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1 Inter and intraspecific variability of dieldrin
2 accumulation in *Cucurbita* fruits: new perspectives
3 for food safety and phytomanagement of
4 contaminated soils

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13 **Abstract**

14 Due to past agricultural practices, it is common to identify arable soils contaminated with persistent
15 and potentially toxic organochlorine pesticides (OCPs). Occurrence of OCPs, including dieldrin,

16 in vegetables can lead to chronic exposure of the consumers. Some market vegetables, particularly
17 the Cucurbitaceae, are known to accumulate high OCP concentrations. Dieldrin concentration in
18 *Cucurbita* fruits can exceed the Maximal Residue Limit (MRL) resulting in cultivation and sale
19 restrictions for market gardeners. To assess the intra- and interspecific variability of Cucurbitaceae
20 species for low dieldrin concentration in fruits could be a solution.

21 Here, 24 varieties from seven Cucurbitaceae species were cultivated outdoors in large pots, until
22 fruiting, in soils historically contaminated with dieldrin. More than 330 fruits were harvested and
23 analyzed for determining the inter and intraspecific variability of dieldrin accumulation.
24 Significant interspecific differences occurred with mean fruit concentration ranging between 4.2
25 ± 7.0 and 85.0 ± 19.4 μg dieldrin kg^{-1} fresh weigh (FW) in watermelons (*C. lanatus L.*) and
26 cucumbers (*C. sativus L.*), respectively. Intraspecific differences only occurred for *Cucurbita pepo*
27 L. with mean concentration ranging between 4.9 ± 1.1 and 70.3 ± 3.6 μg dieldrin kg^{-1} FW for the
28 varieties Noire maraîchère and Orélia, respectively. For this plant species, the influence of soil
29 concentration, plant exposure time and biomass on fruit dieldrin concentration depended mainly
30 on varieties.

31

32 **Keywords:** Organochlorine pesticide; *Cucurbita pepo* L., soil contamination, plant exposure time,
33 fruit biomass, soil - plant transfer, food safety

34

35 Introduction

36 Contamination of agricultural soils with organochlorine pesticides (OCPs) is frequently detected
37 world-wide. Even though the use of OCPs is either banished or limited since decades, their
38 persistence is nowadays a continuous challenge for farmers, farmland manager and policy makers
39 (Gavrilescu, 2005; Jorgenson, 2001; Morillo and Villaverde, 2017; Widenfalk, 2002). Indeed,
40 depending on plant species and cultivars, the impregnation of several edible plant parts by OCPs
41 and other Persistent Organic Pollutants (POPs) can occur (Namiki et al., 2018; Otani et al., 2007;
42 Singh and Singh, 2017). Plant species from the Cucurbitaceae family are known to absorb,
43 translocate and accumulate OCPs (Donnarumma et al., 2009; Hashimoto, 2005; Mattina et al.,
44 2004, 2000, 1999; Otani et al., 2007). Consequently, some plant products can contain OCP
45 concentrations above the legal European Maximum Residue Limits (EU MRLs) (Official Journal
46 of the European Union, 2008) and thus cannot be sold and must be destroyed. Finding strategies
47 to decrease OCP bioavailability and/or uptake is therefore a main concern. However, fast, cheap
48 and efficient processes allowing a bioavailability decrease are not yet available (Morillo and
49 Villaverde, 2017). Consequently, short-term and ready to use solutions are needed to enable
50 cultivation of cucurbits on agricultural soils contaminated with OCPs.

51 Several authors have highlighted inter and intraspecific variabilities of OCP accumulation in
52 *Cucurbita* roots, stems, leaves, and fruits (Clostre et al., 2014; Otani et al., 2007; White, 2002;
53 White et al., 2003b). Therefore, knowing the genotypic variability but also environmental factors
54 influencing OCP accumulation may help identifying agronomical solutions to produce *Cucurbita*
55 fruits with low OCP concentrations.

56 In addition, other factors can affect OCP concentration and distribution in plant parts for a same
57 cucurbit variety. Previous studies evidenced the effect of soil contaminant concentration (Clostre

58 et al., 2014; Mattina et al., 2004), plant exposure time (Donnarumma et al., 2009; Kelsey et al.,
59 2006) and root to fruit distance (Whitfield Åslund et al., 2008, 2007) on POP concentration in
60 different tissues from Cucurbits. For instance, an increase of chlordane concentration in the soil
61 led to a higher chlordane concentration in *Cucurbita pepo* roots, stems, leaves and fruits. However,
62 the increase in the fruit was lower than in the other plant parts (Mattina et al., 2004). A similar
63 behavior was reported concerning the chlordecone concentration in pumpkin, cucumber, and
64 christophine (chayote, *Sechium edule* (Jacq.) Sw.) fruits, for which the contaminant concentration
65 increased in the fruit with the soil contamination level, although a generally poor relationship was
66 found (Clostre et al., 2014). However, behavior and accumulation in plant may be contaminant
67 dependent (Namiki et al., 2018), preventing extrapolation of the above results to other OCPs.

68 This in-situ study aimed at determining the inter and intraspecific variability of dieldrin
69 accumulation in fruits of a wide range of *Cucurbita* species commercially available, cultivated at
70 a French market garden field with a historically OCP-contaminated soil, located in a peri-urban
71 perimeter, which supports and protects agricultural areas, near the Bordeaux agglomeration
72 (France). Moreover, several plant and soil factors, which may influence the dieldrin concentration
73 in fruits of *C. pepo* varieties were investigated, with a focus on soil contamination level, plant
74 exposure time, and fruit biomass.

75 **Experimental section**

76 **Plant selection and cultivation**

77 **Selection of plant species and varieties**

78 Based on a survey conducted among French market gardeners, highlighting the most important
79 Cucurbitaceae species locally used, 24 varieties included in seven plant species (*C. pepo* L.,
80 *Cucurbita moschata* Duchesne, *Cucurbita maxima* Duchesne, *Cucumis sativus* L., *Momordica*
81 *charantia* L., *Melothria scabra* Naudin, and *Citrullus lanatus* (Thunb.) Matsum. & Nakai) were
82 selected (Table 1). This selection especially focused on *C. pepo* (11 varieties), because this crop is
83 one of the most important for local market gardeners, allowing investigation over a wide range of
84 phenotypic characters. The seeds were obtained thanks to local suppliers and market gardeners.

85 **Seedlings, transplantation in field, and crop maintenance**

86 Crops were cultivated during spring/summer 2021. Following local farmer's practices, sowing was
87 conducted in an uncontaminated substrate (Motte Bio 20, Proveen Substrates) and plantlets were
88 grown in a greenhouse for a month before transplantation in situ. The field, located in the Bordeaux
89 agglomeration presents mean climatic conditions (for the period 1981-2010) for the cultivation
90 duration (June, July and August 2021) as follow: total precipitations of 62.2, 49.9 and 56.0 mm,
91 mean minimal temperatures of 14.1, 15.8 and 15.7 °C, mean maximal temperatures of 24.5, 26.9
92 and 27.1 °C, mean temperatures of 19.3, 21.3 and 21.4 °C for June, July and August respectively.
93 The field has a sandy soil (85 % sand, 5 % silt, and 10 % clay) classified as Umbrisol (Food and
94 Agriculture Organization of the United Nations, 2015) with a neutral soil pH (6.83), a high organic
95 matter content (9.7 %), and high cation exchange capacity (27 cmol⁺ kg⁻¹). The soil dieldrin
96 contamination (in average 123 ± 32 µg kg⁻¹) would mainly result from the past chronic use of
97 insecticides against in particular the Colorado potato beetle (*Leptinotarsa decemlineata* Say).

