

#### Sequencing, annotation, and pangenomics in wheat Frédéric Choulet

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AFBV webinar, May 10th 2022

# Sequencing, annotation, and pangenomics in wheat

#### Frédéric Choulet GDEC, Clermont-Ferrand, France





# 1. Introduction

- Wheat/*Triticeae*
- Pangenomics







#### Complex genome

- o 15 Gb
- Hexaploid AA-BB-DD
- 85% TEs
- Ref cultivar: Chinese Spring



Assembly (v1) 21 chromosomes (14.5 Gb)

Annotation (v1.1) o 107k genes

• 4 million TEs



**IWGSC 2018** 





#### RQAs: Reference-Quality Assemblies of Triticeae genomes



*T. aestivum:*Chinese Spring + **17** accessions

• **SNP** genotyping of **4500** accessions representing world-wide diversity





# Tettelin et al. PNAS 2005 8 genomes of *S. agalactiae*





# Structural Variations (SVs)





1

core

N

# organisms

Koonin and Wolf NAR 2008  $\geq$ bacteria, archea (~700 genomes)



*De novo* assembly of soybean wild relatives for pan-genome analysis of diversity and agronomic traits

Ying-hui Li<sup>1-3,11</sup>, Guangyu Zhou<sup>4,10,11</sup>, Jianxin Ma<sup>5,11</sup>, Wenkai Jiang<sup>4,11</sup>, Long-guo Jin<sup>1-3</sup>, Zhouhao Zhang<sup>4</sup>, Yong Guo<sup>1-3</sup>, Jinbo Zhang<sup>4</sup>, Yi Sui<sup>1-3</sup>, Liangtao Zheng<sup>4</sup>, Shan-shan Zhang<sup>1-3</sup>, Qiyang Zuo<sup>4</sup>, Xue-hui Shi<sup>1-3</sup>, Yan-fei Li<sup>1-3</sup>, Wan-ke Zhang<sup>6</sup>, Yiyao Hu<sup>4</sup>, Guanyi Kong<sup>4</sup>, Hui-long Hong<sup>1-3</sup>, Bing Tan<sup>1-3</sup>, Jian Song<sup>1-3</sup>, Zhang-xiong Liu<sup>1-3</sup>, Yaoshen Wang<sup>4</sup>, Hang Ruan<sup>4</sup>, Carol K L Yeung<sup>4</sup>, Jian Liu<sup>4</sup>, Hailong Wang<sup>4</sup>, Li-juan Zhang<sup>1-3</sup>, Rong-xia Guan<sup>1-3</sup>, Ke-jing Wang<sup>1-3</sup>, Wen-bin Li<sup>7</sup>, Shou-yi Chen<sup>6</sup>, Ru-zhen Chang<sup>1-3</sup>, Zhi Jiang<sup>4</sup>, Scott A Jackson<sup>8</sup>, Ruiqiang Li<sup>4,9</sup> & Li-juan Qiu<sup>1-3</sup>

NATURE BIOTECHNOLOGY VOLUME 32 NUMBER 10 OCTOBER 2014

7 genomes of *Glycine soja* dispensable: ~20% genes

# Extensive gene content variation in the *Brachypodium distachyon* pan-genome correlates with population structure

Sean P. Gordon<sup>1</sup>, Bruno Contreras-Moreira <sup>2,3,4</sup>, Daniel P. Woods<sup>5,6</sup>, David L. Des Marais <sup>7,17</sup>, Diane Burgess<sup>8</sup>, Shengqiang Shu<sup>1</sup>, Christoph Stritt<sup>9</sup>, Anne C. Roulin<sup>9</sup>, Wendy Schackwitz<sup>1</sup>, Ludmila Tyler<sup>10</sup>, Joel Martin <sup>10</sup>, Anna Lipzen<sup>1</sup>, Niklas Dochy <sup>11</sup>, Jeremy Phillips<sup>1</sup>, Kerrie Barry<sup>1</sup>, Koen Geuten <sup>11</sup>, Hikmet Budak <sup>12</sup>, Thomas E. Juenger<sup>13</sup>, Richard Amasino <sup>5,6</sup>, Ana L. Caicedo <sup>10</sup>, David Goodstein <sup>1</sup>, Patrick Davidson<sup>1</sup>, Luis A. J. Mur <sup>14</sup>, Melania Figueroa <sup>15</sup>, Michael Freeling<sup>8</sup>, Pilar Catalan <sup>4,16</sup> & John P. Vogel <sup>18</sup>

