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Nutritional modulation of associations between prenatal exposure to persistent organic pollutants and childhood obesity: a prospective cohort study --Manuscript Draft--

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Corresponding Author:	Maribel Casas, PhD Instituto de Salud Global Barcelona Barcelona, Barcelona SPAIN			
Corresponding Author Secondary Information:				
Corresponding Author's Institution:	Instituto de Salud Global Barcelona			
First Author:	German Cano-Sancho			
First Author Secondary Information:				
Order of Authors:	German Cano-Sancho			
	Charline Warembourg			
	Nuria Güil			
	Nikos Stratakis			
	Aitana Lertxundi			
	Amaia Irizar			
	Sabrina Llop			
	Maria-Jose Lopez-Espinosa			
	Xavier Basagaña			
	Juan Ramon González			
	Xavier Coumoul			
	Silvia Fernandez			
	Jean-Philippe Antignac			
	Martine Vrijheid			
	Maribel Casas			
Order of Authors Secondary Information:				
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Abstract:	Background : Prenatal exposure to persistent organic pollutants (POPs) may contribute to the development of childhood obesity and metabolic disorders. However, little is known about whether the maternal nutritional status during pregnancy can modulate these associations. Objectives : The main objective was to characterize the joint associations and interactions between prenatal levels of POPs and nutrients on childhood obesity. Methods : We used data from to the Spanish INMA birth cohort, on POPs and nutritional biomarkers measured in maternal blood collected at first trimester of pregnancy and child anthropometric measurements at 7 years. Six organochlorine compounds (OCs) (dichlorodiphenyldichloroethylene, hexachlorobenzene [HCB], β -hexachlorocyclohexane [β -HCH] and polychlorinated biphenyls 138, 153, 180) and four			

	B12 and tolate), polyunsaturated fatty acids (PUFAs), and dietary carotenoids. Two POPs-nutrients mixtures datasets were established: (i) OCs, PFAS, vitamins, and carotenoids (n=660) and (ii) OCs, PUFAs, and vitamins (n=558). Cumulative effects of mixtures on obesity were characterized using Bayesian Kernel Machine Regression (BKMR). Relative importance of biomarkers and 2-way interactions were identified using Gradient Boosting Machine, hierarchical group-lasso regularization, and BKMR. Interactions were further characterized using multivariate regression models in the multiplicative and additive scale. Results : Forty percent of children were overweight or obese. We observed a positive overall cumulative effect of both POPs-nutrients mixtures on overweight/obesity risk, being HCB and vitamin B12 the biomarkers contributing the most. Recurrent interactions were found between HCB and vitamin B12 across screening models. Relative risk and 95% confidence interval (95% CI) for a log increase of HCB was 1.31 (1.11-1.54, p interaction =0.02) in the tertile 2 of vitamin B12 and in the additive scale a Relative Excess Risk due to Interaction of 0.11 (95% CI, 0.02-0.20) was found. Interaction between perfluorooctane sulfonate and b-cryptoxanthin suggested a protective effect of the antioxidant on overweight/obesity risk. Conclusion : These results support that maternal nutritional status may modulate the effect of prenatal exposure to POPs on childhood overweight/obesity. These findings may help to develop biological hypothesis for future toxicological studies and to better interpret inconsistent findings in epidemiological studies.					
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3 Authors:

German Cano-Sancho^a, Charline Warembourg^{b,c,d,e}, Nuria Güil^{b,c,d}, Nikos Stratakis^b, Aitana
Lertxundi^{d,f,g}, Amaia Irizar^{d,f,g}, Sabrina Llop^{d,h}, Maria-Jose Lopez-Espinosa^{d,h,i}, Xavier
Basagaña^{b,c,d}, Juan Ramon González^{b,c,d}, Xavier Coumoul^j, Sílvia Fernandez^{b,c,d}, Jean-Philippe
Antiene d^a Martine Maille d^{b,c,d}, Maria Courab^{6,d}

- 7 Antignac^a, Martine Vrijheid^{b,c,d}, Maribel Casas^{b,c,d}
- 8

9 Affiliations :

- 10 ^aLABERCA, Oniris, INRAE, Nantes, France
- ^b ISGlobal, Barcelona, Spain
- ^c Pompeu Fabra University, Barcelona, Spain
- ¹³ ^d Spanish Consortium for Research on Epidemiology and Public Health (CIBERESP), Madrid,
- 14 Spain
- 15 ^eUniv Rennes, Inserm, EHESP, Irset (Institut de recherche en santé, environnement et travail)
- 16 UMR_S 1085, F-35000 Rennes, France
- 17 ^fBiodonostia, Unidad de Epidemiologia Ambiental y Desarrollo Infantil. San Sebastian,
- 18 Gipuzkoa, Spain
- 19 ^gFacultad de Medicina, UPV-EHU. Leioa, Bizkaia, Spain
- 20 ^hEpidemiology and Environmental Health Joint Research Unit, Foundation for the Promotion
- 21 of Health and Biomedical Research in the Valencian Region, FISABIO-Public Health,
- 22 FISABIO–Universitat Jaume I–Universitat de València, Av. Catalunya 21, 46020, Valencia,
- 23 Spain
- ¹Faculty of Nursing and Chiropody, University of Valencia, Valencia, Spain.
- 25 ^jUniversité de Paris, INSERM UMR-S1124, Paris, France
- 26
- Corresponding author: Maribel Casas, Barcelona Institute for Global Health (ISGlobal) Campus MAR Barcelona Biomedical Research Park (PRBB) Doctor Aiguader, 88 08003
 Barcelona, Spain Barcelona, Spain. Email: maribel.casas@isglobal.org
- 30

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32 The authors declare that they have no conflict of interest

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61 Abstract

Background: Prenatal exposure to persistent organic pollutants (POPs) may contribute to the
development of childhood obesity and metabolic disorders. However, little is known about
whether the maternal nutritional status during pregnancy can modulate these associations.

Objectives: The main objective was to characterize the joint associations and interactions
between prenatal levels of POPs and nutrients on childhood obesity.

67 Methods: We used data from to the Spanish INMA birth cohort, on POPs and nutritional 68 biomarkers measured in maternal blood collected at first trimester of pregnancy and child 69 anthropometric measurements at 7 years. Six organochlorine compounds (OCs) 70 (dichlorodiphenyldichloroethylene, hexachlorobenzene [HCB], β-hexachlorocyclohexane [β-71 HCH] and polychlorinated biphenyls 138, 153, 180) and four per-/polyfluoroalkyl substances 72 (PFAS) were measured. Nutrients included vitamins (D, B12 and folate), polyunsaturated fatty 73 acids (PUFAs), and dietary carotenoids. Two POPs-nutrients mixtures datasets were 74 established: (i) OCs, PFAS, vitamins, and carotenoids (n=660) and (ii) OCs, PUFAs, and 75 vitamins (n=558). Cumulative effects of mixtures on obesity were characterized using Bayesian Kernel Machine Regression (BKMR). Relative importance of biomarkers and 2-way 76 interactions were identified using Gradient Boosting Machine, hierarchical group-lasso 77 78 regularization, and BKMR. Interactions were further characterized using multivariate 79 regression models in the multiplicative and additive scale.

Results: Forty percent of children were overweight or obese. We observed a positive overall
cumulative effect of both POPs-nutrients mixtures on overweight/obesity risk, being HCB and
vitamin B12 the biomarkers contributing the most. Recurrent interactions were found between
HCB and vitamin B12 across screening models. Relative risk and 95% confidence interval
(95% CI) for a log increase of HCB was 1.31 (1.11-1.54, *p*interaction=0.02) in the tertile 2 of
vitamin B12 and in the additive scale a Relative Excess Risk due to Interaction of 0.11 (95%)

86 CI, 0.02-0.20) was found. Interaction between perfluorooctane sulfonate and β-cryptoxanthin
87 suggested a protective effect of the antioxidant on overweight/obesity risk.

88 **Conclusion**: These results support that maternal nutritional status may modulate the effect of 89 prenatal exposure to POPs on childhood overweight/obesity. These findings may help to 90 develop biological hypothesis for future toxicological studies and to better interpret 91 inconsistent findings in epidemiological studies.

93 **1. Introduction**

94 Growing evidence supports that the environment and nutrition during the early stages of 95 development may impact the subsequent health during childhood, including obesity and 96 metabolic diseases (Gluckman and Hanson 2004; Inadera 2013). Childhood obesity remains a 97 public health priority due to the high prevalence, associated risk of comorbidities and high 98 societal costs (Lin and Li 2021; NCD-Risk 2017), which has been linked to a growing list of 99 environmental factors including synthetic chemicals (Güil-Oumrait et al. 2021; Legler et al. 100 2015). Persistent organic pollutants (POPs) represent a vast family of chemicals characterized 101 by their hydrophobicity, stability, and capacity to bioaccumulate across the trophic chains and 102 widespread in fatty tissues of populations from across the globe (Jones and De Voogt 1999; 103 UNEP 2017; WHO 2010). Some POPs like the pesticide dichlorodiphenyltrichloroethane 104 (*p*,*p*'-DDT), its main metabolite dichlorodiphenyldichloroethane (*p*,*p* '-DDE), or 105 hexachlorobenzene (HCB) have been associated with obesity or metabolic disruption in human 106 prospective studies and supported by several experimental studies (Iszatt et al. 2015; Nadal et 107 al. 2017; Ren et al. 2020; Stratakis et al. 2021; Valvi et al. 2012).

