



HAL
open science

Process formulations and controlling factors of pesticide dissipation in artificial ponds: A critical review

Aya Bahi, Sabine Sauvage, Sylvain Payraudeau, Gwenael Imfeld, José-Miguel Sánchez-Pérez, Betty Chaumet, Julien Tournebize

► To cite this version:

Aya Bahi, Sabine Sauvage, Sylvain Payraudeau, Gwenael Imfeld, José-Miguel Sánchez-Pérez, et al.. Process formulations and controlling factors of pesticide dissipation in artificial ponds: A critical review. *Ecological Engineering*, 2023, 186, pp.106820. 10.1016/j.ecoleng.2022.106820 . hal-04062745

HAL Id: hal-04062745

<https://hal.inrae.fr/hal-04062745>

Submitted on 13 Nov 2023

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 List of abbreviations

2

Abbreviation	Description
TSS	Total suspended solids
DP	Dissolved pesticides
PP	Particulate pesticides
OM	Organic matter
OC	Organic carbon
HRT	Hydraulic residential time
DOC	Dissolved organic carbon
POC	Particulate organic carbon
Re	Reynolds number
vp	Vapor pressure
FWS	Free water surface
OECD	Organization for Economic Co-operation and Development
PPDB	Pesticide Properties
NER	Non-extractable residue
SET	Settling
RES	Resuspension
VOL	Volatilization
SOR	Sorption
BIO	Biotransformation
PHO	Photolysis
HYD	Hydrolysis

3

Process formulations and controlling factors of pesticide dissipation in artificial ponds: a critical review

Aya Bahi^{a*}, Sabine Sauvage^b, Sylvain Payraudeau^c, Gwenaël Imfeld^c, José-
Miguel Sánchez-Pérez^b, Betty Chaumet^b, Julien Tournebize^{a*}

^a INRAE, French National Research Institute for Agriculture, Food and the Environment, University of Paris-Saclay, CS 10030, F-92761 Antony, France

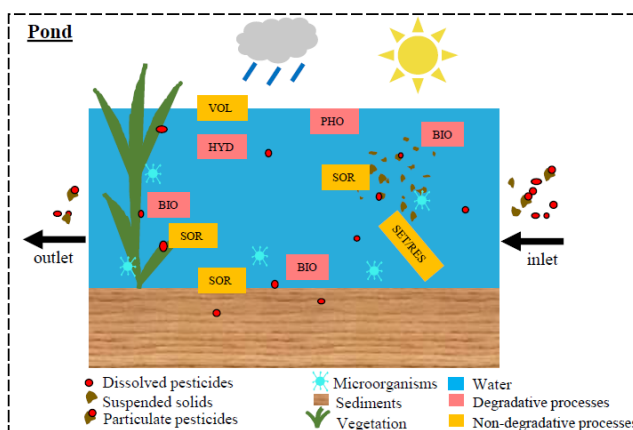
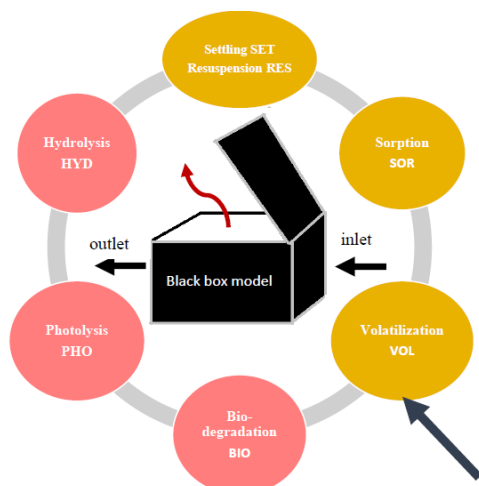
^b Laboratory of functional Ecology and Environment, University of Toulouse, CNRS, UPS, Toulouse INP, ENSAT campus, F-31326 Toulouse, France

^c ITES, Institut Terre et Environnement de Strasbourg (ITES), University of Strasbourg /ENGEES, CNRS UMR 7063, F-67084 Strasbourg, France

* Corresponding authors.

E-mail addresses: aya.bahi@inrae.fr (A. BAHI); julien.tournebize@inrae.fr

Graphical abstract:



17

18

19 **Abstract**

20 Pesticides are a non-point source of pollution affecting the quality of drinking water supplies
21 and aquatic environments. Many facilities are set up to reduce the transfer of pesticides from
22 agricultural parcels into the environment, preventing their input into water bodies. Artificial
23 ponds collecting runoff and erosion fluxes offer a complementary tool to mitigate pesticide
24 transfer. Ponds are free-water surface-constructed wetlands, rarely dried out and not
25 exceeding two meters in depth. Their main compartments include a water column, a sediment
26 layer, living organisms, and suspended particles. The dissipation potential of ponds is due to a
27 combination of physical, chemical, and biological processes that pesticides undergo in an
28 interplay between different pond compartments. Although many studies on the dissipation
29 potential of ponds focus on nitrates and suspended sediments, very little is known about the
30 behavior of pesticides. Also, most studies evaluate a process individually without
31 emphasizing its eventual interaction with other processes. Few papers address and compare
32 mathematical formulations of pesticide transfer and transformation processes for artificial
33 pond modeling. In this review, we investigate the interactions and contribution of multiple
34 processes to artificial pond dissipation function by comparing their rates evaluated at different
35 scales. We also discuss the mathematical formulations of processes and their relationship with
36 pesticide properties. Our evaluation suggests that quantifying the contribution of each process
37 to pesticide dissipation in artificial ponds is a major tool for hierarchizing dissipation
38 processes. We believe that quantification and hierarchization may improve the overall
39 artificial pond efficiency estimation. We anticipate this work to be the basis for developing a
40 process-based model of pesticide fate in artificial ponds. The model will be particularly
41 helpful in setting up the dimensioning criteria to design performant and efficient ponds to
42 mitigate pesticide transfer into the environment.

43 **Keywords:**

44 Artificial pond; pesticide; dissipation; process; model, review.

45

46 Highlights:

- 47 • Identification and formalisms of dissipation processes of pesticides in artificial ponds
- 48
- 49 • Sorption as a key process for lowly soluble and hydrophobic pesticide dissipation
- 50
- 51 • Biotransformation enhanced in water-substrate interfaces favoring microbial activity
- 52
- 53 • Volatilization and hydrolysis can be mostly neglected
- 54
- 55 • Processes interplay contributes more significantly to the dissipation of pesticides
- 56
- 57 • Added value of descriptive process-based model of pesticide behavior in artificial
- 58 ponds
- 59

60

61

62

63

64		
65	List of abbreviations	1
66	1. Introduction and definitions	6
67	1.1 Wetlands and artificial ponds	6
68	1.2 Processes governing the fate of pesticides in artificial ponds	7
69	1.3 Hydraulics of artificial ponds	8
70	2. Description and modeling of dissipation processes in artificial ponds	9
71	2.1 Transport of pesticides	9
72	2.1.1 Settling	10
73	2.1.2 Resuspension	11
74	2.1.3 Advection, dispersion and diffusion	12
75	2.2 Transfer of pesticides	12
76	2.2.1 Sorption process	12
77	2.2.1.1 Sorption theory	13
78	2.2.1.2 Sorption of pesticides in artificial ponds	14
79	2.2.2 Volatilization	15
80	2.3 Transformation of pesticides	17
81	2.3.1 Biotransformation	17
82	2.3.2 Photodegradation	20
83	2.3.3 Hydrolysis	21
84	3. Discussion	23
85	3.1 Controlling factors of the processes	23
86	3.2 Parameter estimation for modeling	24
87	3.3 Hierarchization of dissipation processes	26
88	4. Conclusion	30
89	Appendix A.	33
90	Methodology	44
91	References	47
92		
93		

94

95 **1. Introduction and definitions**

96 **1.1 Wetlands and artificial ponds**

97 In agriculture, various pests (e.g., weeds, insects, rodents, fungi, molds.) can significantly
98 deteriorate the quantity and quality of agricultural products. In order to protect agricultural
99 production, chemicals called "pesticides" are commonly used. Pesticides cover many
100 compounds, including insecticides, fungicides, herbicides, rodenticides, molluscicides,
101 nematicides, plant growth regulators, and others (Aktar et al., 2009). Due to their toxicity,
102 pesticides represent a non-point source of pollution for the ecosystem and affect the quality of
103 drinking water supplies and aquatic environments. Material solutions can be implemented as a
104 complementary tool to safeguard water resources quality and mitigate pesticide input into
105 water bodies, such as edge-of-field and riparian buffer strips, vegetated ditches, wetlands, and
106 artificial ponds (Vymazal et al., 2015). In this paper, we will focus on constructed wetlands,
107 especially artificial ponds, since they have the advantages of needing minimal operations, i.e.,
108 being inexpensive to operate and construct and providing wildlife habitat (Sudarsan et al.,
109 2021). A neat definition of wetlands and ponds and a description of their different types can
110 be found in (Kadlec et al., 2008).

111 Artificial ponds are free water surface constructed wetlands, often kept wet and not exceeding
112 2m depth (Z. Zhang et al., 2021), and they serve different purposes. Artificial ponds can be
113 used as detention ponds to store water for irrigation purposes (Chaumet et al., 2022;
114 Tournebize et al., 2015) or stormwater ponds to prevent flooding (Maillard et al., 2012).
115 Artificial ponds are also used as complementary tools to treat contaminated agricultural water
116 (Tournebize et al., 2017). These artificial ponds intercept agricultural water through runoff or
117 subsurface drainage from cultivated agricultural plots. Artificial ponds are constructed upon
118 an impermeable layer of sediments. The main artificial pond compartments are a water
119 column, a sediment layer, living organisms (i.e., vegetation and microorganisms), and
120 suspended particles. Since the late '80s, artificial ponds have been generally applied to treat
121 various wastewaters, including agricultural water (Chouinard et al., 2014; Grinham et al.,
122 2018; Wang et al., 2018; Zamora et al., 2019; Zhuang et al., 2019). It was widely reported that
123 artificial ponds are able to reduce the transfer of chemicals (pesticides, organics, and
124 inorganics pollutants), physical components (solid suspended particulates), and biological
125 elements (bacteria) to natural water resources and preserve their quality (Kasak et al., 2018;
126 Zheng et al., 2018). For example, artificial ponds had an important removal efficiency for
127 organic matter, solids, and nitrogen up to 90% (de Oliveira et al., 2020; Tournebize et al.,
128 2015). For pesticides, artificial pond efficiency can reach 97% for the organochlorine
129 chemical group (Matamoros et al., 2006; Rose et al., 2006), 96% for the strobilurin/strobin
130 group, and 84% for pyrethroids (Vymazal et al., 2015).

131 In that matter, significant research effort has been dedicated to understanding the fate of
132 pesticides in artificial ponds and their link to the pond and pesticide properties (Bloomfield et
133 al., 2006). In artificial ponds, pesticides are dissolved in water, attached to suspended
134 particles, and sorbed on sediments and vegetation. The dissipation potential of artificial ponds
135 results from a combination of transport, transfer, and transformation processes of pesticides.
136 Transport processes carry pesticides following a water flow. Transfer processes allow the
137 pesticide molecule to move from one phase to another (i.e., dissolved phase to particulate

138 phase or dissolved phase to gas phase), and transformation degrades and modifies the original
139 chemical structure of pesticides.

140 Most studies could not distinguish the extent of removal processes because they depend on
141 each other (Gregoire et al., 2009; Liu et al., 2019; Sahin et al., 2020) and thus could not model
142 their impact on pesticide behavior. Additionally, previous reviews and studies (Stehle et al.,
143 2011; Vymazal et al., 2015) focused on the efficiency of artificial ponds to remove pesticides
144 and the ecological risk assessment of pesticides (Haith, 2010). However, they did not detail
145 the processes behind pesticide dissipation, and they could not identify a direct link between
146 ponds' efficiency and the pesticides' properties. Therefore, this paper investigates the
147 relationship between dissipation processes and the main characteristics of pesticides in
148 artificial ponds. Contrarily to the studies reviewed in this paper, this survey will evaluate
149 plentiful pesticides with contrasting physicochemical properties and present the mathematical
150 formulations of multiple processes evaluated from lab-scale to field-scale. These formulations
151 are useful for modeling that help to estimate artificial pond efficiency and to optimize their
152 implantation in agricultural areas.

153 **1.2 Processes governing the fate of pesticides in artificial ponds**

154 The dissipation of pesticides in artificial ponds results from the synergy between bio-
155 physicochemical processes detailed thereafter. Several studies have proved the possibility of
156 quantifying this dissipation and predicting the fate of pesticides by a black-box model
157 (Braschi et al., 2022). However, a black box approach does not explicit the origin of pesticide
158 dissipation and does not detail the contribution of each process since it only considers a single
159 dissipation rate k_{diss} (d^{-1}) (Yuan et al., 2020).

160 In this review, we will describe and present different estimation methods of processes behind
161 pesticide dissipation in artificial ponds. The form under which pesticides are introduced to the
162 artificial pond depends on how agricultural water is drained into the artificial pond. If the
163 watershed is drained with significant subsurface flow, pesticides are more likely to be
164 dissolved rather than particulate (Braschi et al., 2022; Le Cor et al., 2021). In contrast,
165 pesticides can be present in dissolved and particulate phases if runoff occurs. Gramlich et al.
166 (2018) assumed that subsurface drainage reduces erosion risk, while runoff may increase it.
167 Dissolved Pesticides (DP) in water bodies are widely studied in the literature, whereas few
168 studies focus on Particulate Pesticides (PP) (Climent et al., 2019). PP are transported within
169 the artificial pond along with TSS following the water flow. According to the hydraulic
170 regime of the artificial pond, PP can settle on the sediment bed or be suspended back to the
171 water. In contrast, DP can be bound to the solid surfaces available in the artificial pond
172 through sorption. Sorbed DP can be released back into water through desorption. DP can also
173 be transported to the atmosphere when volatilized or uptakes by the plants growing in the
174 artificial pond. On the other hand, pesticides can be a subject of transformation processes,
175 whose prevailing drivers are microorganisms, solar radiation, and hydrolysis.

176 Processes involved in pesticides dissipation in artificial ponds depend on many factors such as
177 the characteristics of the soil (i.e., Organic matter (OM), organic carbon (OC), and clay
178 content), water physicochemical properties (pH, temperature (T) and redox conditions),
179 vegetation, and the hydraulic conditions of the artificial pond (Papaevangelou et al., 2017;
180 Vymazal et al., 2015). The mitigation of pesticides in artificial ponds also depends on the
181 pesticide's physicochemical properties. The main physicochemical properties of a pesticide
182 include mobility, hydrophobicity, the solubility in water (S), and the pesticide half-life (DT_{50})

183 - the time required for the dissipation of 50% of the concerned substance (Gregoire et al.,
184 2009).

185 The properties of pesticides and the processes behind their dissipation in artificial ponds will
186 be detailed afterward.

187 **1.3 Hydraulics of artificial ponds**

188 The hydraulic performance is a key factor controlling the pesticide dissipation function of a
189 artificial pond. The hydraulic performance of artificial ponds includes lowering peak flow and
190 limiting sediment export from agricultural parcels to natural water bodies (Passeport et al.,
191 2010; Tournebize et al., 2013). Artificial ponds hydraulic parameters include (i) The hydro
192 period (Bojcevska et al., 2007; Convertino et al., 2013; Liu et al., 2019; Prochaska et al.,
193 2007) and (ii) The hydraulic residence time (HRT) of the water within the artificial pond (J.
194 Holland et al., 2004; Kjellin et al., 2007).

195 The hydraulic performance of artificial ponds depends on design parameters such as depth,
196 the layout of the inlet and outlet, and plant spacing. In this subsection, we will describe the
197 impact of design parameters on the hydraulic performance of artificial ponds.

198 Multi tracer experiments showed that shallow water depth was an essential factor enhancing
199 sorption and that pesticide load reduction was more significant in shallow artificial ponds than
200 in deep ones (Lange et al., 2011). A low water depth allows a uniform fluid distribution
201 compared to higher depths. The uniformity of the fluid flow decreases the hydraulic dead
202 zones and increases the effective volume, which is the artificial pond volume used for
203 pesticides dissipation (Rayen et al., 2019).

204 Globally, an artificial pond volume can be divided into three hydraulic zones (C. J. Martinez
205 et al., 2003; Werner et al., 2000):

- 206 1. The main channel with the preferential flow.
- 207 2. The mixing zone is a temporary storage zone showing exchanges with the main
208 channel.
- 209 3. The dead zone is more isolated from the main channel than the mixing zone.

210 Tracing studies showed that water is transported mainly by convection in the main channel,
211 while it is transported by dispersion in the isolated zones (mixed and dead zones). The HRT in
212 isolated zones is longer than in the main channel. Thus, isolated zones could enhance
213 dissipation processes such as sorption, settling, and degradation (Gaulhier et al., 2018). The
214 main channel is where short circuits in the water flow can occur, causing the transport of
215 water from the inlet to the outlet by preferential flow, which decreases the HRT and effective
216 volume of the pond (Vieira et al., 2018). Another controlling parameter of the artificial pond
217 hydraulic performance is the inlet/outlet design. Shilton et al. (2003) reported that positioning
218 the outlet in an isolated zone from the main channel avoids short-circuiting issues and
219 maintains the maximum hydraulic performance of artificial ponds. In addition, vegetation
220 density slows down water velocity and increases the HRT (Gu et al., 2017). Increasing the
221 hydraulic residence time (HRT) increases the contact time between pesticides and pond
222 substrates, giving pesticides more time to be sorbed, settled, or degraded.

223 A good hydraulic performance also guarantees an optimistic interception of agricultural water
224 by catching the highest concentrations of pesticides (during the post-application period) in
225 minimum water volumes through off/on stream artificial ponds (Passeport et al., 2013;
226 Passeport et al., 2010).

227

228 Usually, the risk of transfer of a pesticide is evaluated by the FOCUS models to decide
229 whether to be authorized for use or not. However, since pesticides are still transferred to the
230 environment after their application and their concentration in water resources than the RAC
231 (regulatory acceptable concentrations) (Gassmann et al., 2015; Halbach et al., 2021; Tang et
232 al., 2012), this theoretical study can be improved. Therefore, this work is based on a detailed
233 bibliographic analysis to present the required knowledge to understand the behavior of
234 pesticides after being applied on agricultural plots and intercepted by artificial ponds. This
235 knowledge is useful for developing a simple and robust conceptual model to simulate the fate
236 of pesticides in artificial ponds and then at the catchment scale, like the models developed for
237 nutrients (Hantush et al., 2013; Kalin et al., 2017; Walker Jr et al., 2011). Modeling can also
238 improve the understanding and hierarchization of the dissipation process and optimize its
239 implantation in agricultural areas. Therefore, the next section will describe the key processes
240 behind pesticide dissipation in artificial ponds and their mathematical formulations used in
241 modeling.

242 **2. Description and modeling of dissipation processes in artificial ponds**

243 Modeling is an efficient, comprehensive tool for assessing and setting up facilities to treat
244 contaminated environments (Warren et al., 2002). There are mainly two types of models that
245 describe pesticide fate in the environment, (i) Physical 3D and 2D models that simulate the
246 hydraulic performance of wetlands by considering water velocity profiles and the vegetation
247 impact of the flow (Carleton et al., 2010; Luo et al., 2016; Christopher J. Martinez et al.,
248 2003; Tsavdaris et al., 2013). (ii) Conceptual models that simulate the biogeochemistry and
249 the dissipation performance of the wetland for a large-scale application (Gobas et al., 2018;
250 Willkommen et al., 2018). The asset of this paragraph is to identify and describe processes
251 behind pesticide dissipation in artificial ponds, along with their mathematical formulations
252 established as part of conceptual models.

253 Our literature investigation showed that researchers had widely discussed the fate of
254 pesticides in artificial ponds. Although, most studies are generally limited to one pesticide or
255 one specific process evaluation and are rarely supported by mathematical formulae. However,
256 we assume that mathematical formulations allow the integration of processes into a
257 computational model to quantify and predict the fate of contaminants in artificial ponds. The
258 originality of this review lies in the description of the main processes influencing the behavior
259 of multiple pesticides in artificial ponds, namely transport, sorption, and transformation
260 processes. The evaluation and formulation of these processes are necessary to develop
261 predictive models for pesticides dissipation in artificial ponds (Papaevangelou et al., 2017;
262 Saaristo et al., 2018). In this section, we will investigate the behavior of multiple pesticides
263 with contrasting physicochemical properties and describe different methods currently
264 employed to evaluate and formulate the processes controlling dissipation in artificial ponds.

265 **2.1 Transport of pesticides**

266 Pesticides reaching artificial ponds can be dissolved in water or associated with dissolved
267 organic carbon (DOC). They can also be bound to the organic fraction of suspended matter
268 (i.e., particulate organic carbon (POC)), featuring pesticides in a particulate form. PP are
269 carried along with suspended matter throughout the pond artificial pond due to water
270 movements. PP can be deposited on the sediment bed due to settling and released back to
271 water through resuspension. Settling and resuspension are transport processes that strongly
272 depend on the particle size and the strength of the water flow in pond artificial ponds.

273 Pesticides can also be the subject of advection, dispersion, and diffusion processes within the
274 water column and pore water of the sediment layer. According to the Hjulstrom curve, the
275 water velocity in the artificial pond can determine which transport process is prevailing
276 (Hjulstrom, 1935).

277 **2.1.1 Settling**

278 Settling is a physical transport process through which suspended particles in water settle on
279 the sediment bed of artificial ponds. Settling depends on the particle size of pesticides, water
280 velocity and turbulence, temperature, and water column depth (Kadlec et al., 2008).

281 Settling has been efficient in the water-quality improvement function of artificial ponds,
282 especially in lower flows, since suspended particles, on which pesticides may be attached, can
283 leave the water column while settling on the sediment bed (O'Geen et al., 2010). Settling is
284 enhanced when vegetation covers a significant surface of the artificial pond; emergent
285 macrophytes slow down water flow and amplify settling (Vymazal, 2013). In addition, stems
286 and leaves in the water column obstruct water flow and dissipate the energy required to
287 support particle suspension (Schmid et al., 2005). Rose et al. (2008) pond (0.021 h^{-1}). A small
288 water depth was also reported that the settling rate in a vegetated artificial pond (0.042 h^{-1})
289 was double the rate estimated in a non-vegetated artificial to promote the settling process; it
290 fosters plant growth and shortens the settling distance for particles (Lange et al., 2011). It is
291 valid for water depths between 20 cm and 50 cm (Braskerud, 2002). Pesticides associated
292 with fine suspended particles may hardly be retained in ponds since fine particles (clay and
293 silts) are highly transported across preferential pathways where the HRT is low (see section
294 1.3) (Maynard, 2009). The settling of PP is more likely to occur in hydraulically isolated
295 zones with lower water flow and higher HRT (Gauillier et al., 2020; Passeport et al., 2010).
296 Ponds with the highest settling rate are the ones receiving agricultural surface runoff (Fiener
297 et al., 2003; Zhang et al., 2011), where the intercepted agricultural water is heavily loaded
298 with coarser ($>200 \mu\text{m}$) particles from the fields (Li et al., 2017). It is due to the higher TSS
299 load from erosion that is less available in artificial ponds collecting subsurface drainage water
300 (Koskiaho, 2003; Maillard et al., 2011). It was also suggested that coarse particles settle
301 before the water is distributed across pond compartments, unlike fine particles requiring a
302 longer time to settle. As a result, fine particles become more concentrated downstream of the
303 pond (Gan et al., 2005) because gravity induces the settling of heavy particles ($>36 \mu\text{m}$) (i.e.,
304 sand) more than lighter particles ($<2 \mu\text{m}$) (i.e., clay and OM) (Fiener et al., 2003). Several
305 studies suggested that a large proportion of highly hydrophobic organic substances in artificial
306 ponds were retained by settling pesticide-laden solids (sorbed on suspended solids) (Luo et
307 al., 2009; Matamoros et al., 2006; Rose et al., 2008). Therefore, settling may not be sufficient
308 to trap pesticides with a stronger affinity to fine suspended particles in ponds. Consequently, a
309 higher HRT and vegetation are required to increase the settling of fine PP (Budd et al., 2009,
310 2011; Fiener et al., 2003).

311 Settling was commonly integrated with an accumulation rate expressed empirically in
312 pesticide dissipation models. The empirical formulations of settling rates can be found in
313 Table A.1. The first formulae (i) estimates the settling rate v_s according to Stokes equation for
314 laminar conditions (Reynolds number $Re < 500$). Fiener et al. (2003) computed the settling
315 rates of particles of different sizes in a vegetated wetland. It was observed that particles >63
316 μm (i.e., sand) were fully sedimented, whereas particles $2 \mu\text{m} < x < 63 \mu\text{m}$ (i.e., silt) were
317 moderately sedimented (up to 26%). In contrast, only 4% of fine particles $<2 \mu\text{m}$ (i.e., clay)

318 settled on the sediment bed. It was also noticed that settling primarily occurred on the side-
319 slopes with small Re . The second formulation (ii) suggested by Kadlec et al. (2008) allows
320 calculating the settling rate in both laminar and turbulent flow based on a drag coefficient C_D
321 that depends on Re . Formula (ii) assumes that settling results from density difference between
322 suspended particles and water, so it expresses the rate of a single and isolated spherical
323 particle. The limit of the first (i) and second method (ii) are the required parameters since in
324 artificial ponds, neither the density nor the particle size is known. In addition, particles are not
325 spheres or discs and can agglomerate to a larger size. The formula (iii) estimates the settling
326 rate of suspended particles in wetlands (Hawley et al., 2014). This formulation required more
327 parameters such as the water kinematic viscosity since it is adapted for also non-spherical
328 particles. However, it was often applied to large size particles.

329 Overall, the formula (i) could be sufficient and easy to use for a simple simulation of PP
330 settling in laminar flows. If the flow is turbulent with a known Re , formula (ii) would be more
331 suitable for its accuracy. For formula (ii), particle sizes are often approximated according to
332 their nature (i.e., clay, silt, sand, and OM). Otherwise, the settling rate could be calibrated if
333 the required data is available. But still, the majority of settling studies are done on nutrients,
334 and little is known about pesticides (Hantush et al., 2013; Kalin et al., 2017; Walker Jr et al.,
335 2011). Regarding behavior, settling seems to be a key process of pesticide accumulation in
336 pond artificial ponds alimented by surface runoff in erosive areas, especially for hydrophobic
337 pesticides and coarse PP.