NATURE COMMUNICATIONS | 8:2184 | DOI: 10.1038/s41467-017-02292-8 | www.nature.com/naturecommunications 2017

#### 54 genomes of Brachypodium distachyon



# The pangenome of an agronomically important crop plant *Brassica oleracea*

Agnieszka A. Golicz<sup>1</sup>, Philipp E. Bayer<sup>2</sup>, Guy C. Barker<sup>3</sup>, Patrick P. Edger<sup>4</sup>, HyeRan Kim<sup>5</sup>, Paula A. Martinez<sup>1</sup>, Chon Kit Kenneth Chan<sup>2</sup>, Anita Severn-Ellis<sup>2</sup>, W. Richard McCombie<sup>6</sup>, Isobel A.P. Parkin<sup>7</sup>, Andrew H. Paterson<sup>8</sup>, J. Chris Pires<sup>9</sup>, Andrew G. Sharpe<sup>10</sup>, Haibao Tang<sup>11</sup>, Graham R. Teakle<sup>3</sup>, Christopher D. Town<sup>12</sup>, Jacqueline Batley<sup>2</sup> & David Edwards<sup>2</sup>

NATURE COMMUNICATIONS | 7:13390 | DOI: 10.1038/ncomms13390 | www.nature.com/naturecommunications 2016

9 genomes of *Brassica oleracea* dispensable: ~20% genes

# Pan-genome analysis highlights the extent of genomic variation in cultivated and wild rice

Qiang Zhao<sup>1</sup>, Qi Feng<sup>1</sup>, Hengyun Lu<sup>1</sup>, Yan Li<sup>1</sup>, Ahong Wang<sup>1</sup>, Qilin Tian<sup>1</sup>, Qilin Zhan<sup>1</sup>, Yiqi Lu<sup>1</sup>, Lei Zhang<sup>1</sup>, Tao Huang<sup>1</sup>, Yongchun Wang<sup>1</sup>, Danlin Fan<sup>1</sup>, Yan Zhao<sup>1</sup>, Ziqun Wang<sup>1</sup>, Congcong Zhou<sup>1</sup>, Jiaying Chen<sup>1</sup>, Chuanrang Zhu<sup>1</sup>, Wenjun Li<sup>1</sup>, Qijun Weng<sup>1</sup>, Qun Xu<sup>2</sup>, Zi-Xuan Wang<sup>1</sup>, Xinghua Wei<sup>2</sup>, Bin Han<sup>1</sup> and Xuehui Huang<sup>13\*</sup>

NATURE GENETICS | VOL 50 | FEBRUARY 2018 | 278-284 | www.nature.com/naturegenetics

67 genomes of *Oryza sativa* + *rufipogon* ~20% genes

Core 26k Dispensable 16k

The Plant Cell, Vol. 26: 121-135, January 2014, www.plantcell.org © 2014.

Insights into the Maize Pan-Genome and Pan-Transcriptome<sup>MOPEN</sup>

Candice N. Hirsch,<sup>a,b,1</sup> Jillian M. Foerster,<sup>c,2</sup> James M. Johnson,<sup>c,3</sup> Rajandeep S. Sekhon,<sup>c,d</sup> German Muttoni,<sup>c,4</sup> Brieanne Vaillancourt,<sup>a,b</sup> Francisco Peñagaricano,<sup>e</sup> Erika Lindquist,<sup>†</sup> Mary Ann Pedraza,<sup>f</sup> Kerrie Barry,<sup>†</sup> Natalia de Leon,<sup>c,d</sup> Shawn M. Kaeppler,<sup>c,d</sup> and C. Robin Buell<sup>a,b,5</sup>

#### 503 maize accessions (RNASeq) => ~8700 novel transcripts

Darracq et al. BMC Genomics (2018) 19:119 DOI 10.1186/s12864-018-4490-7

Sequence analysis of European maize inbred line F2 provides new insights into molecular and chromosomal characteristics of presence/absence variants

