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# Sequencing the genome of the French wheat variety Renan

Frédéric Choulet

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Submitted on 5 May 2023

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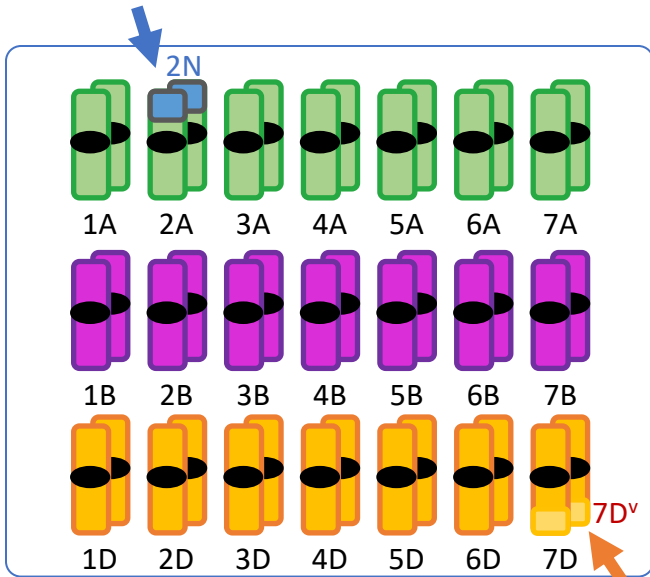
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# Sequencing the genome of the French wheat variety Renan