98 Other OCPs (aldrin, hexachlorobenzene, and chlordanes) were detected in the soil with maximal
99 concentrations of $6 \mu\text{g kg}^{-1}$, leading to concentrations in the fruits below the limit of quantification
100 (LQ): 0.5, 0.7, 0.4, and $0.4 \mu\text{g kg}^{-1}$ for hexachlorobenzene, aldrin and cis- and trans-chlordane,
101 respectively. Transplantation into the field was performed as follows. For each plant, soil was
102 sampled with a stainless spade in the 0-30 cm soil layer, sieved (1 cm), and homogenized before
103 its introduction into a 25 L plastic pot. During this step, for each pot, soil samples (about 90 g)
104 were collected for dieldrin quantification (results for each pot in SI 1). For each tested plant variety,
105 four plant replicates were set up in individual pot, leading to a total of 112 plants. The 112 pots
106 were placed at least 1 m apart in the field where the soil was sampled. They were buried at two
107 third of their height to limit the increase of soil temperature in the pots and regularly inspected to
108 cut off any roots that might emerge from the drainage holes. Depending on the climatic conditions,
109 especially rainfall events, the crops were watered two or three times a week with local groundwater
110 (dieldrin concentration in the well water was below $0.1 \mu\text{g L}^{-1}$). Once a week, the crops were
111 fertilized with an organic fertilizer (Bio-grow, Biobizz®), with NPK values of 4-3-6, diluted at
112 1:1000 (v:v) with groundwater.

113 **Cucurbit fruit harvest, physiological data collection, and crop termination**

114 The first fruits were harvested one month after the transplantation and then three times a week.
115 They were harvested when they reached the commercial size or when no growth was observed for
116 a few days. For each harvested fruit, the following parameters were recorded: pot number, harvest
117 date, fresh weight (FW), distance to the root collar, length, and circumference. The crops were
118 terminated 3.5 months after transplantation.

119 **Dieldrin analysis in soil and fruits**

120 Concerning the soil, dieldrin was analyzed after n-heptane (n-heptane, 99+ %, residues analysis,
121 ACROS Organic, UK) extraction following the protocol described in Colin et al. (2022). This
122 protocol was adapted to quantify dieldrin in fruits. Each fruit was analyzed individually, except
123 for Mexican cucumbers, which were gathered and analyzed by plant and harvest date due to their
124 small size (about 3 g FW per fruit). After harvest, each fruit was first washed with tap water for
125 removing any soil particle and then wiped with paper towel. The entire fruit (flesh and skin
126 together, even for fruits with non-edible peel) was then cut and stored at $-18\text{ }^{\circ}\text{C}$. Frozen fruits
127 were ground with a knife mill (Pulverisette 11, Fritsch) to obtain a homogenized puree. The
128 following grinding cycle was applied: 5 s at 2000 rpm in reverse mode, 5 s at 2000 rpm, 10 s at
129 6000 rpm and 10 s at 10000 rpm. Liquid-liquid extractions were then performed with 20 g FW of
130 this puree mixed with 10 mL of n-heptane into a 50 mL glass flask and shaken during 48 h (Roller
131 10, IKATM). Supernatant was recovered after 5 min of centrifugation at 1500 rpm (JOUANTM
132 C412) and placed in a 2 mL vial. Prior analysis, a few grains of sodium sulfate anhydrous (Fisher
133 Scientific), used to remove residual water, and 50 ng of phenanthrene d-10 (Restek, France), used
134 as internal standard, were added to the sample. Analyses were conducted as presented in Colin et
135 al. (2022). Validation of this protocol for *Cucurbita* fruits is presented in SI 2

136 **Statistical analysis**

137 Calculations and statistical analyses were performed using the R open source software
138 environment (version 4.2.1, (R Core Team, 2021)) and functions available in the ‘stats’, ‘rstatix’,
139 ‘outliers’ and ‘agricolae’ packages (Alboukadel, 2021; De Mandiburu, 2021; Komsta, 2022; R
140 Core Team, 2021). First, outliers were identified using Grubbs test and removed from the data set,
141 then data normality was checked using the Shapiro-Wilk test. As data were not normal, mean

142 comparisons were made with a Kruskal-Wallis test. Relationships between fruit dieldrin
143 concentrations and other parameters such as soil dieldrin concentration, plant exposure time, and
144 fruit biomass were determined using linear and quadratic models as well as Spearman's correlation
145 tests. The later assessed monotonic relationships, which can be linear or not.

146 **Results and discussion**

147

148 **Fruit biomass and harvest**

149 In total, 334 fruits were harvested. Zucchini's were the most productive (Table 1) with an important
150 discrepancy between varieties producing 1.5 (Adrielle) and 9.75 (Mirza) fruits per plant, in
151 average. Harvest for cucumbers and watermelon was scarce, with small fruits due to the 2021
152 climatic conditions (rainfall higher than 1981-2010 seasonal standards in June, 141 mm vs. 62 mm,
153 and mean temperatures 0.3 and 0.8 °C lower than mean seasonal values of 21.3 and 21.4 in July
154 and August, respectively), promoting plant diseases and limiting growth for those plant species
155 needing warm temperatures. Squash and pumpkin varieties produced in average only one fruit per
156 plant, due to the limitation of the pot cultivation.

157

158 **Intra and interspecific variability of dieldrin accumulation in fruits**

159 Dieldrin concentrations in *Cucurbita* fruits are presented in Figure 1, with significant interspecific
160 differences. Results are expressed on the fruit FW basis as water content variability in *Cucurbita*
161 fruits is low (90 ± 5 %, Kusumiyati et al., 2021) but also to be able to compare with EU MRL. The
162 mean concentrations ranged from 4.2 ± 7.0 in watermelons (*C. lanatus*) to 85.0 ± 19.4 µg dieldrin
163 kg⁻¹ FW in cucumbers (*C. sativus*). Dieldrin concentration ranked in the decreasing order: *C.*

164 *sativus* > *C. maxima* = *M. scabra* > *C. pepo* = *C. moschata* > *M. charantia* = *C. lanatus*. This
165 differs from previous reports where zucchinis contained higher (Donnarumma et al., 2009) or
166 similar (fruits from the market, Khandekar et al., 1982; historically contaminated soil, Saito et al.,
167 2012) dieldrin concentrations than cucumbers. It may be explained by the use of different zucchini
168 varieties, soil types, agricultural practices, root distribution in the soil profile, and dieldrin
169 distribution in the soil bearing phases.

170 *Momordica charantia* and *C. lanatus* produced fruits with the lowest dieldrin concentrations.
171 Khandekar et al. (1982) and Saito et al. (2012) reported similar results. Saito et al. (2012) grew
172 two watermelon varieties on a historically contaminated soil (72 µg dieldrin kg⁻¹ dry weight (DW))
173 and analyzed the peeled fruits: for both varieties, dieldrin concentrations were lower than 1 µg kg⁻¹
174 FW. However, extraction methods being different in Saito et al. (2012) and this study, it was
175 difficult to compare the absolute concentrations. Nevertheless, as in this study, fruits from both
176 watermelon varieties accumulated 4 to 50 times less dieldrin than the other *Cucurbita* species
177 (Saito et al., 2012). The low accumulation level evidenced here would be a trait of the watermelon
178 species rather than a bias due to the tested variety and the protocol used. Regarding *M. charantia*,
179 to the authors knowledge, no other study reported pesticide residues in its fruits with known soil
180 dieldrin concentration. A 3-year Indian survey analyzing market vegetables showed OCP
181 concentration under the detection limit in seven bitter melons, while other vegetables contained
182 some (Khandekar et al., 2009). However, soil and cultivation conditions can highly differ between
183 the bitter melons and other vegetables analyzed.