Aude Darracq<sup>1†</sup>, Clémentine Vitte<sup>1†</sup>, Stéphane Nicolas<sup>1</sup>, Jorge Duarte<sup>2</sup>, Jean-Philippe Pichon<sup>2</sup>, Tristan Mary-Huard<sup>1,3</sup>, Céline Chevalier<sup>1</sup>, Aurélie Bérard<sup>4</sup>, Marie-Christine Le Paslier<sup>4</sup>, Peter Rogowsky<sup>5</sup>, Alain Charcosset<sup>1</sup> and Johann Joets<sup>1\*</sup><sup>•</sup>

( CrossMark

#### Dispensable vs core genes:

- shorter, fewer introns
- less expressed, fewer conditions
- evolve more rapidly
- enriched in functions related to adaptation, response to env

# High-resolution genetic mapping of maize pan-genome sequence anchors

Fei Lu<sup>1</sup>, Maria C. Romay<sup>1</sup>, Jeffrey C. Glaubitz<sup>1</sup>, Peter J. Bradbury<sup>2</sup>, Robert J. Elshire<sup>1</sup>, Tianyu Wang<sup>3</sup>, Yu Li<sup>3</sup>, Yongxiang Li<sup>3</sup>, Kassa Semagn<sup>4</sup>, Xuecai Zhang<sup>5</sup>, Alvaro G. Hernandez<sup>6</sup>, Mark A. Mikel<sup>6,7</sup>, Ilya Soifer<sup>8</sup>, Omer Barad<sup>8</sup> & Edward S. Buckler<sup>1,2</sup>

NATURE COMMUNICATIONS | 6:6914 | DOI: 10.1038/ncomms7914 | www.nature.com/naturecommunications 2015



=> "PAV SNPs exhibit enriched associations with traits"

# The tomato pan-genome uncovers new genes and a rare allele regulating fruit flavor

Lei Gao <sup>©</sup><sup>1,10</sup>, Itay Gonda<sup>1,2,10</sup>, Honghe Sun<sup>1</sup>, Qiyue Ma<sup>1</sup>, Kan Bao<sup>1</sup>, Denise M. Tieman<sup>3</sup>, Elizabeth A. Burzynski-Chang<sup>4</sup>, Tara L. Fish<sup>5</sup>, Kaitlin A. Stromberg<sup>1</sup>, Gavin L. Sacks<sup>©</sup><sup>4</sup>, Theodore W. Thannhauser<sup>5</sup>, Majid R. Foolad<sup>6</sup>, Maria Jose Diez<sup>7</sup>, Jose Blanca<sup>7</sup>, Joaquin Canizares<sup>7</sup>, Yimin Xu<sup>1</sup>, Esther van der Knaap<sup>8</sup>, Sanwen Huang<sup>®</sup><sup>9</sup>, Harry J. Klee<sup>3</sup>, James J. Giovannoni<sup>©</sup><sup>1,5\*</sup> and Zhangjun Fei<sup>©</sup><sup>1,5\*</sup>

NATURE GENETICS | VOL 51 | JUNE 2019 | 1044-1051 | www.nature.com/naturegenetics

# 725 genomes of *Solanum cultivated* + *3 wild species* core: **74%** genes

"The most striking feature of the tomato pan-genome was its **high core** gene content (74.2%), as compared with those of <u>Arabidopsis thaliana</u> (70%), <u>Brassica napus</u> (62%), <u>bread</u> wheat (64%), <u>rice</u> (54%), wild <u>soybean</u> (49%) and <u>Brachypodium</u> <u>distachyon</u> (35%)"

#### The pangenome of hexaploid bread wheat

Juan D. Montenegro<sup>1,†</sup>, Agnieszka A. Golicz<sup>1,2,†,‡</sup>, Philipp E. Bayer<sup>2,†</sup>, Bhavna Hurgobin<sup>1,2</sup>, HueyTyng Lee<sup>1,2</sup>, Chon-Kit Kenneth Chan<sup>2</sup>, Paul Visendi<sup>1</sup>, Kaitao Lai<sup>3</sup>, Jaroslav Doležel<sup>4</sup>, Jacqueline Batley<sup>1,2,5</sup> and David Edwards<sup>1,2,5,</sup>\*