- *Frédéric CHOULET*  
*GDEC, INRAE-Univ Clermont-Ferrand*

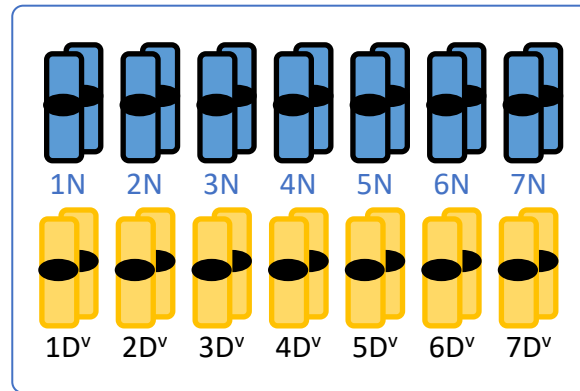


Yr17 Lr37 Sr38 Cre5

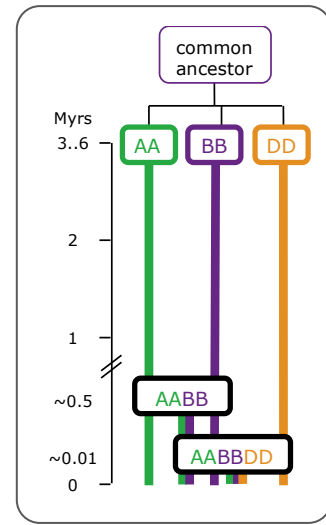


RENAN

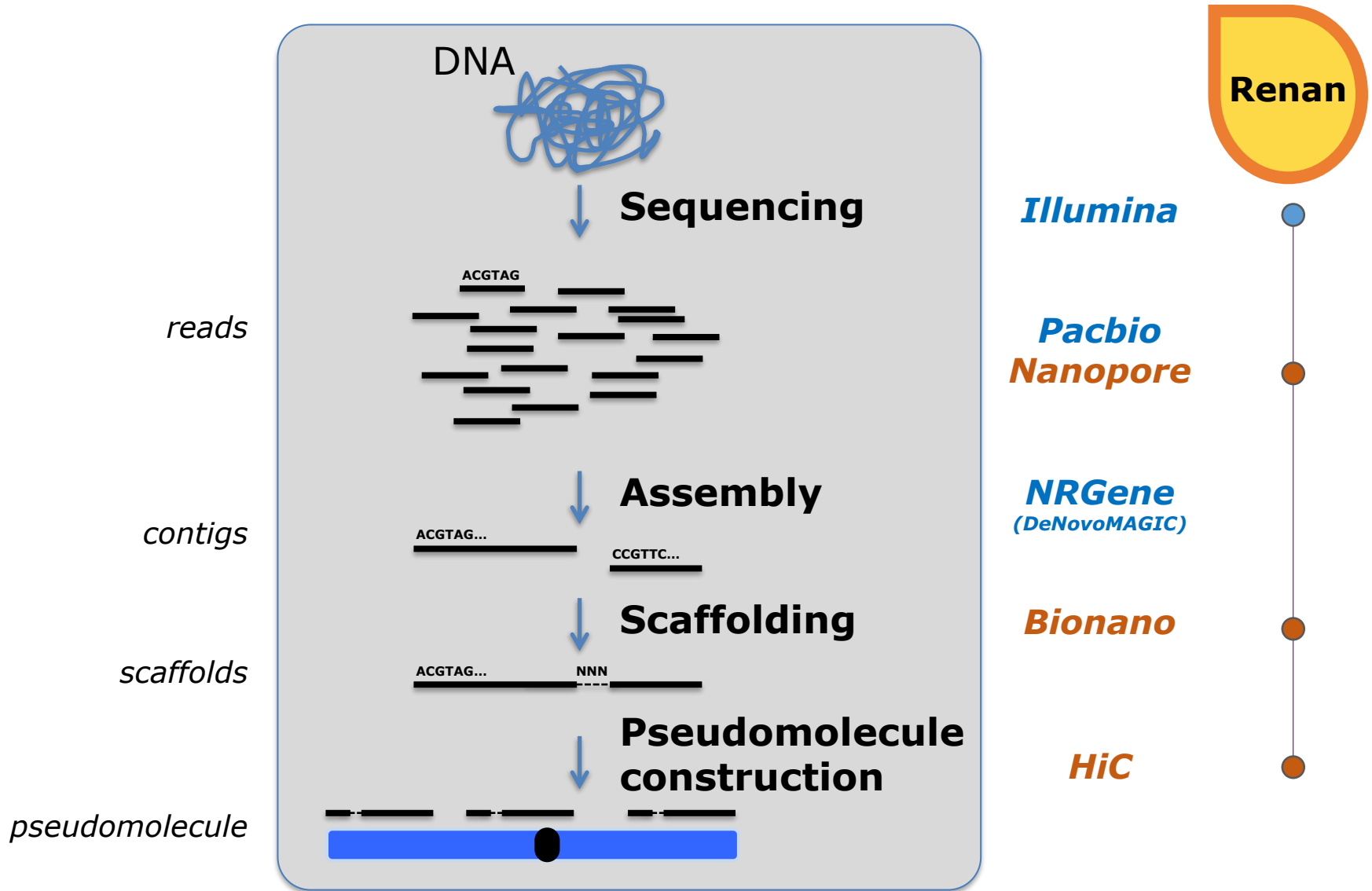
Pch1



*Ae. ventricosa*



# □ Genome assembly: steps and techno



# ❑ *Triticeae* refseq current status (09/2022)

## Reference-quality genome assemblies

- *T. aestivum*
  - Chinese Spring (NRgene)
  - ArinaLrFor Jagger Julius CDC\_Lancer CDC\_Landmark Mace-Norin61 Stanley SY-Mattis Spelta (NRGene)
  - Zang1817 (NRGene)
  - Fielder (Pacbio HiFi)
  - Kariega (Pacbio HiFi)
  - Renan (ONT)
  - Attraktion (Pacbio HiFi)
- *T. urartu* AA
- *T. durum* AABB
- *T. dicoccoides* AABB
- *Ae. tauschii* DD
- *Ae. searsii* DD
- *Ae. bicornis* DD
- *Ae. longissima* DD
- *Ae. sharonensis* DD
- *Ae. speltoides* SS

Article | Open Access | Published: 08 October 2020

### Origin and adaptation to high altitude of Tibetan semi-wild wheat

Weilong Guo, Mingming Xin, Zihao Wang, Yingyin Yao, Zhaorong Hu, Wanjun Song, Kuohai Yu, Yongming Chen, Xiaobo Wang, Panfeng Guan, Rudi Appels, Huiru Peng, Zhongfu Ni & Qixin Sun

Nature Communications 11, Article number: 5085 (2020) | Cite this article

DNA Research, 2021, 00(0), 1-7  
doi: 10.1093/dnares/dsab008  
Advance Access Publication Date: 12 July 2021  
Resource Article: Genomes Explored

Resource Article: Genomes Explored

### Chromosome-scale genome assembly of the transformation-amenable common wheat cultivar 'Fielder'

Kazuhiro Sato<sup>1\*</sup>, Fumitaka Abe<sup>2</sup>, Martin Mascher<sup>3,4</sup>, Georg Haberer<sup>5</sup>, Heidrun Gundlach<sup>5</sup>, Manuel Spannagl<sup>5</sup>, Kenta Shirasawa<sup>6</sup>, and Sachiko Isobe<sup>6</sup>

nature genetics LETTERS  
https://doi.org/10.1038/s41588-022-01022-1

OPEN

### Long-read genome sequencing of bread wheat facilitates disease resistance gene cloning

Abrouk<sup>1,2</sup>, Willem H. P. Boshoff<sup>2,7</sup>, Stéphane Cauet<sup>1,8</sup>, Mohamed Mohammed<sup>1</sup>, Jan Bettgenhaeuser<sup>1</sup>, Kirsty S. Botha<sup>1,9</sup>, René Prins<sup>1,4,5,10</sup> and Simon G. Krattinger<sup>1,10</sup>

Molecular Plant Research Article

CellPress Partner Journal

### Genome sequences of five Sitopsis species of *Aegilops* and the origin of polyploid wheat B subgenome

Lin-Feng Li<sup>1,2,5\*</sup>, Zhi-Bin Zhang<sup>1,3,5</sup>, Zhen-Hui Wang<sup>4</sup>, Ning Li<sup>1</sup>, Yan Sha<sup>1</sup>, Xin-Feng Wang<sup>2</sup>, Ning Ding<sup>2</sup>, Yang Li<sup>1</sup>, Jing Zhao<sup>1</sup>, Ying Wu<sup>1</sup>, Lei Gong<sup>1</sup>, Fabrizio Mafessoni<sup>3</sup>, Avraham A. Levy<sup>3,\*</sup> and Bao Liu<sup>1,\*</sup>

QAO

doi: 10.1111/gpb.13841

QAO

doi: 10.1111/gpb.13841

### Gene homologs and reference sequence support resistance gene mapping in winter wheat

Sandip M. Kale<sup>1,1</sup>, Albert W. Schulthess<sup>1,1</sup>, Sudharsan Padmarasu<sup>1</sup>, Philipp H. G. Boever<sup>2</sup>, Johannes Schacht<sup>2</sup>, Axel Himmelbach<sup>1</sup>, Burkhard Steuernagel<sup>1</sup>, Brande B. H. Wulff<sup>3,4</sup>, Jochen C. Reif<sup>1</sup>, Nils Stein<sup>1,5,\*</sup> and Martin Mascher<sup>1,5,\*</sup>

# Long-read and chromosome-scale assembly of the hexaploid wheat genome achieves high resolution for research and breeding