184

185 Mean fruit dieldrin concentrations for all the *Cucurbita* varieties tested are presented in Figure
186 1. No significant intraspecific difference was found for *C. sativus* (four varieties), *C. maxima* (two

187 varieties), and *C. moschata* (four varieties). For those plant species, all the tested varieties
188 displayed mean dieldrin concentrations above the EU MRLs ($20 \mu\text{g kg}^{-1}$ FW for cucurbits with
189 edible peel except zucchinis, $30 \mu\text{g kg}^{-1}$ FW for cucurbits with non-edible peel). However, it
190 cannot be ruled out that other varieties (untested here) with low accumulation pattern may exist
191 for these three *Cucurbita* species.

192 Mean fruit dieldrin concentrations of *C. pepo* (11 varieties) showed significant differences.
193 Intraspecific variability was important as the mean dieldrin concentration in zucchinis (in $\mu\text{g kg}^{-1}$
194 FW) ranged from 4.9 ± 1.1 for Noire maraîchère to 70.3 ± 3.6 for Orélia. Several zucchini varieties
195 accumulated, in average, lower dieldrin concentrations than the EU MRL ($50 \mu\text{g kg}^{-1}$ FW). The
196 percentage of fruits with dieldrin concentration below the EU MRL was 100 % for Noire
197 maraîchère, more than 90 % for Mirza and Kopana, and between 65 % to 75 % for Anissa, Ola
198 escaladora, and Gold Rush. The Noire maraîchère variety, which displayed lower dieldrin
199 concentration in the fruits, as well as Mirza, which was the most productive variety out of the 11
200 zucchini tested, and presented dieldrin concentration mainly under the EU MRL, are relevant
201 candidates for cultivation in dieldrin-contaminated soils.

202 It is worth mentioning the relationship between the fruit's skin color and fruit dieldrin
203 concentration (SI 3). Among the zucchini varieties with mean dieldrin concentrations below the
204 EU MRL, only one was not dark green, while the opposite was true for the varieties exceeding the
205 EU MRL. The Orelia and Floridor varieties produced both yellow fruits, Adrielle pale green fruits,
206 and Verte non coureuse d'Italie striped clear and dark green fruits. This difference of dieldrin
207 content in fruits could thus be related to their carotenoids content. Indeed, previous studies showed
208 that yellow and orange fruits contained more carotenoids than green ones (Houhou et al., 2021;
209 Kim et al., 2012; Xu et al., 2021). Carotenoids, as well as other lipophilic compounds, are

210 sequestered into chromoplasts to prevent the detrimental effect of their excess on cellular
211 functions (Egea et al., 2010). Houhou et al. (2021) highlighted that, triggering the transformation
212 of chloroplasts to chromoplasts allowed to change zucchini peel color from green to yellow,
213 suggesting that yellow fruits contained more chromoplasts than green ones. Dieldrin and other
214 lipophilic organic contaminant might thus be sequestered and therefore accumulated in
215 chromoplasts with carotenoids. However, more investigations are needed to confirm this
216 observation and hypothesis.

217

218 The bioconcentration factor (BCF), which is the ratio between dieldrin concentration in the fruit
219 and that in the soil, was calculated to confirm that the intraspecific variability was not due to
220 differences in soil contamination level (ranging between 58 and 192 $\mu\text{g dieldrin kg}^{-1}$), as the soil
221 dieldrin concentration slightly differed for each pot (SI 2). The results showed a similar trend as
222 the one evidenced for the mean values of fruit dieldrin concentration (SI 4). This suggested that
223 intraspecific variability would be merely due to phenotypic differences, with possible genome x
224 environment interaction, and not to small changes in the soil dieldrin concentration. Previous
225 studies suggested that changes in POP accumulation in *Cucurbitaceae* plants can be related to
226 differences in (i) contaminant solubilization from the soil involving the occurrence of citric acid
227 in the root exudates (White et al., 2006b, 2003; White and Kottler, 2002; Yoshihara et al., 2014),
228 and (ii) in the POP translocation, involving the major latex-like proteins (MLP) (Fujita and Inui,
229 2021). Differences in MLP gene expression or the affinity of the MLP produced for the POP (Inui
230 et al., 2013; Iwabuchi et al., 2020), as well as root exudate composition and flux might be involved
231 in the inter and intraspecific variabilities found in this study.

232 Based on Figure 2, dieldrin transfer from the soil to the zucchini fruit could be minimized by
233 selecting zucchini varieties characterized by low fruit dieldrin concentration. As for metal(loid)s
234 (Yang et al., 2010; Zeb et al., 2022), selection and breeding of cultivars adapted to the
235 phytoavailable contaminant can be a suitable option for food safety, especially considering the
236 persistent problems in line with OCP residues in arable lands (Tzanetou and Karasali, 2022).
237 However, deeper investigations are needed to determine if low dieldrin accumulation character
238 could be extrapolated to other OCPs. According to Namiki et al. (2018), the accumulation
239 capacities in the shoots and roots of a plant species depend on the molecule considered.

240 For a same *C. pepo* variety, fruit dieldrin concentration varied (Figure 1), with, in some cases,
241 maximal concentrations three to four times higher than minimal concentrations. To explain this
242 observation, the influence of soil dieldrin concentration, plant exposure time, and fruit biomass on
243 the dieldrin concentration of zucchini fruits was investigated.

244

245 **Influence of soil dieldrin concentration on dieldrin concentration in zucchini** 246 **fruits**

247 The effect of soil dieldrin concentration on the fruit dieldrin concentration of *C. pepo* is
248 presented in Figure 3. This was only carried out for this plant species, excluding the Adrielle
249 variety, as the number of fruits harvested per pot, and therefore per soil dieldrin concentration, was
250 too limited (< 3) to be relevant for the other plant species. The dieldrin concentration in potted
251 soils cultivated with *C. pepo* plants varied from 60 to 180 $\mu\text{g kg}^{-1}$ DW. For the range of soil
252 dieldrin concentrations considered, it had a low effect on the fruit dieldrin concentration. Only
253 three zucchini varieties showed a significant positive linear relationship and a quadratic
254 relationship could also be fitted for two of them (details of R^2 , p-value, and equation in SI 5),

255 although the slopes were weak (< 0.4) between dieldrin concentration in fruits and soil: B lor
256 (0.155), Floridor (0.362), and Gold Rush (0.244). For other zucchini varieties (Anissa, Noire
257 mara ch re, Or lia, and Verte non coureuse d'Italie), dieldrin concentration in fruits did not show
258 significant changes with increasing soil dieldrin concentrations, which could correspond to a
259 plateau already reached. This is for instance a frequent plant response to excess of metal(loid)s
260 (Affholder et al., 2020; Baker, 1981). For the last three zucchini varieties (Mirza, Kopana, and Ola
261 escaladora), the range of soil dieldrin concentration was narrower, making it difficult to evidence
262 any possible relationship. Determination of a threshold concentration of dieldrin in soil to obtain
263 zucchini under the EU MRL was therefore not possible for most varieties. For the three zucchini
264 varieties displaying a linear relationship, soil dieldrin concentrations lower than 60, 75 and
265 150 $\mu\text{g kg}^{-1}$ DW (using the same extraction protocol) resulted in mean values of fruit dieldrin
266 concentration under the EU MRL for B lor, Floridor, and Gold Rush, respectively.