The Plant Journal (2017) 90, 1007-1013

16 Australian wheat cultivars (Illumina reseq) dispensable: ~35% of genes

#### 15 wheat genomes assembled

- "12% of genes showed PAVs"
- "26% of the projected genes were found in tandem duplications, indicating that CNV is a strong contributor of genetic variation in wheat"



# Multiple wheat genomes reveal global variation in modern breeding

https://doi.org/10.1038/s41586-020-2961-x

Sean Walkowiak<sup>1,2,41</sup>, Liangliang Gao<sup>3,41</sup>, Cecile Monat<sup>4,41</sup>, Georg Haberer<sup>5</sup>, Mulualem T. Kassa<sup>6</sup>, Jemima Brinton<sup>7</sup>, Ricardo H. Ramirez-Gonzalez<sup>7</sup>, Markus C. Kolodziej<sup>8</sup> Emily Delorean<sup>3</sup>, Dinushika Thambugala<sup>9</sup>, Valentyna Klymiuk<sup>1</sup>, Brook Byrns<sup>1</sup>, Heidrun Gundlach<sup>5</sup>, Venkat Bandi<sup>10</sup>, Jorge Nunez Siri<sup>10</sup>, Kirby Nilsen<sup>111</sup>, Catharine Aquino<sup>12</sup>, Axel Himmelbach<sup>4</sup>, Dario Copetti<sup>13,14</sup>, Tomohiro Ban<sup>15</sup>, Luca Venturini<sup>16</sup>, Michael Bevan<sup>7</sup>, Bernardo Clavijo17, Dal-Hoe Koo3, Jennifer Ens1, Krystalee Wiebe1, Amidou N'Diaye1, Allen K. Fritz<sup>3</sup>, Carl Gutwin<sup>10</sup>, Anne Fiebig<sup>4</sup>, Christine Fosker<sup>17</sup>, Bin Xiao Fu<sup>2</sup>, Gonzalo Garcia Accinelli<sup>17</sup>, Keith A. Gardner<sup>18</sup>, Nick Fradgley<sup>18</sup>, Juan Gutierrez-Gonzalez<sup>19</sup>, Gwyneth Halstead-Nussloch<sup>13</sup>, Masaomi Hatakeyama<sup>12,13</sup>, Chu Shin Koh<sup>20</sup>, Jasline Deek<sup>21</sup>, Alejandro C. Costamagna<sup>22</sup>, Pierre Fobert<sup>6</sup>, Darren Heavens<sup>17</sup>, Hiroyuki Kanamori<sup>23</sup>, Kanako Kawaura<sup>15</sup>, Fuminori Kobayashi<sup>23</sup>, Ksenia Krasileva<sup>17</sup>, Tony Kuo<sup>24,25</sup>, Neil McKenzie<sup>7</sup> Kazuki Murata<sup>26</sup>, Yusuke Nabeka<sup>26</sup>, Timothy Paape<sup>13</sup>, Sudharsan Padmarasu<sup>4</sup>, Lawrence Percival-Alwyn<sup>18</sup>, Sateesh Kagale<sup>6</sup>, Uwe Scholz<sup>4</sup>, Jun Sese<sup>25,27</sup>, Philomin Juliana<sup>28</sup>, Ravi Singh<sup>28</sup>, Rie Shimizu-Inatsugi<sup>13</sup>, David Swarbreck<sup>17</sup>, James Cockram<sup>18</sup>, Hikmet Budak<sup>29</sup>, Toshiaki Tameshige<sup>15</sup>, Tsuyoshi Tanaka<sup>23</sup>, Hiroyuki Tsuji<sup>15</sup>, Jonathan Wright<sup>17</sup>, Jianzhong Wu<sup>23</sup>, Burkhard Steuernagel<sup>7</sup>, Ian Small<sup>30</sup>, Sylvie Cloutier<sup>31</sup>, Gabriel Keeble-Gagnère<sup>32</sup>, Gary Muehlbauer<sup>19</sup>, Josquin Tibbets<sup>32</sup>, Shuhei Nasuda<sup>26</sup>, Joanna Melonek<sup>30</sup>, Pierre J. Hucl<sup>1</sup>, Andrew G. Sharpe<sup>20</sup>, Matthew Clark<sup>16</sup>, Erik Legg<sup>33</sup>, Arvind Bharti<sup>33</sup>, Peter Langridge<sup>34</sup>, Anthony Hall<sup>17</sup>, Cristobal Uauy<sup>7</sup>, Martin Mascher<sup>4,35</sup>, Simon G. Krattinger<sup>8,36</sup>, Hirokazu Handa<sup>23,37</sup>, Kentaro K. Shimizu<sup>13,15</sup>, Assaf Distelfeld<sup>38</sup>, Ken Chalmers<sup>34</sup>, Beat Keller<sup>8</sup>, Klaus F. X. Mayer<sup>5,39</sup>, Jesse Poland<sup>3</sup>, Nils Stein<sup>4,40</sup>, Curt A. McCartney<sup>9</sup>, Manuel Spannagl<sup>5</sup>, Thomas Wicker<sup>8</sup> & Curtis J. Pozniak<sup>1</sup>

