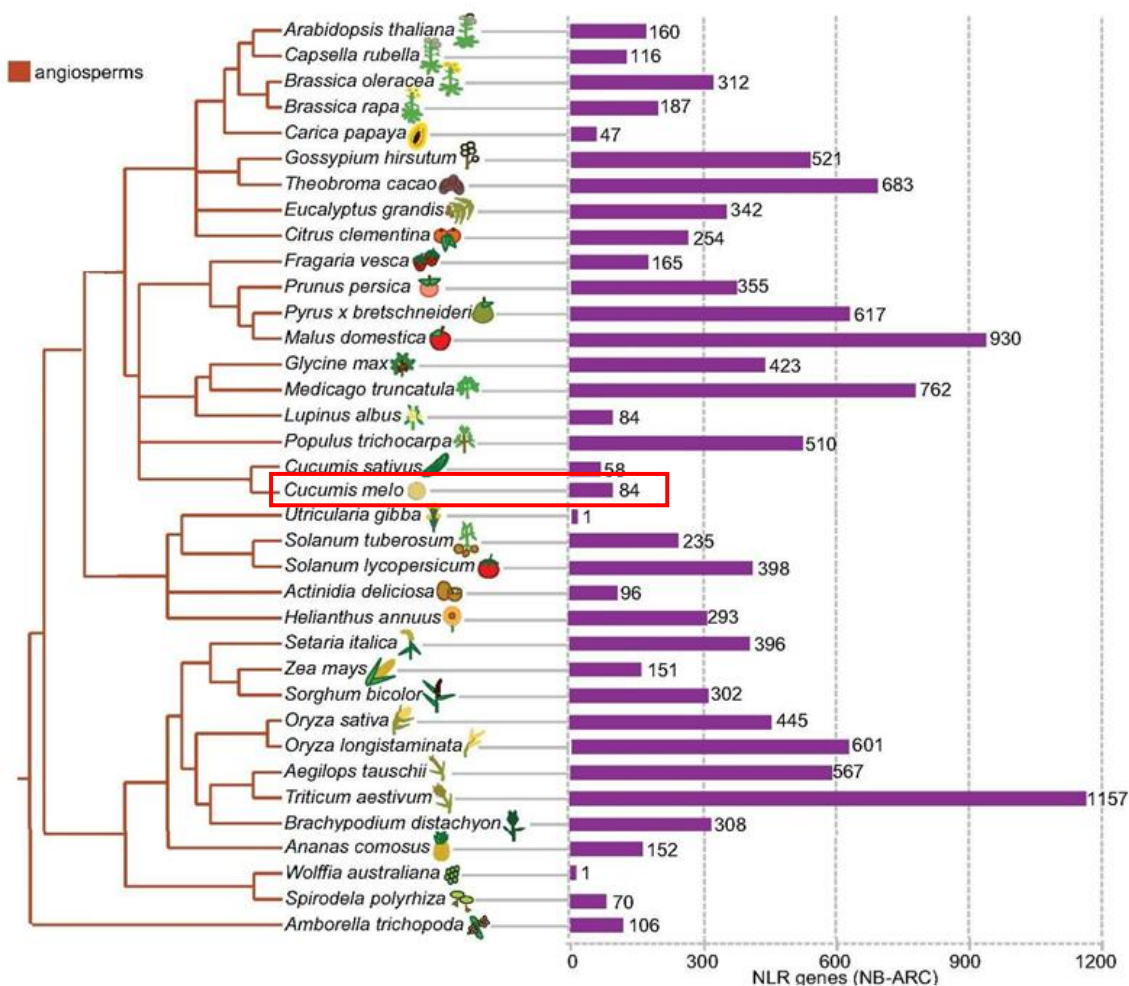
Major family of plant-resistance (R) genes

Broad range of **resistance**



Adapted from (Gottin et al., 2021)

➤ Variable number of NLR genes between plant species



Usually grouped into **clusters**

High level of presence/absence polymorphisms (PAV)

Low frequency in the Cucurbitaceae family



C. melo variety Anso77

84 NLR genes in 15 clusters

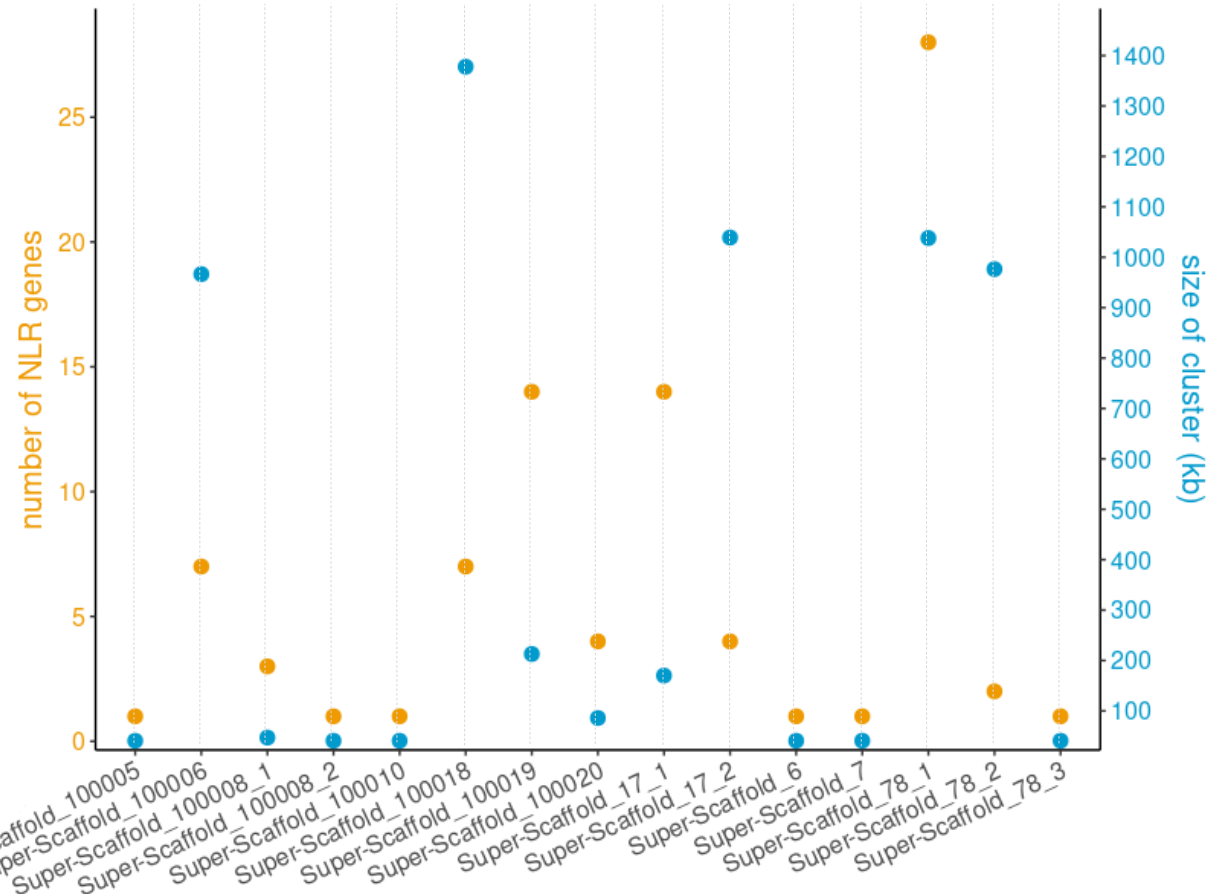
NLRGenomeSweeper (Toda et al., 2020)

Adapted from (Barragan & Weigel, 2021)



➤ Our target regions: 15 NLR-gene clusters

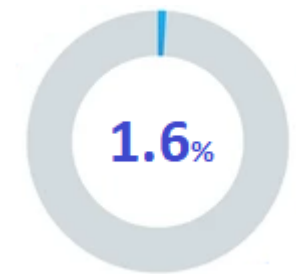
Reference genome → Variety **Anso77** (draft genome produced at INRAE-GAFL/EPGV)



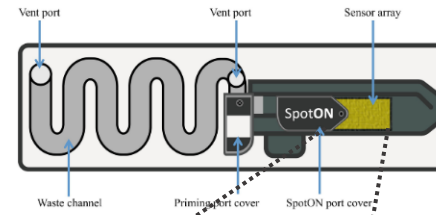
Reference genome (.fasta)
+
Target regions (.bed)

Clusters:
NLR genes + 20kb-extra flanking
Variable in size
Variable in number of genes