Jean-Marc Aury<sup>1,\*</sup>, Stefan Engelen<sup>1</sup>, Benjamin Istace<sup>1</sup>, Cécile Monat<sup>2</sup>, Pauline Lasserre-Zuber<sup>2</sup>, Caroline Belser<sup>1</sup>, Corinne Cruaud<sup>3</sup>, Hélène Rimbert<sup>2</sup>, Philippe Leroy<sup>2</sup>, Sandrine Arribat<sup>4</sup>, Isabelle Dufau<sup>4</sup>, Arnaud Bellec<sup>4</sup>, David Grimbichler<sup>5</sup>, Nathan Papon<sup>2</sup>, Etienne Paux<sup>2</sup>, Marion Ranoux<sup>2</sup>, Adriana Alberti<sup>1,6</sup>, Patrick Wincker<sup>1</sup> and Frédéric Choulet<sup>2,\*</sup>

<sup>1</sup>Génomique Métabolique, Genoscope, Institut François Jacob, CEA, CNRS, Univ Evry, Université Paris-Saclay, 91057 Evry, France

<sup>2</sup>GDEC, Université Clermont Auvergne, INRAE, UMR1095, 63000 Clermont-Ferrand, France

<sup>3</sup>Commissariat à l'Énergie Atomique (CEA), Institut François Jacob, Genoscope, F-91057 Evry, France

<sup>4</sup>INRAE, CNRGV French Plant Genomic Resource Center, F-31320, Castanet Tolosan, France

<sup>5</sup>Mésocentre Clermont Auvergne, DOSI / Bâtiment Turing, 7 avenue Blaise Pascal, 63178 Aubière, France

<sup>6</sup>Present address: Université Paris-Saclay, CEA, CNRS, Institute for Integrative Biology of the Cell (I2BC), 91198, Gif-sur-Yvette, France

\*Correspondence address. Jean-Marc Aury, Génomique Métabolique, Genoscope, Institut François Jacob, CEA, CNRS, Univ Evry, Université Paris-Saclay, 91057 Evry, France. E-mail: [jmaury@genoscope.cns.fr](mailto:jmaury@genoscope.cns.fr); Frédéric Choulet, GDEC, Université Clermont Auvergne, INRAE, UMR1095, 63000 Clermont-Ferrand, France. E-mail: [frederic.choulet@inrae.fr](mailto:frederic.choulet@inrae.fr)

## Abstract

**Background:** The sequencing of the wheat (*Triticum aestivum*) genome has been a methodological challenge for many years owing to its large size (15.5 Gb), repeat content, and hexaploidy. Many initiatives aiming at obtaining a reference genome of cultivar Chinese Spring have been launched in the past years and it was achieved in 2018 as the result of a huge effort to combine short-read sequencing with many other resources. Reference-quality genome assemblies were then produced for other accessions, but the rapid evolution of sequencing technologies offers opportunities to reach high-quality standards at lower cost.

**Results:** Here, we report on an optimized procedure based on long reads produced on the Oxford Nanopore Technology PromethION device to assemble the genome of the French bread wheat cultivar Renan.

**Conclusions:** We provide the most contiguous chromosome-scale assembly of a bread wheat genome to date. Coupled with an annotation based on RNA-sequencing data, this resource will be valuable for the crop community and will facilitate the rapid selection of agronomically important traits. We also provide a framework to generate high-quality assemblies of complex genomes using ONT.

**Keywords:** wheat, hexaploid genome, long-reads, nanopore sequencing, genome assembly, haplotype characterization, introgressions

## □ **Step#1: sequencing**

NovaSeq  
**30x**

*Illumina*

**Renan**

MinION PromethION  
2 runs 18 runs

*Nanopore*

- Total 63x
- Select **30x** with longer reads  
(reads>50kb → 15x)

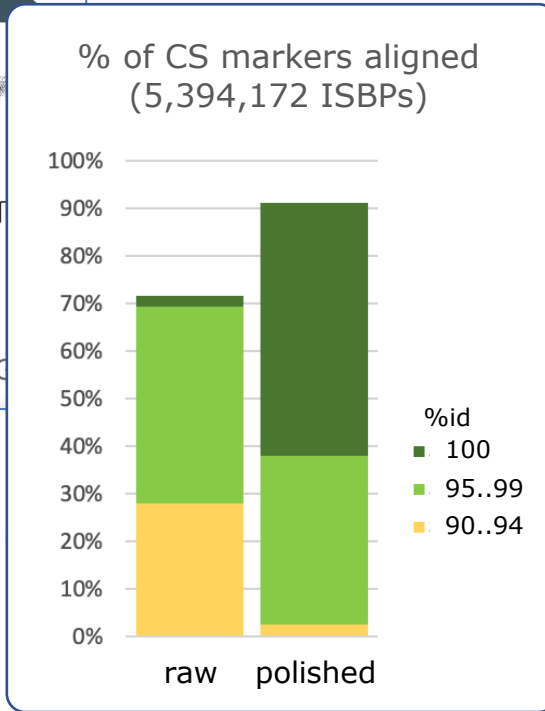
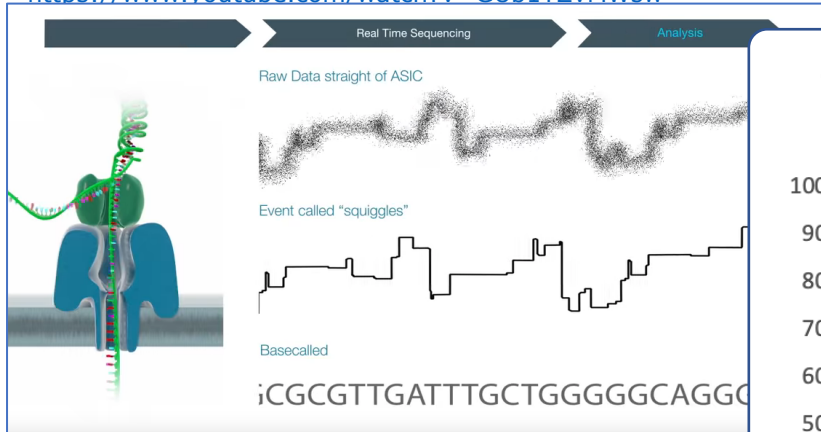
*Bionano*