Build and visualize pangenomes





GenomicusPlants - database version: 49.01 / Web-code version: 2021-08-15 - Dyogen Team

#### Gautereau et al. Plos Comput. Biol. 2020

pangenome graph of ~3000 Acinetobacter PPanGGOLiN



# • Practical Haplotype Graph

 A sorghum practical haplotype graph facilitates genome-wide imputation and cost-effective genomic prediction

 Sarah E. Jensen<sup>1</sup> | Jean Rigaud Charles<sup>2</sup> | Kebede Muleta<sup>3</sup> |

 Peter J. Bradbury<sup>4,6</sup> | Terry Casstevens<sup>4</sup> | Santosh P. Deshpande<sup>5</sup> |

 Michael A. Gore<sup>1</sup> | Rajeev Gupta<sup>5</sup> | Daniel C. Ilut<sup>1</sup> | Lynn Johnson<sup>4</sup> |

 Roberto Lozano<sup>1</sup> | Zachary Miller<sup>4</sup> | Punna Ramu<sup>4</sup> | Abhishek Rathore<sup>5</sup> |

 M. Cinta Romay<sup>4</sup> | Hari D. Upadhyaya<sup>5</sup> | Rajeev K. Varshney<sup>5</sup> |

 Geoffrey P. Morris<sup>3</sup> | Gael Pressoir<sup>2</sup> | Edward S. Buckler<sup>1,4,6</sup> |

 Guillaume P. Ramstein<sup>4</sup> |

(a) (b) Reference Taxon 1 Taxon 2 Taxon 3 Taxon 4 Taxon 5 Taxon 6 Taxon 7 Taxon 8 Taxon 9 Taxon 10 (c) (d) Reference Consensus 1 Consensus 3 (c) (d) Reference Consensus 2 Consensus 3 Consensus 4 Consensus 4 Consensus 5 Consensus 4 Consensus 4 Consensus 5 Consensus 4 Consensus 5 Consensus 5 Consensus 4 Consensus 5 Consensus 5 Consensus 5 Consensus 5 Consensus 6 Consensus 6 Consensus 7 Consensus

- for breeding programs
- based on a limited number of founder lines
- 0.01x skim-seq ->call SNPs with 5% errors

#### Development of the Wheat Practical Haplotype Graph database as a resource for genotyping data storage and genotype imputation

Katherine W. Jordan (D), <sup>1,2,†</sup> Peter J. Bradbury (D), <sup>3</sup> Zachary R. Miller,<sup>4</sup> Moses Nyine (D), <sup>1</sup> Fei He,<sup>1</sup> Max Fraser,<sup>5</sup> Jim Anderson,<sup>5</sup> Esten Mason,<sup>6</sup> Andrew Katz,<sup>6</sup> Stephen Pearce (D), <sup>6</sup> Arron H. Carter,<sup>7</sup> Samuel Prather,<sup>7</sup> Michael Pumphrey,<sup>7</sup> Jianli Chen,<sup>8</sup> Jason Cook,<sup>9</sup> Shuyu Liu, <sup>10</sup> Jackie C. Rudd, <sup>10</sup> Zhen Wang,<sup>10</sup> Chenggen Chu (D), <sup>10</sup> Amir M. H. Ibrahim, <sup>10</sup> Jonathan Turkus, <sup>11</sup> Eric Olson,<sup>11</sup> Ragupathi Nagarajan,<sup>12</sup> Brett Carver,<sup>12</sup> Liuling Yan,<sup>12</sup> Ellie Taagen,<sup>4</sup> Mark Sorrells,<sup>4</sup> Brian Ward,<sup>13</sup> Jie Ren,<sup>1,14</sup> Alina Akhunova, <sup>1,14</sup> Guihua Bai,<sup>2</sup> Robert Bowden,<sup>2</sup> Jason Fiedler,<sup>15</sup> Justin Faris (D), <sup>15</sup> Jorge Dubcovsky (D), <sup>16</sup> Mary Guttieri,<sup>2</sup> Gina Brown-Guedira,<sup>13</sup> Ed Buckler,<sup>3</sup> Jean-Luc Jannink (D), <sup>3</sup> and Eduard D. Akhunov (D), <sup>1,\*</sup>