Genome length ~380Mb
Target regions length ~6.16Mb



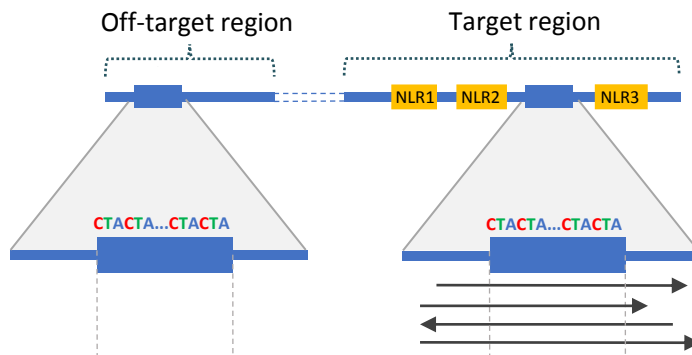
➤ Our first experience: Variety Anso-77 (same as provided reference)



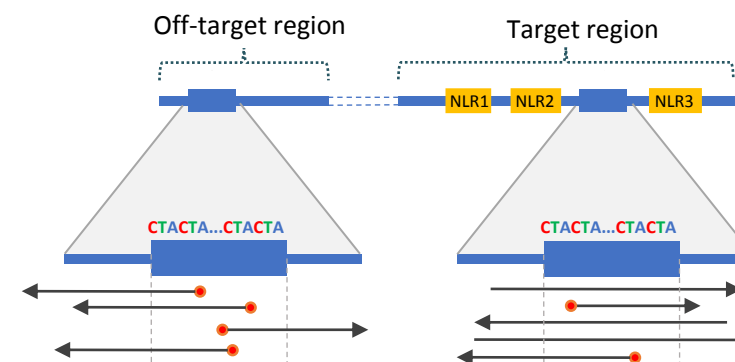
NAS

Standard sequencing

Masking repetitive elements >200kb



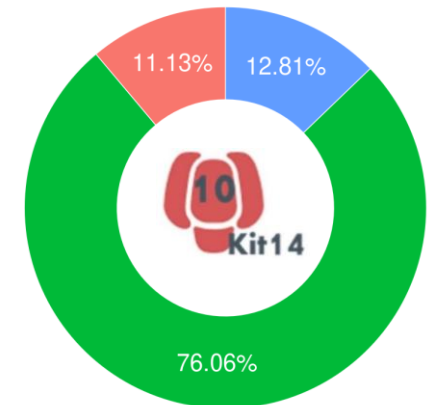
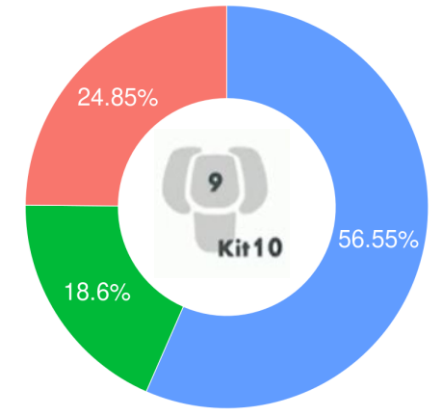
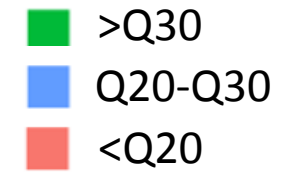
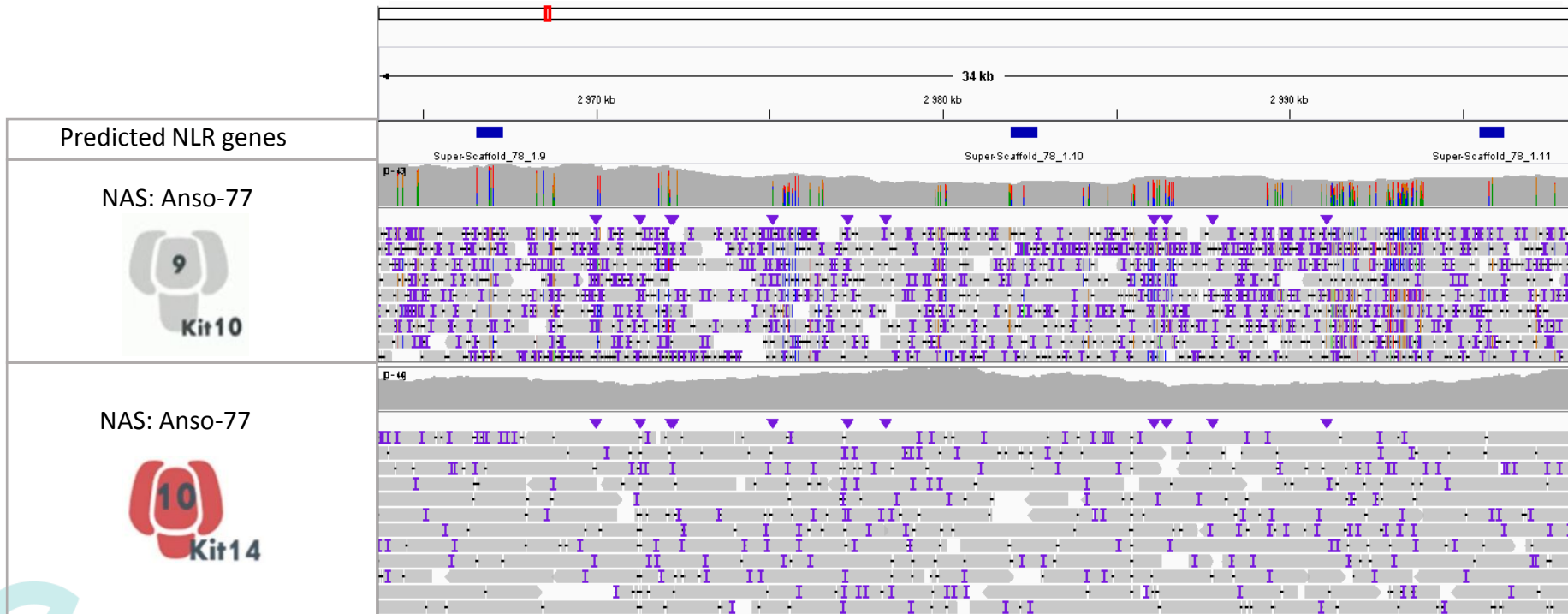
Not masking repetitive elements



➤ Nanopore kit evolution: From kit10 to kit14

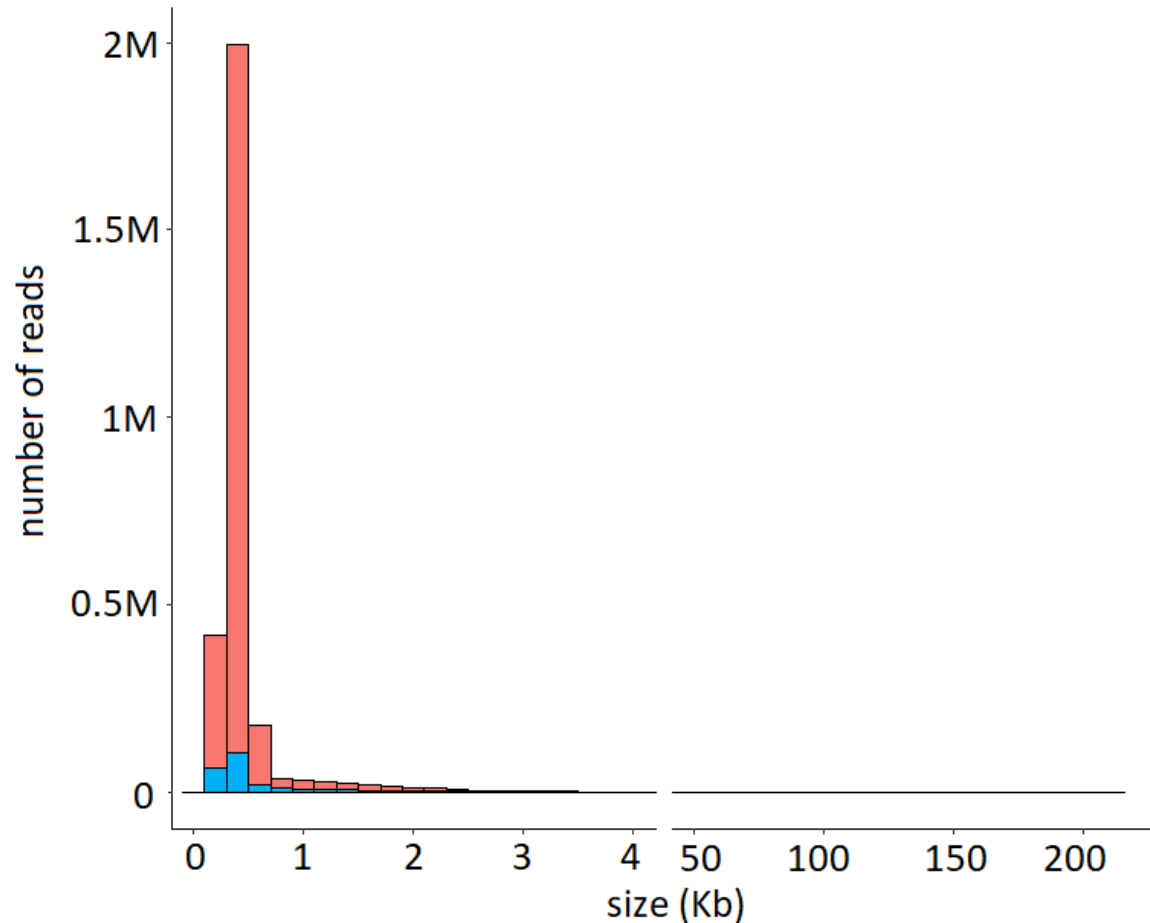


- **Increased accuracy**, improved sequencing **speed**
- **Reactive consumption reduced**, adaptors capture improved
- Duplex sequencing, **better throughput**