*HiC*



# ❑ Step#2: read assembly

<https://www.youtube.com/watch?v=GUb1TZvMWsw>



Early stages

- > error rate in reads: 12%
- > error rate in assembly: 4%

- basecalling
- polishing



Illumina  
30x

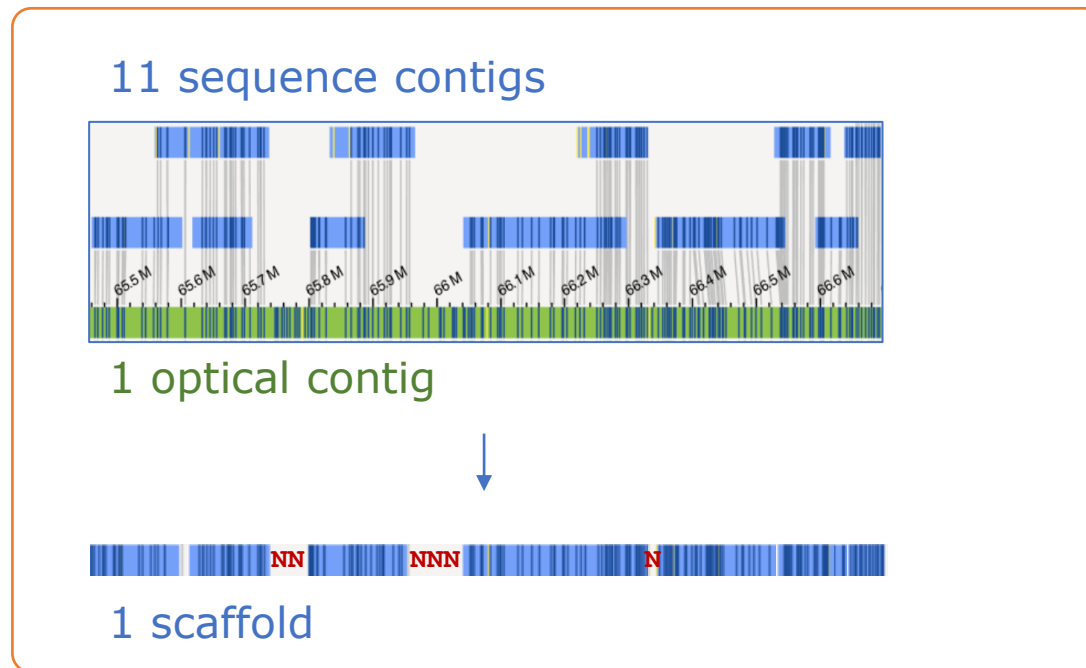
ACGTAGACGTACGGCTGCATCGCATTTCATCGGCCGGGGGGGGCTGCTCGTCGCTCCTCGT

assembly





## □ **Step#3: scaffolding**



*Illumina*

*Nanopore*

***Bionano***

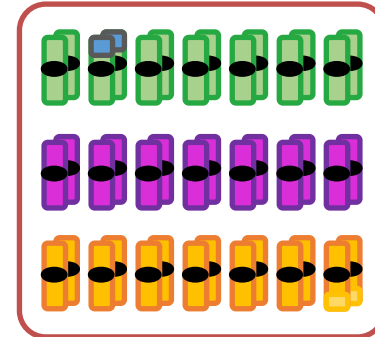
*HiC*



## □ **Assembly metrics**

### ❖ **Metrics**

- Genome size=14.3 Gb
- 2904 scaffolds (=138/chr)



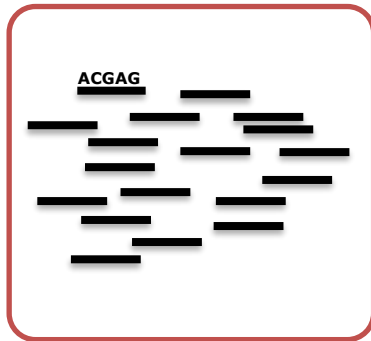
### ❖ **Quality Assessment**

Genes: 98% CS genes found

TEs: 92% CS ISBPs found

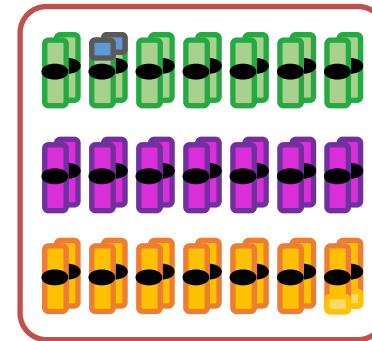
QV: lower (99.95% accuracy) than NRGene-based assemblies (99.99%)