G3, 2022, 12(2), jkab390

- 65 wheat lines, exome capture
- 0.01x -> imputations with 8% errors

# 2. Wheat genome sequencing, annotation, A-B-D comparative genomics

#### Shifting the limits in wheat research and breeding using a fully annotated reference genome

International Wheat Genome Sequencing Consortium (IWGSC)\*

Science 361, 661 (2018) 17 August 2018

## $\circ$ $\,$ Predicting genes is still NOT routine

**GDEC** Clermont, France – *Rimbert Leroy Choulet et al.* **PGSB** Munich, Germany – *Spannagl Twardziok et al.* **EI** Norwich, UK – *Swarbreck Venturini et al.* 

#### IWGSC RefSeq (Chinese Spring)

• 107,891 genes







⊗ HighConfidenceGenesv1.1 (GOMAP annotation available)	
Sorghum bicolor	•
Hordeum vulgare var. distichum	
🔇 Aegilops tauschii	
S Triticum urartu	
⊗ Brachypodium distachyon	
S Oryza barthii	
S Zea mays +	
⊗ Triticum aestivum RNA-Seq (Pingault et al. 2014)	
S EMBL EST of Triticeae	+   
S FgeneSH predictions	

- Impact of annotation methods on knowledge
- ➢ in 2017: 3 versions of Chinese Spring chr3B

#### chr3B BAC-by-BAC



Choulet et al. Science 2014

TGAC\_v1



Clavijo et al. Genom Biol 2017

Chr. Survey Seq



IWGSC Science 2014

7264 genes

5728 genes -> (**26**% identical) 5862 genes (**12**% identical)



39k homeologous groups [including 29k LC genes]

[ 27% of genes are duplicates ] ex: 207 PPRs

## Polyploidy + single gene duplications



# 3. A-B-D Comparative Analysis of **TEs**

Micker et al. Genome Biology (2018) 19:03 https://doi.org/10.1186/s13059-018-1479-0 RESEARCH Open Access Impact of transposable elements on genome structure and evolution in bread wheat Thomas Wicker<sup>1+</sup>, Heidrun Gundlach<sup>2+</sup>, Manuel Spannagl<sup>2</sup>, Cristobal Uauy<sup>3</sup>, Philippa Borrill<sup>3</sup>, Ricardo H. Ramírez-González<sup>3</sup>, Romain De Oliveira<sup>4</sup>, International Wheat Genome Sequencing Consortium<sup>5</sup>, Klaus F. X. Mayer<sup>26</sup>, Etienne Paux<sup>4</sup> and Frédéric Choulet<sup>4\*</sup>

# $\circ\,$ TE modeling with CLARI-TE and ClariTeRep

https://github.com/jdaron/CLARI-TE



Daron et al. Genom Biol 2015





#### Near-complete **TE turnover** since A-B-D divergence

## o % TEs



Families

- Abundant families are the same on A-B-D
- > **0** family specific to 1 subgenome
- 76% of the fam. account for similar % on A-B-D (<2 fold-change)</p>

## $\circ$ TE trees



Independent TE evolution in the diploids AA, BB, DD

Some rare cases of TE amplif in the tetraploid AABB

# $_{\odot}\,$ TE content around genes



# $\circ$ TE dynamics

A-B-D last common ancestor





- Most families were active
- Equilibrium
- -> Hypothesis of a structural role of TEs likely under selection pressure?