108 Such hydrophobic pollutants present physicochemical commonalities with some nutrients, 109 sharing mechanisms of uptake, transport, and metabolism, and/or targeting similar molecular 110 pathways, which results in a large potential to interact in multiple health outcomes (Cano-111 Sancho and Casas 2021). Specific dietary patterns and nutritional status can modulate the effect 112 of toxicants, and thus it can become a source of heterogeneity in environmental 113 epidemiological research, if not properly addressed (Cano-Sancho and Casas 2021; Hennig et 114 al. 2012). For instance, in a seminal study where nutritional confounding was first reported, the neurotoxic effects of prenatal methyl-mercury exposure were strengthened when 115 polyunsaturated fatty acids (PUFA) were included in the regression models (Choi et al. 2008). 116 The interactive effect of POPs and nutritional compounds in metabolic disorders has been 117

observed in experimental studies; for instance, counteracting the beneficial effects of PUFA ω-118 3 on the prevention of insulin resistance and obesity (Ibrahim et al. 2011; Ruzzin et al. 2010). 119 120 Previous studies within the Spanish longitudinal INMA (INfancia y Medio Ambiente -121 Environment and Childhood) birth cohort have reported positive associations between prenatal 122 exposure to organochlorine compounds (OCs) with higher offspring obesity risk (Agay-Shay 123 et al. 2015; Güil-Oumrait et al. 2021; Valvi et al. 2012; Valvi et al. 2014), but mild or null 124 associations for per- and polyfluoroalkyl substances (PFAS) (Manzano-Salgado et al. 2017). 125 To the best of our knowledge, no previous studies have been conducted to assess the joint effect of prenatal POPs and nutrients to date. Thus, built on the hypothesis that the health effects of 126 127 toxicants may be modulated by the nutritional status (Cano-Sancho and Casas 2021), this study 128 extends the previous INMA work to characterize joint effect of prenatal POPs and nutrients on 129 childhood obesity risk. To this end, we conducted a comprehensive multi-step framework 130 intended to answer major questions in epidemiological mixture analyses including (Barrera-131 Gómez et al. 2017; Braun et al. 2016; Knol and VanderWeele 2012; Lazarevic et al. 2019): 1) what is the effect of individual POPs when other POPs are considered in the model?; 2) what 132 133 is the cumulative effect of the mixture of POPs and nutrients on the specific outcome?; 3) are 134 there interactions between POPs and nutrients within the mixture? and 4) if exist, how these 135 interactions affect the POPs-obesity effect estimates?

136

137 **2. Methods**

138 2.1. Study Population

Data from the Spanish INMA birth cohort were used for the present analysis, extensively described elsewhere (Guxens et al. 2012). Briefly, a total of 2,150 pregnant women from the regions of Gipuzkoa, Sabadell, and Valencia were recruited at first trimester of pregnancy (weeks 10-13 of gestation) from 2003 to 2008. To be eligible, women must be at least 16 years 143 old and present singleton pregnancy, no communication barrier, no reproductive assistance and 144 giving birth in the reference hospital (Guxens et al. 2012). The study was approved by the 145 ethical committees of the centers involved in the study. Written informed consent was obtained 146 from the parents of all children. In the present analysis, we included mother-child pairs with information on blood biomarkers of POPs (OCs and PFAS) exposure and nutrient intake 147 148 (vitamins, PUFAs, and carotenoids) during pregnancy and child anthropometric measurements at 7 years of age (Figure S1). A total of 1241 mothers had information on OCs and vitamins 149 150 and child obesity outcomes (Figure S1). However, since not all women had available 151 information of all POPs and nutrients, we generated two consolidated datasets for the mixture 152 analysis: the ANTIOX dataset with OCs, PFAS, vitamins and carotenoids (n=660 with 30 153 variables) and the PUFA dataset consisting of OCs, vitamins and PUFAs (n=558 with 14 154 variables), see details in Table 1. Among both datasets there is an overlap of 241 mother-child 155 pairs. The list of compounds included within each dataset is detailed in Table 1.

156 2.2. Prenatal POPs determination

157 Blood samples from mothers were collected at recruitment at the end of first trimester of gestation (weeks 10-13 of gestation), aliquoted in 1.5 mL cryotubes and stored at -80 °C until 158 their analysis. Concentrations of HCB, β -hexachlorocyclohexane (β -HCH), *p*,*p*'-DDE, and 159 160 polychlorinated biphenyl (PCB) congeners 138, 153, and 180 were determined in serum 161 samples using gas chromatography equipped coupled to electron capture detector or mass 162 spectrometer as previously described (Goñi et al. 2007; Grimalt et al. 2010; Mendez et al. 2011; 163 Valvi et al. 2012). Concentrations of OCs were adjusted to total serum lipids calculated with 164 the reduced equation using cholesterol and triglycerides determined enzymatically (Phillips et 165 al. 1989). Concentrations of perfluorooctanoate (PFOA), perfluorooctane sulfonate (PFOS), perfluorohexane sulfonate (PFHxS), and perfluorononanoate (PFNA) were determined in 166 167 plasma samples by column-switching liquid chromatography coupled with tandem mass

168 spectrometry at the Institute for Occupational Medicine, RWTH Aachen University (Aachen,

169 Germany), as described previously (Manzano-Salgado et al. 2015; Manzano-Salgado et al.

170 2017). Limits of detection (LOD) ranged between 0.01 and 0.07 ng/ml for OCs and between

171 0.05 and 0.20 ng/mL for PFAS.

172 2.3. Nutritional biomarkers determination

173 Biomarkers of vitamins, PUFAs, and carotenoids were determined in maternal blood using validated analytical methods described elsewhere (Montes et al. 2013; Morales et al. 2015; 174 175 Vioque et al. 2013). Briefly, serum levels of vitamin B12 and folate were measured using a 176 commercially available radioassay (Vioque et al. 2013). Levels of 25-hydroxyvitamin D3 were 177 measured in plasma by high-performance liquid chromatography (HPLC) using a BIO-RAD 178 kit (BIO-RAD Laboratories GmbH, Munchen, Germany) as measure of vitamin D status 179 (Morales et al. 2015). Concentrations of long-chain PUFAs were determined in maternal 180 plasma using fast-gas chromatography (Montes et al. 2013). The levels of carotenoids (α - and 181 β -tocopherol, β -cryptoxanthin, α - and β -carotene, lutein, lycopene, zeaxanthin, and retinol) 182 were measured in serum using HPLC with diode array detection and UV detection at 292 nm, in case of α -tocopherol (Vioque et al. 2013). All biomarkers showed coefficients of variation 183 184 below 10% in inter-assays and 5% intra-assays.

185 2.4. Childhood obesity outcomes

Trained nurses measured weight and height from children at the follow-up visit at 7 years (mean: 7.7, standard deviation (SD): 0.23 - ANTIOX dataset), using standard protocols. Ageand sex-specific body mass index (BMI) z-scores (zBMI) were calculated based on the WHO standard reference curves (de Onis et al. 2007). Overweight and obesity were defined as the proportion of children with values zBMI over >1SDs and >2SDs, respectively (de Onis et al. 2010; Vrijheid et al. 2020).

192 *2.5. Covariates*

193 Information on socio-demographic (age, parity, education,) and lifestyle characteristics 194 (smoking) of the mothers was collected by questionnaires administered to mothers during the 195 first trimester of pregnancy. Measured maternal height and reported weight by the mother at 196 the first trimester visit was used to calculate pre-pregnancy BMI (kg/m²). Data regarding the 197 maternal health status during pregnancy was directly collected from clinical records. 198 Confounding variables were selected on the basis of published literature on established 199 determinants of maternal levels of POPs and childhood obesity risk (Ibarluzea et al. 2011; Llop 200 et al. 2010). All models were thus adjusted for maternal pre-pregnancy BMI (kg/m²), age of 201 the mother (years), education level (primary or without education, secondary, university), 202 smoking during pregnancy (nonsmoking, any smoking during pregnancy), region of residence 203 (Gipuzkoa, Sabadell, Valencia), and child's sex (female, male).