267

268 **Influence of physiological factors on dieldrin concentration in zucchini fruits**

269 Figure 4 presents the influence of plant exposure time on dieldrin concentration in *C. pepo* fruit.
270 For five out of the 10 zucchini varieties tested, a significant linear relationship occurred between
271 fruit dieldrin concentration and the plant exposure time. Significant quadratic relationships were
272 also positive for two of these varieties (details of R^2 , p-value, and equation in SI 5). Correlations
273 were positive for Anissa, Floridor, Noire mara ch re, and Verte non coureuse d'Italie, and negative
274 for B lor. This agreed with previous studies highlighting significant differences in fruit dieldrin
275 concentration of *C. pepo* collected at three development stages (Donnarumma et al., 2009). With
276 the plant development, the root system increases, which could result in a more efficient nutrient
277 but also contaminant uptake. Moreover, several plant species (rice, lupine, and maize) display

278 changes in root exudation depending on the growth stage and the variety (Aulakh et al., 2001;
279 Gransee and Wittenmayer, 2000; Lucas García et al., 2001). This may contribute to explain that
280 *C. pepo* varieties were not behaving the same way in this study. Root exudation flux and
281 composition could affect the desorption, uptake (White et al., 2006a, 2003; White and Kottler,
282 2002) and translocation of OCPs (Fujita and Inui, 2021). It could lead either to an increase or a
283 decrease of OCP concentration in the fruit and other plant parts. Indeed, White et al. (2006)
284 reported changes in p,p-DDE translocation as well as accumulation in roots, stems and leaves with
285 plant exposure time for two *C. pepo* subspecies. In addition, the shoots are also growing, increasing
286 the root-to-fruit distance, which was correlated to the plant exposure time, except for the climbing
287 variety, Ola escaladora (SI 6). Whitfield Åslund et al. (2008, 2007) showed that polychlorinated
288 biphenyl (PCB) concentration in stems and leaves decreased with increasing distance from the
289 roots in pumpkin (*C. pepo* spp. *pepo*). As zucchini plants usually have a smaller internodal distance
290 than pumpkin plants, it would probably minimize the root distance effect. However, the climbing
291 variety Ola escaladora was assessed and no correlation was found between fruit dieldrin
292 concentration and the root-to-fruit distance (SI 7), even with fruits harvested up to 2 m height. This
293 suggested that dieldrin translocation to fruits differs from translocation to the above ground
294 vegetative parts.

295 The fruit biomass significantly influenced the fruit dieldrin concentration only for the Floridor
296 variety (Figure 5, SI 5). Nonetheless, this zucchini variety was the only tested one that produced
297 yellow, round-shaped fruits. This could confirm that dieldrin concentration in zucchini was higher
298 in skin than in flesh (Saito et al., 2012). Contrarily to the long-shaped fruits, when the biomass of
299 the round-shaped fruit increases, the skin to flesh ratio decreases. Consequently, the whole fruit

300 dieldrin concentration (determined here) decreased. For the other zucchini varieties, no dilution
301 effect was observed with the increase of the fruit biomass.

302

303 Conclusion

304 To conclude, this study highlighted interspecific variability of dieldrin concentration in *Cucurbita*
305 fruits, as well as intraspecific variability in zucchini fruits, some zucchini varieties producing fruits
306 under the EU MRL (e.g. Noire maraîchère and Mirza). Further investigations would be needed to
307 evidence the mechanisms leading to lower fruit dieldrin accumulation for these varieties such as
308 i) the root exudate composition and flux and their impact on dieldrin bioavailability and uptake,
309 ii) the MLP type and gene expression as *C. pepo* contains 21 MLP genes, which can have different
310 affinity for POP (Fujita et al., 2022), and iii) pigment content such as carotenoids in fruits and its
311 potential relationship with dieldrin concentration.

312 In addition, significant differences in the dieldrin concentration of zucchini fruits were evidenced
313 for a same variety and even individual plant. However, out of the three soil and plant factors
314 investigated, none had a ubiquitous effect on the varieties tested, suggesting that other parameters
315 could be involved. For instance, MLP and carotenoid biosynthesis is regulated by environmental
316 signals such as light, temperature, and water availability, which are changing during the cultivation
317 period (Inui et al., 2013; Sun et al., 2010; Wang et al., 2016; Yuan et al., 2015).

318 Finally, considering the biomass that *Cucurbita* fruits represent and their dieldrin concentration,
319 as well as the potential accumulation in the other plant parts (leaves, stems, and roots), the potential
320 for dieldrin phytoextraction and dieldrin dissipation from the soil by the whole plant will be
321 presented in a forthcoming paper.

322

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470

471 Table 1: number of fruits harvested by variety, plant species, and average biomass of fruits per
472 variety. (mean value \pm SE)

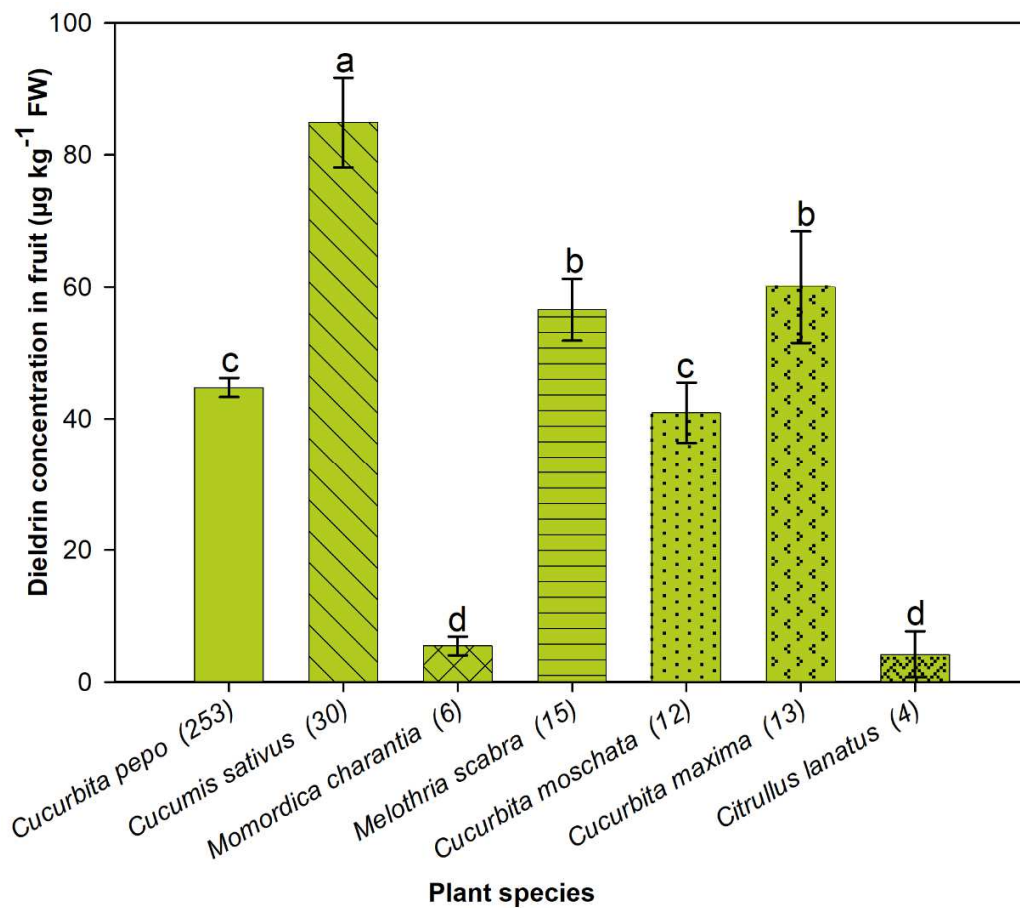
Plant species <i>Latin name</i> Common name	Variety	Morphological traits (color, shape, other)	Number of fruits harvested		Fruit biomass (g FW)
			Total/variety	Mean/plant	
<i>Cucurbita pepo</i> Zucchini	Adrielle	Clear green, long	3	1.50 ± 0.50	401 ±19
	Anissa	Dark green, long	32	8.00 ±2.04	349 ±27
	Bélor	Dark green, long	25	6.25 ±1.75	333 ±38
	Floridor	Yellow, round	16	4.00 ±0.91	382 ±31
	Gold Rush	Golden Yellow, long	26	6.25 ±1.38	318 ±21
	Kopana	Dark green, long	30	7.50 ±1.71	326 ±26
	Mirza	Dark green, long	39	9.75 ±0.48	303 ±17
	Noire maraîchère	Very dark green, long	21	5.25 ±1.38	377 ±27
	Ola escaladora	Dark green, climbing plant	12	3.25 ±1.31	339 ±34
	Orélia	Yellow, long	35	8.75 ±2.78	256 ±16
	Verte non coureuse d'Italie	Dark and clear green stripes, long	15	4.00 ±2.12	397 ±46
<i>Cucumis sativus</i>	Rocker	Green, short with spikes	11	2.75 ±0.63	189 ±18