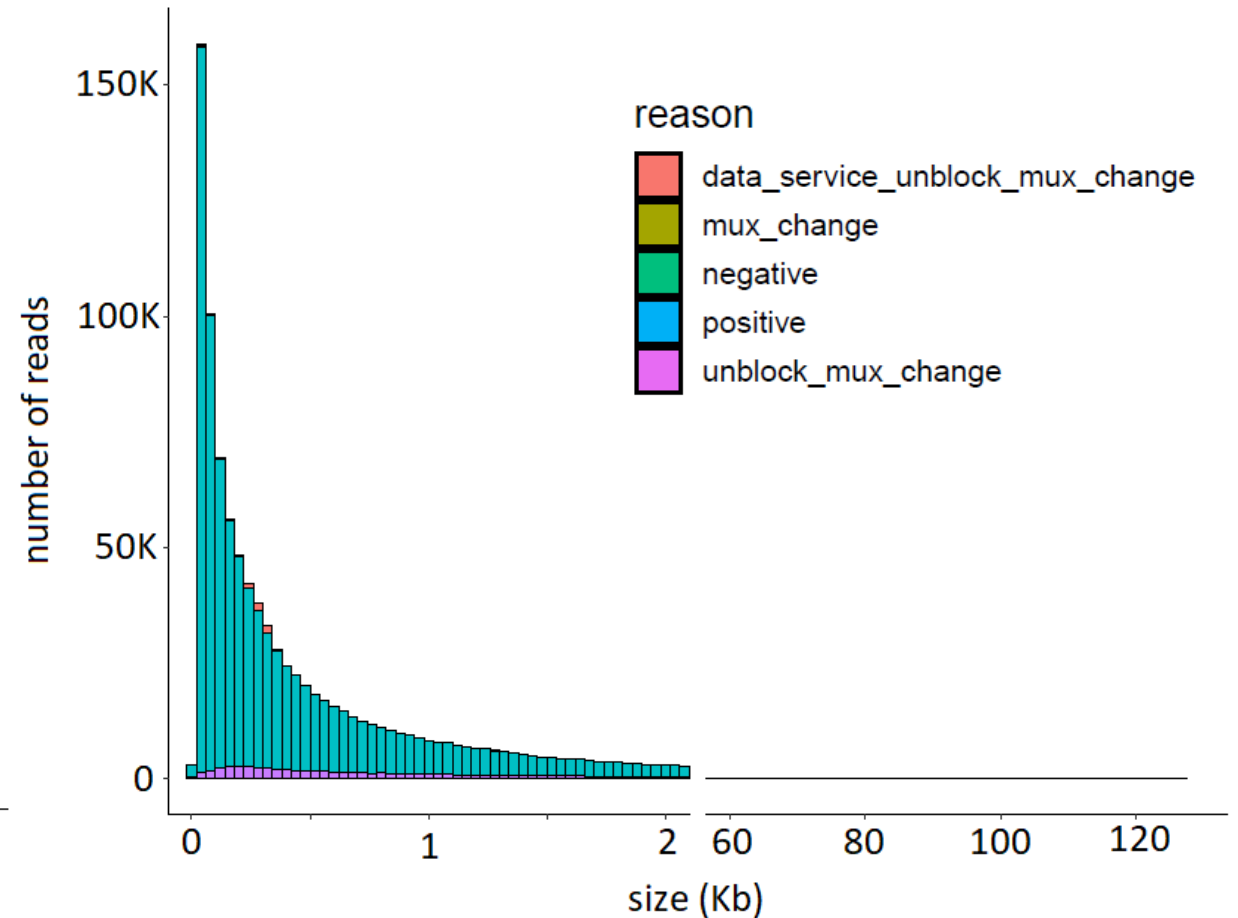


➤ NAS: A characteristic distribution of read lengths

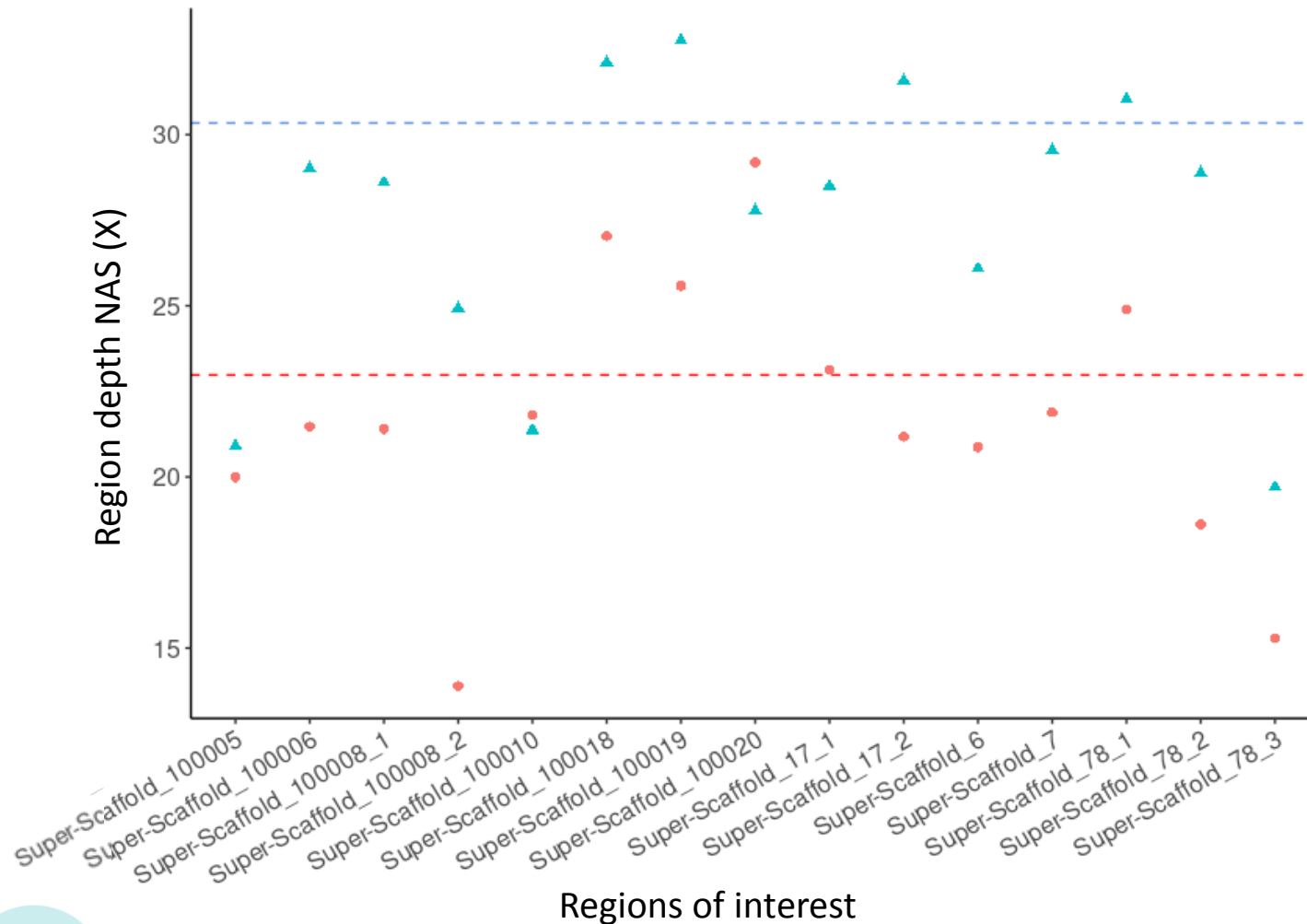
Nanopore adaptive sampling



Standard sequencing



➤ Statistics: regions depth

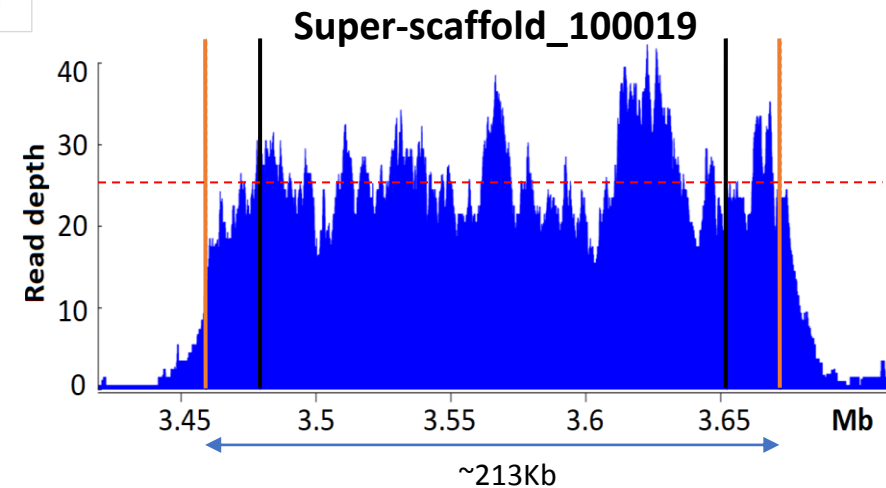
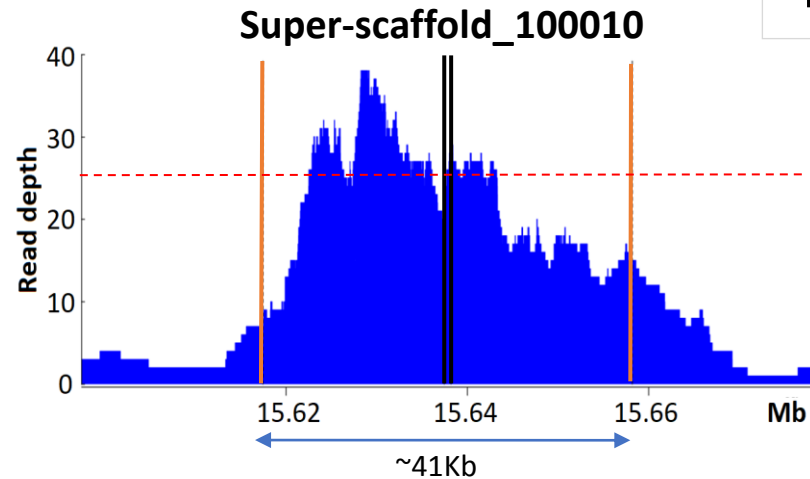
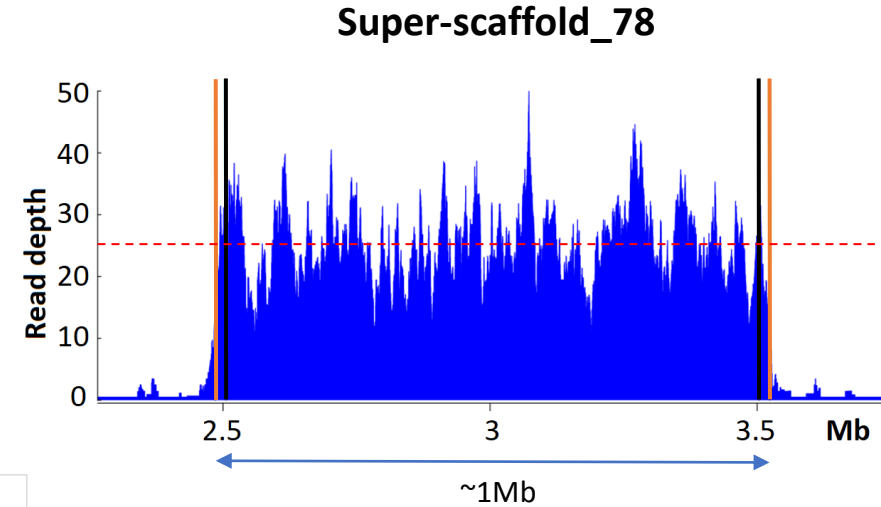
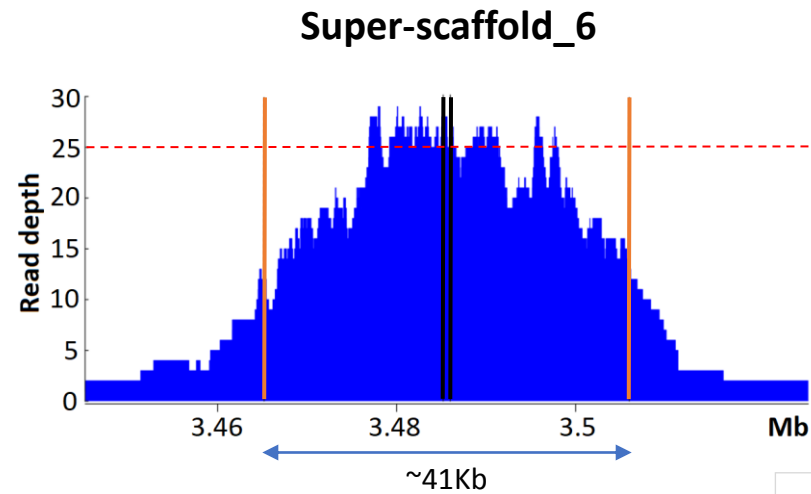