## □ *Step#4: pseudomolecule construction*



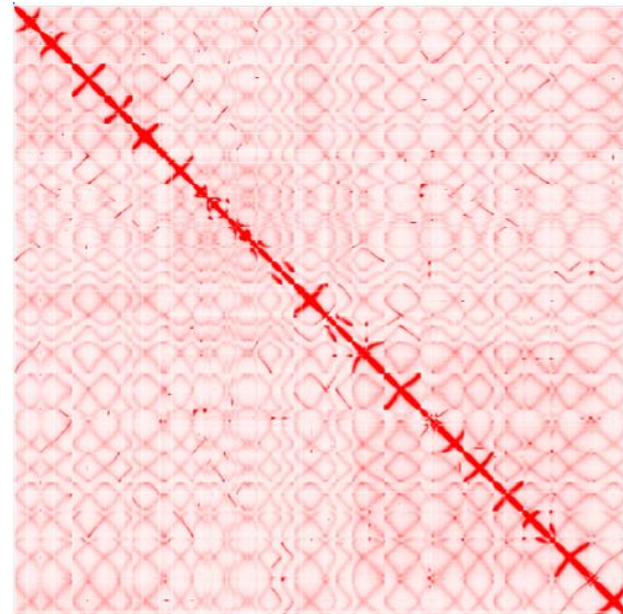
2904 scaff

1. CS-guided
2. HiC

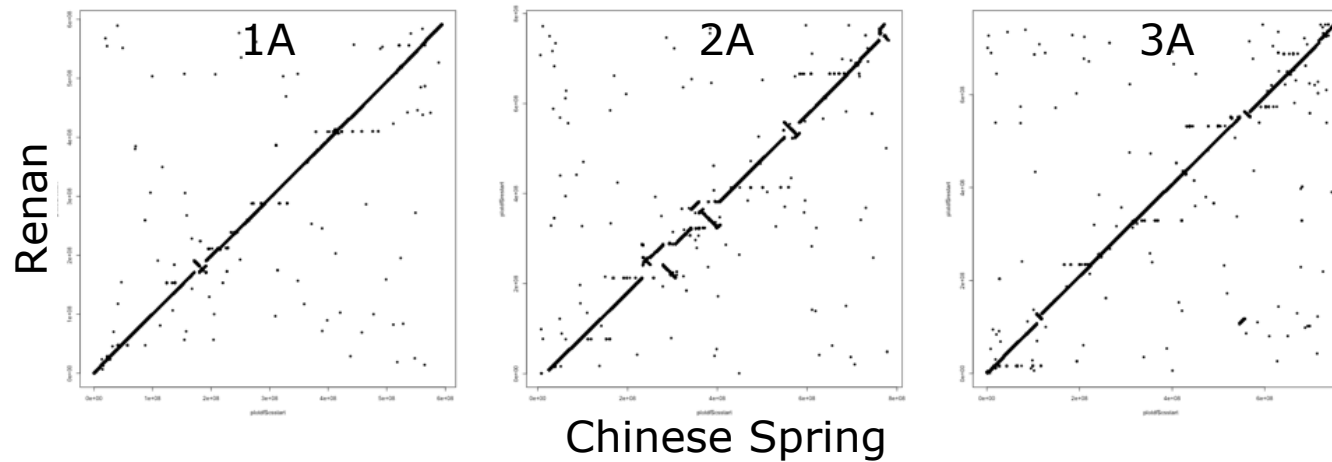
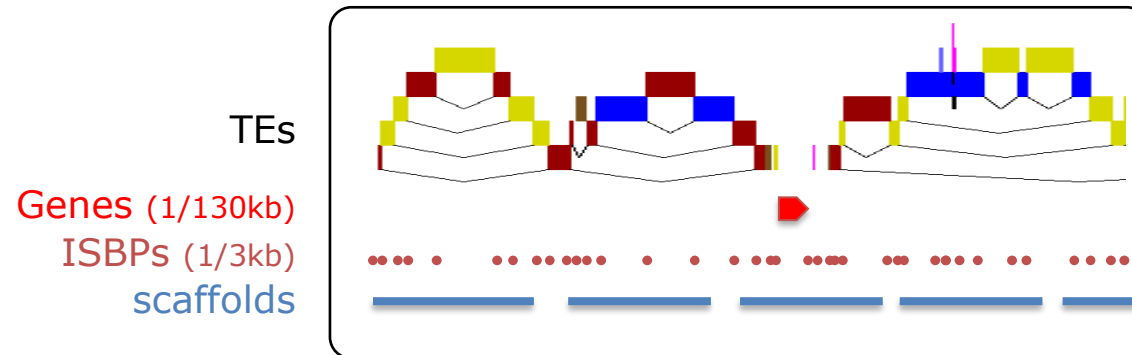