Cucumber	Diapason	Green, long, smooth	6	1.50 ±0.65	182 ±22
	Le Génereux	Green, short with spikes	5	1.25 ±0.25	140 ±24
	Marketer	Green, short with spikes	8	2.25 ±0.48	215 ±30
<i>Melothria scabra</i> Mexican cucumber	Mexican	Very small fruits (2/3 cm), with dark green and clear green stripes	15	3.75 ±0.25	70 ±11
<i>Momordica charantia</i> Bitter melon	Moonlight	Clear green, bumpy skin	6	1.50 ±0.29	128 ±36
<i>Cucurbita moschata</i> Pumpkin	Longue de Nice	Dark green, long, old French variety	3	0.75 ±0.25	446 ±42
	Sucrine berry	Small, clear orange skin, dark orange flesh, old French variety	4	1.00 ±0.00	559 ±46
	Waltham Butternut	Camel skin	3	1.00 ±0.00	520 ±113
	Hercules butternut	Camel skin, fast growth	2	1.00 ±0.00	393 ±97
<i>Cucurbita maxima</i>	Orange summer	Small, dark orange skin and flesh	9	2.25 ±0.00	444 ±60

Squash	Rouge d'Etampes	Big, red skin, orange flesh	4	1.00 ±0.00	1922 ±213
<i>Citrullus lanatus</i> Watermelon	Sugar baby	Dark green skin, dark pink flesh, fast growing	4	1.00 ±0.00	368±108
TOTAL : 7	24		334	3.56	

473

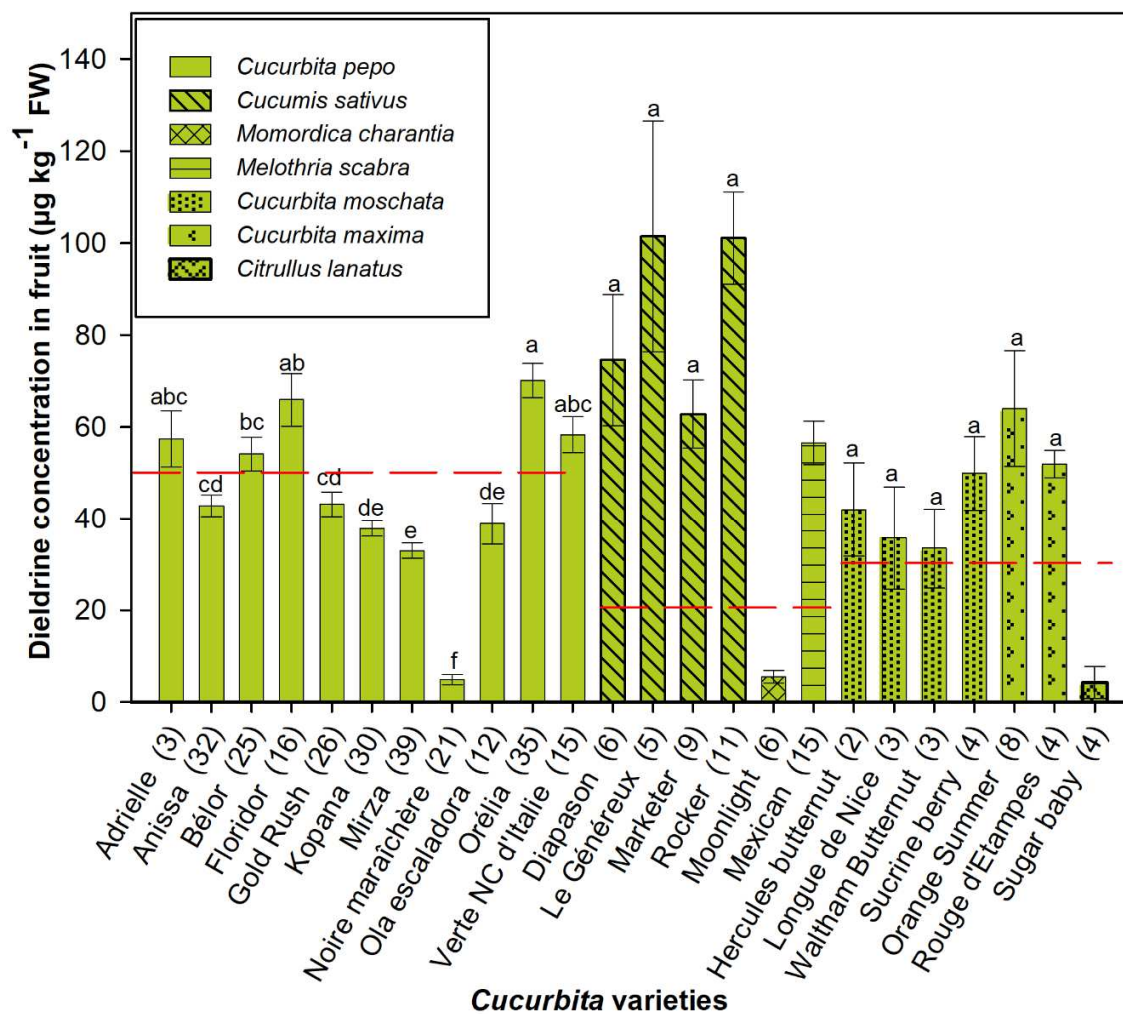
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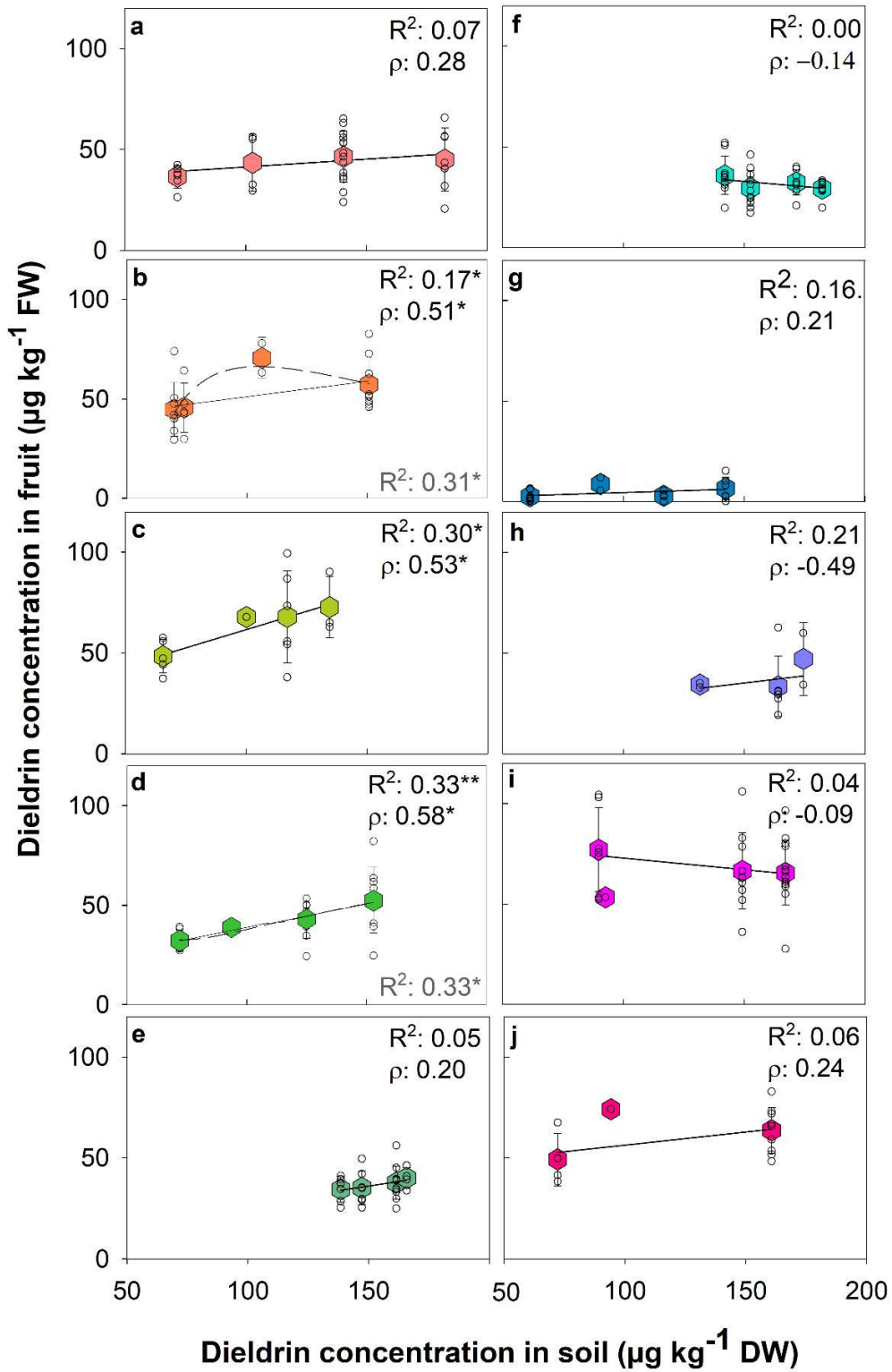
476 Figure 1: Mean values of fruit dieldrin concentration ($\mu\text{g kg}^{-1}$ FW) depending on *Cucurbita* species
 477 (mean \pm standard error (SE)). Mean values with different letters are significantly different
 478 (Kruskal-Wallis, $p < 0.05$). The number of fruits analyzed for each species is presented in brackets.