# 4. Structural Variations in *Triticum* using resequencing data

PhD Romain De Oliveira



Frontiers in Genetics | www.frontiersin.org

August 2020 | Volume 11 | Article 891

Structural Variations Affecting Genes and Transposable Elements of Chromosome 3B in Wheats

Romain De Oliveira<sup>1</sup>, Hélène Rimbert<sup>1</sup>, François Balfourier<sup>1</sup>, Jonathan Kitt<sup>1</sup>, Emeric Dynomant<sup>1</sup>, Jan Vrána<sup>2</sup>, Jaroslav Doležel<sup>2</sup>, Federica Cattonaro<sup>3</sup>, Etienne Paux<sup>1</sup> and Frédéric Choulet<sup>1\*</sup>



Illumina reseq ~30x



De Oliveira et al. Front. Genet. 2020





Chinese Spring chr3B **~500** TE families

## $\circ$ TE CNVs

#### TE% = **84%** in all 45 accessions



No major difference of the number of TE copies for each superfamily

## $\circ$ TE CNVs



0 fam with log2ratio > 2
➤ At the family level: globally extremely conserved proportions



#### **RESULTS for 45** *Triticum* accessions:

- from 7% to 19% TE-PAVs
- cumulated variable: 60% ISBPs

## **•** TE PAVs – distribution of polymorphisms



# 5. Genomic variability affecting **TEs** in wheat **using assembled genomes**

PhD Nathan Papon (2019-2022)

- *Methodology for comparative genomics of complex genomes with Gbps of repeats*
- Extent of genomic variability?
- Active families? Amount of transposition events?
- Did polyploidization trigger a TE burst?



#### Multiple RefSeq now available

Ling et al. 2018









#### Results:

- Variable regions represent (pairwise):
  - inter-specific: 8-34% of the sequences
  - intra-specific: 4-6% of the sequences



Extent of variability due to recent insertions?

- $\sim \frac{5k-10k}{10k}$  transposition events / subgenome (<1% of the genome)
- 238 active families

#### Impact of polyploidy?

➢ none

			Recent TE insertions		
	Top	10 families	T. aestivum AABBDD	T. durum AABB	
RLC_famc1	А	B D	3032	3239	
DTC_famc2			177	191	
RLG_famc2			118	133	
RLG_famc1			259	228	
RLG_famc7			521	537	
RLG_famc5			67	66	
RLG_famc3			2312	2515	
RLG_famc4			36	27	
RLG_famc11			157	168	
RLG_famc10			104	106	

> Equilibrium

# 6. Sequencing the Renan genome



GigaScience, 2022, 11, 1–18 DOI: 10.1093/gigascience/giac034

RESEARCH

Long-read and chromosome-scale assembly of the hexaploid wheat genome achieves high resolution for research and breeding

Jean-Marc Aury <sup>1</sup>, Stefan Engelen <sup>1</sup>, Benjamin Istace <sup>1</sup>, Cécile Monat <sup>2</sup>, Pauline Lasserre-Zuber<sup>2</sup>, Caroline Belser <sup>1</sup>, Corinne Cruaud <sup>3</sup>, Hélène Rimbert <sup>2</sup>, Philippe Leroy <sup>2</sup>, Sandrine Arribat <sup>4</sup>, Isabelle Dufau<sup>4</sup>, Arnaud Bellec <sup>4</sup>, David Grimbichler <sup>5</sup>, Nathan Papon<sup>2</sup>, Etienne Paux <sup>2</sup>, Marion Ranoux<sup>2</sup>, Adriana Alberti <sup>3</sup>, Patrick Wincker <sup>1</sup> and Frédéric Choulet <sup>2</sup>,\*





G. Doussinault, J. Jahier, M. Bernard et al. (1980's 90's 00's)

Genome Assembly





## $\circ$ Annotation of genes

107k HC genes Chinese Spring



#### ~12% ambiguous mapping

- *denovo* gene annotation would generate to many differences
- gene mapping from a reference is NOT satisfying
  - Develop MAGATT

□ MAGATT (Marker-Assisted Gene Annotation Transfer for Triticeae)





# □ Introgressions





## • Conclusions

- Wheat has reached the pangenomic area
- Annotation matters
- Perspectives
- Build a wheat pangenome based on 8-12 founder lines
  - IWGSC/NSF proposal under review
  - GDEC, IPS2







Balfourier et al. 2018

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