21 chromosomes

**X** not efficient enough to build a chromosome-scale assembly

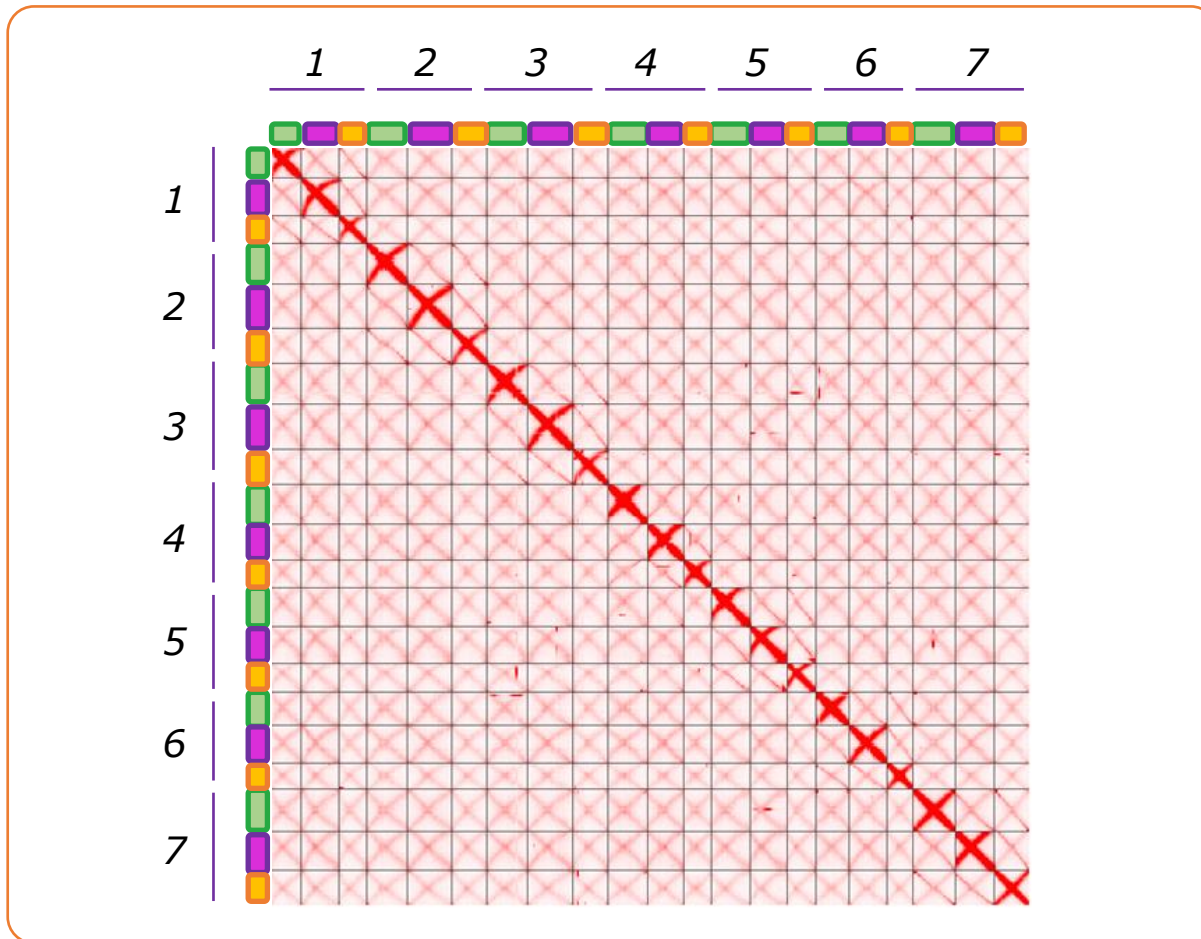


## □ **Step#4-1: CS-guided pseudomol construction**



⇒ 14.2 Gb (0.1 Gb unanchored)

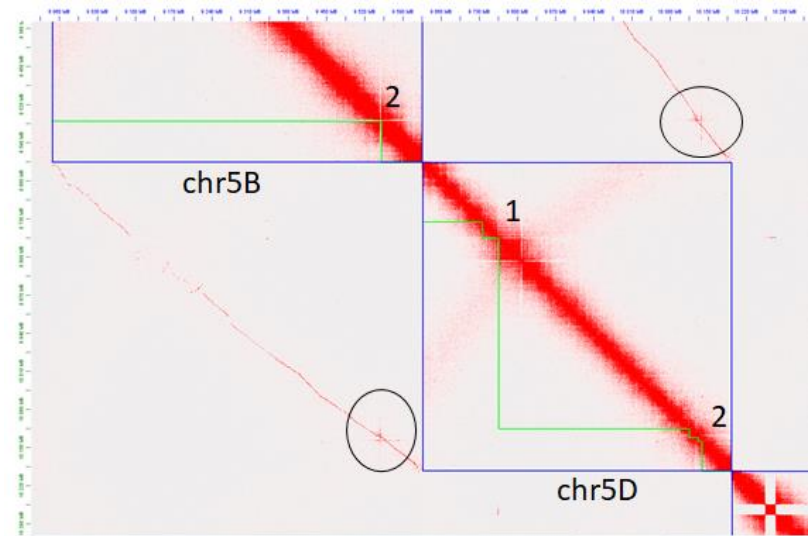
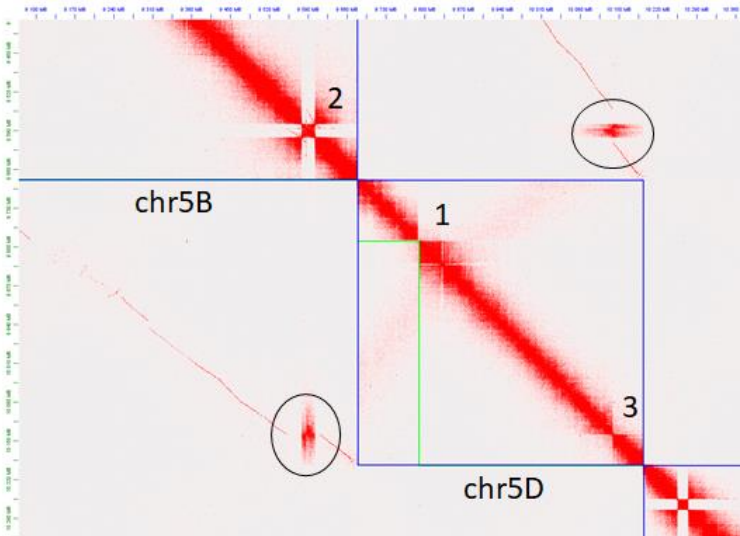
- ❑ *Step#4-1: CS-guided pseudomol construction*
- ❑ **Step#4-2: HiC corrections**