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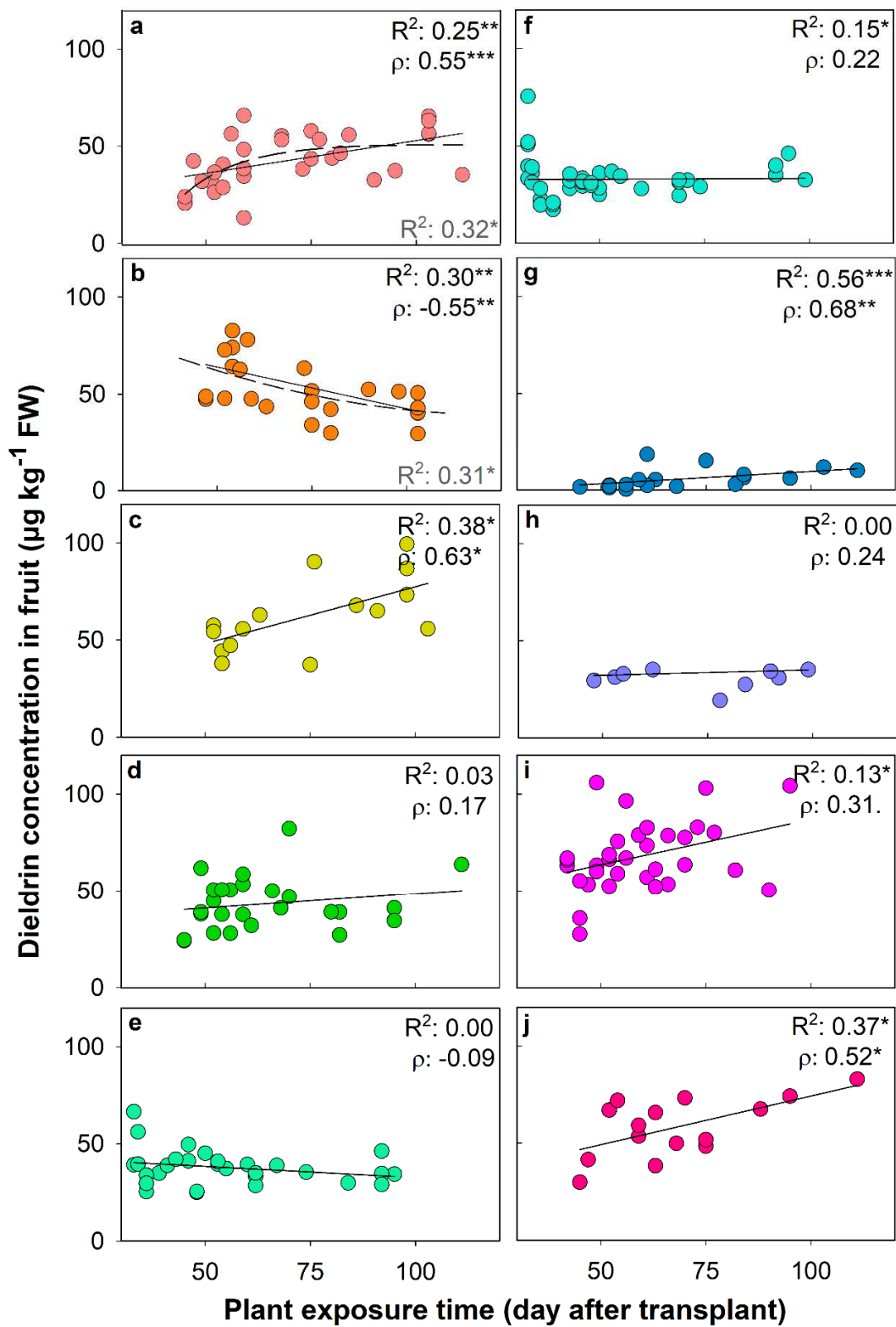
480
 481 Figure 2: mean values (\pm SE) of fruit dieldrin concentrations ($\mu\text{g kg}^{-1}$ FW) depending on *Cucurbita*
 482 varieties. For a same plant species, mean values with different letters significantly differed
 483 (Kruskal-Wallis, $p < 0.05$). Dashed red lines represent the EU MRLs for dieldrin corresponding to
 484 the various plant species: $50 \mu\text{g kg}^{-1}$ FW for *C. pepo*, $20 \mu\text{g kg}^{-1}$ FW for Cucurbits with edible
 485 peel, and $30 \mu\text{g kg}^{-1}$ FW for Cucurbits with non-edible peel. The number of fruits analyzed for
 486 each variety is presented in brackets.

487



489 Figure 3: Fruit dieldrin concentration ($\mu\text{g kg}^{-1}$ FW) depending on the soil dieldrin concentration
490 ($\mu\text{g kg}^{-1}$ DW), for various zucchini varieties: a) Anissa, b) B  lor, c) Floridor, d) Gold Rush, e)
491 Kopana, f) Mirza, g) Noire mara  ch  re, h) Ola escaladora, i) Or  lia, and j) Verte non coureuse
492 d'Italie. Each hexagon corresponds to the mean dieldrin concentration of fruits (\pm SE) from one
493 pot. Dots represent the dieldrin concentration for individual fruit harvested on the corresponding
494 pot. R^2 : coefficient of determination for linear model plotted as a black solid line, ρ : Spearman's
495 rank correlation coefficient. When significant, quadratic regression is plotted as a dashed black
496 line, the corresponding R^2 is presented in grey. Significance levels: $p < 0.001$: ***; $p < 0.01$: **;
497 $p < 0.05$: *; and $p < 0.1$: .