Variable between clusters

Consistent between experiences

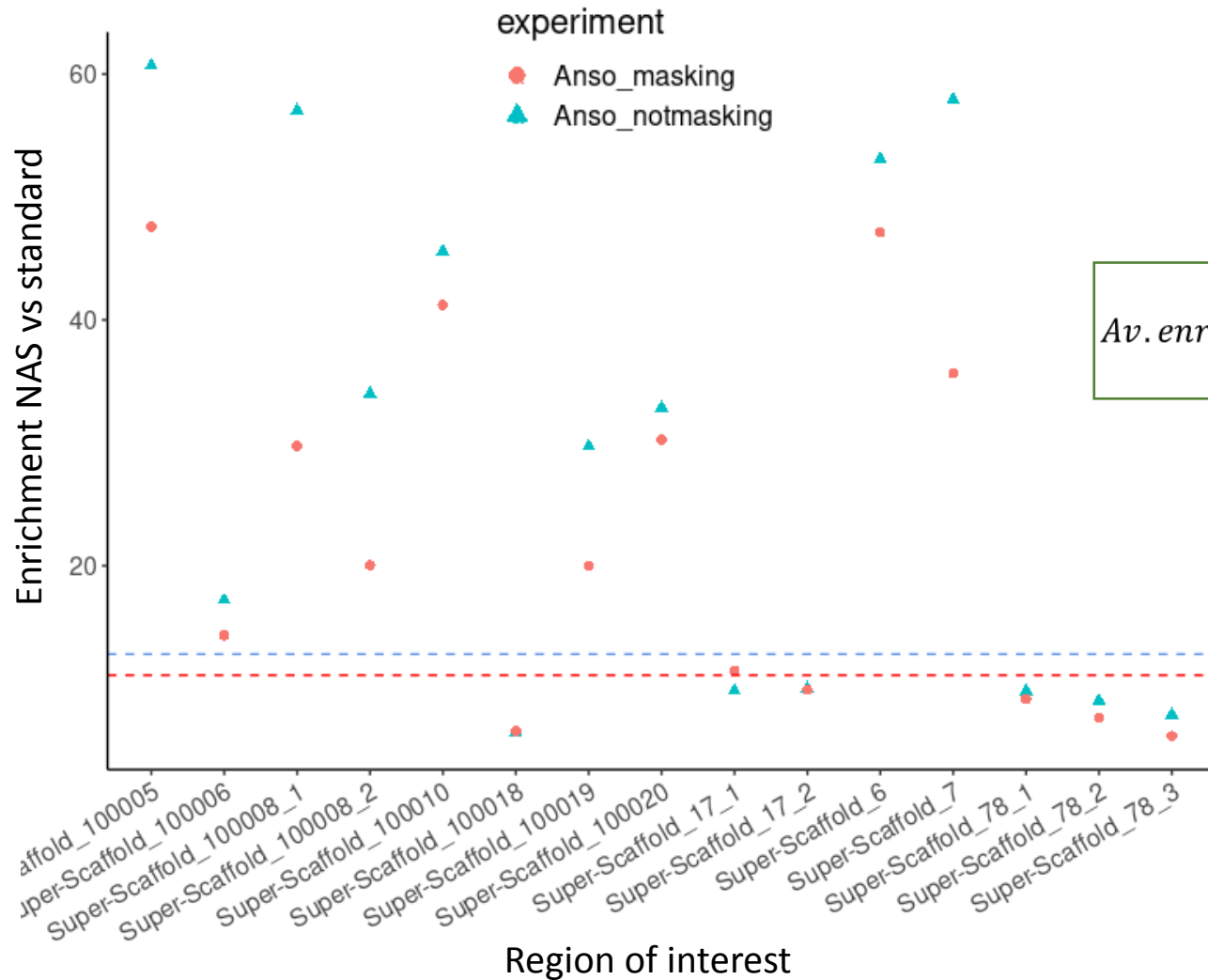
Mostly higher not masking

➤ Visual enrichment of our target regions



— ROI $\pm 20\text{kb}$
— ROI delimiter

➤ Good enrichment of our target regions



$$Enrichment_{region} = \frac{\frac{region_cov_NAS}{chrom_cov_NAS}}{\frac{region_cov_standard}{chrom_cov_standard}}$$