204 2.6. Data analysis

205 The multi-step workflow for data-analysis is illustrated in Figure 1 covering major questions 206 about the effect of mixtures: an exploratory analysis (step 1) to identify the correlations 207 between POPs and nutrients; a preliminary characterization of associations between individual 208 biomarkers and obesity outcomes without accounting for the rest of biomarkers (step 2); a 209 ranking of biomarker importance accounting for the co-exposure in multipollutant models (*step* 210 3); an estimation of the cumulative effect of the mixtures on obesity outcomes (step 4); a 211 screening of 2-way interactions to select suspected pairs (*step 5*); and a refined characterization 212 of those interactions using conventional regression methods to facilitate the interpretation in 213 terms of risk estimation (*step 6*). To this end, we applied a battery of complementary algorithms 214 developed to integrate multiple correlated exposure variables to identify joint-effects and 215 interactions (Barrera-Gómez et al. 2017; Lazarevic et al. 2020).

- 216 2.6.1. Data pre-processing and unsupervised exploratory analysis
- 217

218 The distribution profiles of POPs and nutrients were explored in order to characterize the 219 skewness and identify extreme values, left-censored data, and missing data. Biomarkers with 220 detection frequencies below 75% were removed from the analysis (*p*,*p*'-DDT and PCB118); 221 only complete exposure data was used in the statistical analysis. Multiple multivariate 222 imputation procedure was applied to covariate variables with some missing data following the 223 workflow described elsewhere for data missing at random (van Buuren and Groothuis-224 Oudshoorn 2011). Batches of 15 variables were considered in the multivariable imputation 225 models using the R package mice, developed specifically for each dataset. Pre-processing of 226 continuous data included log-transformation and scaling to the SD to improve model fit, if 227 necessary. Exploratory analysis included Spearman's rank correlation analysis to identify 228 correlation patterns between pairs of biomarkers and support the biomarker grouping in the 229 Bayesian kernel machine regression (BKMR) (see section 2.5.3.) and interpretation of results.

230 2.5.2. Single-biomarker outcome associations

231 Multivariate linear and Poisson regression (MLR) with robust variance were used to 232 characterize the associations between individual biomarkers (POPs and nutrients) and continuous (zBMI) or binary outcomes (normal weight vs overweight and obese combined), 233 respectively. Confounding variables were included in the model as covariates, allowing a 234 235 flexible estimation of marginal effects. Modified Poisson regression with robust variance was 236 computed using the "sandwich" approach, which is considered to provide unbiased estimates 237 of risk ratios under potential model misspecification (Chen et al. 2018). The method was 238 implemented in R software using the *coeftest* function with 'sandwich' and 'lmtest' packages.

239 2.5.3. Biomarker importance from multipollutant models

Three different statistical methods to examine multipollutant associations were selected based on their complementarity to characterize linear and non-linear associations, with the capacity to manage confounding data and identify potential interactions. These models also allow the characterization of rankings of associations for individual exposures, thus providing a measure
of their relative importance within each model, while accounting for the rest of biomarker
effects and the identification of interactions (see next sub-section). A summary of main features
from each model supporting the complementarity are displayed in Table S2.

247 - Group Lasso Interaction Network (Glinternet) is a flexible regularization algorithm 248 designed to identify pairwise interactions in regression models imposing the group-lasso (L1) 249 penalties with strong hierarchy (Lim and Hastie 2015). Thus, if an interaction coefficient is 250 estimated to be nonzero, then its two associated main associations also have nonzero estimated 251 coefficients, controlled by the parameter λ . Optimal λ can be selected by cross validation in 252 order to identify an adequate bias-variance trade-off. In the present study, 10-fold cross-253 validation was used to identify the λ exhibiting the lowest mean squared error, computed with 254 the R package "glinternet". To increase the robustness of findings, the models were fitted to 255 100 bootstrap samples, as described elsewhere (Havard et al. 2019). This feature allowed the 256 measurement of coefficient variability and the frequency of interaction detection across 257 bootstrap samples. Averaged model coefficients were used as variable importance scores 258 considering all of them were at the same scale. The approach is intuitive, may handle a large number of independent variables (e.g., up to 10^5) and their interactions, computationally 259 260 efficient and the results are straightforward to interpret. The method may be limited to 261 characterize non-linear associations, and the built-in package does not allow forcing 262 confounding variables out of model penalties.

Gradient boosting machine (GBM) is one of the first approaches proposed to evaluate
the joint associations of environmental exposures and their interactions (Lampa et al. 2014).
Boosting machines combines a large number of simple regression tree models throughout an
iterative process of simpler models' combination (boosting) to improve the overall model fit.
We implemented the method using the R packages "gbm" and "dismo" (Elith et al. 2008). The

input settings used in the gbm.step function included a learning rate of 0.001, tree complexity 268 269 of 4, bag fraction at 0.8, and 10-fold cross-validation. Learning rate is a weight applied to the 270 parameter that minimizes the loss function; thus, slower learning rates (smaller values) require 271 more iterations to achieve local minima. Tree complexity (interaction depth) is the number of nodes in a tree and should be sufficiently large to capture potential interactions (interactions 272 273 between predictors are also evaluated after running a model). The bag fraction is the proportion 274 of data randomly selected to propose the next expansion in a tree. In order to obtain robust 275 estimates, we replicated the model 100 times and extracted the most frequently detected 276 variables and their interactions across the replicates (>50 %). The relative contribution of 277 biomarkers to the overall model fit were used as variable importance metric. Strengths of GBM includes the capacity to detect non-linear associations and high-order interactions, at a 278 279 moderate computational cost, however the interpretability is often challenging, requiring a 280 second modeling step.

281 - Bayesian kernel machine regression (BKMR). The BKMR framework is a flexible non-282 parametric approach that allows the estimation of the overall effect estimate of multiple 283 correlated exposures accounting for confounding variables (Bobb et al. 2015). The method was 284 implemented with the R package "bkmr" using 10,000 iterations (Bobb et al. 2018). All variables were included in the model using the variable selection mode which allows the 285 286 computation of posterior inclusion probabilities (PIPs) to support the selection of most relevant 287 variables and rank the variables according to their probability to be included in the model as 288 approximate measure of variable importance. Important assets of BKMR compared to the 289 previous methods include the unique capacity to measure the cumulative effect of the mixture 290 and the model structure specifically accounting for confounding variables. Despite the method 291 can efficiently account for complex interactions, the identification process involves graphical 292 inspection that can become sometimes conflicting.

293 2.5.4. Cumulative effect of mixtures

The joint effect of the mixture composed by those POPs and nutrients with higher PIPs was characterized using BKMR with hierarchical variable selection, as described above and using the OverallRiskSummaries function from the "*bkmr*" R package. The summary estimate displayed the overall effect of biomarkers as the comparison of the predicted outcome when all biomarkers are fixed at given percentiles with the predicted outcome when all biomarkers are fixed at the 10th percentile.

300 2.5.5. Screening of 2-way interactions

301 In this step the 2-way interactions included in the multipollutant models were identified. With 302 Glinternet, two-way interactions were identified through the inherent strong hierarchical fitting 303 process, together with those main-effects likely to be nonzero. In the case of GBM, as other 304 tree-based models, interactions between predictors are inherently included, as some of the 305 regression trees used are likely to be asymmetric (thus inducing interactions between 306 variables), where the response of one variable depends on the others higher in the tree. The 307 presence of interactions was assessed using the gbm.interactions routine from the "dismo" 308 package. Briefly, the interactions were identified if departures of model predictions for linear 309 combinations of pair of variables was elucidated (Elith et al. 2008). For BKMR, two-way 310 interactions were graphically explored using the cross-section plots depicting the exposure-311 response function for a given exposure when another other exposure was fixed at the 25th, 312 50th, or 75th percentile fixing the rest of exposures to the median.

In order to develop a priority list of interactions for refined analysis, we selected those chemicals pairs considering the criteria of being a pair of POPs and nutrients and either exhibiting: the most influential or largest strength estimates in at least one approach (e.g. contribution for GBM, coefficients for Glinternet, clear visual trends for BKMR) or weak/moderate strength estimates in at least two approaches or obesity outcomes (i.e.

318 continuous or binary). For those methods involving bootstrapping samples (GBM and
319 Glinternet), a frequency detection threshold was defined based on the number of interactions
320 identified (e.g. 50% large number of interactions or 25% low number of interactions).

321 2.5.6. Characterization of interactions for selected pairs

322 In the latter step, we aimed to characterize the joint effect of pairs biomarkers (i.e. POPs and 323 nutrients) identified in the previous screening phase to facilitate interpretation. We first built 324 the generalized additive models (GAM) including the interaction product term for the selected 325 pairs of variables, considering the POPs in continuous scale and nutrients in categorical scale 326 (tertiles), adjusted for the abovementioned covariates. Interaction plots were inspected in order 327 to characterize the shape of the exposure-response functions and refine the regression models. 328 Unlike distributions of nutrients broken in tertiles for a better interpretation of interactions, 329 POPs were categorized in quartiles if departure from linearity was graphically visualized. Risk 330 estimates of POPs were then evaluated across the different tertiles of nutrients using linear or 331 robust Poisson regression models for zBMI or overweight/obesity risk, respectively, as detailed 332 above. In order to formally evaluate the departures from additive joint effects we also estimated the relative excess risk of overweight/obesity due to interaction (RERI) with the regression 333 334 models' coefficients from Poisson regression (Knol et al. 2007; Knol and VanderWeele 2012). 335 The additive interaction is present when the RERI (95% confidence interval) >0 if positive or 336 <0 if negative.