498

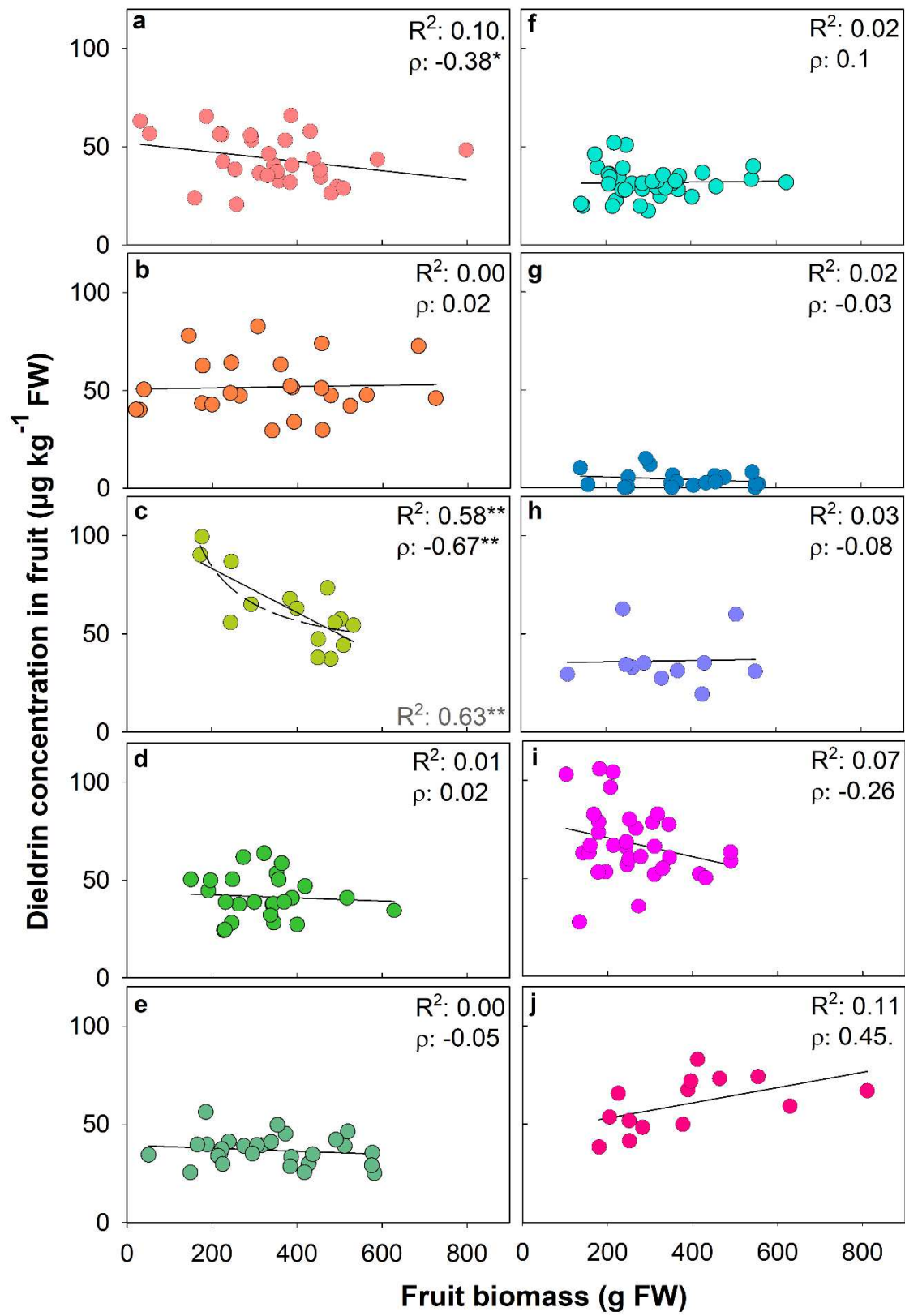


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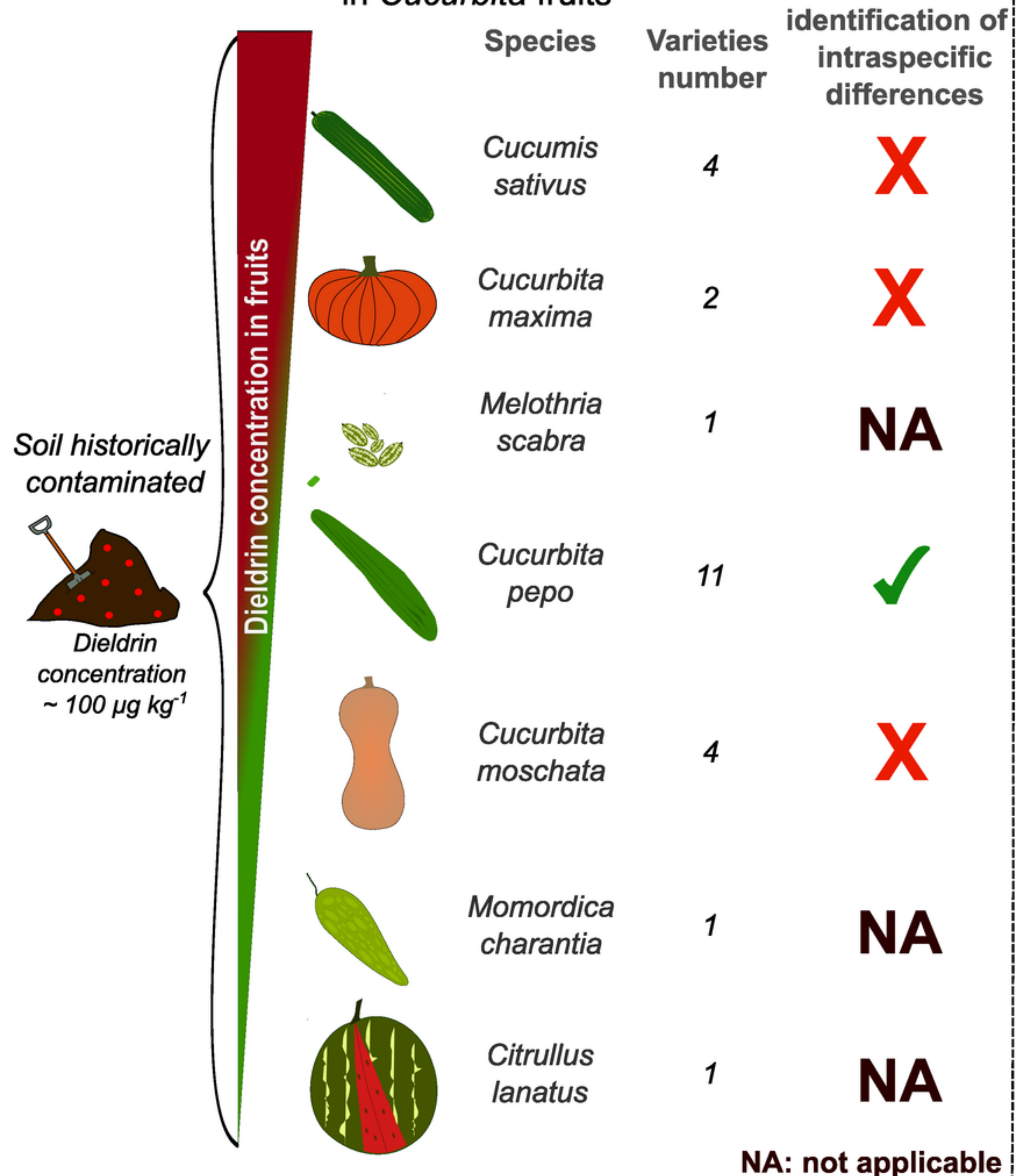
502 Figure 4 : Fruit dieldrin concentration ($\mu\text{g kg}^{-1}$ FW) depending on the plant exposure time (days
503 after transplant) for various varieties of *C. pepo*: a) Anissa, b) B lor, c) Floridor, d) Gold Rush, e)
504 Kopana, f) Mirza, g) Noire mara ch re, h) Ola escaladora, i) Or lia, and j) Verte non coureuse
505 d'Italie. Each dot corresponds to one fruit. R^2 : coefficient of determination for linear model plotted
506 as a black solid line, ρ : Spearman's rank correlation coefficient. When significant, quadratic
507 regression is plotted as a dashed black line, the corresponding R^2 is presented in grey. Significance
508 levels: $p < 0.001$: ***; $p < 0.01$: **; $p < 0.05$: *; and $p < 0.1$: ..