338 2.1.2 Resuspension

339 Resuspension is the opposite process of settling, through which PP are transported from the
340 sediment bed to water. Resuspension depends mainly on the water flow and the water depth in
341 artificial ponds.

342 Resuspension is likely to occur when water turbulence is important (Siobhan Fennessy et al.,
343 1994), and high water flow can remobilize settled PP back to the water (Collins, 2004;
344 Jamieson et al., 2005; Vallet, 2011; Wilkes et al., 2009). It was proved that resuspension was
345 highly affected by the type and density of vegetation in artificial ponds (Liu et al., 2019;
346 Schulz et al., 2001). Braskerud (2001) presumed that an increase of 30% in a vegetative
347 cover, initially less than 20%, could reduce resuspension rates from 40% up to 100%. Rose et
348 al. (2008) showed that the resuspension rate was divided in half by switching to a vegetated
349 artificial pond. Some studies suggested that deeper FWS wetlands enhanced settling and
350 burial rather than resuspension (Brueske et al., 1994; Budd et al., 2011). Resuspension is
351 likely to occur in the main channel where the water velocity is fast (see section 1.3) (Gaullier
352 et al., 2020) and is mainly driven by wave action generated by the wind (Hawley et al., 2014).
353 Other agitation factors may be responsible for particle resuspension, such as wildlife
354 movements within the artificial pond (i.e., bioturbation), although quantitative studies on this
355 topic remain scarce. Resuspension was significantly less considered in the literature than
356 settling. However, resuspension occurred with a slow rate of remobilization ($v_r \sim 10^{-6}$ mm/s)
357 (Hawley et al., 2014; Sharifi et al., 2013), while Rijn (1984) suggested that the resuspension
358 rate v_r reached the quarter of the settling rate v_s .

359 Globally, significantly lower concentrations of PP measured at the outlet of wetlands than at
360 the inlet suggests that the contribution of resuspension in pesticides behavior might be
361 negligible (Budd et al., 2009; Cryder et al., 2021; Supowit et al., 2016), also resuspension is
362 more dominant in systems with stronger water flows (i.e., rivers) rather than artificial ponds.

363 In addition, high water flow events are usually short and have a long return period, which
364 decreases the occurrence of resuspension.

365 **2.1.3 Advection, dispersion and diffusion**

366 Among the processes behind pesticide transport within the artificial pond are advection,
367 dispersion including diffusion. Advection carries pesticides in the same direction as the water
368 flow. Dispersion results from heterogeneities in flow patterns and is a macroscopic process. In
369 contrast, diffusion is a microscopic movement of pesticide molecules due to a random
370 molecular motion (i.e., Brownian motion) (Hamdi, 2007). Dispersion and diffusion processes
371 result from a concentration gradient of pesticides in artificial pond water and carry pesticides
372 from highly concentrated areas to low concentrations. Advection, dispersion, and diffusion
373 allow the homogenization of DP concentrations in the artificial pond.

374 Gaullier et al. (2019) showed that water transport is mainly governed by advection at a high
375 flow rate. While at low flow rates, both water transport and pesticide mitigation are governed
376 by hydrodynamic dispersion, including molecular diffusion, resulting in a longer contact time
377 between pesticides and substrate. Without agitation, pesticide transport is only influenced by
378 diffusion, while agitation enhances advection and dispersion (Gaullier et al., 2018). Thus,
379 advection/dispersion is more important in the main channel, while diffusion is more likely to
380 occur in the dead zones (section 1.3). Moreover, diffusion is slower in the sediment layer pore
381 water because of its tortuosity and the increased length of the water pathway between pores
382 (Beulke et al., 2004). When the hydraulic regime is laminar, the diffusion is on the molecular
383 scale, with a thermal diffusion of the order $1.4 \times 10^{-7} \text{ m}^2 \text{ s}^{-1}$ in pure water. In river water
384 systems, the water is always turbulent. Thus, the effective diffusion is several orders of
385 magnitude greater. Still, the given thermal diffusion is considered in wetlands, where water
386 disperses horizontally, and the flow rate is lower than in rivers (Damalas et al., 2011). For
387 pesticide molecules, the diffusion coefficients usually range from 10^{-11} to $10^{-10} \text{ m}^2/\text{s}$
388 (Kipkoech, 2019). Dispersion, including diffusion, also depend on the internal structure of the
389 artificial pond. For instance, the dispersion was proved to be greater in artificial ponds with a
390 larger width (Gaullier et al., 2019), while macrophytes enhance diffusion and the hydraulic
391 performance of the artificial pond (J. F. Holland et al., 2004).

392 In a pesticide transport equation, advection is expressed by the term $-v \frac{\partial C_w}{\partial x}$ and dispersion is
393 expressed by $D_L \frac{\partial^2 C_w}{\partial x^2}$. Where v [$\text{L} \cdot \text{T}^{-1}$] is the water velocity; C_w [$\text{M} \cdot \text{L}^{-3}$] is the pesticide
394 concentration in water; D_L [$\text{L}^2 \cdot \text{T}^{-1}$] is the hydrodynamic dispersion coefficient; with
395 $D_L = D_e + \alpha_L v$, where D_e the effective diffusion coefficient, and α_L [L] the dispersivity.

396

397 **2.2 Transfer of pesticides**

398 Once pesticides reach the artificial pond, they are distributed among water, sediments, TSS,
399 and vegetation are transferred among artificial pond compartments. DP can be exchanged
400 between the water column and the solid compartments due to “Sorption” and the atmosphere
401 during “Volatilization”. Also, DP can become PP when sorbed to TSS.

402 **2.2.1 Sorption process**

403 Sorption was commonly reported as an important process in the fate of pesticides in artificial
404 ponds (Cryder et al., 2021; Fernández-Pascual et al., 2020; Hand et al., 2001). In the
405 following two subsections, we will first present the sorption theory and analyze study cases of
406 pesticide sorption in artificial ponds. This review will focus on sorption information directly
407 related to pesticides' fate in artificial ponds; more detailed knowledge of sorption can be

408 found in the literature (Huang et al., 2003; M. McBride, 1994; Pignatello, 1989; Sposito,
409 2008)

410 2.2.1.1 Sorption theory

411 Physically, *sorption* refers to all exchange processes between a dissolved substance and solid
412 matter in a solution. Among these exchanges, adsorption and desorption remain the most
413 studied for pesticide fate in the environment. Adsorption is a physical transfer process of
414 retaining dissolved substances in water on solid surfaces. The dissolved substances are the
415 “sorbate,” and the solid surface to which they are bound is called “sorbent” or “substrate.”
416 *Desorption* is when the substrate releases the sorbate and returns to water (Sposito, 2008).
417 From now on, the term “sorption” will be used to refer to adsorption plus desorption.

418 Most of the sorption studies are conducted in the laboratory and adapted from OECD 106
419 guidelines for individual molecules, in which sorption experiments are performed using a
420 batch equilibration technique (OCDE, 2000). The retention and release of the sorbate on a
421 substrate at various concentrations and a constant temperature are described by a curve called
422 “adsorption isotherm” and “desorption isotherm,” respectively. The isotherms represent the
423 relationship $C_{se}=f(C_{we})$ during adsorption and desorption equilibrium, where C_{se} ($\mu\text{g}\cdot\text{g}^{-1}$) and
424 C_{we} ($\mu\text{g}\cdot\text{L}^{-1}$) are the sorbate concentration in the substrate and the sorbate concentration in
425 the solution at the equilibrium state, respectively.

426 Based on the thermodynamic equilibria theory that assumes complete reversibility of the
427 chemical reactions, the adsorption and desorption isotherms should be the same (Strawn et al.,
428 1999). However, sorption mechanisms are driven by different kinetically controlled reactions
429 or physical phenomena with a large variability of reaction times going from seconds to years,
430 emphasizing the kinetic character of sorption (Beulke et al., 2004; Caceres-Jensen et al., 2019;
431 Cara et al., 2021; Kaur et al., 2018; Mamy et al., 2007; Sparks, 2000; Villaverde et al., 2009;
432 Wu et al., 2017).

433 A difference between adsorption and desorption kinetic rates to reach the equilibrium is
434 translated by a mismatch between the adsorption and desorption isotherms curves. This
435 mismatch is called “pseudo-hysteresis” or “kinetic hysteresis” (Strawn et al., 1999), also
436 referred to as an “apparent irreversibility” (Bladel et al., 1967; M. B. McBride, 1994). It is
437 important to clarify that the term “irreversibility” does not mean that the sorbate will not
438 desorb after its adsorption. Still, it will take a long time to reach the equilibrium state
439 (Limousin et al., 2007). In other terms, hysteresis means that desorption requires a more
440 significant amount of energy than needed by adsorption (Bowman et al., 1985; Koskinen et
441 al., 1979; Wu et al., 1988), which could be caused by experimental artifacts or transformation
442 of the sorbent (Pignatello et al., 1996).

443 The evaluation of sorption requires the determination of its equilibrium and kinetic properties
444 (Douven et al., 2015). Sorption kinetics are studied with the most applied chemisorption
445 models: Pseudo-first-order (PFO), pseudo-second-order (PSO), and Elovich models (Azizian,
446 2004; Ho, 2006; Ho et al., 1998, 1999; Yuh-Shan, 2004), to describe the temporal evolution
447 of the sorbed amount of the sorbate. At the same time, sorption equilibrium is commonly
448 modeled by the Freundlich and Langmuir isotherm models (Azizian et al., 2018; Foo et al.,
449 2010; McKay, 1995). As the equilibrium of kinetic sorption is reached, isotherm models are
450 applied. The corresponding equations of kinetic and isotherm models can be found in Table
451 A.2.

452 2.2.1.2 Sorption of pesticides in artificial ponds

453 In the case of artificial ponds, the sorbates are dissolved pesticides (DP), and substrates are
 454 total suspended solids (TSS) with organic and inorganic compounds, sediments, and
 455 vegetation (Gregoire et al., 2009; Sahin et al., 2020), which represent development support for
 456 OM and biofilm. The Freundlich and the linear model are commonly used isotherm models to
 457 describe pesticide sorption on artificial pond substrates (Liu et al., 2019; Takagi et al., 2012;
 458 Watanabe et al., 2006; Zhang et al., 2018). Pesticide adsorption is widely described by the
 459 adsorption coefficient normalized to substrate organic carbon content K_{foc} and K_{oc} for the
 460 linear isotherm:

$$461 K_{foc} = \frac{K_{fads}}{OC} \quad (1)$$

$$462 K_{oc} = K_d/OC \quad (2)$$

463 Where OC (%) is the organic carbon content of the pond substrates, K_{fads} is the Freundlich
 464 adsorption coefficient, and K_d is the distribution coefficient (Table A.2).

465 Vallée (2015) and Passeport et al. (2011b) studied the sorption of 12 pesticides with
 466 contrasting physicochemical properties on pond substrates (i.e., sediments, vegetation, and
 467 TSS). Sorption coefficients can be found in Table A.3. Based on the K_{fads} values,
 468 epoxiconazole was the highly adsorbed pesticide, followed by prochloraz. Epoxiconazole and
 469 prochloraz have a K_{foc} range of $702\text{--}1962 \text{ mg}^{1-n}_f \cdot \text{L}^n_f \cdot \text{kg}^{-1}$ and $1222\text{--}8654 \text{ mg}^{1-n}_f \cdot \text{L}^n_f \cdot \text{kg}^{-1}$,
 470 respectively (Lewis et al., 2016), which, according to the Soil Survey and Land Research
 471 Centre (SSLRC) soil persistence classification system, means that they are lowly mobile
 472 molecules (PSD, 2005). In contrast, the lowest sorption concerned isoproturon, with the
 473 lowest K_{foc} values $55\text{--}60 \text{ mg}^{1-n}_f \cdot \text{L}^n_f \cdot \text{kg}^{-1}$ (Vallée, 2015), followed by MCPA. The low
 474 adsorption of MCPA could be due to its anionic form, inducing an electrostatic repulsion of
 475 negatively charged particles (i.e., clay and OM), as supported by Thorstensen et al. (2001).
 476 Likewise, Poissant et al. (2008) and Wu et al. (2004) reported that pesticides with higher K_{oc}
 477 values ($K_{oc} > 500 \text{ L} \cdot \text{kg}^{-1}$) are most likely to be sorbed to pond solid particles, agreeing with
 478 previous findings (Papp, 2009; Saripalli, 1997). The low solubility of epoxiconazole and
 479 prochloraz also favored their retention on pond substrates. The literature widely reported that
 480 low solubility enhances pesticide adsorption (Blachnio et al., 2020; Brusseau et al., 2019; Kah
 481 et al., 2006). Sorption also depends on the hydrophobicity of the pesticide expressed by the
 482 octanol-water partition coefficient K_{ow} . Hijosa-Valsero et al. (2016) analyzed fifty-nine
 483 compounds in water and sediment samples. They found that herbicides in the sediments had
 484 an average $S=194 \text{ mg} \cdot \text{L}^{-1}$ and a $\log K_{ow} = 3.7$, while herbicides in the water phase were highly
 485 soluble $S = 1395 \text{ mg} \cdot \text{L}^{-1}$ with an average $\log K_{ow} = 2.24$. These results support that lowly
 486 soluble, and hydrophobic pesticides ($\log K_{ow} > 3$) are likely to be sorbed on pond sediments,
 487 which agrees with O'Geen et al. (2010), that observed a 50% reduction in pesticide
 488 concentration with $\log K_{ow} > 4.2$. Likewise, Tang et al. (2017) found that adsorption rates of
 489 hydrophobic pesticides were significantly higher on the artificial pond vegetation ($5.52\text{--}8.84$
 490 $\text{g m}^{-2} \text{ d}^{-1}$) than hydrophilic pesticides ($\log K_{ow} < 3$). These results suggest that sorption may
 491 be the primary driving process of removing hydrophobic pesticides from the artificial pond
 492 water.

493 On the other hand, vegetation, biofilm, and the OM and clay content of the substrate have a
 494 significant role in pesticide sorption in artificial ponds. Barchanska et al. (2019) observed that
 495 the artificial pond remediation of pesticides due to adsorption on vegetation reached 30%. An

496 extensive vegetation cover can enhance adsorption by increasing the HRT [Gaullier et al.](#)
497 [\(2020\)](#). A higher HRT increases the contact time between pesticides and pond substrates,
498 favoring adsorption and generally decreasing desorption, as observed by [\(Mamy et al., 2007;](#)
499 [Olvera-Velona et al., 2008\)](#).

500 [Passeport et al. \(2011b\)](#) and [Vallée \(2015\)](#) observed higher K_{fads} on vegetation than in
501 sediments (Table A.3), showing greater retention on OC-rich substrates. [Moore et al. \(2007\)](#)
502 and [Sahin et al. \(2020\)](#) also proved that diazinon preferentially sorbed on vegetation surfaces
503 rather than sediments. In like manner, [Rogers et al. \(2009\)](#) showed that the lowly mobile
504 chlorpyrifos ($K_{oc} = 5509 \text{ L.kg}^{-1}$ [\(Lewis et al., 2016\)](#)) adsorption was ten times higher on
505 vegetation than on sediments, and several other papers demonstrated that adsorption is more
506 dominant in OC-rich substrates, including vegetation ([Guo et al., 2000; Mahabali et al., 2014;](#)
507 [Moore et al., 2007; Mugni et al., 2011\)](#). Biofilm was also proven to enhance adsorption for
508 certain pesticides ([Agudelo et al., 2012; Beecraft et al., 2021\)](#).

509 Desorption was also evaluated and was reported to be less dominant in sediments with high
510 clay and OC content, especially for lowly soluble pesticides ([Gramlich et al., 2018; Olvera-](#)
511 [Velona et al., 2008; Passeport et al., 2011b; Song et al., 2021; Vallée, 2015\)](#). In comparison,
512 greater desorption was observed on some highly soluble pesticides ([Olvera-Velona et al.,](#)
513 [2008\)](#). Hence, hysteresis was generally observed on OC and clay-rich substrates from which
514 hydrophobic and lowly soluble pesticides are hardly desorbed ([Hand et al., 2001; Li et al.,](#)
515 [2003; Vagi et al., 2010\)](#).

516 In addition, kinetic sorption rates showed that desorption was slower than adsorption for six
517 evaluated pesticides, except for picloram. The reviewed kinetic rates of adsorption and
518 desorption can be found in Table A.4. The picloram exception can be explained because its
519 sorption was evaluated in lab conditions. Since, in the experimental sample, there was no
520 consideration of other processes that may compete with desorption (i.e., transformation). The
521 interaction between desorption and transformation was also observed by [Sørensen et al.](#)
522 [\(2006\)](#), noticing that slower desorption of MCPA (coincided with higher microbial
523 transformation. In fact, unlike laboratory-batch sorption observations, sorption and
524 transformation processes co-occur under field conditions, favoring the occurrence of
525 hysteresis in the presence of other competitive processes like transformation and diffusion
526 into sediment layer micropores ([Mamy et al., 2007; Paszko et al., 2016; Takagi et al., 2012;](#)
527 [Uchimiya et al., 2012\)](#). In contrast, advection/dispersion enhance sorption by facilitating the
528 access of DP to the pond substrates ([Gaullier et al., 2018\)](#).

529 Overall, we assume that both adsorption and desorption depend on the clay and OC content of
530 pond substrates and the properties of pesticides. Adsorption is more dominant for
531 hydrophobic, lowly mobile, and lowly soluble pesticides, and desorption hysteresis is more
532 accentuated in clayey and OC-rich pond substrates, especially for highly soluble pesticides.

533 **2.2.2 Volatilization**

534 Volatilization is a process through which pesticides are transferred outside the pond, from the
535 water to the atmosphere. This non-degradative removal process strongly depends on the
536 pesticide molecule's vapor pressure.

537 There are two regimes controlling volatilization: the atmospherically controlled regime and
538 the regime controlled by water properties. Most pesticides (80-90%) belong to the
539 atmospherically controlled regime, where the gas exchange between the water and the

540 atmosphere is only sensitive to the parameterization of atmospheric turbulence, and the
 541 majority of pesticides belong to it; therefore, we will focus on the atmospherically controlled
 542 regime. In the atmospherically controlled regime, at a very low dimensionless Henry constant
 543 ($<10^{-7}$) (equivalent to a vapor pressure $vp <10^{-5}$ Pa), the volatilization rate is so slow that the
 544 volatilization half-life is tens to hundreds of years. In that case, volatilization becomes
 545 insignificant compared to other processes (Jacobs et al., 2012). In that respect, Höllrigl-Rosta
 546 (2017) classified of volatilization according to the vapor pressure of the pesticide molecule
 547 (Table A.5).

548 In modeling, volatilization is often described by a first-order kinetic equation (Barchanska et
 549 al., 2019; Comoretto et al., 2008; Gobas et al., 2018; Inao et al., 1999; Singh et al., 1991;
 550 Watanabe et al., 2006) :

$$551 \quad \frac{dMw}{dt} = -k_v * Mw \quad (3)$$

552 Where Mw is the mass of the DP, and k_v (d^{-1}) is the pesticide rate constant of volatilization to
 553 the atmosphere. Volatilization can also be represented by its corresponding half-life DT_{50v} (d):

$$554 \quad DT_{50v} = \frac{\ln(2)}{k_v} \quad (4)$$

555 Since this transfer process depends on other factors, the volatilization rate may be expressed
 556 empirically as a function of both pesticide properties and climate conditions. Table A.6 are
 557 simplified form of the rate of volatilization k_v used in descriptive models of the fate of
 558 pesticides. In other studies, there are many complicated formulations (Bunyakan et al., 2006;
 559 Comoretto et al., 2008), in which k_v depends on diffusion coefficients, air-water side, water-
 560 air side mass transfer coefficient, and friction. However, they are usually left for
 561 volatilization-specific studies and do not model the fate of pesticides in artificial ponds.

562 A bibliographic review of calculated volatilization half-lives DT_{50v} (d) of certain pesticides
 563 can be found in Table A.7. For most pesticides in artificial ponds, the vapor pressure is lower
 564 than 10^{-5} Pa, which enables to neglect the contribution of volatilization to the artificial pond
 565 dissipation function according to the classification in Table A.5. However, it was
 566 demonstrated that some pesticides were efficiently removed from water by volatilization
 567 (Wang et al., 2014; Wang et al., 2019). In an aquatic microcosm under semi-field conditions,
 568 Laabs et al. (2007) showed that volatilization was a primary reason for high pesticide losses
 569 from the water shortly after application, especially for non-polar pesticides (i.e., endosulfan,
 570 chlorpyrifos, and trifluralin). Similarly, Comoretto et al. (2008) reported that oxadiazon had a
 571 DT_{50v} of 23 days in a rice field, making it more vulnerable to volatilization when compared to
 572 stable MPCA and pretilachlor. k_v rates were either calculated using the empirical formulas in
 573 Table A.6 or estimated in laboratory experiments. Empirical formulations of k_v show that
 574 volatilization rates are mainly dependent on water depth, a mass water-air transfer coefficient
 575 that is a function of the wind speed, the Henry constant (or vapor pressure and solubility), and
 576 the molecular weight of the pesticide. Pesticides with a vp lower than 10^{-5} require years to be
 577 volatilized according to volatilization half-lives in Table A.7, making them virtually non-
 578 volatile. In contrast, other pesticides such as alphamethrin, molinate, endosulfan, and dieldrin
 579 need a few days to volatilize, while oxadiazon requires more days to volatilize. With careful
 580 consideration of the vp of these substances, we assume that these assumptions agree with the
 581 classification in Table A.5. These observations showed that most pesticides are non-volatile

582 due to their low vapor pressure. However, there are still some for which volatilization takes
583 part in their dissipation in artificial ponds.

584 **2.3 Transformation of pesticides**

585 In addition to the transport and transfer processes described in sections 2.1 & 2.2, pesticides
586 face other processes that transform their original chemical structure. They are also designated
587 as “degradative” processes when attempting to degrade pesticides to non-organic products
588 (Cessna et al., 2015; Imfeld et al., 2021). The importance of transformation processes in the
589 artificial pond dissipation efficiency was widely proved. Pesticides can be transformed into
590 less toxic products or, in the ideal case, wholly degraded to simple mineral products under the
591 action of the chemical, photochemical, and biological processes (Tournebize et al., 2017).
592 This section describes the main biotic and abiotic transformation processes to which
593 pesticides are exposed in the different compartments of the artificial pond.

594 **2.3.1 Biotransformation**

595 Biotransformation is a process by which organic substances are transformed by
596 microorganisms (bacteria, archaea, and fungi) into chemical substances called metabolites,
597 resulting in some cases in their complete mineralization (Garcia-Rodríguez et al., 2014). If the
598 microbial transformation is complete, it becomes a transformation, and pesticides are
599 mineralized to CO₂ and CH₄. Otherwise, co-metabolism occurs, during which microorganisms
600 degrade OM substrates, the primary source of nutrients of the bacteria, primarily to produce
601 enzymes that will break down sequentially the pesticide molecule, representing a secondary
602 source of nutrients of the bacteria (Arcangeli et al., 1997), resulting in a metabolite.

603 Recently, in a artificial pond system context, some metabolites were less toxic than their
604 parent pesticides (Kang et al., 2020). However, it remains unknown if other metabolites are
605 less or more toxic than the mother compounds. Nonetheless, biotransformation has been
606 extensively shown as a significant removal process for micro-organic pollutants (Hijosa-
607 Valsero et al., 2010; Langergraber, 2008; Llorens et al., 2011; Lyu et al., 2018; Samsó et al.,
608 2013; Yang et al., 2017; Zhang et al., 2017).

609 Biotransformation occurs in both aerobic and anaerobic conditions. Aerobic
610 biotransformation occurs in oxic conditions, mainly near the plant roots or the water surface,
611 where the amount of dissolved oxygen is high enough. Biotransformation is anaerobic in deep
612 water and sediment depths with generally dissolved oxygen concentrations below 0.5 mg.L⁻¹
613 (U.S.G.S, 2020). The rhizosphere can also be considered as an aerobic zone since oxygen can
614 be transported from the plant body to the roots (Maltais-Landry et al., 2009), but researchers
615 showed that the amount of oxygen released by the plant was too little to be considered in
616 artificial pond fate models (Bezbaruah et al., 2005; Llorens et al., 2011). Artificial ponds are
617 mostly aerobic in the water column surface, with anaerobic conditions occurring at the bottom
618 sediment layer and deep water (Vymazal et al., 2015). The sediment layer was proved to be
619 characterized by a sharp decrease in oxygen (Martínez-Lavanchy et al., 2015; Stottmeister et
620 al., 2003; Thomas et al., 2011; Zhang et al., 2010). Karpuzcu et al. (2013), Paszko et al.
621 (2016) and (Mulligan et al., 2016); Paszko et al. (2016) demonstrated that anaerobic
622 conditions would result in lower biotransformation rates than under aerobic conditions. It
623 agrees with the recent study of commonly used herbicides in which Droz et al. (2021)
624 indicated that acetochlor and S-metolachlor transformation prevailed in the water phase and
625 seemed insignificant in the sediment.

626 Microbial transformation also depends on temperature and radiation (Burrows et al., 2002),
627 pH, OM content, and microbial population (Vink et al., 1997). Sahin et al. (2020) reported
628 that humic substances increase biotransformation rates since they can carry microorganisms to
629 the pesticide. This effect has been widely investigated (Cai et al., 2017; Ortega-Calvo et al.,
630 1998; Smith et al., 2009; Tejeda-Agredano et al., 2014), mainly for hydrophobic pesticides.
631 For instance, atrazine was highly biodegraded when sorbed onto clay-humic acid complexes
632 (Besse-Hoggan et al., 2009). Likewise, Rong et al. (2019) found that the interfacial reactions
633 on clay surfaces increased the biotransformation rates of methyl parathion. The exposure
634 history to pesticides of artificial ponds also influences biotransformation. Johnson et al.
635 (2004) demonstrated that sites with a higher pesticide exposure history had higher
636 biotransformation rates due to microbial acclimation and microbial community adaptation.

637 The water level is another controlling factor of biotransformation in the sediment
638 compartment of artificial ponds. Ma et al. (2018) speculated that a decline in the water level
639 enhances microbial growth and activity and would modify the structure and function of
640 microbial communities. The study also emphasized that a water volume drop-off would
641 enhance aerobic conditions, induce plant succession to the artificial pond center, and stimulate
642 OM accumulation and mineralization.