337

338 **3. Results**

339 *3.1. Unsupervised exploratory analysis*

The prevalence of overweight including obesity was 43% and 41% in the ANTIOX and PUFA datasets, respectively (Table S1). The respective mean age of children ranged was 7.7 and 7.3 years old, having an equal proportion of girls and boys in both datasets (Table S1). Mothers 343 had a mean age of 30 years at recruitment with a presence of about 30% of smokers at some 344 moment during pregnancy. Levels of OCs and vitamins were similar between datasets (Table 1). The correlation analysis showed high positive correlations ($\rho > 0.5$) between POPs within 345 346 families (Figure 2A and 2B), and mild or no correlation between families. In turn, p,p'-DDE 347 was not correlated with PFAS and moderately correlated (ρ : 0.3-0.2) with PCBs, HCB, and β -348 HCH. Regarding the fatty acids, arachidonic acid (AA) and docosahexaenoic acid (DHA), both 349 showed positive correlations with PCBs and negative correlation with β -HCH (ρ : 0.3 and -0.1, 350 respectively). Similar but weaker association profiles were shown for eicosapentaenoic acid 351 (Figure 2B), and LA was negatively correlated with PCBs and the rest of PUFAs. Most OCs 352 and PFAS congeners were not correlated with dietary carotenoids and vitamins with few exceptions. For instance PFHxS showed mild positive correlations with most nutrients (ρ : 0.1-353 354 0.2), whereas p,p'-DDE was negatively associated with retinol (ρ : -0.3) (Figure 2B).

355 *3.2. Single-biomarkers outcome associations*

356 The contributions and associations of individual biomarkers with overweight/obesity risk are 357 summarized in Figure 3 (ANTIOX dataset) and Figure S3 (PUFA dataset); whereas the 358 estimates for zBMI scores can be found in Figures 2 (ANTIOX dataset) and 4 (PUFA dataset). 359 Numeric results are reported in Tables S3 and S4. Single-pollutant models showed consistent 360 statistically significant associations of prenatal exposure to HCB and β -HCH with obesity 361 outcomes in children (e.g. adjusted Relative Risk (RR): 1.18 [95% confidence interval (CI): 1.07, 1.31] per 1-SD increase in log HCB ANTIOX dataset, Figure 3). PFNA was positively 362 363 associated with overweight/obesity risk (e.g., RR 1.10 [95% CI: 1.01, 1.20] per 1-SD increase 364 in log PFNA *p*=0.03, Figure 3) but not with zBMI (Figures S2 and S4). Null associations were 365 found for the rest of POPs, nonetheless PCB153, PCB180 and PFOA were positively associated 366 with overweight/obesity risk but at limit of conventional statistical significance (p=0.06-0.10, 367 Table S3, S4). Estimates for most nutrients were also null with the exception of β - 368 cryptoxanthin, zeaxanthin or α-tocopherol, positively associated with zBMI, whereas
369 docosahexaenoic acid (DHA) showed a negative association.

370 *3.3. Biomarker importance from multipollutant models*

371 Variable selection methods (Glinternet, GBM, BKMR) consistently indicated that HCB and vitamin B12 had a high relative importance in the multipollutant models of overweight/obesity 372 373 risk (Figures 3 and Figure S3) and zBMI (Figures S2 and 4). In turn, β-HCH, PFNA, PFOA 374 and PFOS also scored high, but the ordering was less consistent across datasets, models, and 375 obesity outcomes. Similarly, the ordering of PUFAs in the importance ranking was less consistent between obesity outcomes. For instance, AA was identified among most 376 377 contributing variables on overweight/obesity risk (Figure S3), whereas DHA, LA and ALA 378 appeared among the most important contributors on zBMI models (Figure S4). Among the 379 carotenoids, β -cryptoxanthin was the most important biomarker across models and outcomes, 380 followed by α -tocopherol and γ -tocopherol. In turn, folate showed one of the most inconsistent 381 behaviors across datasets, models, and obesity outcomes, being among the top 5 PIPs from 382 BKMR in one overweight/obesity model (ANTIOX dataset, Figure 3) but scoring low in the 383 rest of the models.

384 *3.4. Cumulative effect of mixtures*

The profile of most relevant chemicals in the mixtures is depicted by the conditional PIPs discussed in the previous section. The hierarchical variable selection approach, built on groups of biomarker nature (i.e. OCs, PFAS, vitamin, PUFA, and antioxidant), resulted in a balanced composition of top ranked biomarkers with representatives from each family. The overall cumulative effect of the mixture was positive, mostly linear and significant, for overweight/obesity risk (Figure 4A) and zBMI (Figure 4B). Details of most relevant biomarkers in the mixtures can be found in Figures S9-12.

392 3.5. Screening of 2-way interactions

393 The screening of interactions relied in three computational methods (Glinternet, GBM, and 394 BKMR) with complementary features to fit the data and identify interactions. For that reason 395 and in order to increase the robustness of findings, we selected interactions with the largest 396 strength present in at least a single model or weaker strength but selected by multiple models. 397 Results from each model and outcome are summarized in tables and interaction network plots 398 depicting the bivariate interactions in the ANTIOX (Table S5 and Figures S5-6) and PUFA 399 datasets (Table S5 and Figures S7-8). Specific interactions outputs from GBM (bootstrap 400 interaction contributions) and Glinternet (bootstrap interaction coefficients) can be found in 401 Figures S13-14 for overweight/obesity risk and Figures S15-16 for zBMI. The frequency of 402 interactions and the variability of strength attributed to each interaction across bootstrap 403 samples is depicted with the raw shaded points and summarized with the means depicted with 404 the white dot.

405 Overall, we observed larger uncertainty for interaction coefficients in Glinternet models than 406 model contributions from GBM models. Detection frequency of interactions across bootstrap 407 samples was higher among the findings from PUFA dataset (threshold set up 50%) than 408 ANTIOX dataset (threshold set up 25%). Detailed graphical output of bivariate cross-section 409 from BKMR can be found in Figures S9-12. For the present study we focused on interactions 410 between POPs and nutrients; however, a list of POP-POP and nutrient-nutrient interactions 411 were also automatically identified from GBM and Glinternet, and represented in the interaction 412 network plots (Figures S5-8). In general, interactions between POPs and nutrients were weak 413 and inconsistent across models, with presence of more but weaker interactions within the ANTIOX dataset, whereas in the PUFA dataset, we found less but stronger interactions. 414

In order to conduct a more refined analysis, ten interactions were discerned due to their largest strength/contribution in models, or because they were frequently detected across models (highlighted in bold in Table S5), as detailed in the prioritization criteria. For instance, the 418 interaction between HCB and vitamin B12 showed the largest contribution in the GBM models 419 for overweight/obesity risk but also graphically suggested in the BKMR model (Table S5). 420 Similarly, but to lesser extent, the interaction between β -HCH and vitamin B12 was found in 421 the ANTIOX dataset (Table 2). The pesticide β -HCH appeared to interact with folate, 422 recurrently detected by GBM in the ANTIOX dataset and Glinternet in the PUFA dataset 423 (Table S5). In turn, β -HCH also showed frequently detected interactions with ALA and LA 424 (Table S5). The interaction between PCB138 and LA also exhibited the highest coefficient and 425 largest detection rates in the Glinternet model for overweight/obesity risk. The interactions 426 between PFOA and vitamin B12, and between PFOS and γ -tocopherol, β -cryptoxanthin, and 427 retinol were also considered of priority interest based on detection frequency and/or strength 428 (Table S5).

429 3.6. Characterization of interactions for selected pairs of POPs and nutrients

For the selected ten interactions, we further conducted a regression analysis using GAM models and supported by graphical summaries (interaction plots) in order to identify non-linearity and the direction of interactions. This visualization allowed for instance to identify the potential synergic effect of HCB and vitamin B12 (Figure 5A) and the potential protective effect of β cryptoxanthin on PFOS (Figure 5B). To further characterize the impact of the identified interactions between OCs and vitamins, we used the dataset with complete data on OCs and vitamins (n=1,241) as depicted in Figure S1.