510 Figure 5: Fruit dieldrin concentration ($\mu\text{g kg}^{-1}$ FW) depending on the fruit biomass (g FW) for
511 various varieties of *C. pepo*: a) Anissa, b) B lor, c) Floridor, d) Gold Rush, e) Kopana, f) Mirza,
512 g) Noire mara ch re, h) Ola escaladora, i) Or lia, and j) Verte non coureuse d'Italie. Each dot
513 corresponds to one fruit. R^2 : coefficient of determination for linear model plotted as a black solid
514 line, ρ : Spearman's rank correlation coefficient. When significant, quadratic regression is plotted
515 as a dashed black line, the corresponding R^2 is presented in grey. Significance levels: $p < 0.001$:
516 ***; $p < 0.01$: **; $p < 0.05$: *; and $p < 0.1$: ..

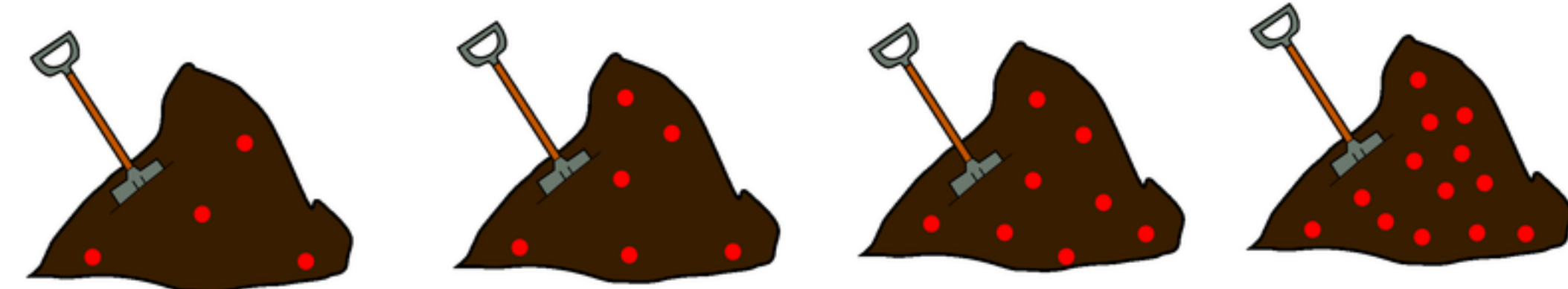
517

1. Inter and intraspecific differences of dieldrin concentration in *Cucurbita* fruits



2. Effect of soil, plant exposure time and fruit biomass on dieldrin concentration in zucchinis

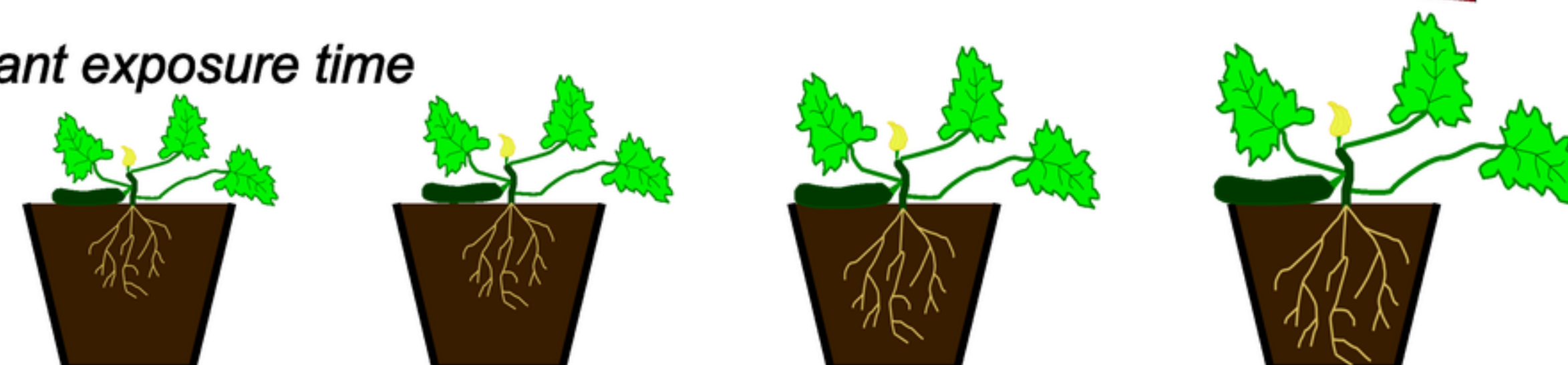
a. Soil dieldrin concentration



Dieldrin concentration in fruits

Varieties significantly affected
Gold Rush
Floridor
Bélor

b. Plant exposure time



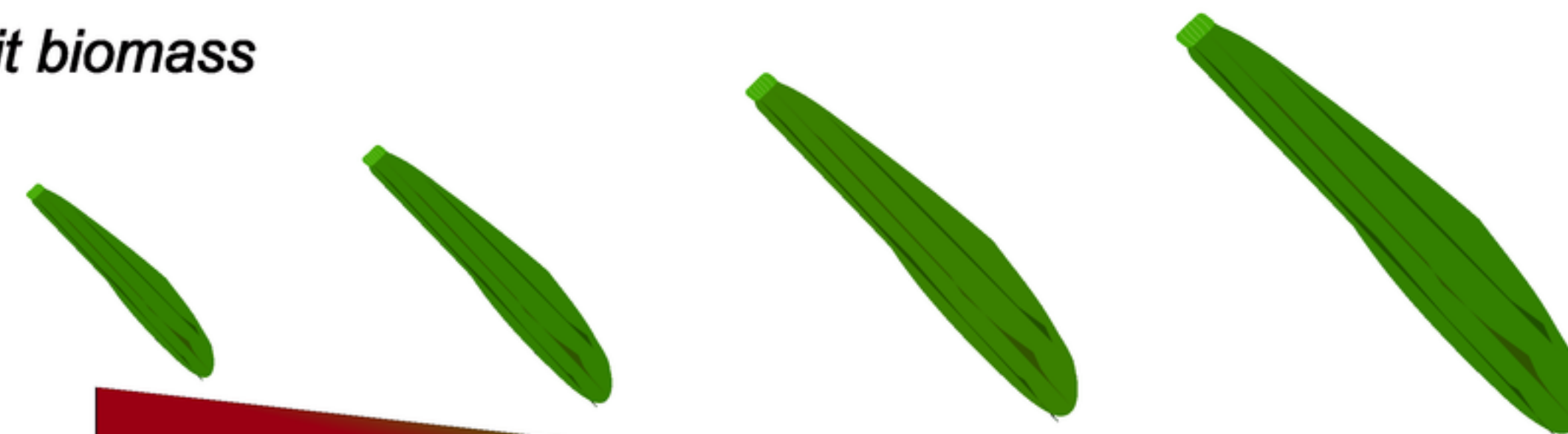
Dieldrin concentration in fruits

Varieties significantly affected
Anissa
Floridor
Noire maraichère
Verte Italie

Dieldrin concentration in fruits

Bélor

c. Fruit biomass



Dieldrin concentration in fruits

Floridor