643 The aging of sediments also intensifies the sequestration of PP after their settling, thus
644 decreasing their bioavailability and hindering microbial transformation (Ahmad et al., 2004).
645 Budd et al. (2011) proved that once pyrethroids were settled, they were stable in dry and
646 aerobic sediments between irrigation seasons. They were moderately persistent under flooded
647 and anaerobic conditions, with a DT_{50} between 106-353 days. This agrees with the
648 observation of Cryder et al. (2021), showing that adsorption and subsequent settling were the
649 dominant processes governing the fate of pyrethroids in ponds instead of biotransformation.

650 Vegetation has been widely recognized as an important stimulator of biotransformation
651 (Moore et al., 2007; Rai, 2009; Stang et al., 2016; Sur et al., 2003; Wang et al., 2017). For
652 instance, chlorpyrifos removal rate reached $0.1 \text{ mg}\cdot\text{m}^{-1}\cdot\text{day}^{-1}$ due to biotransformation in
653 vegetated mesocosms (Tang et al., 2019). Submerged stems and leaves of macrophytes are an
654 essential habitat for biofilms (Brix, 2003; C. Zhang et al., 2021). These biofilms, along with
655 those on all other immersed solid surfaces in the artificial pond system - including dead
656 macrophytes – are stimulators of microbial processing in artificial ponds (Brix, 2003; Dash et
657 al., 2020; Lv et al., 2017). Several kinds of phototrophic bacteria, algae, and biofilms in the
658 water-sediment interface and on the surface of macrophytes were proved to be important
659 agents of pesticides transformation in artificial ponds (Katagi et al., 2016). Although,
660 biotransformation in the biofilm can be limited sometimes due to the slow kinetics of mass
661 transfer or enzymatic reactions (Chen et al., 2015).

662 In modeling, biotransformation is often described by a first-order kinetic model (Beringer et
663 al., 2021; Green et al., 1990; Mamy et al., 2007; Pignatello, 1999; Scow et al., 1996; Torabi et
664 al., 2020). Where k_{bio} (d^{-1}) is the biotransformation kinetic constant in the compartment in
665 question. Biotransformation can be represented either by the kinetic constant k_{bio} (d^{-1}) or by
666 the corresponding half-life $DT_{50\text{bio}}$ (d) (see Eq.(3) and Eq.(4)).

667 Several studies managed to estimate the biotransformation rate constants in artificial ponds in
668 laboratory conditions, through incubation manipulations. Incubation experiments aims to
669 calculate pesticide concentrations in the different compartments of a contaminated sample

670 (water and sediments). Then, the concentrations are fitted to a first-order kinetic decay model
671 to estimate the rate constants k_{bio} and half-lives $DT_{50\text{bio}}$. Table A.8 presents a literature review
672 of estimated biotransformation half-lives $DT_{50\text{bio}}$.

673 The values of biotransformation half-lives (Table A.8) show that hydrophobic pesticides
674 pretilachlor and fenthion were rapidly biodegraded in sediments; we suggest that it may be
675 related to the fact that they were mainly sorbed on substrates easily accessible to
676 microorganisms. Pretilachlor was more rapidly degraded in water ($DT_{50\text{bio,w}} = 9.8$ d). Even
677 though being hydrophobic and lowly mobile ($K_{\text{oc}} = 5509$ L.kg⁻¹), chlorpyrifos was more
678 persistent in sediments than in water columns, and was favorable to aerobic conditions
679 (O'Geen et al., 2010). Comparing the behavior of pretilachlor, fenthion, and chlorpyrifos, that
680 hydrophobicity, and low mobility may not necessarily favor biotransformation in sediments
681 rather than water. On the other hand, biotransformation in sediments seemed relatively slower
682 for more mobile pesticides (i.e., 2, 4-D, clothianidin, acetochlor, metazachlor, and S-
683 metolachlor), except for the MCPA. We suggest that MCPA was not expected to sorb on TSS
684 and sediments due to its low K_{oc} . Its transformation might be more favorable under aerobic
685 conditions (on the water surface) (PubChem, 2021), explaining its short half-life in the water
686 phase. S-metolachlor was moderately persistent in both water and sediments; this may be
687 explained by its complex chemical structure that forbids access to enzymes and prevents its
688 transformation under aerobic and anaerobic conditions (Droz et al., 2021). Finally, $DT_{50\text{bio}}$
689 values showed that biotransformation was faster in oxic areas, except for acetochlor ($DT_{50\text{bio,w}}$
690 = 47 d) and clothianidin ($DT_{50\text{bio,w}} = 27.2$ d) that were rapidly transformed under anaerobic
691 conditions. Although, it is not possible to infer any direct link between the OC content and the
692 biotransformation rate since sites were in the same range of OC.

693 Compared to the values of the Pesticide Properties Database (PPDB) (Lewis et al., 2016),
694 biotransformation half-lives were faster when estimated in artificial ponds, and for now, we
695 cannot make any conclusion about this difference since the conditions to estimate $DT_{50\text{bio}}$ in
696 PPDB have not been detailed.

697 Overall, biotransformation is commonly expressed by a first-kinetic order model and
698 enhanced by increased vegetation within the artificial pond providing OM that supports
699 microbes and increases the HRT. Therefore, we suggest introducing a modulation coefficient
700 according to vegetation cover and OC content in water and sediments of a specific site will
701 increase k_{bio} representability. At this time, no rule has been set up for pesticides
702 biotransformation in artificial ponds: neither a direct link with the pesticide properties has
703 been highlighted because biotransformation does not act alone, but simultaneously with other
704 processes. In the future, a more profound exam of biotransformation rates of different
705 pesticides accounting for their interaction with other processes (i.e., adsorption and settling,
706 adsorption/desorption, and dispersion/diffusion) is needed. There is a lack of information
707 concerning biotransformation in vegetation/biofilm compartments. Therefore further studies
708 should be carried out to estimate corresponding transformation rates and account for a
709 possible lag phase reflecting microbial acclimation, as shown by Passeport et al. (2011a). In
710 pesticides databases (i.e., PPDB) there is no distinction between the different compartments
711 (water, sediments, and vegetation) for biotransformation half-lives, making it hard to compare
712 and confront the rates measured as a part of artificial pond surveys with those in pesticides
713 databases.

714 Globally, biotransformation remains a highly changing phenomenon that depends on both the
715 pesticide properties and the site conditions in which microorganisms can act. Defining

716 specific biotransformation half-life for artificial ponds, would be more informative than the
717 PPDB $DT_{50\text{bio}}$ values to estimate the contribution of biotransformation to overall dissipation in
718 artificial ponds.

719 **2.3.2 Photodegradation**

720 Photodegradation or photolysis transforms pesticides exposed to sunlight, especially at
721 wavelengths below 400 nm. The products formed by sunlight-induced transformations are
722 known as photoproducts (Gerecke et al., 2001). Photodegradation is one of the significant
723 abiotic degradation processes in artificial ponds (Lin et al., 2005; Papaevangelou et al., 2017;
724 Stangroom et al., 2000).

725 Photolysis can be direct or indirect. During direct photolysis, pesticides absorb light energy
726 and become excited; they can be transformed depending on the availability of the activation
727 energy. In indirect photolysis, the pesticide reacts with other species produced
728 photochemically (i.e., $CO_3^{\bullet-}$, hydroxyl radical OH^{\bullet} , singlet oxygen O^1_2 , and triplet-excited
729 state dissolved OM (DOM)), causing degradation or conversion into other products (Katagi et
730 al., 2016; Miller et al., 2005; Pohlman et al., 1983; Zeng et al., 2013; Zepp et al., 1987). OH^{\bullet}
731 can be produced by nitrate photolysis, even at a concentration as low as 0.02 mg.L^{-1} (Vione et
732 al., 2014). DOM generates DOM*, the precursor of singlet molecular oxygen (O^1_2) in surface
733 waters. DOM* and O^1_2 can react with pesticides (Drouin et al., 2021). The most reactive
734 species is OH^{\bullet} attacking C-H bonds and mainly controls pesticide photodegradation.

735 Because of the non-ability of solar radiation to penetrate deeper into the water, direct
736 photolysis remains efficient only in the first centimeters of the artificial pond (Frank et al.,
737 2002). The effectiveness of photodegradation strongly depends on the shading capacity of the
738 vegetation canopy and turbidity within the water column, which may attenuate the penetration
739 of solar radiation (MacIntyre et al., 2006; Rose et al., 2008). This transformation process was
740 also reported to be positively correlated to the artificial pond water pH (Le Person et al.,
741 2016). In natural waters, DOM and inorganic compounds are essential absorbers of sunlight
742 and influence the photochemistry of pesticides in artificial pond (Burrows et al., 2002). In
743 addition, particulate matter such as sediment particles and microorganisms suspended in the
744 water column may scatter incident light and prevent the light from penetrating the artificial
745 pond. This phenomenon is called the quenching effect of OM (Larson et al., 1994; Pattanayak
746 et al., 2018; Ruiz-Medina et al., 2018). Photochemical degradation would be more efficient in
747 an open water artificial pond with low turbidity (O'Geen et al., 2010; Vymazal et al., 2015). In
748 DOC-rich ponds ($DOC > 20 \text{ mg.L}^{-1}$), photolysis is unlikely to be operative in pesticide
749 dissipation since DOC content massively attenuates UV light transmission to DP (Waiser et
750 al., 2004). Lagunas-Allué et al. (2010) observed less than an 8% decrease of boscalid caused
751 by direct photolysis, while the main removal processes were adsorption on OM and plant
752 uptake. This observation can be explained by the high OM content that prevents light
753 penetration to pesticides and limits direct photolysis.

754 On the other hand, several herbicide were efficiently removed by indirect photolysis, such as
755 atrazine, diuron, and chlorpyrifos in a study conducted by Zeng et al. (2013). Mathon et al.
756 (2019) evaluated the photodegradation of 23 pesticides using in-situ photo-reactors. They
757 found that 15 pesticides were photodegraded in the summer against 12 in winter, which
758 showed that photolysis was not directly linked to the season. However, when Mathon et al.
759 (2019) compared each of the 23 pesticides photolysis direct and direct+indirect experimental

760 half-lives, they found that indirect photolysis enhances the degradation compared to direct
761 photolysis. Additionally, the contribution of direct and indirect photolysis was studied for 60
762 pesticides. Direct photolysis predominated for dinitroaniline herbicides, although more than
763 50% of the total photodegradation rate could be attributed to indirect photolysis for the other
764 pesticides. This observation supports the relevance of indirect photolysis in artificial ponds
765 when light attenuation is minimal (Katagi et al., 2016; Zeng et al., 2013).

766 As with most transformation processes, photodegradation was commonly computed with a
767 first-order kinetic model (Green et al., 1990; Lam et al., 2003; Rose et al., 2008; Watanabe et
768 al., 2006; Zeng et al., 2013), where k_{photo} (d^{-1}) is the photodegradation kinetic constant,
769 assumed to be the sum of direct and indirect photolysis kinetic rates (Mathon et al., 2019), and
770 $\text{DT}_{50\text{photo}}$ (d^{-1}) is the photodegradation half-life. Photolysis half-lives that fitted a first-order
771 kinetic model in pesticide fate models are presented in Table A.9.

772 Photodegradation half-lives show that the slowest pesticides to be photodegraded were mobile
773 and moderately mobile pesticides, according to their K_{oc} values. In contrast, while lowly
774 mobile pesticides were rapidly photodegraded (Table A.9). Since lowly pesticides are likely
775 to be sorbed (see section 2.2.1), we propose that they might be indirectly photodegraded once
776 sorbed on sediments, suspended solids, or plants through chemical reactions with free radicals
777 in the water-substrate interface.

778 Overall, since the penetration of solar radiation to deeper water is limited, direct photolysis is
779 considered only in the first centimeters of the artificial pond, depending on the DOC content.
780 In contrast, indirect photolysis involving excited radicals may be more relevant through
781 photochemical degradation and is well described by a first-order kinetic rate. We suggest that
782 introducing a modulating factor taking into consideration vegetation cover and OM limit light
783 penetration will increase the accuracy of photolysis modeling.

784 2.3.3 Hydrolysis

785 Another abiotic transformation process is the hydrolysis of DP, which depends on the pH and
786 temperature of artificial ponds. Hydrolysis was commonly considered a minor dissipation
787 process when compared to adsorption, microbial and photochemical degradation. However, in
788 some cases, hydrolysis played a significant role in the dissipation function of artificial ponds,
789 especially under alkaline conditions (Bondarenko et al., 2004; Liu et al., 2001; Papaevangelou
790 et al., 2017; Yu et al., 2019).

791 For instance, chlorpyrifos was proved sensitive to hydrolysis (Liu et al., 2001; Wu et al.,
792 2002). Experimental studies showed that higher pH (> 8) increased the hydrolysis rate of
793 chlorpyrifos (Agudelo et al., 2010; Baskaran et al., 2003; Sardar et al., 2005), corresponding
794 to a half-life of 50 days (Liu et al., 2019; Mackay et al., 2006). Similarly, carbaryl insecticide
795 was mainly degraded by photolysis and hydrolysis in a pond (Wolfe et al., 1978). Hydrolysis
796 mainly contributed to endosulfan dissipation (Walse et al., 2002) and dichlorvos had a
797 hydrolysis half-life of 3 days (Tomlin, 2009). In contrast, hydrolysis contributed to less than
798 2% of boscalid degradation (Lagunas-Allué et al., 2010). Similarly, the hydrolysis half-life of
799 dieldrin exceeded three months (Singh et al., 1991) and reached 115 and 138 days for
800 diazinon and fenthion, respectively (Ibanez et al., 2006; Mackay et al., 2006; Sakellarides et
801 al., 2003). S-Metolachlor and butachlor were also proved to be persistent to hydrolysis with a
802 half-life of 200-1155 days at different pH (Masbou et al., 2018; Zheng et al., 2001).
803 Sometimes, change in diurnal pH due to photosynthesis by aquatic algae and macrophytes

804 may result in photo-synthetically-driven alkaline hydrolysis of some pesticides (Katagi, 2006,
805 2013).

806 Likewise, hydrolysis can be estimated with a first-order kinetic model (Rose et al., 2008;
807 Wang et al., 2019), where $k_h(d^{-1})$ is the hydrolysis kinetic constant and $DT_{50h}(d^{-1})$ is the
808 hydrolysis half-life. It was reported that the main controlling factor of hydrolysis is pH (Deer
809 et al., 2001; Farran et al., 1988; Sereshti et al., 2021; Stangroom et al., 2000).

810 Overall, most pesticides that threaten water quality were considered insensitive to hydrolysis.
811 It may be due to their chemical structure, such as the lack of methyl ester linkage for
812 sulfosulfuron herbicide (Sabadie, 2002; Saha et al., 2002) and phosphodiester linkages for
813 organophosphate pesticides (Sahin et al., 2020). In addition, artificial pond water might
814 generally be beyond the range of alkaline or acidic conditions, making hydrolysis a very slow
815 process (Rose et al., 2008). Therefore, we assume that hydrolysis could be neglected in
816 modeling pesticide dissipation in artificial ponds.

817

818 3. Discussion

819 After reviewing various research works about pesticide dissipation in artificial ponds, it
 820 appears that some studied different processes, while others only focused on one specific
 821 process. Similarly, some surveys investigated the behavior of multiple pesticides while others
 822 only evaluated one or multiple pesticides from the same chemical group (see section 2).
 823 Papers evaluating multiple processes often studied one process. Therefore for a better
 824 understanding of artificial pond processing, this review combines the investigation of multiple
 825 pesticides behavior with contrasting physicochemical properties and evaluates transport,
 826 transfer, and transformation processes occurring in the artificial pond compartments (water,
 827 sediments, and vegetation). In the following subsections, we discuss this critical review's
 828 main assumptions regarding the processes' controlling factors, the estimation of parameters
 829 required to model processes, and the hierarchization of processes.

830 3.1 Controlling factors of the processes

831 As described in section 2, the surveyed processes depend on several factors:

- 832 • **The environmental conditions:** hydraulic parameters, temperature, sediments and
 833 TSS composition (OC, clay content), vegetation cover, solar radiation, pH, and redox
 834 conditions.
- 835 • **The pesticide physicochemical properties:** mobility K_{oc} , hydrophobicity K_{ow} and
 836 vapor pressure vp .

837 The hydraulic regime is responsible for the residence time. A long HRT gives time for the
 838 pesticide to settle, sorb, desorb, or be transformed (Liu et al., 2019; Sahin et al., 2020).
 839 Another hydraulic parameter is the water column depth; a shallow water depth enhances
 840 photolysis and biotransformation by increasing the oxygen concentration in artificial ponds.

841 Sediments and TSS composition interfere in the sorption process. As presented in section
 842 2.2.1, OC content adsorption and clay content increase the trapping of pesticides in sediments
 843 by favoring desorption hysteresis. Thus, OC and clay content increase the removal of
 844 pesticides from the artificial pond water. There was no direct link between biotransformation
 845 half-lives DT_{50bio} in substrates and their OC content (Table A.11). $DT_{50bio,s}$ values show that
 846 the extent of the studied pesticides was not resistant to biotransformation in the sediment
 847 regardless of the OC content.

848 However, biotransformation half-lives in the water $DT_{50bio,w}$, and sediments $DT_{50bio,s}$ varied
 849 according to pesticide properties. However, no direct link was observed between half-life
 850 (DT_{50bio}) and pesticide hydrophobicity (K_{oc}) (Table A.8). This observation supports our
 851 hypothesis of process synergy, and that dissipation is explained by the interaction between
 852 processes (i.e., sorption and transformation) and not a single process.

853 For pesticides properties, the most affecting ones are the K_{oc} and K_{ow} , since they control the
 854 distribution, mobility, and availability of pesticides in the different artificial pond
 855 compartments. We observed that K_{oc} and K_{ow} were determining factors for sorption,
 856 biotransformation, and photodegradation.

857 We have also noticed that biotransformation half-lives were faster in artificial ponds than
 858 half-lives in pesticide databases (i.e., PPDB). It should also be noticed that biotransformation
 859 half-lives are faster in artificial ponds sediments than in agricultural soils (Bolan et al., 1996;
 860 Passeport et al., 2011a), which support the efficiency and usefulness of artificial ponds.

861 Besides biotransformation, the K_{oc} and $\log K_{ow}$ also affect the sorption process. Once lowly
862 mobile and hydrophobic pesticides are sorbed onto a substrate ($K_{oc} > 1000 \text{ L.kg}^{-1}$ and $\log K_{ow}$
863 > 3), they become harder to release back into the water. In comparison, more mobile pesticides
864 ($K_{oc} < 1000 \text{ L.kg}^{-1}$) are continuously transported within the artificial pond. However, it is
865 unknown whether sorbed pesticides will be trapped in the sediment, suspended solid or
866 vegetation, or desorbed afterward.

867 Numerous studies proved that vegetation is a determining agent in the fate of pesticides as it
868 intervenes implicitly in the removal function of artificial ponds. It provides habitat to
869 microorganisms responsible for pesticide transformation, decreases water velocity to increase
870 settling, increases the OC content and composition diversity to enhance adsorption, and brings
871 oxygen to sediments to promote aerobic transformation. In rare cases, vegetation was also
872 responsible for trapping pesticides due to plant uptake. Nevertheless, it remains complex to
873 estimate and quantify the effect of plant uptake on the fate of pesticides (Jorda et al., 2021).
874 Another phytoremediation function of vegetation is through entophytic bacteria living in the
875 interior tissue of plants that can potentially degrade pesticides. However, the fraction of
876 pesticides collated in plants were often insignificant ($< 10\%$) (Butkovskiy et al., 2021; Singh
877 et al., 2021).

878 Temperature is a stimulator for transformation that increases the microbial activity behind
879 biotransformation, solar radiation is responsible for photodegradation and enhances
880 hydrolysis.

881

882 We suggest that process interactions affect their contribution to pesticide fate. The strong
883 relationship between a process and pesticide properties may explain the choice of studying a
884 specific process and pesticide. For instance, sorption is more involved in hydrophobic
885 pesticide dissipation. At the same time, volatilization contributes more to the behavior of
886 pesticides with a significant vapor pressure (Table A.7). On the other hand, PPDB half-life
887 values can be used to model the behavior of some pesticides if they are close to the half-lives
888 estimated in laboratory conditions or give satisfactory simulation results close to the
889 observations.

890 **3.2 Parameter estimation for modeling**

891 After the analysis of different formulations, we suggest that processes can be quantified and
892 hierarchized according to:

- 893 • Kinetic rates k (d^{-1}) for the transport and transfer processes: adsorption-desorption;
894 settling-resuspension; and volatilization.
- 895 • Half-lives DT_{50} (d) for the transformation processes: biotransformation, photolysis,
896 and hydrolysis.

897 The different methods used to evaluate the previously described processes and their
898 corresponding parameters are summarized in Table 1.

899

900 **Table 1**

901 Commonly established methods in the literature to quantify and parametrize a process for a pesticide dissipation
 902 modeling approach in artificial ponds.

Process	Method	Parameter	Units
Transport			
Settling-Resuspension	Empirical formulas	$v_s; v_r$	m.day^{-1}
Transfer			
Adsorption-Desorption	Batch experiments + Kinetic model fitting	$k_{ads}; k_{des}$	day^{-1}
Volatilization	Empirical formulas	k_v	day^{-1}
Transformation			
Biotransformation	Incubation experiments + Kinetic model fitting	DT_{50bio}	day^{-1}
Photolysis	Lab experiments	$DT_{50photo}$	day^{-1}
Hydrolysis	Empirical formulas	DT_{50h}	day^{-1}

903

904 Most studies were conducted on a laboratory scale (batch, microcosms, and mesocosms).
 905 However, laboratory experiments are less advantageous than the rarely conducted in-situ
 906 manipulations (Fig. 1). More representative; in-situ manipulations imply a better description
 907 of the processes and the fate of pesticides while considering the interactions and links
 908 between the different processes (i.e., between sorption and transformation, sorption and
 909 dispersion/diffusion), which is not easy to bring out in laboratory conditions. In fact, in
 910 laboratory experiments, only one process is emphasized and examined independently of other
 911 processes that usually co-occur in field conditions. This often results in the overestimation of
 912 dissipation in lab-scale studies.

913 Additionally, not all estimated half-lives are compatible with those available in the PPDB,
 914 especially biotransformation (Table A.8). This incompatibility may be explained by the
 915 different sites where parameters were estimated, whereas, for the accessible PPDB data, there
 916 is no site specification. This observation questions the validity of parameters in the free PPDB
 917 data and their applicability in a process-based model that aims to describe what is happening
 918 in the field accurately. Using PPDB values in models increases the uncertainty; therefore, we
 919 suggest defining a DT50 specific to ponds for each process to use in models. The fact that
 920 DT50 values in the PPDB are higher than those calculated in-situ shows a greater dissipation
 921 in pond artificial ponds. Because certain photodegradation half-lives in the literature present
 922 better compatibility with the PPDB values (Table A.8), they could be a good model fit.

923 Table 2 summarizes the studied processes for each pesticide described in this review and the
 924 equation used to describe it. We noticed a lack of studies quantifying and modeling
 925 dissipation processes in artificial ponds because studies are often limited to observing the
 926 process without a modeling approach. A considerable number of pesticides (55) were studied
 927 to evaluate their behavior according to previously described processes. However, many of

928 these pesticides have only been evaluated once. Hence, more studies are needed on these
929 pesticides to compare their behavior and draw more robust conclusions.

930 Some pesticides (i.e., boscalid, chlorpyrifos, and pretilachlor) were evaluated according to
931 multiple processes, but not necessarily with a modeling perspective (Agudelo et al., 2010;
932 Laabs et al., 2007; Lagunas-Allué et al., 2010; Rogers et al., 2009; Tang et al., 2019;
933 Karpuzcu et al., 2013; Mackay et al., 2006; Yu et al., 2019). Only, 57 of the 252 reviewed
934 articles concern modeling.

935 Little is known about the estimation of biotransformation half-life in artificial pond
936 compartments, except that macrophytes and algae enhance both adsorption and
937 biotransformation of DP (Katagi et al., 2016). Similarly, anaerobic biotransformation may
938 occur in the sediment layer yet is still under-researched (Maillard et al., 2014; Vandermaesen
939 et al., 2016). Globally, it remains challenging to determine a $DT_{50\text{bio}}$ in each compartment
940 with lab experiments due to the different behavior a pesticide may have in each of them. It is
941 also challenging because laboratory experiments can hardly mimic reproducibly process
942 interactions occurring in the field. For sorption, vegetation remains the least exploited
943 compartment since there were few surveys to estimate sorption kinetics on vegetation, its
944 biotransformation rates, or its uptake rates.

945

946 We propose that the previously mentioned controlling factors (temperature, OM, and
947 vegetation cover) and their interactions are considered for better accuracy by integrating site-
948 specific modulating factors in the development of pesticide fate models. Modulating factors
949 also highlight the temporal variation of process efficiency, such as biotransformation
950 prevailing during the plant growth period (Vandermaesen et al., 2016). So then the question is
951 whether it is necessary to include this temporal variation in a process-based model to improve
952 the accuracy or if, by doing so, the complexity increases.

953

954 **3.3 Hierarchization of dissipation processes**

955 This review provides a primary idea about the hierarchization of processes according to their
956 contribution to the removal function of artificial ponds.

957 As adsorption can only trap pesticides in artificial ponds, degradation (i.e., complete
958 biotransformation, photolysis, and hydrolysis) contributes more to the artificial pond removal
959 function since it can permanently eliminate toxic substances from the environment. Although
960 little is known about whether sorbed pesticides will be degraded or released back into the
961 environment over time, we assume that sorption is still a key process, as it determines the
962 distribution and availability of pesticides in artificial pond compartments.

963 Among the transformation processes, the most rapid was biotransformation compared to
964 photodegradation for some pesticides. By comparing the different half-lives, volatilization
965 was the slowest process, which is in line with the fact that most pesticides are non-volatile due
966 to their low vapor pressure ($<10^{-5}$ Pa). Lastly, hydrolysis appears negligible of most
967 pesticides.