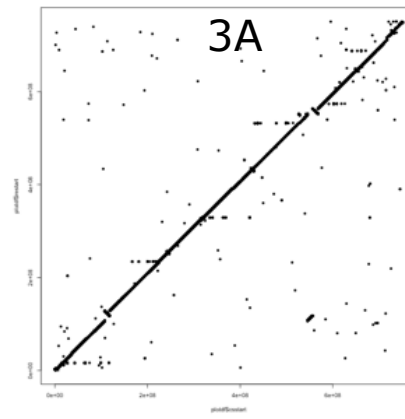
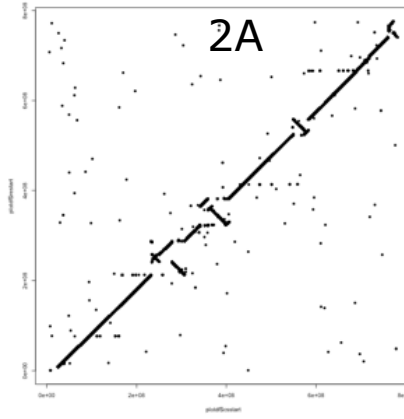
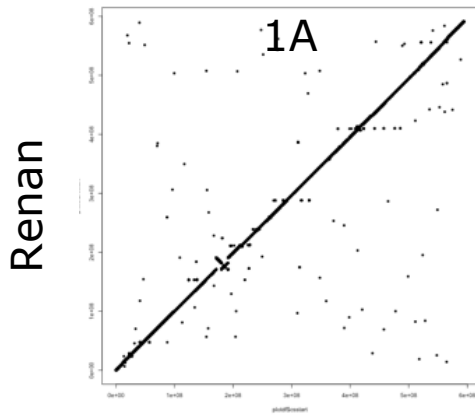
*HiC*



## □ **Step#4-2: HiC corrections**



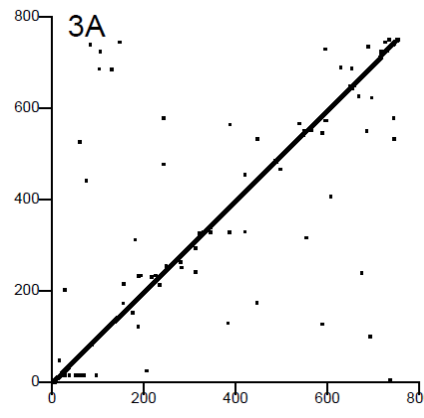
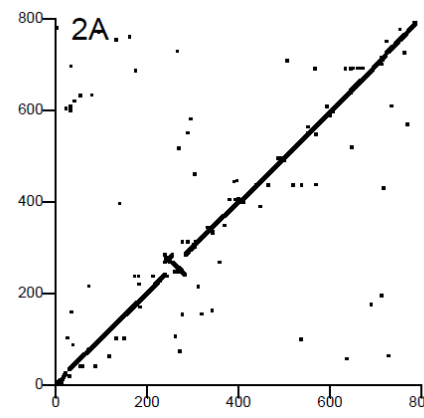
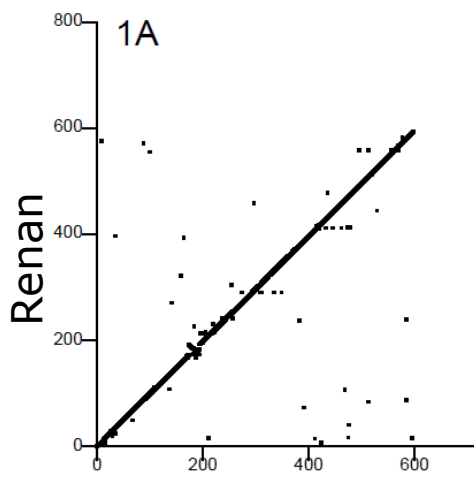
- **18** chimeric scaff
  - **198** scaff with correction of location and/or orientation
- ↓
- 21 chr + chrUn 61 Mb (<1%)