The regression analysis confirmed those effects, being the associations between HCB and childhood overweight/obesity risk and zBMI strengthened at higher levels of vitamin B12 (Table 3). For instance, the associations between HCB and overweight/obesity risk at tertile 2 of vitamin B12 showed a RR 95%CI of 1.31 (1.11-1.54), whereas at tertile 1 were close to the null 0.99 (0.85-1.14) (p_{int} =0.02). Those trends were also observed in the additive scale, with corresponding RERIs 95%CI of 0.11 (0.02-0.20) for tertile 2 and 0.12 (0.03-0.21) for tertile 3. 443 A similar but weaker trend was also observed for the interaction between β -HCH and vitamin 444 B12 (Table S6) and between β -HCH and folate (Table 3). In the latter case, despite being 445 statistically not significant at the multiplicative scale, a synergism was suggested in the additive 446 scale between β -HCH and tertile 2 folate with a RERI 95%CI of 0.11 (0.01-0.21).

447 Synergistic interactions were also noticed between PFOS and retinol on the associations with 448 overweight/risk and zBMI, yet at the limit of statistical significance (p_{int} =0.069) at the 449 multiplicative scale (Table 3). For example, at the highest concentration of retinol, the RR 450 95%CI of overweight/obesity of PFOS was 1.42 (0.96-2.11) for quartile 3 vs 1. Likewise, in 451 the additive scale the estimates supported the synergism with a RERI of 0.58 (0.11-1.05) for 452 the same contrast. Similar associations were also observed between PFOS and γ-tocopherol, 453 with a RERI of 0.60 (0.06-1.15) at highest levels of PFOS and γ-tocopherol.

In the opposite direction, a negative interaction was observed between PFOS and β cryptoxanthin. In this case, the associations between PFOS and overweight/obesity or zBMI, were substantially higher at the lowest tertile of β -cryptoxanthin, reaching a RR 95%CI of 1.59 (1.02-2.05) for quartile 4 vs 1 of PFOS. Whereas the interaction was not statistically significant in the multiplicative scale (p_{int} >0.1), the RERIs supported antagonisms in the additive scale (Table 3).

460 Finally, the synergistic interactions suggested between POPs and PUFAs appeared to be mostly
461 weak and non-statistically significant in both scales (Table S6) with the exception of PCB138
462 and linoleic acid (LA) that showed a RERI 95% of 0.18 (0.02-0.33) in the third tertile.

463 **4. Discussion**

In the present study we have attempted to develop and apply a comprehensive statistical framework to evaluate the mixture effect of prenatal exposure to POPs and nutrients on childhood overweight/obesity. This approach, applied to the population-based birth cohort study INMA, confirmed findings from previous studies in this cohort, highlighting the role of

prenatal exposure to HCB (Agay-Shay et al. 2015; Güil-Oumrait et al. 2021; Valvi et al. 2012) 468 469 and β -HCH (Agay-Shay et al. 2015) on childhood obesity risk and providing evidence of a 470 positive cumulative effect of the mixture of POPs and nutrients. Screening for interactions 471 using advanced approaches highlighted a number of potential combinations. Conventional 472 regression models allowed to translate those interactions in more meaningful effect estimates 473 in terms of inferential interpretation. Among the ten POPs-nutrient interactions retained in the 474 screening step, HCB x vitamin B12 was the most consistent across models and outcomes, and 475 the regression models suggested a potential synergistic effect. Interaction between PFOS and 476 β -cryptoxanthin suggested a protective effect of the antioxidant on overweight/obesity risk, 477 with a higher risk associated with PFOS exposure only being observed at lower concentrations 478 of β -cryptoxanthin.

479 Despite the raise of multipollutant modelling approaches, few studies have considered mixtures 480 of biomarkers others than pollutants (Lazarevic et al. 2019). In fact, some dietary nutrients have 481 the potential to counterbalance the effects of environmental pollutants, highlighting the interest 482 of accounting them in the mixture model (Cano-Sancho et al. 2020; Cano-Sancho and Casas 483 2021). The mixture analysis allowed the identification of an unexpected synergistic effect of 484 vitamin B12, strengthening the associations of HCB. Vitamin B12 is an essential hydrophilic 485 vitamin mainly found in animal food products, being dairy products and meat the major 486 contributors exhibiting a short half-life in the body (Obeid et al. 2019). Despite the lack of 487 international consensus on the optimal levels during pregnancy, there is some agreement that 488 the range between 220 and 850 pmol/L would be adequate (Sukumar et al. 2016). Thus, 489 considering that median levels of vitamin B12 within our population were 350 pmol/L 490 (Interquartile range 279-435 pmol/L) most population would fall within that optimal range, 491 with only 12 participants exceeding the upper threshold. Considering that sources of vitamin 492 B12 in humans is exogenous, the blood levels may be determined either by the dietary intake

493 or as result of their metabolism. Thus, we draw different hypothesis that could help to explain 494 the interaction with HCB on obesity risk. First, there is the possibility that vitamin B12 is 495 confounding the true effect of some other concomitant nutrients of animal origin (e.g. total fat, 496 saturated fat, branched amino acids), or specific food items contributing to poor diet quality 497 (e.g. meat) associated with childhood obesity (Chen et al. 2021; Fernández-Barrés et al. 2016). 498 In turn, some authors have suggested that meat could be a determinant of HCB intake (Gasull 499 et al. 2011); however, our previous analysis rule out that hypothesis in the INMA population 500 (Ibarluzea et al. 2011; Llop et al. 2010). The second hypothesis is the actual joint effects, 501 considering that vitamin B12 intake during pregnancy could also be due to supplementation, 502 often found in multivitamin complexes, as previously reported in the same cohort (Navarrete-503 Muñoz et al. 2015). Despite the excessive use of supplements as upstream source of highest 504 levels of vitamins remains to be explored, several mechanistic hypotheses can be developed to 505 explain the synergistic effects with HCB on fetal metabolic programming and increasing the 506 risk of obesity later in childhood. We observed a similar trend for folate on the associations 507 between β-HCH and higher overweight/obesity risk and zBMI, being the associations 508 strengthened among women with higher levels of folate. Interestingly, studies in mice showed 509 that supplementation with high doses of folate during pregnancy was associated with offspring 510 metabolic disruption and obesity related phenotypes (Huang et al. 2014; Kintaka et al. 2020). 511 Modes of action supporting the joint effect of OCs and vitamin B12 or folate on overweight 512 risk can involve epigenetic programming (McKay et al. 2012; Ouidir et al. 2020). A maternal 513 intake of methyl-group donors (e.g. folates, vitamin B12) could also alter the DNA methylation 514 profiles of offspring's metabolic genes (Pauwels et al. 2017). In turn, HCB and vitamin B12 515 can both individually impact the metabolic programing of adipocytes during differentiation, or 516 their DNA methylation profiles (Bastos Sales et al. 2013; Green et al. 2016). Indeed, vitamin 517 B12 plays a crucial role in humans as a cofactor of methionine synthase, which is actively

518 involved in methionine biosynthesis via the re-methylation of total homocysteine. 519 Interestingly, enzymes involved s-adenosyl methionine synthesis two on 520 (phosphatidylethanolamine N-methyltransferase and glycine N-methyltransferase), are 521 transcriptional target of the aryl hydrocarbon receptor (Kim et al. 2018), which is activated by 522 HCB (Chiappini et al. 2022). Thus, we may hypothesize that a maternal intake of methyl-group 523 donors (i.e. vitamin B12), together with a higher HCB exposure contribute to an increased 524 lipogenesis. Considering the active research to establish more accurate recommendations and 525 thresholds of vitamin supplementation during pregnancy (Maruvada et al. 2020), future studies 526 should consider the concomitant presence of environmental pollutants. A third hypothesis 527 could be developed around the fact that higher levels of vitamin B12 may also reflect a 528 metabolic alteration of one-carbon metabolism, as shown in some hepatic disorders (Ermens 529 et al. 2003). Under this scenario, our findings could reflect an effect of HCB on maternal 530 mitochondrial dysfunction (Park et al. 2021), having a direct impact on one-carbon metabolism 531 pathways due to their coupling to the respiratory chains (Bao et al. 2016). This can be 532 manifested by an alteration of cell uptake and utilization of vitamins resulting in imbalanced blood levels of vitamin B12 (Lyon et al. 2020). High levels of vitamin B12 and folate among 533 534 pregnant women have been associated with metabolic disruption and gestational diabetes 535 probably due to a mild liver dysfunction (Chen et al. 2021).