968 Previous reviews, such as Stehle et al. (2011) and Vymazal et al. (2015), did not find a direct
969 link between pesticide dissipation or accumulation in artificial ponds and pesticide properties
970 (i.e., K_{oc} vs sorption). We suggest that it is because the fate of pesticides in artificial ponds
971 does not result from a single process but a combination of different transport, transfer, and

972 transformation processes involving multiple artificial pond compartments. This combination
973 depends on different conditions (i.e., hydrology, climate, physico-chemical factors, and
974 vegetation cover). We assume that the most dependent processes are sorption and
975 biotransformation. For some moderately mobile and hydrophilic pesticides, biotransformation
976 can be the main dissipation process in a artificial pond. After 42 days, up to 86% of diuron
977 ($K_{oc} = 600 \text{ L.kg}^{-1}$), atrazine ($K_{oc} = 100 \text{ L.kg}^{-1}$) and terbuthylazine ($K_{oc} = 318 \text{ L.kg}^{-1}$) were
978 degraded (Anderson et al., 2002; Gregoire et al., 2009; Hijosa-Valsero et al., 2016). Also, the
979 removal of the moderately mobile triazophos ($K_{oc} = 308 \text{ L.kg}^{-1}$) was mostly achieved by
980 biotransformation rather than adsorption (Cheng et al., 2007; Imfeld et al., 2009; Meng et al.,
981 2014; Rani et al., 2001). In contrast, for the highly hydrophobic butachlor ($\log K_{ow} = 4.5$),
982 adsorption was the main process contributing to its dissipation in microcosm experiments.
983 Many researchers ascertained that strong adsorption to sediments and limited desorption (also
984 known as “aging”) might decrease the bioavailability of pesticides and thus limit their
985 biotransformation (Ahmad et al., 2004; Budd et al., 2011; Chaumet et al., 2021; Lee et al.,
986 2004; Mulligan et al., 2016). All of these results emphasize the dependency between sorption
987 and biotransformation.

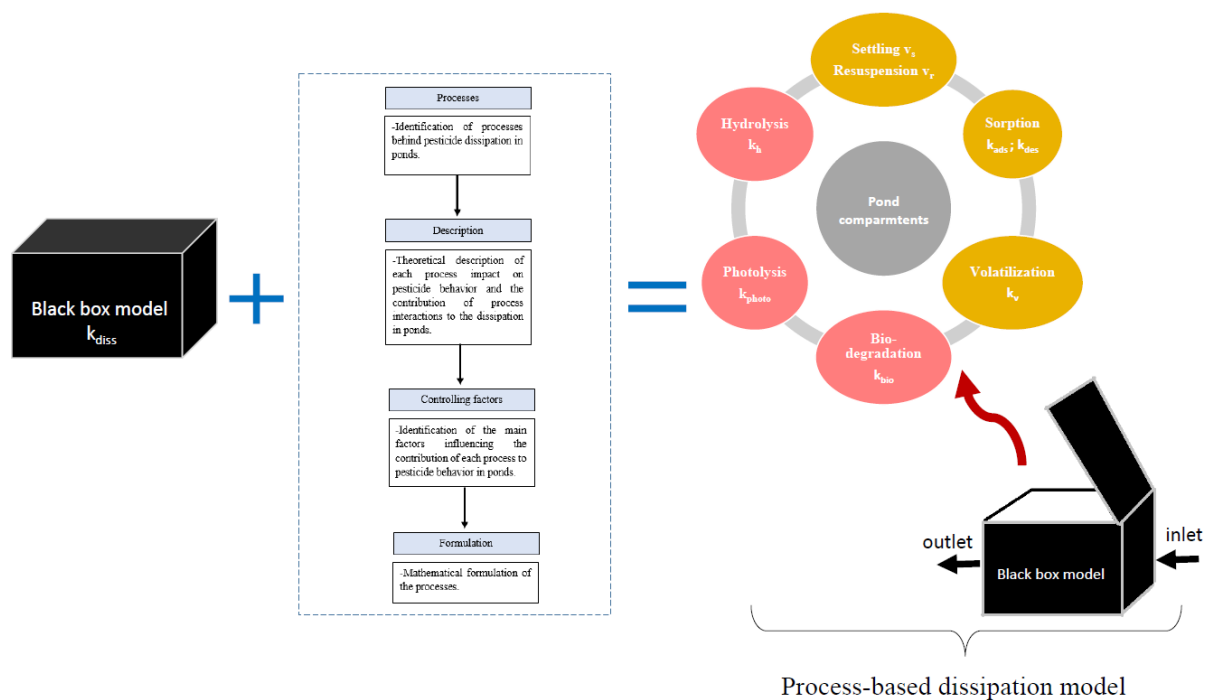
988 When examining citation frequency, some processes (i.e., sorption, biotransformation) were
989 more cited than others (i.e., settling, resuspension, and volatilization). When a process is not
990 frequently cited, this could be explained by the difficulty level of its evaluation in both
991 laboratory and field conditions.

992 The previous conclusions were based on observing removal rates of different processes and
993 pesticides. The limitations of this approach are the relevance of the statements made based on
994 a single case study. Nevertheless, describing one survey means that no other case study was
995 found. The robustness of the conclusions made in this review will be stronger if the same
996 processes are tested on multiple pesticides and sites with contrasting properties to highlight
997 the link between the impact of processes and the properties of the pesticides and artificial
998 ponds.

999
1000 Identifying the processes with a higher impact can be used to hierarchize processes based on
1001 their contribution to the dissipation function of artificial ponds. We believe that a
1002 hierarchization of relevant processes may improve the estimation of the overall efficiency of
1003 artificial ponds in reducing pesticide transfer into the environment.

1004
1005 In addition, the black box model does not consider the processes mentioned above and
1006 describes pesticide behavior with an overall dissipation rate k_{diss} . Although easy to apply, the
1007 black box approach does not provide enough information on the behavior of pesticides and
1008 the contribution of each process to the dissipative function of artificial ponds. Therefore, this
1009 paper discusses the required elements to develop a simple process-based model involving the
1010 transfer, transport, and transformation processes in an interplay of artificial pond
1011 compartments to open the black box (Fig. 3). We assume that a model with multiple process
1012 rates shows the complexity of modeling the fate of pesticides in artificial ponds; however, it
1013 can describe and predict pesticide behavior more realistically. We believe this work will
1014 contribute to a better understanding of artificial pond processing and optimize pesticide fate
1015 models.

1016



1017
 1018 **Fig. 3.** Summarized elements and steps detailed in this work in order to develop a process-based model, that
 1019 contrarily to black models, involves multiple processes in an interplay between different artificial pond
 1020 compartments.

1021

1022

1023

1024

1025 **Table 2**

1026 List of the surveyed processes for each cited pesticide in the reviewed papers, and the equation used to estimate
 1027 the rate of processes. NA means that the process was evaluated without a modeling approach.

Pesticides	Processes							Source
	SET	RES	SOR	VOL	BIO	PHO	HYD	
Acetochlor					Eq. (13)			(Droz et al., 2021)
Aclonifen					NA			(Passeport et al., 2013)
Alphamethrin				Eq. (11)				(Comoretto et al., 2008)
Atrazine			NA		NA			(Besse-Hoggan et al., 2009; Moore et al., 2013; Zeng et al., 2013)
Bifenthrin	NA							(Cryder et al., 2021)
Boscalid			NA		NA	NA	Eq. (17)	(Lagunas-Allué et al., 2010; Vallée, 2015)
Butachlor			NA		NA		NA	(Masbou et al., 2018; Torabi et al., 2020; Zheng et al., 2001)
Carbaryl							NA	(Wolfe et al., 1978)
Chlorothalonil					NA			(Passeport et al., 2013)
Chlorotoluron					NA			(Passeport et al., 2013)
Chlorpyrifos			NA	NA	Eq. (13)		NA	(Agudelo et al., 2010; Laabs et al., 2007; Sardar et al., 2005; Rogers et al., 2009; Tang et al., 2019; Wang et al., 2019; Karpuzcu et al., 2013; Liu et al., 2019; Mackay et al., 2006; Yu et al., 2019)
Clothianidin					Eq. (13)			(Beringer et al., 2021)
Cyfluthrin	NA							(Cryder et al., 2021)
Cyproconazole					NA			(Passeport et al., 2013)
Diazinon			NA		Eq. (13)		Eq. (17)	(Ibáñez et al., 2006; Mackay et al., 2006; Sahin et al., 2020; Sakellarides et al., 2003)
Dichlorvos					NA		Eq. (17)	(Sahin et al., 2020; Tomlin, 2009)
Dieldrin				Eq. (11)			Eq. (17)	(Singh et al., 1991)
Diuron					NA			(Zeng et al., 2013)
Endosulfan				Eq. (11)		Eq. (15)	Eq. (17)	(Rose et al., 2008; Singh et al., 1991; Walse et al., 2002)
Epoxiconazole			NA		NA			(Passeport et al., 2011; Passeport et al., 2013)
Ethofumesate					NA			(Passeport et al., 2013)
Fenitrothion						Eq. (15)		(Sakellarides et al., 2003)
Fenpropidine					Eq. (13)			(Passeport et al., 2013)
Fenthion					NA	Eq. (15)	NA	(Sahin et al., 2020; Sakellarides et al., 2003)
Fipronil	NA							(Cryder et al., 2021)
Fludioxonil			NA			Eq. (15)		(Katagi et al., 2016; Maillard et al., 2014)
Fluometuron			NA			Eq. (15)	Eq. (17)	(Hand et al., 2001; Rose et al., 2008)
Glyphosate			NA					(Maillard et al., 2014)
Imazosulfuron				Eq. (8)				(Takagi et al., 2012)
Imidacloprid			NA					(Mahabali et al., 2014)
Isoproturon			NA		NA			(Passeport et al., 2011; Passeport et al., 2013; Vallée, 2015)
Kresoxim-methyl				Eq. (11)				(Gobas et al., 2018)
Lindane			Eq. (8)					(Krishna et al., 2008)
MCPA			NA		Eq. (13)			(Comoretto et al., 2008; Sørensen et al., 2006; Thorstensen et al., 2001)
Mefenacet			Eq. (8)	Eq. (11)				(Bolan et al., 1996; Watanabe et al., 2006; Yoshida et al., 2000)
Mefenpyr-diethyl				NA				(Passeport et al., 2013)
Metazachlor			NA		Eq. (13)			(Passeport et al., 2011; Passeport et al., 2013)

1028

1029 **Table 2** (continued)

Pesticides	Processes							Source
	SET	RES	SOR	VOL	BIO	PHO	HYD	
Methyl parathion		NA	NA					(Rong et al., 2019; Schulz et al., 2003)
Molinate			Eq. (8)	Eq. (11)		Eq. (15)		(Inao et al., 1999)
Napropamide			NA		NA			(Passeport et al., 2013; Vallée, 2015)
Oxadiazon				Eq. (11)				(Comoretto et al., 2008)
Parathion-Methyl						Eq. (15)		(Sakellarides et al., 2003)
Picloram			Eq. (8)					(McCall et al., 1985)
Pretilachlor			Eq. (8)	Eq. (11)	Eq. (13)	Eq. (15)		(Vallée, 2015)
Prochloraz			NA		NA			(Passeport et al., 2013)
Prosulfocarb					NA			(Gobas et al., 2018)
Pyraclostrobin				Eq. (11)				(Maillard et al., 2014)
Pyrimethanil			NA					(Droz et al., 2021; Masbou et al., 2018; Passeport et al., 2013)
S-Metolachlor					Eq. (13)	Eq. (15)	NA	(Comoretto et al., 2008; Nakano et al., 2004; Watanabe et al., 2006; Yoshida et al., 2000)
Spiroxamine			NA					(Maillard et al., 2014)
Tebuconazole			NA		NA			(Maillard et al., 2014; Passeport et al., 2011; Passeport et al., 2013; Vallée, 2015)
Terbutylazine					NA			(Hijosa-Valsero et al., 2016)
Triazophos					NA			(Cheng et al., 2007)
Trifluralin				NA				(Laabs et al., 2007)

1030

1031 **4. Conclusion**

1032 The scientific community has widely proved the role and efficiency of artificial ponds in
 1033 reducing pesticide transfer into the environment. However, few papers have identified the set
 1034 of processes behind the behavior of pesticides in artificial ponds and opened the black box.
 1035 The relationship between the dissipation potential of processes and the properties of artificial
 1036 ponds and pesticides has yet to receive little attention.

1037 When reviewing the literature, we assumed that pesticide behavior is due to multiple
 1038 processes acting to degrade or trap them in artificial ponds. From this, we can conclude that
 1039 the fate of pesticides results from the interaction of different processes. These processes
 1040 depend on several hydraulic, climatic, and physicochemical factors and vegetation cover.
 1041 Since previous studies have generally focused on a single pesticide or dissipation process, this
 1042 review combines research work on pesticides with contrasting properties. This extends to
 1043 pesticide behavior, reflecting several processes involved in the dissipation of pesticides in a
 1044 large variety of artificial ponds. We also describe and compare the methods used to quantify
 1045 processes behind pesticide dissipation or accumulation in artificial ponds.

1046 Based on the bibliometric analysis, we explain the citation frequency of a process by the
 1047 difficulties of evaluating its contribution in the field or the laboratory. Moreover, investigating
 1048 pesticide behavior in an interplay between transfer, transport, and transformation processes
 1049 can be used to hierarchize processes based on their contribution to the dissipation function of
 1050 pond artificial ponds. We suggest that this hierarchization may also improve the estimation of
 1051 the overall efficiency of pond artificial ponds in pesticide dissipation.

1052 This work is a detailed analysis of the literature to improve our understanding of the potential
1053 of artificial ponds to dissipate incoming pesticide loads and thus lead to better management
1054 and, therefore, water quality improvement. This work paves the way for the scientific
1055 community to develop generic process-based models that, contrary to black-box models, will
1056 predict pesticide behavior at the artificial pond scale while integrating the key dissipation
1057 processes.

1058

1059 **Author Contributions:**

1060 Conceptualization: A.B, J.T, S.S, S.P, G.I.; Writing - Original Draft: A.B; Writing - Review
1061 & Editing: J.T, S.S, S.P, G.I, J.P, B.C; Supervision: J.T, S.S; Project administration: G.I;
1062 Funding acquisition: G.I, J.T, S.S, S.P, J.P.

1063 **Acknowledgments:**

1064 This work was financially supported by the French National Research Agency ANR [ANR-
1065 18-CE32-0007], as a part of the project PESTIPOND, and the French National Research
1066 Institute of Agriculture, Alimentation and Environment.

1067 **Declaration of competing interest:**

1068 The authors declare that they have no known competing financial interests or personal
1069 relationships that could have appeared to influence the work reported in this paper.

1070

1071

Appendix A.

1072

1073

1074 **Table A.1**

1075 Formulation of settling rates of suspended particles in wetlands and associated parameters.

Settling rate v_s (m.s ⁻¹)	Parameters	Conditions of application	Source
$\frac{2}{9} * \frac{r_p^2 * (\rho_p - \rho_w) * g}{\mu}$ (i)	<ul style="list-style-type: none"> • v_s (m.s⁻¹): Settling rate. • r_p (m): Average radius of the particle. • ρ_p (kg.m⁻³): Particles density. • ρ_w (kg.m⁻³): Water density. • g (m.day⁻²): Gravity. • μ (kg.m⁻¹.day⁻¹): Water viscosity. 	Reynolds number < 500	(Fiener et al., 2003; Marois et al., 2016; Reddy et al., 2008)
$\frac{4}{3} * \frac{g * d}{C_D} * \frac{(\rho_s - \rho_w)^{\frac{1}{2}}}{\rho_w}$ (ii)	<ul style="list-style-type: none"> • v_s (m.s⁻¹): Settling rate. • d (m): Particles diameter. • ρ_s (kg.m⁻³): Solid density. • ρ_w (kg.m⁻³): Water density. • g (m.j⁻²): Gravity. • $C_D = f(\text{Rep})$, Rep : Reynolds number of the particle. 	Spherical particles in water at 20°C	(Kadlec et al., 2008)
$\mu * d^{-1} * ((25 + 1,2 * d^{*2})^{0,5} - 5)^{1,5}$ $d^* = (d * (g * \mu^{-2} * (s - 1))^{1/3})$ (iii)	<ul style="list-style-type: none"> • v_s (m.s⁻¹): Settling rate. • d (m): Particles diameter. • g (m.j⁻²): Gravity. • μ (kg.m⁻¹.day⁻¹): Water viscosity. • s: Relative density= $f(\rho_s, \rho_w)$ 	Natural sand particles	(Hawley et al., 2014)

1076

1077

1078 **Table A.2**

1079 Integrated sorption kinetic and isotherm models. Kinetic models describe the temporal evolution of the sorbed
 1080 amount of a dissolved substance. Kinetic rates are estimated from fitting laboratory measurements to the kinetic
 1081 models. Isotherm models describe adsorption and desorption equilibrium. Likewise, isotherm constants are
 1082 calculated from fitting laboratory measurements to isotherm curves.

Kinetic models		
Equations	Parameters	Source
<p><u>Pseudo first order (PFO):</u></p> $C_s = C_{se} - e^{-k_1 * t}$	<ul style="list-style-type: none"> • C_s ($\mu\text{g.g}^{-1}$): Sorbate concentration in the substrate at instant t. • C_{se} ($\mu\text{g.g}^{-1}$): Sorbate concentration in the substrate at the equilibrium. • k_1 (min^{-1}): Kinetic constant. 	<p>(Lin et al., 2009)</p>
<p><u>Pseudo second order (PSO):</u></p> $C_s = \frac{k_2 * C_{se}^2 * t}{1 + k_2 * C_{se} * t}$	<ul style="list-style-type: none"> • C_s ($\mu\text{g.g}^{-1}$): Sorbate concentration in the substrate at instant t. • C_{se} ($\mu\text{g.g}^{-1}$): Sorbate concentration in the substrate at the equilibrium. • k_2 ($\text{g.mg}^{-1}.\text{min}^{-1}$): Kinetic constant rate. 	<p>(Boparai et al., 2011)</p>
<p><u>Elovich:</u></p> $C_s = \frac{1}{\beta} * \ln(1 + \alpha * \beta * t)$	<ul style="list-style-type: none"> • C_s ($\mu\text{g.g}^{-1}$): Sorbate concentration in the substrate at instant t. • α ($\text{g.mg}^{-1}.\text{min}^{-1}$), β (g.mg^{-1}): Kinetic constants. 	<p>(Chien et al., 1980; Ho et al., 2002)</p>
Isotherm models		
Equations	Parameters	Source
<p><u>Freundlich:</u></p> $C_{se} = K_f * C_{we}^{n_f}$	<ul style="list-style-type: none"> • C_{se} ($\mu\text{g.g}^{-1}$): Sorbate concentration in the substrate at the equilibrium. • C_{we} ($\mu\text{g.g}^{-1}$): Sorbate concentration in the water at the equilibrium. • K_f ($\text{mg}^{1-n_f}.\text{L}^n.\text{kg}^{-1}$): Freundlich constant expressing the molecule-substrate affinity. During adsorption K_f is called K_{fads} and K_{fdes} during desorption. • n_f (unitless): Freundlich constant. When $n_f = 1$, the kinetic model becomes linear and K_f becomes the classical distribution coefficient K_d. 	<p>(Mckay, 1995)</p>
<p><u>Langmuir:</u></p> $C_{se} = \frac{K_L * C_{we}}{1 + n_L * C_{we}}$	<ul style="list-style-type: none"> • C_{se} ($\mu\text{g.g}^{-1}$): Sorbate concentration in the substrate at the equilibrium. • C_{we} ($\mu\text{g.g}^{-1}$): Sorbate concentration in the water at the equilibrium. • K_L (L.g^{-1}), n_L (unitless): Langmuir constants. 	<p>(Mckay, 1995)</p>

1084

1085 **Table A.3**

1086 Non-linear Freundlich isotherm adsorption K_{fads} ($\text{mg}^{1-n} \text{kg}^{-1} \text{L}^n$) and desorption K_{fdes} ($\text{mg}^{1-n} \text{kg}^{-1} \text{L}^n$) coefficients.
 1087 Linear isotherm coefficient K_d ($\text{L} \cdot \text{kg}^{-1}$) for metazachlor, isoproturon, epoxiconazole, napropamide, boscalid,
 1088 prochloraz, tebuconazole, glyphosate, pyrimethanil, tetraconazole, fludioxonil and spiroxamine. The sorption
 1089 coefficients were measured in laboratory conditions with batch experiments on different pond substrates:
 1090 Sediments, vegetation, and total suspended solids (TSS) (Maillard et al., 2014; Passeport et al., 2011b; Vallée,
 1091 2015).

Pesticide	K_d/K_f^*	Substrates	K_d	K_{fads}	K_{fdes}	Source
Metazachlor	$K_d = 0.78$	Sediments	3.65	4.5	0.11	
	$K_f = -$	Vegetation	53	60	3,35	
Isoproturon	$K_d = -$	Sediments	5.85	3.5	0.1	
	$K_f = 2.83$	Vegetation	34	44	26.5	
Epoxiconazole	$K_d = -$	Sediments	-	281.5	0.2	
	$K_f = 12.2$	Vegetation	546	902	2.34	(Passeport et al., 2011)
Napropamide	$K_d = 21.5$	Sediments		5	4.8	
	$K_f = 21,6$	Vegetation		77.4	75.2	
Boscalid	$K_d = -$	Sediments		9.05	8.9	
	$K_f = 12.6$	Vegetation	-	191.8	129.5	
Prochloraz	$K_d = -$	Sediments		63.3	10.3	
	$K_f = 38$	Vegetation		442.2	110	
Tebuconazole	$K_d = -$	Sediments		12.3	9	
	$K_f = 12.69$	Vegetation		127.8	98.1	(Vallée, 2015)
Glyphosate	$K_d = 209.4$		Inlet: 87.06			
	$K_f = 226.3$	TSS	Outlet: 18067			
Pyrimethanil	$K_d = -$		Inlet: 833.3			
	$K_f = 8.38$	TSS	Outlet: 414			
Tetraconazole	$K_d = -$		Inlet: 524.8	-		
	$K_f = 145$	TSS	Outlet: 1533			
Fludioxonil	$K_d = -$		Inlet: 944.4			
	$K_f = 3312$	TSS	Outlet: 977			
Spiroxamine	$K_d = -$		Inlet: 1461			
	$K_f = 142.3$	TSS	Outlet: 1681			(Maillard et al., 2014)

1092

1093 (*): Sorption coefficients (K_d , K_f) were extracted from the pesticide database PPDB (Lewis et al., 2016).

1094 (-): Not measured.

1095

1096

1097 **Table A.4**

1098 Adsorption and desorption kinetics (k_{ads} , k_{des}) (d^{-1}) of five selected pesticides (imazosulfuron, lindane,
 1099 mefenacet, molinate and picloram) in different ponds, along with their physicochemical properties S ($mg.L^{-1}$)
 1100 and $K_{oc}(L.kg^{-1})$ (Inao et al., 1999; Krishna et al., 2008; Mccall et al., 1985; Takagi et al., 2012; Yoshida et al.,
 1101 2000). Pesticide properties were extracted from pesticide databases (ChemSpider; Kim et al., 2021; Lewis et al.,
 1102 2016).

Pesticide	Physicochemical properties	Adsorption $k_{ads}(d^{-1})$	Desorption $k_{des}(d^{-1})$	Site	Source
Imazosulfuron	S (mg/l) = 1400 $K_{oc}(L/kg)$ = 754	$1.64 \cdot 10^{-1}$	$7.59 \cdot 10^{-3}$	Paddy field OC = 1% OM = 1.83%	(Takagi et al., 2012)
Lindane	S (mg/l) = 8.5 $K_{oc}(L/kg)$ = 1081	$1.7 \cdot 10^{-4}$	$8.54 \cdot 10^{-6}$	Agricultural pond OC=2% Clay=56%	(Krishna et al., 2008)
Mefenacet	S (mg/l) = 4 $K_{oc}(L/kg)$ = 2964	1.2	$2 \cdot 10^{-1}$	Paddy field OC = 16% OM = 27%	(Yoshida et al., 2000)
Molinate	S (mg/l) = 1100 $K_{oc}(L/kg)$ = 190	$4.87 \cdot 10^{-1}$	$2.9 \cdot 10^{-1}$	Paddy field OC = 2%, OM=3.44% Clay = 47%	(Inao et al., 1999)
Picloram	S (mg/l) = 430 $K_{oc}(L/kg)$ = 100	$1.78 \cdot 10^{-1}$	3.7	Agricultural pond OC= 3%	(McCall et al., 1985)
Pretilachlor	S (mg/l) = 50 $K_{oc}(L/kg)$ = 1159	$1.4 \cdot 10^{-5}$	$2.3 \cdot 10^{-6}$	Paddy field OC = 16% OM = 27%	(Yoshida et al., 2000)

1103

1104

1105

1106 **Table A.5**

1107 Volatilization classes according to the vapor pressure values (Höllrigl-Rosta, 2017).

Vapor pressure (Pa) at 20°C	Volatilization class
$< 10^{-5}$	Non-volatile
$< 10^{-4}$	Non-volatile from soil surfaces, no volatilization from greenhouses
$10^{-5} < \text{---} < 10^{-4}$	Semi volatile
$10^{-4} < \text{---} < 5 \cdot 10^{-3}$	Semi volatile
$\geq 10^{-3}$	Volatile

1108

1109

1110

1111 **Table A.6**

1112 Empirical formulations of volatilization rate $k_v(d^{-1})$ along with their corresponding parameters, based on a 1st
 1113 order kinetic volatilization model.

Volatilization rate k_v	Parameters	Source
$\frac{1}{h_w} * \frac{\frac{H}{RT}}{\frac{1}{720 * \sqrt{\frac{18}{MW}}} + \frac{1}{4.8 * \sqrt{\frac{44}{MW}}}}$	<ul style="list-style-type: none"> • H (Pa.m³.mol⁻¹): Henry coefficient at 25°C. • R (Pa.m³.mol⁻¹ K⁻¹): Universal gas constant. • T (K): Absolute temperature. • MW (g.mol⁻¹): Molecular Weight of the chemical compound. • hw (m): Water depth. 	(Jacobs et al., 2012)
$\frac{1}{0,0036 + 0,01 * u^{1,2} + \frac{1}{H * (3,6 + 5 * u^{1,2})}}$	<ul style="list-style-type: none"> • u (m.s⁻¹): Wind speed. • H (Pa m³ g⁻¹): Henry constant. • T(°C) : Temperature. 	(Rose et al., 2008)

1114

1115

1116

1117 **Table A.7**

1118 Review of the volatilization half-lives DT_{50v} (d) and the corresponding volatilization rates k_v (d^{-1}) in ponds, of
 1119 different pesticides along with their corresponding vapor pressure vp (Pa) and their volatilization classification
 1120 according to the table A.5.