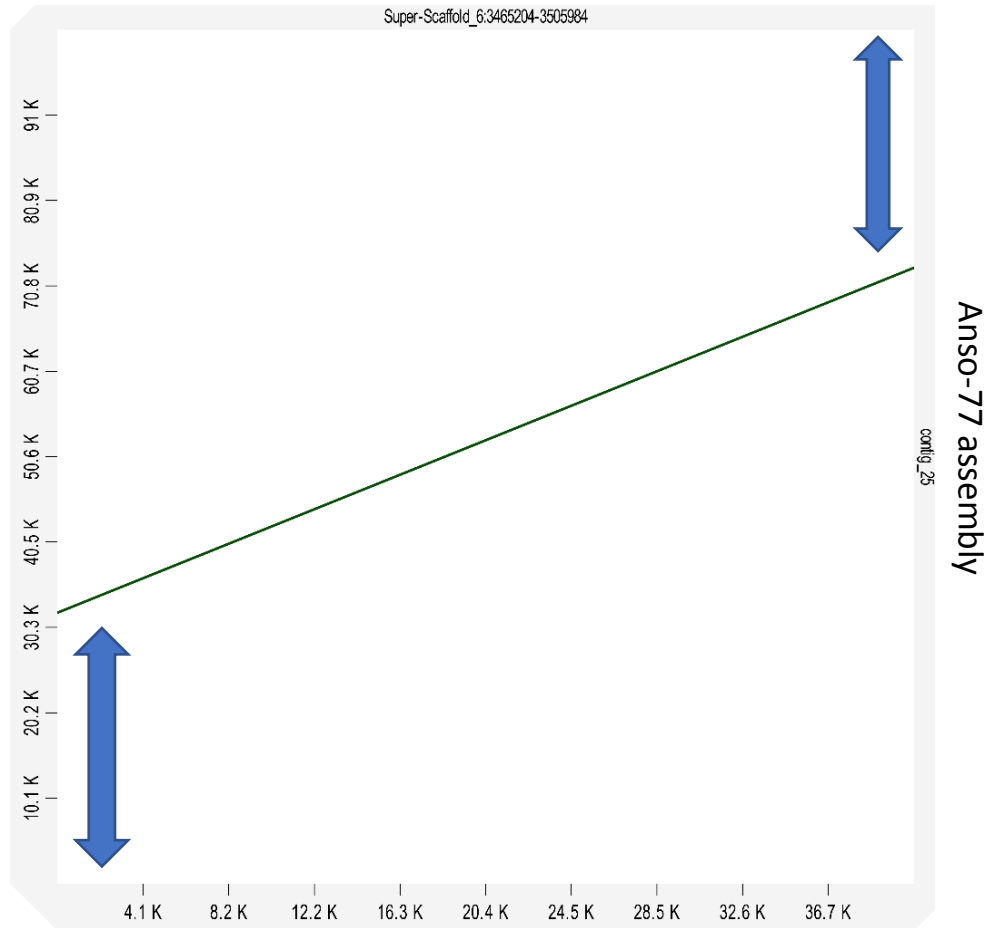
$$Av.enrichment = \sum_{region=1}^N \frac{enrichment_{region}(X) * region_size(bp)}{total_size_regions(bp)}$$

- Variable between clusters
- Variable between experiences
- But consistent between experiences for clusters under 20X
- Average ~equal between experiences