The protective effect of β -cryptoxanthin on the association between PFOS and childhood obesity also deserve attention. β -cryptoxanthin is a naturally occurring carotenoid. It is found in many foods of plant and animal origin (e.g. oranges, apples, egg yolk). It is closely related to β -carotene and has antioxidant properties. Conversely, PFOS is known for its prooxidative activity (Chen et al. 2014) and increase adipogenesis in vitro (Modaresi et al. 2022), but epidemiological studies are globally inconsistent. A study in the same INMA cohort showed mild or null associations (Manzano-Salgado et al. 2017); other previous studies have generally 543 shown inconsistent findings (Lee et al. 2021) and even proposed as anti-obesogens by some authors (Di Gregorio et al. 2019). For the first time, we were able to observe a higher risk of 544 545 obesity among children exposed to higher levels of PFOS and lower levels of this antioxidant 546 during gestation. Current evidence with adult women has shown that concentrations of carotenoids, including β-cryptoxanthin, are inversely associated with BMI and waist 547 548 circumference, with major effect modification by exposure to toxicants like smoking (Kabat et 549 al. 2016). In experimental studies, β -cryptoxanthin exerted and anti-obesogenic effect reducing 550 the body fat of mice and increasing the expression of uncoupling protein 1 (UCP1) in adipose 551 tissue via the retinoic acid receptor (RAR) (Hara et al. 2019). In turn, PFOA and PFOS has 552 been shown also to activate UCP1 in brown adipose tissue, which can modulate the food intake 553 and body weight (Di Gregorio et al. 2019), but our findings suggest the presence of other 554 potential mechanisms to explain the obesogenic effects of PFOS. For instance, PFOS and 555 PFOA are activators of the Peroxisome Proliferator-Activated Receptor-alpha (PPAR-α) in 556 humans. PPAR- α and RAR share a common dimerization partner, the Retinoid X Receptor 557 (RXR). The activation of both receptors (PPAR- α by PFOS) and (RAR by β -cryptoxanthin) 558 could lead to a competitive effect towards this partner (RXR).

559 Biomonitoring studies during perinatal periods supports the fact that women are exposed to 560 multiple environmental chemicals during pregnancy and lactation periods, as critical windows 561 for offspring development (Cano-Sancho et al. 2020; Haug et al. 2018). This exposure 562 paradigm has stimulated the increasing interest in characterizing the joint effect of 563 environmental chemicals during pregnancy on offspring's health outcomes, raising the 564 development and implementation of statistical approaches to address mixture related questions 565 (Lazarevic et al. 2019). Whereas the apparel of algorithms and statistical methods has been growing during the last few years, there is no specific method consistently outperforming the 566 567 others as assessed in simulation studies (Agier et al. 2016; Barrera-Gómez et al. 2017;

568 Lazarevic et al. 2020). For this reason, we have conceived an approach combining multiple 569 models in order to strengthen the robustness of our findings supported by the specific features 570 from each algorithm. The method selection included Glinternet, GBM, and BKMR, based on 571 previous literature supporting their relatively high statistical performance and capacity to characterize the joint associations of correlated variables accounting for their interactions 572 573 (Barrera-Gómez et al. 2017; Lampa et al. 2014). A simulation study showed that BKMR in 574 case of non-monotonic exposure-response relationships, may outperform penalized regression 575 methods that assume linearity (Lazarevic et al. 2020), a group of methods that includes 576 Glinternet. Identifying relevant components of the mixtures remains a major question in terms 577 of public health but also regulatory decision-making. Statistically, this is commonly 578 accomplished by using variable selection, a process that becomes specially challenging as 579 correlation between variables increases (Lenters and Vermeulen 2018). Whereas data-driven 580 approaches such as BKMR may improve the predictive performance of models, those may fail 581 to attribute the true effect to right candidates within the correlated cluster (Braun et al. 2016). 582 Probably, this fact together with the different nature of variable selection method (e.g. 583 Glinternet and GBM), may help to explain some inconsistencies in the variable importance 584 rankings between models. An alternative way to leverage this issue would be to benefit of a 585 priori toxicological knowledge to inform the variable selection process, especially in 586 exploratory contexts where improving predictive performance falls out of scope. Selection of 587 relevant interactions follows a similar process than main effects, based on likelihood 588 penalization in case of Glinternet or 'spike-and-slab' priors in case of BKMR (Bobb et al. 2015; Lim and Hastie 2015). We noticed that detection of interactions becomes specially challenging 589 590 and inconsistent between models when the interaction is weak (Brookes et al. 2004), supported 591 by the fact that low powered studies prone to false positive detection (Christley 2010). For that reason, the agreement criteria across screening methods could be a solution to attenuate the 592

false positive as observed in the case of the strongest interaction between HCB and vitamin B12. In order to increase the robustness of findings from Glinternet and GBM we applied a bootstrapping approach with 100 replicates, allowing the identification of interactions in terms of frequency of detection and relative contribution or strength. The findings also highlighted the presence of other interactions (i.e. pollutant:pollutant or nutrient:nutrient) not discussed in the present manuscript, which may help to illustrate the complex interplay of chemicals within the mixture.

600 The present study should be considered with caution in the light of some study limitations. 601 First, the sample size is relatively small (n=558-1241) for the exploration and characterization 602 of interactions, which might have resulted in low power to detect interacting effects. Second, 603 we applied a data-driven approach to explore potential interactions with biological meaning. 604 The high correlation between some pollutants and our lack of congener-specific knowledge 605 about their obesogenic potential, may increase the risk of exposure misclassification, thus 606 attributing the interactive effect to the wrong chemical within the clusters of highly correlated 607 variables. Current *in vitro* and *in vivo* studies about obesogenic effects of POPs are relatively limited to few congeners, which in turn, can be highly correlated in biological matrices. For 608 609 simplicity and due to the limited sample size, we have focused the study to characterize 2-way 610 interactions; however, higher order interactions cannot be neglected, either between pollutants 611 and nutrients but also, with other individual characteristics like maternal smoking or child's 612 sex, as previously observed (Casas et al. 2015). We may have also failed to accurately measure 613 vitamin levels representative of all pregnancy as these nutrient biomarkers are reflecting the 614 current intakes or relatively short time-frames (Burri et al. 2001; Gregory et al. 1998). 615 However, for POPs, a single spot blood measurement is considered to be indicative of long-616 term exposure, due to their long elimination half-lives as proved also in the INMA cohort 617 (Lopez-Espinosa et al. 2016; Manzano-Salgado et al. 2015). The exploration of interactions is

an emerging and active field of methodological research, and other novel approaches 618 619 accommodating the complexities of pollutant datasets could be considered in future studies 620 (Ferrari and Dunson 2020, 2021). Finally, the biological interpretation of statistical interactions 621 should be considered with caution in part due to the different implication of interaction scales 622 or the definitions used in different fields (Howard and Webster 2013). Following current 623 recommendations, we reported the interactions in multiplicative and additive scale (Knol and 624 VanderWeele 2012), and we believe that our findings may help to develop biological 625 hypothesis for future toxicological studies and better interpret inconsistent findings in 626 epidemiological studies.

627 To sum up, the present study supports the hypothesis that nutritional status during pregnancy 628 can modify the effect of environmental pollutants on child health. Specifically, we have found 629 that high levels of vitamin B12 may strengthen the associations between prenatal exposure to 630 HCB and childhood obesity. In the opposite direction, the dietary antioxidant β -cryptoxanthin 631 might have a protective effect against the obesogenic effects of PFOS. Our findings suggest 632 that independent models may fail to identify weak interactions between pollutants and 633 nutrients; thus combining complementary models may be a more powerful approach to 634 consider. In the light of the public health implications of these findings, further observational 635 and experimental research will be required for confirmation and gaining insight on the complex 636 interplay between pollutants and nutrients during pregnancy on the metabolic programing of 637 the offspring. As highlighted, these interactions may uncover sub-populations at risk for 638 specific chemicals under regulatory policies; but also support more accurate nutritional 639 guidelines during pregnancy.

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Table 1. Distribution (median $[25^{th} - 75^{th} \text{ percentile}]$) of persistent organic pollutants and 897 nutrients in maternal blood for each dataset.

Matamal Biamanhana (ANTIOX	PUFA
Maternal Biomarkers (units)	N=660	N=558
Organochlorine Compounds		
HCB (ng/g lipid)	55.2 [31.3 - 91.0]	39.0 [29.2 - 78.7]
β -HCH (ng/g lipid)	23.6 [6.71 - 35.1]	30.5 [15.3 - 38.4]
<i>p,p</i> '-DDE (ng/g lipid)	134.0 [85.5 - 234]	122.0 [79.7 - 202]
PCB138 (ng/g lipid)	33.5 [23.5 - 45.6]	25.5 [16.0 - 37.6]
PCB153 (ng/g lipid)	52.5 [37.3 - 71.6]	42.9 [29.0 - 61.8]
PCB180 (ng/g lipid)	39.2 [26.7 - 55.6]	30.2 [20.2 - 43.1]
Per- and polyfluoroalkyl substances		
PFOA (ng/mL)	2.10 [1.47 - 2.91]	-
PFOS (ng/mL)	5.88 [4.49 - 7.66]	-
PFHxS (ng/mL)	0.483 [0.37 - 0.66]	-
PFNA (ng/mL)	0.582 [0.43 - 0.76]	-
Vitamins		
Vitamin B12 (pmol/L)	397 [310 - 519]	341 [271 - 428]
Vitamin D (mmol/L)	30.3 [22.9 - 37.6]	29.7 [21.2 - 37.5]
Folate (mmol/L)	17.9 [12.7 - 25.7]	14.2 [9.70 - 23.1]
Poly-unsaturated fatty acids		
LA (% of fatty acids)	-	30.9 [25.2 - 34.2]
ALA (% of fatty acids)	-	0.3 [0.2 - 0.3]
EPA (% of fatty acids)	-	0.3 [0.2 - 0.5]
DHA (% of fatty acids)	-	2.9 [2.3 - 4.1]
AA (% of fatty acids)	-	8.0 [6.8 - 9.6]
Carotenoids		
γ-tocopherol (µmol/L)	1.49 [1.24 - 1.87]	-
α -tocopherol (μ mol/L)	30.9 [26.1 - 35.6]	-
β -cryptoxanthin (μ mol/L)	0.17 [0.111 - 0.25]	-
α -carotene (µmol/L)	0.11 [0.07 - 0.18]	-
β -carotene (µmol/L)	0.278 [0.17 - 0.43]	-
Lutein (µmol/L)	0.22 [0.17 - 0.28]	-
Lycopene (umol/L)	0.408 [0.23 - 0.77]	-
Zeaxanthin (umol/L)	0.06 [0.05 - 0.08]	-
Retinol (µmol/L)	1.95 [1.52 - 2.55]	-