Pesticide	vp (Pa)*	Classification	DT_{50v} (d)	k_v (d^{-1})	Source
Alphamethrin	$2.3 \cdot 10^{-3}$	Volatile	0.2	3.01	
Oxadiazon	$1.49 \cdot 10^{-5}$	Semi volatile	24.8	0.03	(Comoretto et al., 2008)
Kresoxim-methyl	$2.29 \cdot 10^{-6}$	Non-volatile	96924	0.00	
Pyraclostrobin	$2.60 \cdot 10^{-8}$	Non-volatile	6850343	0.00	(Gobas et al., 2018)
Molinate	1.3	Volatile	2.6	0.27	(Inao et al., 1999)
Endosulfan	$7.59 \cdot 10^{-1}$	Volatile	3	0.23	
Dieldrin	$3.20 \cdot 10^{-3}$	Volatile	4.4	0.16	(Singh et al., 1991)
Pretilachlor	$1.2 \cdot 10^{-4}$	Semi volatile	346.5	0.00	
Mefenacet	$6.39 \cdot 10^{-7}$	Non-volatile	5941	0.00	(Watanabe et al., 2006)

1121

1122 *: vp (Pa) is the vapor pressure of the pesticide at 20°C. The values were extracted from pesticides properties databases
 1123 PPDB, PubChem and ChemSpider.

1124

1125

1126 **Table A.8**

1127 Biotransformation half-lives in the water column $DT_{50bio,w}$ (d) and sediment surface $DT_{50bio,s}$ (d) of nine
 1128 pesticides in different wetlands, along with their mobility K_{oc} ($L \cdot kg^{-1}$) (Beringer et al., 2021; Bondarenko et al.,
 1129 2004; Comoretto et al., 2008; Droz et al., 2021; Nakano et al., 2004; Passeport et al., 2011a; Passeport et al.,
 1130 2013; Sahin et al., 2020). These half-lives were confronted to the PPDB values.

Pesticide	K_{oc}^+ (L/kg)	$DT_{50bio,w}$ (d)	$DT_{50bio,s}$ (d)	Site	$DT_{50bio,w}$ (d) PPDB	$DT_{50bio,s}$ (d) PPDB	Source
Acetochlor	156	Anaerobic: 59 Aerobic: 31	Anaerobic: 47 Aerobic: 53	Wetland OC=2.3%	40.5	19.7	(Droz et al., 2021)
Chlorpyrifos	5509	-	Anaerobic: 125-746 Aerobic: 1.8 - 4.9	Urban stream sediment OC=1.8%	-	36.5	(Bondarenko et al., 2004)
Clothianidin	123	-	Anaerobic: 27.2 Aerobic: 115	Wetland OC=0.2%	-	-	(Beringer et al., 2020)
Diazinon	165-189	-	Aerobic: 53.32	Lake sediment OC= 2.22%	4.3	10.4	(Sahin et al., 2020)
Fenthion	1500	-	Aerobic: 5.5		-	92	
MCPA	100	-	5.1*	Paddy field OC=0.9%, Clay=47%	-	25	(Comoretto et al., 2008)
Metazachlor	134	216*	20.6*	Wetland OC = -	-	-	(Passeport et al., 2013)
Pretilachlor	1159	9.8*	19.0*	Paddy field OC=2.3%	-	-	(Nakano et al., 2004)
S-Metolachlor	300	Anaerobic: 199 Aerobic: 29	Anaerobic: 182 Aerobic: 151	Wetland OC=2.3%	88	365	(Droz et al., 2021)
Epoxiconazole	300		170	Wetland OC=2%	1000	103.6	(Passeport et al., 2011)

1131

1132 *: It was not specified under which conditions (aerobic or anaerobic) biotransformation half-lives were measured.

1133 +: The K_{oc} values were extracted from pesticides databases (ChemSpider; Kim et al., 2021; Lewis et al., 2016; Zambonin et
 1134 al., 2002).

1135 - : Not available.

1136

1137

1138 **Table A.9**

1139 Photodegradation half-lives in the water column $DT_{50\text{photo}}$ (d) of eight selected pesticides with different
 1140 mobilities K_{oc} ($L \cdot kg^{-1}$). The DT_{50} values were estimated in (Inao et al., 1999; Katagi et al., 2016; Rose et al.,
 1141 2008; Sakellarides et al., 2003).

Pesticide	K_{oc} * (L/kg)	$DT_{50\text{photo}}$ (d)	PPDB	Source
Atrazine	100	385	2.6	(Mathon et al. 2019)
Dimethoate	5.2-50	495	175	(Mathon et al. 2019)
Diuron	680	93.7	43	(Mathon et al. 2019)
Endosulfan	11500	12	-	(Rose et al., 2008)
Fenitrothion	2000	0.5	3.5	(Sakellarides et al., 2003)
Fenthion	1500	1.6	0.4	(Sakellarides et al., 2003)
Fludioxonil	145 600	9	10	(Katagi et al., 2016)
Fluometuron	202	1	stable	(Rose et al., 2008)
Isoproturon	126	41.3	48	(Mathon et al. 2019)
Molinate	190	54	stable	(Inao et al., 1999)
Parathion-Methyl	1500	25	9	(Sakellarides et al., 2003)
Simazine	130	173	1.9	(Mathon et al. 2019)

1142 *: The K_{oc} values were extracted from pesticides databases (ChemSpider; Kim et al., 2021; Lewis et al., 2016; Zambonin et
 1143 al., 2002).

1144

1145

1146 **Table A.10**

1147 Modulating factors applied to the photolysis rate k_{photo} to account for controlling factors (i.e., Temperature,
 1148 depth, and radiation).

Modulating factors of k_{photo}	Parameters	Source
$\frac{1}{h_w} * \frac{I_0}{I_{\max}} * \frac{1}{25 * TSS * e^{-25 * TSS * z}}$	<ul style="list-style-type: none"> • z (m): Depth. • I_0 ($\text{MJ m}^{-2} \text{h}^{-1}$): Actual radiation. • I_{\max} ($\text{MJ m}^{-2} \text{h}^{-1}$): Maximum radiation. • TSS (mg.g^{-1}): Amount of TSS. 	(Rose et al., 2008)
$R_{\text{UVB}-b}$	<ul style="list-style-type: none"> • $R_{\text{UVB}-b}$: Daily UV-B radiation that penetrates the pond. 	(Watanabe et al., 2006)

1149

1150

1151 **Table A.11**

1152 Biotransformation half-lives $DT_{50\text{bio},s}$ (d) of five pesticides with different mobilities K_{oc} and hydrophilicities \log
 1153 K_{ow} , measured in laboratory studies on sediment substrates with different organic carbon content OC (%) (Vallee
 1154 et al., 2016). Along with, biotransformation half-lives at the water-sediment interface in wetlands, extracted from
 1155 the PPDB.

Pesticides	K_{oc} (L/kg)*	$\log(K_{ow})^*$	OC (%)	$DT_{50\text{bio},s}$ (d)	$DT_{50\text{bio},s}$ (d) PPDB
Boscalid	9500	2.96	1.23	39.4	
			1.5	29.8	545
			1.7	9.4	
Isoproturon	122	2.5	1.2	100	
			1.5	61	149
			1.7	63.9	
Napropamide	885	3.3	1.23	82.7	
			1.5	80.1	316
			1.7	54	
Prochloraz	378	4.6	1.2	100	
			1.5	17.2	359
			1.7	10.2	
Tebuconazole	769	3.7	1.2	30.5	
			1.5	30.6	365
			1.7	1.4	

1156

1157 (*): The physicochemical properties K_{oc} and $\log(K_{ow})$ were extracted from the Pesticide Properties DataBase PPDB and
 1158 PubChem.

1159

Appendix B.

1160
1161
1162
1163
1164
1165
1166
1167
1168
1169
1170
1171
1172
1173
1174
1175
1176
1177
1178
1179
1180
1181
1182
1183
1184
1185
1186
1187
1188
1189
1190
1191

Methodology

In this paragraph, we will present the methodology adopted to realize this critical review of process formulations and controlling factors behind pesticide dissipation in ponds. This work started with a bibliometric research. We used the listed research equations in different bibliographic research tools*:

- (constructed wetland) AND ((wastewater OR agricultural) AND treatment)

- pesticide AND (dissipation OR mitigation) AND (wetland OR pond)

- (pesticide OR "Organic pollutant") AND wetland AND model

- (pesticide OR "Organic pollutant") AND wetland AND fate

- pesticide AND *process* AND (pond OR wetland)

- pesticide AND transfer AND (wetland OR pond)

- *process* AND (rate OR model OR kinetic) AND pesticide AND pond

*: Web of Science (1), Scopus (2), Google Scholar (3), EndNote (4), and Science direct (5).

(1):<https://www.webofscience.com/wos/woscc/basic-search>

(2):<https://www.scopus.com/search/form.uri?display=basic#basic>

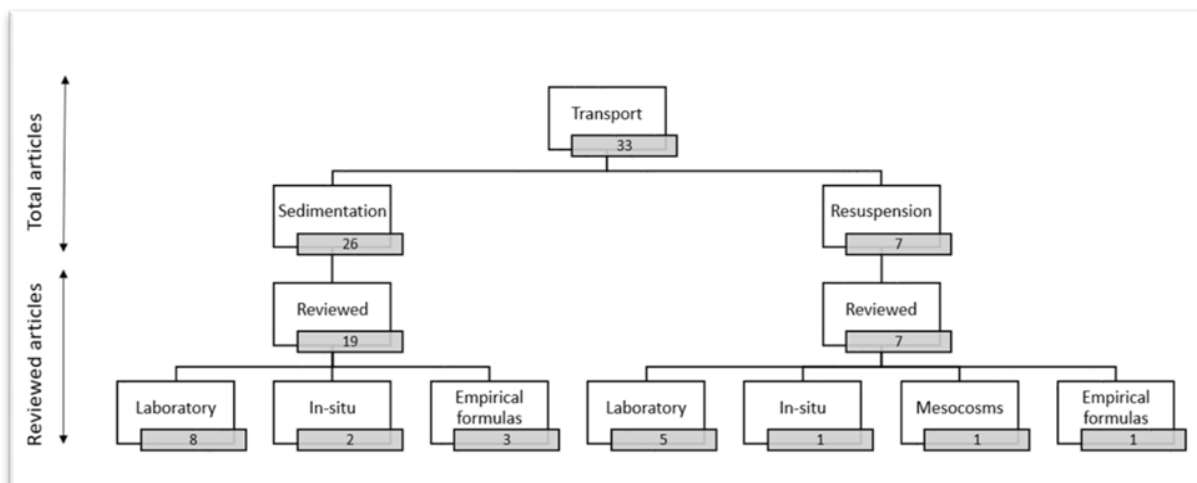
(3): <https://scholar.google.com/schhp?hl=fr>

(4): <https://www.myendnoteweb.com/EndNoteWeb.html>

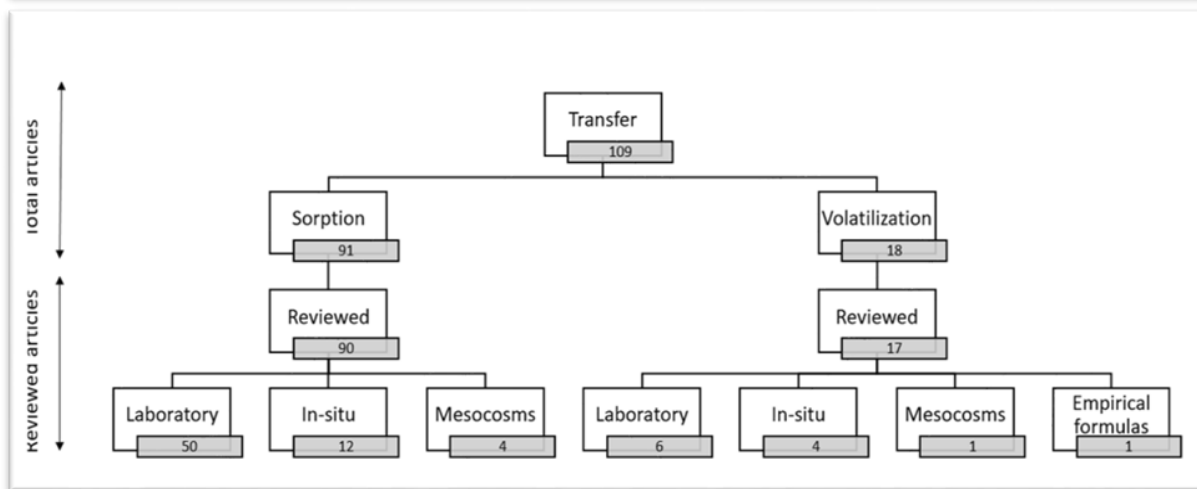
Italic represents the name of the processes (i.e., sorption/adsorption/desorption, biodegradation/degradation, settling/sedimentation/burial, resuspension, photolysis/photodegradation, volatilization, and hydrolysis).

First, we looked for general studies about the fate of pesticides in wetlands to record every potential process behind their dissipation. Afterward, we searched for each process whose effect was investigated in ponds on single or multiple pesticides. We selected theoretical papers for the descriptive part of processes and modeling papers to extract the equations for the formulation part in section 3. In each paper, we identified the pesticides targeted, processes investigated, site, and the scale of their evaluation (i.e., laboratory, in-situ) by focusing more on recent work. Given this, three categories of processes in ponds can be distinguished:

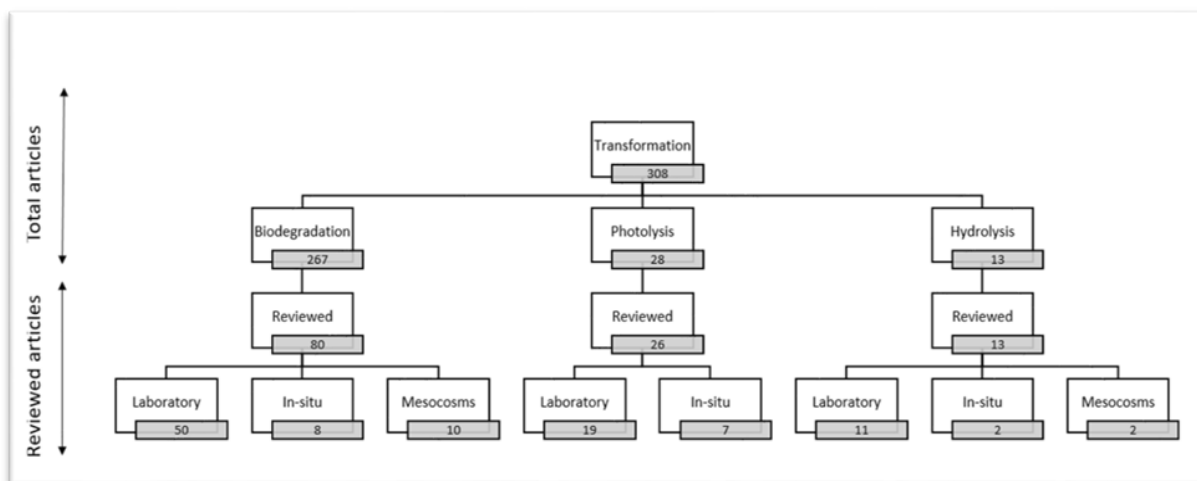
1. Transport is responsible for the fate of pesticides within a compartment of the pond.
2. Transfer controls redistribution of pesticides from a compartment to another.
3. Transformation degrades and modifies the original chemical structure of pesticides.



1192



1193



1194

1195 **Fig A. 1.** Diagram showing the number of searched articles according to the type of processes, and the number
 1196 of papers we reviewed. We reviewed papers with the most information about pesticides and processes in ponds.
 1197 We did not analyze papers with little information, out of context, or redundant. The bottom of the diagram refers
 1198 to the enumeration of papers according to study scale.

1199 Among the set of processes, according to transformation processes, biodegradation was the
 1200 most one surveyed with a total of 267 papers, followed by 91 studies of sorption (Fig. 1).
 1201 Photolysis and settling were less evaluated; only 28 and 26 articles cited them, respectively.
 1202 Finally, resuspension and volatilization had a minor presence in search results. As for
 1203 processes formulations, it was necessary to look for modeling papers that either presented a
 1204 process equation within the framework of a fate model or studied just that specific process.
 1205 Among the 273 reviewed papers, only 20% were about descriptive models of pesticides in

1206 ponds, and over 80% evaluated more than one pesticide. Although, most of the studies
1207 investigated a single process for a group of 2-6 pesticides. Most studies aiming to quantify a
1208 process rate were conducted at a laboratory scale in controlled conditions. They were few
1209 studies conducted in mesocosms* and even less in the field.

1210 *: Mesocosms are experimental water enclosures designed to provide a limited body of water with close to
1211 natural conditions, in which environmental factors can be realistically manipulated ([Berger et al., 2016](#)).

1212

1213

1215 **References**

- 1216 Agudelo, C. R., Jaramillo, M. L., & Penuela, G. (2012). Comparison of the removal of chlorpyrifos and
 1217 dissolved organic carbon in horizontal sub-surface and surface flow wetlands. *Science of the*
 1218 *Total Environment*, 431, 271-277. doi:10.1016/j.scitotenv.2012.05.045
- 1219 Agudelo, R. M., Penuela, G., Aguirre, N. J., Morato, J., & Jaramillo, M. L. (2010). Simultaneous removal
 1220 of chlorpyrifos and dissolved organic carbon using horizontal sub-surface flow pilot wetlands.
 1221 *Ecological Engineering*, 36(10), 1401-1408. doi:10.1016/j.ecoleng.2010.06.019
- 1222 Ahmad, R., Kookana, R. S., Megharaj, M., & Alston, A. M. (2004). Aging reduces the bioavailability of
 1223 even a weakly sorbed pesticide (carbaryl) in soil. *Environ Toxicol Chem*, 23(9), 2084-2089.
 1224 doi:10.1897/03-569
- 1225 Aktar, M. W., Sengupta, D., & Chowdhury, A. (2009). Impact of pesticides use in agriculture: their
 1226 benefits and hazards. *Interdisciplinary toxicology*, 2(1), 1-12. doi:10.2478/v10102-009-0001-7
- 1227 Anderson, K. L., Wheeler, K. A., Robinson, J. B., & Tuovinen, O. H. (2002). Atrazine mineralization
 1228 potential in two wetlands. *Water Res*, 36(19), 4785-4794. doi:10.1016/s0043-1354(02)00209-
 1229 9
- 1230 Arcangeli, J. P., & Arvin, E. (1997). Modeling of the cometabolic biodegradation of trichloroethylene
 1231 by toluene oxidizing bacteria in a biofilm system. *Environmental Science & Technology*,
 1232 31(11), 3044-3052. doi:DOI 10.1021/es9609112
- 1233 Azizian, S. (2004). Kinetic models of sorption: a theoretical analysis. *J Colloid Interface Sci*, 276(1), 47-
 1234 52. doi:10.1016/j.jcis.2004.03.048
- 1235 Azizian, S., Eris, S., & Wilson, L. D. (2018). Re-evaluation of the century-old Langmuir isotherm for
 1236 modeling adsorption phenomena in solution. *Chemical Physics*, 513, 99-104.
 1237 doi:10.1016/j.chemphys.2018.06.022
- 1238 Barchanska, H., Plonka, J., Jaros, A., & Ostrowska, A. (2019). Potential application of Pistia stratiotes
 1239 for the phytoremediation of mesotrione and its degradation products from water. *Int J*
 1240 *Phytoremediation*, 21(11), 1090-1097. doi:10.1080/15226514.2019.1606780
- 1241 Baskaran, S., Kookana, R. S., & Naidu, R. (2003). Contrasting behaviour of chlorpyrifos and its primary
 1242 metabolite, TCP (3,5,6-trichloro-2-pyridinol), with depth in soil profiles. *Australian Journal of*
 1243 *Soil Research*, 41(4), 749-760. doi:10.1071/Sr02062
- 1244 Beecraft, L., & Rooney, R. (2021). Bioconcentration of glyphosate in wetland biofilms. *Science of the*
 1245 *Total Environment*, 756, 143993. doi:10.1016/j.scitotenv.2020.143993
- 1246 Berger, S. A., & Nejtgaard, J. C. (2016). A Worldwide Web-portal for Aquatic Mesocosm Facilities:
 1247 WWW. MESOCOSM. EU. *American Geophysical Union, 2016*, IS34A-2323.
- 1248 Beringer, C. J., Goynes, K. W., Lerch, R. N., Webb, E. B., & Mengel, D. (2021). Clothianidin
 1249 decomposition in Missouri wetland soils. *J Environ Qual*, 50(1), 241-251.
 1250 doi:10.1002/jeq2.20175
- 1251 Besse-Hoggan, P., Alekseeva, T., Sancelme, M., Delort, A. M., & Forano, C. (2009). Atrazine
 1252 biodegradation modulated by clays and clay/humic acid complexes. *Environ Pollut*, 157(10),
 1253 2837-2844. doi:10.1016/j.envpol.2009.04.005
- 1254 Beulke, S., Brown, C. D., Fryer, C. J., & van Beinum, W. (2004). Influence of kinetic sorption and
 1255 diffusion on pesticide movement through aggregated soils. *Chemosphere*, 57(6), 481-490.
 1256 doi:10.1016/j.chemosphere.2004.06.026
- 1257 Bezbaruah, A. N., & Zhang, T. C. (2005). Quantification of oxygen release by bulrush (*Scirpus validus*)
 1258 roots in a constructed treatment wetland. *Biotechnol Bioeng*, 89(3), 308-318.
 1259 doi:10.1002/bit.20332
- 1260 Blachnio, M., Derylo-Marczewska, A., & Seczkowska, M. (2020). Influence of Pesticide Properties on
 1261 Adsorption Capacity and Rate on Activated Carbon from Aqueous Solution.
 1262 doi:10.5772/intechopen.88726
- 1263 Bladel, R. V., & Laudelout, H. (1967). APPARENT IRREVERSIBILITY OF ION-EXCHANGE REACTIONS IN
 1264 CLAY SUSPENSIONS *Soil Science* (Vol. 104).
- 1265 Bloomfield, J. P., Williams, R. J., Gooddy, D. C., Cape, J. N., & Guha, P. (2006). Impacts of climate
 1266 change on the fate and behaviour of pesticides in surface and groundwater--A UK
 1267 perspective. *Science of the Total Environment*, 369(1-3), 163-177.
 1268 doi:10.1016/j.scitotenv.2006.05.019

- 1269 Bojcevska, H., & Tonderski, K. (2007). Impact of loads, season, and plant species on the performance
1270 of a tropical constructed wetland polishing effluent from sugar factory stabilization ponds.
1271 *Ecological Engineering*, 29(1), 66-76. doi:10.1016/j.ecoleng.2006.07.015
- 1272 Bolan, N. S., & Baskaran, S. (1996). Biodegradation of 2,4-D herbicide as affected by its adsorption-
1273 desorption behaviour and microbial activity of soils. *Australian Journal of Soil Research*,
1274 34(6), 1041-1053. doi:10.1071/Sr9961041
- 1275 Bondarenko, S., Gan, J., Haver, D. L., & Kabashima, J. N. (2004). Persistence of selected
1276 organophosphate and carbamate insecticides in waters from a coastal watershed. *Environ*
1277 *Toxicol Chem*, 23(11), 2649-2654. doi:10.1897/03-436
- 1278 Boparai, H. K., Joseph, M., & O'Carroll, D. M. (2011). Kinetics and thermodynamics of cadmium ion
1279 removal by adsorption onto nano zerovalent iron particles. *J Hazard Mater*, 186(1), 458-465.
1280 doi:10.1016/j.jhazmat.2010.11.029
- 1281 Bowman, B., & Sans, W. (1985). *Partitioning Behavior of Insecticides in Soil-Water Systems: I.*
1282 *Adsorbent Concentration Effects* (0047-2425). Retrieved from
- 1283 Braschi, I., Blasioli, S., Lavrnic, S., Buscaroli, E., Di Prodi, K., Solimando, D., & Toscano, A. (2022).
1284 Removal and fate of pesticides in a farm constructed wetland for agricultural drainage water
1285 treatment under Mediterranean conditions (Italy). *Environ Sci Pollut Res Int*, 29(5), 7283-
1286 7299. doi:10.1007/s11356-021-16033-4
- 1287 Braskerud, B. C. (2001). The influence of vegetation on sedimentation and resuspension of soil
1288 particles in small constructed wetlands. *J Environ Qual*, 30(4), 1447-1457.
1289 doi:10.2134/jeq2001.3041447x
- 1290 Braskerud, B. C. (2002). Design considerations for increased sedimentation in small wetlands treating
1291 agricultural runoff. *Water Science and Technology*, 45(9), 77-85. doi:10.2166/wst.2002.0209
- 1292 Brix, H. (2003). *Plants used in constructed wetlands and their functions*. Paper presented at the 1st
1293 International Seminar on the use of Aquatic Macrophytes for Wastewater Treatment in
1294 Constructed Wetlands, edit. Dias V., Vymazal J. Lisboa, Portugal.
- 1295 Brueske, C. C., & Barrett, G. W. (1994). Effects of Vegetation and Hydrologic Load on Sedimentation
1296 Patterns in Experimental Wetland Ecosystems. *Ecological Engineering*, 3(4), 429-447. doi:10.1016/0925-8574(94)00011-5
- 1297
- 1298 Brusseau, M. L., & Chorover, J. (2019). Chemical Processes Affecting Contaminant Transport and
1299 Fate. 113-130. doi:10.1016/b978-0-12-814719-1.00008-2
- 1300 Budd, R., O'Geen, A., Goh, K. S., Bondarenko, S., & Gan, J. (2009). Efficacy of constructed wetlands in
1301 pesticide removal from tailwaters in the Central Valley, California. *Environ Sci Technol*, 43(8),
1302 2925-2930. doi:10.1021/es802958q
- 1303 Budd, R., O'Geen, A., Goh, K. S., Bondarenko, S., & Gan, J. (2011). Removal mechanisms and fate of
1304 insecticides in constructed wetlands. *Chemosphere*, 83(11), 1581-1587.
1305 doi:10.1016/j.chemosphere.2011.01.012
- 1306 Bunyakan, C., Seaoung, S., Intamane, J., & Chungsiriporn, J. (2006). Factors affecting the
1307 volatilization of volatile organic compounds from wastewater *Songklanakarinn J Sci Technol*
1308 (Vol. 28, pp. 1083-1093).
- 1309 Burrows, H. D., Canle, L. M., Santaballa, J. A., & Steenken, S. (2002). Reaction pathways and
1310 mechanisms of photodegradation of pesticides. *J Photochem Photobiol B*, 67(2), 71-108.
1311 doi:10.1016/s1011-1344(02)00277-4
- 1312 Butkovskiy, A., Jing, Y., Bergheim, H., Lazar, D., Gulyaeva, K., Odenmarck, S. R., . . . Eggen, T. (2021).
1313 Retention and distribution of pesticides in planted filter microcosms designed for treatment
1314 of agricultural surface runoff. *Science of the Total Environment*, 778, 146114.
1315 doi:10.1016/j.scitotenv.2021.146114
- 1316 Caceres-Jensen, L., Rodriguez-Becerra, J., Sierra-Rosales, P., Escudey, M., Valdebenito, J., Neira-
1317 Albornoz, A., . . . Villagra, C. A. (2019). Electrochemical method to study the environmental
1318 behavior of Glyphosate on volcanic soils: Proposal of adsorption-desorption and transport
1319 mechanisms. *J Hazard Mater*, 379, 120746. doi:10.1016/j.jhazmat.2019.120746
- 1320 Cai, D., Yang, X., Wang, S., Chao, Y., Morel, J. L., & Qiu, R. (2017). Effects of dissolved organic matter
1321 derived from forest leaf litter on biodegradation of phenanthrene in aqueous phase. *J Hazard*
1322 *Mater*, 324(Pt B), 516-525. doi:10.1016/j.jhazmat.2016.11.020