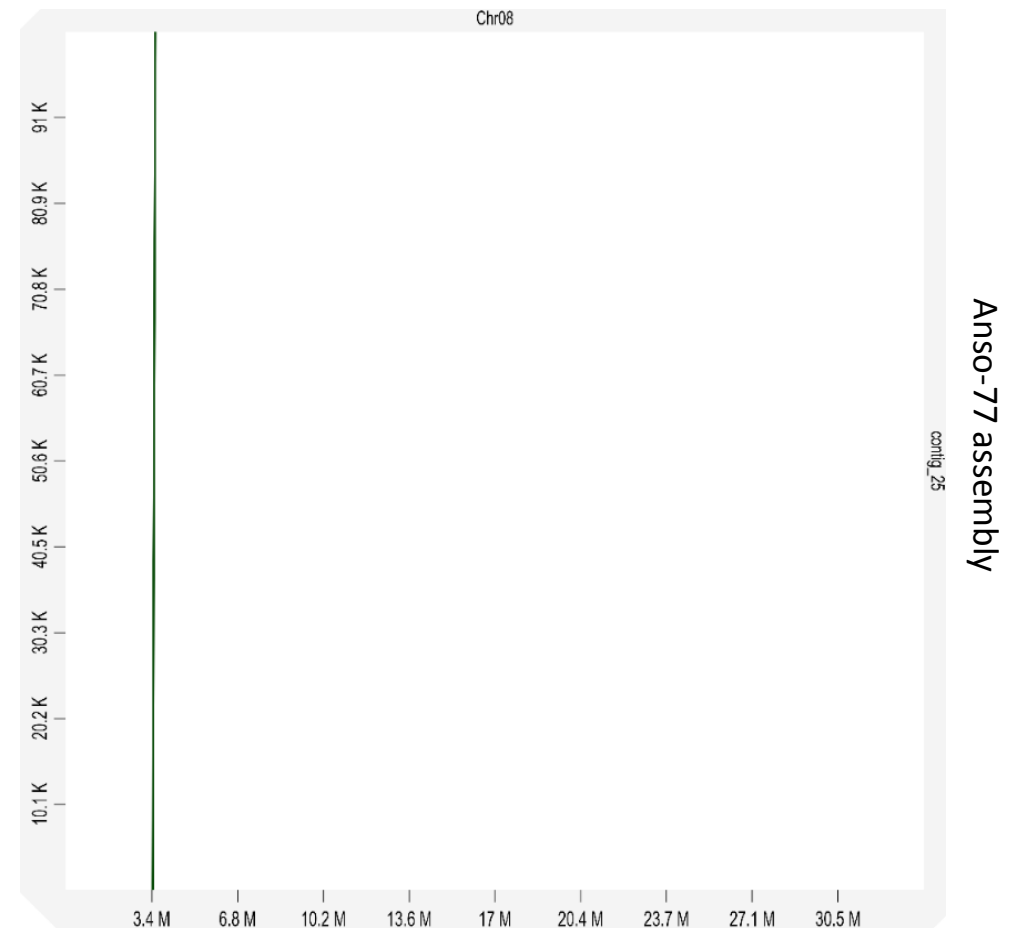


➤ Genome assemblies of target regions: statistics

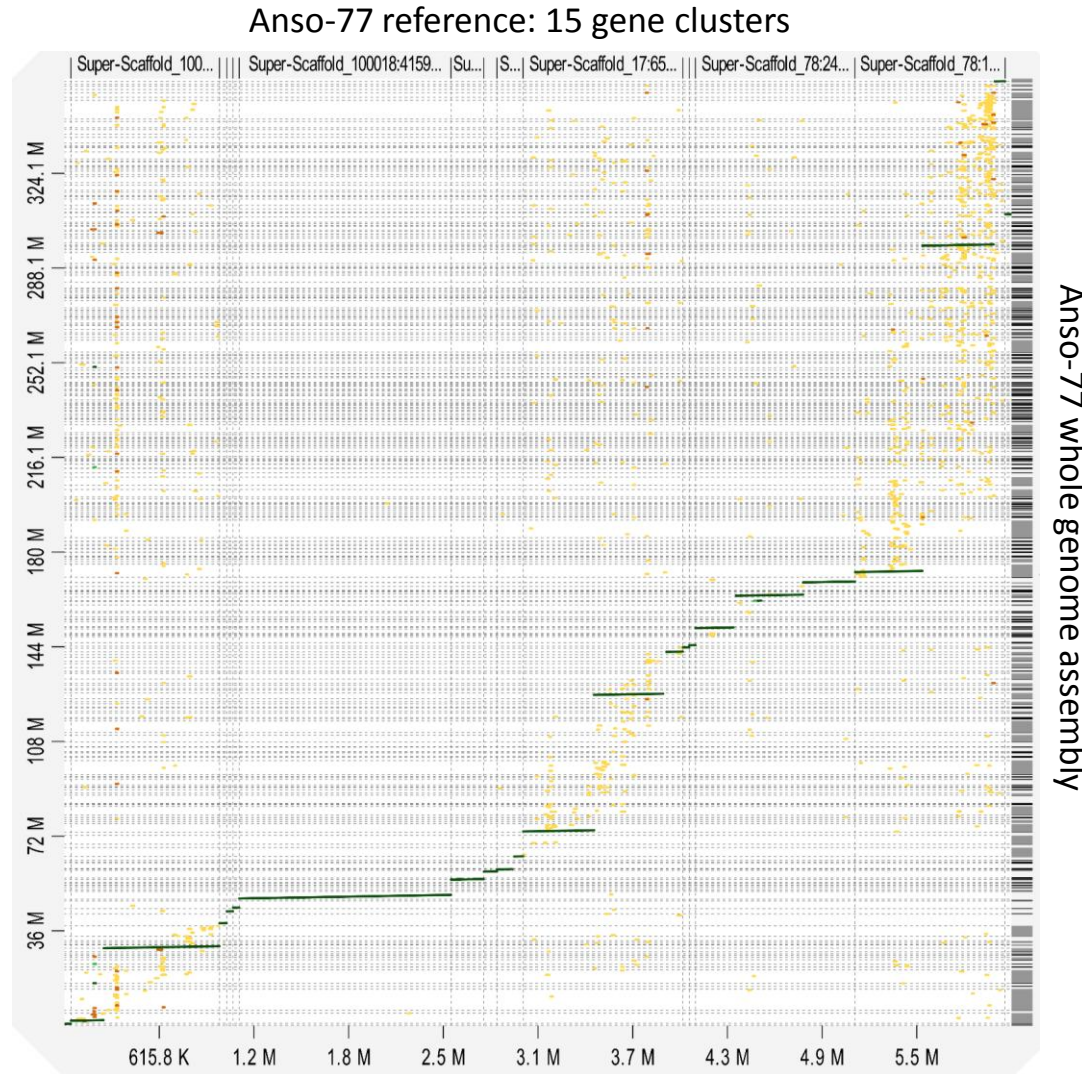
Anso-77 reference: one gene cluster



Anso-77 reference: complete chromosome



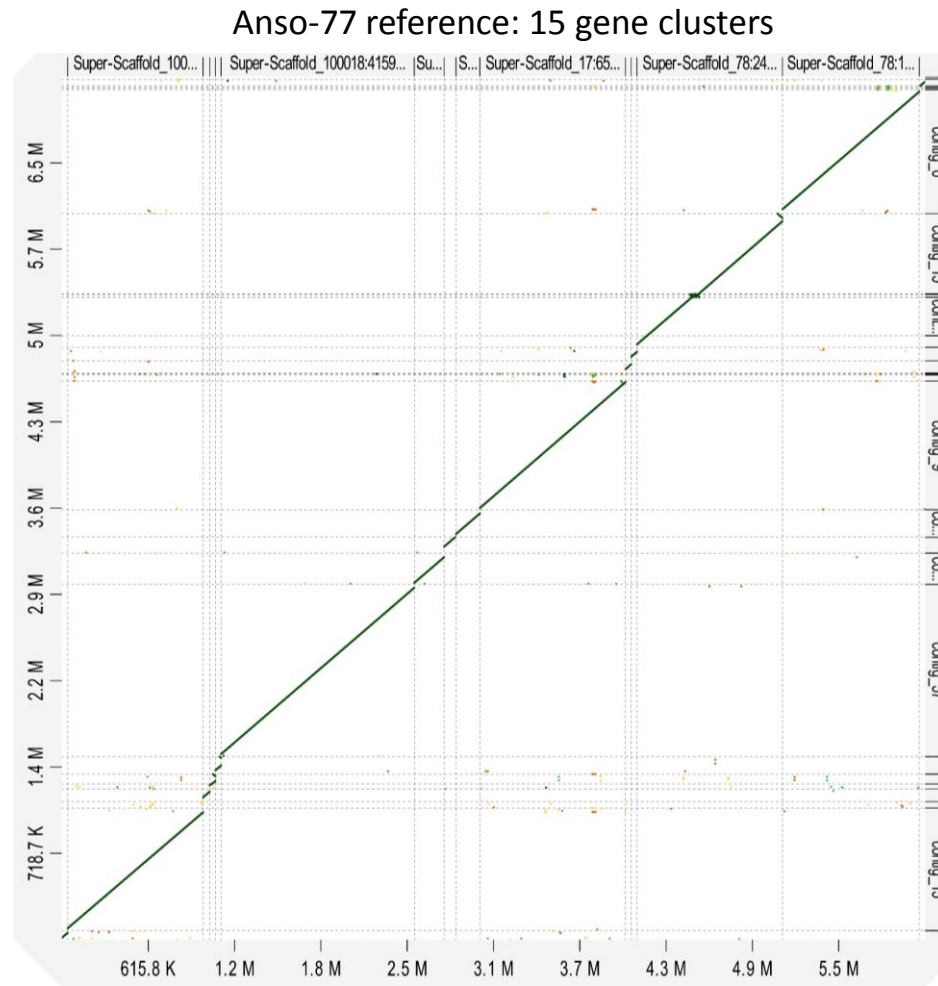
➤ Genome assemblies: Good assembly of target regions



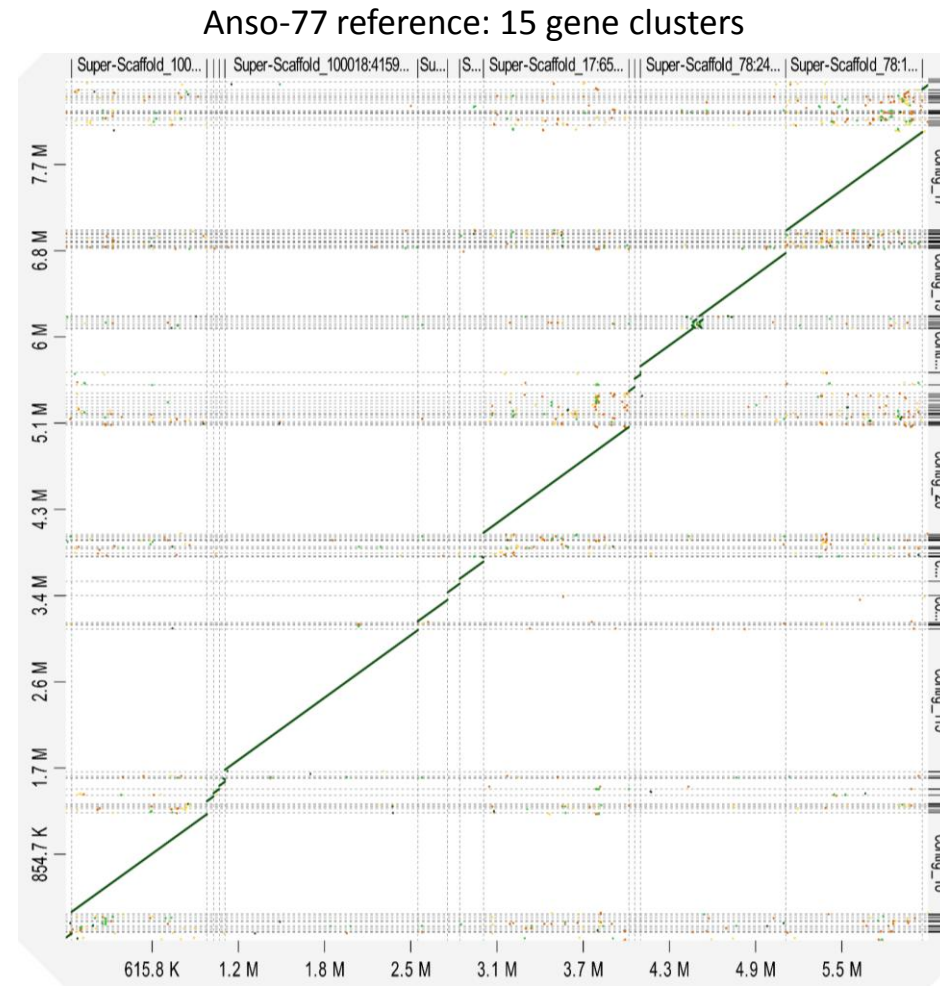
Assembly using WGS data:

- Target regions **fragmented** into multiple contigs
- Target regions **poorly covered** compared to adaptive sampling

➤ Genome assemblies: Good assembly of target regions



Anso-77 assembly **not** masking



Anso-77 assembly **masking**

Target regions fully assembled with a good depth



➤ Genome assemblies: cluster Vat

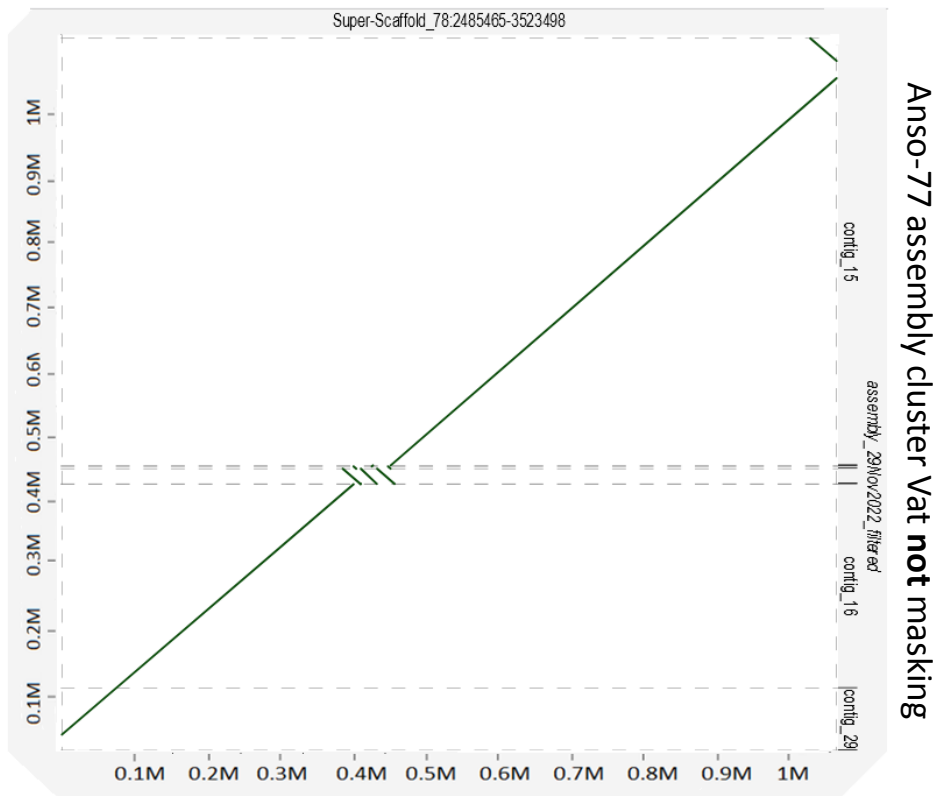
Cluster Vat: Most complex cluster of NLR genes in melon

28 NLR genes

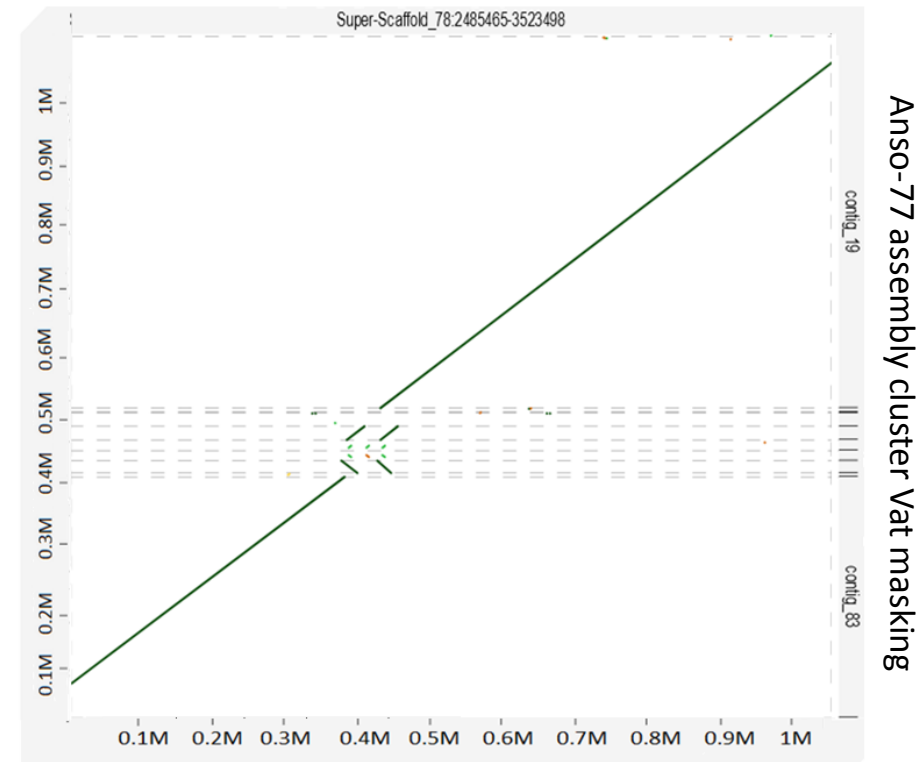
1Mb length

But also the most studied one (INRAE-GAFL)