Abbreviations: AA, arachidonic acid; ALA, alpha linolenic acid; β-HCH, β-hexachlorocyclohexane; DHA,
docosahexaenoic acid; EPA, eicosopentaenoic acid; Folate, Folic acid; HCB, hexachlorobenzene; LA, linoleic
acid; PCB138, 2,2',3,4,4',5'-hexachlorobiphenyl; PCB153, 2,2',4,4',5,5'-hexachlorobiphenyl; PCB180,
2,2',3,4,4',5,5'-Heptachlorobiphenyl; PFOA, perfluorooctanoate; PFOS, perfluorooctane sulfonate; PFNA,

903 perfluorononanoate; PFHxS, perfluorohexane sulfonate; *p*,*p*'-DDE, dichlorodiphenyldichloroethylene.

906 Table 2. Summary estimates for the associations between selected POPs across tertiles of nutrients with obesity outcomes from Poisson/lineal 907 regression models with a cross-product interaction term. Additive interactions on overweight/obesity risk are depicted by the relative excess risk 908 due to interaction (RERI) and respective 95% confidence intervals (95%CI). All models were adjusted for maternal age, pre-pregnancy body mass 909 index, smoking during pregnancy, education and region of residence, in addition models on overweight/obesity risk were further adjusted for child 910 sex and age.

			0	verweigh	t/obesity			zBMI	
POP	Nutrient tertiles	RR	95% CI	$p_{ m int}$	RERI	95% CI	β	95% CI	$p_{ m int}$
HCB (log) ^a	Vitamin B12 (<302 pmol/L)	0.99	(0.85; 1.14)	0.022			-0.07	(-0.21; 0.07)	0.002
	Vitamin B12 (302-339 pmol/L)	1.31	(1.11; 1.54)		0.11	(0.02; 0.20)	0.26	(0.12; 0.40)	
	Vitamin B12 (>339 pmol/L)	1.21	(1.06; 1.37)		0.12	(0.03; 0.21)	0.16	(0.05; 0.28)	
β-HCH (log) ^a	Folate (<11.4 mmol/dl)	1.00	(0.88; 1.15)	0.260			0.00	(-0.12; 0.12)	0.110
	Folate (11.4-19.0 mmol/dl)	1.16	(1.02; 1.32)		0.11	(0.01; 0.21)	0.16	(0.06; 0.27)	
	Folate (>19.0 mmol/dl)	1.13	(1.02; 1.25)		0.09	(-0.02; 0.20)	0.12	(0.03; 0.21)	
PFOS Q2 ^b	Retinol (<1.7 µmol/L)	0.90	(0.59; 1.38)	0.069			-0.34	(-0.80; 0.13)	0.076
Q3		0.77	(0.48; 1.22)				-0.26	(-0.73; 0.20)	
Q4		0.97	(0.63; 1.49)				-0.13	(-0.59; 0.33)	
PFOS Q2	Retinol (1.7-2.3 µmol/L)	1.20	(0.83; 1.74)		0.12	(-0.23; 0.81)	0.37	(-0.05; 0.78)	
Q3		1.09	(0.74; 1.61)		0.32	(-0.19; 0.84)	0.26	(-0.17; 0.69)	
Q4		1.16	(0.79; 1.69)		0.18	(-0.36; 0.72)	0.23	(-0.19; 0.64)	
PFOS Q2	Retinol (>2.3 µmol/L)	0.62	(0.35; 1.10)		0.12	(-0.80; 0.38)	-0.30	(-0.72; 0.12)	
Q3		1.42	(0.96; 2.11)		0.58	(0.11; 1.05)	0.35	(-0.06; 0.76)	
Q4		1.10	(0.69; 1.74)		0.11	(-0.45; 0.67)	0.07	(-0.35; 0.49)	
PFOS Q2 ^b	β -cryptoxanthin (<0.1 μ mol/L)	1.32	(0.82; 2.11)	0.244			0.11	(-0.30; 0.52)	0.367
Q3		1.34	(0.85; 2.13)				0.38	(-0.07; 0.82)	
Q4		1.59	(1.02; 2.50)				0.26	(-0.17; 0.69)	
PFOS Q2	β-cryptoxanthin (0.1-0.2 µmol/L)	0.79	(0.49; 1.28)		0.12	(-1.48; 0.30)	-0.02	(-0.45; 0.41)	
Q3		1.05	(0.70; 1.58)		-0.28	(-1.09; 0.54)	0.12	(-0.31; 0.55)	
Q4		1.12	(0.74; 1.69)		-0.44	(-1.32; 0.45)	0.28	(-0.14; 0.70)	
PFOS Q2	β -cryptoxanthin (>0.2 μ mol/L)	0.75	(0.50; 1.11)		0.12	(-1.75; 0.20)	-0.32	(-0.77; 0.12)	
Q3		0.86	(0.60; 1.23)		-0.59	(-1.50; 0.32)	-0.19	(-0.61; 0.24)	
Q4		0.71	(0.47; 1.07)		-1.12	(-2.19; -0.05)	-0.35	(-0.79; 0.09)	
PCB138 (log) ^c	Linoleic acid (<27.6% fatty acids)	0.96	(0.75; 1.24)	0.297			-0.09	(-0.35; 0.16)	0.372
•	Linoleic acid (27.6-33.4% fatty acids)	1.10	(0.86; 1.41)		0.10	(-0.13; 0.33)	0.03	(-0.21; 0.27)	
	Linoleic acid (>33.4% fatty acids)	1.25	(0.99; 1.58)		0.18	(0.02; 0.33)	0.14	(-0.08; 0.36)	

Abbreviations: *p*_{int}, p-value from interaction testing; RERI, relative excess risk due to interaction; RR, adjusted relative risks; SD, Standard Deviation; zBMI, child body mass

- index z-score. Details of chemical abbreviations are provided in Table 1.
- ^a Population sample size n=1241 (see details in the study flowchart in Figure S1)
- 911 912 913 914 ^b Population sample size n=660 (ANTIOX Dataset)
- 915 ^c Population sample size n=558 (PUFA Dataset)
- 916
- 917

918

919 **FIGURE LEGENDS**

920

Figure 1. Statistical Workflow. Abbreviations: BKMR; Bayesian Kernel Machine Regression;
 GAM, Generalized Additive Models; GBM, Gradient Boosting Machine; MLR, multivariate
 linear/Poisson regression analysis; relative excess risk due to interaction (RERI)

924

Figure 2. Spearman's correlation plots depicting the association strength between POPs and nutrients in the ANTIOX (Panel B, n=660) and PUFA (Panel A, n=558) datasets. Details of chemical abbreviations are provided in Table 1.

928

929 Figure 3. Associations between persistent organic pollutants and nutrients with childhood 930 overweight/obesity within the ANTIOX dataset (n=660). The forest plots depict the 931 associations between individual prenatal exposures (log scaled) and risk of childhood 932 overweight/obesity (dashed panel). Summary estimates from single-biomarker models based 933 on multivariate robust Poisson regression are depicted by adjusted relative risk (RR) and 934 respective 95% confidence intervals (95% CI). Variable importance plots (non-dashed panels) 935 depict the rank of variables based on their relative importance in multipollutant models using the absolute coefficients for Glinternet; model contribution for gradient boosting machine 936 937 regression (GBM); and posterior inclusion probabilities (PIPs) for Bayesian kernel machine regression (BKMR). All models were adjusted for maternal age, pre-pregnancy body mass 938 939 index, smoking during pregnancy, region of residence, education, child sex, and age. Details 940 of chemical abbreviations are provided in Table 1.