- 1323 Cara, I. G., Filip, M., Bulgariu, L., Raus, L., Topa, D., & Jitareanu, G. (2021). Environmental Remediation
 1324 of Metribuzin Herbicide by Mesoporous Carbon-Rich from Wheat Straw. *Applied Sciences-
 1325 Basel, 11*(11), 4935. doi:ARTN 4935
- 1326 10.3390/app11114935
- 1327 Carleton, J. N., & Montas, H. J. (2010). An analysis of performance models for free water surface
 1328 wetlands. *Water Research, 44*(12), 3595-3606.
 1329 doi:<https://doi.org/10.1016/j.watres.2010.04.008>
- 1330 Cessna, A. J., Donald, D. B., Bailey, J., & Waiser, M. (2015). Persistence of the Sulfonylurea Herbicides
 1331 Sulfosulfuron, Rimsulfuron, and Nicosulfuron in Farm Dugouts (Ponds). *J Environ Qual, 44*(6),
 1332 1948-1955. doi:10.2134/jeq2014.11.0503
- 1333 Chaumet, B., Probst, J.-L., Payré-Suc, V., Granouillac, F., Riboul, D., & Probst, A. (2022). Pond
 1334 mitigation in dissolved and particulate pesticide transfers: Influence of storm events and
 1335 seasonality (Auradé agricultural catchment, SW-France). *Journal of Environmental
 1336 Management, 320*, 115911. doi:10.1016/j.jenvman.2022.115911
- 1337 Chaumet, B., Probst, J. L., Eon, P., Camboulive, T., Riboul, D., Payre-Suc, V., . . . Probst, A. (2021). Role
 1338 of Pond Sediments for Trapping Pesticides in an Agricultural Catchment (Aurade, SW France):
 1339 Distribution and Controlling Factors. *Water, 13*(13), 1734. doi:ARTN 1734
- 1340 10.3390/w13131734
- 1341 ChemSpider. Retrieved from <https://www.chemspider.com/>
- 1342 Chen, Y., Wen, Y., Zhou, J., Zhou, Q., Vymazal, J., & Kuschik, P. (2015). Transformation of chloroform
 1343 in model treatment wetlands: from mass balance to microbial analysis. *Environ Sci Technol,
 1344 49*(10), 6198-6205. doi:10.1021/es506357e
- 1345 Cheng, S., Xiao, J., Xiao, H., Zhang, L., & Wu, Z. (2007). Technical note phytoremediation of triazophos
 1346 by *Canna indica* Linn. in a hydroponic system. *Int J Phytoremediation, 9*(6), 453-463.
 1347 doi:10.1080/15226510701709531
- 1348 Chien, S. H., & Clayton, W. R. (1980). Application of Elovich Equation to the Kinetics of Phosphate
 1349 Release and Sorption in Soils. *Soil Science Society of America Journal, 44*(2), 265-268. doi:DOI
 1350 10.2136/sssaj1980.03615995004400020013x
- 1351 Chouinard, A., Balch, G. C., Wootton, B. C., Jørgensen, S. E., & Anderson, B. C. (2014). Chapter 19 -
 1352 SubWet 2.0. Modeling the Performance of Treatment Wetlands. *Developments in
 1353 Environmental Modelling, 26*, 519-537. doi:[https://doi.org/10.1016/B978-0-444-63249-
 1354 4.00021-X](https://doi.org/10.1016/B978-0-444-63249-4.00021-X)
- 1355 Climent, M. J., Herrero-Hernandez, E., Sanchez-Martin, M. J., Rodriguez-Cruz, M. S., Pedreros, P., &
 1356 Urrutia, R. (2019). Residues of pesticides and some metabolites in dissolved and particulate
 1357 phase in surface stream water of Cachapoal River basin, central Chile. *Environ Pollut, 251*, 90-
 1358 101. doi:10.1016/j.envpol.2019.04.117
- 1359 Collins, R. (2004). Fecal contamination of pastoral wetlands. *J Environ Qual, 33*(5), 1912-1918.
 1360 doi:10.2134/jeq2004.1912
- 1361 Comoretto, L., Arfib, B., Talva, R., Chauvelon, P., Pichaud, M., Chiron, S., & Hohener, P. (2008). Runoff
 1362 of pesticides from rice fields in the Ile de Camargue (Rhône river delta, France): field study
 1363 and modeling. *Environ Pollut, 151*(3), 486-493. doi:10.1016/j.envpol.2007.04.021
- 1364 Convertino, M., Nardi, F., Kiker, G. A., Muñoz-Carpena, R., Troccoli, A., & Linkov, I. (2013). Epitomes
 1365 of Bottom-Up Hydro-Geo-Climatological Analysis to Face Sea Level Rise in Complex Coastal
 1366 Ecosystems. 267-282. doi:10.1016/b978-0-12-384703-4.00502-5
- 1367 Cryder, Z., Wolf, D., Carlan, C., & Gan, J. (2021). Removal of urban-use insecticides in a large-scale
 1368 constructed wetland. *Environ Pollut, 268*(Pt A), 115586. doi:10.1016/j.envpol.2020.115586
- 1369 Damalas, C. A., & Eleftherohorinos, I. G. (2011). Pesticide exposure, safety issues, and risk assessment
 1370 indicators. *International Journal of Environmental Research and Public Health, 8*(5), 1402-
 1371 1419.
- 1372 Dash, D. M., & Osborne, W. J. (2020). Rapid biodegradation and biofilm-mediated bioremoval of
 1373 organophosphorus pesticides using an indigenous *Kosakonia oryzae* strain -VITPSCQ3 in a
 1374 Vertical-flow Packed Bed Biofilm Bioreactor. *Ecotoxicol Environ Saf, 192*, 110290.
 1375 doi:10.1016/j.ecoenv.2020.110290

- 1376 de Oliveira, M., Frihling, B. E. F., Velasques, J., Filho, F., Cavalheri, P. S., & Migliolo, L. (2020).
1377 Pharmaceuticals residues and xenobiotics contaminants: Occurrence, analytical techniques
1378 and sustainable alternatives for wastewater treatment. *Science of the Total Environment*,
1379 705, 135568. doi:10.1016/j.scitotenv.2019.135568
- 1380 Deer, H. M., & Beard, R. (2001). *Effect of water pH on the chemical stability of pesticides*. Retrieved
1381 from
1382 https://digitalcommons.usu.edu/cgi/viewcontent.cgi?article=1074&context=extension_histal
1383 [!](#)
- 1384 Douven, S., Paez, C. A., & Gomme, C. J. (2015). The range of validity of sorption kinetic models. *J*
1385 *Colloid Interface Sci*, 448, 437-450. doi:10.1016/j.jcis.2015.02.053
- 1386 Drouin, G., Droz, B., Leresche, F., Payraudeau, S., Masbou, J., & Imfeld, G. (2021). *Direct and indirect*
1387 *photodegradation of atrazine and S-metolachlor in agriculturally impacted surface water and*
1388 *associated C and N isotope fractionation*. Cambridge University Press (CUP). Retrieved from
1389 <https://dx.doi.org/10.33774/chemrxiv-2021-sg66n>
- 1390 Droz, B., Drouin, G., Maurer, L., Villette, C., Payraudeau, S., & Imfeld, G. (2021). Phase Transfer and
1391 Biodegradation of Pesticides in Water-Sediment Systems Explored by Compound-Specific
1392 Isotope Analysis and Conceptual Modeling. *Environ Sci Technol*, 55(8), 4720-4728.
1393 doi:10.1021/acs.est.0c06283
- 1394 Farran, A., De Pablo, J., & Barcelo, D. (1988). Identification of organophosphorus insecticides and
1395 their hydrolysis products by liquid chromatography in combination with UV and
1396 thermospray-mass spectrometric detection. *Journal of Chromatography A*, 455, 163-172.
1397 doi:10.1016/s0021-9673(01)82115-4
- 1398 Fernández-Pascual, E., Bork, M., Hensen, B., & Lange, J. (2020). Hydrological tracers for assessing
1399 transport and dissipation processes of pesticides in a model constructed wetland system.
1400 *Hydrology & Earth System Sciences*, 24(1). doi:<https://doi.org/10.5194/hess-24-41-2020>
- 1401 Fiener, P., & Auerswald, K. (2003). Effectiveness of grassed waterways in reducing runoff and
1402 sediment delivery from agricultural watersheds. *J Environ Qual*, 32(3), 927-936.
1403 doi:10.2134/jeq2003.9270
- 1404 Foo, K. Y., & Hameed, B. H. (2010). Insights into the modeling of adsorption isotherm systems.
1405 *Chemical Engineering Journal*, 156(1), 2-10. doi:10.1016/j.cej.2009.09.013
- 1406 Frank, M. P., Graebing, P., & Chib, J. S. (2002). Effect of soil moisture and sample depth on pesticide
1407 photolysis. *J Agric Food Chem*, 50(9), 2607-2614. doi:10.1021/jf0115746
- 1408 Gan, J., Lee, S. J., Liu, W. P., Haver, D. L., & Kabashima, J. N. (2005). Distribution and persistence of
1409 pyrethroids in runoff sediments. *J Environ Qual*, 34(3), 836-841. doi:10.2134/jeq2004.0240
- 1410 Garcia-Rodríguez, A., Matamoros, V., Fontàs, C., & Salvadó, V. (2014). The ability of biologically based
1411 wastewater treatment systems to remove emerging organic contaminants—a review.
1412 *Environmental Science and Pollution Research*, 21(20), 11708-11728. doi:10.1007/s11356-
1413 013-2448-5
- 1414 Gassmann, M., Olsson, O., Stamm, C., Weiler, M., & Kümmerer, K. (2015). Physico-chemical
1415 characteristics affect the spatial distribution of pesticide and transformation product loss to
1416 an agricultural brook. *Science of The Total Environment*, 532, 733-743.
1417 doi:<https://doi.org/10.1016/j.scitotenv.2015.06.068>
- 1418 Gaullier, C., Baran, N., Dousset, S., Devau, N., Billet, D., Kitzinger, G., & Coisy, E. (2019). Wetland
1419 hydrodynamics and mitigation of pesticides and their metabolites at pilot-scale. *Ecological*
1420 *Engineering*, 136, 185-192. doi:<https://doi.org/10.1016/j.ecoleng.2019.06.019>
- 1421 Gaullier, C., Dousset, S., Baran, N., Kitzinger, G., & Coureau, C. (2020). Influence of hydrodynamics on
1422 the water pathway and spatial distribution of pesticide and metabolite concentrations in
1423 constructed wetlands. *J Environ Manage*, 270, 110690. doi:10.1016/j.jenvman.2020.110690
- 1424 Gaullier, C., Dousset, S., Billet, D., & Baran, N. (2018). Is pesticide sorption by constructed wetland
1425 sediments governed by water level and water dynamics? *Environ Sci Pollut Res Int*, 25(15),
1426 14324-14335. doi:10.1007/s11356-017-9123-1
- 1427 Gerecke, A. C., Canonica, S., Muller, S. R., Scharer, M., & Schwarzenbach, R. P. (2001). Quantification
1428 of dissolved natural organic matter (DOM) mediated phototransformation of phenylurea
1429 herbicides in lakes. *Environ Sci Technol*, 35(19), 3915-3923. doi:10.1021/es010103x

- 1430 Gobas, F., Lai, H. F., Mackay, D., Padilla, L. E., Goetz, A., & Jackson, S. H. (2018). AGRO-2014: A time
1431 dependent model for assessing the fate and food-web bioaccumulation of organic pesticides
1432 in farm ponds: Model testing and performance analysis. *Science of the Total Environment*,
1433 639, 1324-1333. doi:10.1016/j.scitotenv.2018.05.115
- 1434 Gramlich, A., Stoll, S., Stamm, C., Walter, T., & Prasuhn, V. (2018). Effects of artificial land drainage on
1435 hydrology, nutrient and pesticide fluxes from agricultural fields - A review. *Agriculture
1436 Ecosystems & Environment*, 266, 84-99. doi:10.1016/j.agee.2018.04.005
- 1437 Green, R. E., & Karickhoff, S. W. (1990). Sorption Estimates for Modeling *Pesticides in the Soil
1438 Environment: Processes, Impacts and Modeling* (pp. 79-101).
- 1439 Gregoire, C., Elsaesser, D., Huguenot, D., Lange, J., Lebeau, T., Merli, A., . . . Schütz, T. (2009).
1440 Mitigation of agricultural nonpoint-source pesticide pollution in artificial wetland
1441 ecosystems. *Environmental Chemistry Letters*, 7(3), 205-231.
1442 doi:<https://doi.org/10.1007/s10311-008-0167-9>
- 1443 Grinham, A., Albert, S., Deering, N., Dunbabin, M., Bastviken, D., Sherman, B., . . . Evans, C. D. (2018).
1444 The importance of small artificial water bodies as sources of methane emissions in
1445 Queensland, Australia. *Hydrology and Earth System Sciences*, 22(10), 5281-5298.
1446 doi:10.5194/hess-22-5281-2018
- 1447 Gu, L., Dai, B., Zhu, D. Z., Hua, Z. L., Liu, X. D., van Duin, B., & Mahmood, K. (2017). Sediment
1448 modelling and design optimization for stormwater ponds. *Canadian Water Resources Journal*,
1449 42(1), 70-87. doi:10.1080/07011784.2016.1210542
- 1450 Guo, L., Jury, W. A., Wagenet, R. J., & Flury, M. (2000). Dependence of pesticide degradation on
1451 sorption: nonequilibrium model and application to soil reactors. *Journal of Contaminant
1452 Hydrology*, 43(1), 45-62. doi:Doi 10.1016/S0169-7722(99)00097-2
- 1453 Haith, D. A. (2010). Ecological risk assessment of pesticide runoff from grass surfaces. *Environmental
1454 Science & Technology*, 44(16), 6496-6502.
- 1455 Halbach, K., Möder, M., Schrader, S., Liebmann, L., Schäfer, R. B., Schneeweiss, A., . . . Reemtsma, T.
1456 (2021). Small streams—large concentrations? Pesticide monitoring in small agricultural
1457 streams in Germany during dry weather and rainfall. *Water Research*, 203, 117535.
1458 doi:<https://doi.org/10.1016/j.watres.2021.117535>
- 1459 Hamdi, A. (2007). Identification of point sources in two-dimensional advection-diffusion-reaction
1460 equation: application to pollution sources in a river. Stationary case. *Inverse Problems in
1461 Science and Engineering*, 15(8), 855-870.
- 1462 Hand, L. H., Kuet, S. F., Lane, M. C., Maund, S. J., Warinton, J. S., & Hill, I. R. (2001). Influences of
1463 aquatic plants on the fate of the pyrethroid insecticide Lambda-cyhalothrin in aquatic
1464 environments. *Environmental Toxicology and Chemistry: An International Journal*, 20(8),
1465 1740-1745. doi:10.1897/1551-5028(2001)020<1740:ioapot>2.0.co;2
- 1466 Hantush, M. M., Kalin, L., Isik, S., & Yucekaya, A. (2013). Nutrient Dynamics in Flooded Wetlands. I:
1467 Model Development. *Journal of Hydrologic Engineering*, 18(12), 1709-1723.
1468 doi:10.1061/(Asce)He.1943-5584.0000741
- 1469 Hawley, N., Redder, T., Beletsky, R., Verhamme, E., Beletsky, D., & DePinto, J. V. (2014). Sediment
1470 resuspension in Saginaw Bay. *Journal of Great Lakes Research*, 40, 18-27.
1471 doi:10.1016/j.jglr.2013.11.010
- 1472 Hijosa-Valsero, M., Becares, E., Fernandez-Alaez, C., Fernandez-Alaez, M., Mayo, R., & Jimenez, J. J.
1473 (2016). Chemical pollution in inland shallow lakes in the Mediterranean region (NW Spain):
1474 PAHs, insecticides and herbicides in water and sediments. *Science of the Total Environment*,
1475 544, 797-810. doi:10.1016/j.scitotenv.2015.11.160
- 1476 Hijosa-Valsero, M., Matamoros, V., Sidrach-Cardona, R., Martin-Villacorta, J., Becares, E., & Bayona, J.
1477 M. (2010). Comprehensive assessment of the design configuration of constructed wetlands
1478 for the removal of pharmaceuticals and personal care products from urban wastewaters.
1479 *Water Res*, 44(12), 3669-3678. doi:10.1016/j.watres.2010.04.022
- 1480 Ho, Y. S. (2006). Review of second-order models for adsorption systems. *J Hazard Mater*, 136(3), 681-
1481 689. doi:10.1016/j.jhazmat.2005.12.043
- 1482 Ho, Y. S., & McKay, G. (1998). A comparison of chemisorption kinetic models applied to pollutant
1483 removal on various sorbents. *Process Safety and Environmental Protection*, 76(B4), 332-340.
1484 doi:Doi 10.1205/095758298529696

- 1485 Ho, Y. S., & McKay, G. (1999). Batch lead (II) removal from aqueous solution by peat: Equilibrium and
1486 kinetics. *Process Safety and Environmental Protection*, 77(B3), 165-173. doi:Doi
1487 10.1205/095758299529983
- 1488 Ho, Y. S., & McKay, G. (2002). Application of kinetic models to the sorption of copper(II) on to peat.
1489 *Adsorption Science & Technology*, 20(8), 797-815. doi:Doi 10.1260/026361702321104282
- 1490 Holland, J., M, B., T, B., S, S., S.R, T., Alexander, C., & Thomas, C. F. (2004). The spatial dynamics and
1491 movement of *Pterostichus melanarius* and *P. madidus* (Carabidae) between and within
1492 arable fields in the UK. *International Journal of Ecology and Environmental Sciences*, 30, 35-
1493 50.
- 1494 Holland, J. F., Martin, J. F., Granata, T., Bouchard, V., Quigley, M., & Brown, L. (2004). Effects of
1495 wetland depth and flow rate on residence time distribution characteristics. *Ecological*
1496 *Engineering*, 23(3), 189-203. doi:<https://doi.org/10.1016/j.ecoleng.2004.09.003>
- 1497 Höllrigl-Rosta. (2017). Establishment of a concept for comparative risk assessment of plant protection
1498 products with special focus on the risks to the environment.
- 1499 Huang, W. L., Ping, P. A., Yu, Z. Q., & Fu, H. M. (2003). Effects of organic matter heterogeneity on
1500 sorption and desorption of organic contaminants by soils and sediments. *Applied*
1501 *Geochemistry*, 18(7), 955-972. doi:Doi 10.1016/S0883-2927(02)00205-6
- 1502
- 1503 Ibanez, M., Sancho, J. V., Pozo, O. J., & Hernandez, F. (2006). Use of liquid chromatography
1504 quadrupole time-of-flight mass spectrometry in the elucidation of transformation products
1505 and metabolites of pesticides. Diazinon as a case study. *Anal Bioanal Chem*, 384(2), 448-457.
1506 doi:10.1007/s00216-005-0167-6
- 1507 Imfeld, G., Braeckevelt, M., Kuschik, P., & Richnow, H. H. (2009). Monitoring and assessing processes
1508 of organic chemicals removal in constructed wetlands. *Chemosphere*, 74(3), 349-362.
1509 doi:10.1016/j.chemosphere.2008.09.062
- 1510 Imfeld, G., Payraudeau, S., Tournebize, J., Sauvage, S., Macary, F., Chaumont, C., . . . Probst, J. L.
1511 (2021). The Role of Ponds in Pesticide Dissipation at the Agricultural Catchment Scale: A
1512 Critical Review. *Water*, 13(9), 1202. doi:ARTN 1202
- 1513 10.3390/w13091202
- 1514 Inao, K., & Kitamura, Y. (1999). Pesticide paddy field model (PADDY) for predicting pesticide
1515 concentrations in water and soil in paddy fields. *Pesticide Science*, 55(1), 38-46. doi:DOI
1516 10.1002/ps.2780550108
- 1517 Jacobs, C. M. J., & Adriaanse, P. (2012). *Pesticide volatilization from small surface waters: rationale of*
1518 *a new parameterization for TOXSWA (1566-7197)*. Retrieved from
1519 <https://edepot.wur.nl/213995>
- 1520 Jamieson, R. C., Joy, D. M., Lee, H., Kostaschuk, R., & Gordon, R. J. (2005). Resuspension of sediment-
1521 associated *Escherichia coli* in a natural stream. *J Environ Qual*, 34(2), 581-589.
1522 doi:10.2134/jeq2005.0581
- 1523 Johnson, A., Llewellyn, N., Smith, J., van der Gast, C., Lilley, A., Singer, A., & Thompson, I. (2004). The
1524 role of microbial community composition and groundwater chemistry in determining
1525 isoproturon degradation potential in UK aquifers. *FEMS Microbiol Ecol*, 49(1), 71-82.
1526 doi:10.1016/j.femsec.2004.03.015
- 1527 Jorda, H., Huber, K., Kunkel, A., Vanderborght, J., Javaux, M., Oberdorster, C., . . . Schnepf, A. (2021).
1528 Mechanistic modeling of pesticide uptake with a 3D plant architecture model. *Environ Sci*
1529 *Pollut Res Int*, 28(39), 55678-55689. doi:10.1007/s11356-021-14878-3
- 1530 Kadlec, R. H., & Wallace, S. (2008). *Treatment wetlands*: CRC press.
- 1531 Kah, M., & Brown, C. D. (2006). Adsorption of Ionisable Pesticides in Soils. In G. W. Ware, D. M.
1532 Whitacre, L. A. Albert, P. de Voogt, C. P. Gerba, O. Hutzinger, J. B. Knaak, F. L. Mayer, D. P.
1533 Morgan, D. L. Park, R. S. Tjeerdema, R. S. H. Yang, & F. A. Gunther (Eds.), *Reviews of*
1534 *Environmental Contamination and Toxicology: Continuation of Residue Reviews* (pp. 149-
1535 217). New York, NY: Springer New York.
- 1536 Kalin, L., & Hantush, M. M. (2017). Special Issue on Advances in Wetland Hydrology and Water
1537 Quality Processes Modeling. *Journal of Hydrologic Engineering*, 22(1), D2016001.
1538 doi:10.1061/(asce)he.1943-5584.0001483

- 1539 Kang, D., Doudrick, K., Park, N., Choi, Y., Kim, K., & Jeon, J. (2020). Identification of transformation
1540 products to characterize the ability of a natural wetland to degrade synthetic organic
1541 pollutants. *Water Res*, 187, 116425. doi:10.1016/j.watres.2020.116425
- 1542 Karpuzcu, M. E., Sedlak, D. L., & Stringfellow, W. T. (2013). Biotransformation of chlorpyrifos in
1543 riparian wetlands in agricultural watersheds: implications for wetland management. *J Hazard*
1544 *Mater*, 244-245, 111-120. doi:10.1016/j.jhazmat.2012.11.047
- 1545 Kasak, K., Kill, K., Parn, J., & Mander, U. (2018). Efficiency of a newly established in-stream
1546 constructed wetland treating diffuse agricultural pollution. *Ecological Engineering*, 119, 1-7.
1547 doi:10.1016/j.ecoleng.2018.05.015
- 1548 Katagi, T. (2006). Behavior of pesticides in water-sediment systems. *Reviews of Environmental*
1549 *Contamination and Toxicology*, Vol 187, 187, 133-251. doi:Doi 10.1007/0-387-32885-8_4
- 1550 Katagi, T. (2013). Aerobic microbial transformation of pesticides in surface water. *Journal of Pesticide*
1551 *Science*, 38(1-2), 10-26. doi:10.1584/jpestics.D12-053
- 1552 Katagi, T., & Tanaka, H. (2016). Metabolism, bioaccumulation, and toxicity of pesticides in aquatic
1553 insect larvae. *J Pestic Sci*, 41(2), 25-37. doi:10.1584/jpestics.D15-064
- 1554 Kaur, P., & Kaur, P. (2018). Time and temperature dependent adsorption-desorption behaviour of
1555 pretilachlor in soil. *Ecotoxicol Environ Saf*, 161, 145-155. doi:10.1016/j.ecoenv.2018.05.081
- 1556 Kim, S., Chen, J., Cheng, T., Gindulyte, A., He, J., He, S., . . . Bolton, E. E. (2021). PubChem in 2021:
1557 new data content and improved web interfaces. *Nucleic Acids Res*, 49(D1), D1388-D1395.
1558 doi:10.1093/nar/gkaa971
- 1559 Kipkoech, W. (2019). *Profiling of pesticide residues in Mobego-Kabianga wetland ecosystem and its*
1560 *retention efficiency*. University of Kabianga.
- 1561 Kjellin, J., Hallin, S., & Worman, A. (2007). Spatial variations in denitrification activity in wetland
1562 sediments explained by hydrology and denitrifying community structure. *Water Res*, 41(20),
1563 4710-4720. doi:10.1016/j.watres.2007.06.053
- 1564 Koskiaho, J. (2003). Flow velocity retardation and sediment retention in two constructed wetland-
1565 ponds. *Ecological Engineering*, 19(5), 325-337. doi:Pii S0925-8574(02)00119-2
- 1566 Doi 10.1016/S0925-8574(02)00119-2
- 1567 Koskinen, W., O'Connor, G., & Cheng, H. (1979). Characterization of hysteresis in the desorption of 2,
1568 4, 5-T from soils. *Soil Science Society of America Journal*, 43(5), 871-874.
1569 doi:<https://doi.org/10.2136/sssaj1979.03615995004300050010x>
- 1570 Krishna, K. R., & Philip, L. (2008). Adsorption and desorption characteristics of lindane, carbofuran
1571 and methyl parathion on various Indian soils. *Journal of hazardous materials*, 160(2-3), 559-
1572 567. doi:10.1016/j.jhazmat.2008.03.107
- 1573 Laabs, V., Wehrhan, A., Pinto, A., Dores, E., & Amelung, W. (2007). Pesticide fate in tropical wetlands
1574 of Brazil: an aquatic microcosm study under semi-field conditions. *Chemosphere*, 67(5), 975-
1575 989. doi:10.1016/j.chemosphere.2006.10.067
- 1576 Lagunas-Allué, L., Martínez-Soria, M.-T., Sanz-Asensio, J., Salvador, A., Ferronato, C., & Chovelon, J.
1577 M. (2010). Photocatalytic degradation of boscalid in aqueous titanium dioxide suspension:
1578 Identification of intermediates and degradation pathways. *Applied Catalysis B:*
1579 *Environmental*, 98(3-4), 122-131. doi:<https://doi.org/10.1016/j.apcatb.2010.05.020>
- 1580 Lam, M. W., Tantuco, K., & Mabury, S. A. (2003). PhotoFate: A new approach in accounting for the
1581 contribution of indirect photolysis of pesticides and pharmaceuticals in surface waters.
1582 *Environmental Science & Technology*, 37(5), 899-907. doi:10.1021/es025902+
- 1583 Lange, J., Schuetz, T., Gregoire, C., Elsasser, D., Schulz, R., Passeur, E., & Tournebize, J. (2011).
1584 Multi-tracer experiments to characterise contaminant mitigation capacities for different
1585 types of artificial wetlands. *International journal of environmental analytical chemistry*, 91(7-
1586 8), 768-785. doi:10.1080/03067319.2010.525635
- 1587 Langergraber, G. (2008). Modeling of processes in subsurface flow constructed wetlands: A review.
1588 *Vadose Zone Journal*, 7(2), 830-842. doi:10.2136/vzj2007.0054
- 1589 Larson, R. A., & Weber, E. J. (1994). *Reaction mechanisms in environmental organic chemistry*: CRC
1590 press.
- 1591 Le Cor, F., Slaby, S., Dufour, V., Iuretig, A., Feidt, C., Dauchy, X., & Banas, D. (2021). Occurrence of
1592 pesticides and their transformation products in headwater streams: Contamination status