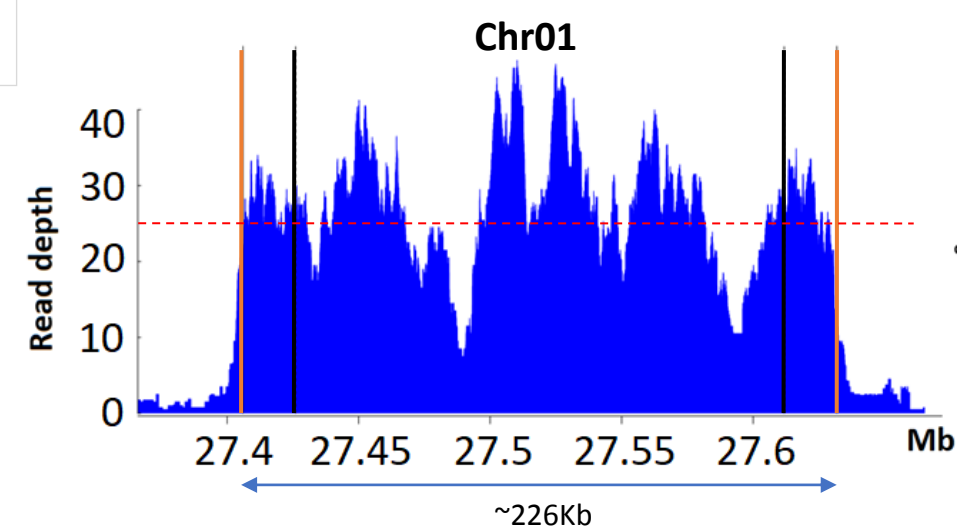
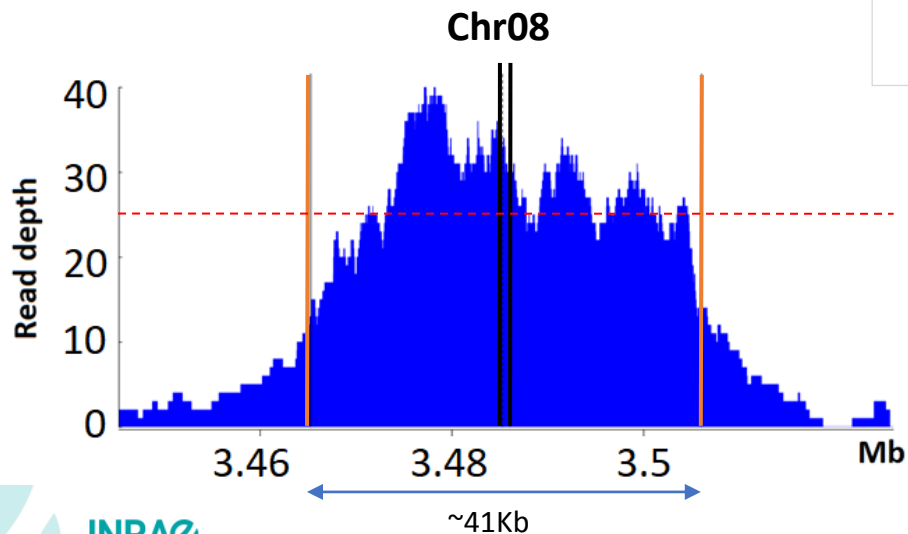
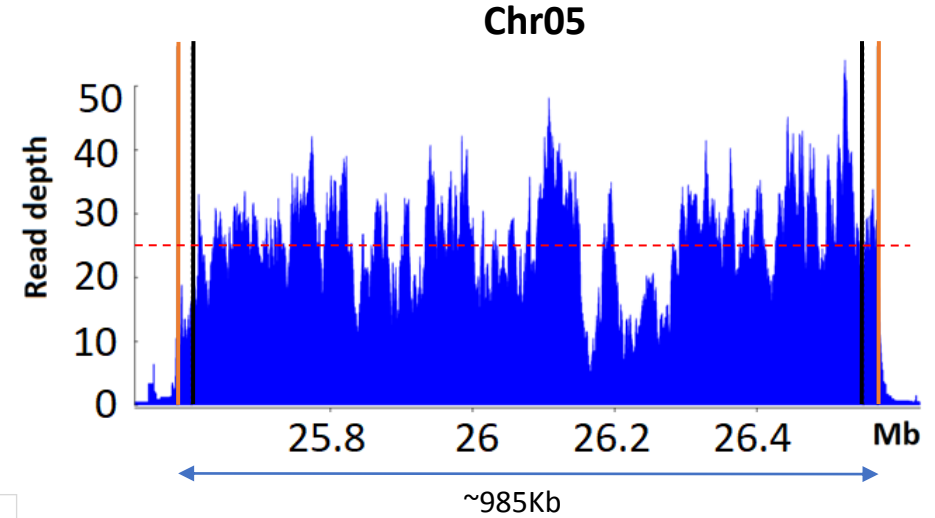
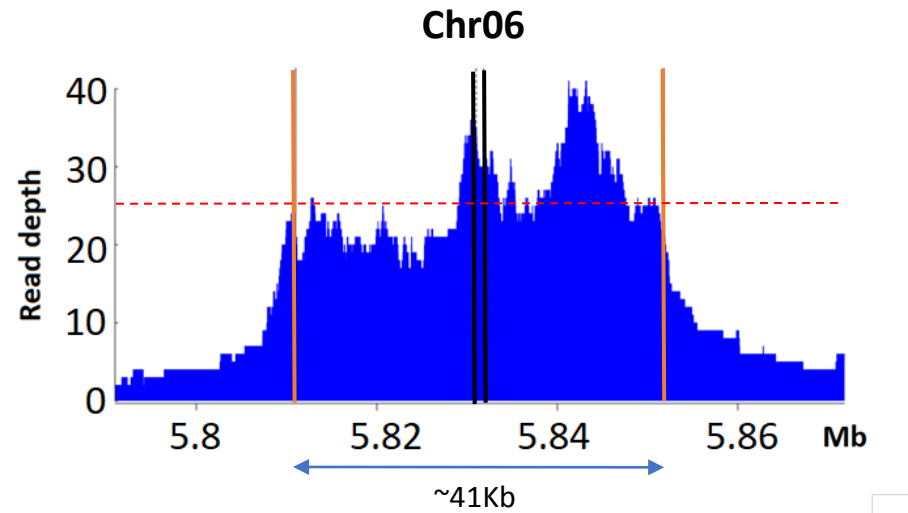
Anso-77 reference: cluster Vat



Anso-77 reference: cluster Vat



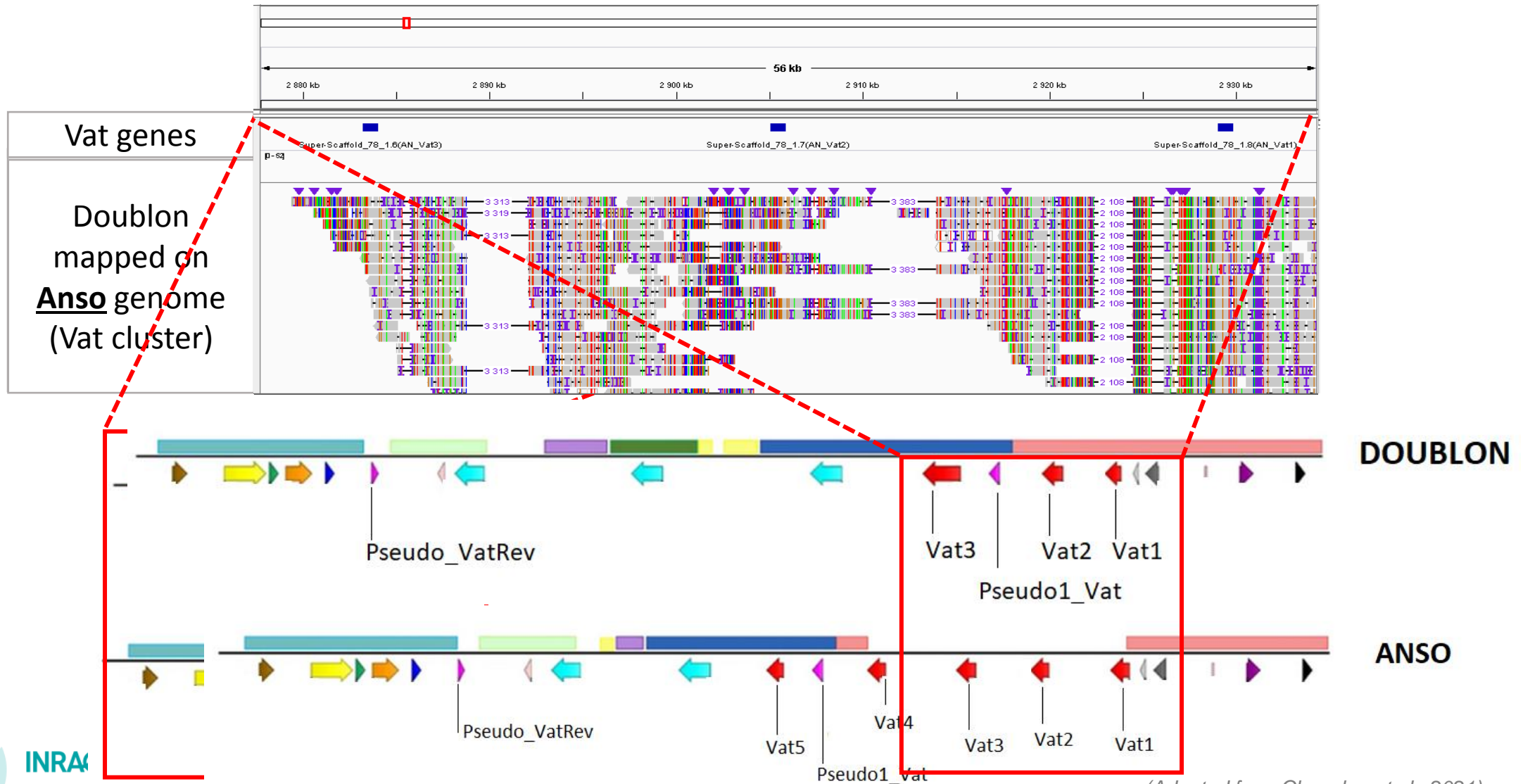
➤ Preliminary results using a different variety: Doublon



— ROI ±20kb
— ROI delimiter



➤ Preliminary results using a different variety: Doublon



> Perspectives

Short-term perspectives:

1. Run on PromethION using a variety **genetically distant** from the provided reference: 'Canton'
2. **Multiplexing** on PromethION using **two barcodes**: Varieties 'Anso' + 'Doublon'

Middle-term perspectives:

1. **Increase the number of barcodes** on PromethION: Selective sequencing of 150 varieties.
2. Evaluate the ability to detect **structural variants** and **copy number variations** between varieties.

Long-term perspectives:

1. Perform a similar experience using a **diploid heterozygote** species with a **higher number of NLR genes** and **more complex clusters**: *Vitis vinifera*.

> In conclusion

What Nanopore adaptive sampling offers?

- Nanopore adaptive sampling is a **simple, reliable, efficient** and **cost-saving** approach.
 - ✓ Standard DNA extraction (no need for HMW DNA)
 - ✓ Standard library preparation
 - ✓ No need of probes design/synthesis
 - ✓ No need of short-read polishing with Kit14
 - ✓ Real-time sequencing
 - ✓ Reduced off-target volume of data compared to WGS

What can we do?

1. Adaptive sampling allows to efficiently retrieve (**mapping** and **assembly**) our **15 ROI** in melon.
2. It will allow to **explore the genetic diversity** of melon **NLR genes** using a **large amount of varieties**.

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