941 942

943 Figure 4. Overall effect estimates from Bayesian kernel machine regression (BKMR) on the association between mixtures of POPs and nutrients and childhood overweight/obesity risk 944 945 (Panel A) and body mass index z-score (zBMI) (panel B) for the mixtures of chemicals selected 946 with the hierarchical procedure from ANTIOX dataset (gray) and PUFA dataset (black). 947 Details of most relevant chemicals in the mixtures are depicted in Figures S9-S12. Graphs show 948 the difference in the effect estimates when all exposures are at a particular quantile compared 949 to when all are at the 10th quantile as reference. All models were adjusted for maternal age, 950 pre-pregnancy body mass index, smoking during pregnancy, region of residence and education, 951 child sex and age were also included in overweight/obesity models.

952

Figure 5. Interaction plots on the associations between hexachlorobenzene (HCB, log increase) and tertiles of vitamin B12 (Panel A) and between perfluorooctane sulfonate (PFOS, log increase) and tertiles of β -cryptoxanthin (Panel B) on overweight/obesity risk. All models were adjusted for maternal age, pre-pregnancy body mass index, smoking during pregnancy, region of residence, education, child sex and age.







 HCB- PFOS- VIT_B12- FOLATE- TOCOPHEROI- TOCOPHEROI- TPFNA- TPA					1	BKMF	5	
PFOS- 0 VIT_B12- 0 FOLATE - 0 a_tocopherol - 0 pFNA- 0 b_HCH - 0 b_tryptoxanthin - 0 VIT_D - 0 Lutein - 0 Lycopene - 0 PFNXS - 0 PFNXS - 0 PFOA - 0 g_tocopherol - 0 b_carotene - 0 Retinol - 0 Zeaxanthin - 0 PCB153 - 0 PCB180 - 0 PDDE - 0 PCB138 - 0 0.00	0	HCB -					0	
VIT_B12- 0 FOLATE- 0 a_tocopherol- 0 PFNA- 0 b_HCH- 0 b_cryptoxanthin- 0 VIT_D- 0 Lutein- 0 Lycopene- 0 PFHXS- 0 PFOA- 0 g_tocopherol- 0 b_carotene- 0 Retinol- 0 a_carotene- 0 Zeaxanthin- 0 PCB153- 0 PCB153- 0 PCB138- 0 0.00 0.25 0.50 0.75 1.00 PID		PFOS -				o		
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a_tocopherol - 0 PFNA - 0 b_HCH - 0 b_cryptoxanthin - 0 VIT_D - 0 Lutein - 0 Lycopene - 0 PFHXS - 0 PFOA - 0 g_tocopherol - 0 b_carotene - 0 Retinol - 0 a_carotene - 0 Zeaxanthin - 0 PCB153 - 0 PCB153 - 0 PCB138 - 0 0.00 0.25 0.50 0.75 1.00 PID		FOLATE -			ċ	i -		
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b_HCH- 0 b_cryptoxanthin - 0 VIT_D- 0 Lutein - 0 Lycopene - 0 PFHXS- 0 PFOA- 0 g_tocopherol - 0 b_carotene - 0 Retrool - 0 Accarotene - 0 Zeaxanthin - 0 PCB153 - 0 PCB180 - 0 PCB180 - 0 PCB188 - 0 PCB138 - 0		PFNA-		c	5			
b_cryptoxanthin - 0 VIT_D - 0 Lutein - 0 Lycopene - 0 PFHXS - 0 PFOA - 0 g_tocopherol - 0 b_carotene - 0 Retinol - 0 a_carotene - 0 Zeaxanthin - 0 PCB153 - 0 PCB180 - 0 PCB188 - 0 PCB138 - 0		b_HCH -		. 9	,			
VIT_D- 0 Lutein- 0 Lycopene- 0 PFHXS- 0 PFOA- 0 g_tocopherol- 0 b_carotene- 0 Retinol- 0 a_carotene- 0 Zeaxanthin- 0 PCB153- 0 PCB153- 0 PCB180- 0 PCB180- 0 PCB180- 0 PCB188- 0		b_cryptoxanthin -		0				
Lutein - 0 Lycopene - 0 PFHXS - 0 PFOA - 0 g_tocopherol - 0 b_carotene - 0 Retinol - 0 a_carotene - 0 Zeaxanthin - 0 PCB153 - 0 PCB153 - 0 PCB180 - 0 PCB180 - 0 PCB188 - 0 PCB138 - 0		VIT_D -		0				
Lycopene - O PFHXS - O PFOA - O g_tocopherol - O b_carotene - O Retinol - O a_carotene - O Zeaxanthin - O PCB153 - O PCB153 - O PCB138 - O 0.00 0.25 0.50 0.75 1.00 PIP		Lutein -		0				
PFHXS - 0 PFOA - 0 g_tocopherol - 0 b_carotene - 0 Retinol - 0 a_carotene - 0 Zeaxanthin - 0 PCB153 - 0 PCB180 - 0 PCB188 - 0 PCB138 - 0 0.00 0.25 0.50 0.75 1.00 PIP		Lycopene -	2	0				
PFOA- 0 g_tocopherol- 0 b_carotene- 0 Retinol- 0 a_carotene- 0 Zeaxanthin- 0 PCB153- 0 PCB180- 0 ppDDE - 0 PCB138- 0 0.00 0.25 0.50 0.75 1.00 PIP		PFHXS -	3	0				
g_tocopherol - 0 b_carotene - 0 Retinol - 0 a_carotene - 0 Zeaxanthin - 0 PCB153 - 0 PCB180 - 0 PCB180 - 0 PCB138 - 0 0.00 0.25 0.50 0.75 1.00		PFOA-	0					
b_carotene - 0 Retinol - 0 a_carotene - 0 Zeaxanthin - 0 PCB153 - 0 PCB180 - 0 ppDDE - 0 PCB138 - 0 0.00 0.25 0.50 0.75 1.00		g_tocopherol -	0					
Retinol - 0 a_carotene - 0 Zeaxanthin - 0 PCB153 - 0 PCB180 - 0 PCB188 - 0 PCB138 - 0 0.00 0.25 0.50 0.75 1.00		b_carotene -	0					
a_carotene - O Zeaxanthin - O PCB153 - O PCB180 - O ppDDE - O PCB138 - O 0.00 0.25 0.50 0.75 1.00		Retinol -	0					
Zeaxanthin - O PCB153 - O PCB180 - O ppDDE - O PCB138 - O 0.00 0.25 0.50 0.75 1.00		a_carotene -	0					
PCB153 - 0 PCB180 - 0 PpDDE - 0 PCB138 - 0 0.00 0.25 0.50 0.75 1.00		Zeaxanthin -	ò					
PCB180 - 0 ppDDE - 0 PCB138 - 0 0.00 0.25 0.50 0.75 1.00 PIP		PCB153-	0					
PCB138-0 0.00 0.25 0.50 0.75 1.00		PCB180 -	0					
PCB138-0 0.00 0.25 0.50 0.75 1.00 PIP		ppDDE -	0					
0.00 0.25 0.50 0.75 1.00		PCB138-	0					
)	0.	00	0.	25	0.50 PIP	0.75	1.00

Multipollutant Models GBM VIT_B12+ HCB-0 b_HCH-•• PFNA-0 PFOA-0 PCB138 -0 ppDDE -0 0 PFOSο a_tocopherol -PFHXS-0 0 FOLATE+ g_tocopherol -0 . Lutein -0 VIT_D b_cryptoxanthin - O 0 a_carotene -Lycopene - O PC8153- 0 Retinol - O Zeaxanthin - O b_carotene - O PCB180 - 0 Ó 3 6 Contribution

idividual regre	ssion models	Glinternet
HCB-		HCB- INNERCONTRACTOR
b_HCH-		a_tocopherol - and the design of the
a_tocopherol-		b_HCH- CARCOR AND ST
PFNA-	}⊷⊷i	PFNA - MPOPO
PCB153-	i-o-i	b_cryptoxanthin - CCC a
PFOA-	HOH	PFOA - COmmon
PCB180 -	Ho-1	PFOS - COMP =
b_cryptoxanthin -	HOH !	VIT_B12 - CONSIGNO
Zeaxanthin -	HOH	VIT_D -
ppDDE-	Ho-I	ppDDE - Commission
g_tocopherol-	HOH	Retinal - Color
VIT_B12-	HOH I	g_tocopherol - Chiefe
PCB138-	HOH	PFHXS - COPIe = 1
FOLATE-	HOH	Lycopene - 🗱 👘 👘
VIT_D-	HOH I	PCB138 - CPerie 1 = =
PFOS-	HOH	FOLATE - CORO
PFHXS-	но́н	PCB180 - 00
Lutein -	нòн	Lutein - 🚥
a_carotene -	нон	a_carotene - 📭 a 💿
Retinol -	нон	PC8153 - CON
b_carotene -	ю́н	b_carotene - Official a
Lycopene -	юі	Zeaxanthin - Com
	0.9 1.1 1.3 1.5 RR (95% CI)	0.0 0.1 0.2 0.3 Absolute Coefficients







Supplemental Material

Click here to access/download Supplemental Material Supplemental Materials.v7_Plain.docx