- 1593 and effect of ponds on contaminant concentrations. *Science of the Total Environment*, 788,
1594 147715. doi:10.1016/j.scitotenv.2021.147715
- 1595 Le Person, A., Siampiringue, M., Sarakha, M., Moncomble, A., & Cornard, J.-P. (2016). The photo-
1596 degradation of mesotrione, a triketone herbicide, in the presence of Cu II ions. *Journal of*
1597 *Photochemistry and Photobiology A: Chemistry*, 315, 76-86.
1598 doi:10.1016/j.jphotochem.2015.09.010
- 1599 Lee, S., Gan, J., Kim, J. S., Kabashima, J. N., & Crowley, D. E. (2004). Microbial transformation of
1600 pyrethroid insecticides in aqueous and sediment phases. *Environ Toxicol Chem*, 23(1), 1-6.
1601 doi:10.1897/03-114
- 1602 Lewis, K. A., Tzilivakis, J., Warner, D. J., & Green, A. (2016). An international database for pesticide
1603 risk assessments and management. *Human and Ecological Risk Assessment*, 22(4), 1050-
1604 1064. doi:10.1080/10807039.2015.1133242
- 1605 Li, H., Sheng, G., Teppen, B. J., Johnston, C. T., & Boyd, S. A. (2003). Sorption and Desorption of
1606 Pesticides by Clay Minerals and Humic Acid-Clay Complexes. *Soil Science Society of America*
1607 *Journal*, 67(1), 122-131. doi:10.2136/sssaj2003.1220a
- 1608 Li, J. S., Fei, L. J., Chen, Z., & Sun, X. L. (2017). Particle size distribution and settling velocity of
1609 sediments in water diverted from the Yellow River during border-strip irrigation. *Tecnología y*
1610 *ciencias del agua*, 8(2), 31-41. doi:DOI 10.24850/j-tyca-2017-02-03
- 1611 Limousin, G., Gaudet, J. P., Charlet, L., Szenknect, S., Barthes, V., & Krimissa, M. (2007). Sorption
1612 isotherms: A review on physical bases, modeling and measurement. *Applied Geochemistry*,
1613 22(2), 249-275. doi:10.1016/j.apgeochem.2006.09.010
- 1614 Lin, J. X., & Wang, L. (2009). Comparison between linear and non-linear forms of pseudo-first-order
1615 and pseudo-second-order adsorption kinetic models for the removal of methylene blue by
1616 activated carbon. *Frontiers of Environmental Science & Engineering in China*, 3(3), 320-324.
1617 doi:10.1007/s11783-009-0030-7
- 1618 Lin, K. D., & Yuan, D. X. (2005). Degradation kinetics and products of triazophos in intertidal
1619 sediment. *J Environ Sci (China)*, 17(6), 933-936.
- 1620 Liu, B., McConnell, L. L., & Torrents, A. (2001). Hydrolysis of chlorpyrifos in natural waters of the
1621 Chesapeake Bay. *Chemosphere*, 44(6), 1315-1323. doi:10.1016/s0045-6535(00)00506-3
- 1622 Liu, T., Xu, S., Lu, S., Qin, P., Bi, B., Ding, H., . . . Liu, X. (2019). A review on removal of
1623 organophosphorus pesticides in constructed wetland: Performance, mechanism and
1624 influencing factors. *Science of the Total Environment*, 651(Pt 2), 2247-2268.
1625 doi:10.1016/j.scitotenv.2018.10.087
- 1626 Llorens, E., Saaltink, M. W., Poch, M., & Garcia, J. (2011). Bacterial transformation and
1627 biodegradation processes simulation in horizontal subsurface flow constructed wetlands
1628 using CWM1-RETRASO. *Bioresour Technol*, 102(2), 928-936.
1629 doi:10.1016/j.biortech.2010.09.038
- 1630 Luo, J., Huai, W., & Wang, P. (2016). Contaminant transport in a three-zone wetland: Dispersion and
1631 ecological degradation. *Journal of Hydrology*, 534, 341-351.
1632 doi:<https://doi.org/10.1016/j.jhydrol.2016.01.008>
- 1633 Luo, J. P., Ma, M., Liu, C., Zha, J. M., & Wang, Z. J. (2009). Impacts of particulate organic carbon and
1634 dissolved organic carbon on removal of polycyclic aromatic hydrocarbons, organochlorine
1635 pesticides, and nonylphenols in a wetland. *Journal of Soils and Sediments*, 9(3), 180-187.
1636 doi:10.1007/s11368-009-0081-1
- 1637 Lv, T., Carvalho, P. N., Zhang, L., Zhang, Y., Button, M., Arias, C. A., . . . Brix, H. (2017). Functionality of
1638 microbial communities in constructed wetlands used for pesticide remediation: Influence of
1639 system design and sampling strategy. *Water Res*, 110, 241-251.
1640 doi:10.1016/j.watres.2016.12.021
- 1641 Lyu, T., Zhang, L., Xu, X., Arias, C. A., Brix, H., & Carvalho, P. N. (2018). Removal of the pesticide
1642 tebuconazole in constructed wetlands: Design comparison, influencing factors and modelling.
1643 *Environ Pollut*, 233, 71-80. doi:10.1016/j.envpol.2017.10.040
- 1644 Ma, Y., Li, J., Wu, J., Kong, Z., Feinstein, L. M., Ding, X., . . . Wu, L. (2018). Bacterial and Fungal
1645 Community Composition and Functional Activity Associated with Lake Wetland Water Level
1646 Gradients. *Sci Rep*, 8(1), 760. doi:10.1038/s41598-018-19153-z

- 1647 MacIntyre, M. E., Warner, B. G., & Slawson, R. M. (2006). Escherichia coli control in a surface flow
1648 treatment wetland. *J Water Health*, 4(2), 211-214.
- 1649 Mackay, D., Shiu, W.-Y., & Lee, S. C. (2006). *Handbook of physical-chemical properties and*
1650 *environmental fate for organic chemicals*: CRC press.
- 1651 Mahabali, S., & Spanoghe, P. (2014). Mitigation of Two Insecticides by Wetland Plants: Feasibility
1652 Study for the Treatment of Agricultural Runoff in Suriname (South America). *Water Air and*
1653 *Soil Pollution*, 225(1), 1771. doi:ARTN 1771
- 1654 10.1007/s11270-013-1771-2
- 1655 Maillard, E., & Imfeld, G. (2014). Pesticide mass budget in a stormwater wetland. *Environ Sci Technol*,
1656 48(15), 8603-8611. doi:10.1021/es500586x
- 1657 Maillard, E., Payraudeau, S., Faivre, E., Gregoire, C., Gangloff, S., & Imfeld, G. (2011). Removal of
1658 pesticide mixtures in a stormwater wetland collecting runoff from a vineyard catchment.
1659 *Science of the Total Environment*, 409(11), 2317-2324. doi:10.1016/j.scitotenv.2011.01.057
- 1660 Maillard, E., Payraudeau, S., Ortiz, F., & Imfeld, G. (2012). Removal of dissolved pesticide mixtures by
1661 a stormwater wetland receiving runoff from a vineyard catchment: an inter-annual
1662 comparison. *International journal of environmental analytical chemistry*, 92(8), 979-994.
1663 doi:10.1080/03067319.2011.609935
- 1664 Maltais-Landry, G., Maranger, R., Brisson, J., & Chazarenc, F. (2009). Nitrogen transformations and
1665 retention in planted and artificially aerated constructed wetlands. *Water Res*, 43(2), 535-545.
1666 doi:10.1016/j.watres.2008.10.040
- 1667 Mamy, L., & Barriuso, E. (2007). Desorption and time-dependent sorption of herbicides in soils.
1668 *European Journal of Soil Science*, 58(1), 174-187. doi:10.1111/j.1365-2389.2006.00822.x
- 1669 Marois, D. E., & Mitsch, W. J. (2016). Modeling phosphorus retention at low concentrations in Florida
1670 Everglades mesocosms. *Ecological Modelling*, 319, 42-62.
1671 doi:10.1016/j.ecolmodel.2015.09.024
- 1672 Martínez-Lavanchy, P., Chen, Z., Lünsmann, V., Marin-Cevada, V., Vilchez-Vargas, R., Pieper, D., . . .
1673 Junca, H. (2015). Microbial toluene removal in hypoxic model constructed wetlands occurs
1674 predominantly via the ring monooxygenation pathway. *Applied and Environmental*
1675 *Microbiology*, 81(18), 6241-6252. doi:10.1128/AEM.01822-15
- 1676 Martinez, C. J., & Wise, W. R. (2003). Analysis of constructed treatment wetland hydraulics with the
1677 transient storage model OTIS. *Ecological Engineering*, 20(3), 211-222.
1678 doi:[https://doi.org/10.1016/S0925-8574\(03\)00029-6](https://doi.org/10.1016/S0925-8574(03)00029-6)
- 1679 Martinez, C. J., & Wise, W. R. (2003). Hydraulic analysis of Orlando Easterly Wetland. *Journal of*
1680 *Environmental Engineering*, 129(6), 553-560. doi:10.1061/(Asce)0733-9372(2003)129:6(553)
- 1681 Masbou, J., Drouin, G., Payraudeau, S., & Imfeld, G. (2018). Carbon and nitrogen stable isotope
1682 fractionation during abiotic hydrolysis of pesticides. *Chemosphere*, 213, 368-376.
1683 doi:10.1016/j.chemosphere.2018.09.056
- 1684 Matamoros, V., & Bayona, J. M. (2006). Elimination of pharmaceuticals and personal care products in
1685 subsurface flow constructed wetlands. *Environ Sci Technol*, 40(18), 5811-5816.
1686 doi:10.1021/es0607741
- 1687 Mathon, B., Coquery, M., Miegé, C., Vandycke, A., & Choubert, J. M. (2019). Influence of water depth
1688 and season on the photodegradation of micropollutants in a free-water surface constructed
1689 wetland receiving treated wastewater. *Chemosphere*, 235, 260-270.
1690 doi:10.1016/j.chemosphere.2019.06.140
- 1691 Maynard, J. (2009). *Spatial and temporal dynamics of carbon and nutrient cycling in a eutrophic flow-*
1692 *through wetland*. Ph. D. dissertation. University of California, Davis.
- 1693 McBride, M. (1994). *Environmental Chemistry of Soils 1994*: Oxford University Press: New York.
- 1694 McBride, M. B. (1994). *Environmental Chemistry of Soils* (Vol. 141580): Obtenido de Recuperado
1695 de: [https://d1wqtxts1xzle7.cloudfront.net/35536696](https://d1wqtxts1xzle7.cloudfront.net/35536696....)
- 1696 Mccall, P. J., & Agin, G. L. (1985). Desorption-Kinetics of Picloram as Affected by Residence Time in
1697 the Soil. *Environmental Toxicology and Chemistry*, 4(1), 37-44. doi:10.1002/etc.5620040106
- 1698 Mckay, G. (1995). *Use of Adsorbents for the Removal of Pollutants from Wastewater*: CRC press.

- 1699 Meng, P. P., Pei, H. Y., Hu, W. R., Shao, Y. Y., & Li, Z. (2014). How to increase microbial degradation in
1700 constructed wetlands: Influencing factors and improvement measures. *Bioresource*
1701 *Technology*, *157*, 316-326. doi:10.1016/j.biortech.2014.01.095
- 1702 Miller, P. L., & Chin, Y. P. (2005). Indirect photolysis promoted by natural and engineered wetland
1703 water constituents: processes leading to alachlor degradation. *Environ Sci Technol*, *39*(12),
1704 4454-4462. doi:10.1021/es049111e
- 1705 Moore, M. T., Cooper, C. M., Smith, S., Cullum, R. F., Knight, S. S., Locke, M. A., & Bennett, E. R.
1706 (2007). Diazinon mitigation in constructed Wetlands: Influence of vegetation. *Water Air and*
1707 *Soil Pollution*, *184*(1-4), 313-321. doi:10.1007/s11270-007-9418-9
- 1708 Mugni, H., Demetrio, P., Bulus, G., Ronco, A., & Bonetto, C. (2011). Effect of aquatic vegetation on
1709 the persistence of cypermethrin toxicity in water. *Bull Environ Contam Toxicol*, *86*(1), 23-27.
1710 doi:10.1007/s00128-010-0143-5
- 1711 Mulligan, R. A., Tomco, P. L., Howard, M. W., Schempp, T. T., Stewart, D. J., Stacey, P. M., . . .
1712 Tjeerdema, R. S. (2016). Aerobic versus Anaerobic Microbial Degradation of Clothianidin
1713 under Simulated California Rice Field Conditions. *J Agric Food Chem*, *64*(38), 7059-7067.
1714 doi:10.1021/acs.jafc.6b02055
- 1715 Nakano, Y., Yoshida, T., & Inoue, T. (2004). A study on pesticide runoff from paddy fields to a river in
1716 rural region--2: development and application of a mathematical model. *Water Res*, *38*(13),
1717 3023-3030. doi:10.1016/j.watres.2004.02.014
- 1718 O'Geen, A. T., Budd, R., Gan, J., Maynard, J. J., Parikh, S. J., & Dahlgren, R. A. (2010). Chapter One -
1719 Mitigating Nonpoint Source Pollution in Agriculture with Constructed and Restored
1720 Wetlands. In D. L. Sparks (Ed.), *Advances in agronomy* (Vol. 108, pp. 1-76): Academic Press.
- 1721 OCDE. (2000). *Test No. 106: Adsorption -- Desorption Using a Batch Equilibrium Method*.
- 1722 Olvera-Velona, A., Benoit, P., Barriuso, E., & Ortiz-Hernandez, L. (2008). Sorption and desorption of
1723 organophosphate pesticides, parathion and cadusafos, on tropical agricultural soils.
1724 *Agronomy for Sustainable Development*, *28*(2), 231-238. doi:10.1051/agro:2008009
- 1725 Ortega-Calvo, J. J., & Saiz-Jimenez, C. (1998). Effect of humic fractions and clay on biodegradation of
1726 phenanthrene by a *Pseudomonas fluorescens* strain isolated from soil. *Appl Environ*
1727 *Microbiol*, *64*(8), 3123-3126. doi:10.1128/AEM.64.8.3123-3126.1998
- 1728 Papaevangelou, V. A., Gikas, G. D., Vryzas, Z., & Tshirintzis, V. A. (2017). Treatment of agricultural
1729 equipment rinsing water containing a fungicide in pilot-scale horizontal subsurface flow
1730 constructed wetlands. *Ecological Engineering*, *101*, 193-200.
1731 doi:10.1016/j.ecoleng.2017.01.045
- 1732 Papp, R. (2009). Les propriétés environnementales des substances. *Collège National d'Experts en*
1733 *Environnement de l'Industrie Chimique*, *126*.
- 1734 Passeport, E., Benoit, P., Bergheaud, V., Coquet, Y., & Tournebize, J. (2011a). Epoxiconazole
1735 degradation from artificial wetland and forest buffer substrates under flooded conditions.
1736 *Chemical Engineering Journal*, *173*(3), 760-765. doi:10.1016/j.cej.2011.08.044
- 1737 Passeport, E., Benoit, P., Bergheaud, V., Coquet, Y., & Tournebize, J. (2011b). Selected pesticides
1738 adsorption and desorption in substrates from artificial wetland and forest buffer. *Environ*
1739 *Toxicol Chem*, *30*(7), 1669-1676. doi:10.1002/etc.554
- 1740 Passeport, E., Tournebize, J., Chaumont, C., Guenne, A., & Coquet, Y. (2013). Pesticide contamination
1741 interception strategy and removal efficiency in forest buffer and artificial wetland in a tile-
1742 drained agricultural watershed. *Chemosphere*, *91*(9), 1289-1296.
1743 doi:10.1016/j.chemosphere.2013.02.053
- 1744 Passeport, E., Tournebize, J., Jankowfsky, S., Promse, B., Chaumont, C., Coquet, Y., & Lange, J. (2010).
1745 Artificial Wetland and Forest Buffer Zone: Hydraulic and Tracer Characterization. *Vadose*
1746 *Zone Journal*, *9*(1), 73-84. doi:10.2136/vzj2008.0164
- 1747 Paszko, T., Muszynski, P., Materska, M., Bojanowska, M., Kostecka, M., & Jackowska, I. (2016).
1748 Adsorption and degradation of phenoxyalkanoic acid herbicides in soils: A review. *Environ*
1749 *Toxicol Chem*, *35*(2), 271-286. doi:10.1002/etc.3212
- 1750 Pattanayak, S., Chakraborty, S., Biswas, S., Chattopadhyay, D., & Chakraborty, M. (2018). Degradation
1751 of Methyl Parathion, a common pesticide and fluorescence quenching of Rhodamine B, a
1752 carcinogen using β -d glucan stabilized gold nanoparticles. *Journal of Saudi Chemical Society*,
1753 *22*(8), 937-948. doi:10.1016/j.jscs.2018.02.004

- 1754 Pignatello, J. (1989). Sorption dynamics of organic compounds in soils and sediments. *Reactions and*
 1755 *movement of organic chemicals in soils*, 22, 45-80. doi:10.2136/sssaspecpub22.c3
- 1756 Pignatello, J. J. (1999). The measurement and interpretation of sorption and desorption rates for
 1757 organic compounds in soil media. *Advances in agronomy*, 69, 1-73.
 1758 doi:[https://doi.org/10.1016/S0065-2113\(08\)60946-3](https://doi.org/10.1016/S0065-2113(08)60946-3)
- 1759 Pignatello, J. J., & Xing, B. S. (1996). Mechanisms of slow sorption of organic chemicals to natural
 1760 particles. *Environmental Science & Technology*, 30(1), 1-11. doi:DOI 10.1021/es940683g
- 1761 Pohlman, A. A., & Mill, T. (1983). Peroxy Radical Interactions with Soil Constituents. *Soil Science*
 1762 *Society of America Journal*, 47(5), 922-927. doi:DOI
 1763 10.2136/sssaj1983.03615995004700050016x
- 1764 Poissant, L., Beauvais, C., Lafrance, P., & Deblois, C. (2008). Pesticides in fluvial wetlands catchments
 1765 under intensive agricultural activities. *Science of the Total Environment*, 404(1), 182-195.
 1766 doi:10.1016/j.scitotenv.2008.05.030
- 1767 Prochaska, C. A., Zouboulis, A. I., & Eskridge, K. M. (2007). Performance of pilot-scale vertical-flow
 1768 constructed wetlands, as affected by season, substrate, hydraulic load and frequency of
 1769 application of simulated urban sewage. *Ecological Engineering*, 31(1), 57-66.
 1770 doi:10.1016/j.ecoleng.2007.05.007
- 1771 PSD. (2005). Pesticide Data Requirement Handbook. Retrieved from <http://sitem.herts.ac.uk/aeru/ppdb/>
- 1772 PubChem. (2021). National Institutes of Health (NIH). *National Center for Biotechnology Information*.
 1773 Retrieved from <https://pubchem.ncbi.nlm.nih.gov/search/search.cgi>
- 1774 Rai, P. K. (2009). Heavy Metal Phytoremediation from Aquatic Ecosystems with Special Reference to
 1775 Macrophytes. *Critical Reviews in Environmental Science and Technology*, 39(9), 697-753.
 1776 doi:Pii 914514507
 1777 10.1080/10643380801910058
- 1778 Rani, S., Madan, V. K., & Kathpal, T. S. (2001). Persistence and dissipation behavior of triazophos in
 1779 canal water under Indian climatic conditions. *Ecotoxicol Environ Saf*, 50(1), 82-84.
 1780 doi:10.1006/eesa.2001.2103
- 1781 Rayen, F., Behnam, T., & Dominique, P. (2019). Optimization of a raceway pond system for
 1782 wastewater treatment: a review. *Crit Rev Biotechnol*, 39(3), 422-435.
 1783 doi:10.1080/07388551.2019.1571007
- 1784 Reddy, K. R., & DeLaune, R. D. (2008). *Biogeochemistry of wetlands: science and applications*: CRC
 1785 press.
- 1786 Rijn, L. C. v. (1984). Sediment transport, part II: suspended load transport. *Journal of Hydraulic*
 1787 *Engineering*, 110(11), 1613-1641. doi:[https://doi.org/10.1061/\(ASCE\)0733-9429\(1984\)110:11\(1613\)](https://doi.org/10.1061/(ASCE)0733-9429(1984)110:11(1613))
- 1788 Rogers, M. R., & Stringfellow, W. T. (2009). Partitioning of chlorpyrifos to soil and plants in vegetated
 1789 agricultural drainage ditches. *Chemosphere*, 75(1), 109-114.
 1790 doi:10.1016/j.chemosphere.2008.11.036
- 1791 Rong, X., Zhao, G., Fein, J. B., Yu, Q., & Huang, Q. (2019). Role of interfacial reactions in
 1792 biodegradation: A case study in a montmorillonite, *Pseudomonas* sp. Z1 and methyl
 1793 parathion ternary system. *J Hazard Mater*, 365, 245-251. doi:10.1016/j.jhazmat.2018.11.019
- 1794 Rose, M. T., Crossan, A. N., & Kennedy, I. R. (2008). The effect of vegetation on pesticide dissipation
 1795 from ponded treatment wetlands: quantification using a simple model. *Chemosphere*, 72(7),
 1796 999-1005. doi:10.1016/j.chemosphere.2008.04.059
- 1797 Rose, M. T., Sanchez-Bayo, F., Crossan, A. N., & Kennedy, I. R. (2006). Pesticide removal from cotton
 1798 farm tailwater by a pilot-scale ponded wetland. *Chemosphere*, 63(11), 1849-1858.
 1799 doi:10.1016/j.chemosphere.2005.10.024
- 1800 Ruiz-Medina, A., Soler-Gallardo, M. I., & Llorent-Martinez, E. J. (2018). Enhanced Quenching Effect of
 1801 Neonicotinoid Pesticides on Time-Resolved Terbium Luminescence in Presence of
 1802 Surfactants. *Journal of Chemistry*, 2018, 5378736. doi:Artn 5378736
 1803 10.1155/2018/5378736
 1804

- 1806 Saaristo, M., Brodin, T., Balshine, S., Bertram, M. G., Brooks, B. W., Ehlman, S. M., . . . Arnold, K. E.
1807 (2018). Direct and indirect effects of chemical contaminants on the behaviour, ecology and
1808 evolution of wildlife. *Proc Biol Sci*, 285(1885), 20181297. doi:10.1098/rspb.2018.1297
- 1809 Sabadie, J. (2002). Nicosulfuron: alcoholysis, chemical hydrolysis, and degradation on various
1810 minerals. *J Agric Food Chem*, 50(3), 526-531. doi:10.1021/jf010873s
- 1811 Saha, S., & Kulshrestha, G. (2002). Degradation of sulfosulfuron, a sulfonylurea herbicide, as
1812 influenced by abiotic factors. *Journal of agricultural and food chemistry*, 50(16), 4572-4575.
1813 doi:10.1021/jf0116653
- 1814 Sahin, C., & Karpuzcu, M. E. (2020). Mitigation of organophosphate pesticide pollution in agricultural
1815 watersheds. *Science of the Total Environment*, 710, 136261.
1816 doi:10.1016/j.scitotenv.2019.136261
- 1817 Sakellarides, T. M., Siskos, M. G., & Albanis, T. A. (2003). Photodegradation of selected
1818 organophosphorus insecticides under sunlight in different natural waters and soils.
1819 *International journal of environmental analytical chemistry*, 83(1), 33-50.
1820 doi:10.1080/0306731031000104227
- 1821 Samsó, R., & Garcia, J. (2013). Bacteria distribution and dynamics in constructed wetlands based on
1822 modelling results. *Science of the Total Environment*, 461-462, 430-440.
1823 doi:10.1016/j.scitotenv.2013.04.073
- 1824 Sardar, D., & Kole, R. K. (2005). Metabolism of chlorpyrifos in relation to its effect on the availability
1825 of some plant nutrients in soil. *Chemosphere*, 61(9), 1273-1280.
1826 doi:10.1016/j.chemosphere.2005.03.078
- 1827 Saripalli, P. (1997). *Fate and Transport of Organic Chemicals in the Environment: A Practical Guide*:
1828 JSTOR.
- 1829 Schmid, B. H., Stephan, U., & Hengl, M. A. (2005). Sediment deposition in constructed wetland ponds
1830 with emergent vegetation: laboratory study and mathematical model. *Water Sci Technol*,
1831 51(9), 307-314. doi:10.2166/wst.2005.0342
- 1832 Schulz, R., & Peall, S. K. (2001). Effectiveness of a constructed wetland for retention of nonpoint-
1833 source pesticide pollution in the Lourens River catchment, South Africa. *Environ Sci Technol*,
1834 35(2), 422-426. doi:10.1021/es0001198
- 1835 Scow, K. M., & Johnson, C. R. (1996). Effect of sorption on biodegradation of soil pollutants. *Advances*
1836 *in agronomy*, 58, 1-56. doi:[https://doi.org/10.1016/S0065-2113\(08\)60252-7](https://doi.org/10.1016/S0065-2113(08)60252-7)
- 1837 Sereshti, H., Zarei-Hosseiniabadi, M., Soltani, S., Jamshidi, F., & AliAbadi, M. H. S. (2021). Hydrophobic
1838 liquid-polymer-based deep eutectic solvent for extraction and multi-residue analysis of
1839 pesticides in water samples. *Microchemical Journal*, 167, 106314. doi:ARTN 106314
1840 10.1016/j.microc.2021.106314
- 1841 Sharifi, A., Kalin, L., Hantush, M. M., Isik, S., & Jordan, T. E. (2013). Carbon dynamics and export from
1842 flooded wetlands: A modeling approach. *Ecological Modelling*, 263, 196-210.
1843 doi:10.1016/j.ecolmodel.2013.04.023
- 1844 Shilton, A., & Harrison, J. (2003). Development of guidelines for improved hydraulic design of waste
1845 stabilisation ponds. *Water Sci Technol*, 48(2), 173-180. doi:10.2166/wst.2003.0114
- 1846 Singh, N. C., Dasgupta, T. P., Roberts, E. V., & Mansingh, A. (1991). Dynamics of pesticides in tropical
1847 conditions. 1. Kinetic studies of volatilization, hydrolysis, and photolysis of dieldrin and
1848 alpha.-and. beta.-endosulfan. *Journal of agricultural and food chemistry*, 39(3), 575-579.
1849 doi:10.1021/jf00003a028
- 1850 Singh, T., Awasthi, G., & Tiwari, Y. (2021). Recruiting endophytic bacteria of wetland plants to
1851 phytoremediate organic pollutants. *International Journal of Environmental Science and*
1852 *Technology*. doi:10.1007/s13762-021-03476-y
- 1853 Siobhan Fennessy, M., Brueske, C. C., & Mitsch, W. J. (1994). Sediment deposition patterns in
1854 restored freshwater wetlands using sediment traps. *Ecological Engineering*, 3(4), 409-428.
1855 doi:10.1016/0925-8574(94)00010-7
- 1856 Smith, K. E., Thullner, M., Wick, L. Y., & Harms, H. (2009). Sorption to humic acids enhances polycyclic
1857 aromatic hydrocarbon biodegradation. *Environ Sci Technol*, 43(19), 7205-7211.
1858 doi:10.1021/es803661s

- 1859 Song, B., Jiang, X., Liu, X., Deng, Y., Hu, D., & Lu, P. (2021). Dissipation and sorption-desorption of
1860 benzoisothiazolinone in agricultural soils and identification of its metabolites. *RSC Adv*, *11*(10),
1861 5399-5410. doi:10.1039/d0ra09553b
- 1862 Sørensen, S. R., Schultz, A., Jacobsen, O. S., & Aamand, J. (2006). Sorption, desorption and
1863 mineralisation of the herbicides glyphosate and MCPA in samples from two Danish soil and
1864 subsurface profiles. *Environmental Pollution*, *141*(1), 184-194.
1865 doi:<https://doi.org/10.1016/j.envpol.2005.07.023>
- 1866 Sparks, D. (2000). *Kinetics and mechanisms of soil chemical reactions*.
- 1867 Sposito, G. (2008). *The chemistry of soils*: Oxford university press.
- 1868 Stang, C., Bakanov, N., & Schulz, R. (2016). Experiments in water-macrophyte systems to uncover the
1869 dynamics of pesticide mitigation processes in vegetated surface waters/streams. *Environ Sci*
1870 *Pollut Res Int*, *23*(1), 673-682. doi:10.1007/s11356-015-5274-0
- 1871 Stangroom, S. J., Collins, C. D., & Lester, J. N. (2000). Abiotic behaviour of organic micropollutants in
1872 soils and the aquatic environment. A review: II. Transformation. *Environmental technology*,
1873 *21*(8), 865-882. doi:Doi 10.1080/09593332108618059
- 1874 Stehle, S., Elsaesser, D., Gregoire, C., Imfeld, G., Niehaus, E., Passet, E., . . . Schulz, R. (2011).
1875 Pesticide risk mitigation by vegetated treatment systems: a meta-analysis. *J Environ Qual*,
1876 *40*(4), 1068-1080. doi:10.2134/jeq2010.0510
- 1877 Stottmeister, U., Wiessner, A., Kusch, P., Kappelmeyer, U., Kastner, M., Bederski, O., . . . Moormann,
1878 H. (2003). Effects of plants and microorganisms in constructed wetlands for wastewater
1879 treatment. *Biotechnol Adv*, *22*(1-2), 93-117. doi:10.1016/j.biotechadv.2003.08.010
- 1880 Strawn, D., & Sparks, D. (1999). Sorption kinetics of trace elements in soils and soil materials. p. 1–28.
1881 HM Selim and KI Iskandar (ed.) Fate and transport of heavy metals in the vadose zone. Lewis
1882 Publ., Boca Raton, FL. *Sorption kinetics of trace elements in soils and soil materials*. p. 1–28.
1883 In HM Selim and KI Iskandar (ed.) Fate and transport of heavy metals in the vadose zone.
1884 Lewis Publ., Boca Raton, FL., -.
- 1885 Sudarsan, J. S., & Nithyanantham, S. (2021). An Integrated Constructed Wetland System for Society
1886 (pp. 397-419): Springer International Publishing.
- 1887 Supowit, S. D., Sadaria, A. M., Reyes, E. J., & Halden, R. U. (2016). Mass Balance of Fipronil and Total
1888 Toxicity of Fipronil-Related Compounds in Process Streams during Conventional Wastewater
1889 and Wetland Treatment. *Environ Sci Technol*, *50*(3), 1519-1526. doi:10.1021/acs.est.5b04516
- 1890 Sur, R., & Stork, A. (2003). Uptake, translocation and metabolism of imidacloprid in plants. *Bulletin of*
1891 *insectology*, *56*, 35-40.
- 1892 Takagi, K., Fajardo, F. F., Ishizaka, M., Phong, T. K., Watanabe, H., & Boulange, J. (2012). Fate and
1893 transport of bensulfuron-methyl and imazosulfuron in paddy fields: experiments and model
1894 simulation. *Paddy and water environment*, *10*(2), 139-151. doi:10.1007/s10333-011-0276-0
- 1895 Tang, X., Zhu, B., & Katou, H. (2012). A review of rapid transport of pesticides from sloping farmland
1896 to surface waters: Processes and mitigation strategies. *Journal of Environmental Sciences*,
1897 *24*(3), 351-361. doi:[https://doi.org/10.1016/S1001-0742\(11\)60753-5](https://doi.org/10.1016/S1001-0742(11)60753-5)
- 1898 Tang, X. Y., Yang, Y., Huang, W. D., McBride, M. B., Guo, J. J., Tao, R., & Dai, Y. V. (2017).
1899 Transformation of chlorpyrifos in integrated recirculating constructed wetlands (IRCWs) as
1900 revealed by compound-specific stable isotope (CSIA) and microbial community structure
1901 analysis. *Bioresource Technology*, *233*, 264-270. doi:10.1016/j.biortech.2017.02.077
- 1902 Tang, X. Y., Yang, Y., McBride, M. B., Tao, R., Dai, Y. N., & Zhang, X. M. (2019). Removal of chlorpyrifos
1903 in recirculating vertical flow constructed wetlands with five wetland plant species.
1904 *Chemosphere*, *216*, 195-202. doi:10.1016/j.chemosphere.2018.10.150
- 1905 Tejeda-Agredano, M. C., Mayer, P., & Ortega-Calvo, J. J. (2014). The effect of humic acids on
1906 biodegradation of polycyclic aromatic hydrocarbons depends on the exposure regime.
1907 *Environ Pollut*, *184*, 435-442. doi:10.1016/j.envpol.2013.09.031
- 1908 Thomas, K. A., & Hand, L. H. (2011). Assessing the potential for algae and macrophytes to degrade
1909 crop protection products in aquatic ecosystems. *Environ Toxicol Chem*, *30*(3), 622-631.
1910 doi:10.1002/etc.412
- 1911 Thorstensen, C. W., Lode, O., Eklo, O. M., & Christiansen, A. (2001). Sorption of bentazone,
1912 dichlorprop, MCPA, and propiconazole in reference soils from Norway. *J Environ Qual*, *30*(6),
1913 2046-2052. doi:10.2134/jeq2001.2046

- 1914 Tomlin, C. D. (2009). *The pesticide manual: A world compendium*: British Crop Production Council.
- 1915 Torabi, E., Wiegert, C., Guyot, B., Vuilleumier, S., & Imfeld, G. (2020). Dissipation of S-metolachlor
1916 and butachlor in agricultural soils and responses of bacterial communities: Insights from
1917 compound-specific isotope and biomolecular analyses. *J Environ Sci (China)*, *92*, 163-175.
1918 doi:10.1016/j.jes.2020.02.009
- 1919 Tournebize, J., Chaumont, C., Fesneau, C., Guenne, A., Vincent, B., Garnier, J., & Mander, U. (2015).
1920 Long-term nitrate removal in a buffering pond-reservoir system receiving water from an
1921 agricultural drained catchment. *Ecological Engineering*, *80*, 32-45.
1922 doi:10.1016/j.ecoleng.2014.11.051
- 1923 Tournebize, J., Chaumont, C., & Mander, U. (2017). Implications for constructed wetlands to mitigate
1924 nitrate and pesticide pollution in agricultural drained watersheds. *Ecological Engineering*,
1925 *103*, 415-425. doi:10.1016/j.ecoleng.2016.02.014
- 1926 Tournebize, J., Passeport, E., Chaumont, C., Fesneau, C., Guenne, A., & Vincent, B. (2013). Pesticide
1927 de-contamination of surface waters as a wetland ecosystem service in agricultural
1928 landscapes. *Ecological Engineering*, *56*, 52-60. doi:10.1016/j.ecoleng.2012.06.001
- 1929 Tsavdaris, A., Mitchell, S., & Williams, J. (2013). Use of CFD to Model Emergent Vegetation in
1930 Detention Ponds. *ARPN Journal of Engineering and Applied Sciences (ISSN 1819-6608)*, *8*,
1931 495-503.
- 1932 U.S.G.S. (2020). U.S. Geological Survey. Retrieved from <https://www.usgs.gov/>
- 1933 Uchimiya, M., Wartelle, L. H., & Boddu, V. M. (2012). Sorption of triazine and organophosphorus
1934 pesticides on soil and biochar. *J Agric Food Chem*, *60*(12), 2989-2997. doi:10.1021/jf205110g
- 1935 Vagi, M. C., Petsas, A. S., Kostopoulou, M. N., & Lekkas, T. D. (2010). Adsorption and desorption
1936 processes of the organophosphorus pesticides, dimethoate and fenthion, onto three Greek
1937 agricultural soils. *International journal of environmental analytical chemistry*, *90*(3-6), 369-
1938 389. doi:10.1080/03067310903194980
- 1939 Vallée, R. (2015). *Efficacité de zones tampons humides à réduire les teneurs en pesticides des eaux de*
1940 *drainage*. Université de Lorraine. Retrieved from [https://hal.univ-lorraine.fr/tel-](https://hal.univ-lorraine.fr/tel-01751701/document)
1941 [01751701/document](https://hal.univ-lorraine.fr/tel-01751701/document)
- 1942 Vallee, R., Dousset, S., & Billet, D. (2016). Influence of substrate water saturation on pesticide
1943 dissipation in constructed wetlands. *Environ Sci Pollut Res Int*, *23*(1), 109-119.
1944 doi:10.1007/s11356-015-4367-0
- 1945 Vallet, B. S. (2011). *Modélisation d'un bassin d'orage en vue de l'amélioration de la qualité des*
1946 *rivières par la gestion en temps réel*. Université Laval Québec. Retrieved from
1947 [https://corpus.ulaval.ca/jspui/handle/20.500.11794/22929;jsessionid=9A0144BC40F304793E](https://corpus.ulaval.ca/jspui/handle/20.500.11794/22929;jsessionid=9A0144BC40F304793ED67882C7DA165D?mode=full)
1948 [D67882C7DA165D?mode=full](https://corpus.ulaval.ca/jspui/handle/20.500.11794/22929;jsessionid=9A0144BC40F304793ED67882C7DA165D?mode=full)
- 1949 Vandermaesen, J., Horemans, B., Bers, K., Vandermeeren, P., Herrmann, S., Sekhar, A., . . . Springael,
1950 D. (2016). Application of biodegradation in mitigating and remediating pesticide
1951 contamination of freshwater resources: state of the art and challenges for optimization. *Appl*
1952 *Microbiol Biotechnol*, *100*(17), 7361-7376. doi:10.1007/s00253-016-7709-z
- 1953 Vieira, I. L., Da Silva, J. B., Ide, C. N., & Janzen, J. G. (2018). Effects of scale and Froude number on the
1954 hydraulics of waste stabilization ponds. *Water Sci Technol*, *77*(1-2), 239-247.
1955 doi:10.2166/wst.2017.529
- 1956 Villaverde, J., van Beinum, W., Beulke, S., & Brown, C. D. (2009). The kinetics of sorption by retarded
1957 diffusion into soil aggregate pores. *Environ Sci Technol*, *43*(21), 8227-8232.
1958 doi:10.1021/es9015052
- 1959 Vink, J. P. M., & VanderZee, S. E. A. T. M. (1997). Pesticide biotransformation in surface waters:
1960 Multivariate analyses of environmental factors at field sites. *Water Research*, *31*(11), 2858-
1961 2868. doi:10.1016/S0043-1354(97)00139-5
- 1962 Vione, D., Minella, M., Maurino, V., & Minero, C. (2014). Indirect photochemistry in sunlit surface
1963 waters: photoinduced production of reactive transient species. *Chemistry*, *20*(34), 10590-
1964 10606. doi:10.1002/chem.201400413
- 1965 Vymazal, J. (2013). Emergent plants used in free water surface constructed wetlands: A review.
1966 *Ecological Engineering*, *61*, 582-592. doi:10.1016/j.ecoleng.2013.06.023

- 1967 Vymazal, J., & Brezinova, T. (2015). The use of constructed wetlands for removal of pesticides from
 1968 agricultural runoff and drainage: a review. *Environ Int*, 75, 11-20.
 1969 doi:10.1016/j.envint.2014.10.026
- 1970 Waiser, M. J., & Robarts, R. D. (2004). Photodegradation of DOC in a shallow prairie wetland:
 1971 evidence from seasonal changes in DOC optical properties and chemical characteristics.
 1972 *Biogeochemistry*, 69(2), 263-284. doi:DOI 10.1023/B:BIOG.0000031048.20050.4e
- 1973 Walker Jr, W. W., & Kadlec, R. H. (2011). Modeling phosphorus dynamics in Everglades wetlands and
 1974 stormwater treatment areas. *Critical Reviews in Environmental Science and Technology*,
 1975 41(S1), 430-446. doi:<https://doi.org/10.1080/10643389.2010.531225>
- 1976 Walse, S. S., Shimizu, K. D., & Ferry, J. L. (2002). Surface-catalyzed transformations of aqueous
 1977 endosulfan. *Environ Sci Technol*, 36(22), 4846-4853. doi:10.1021/es0256257
- 1978 Wang, H. Z., Zuo, H. G., Ding, Y. J., Miao, S. S., Jiang, C., & Yang, H. (2014). Biotic and abiotic
 1979 degradation of pesticide Dufulin in soils. *Environ Sci Pollut Res Int*, 21(6), 4331-4342.
 1980 doi:10.1007/s11356-013-2380-8
- 1981 Wang, M., Zhang, D. Q., Dong, J. W., & Tan, S. K. (2018). Application of constructed wetlands for
 1982 treating agricultural runoff and agro-industrial wastewater: a review. *Hydrobiologia*, 805(1),
 1983 1-31. doi:10.1007/s10750-017-3315-z
- 1984 Wang, Q., & Kelly, B. C. (2017). Occurrence, distribution and bioaccumulation behaviour of
 1985 hydrophobic organic contaminants in a large-scale constructed wetland in Singapore.
 1986 *Chemosphere*, 183, 257-265. doi:10.1016/j.chemosphere.2017.05.113
- 1987 Wang, R., Yuan, Y., Yen, H., Grieneisen, M., Arnold, J., Wang, D., . . . Zhang, M. (2019). A review of
 1988 pesticide fate and transport simulation at watershed level using SWAT: Current status and
 1989 research concerns. *Science of the Total Environment*, 669, 512-526.
 1990 doi:10.1016/j.scitotenv.2019.03.141
- 1991 Warren, C. S., Mackay, D., Bahadur, N. P., & Boocock, D. G. (2002). A suite of multi-segment fugacity
 1992 models describing the fate of organic contaminants in aquatic systems: application to the
 1993 Rihand Reservoir, India. *Water Res*, 36(17), 4341-4355. doi:10.1016/s0043-1354(02)00161-6
- 1994 Watanabe, H., Takagi, K., & Vu, S. H. (2006). Simulation of mefenacet concentrations in paddy fields
 1995 by an improved PCPF-1 model. *Pest Manag Sci*, 62(1), 20-29. doi:10.1002/ps.1115
- 1996 Werner, T. M., & Kadlec, R. H. (2000). Wetland residence time distribution modeling. *Ecological
 1997 Engineering*, 15(1-2), 77-90. doi:Doi 10.1016/S0925-8574(99)00036-1
- 1998 Wilkes, G., Edge, T., Gannon, V., Jokinen, C., Lyautey, E., Medeiros, D., . . . Lapen, D. R. (2009).
 1999 Seasonal relationships among indicator bacteria, pathogenic bacteria, Cryptosporidium
 2000 oocysts, Giardia cysts, and hydrological indices for surface waters within an agricultural
 2001 landscape. *Water Research*, 43(8), 2209-2223. doi:10.1016/j.watres.2009.01.033
- 2002 Willkommen, S., Pfannerstill, M., Guse, B., Ulrich, U., & Fohrer, N. (2018). PondR: a process-oriented
 2003 model to simulate the hydrology of drainage ponds. *Journal of Hydroinformatics*, 20(1), 149-
 2004 163. doi:10.2166/hydro.2017.038
- 2005 Wolfe, N., Zepp, R., & Paris, D. (1978). Carbaryl, propham and chlorpropham: a comparison of the
 2006 rates of hydrolysis and photolysis with the rate of biolysis. *Water Research*, 12(8), 565-571.
 2007 doi:[https://doi.org/10.1016/0043-1354\(78\)90134-3](https://doi.org/10.1016/0043-1354(78)90134-3)
- 2008 Wu, J., Li, Z., Wu, L., Zhong, F., Cui, N., Dai, Y., & Cheng, S. (2017). Triazophos (TAP) removal in
 2009 horizontal subsurface flow constructed wetlands (HSCWs) and its accumulation in plants and
 2010 substrates. *Sci Rep*, 7(1), 5468. doi:10.1038/s41598-017-05874-0
- 2011 Wu, J., Zhang, N., Hayes, A., Panoutsopoulou, K., & Oliver, S. G. (2004). Global analysis of nutrient
 2012 control of gene expression in *Saccharomyces cerevisiae* during growth and starvation. *Proc
 2013 Natl Acad Sci U S A*, 101(9), 3148-3153. doi:10.1073/pnas.0308321100
- 2014 Wu, J. G., & Laird, D. A. (2002). Hydrolysis of chlorpyrifos in aqueous and colloidal systems. *Israel
 2015 journal of chemistry*, 42(1), 99-107. doi:Doi 10.1560/A723-N65t-V12a-1axu
- 2016 Wu, S. C., & Gschwend, P. M. (1988). Numerical Modeling of Sorption Kinetics of Organic-Compounds
 2017 to Soil and Sediment Particles. *Water Resources Research*, 24(8), 1373-1383. doi:DOI
 2018 10.1029/WR024i008p01373
- 2019 Yang, M. Y., Lu, M. Z., Bian, H. F., Sheng, L. X., & He, C. G. (2017). Effects of clogging on hydraulic
 2020 behavior in a vertical-flow constructed wetland system: A modelling approach. *Ecological
 2021 Engineering*, 109, 41-47. doi:10.1016/j.ecoleng.2017.09.010

- 2022 Yoshida, T., & Nakano, Y. (2000). Behavior of pesticides in a paddy field with rapid water penetration.
2023 *Kagaku Kogaku Ronbunshu*, 26(6), 842-848. doi:DOI 10.1252/kakoronbunshu.26.842
- 2024 Yu, X., Zhu, H., Yan, B., Xu, Y., Banuelos, G., Shutes, B., . . . Cheng, R. (2019). Removal of chlorpyrifos
2025 and its hydrolytic metabolite 3,5,6-trichloro-2-pyridinol in constructed wetland mesocosms
2026 under soda saline-alkaline conditions: Effectiveness and influencing factors. *J Hazard Mater*,
2027 373, 67-74. doi:10.1016/j.jhazmat.2019.03.052
- 2028 Yuan, C. B., Huang, T., Zhao, X. H., & Zhao, Y. Q. (2020). Numerical Models of Subsurface Flow
2029 Constructed Wetlands: Review and Future Development. *Sustainability*, 12(8), 3498.
2030 doi:ARTN 3498
- 2031 10.3390/su12083498
- 2032 Yuh-Shan, H. (2004). Citation review of Lagergren kinetic rate equation on adsorption reactions.
2033 *Scientometrics*, 59(1), 171-177. doi:10.1023/B:SCIE.0000013305.99473.cf
- 2034 Zambonin, C. G., Losito, I., Cilenti, A., & Palmisano, F. (2002). Solid-phase microextraction coupled to
2035 gas chromatography-mass spectrometry for the study of soil adsorption coefficients of
2036 organophosphorus pesticides. *J Environ Monit*, 4(4), 477-481. doi:10.1039/b201502c
- 2037 Zamora, S., Marin-Muniz, J. L., Nakase-Rodriguez, C., Fernandez-Lambert, G., & Sandoval, L. (2019).
2038 Wastewater Treatment by Constructed Wetland Eco-Technology: Influence of Mineral and
2039 Plastic Materials as Filter Media and Tropical Ornamental Plants. *Water*, 11(11), 2344.
2040 doi:ARTN 2344
- 2041 10.3390/w11112344
- 2042 Zeng, T., & Arnold, W. A. (2013). Pesticide photolysis in prairie potholes: probing photosensitized
2043 processes. *Environ Sci Technol*, 47(13), 6735-6745. doi:10.1021/es3030808
- 2044 Zepp, R. G., Hoigne, J., & Bader, H. (1987). Nitrate-induced photooxidation of trace organic chemicals
2045 in water. *Environ Sci Technol*, 21(5), 443-450. doi:10.1021/es00159a004
- 2046 Zhang, C.-B., Wang, J., Liu, W.-L., Zhu, S.-X., Ge, H.-L., Chang, S. X., . . . Ge, Y. (2010). Effects of plant
2047 diversity on microbial biomass and community metabolic profiles in a full-scale constructed
2048 wetland. *Ecological Engineering*, 36(1), 62-68. doi:10.1016/j.ecoleng.2009.09.010
- 2049 Zhang, C., Li, Q., Zhang, L., & Zhang, J. (2021). Responses of submerged macrophytes *Vallisneria*
2050 *natans* and epiphytic biofilm to floating plants *Eichhornia crassipes* in eutrophic water. *Water*
2051 *Environ Res*, 93(10), 2237-2249. doi:10.1002/wer.1596
- 2052 Zhang, L., Lv, T., Zhang, Y., Stein, O. R., Arias, C. A., Brix, H., & Carvalho, P. N. (2017). Effects of
2053 constructed wetland design on ibuprofen removal - A mesocosm scale study. *Science of the*
2054 *Total Environment*, 609, 38-45. doi:10.1016/j.scitotenv.2017.07.130
- 2055 Zhang, P., Ren, C., Sun, H., & Min, L. (2018). Sorption, desorption and degradation of neonicotinoids
2056 in four agricultural soils and their effects on soil microorganisms. *Science of the Total*
2057 *Environment*, 615, 59-69. doi:10.1016/j.scitotenv.2017.09.097
- 2058 Zhang, X., & Zhang, M. (2011). Modeling effectiveness of agricultural BMPs to reduce sediment load
2059 and organophosphate pesticides in surface runoff. *Science of the Total Environment*, 409(10),
2060 1949-1958. doi:10.1016/j.scitotenv.2011.02.012
- 2061 Zhang, Z., Fluet-Chouinard, E., Jensen, K., McDonald, K., Hugelius, G., Gumbrecht, T., . . . Poulter, B.
2062 (2021). Development of the global dataset of Wetland Area and Dynamics for Methane
2063 Modeling (WAD2M). *Earth System Science Data*, 13(5), 2001-2023. doi:10.5194/essd-13-
2064 2001-2021
- 2065 Zheng, H. H., & Ye, C. M. (2001). Identification of UV photoproducts and hydrolysis products of
2066 butachlor by mass spectrometry. *Environ Sci Technol*, 35(14), 2889-2895.
2067 doi:10.1021/es0016811
- 2068 Zheng, S., Wang, P., Sun, H., Matsiko, J., Hao, Y., Meng, D., . . . Jiang, G. (2018). Tissue distribution
2069 and maternal transfer of persistent organic pollutants in Kentish Plovers (*Charadrius*
2070 *alexandrinus*) from Cangzhou Wetland, Bohai Bay, China. *Science of the Total Environment*,
2071 612, 1105-1113. doi:10.1016/j.scitotenv.2017.08.323
- 2072 Zhuang, L. L., Yang, T., Zhang, J., & Li, X. (2019). The configuration, purification effect and mechanism
2073 of intensified constructed wetland for wastewater treatment from the aspect of nitrogen
2074 removal: A review. *Bioresour Technol*, 293, 122086. doi:10.1016/j.biortech.2019.122086

2075