Chinese Spring

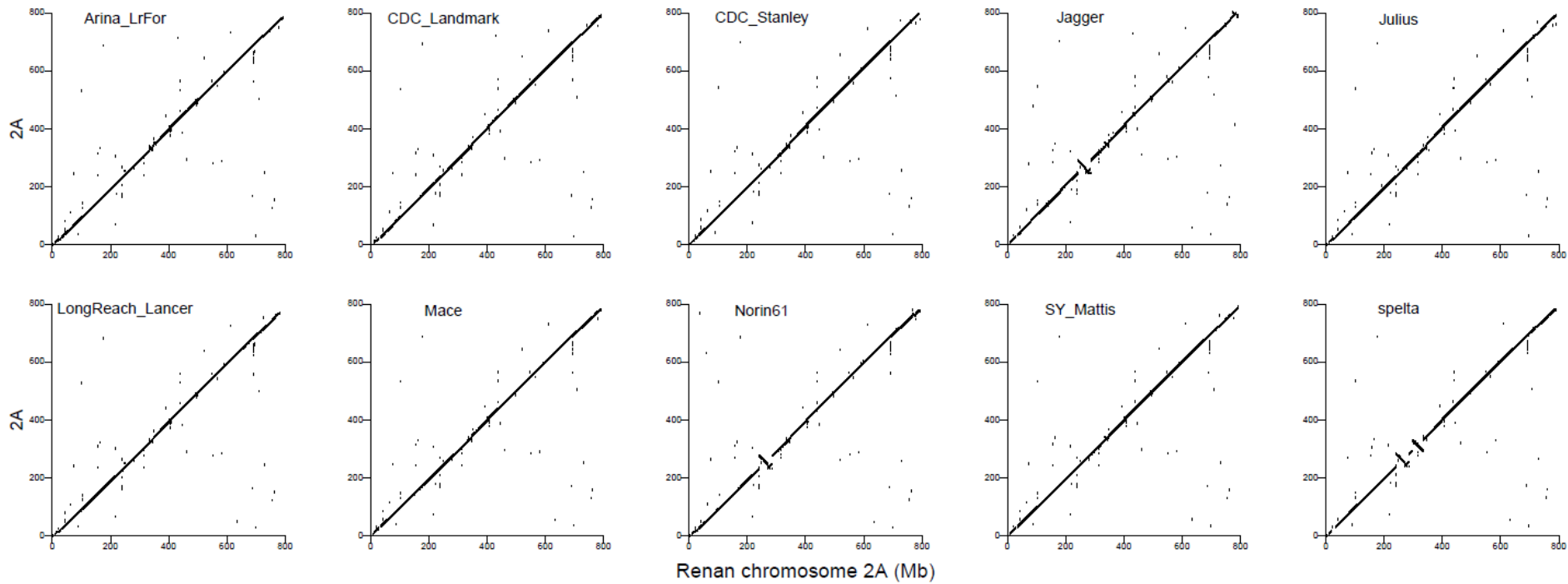


After HiC-based corrections



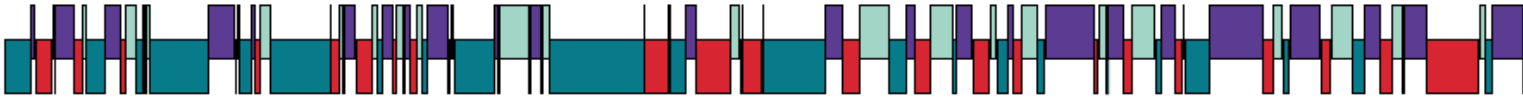
Chinese Spring

# Renan Refseq\_v2 versus 10 wheat genomes





Jagger



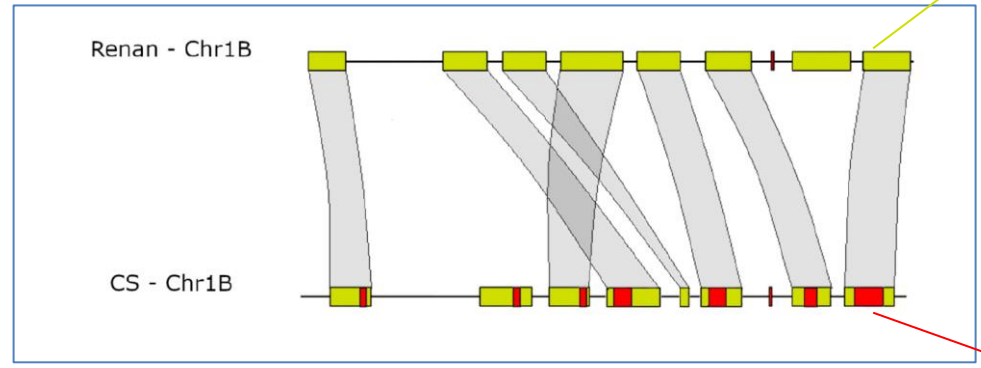
Renan



Contig N50=2 Mb

~1.6 Mb

### ω-gliadin gene cluster



```

ATGAAGACCTTCATCATATTTGTCTCCTTGCTATGGTGATGAGCATCGCCAGTGCCACTAGGCAACTAA
GCCCTAGAAGCAAGGAGTTGCAAAACACCACAAGAACAATCCCCAACAGCAATTTCC TCAACCAACA
AATCTTCCAACAACAATCCCCAACACAACAATCCCCAGCAACCGCAACAATCCCCAACACA
ATCCCGCAACAGCAAAATCCCGCAACAACAATCCCCAACACAACAATCCCCAGCAACCCACAACA
TCCCCCAACAAGCAATTTCCCAACAACAACAATCCCCAGCAACCCACAACAATCCCCCAACA
GCAATTTCCCAACAACAACAATCTCCAGCAACCCACAACAATCCCCAACACAACAATCCCCCA
ACAATTTCCCAACAACAATCCCCGAACA // AACAAATTTCCCAACAAGCAATTTCCCAACAACA
AATTTCCCAACAACAAGTTACCGCAACAACAATTTCCCAACAACAACAATTTCCCGAGCAACCCACA
ACAATTTCCCAACAACAACAATTTCCCAACAACAACAATTTCCCAACAAGCAATTTCCCGAACAACA
TTCCCAACAACAACAATTTCCCAACAACAACAAGTTCCCTCAACAACAACAATTTCCCGCAACAACA
TCCCTGAAACAACAATAATTTCCCAACAACAATTTCCCAACAACAATTTCCCAACAACA
AATTTCCAATACCAATACCCACCCACAACAATCACAAGAACCCTCCCATACCAACAATATCCAACAACA
CCATATGGGAGCAACGTTATAAGTATCAGTGGCCGATGA
  
```

```

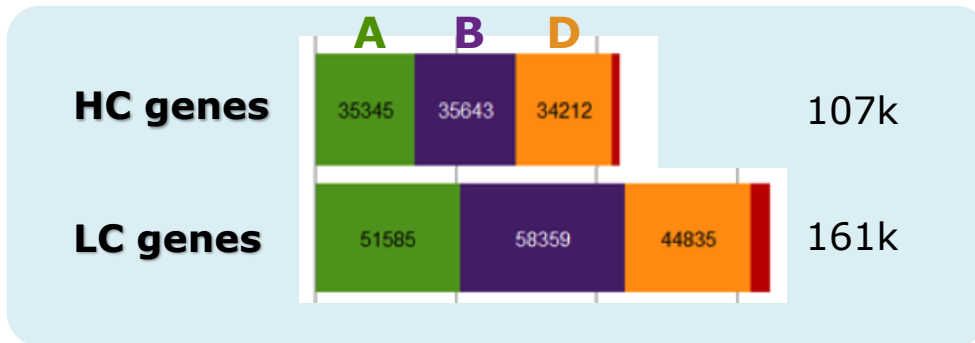
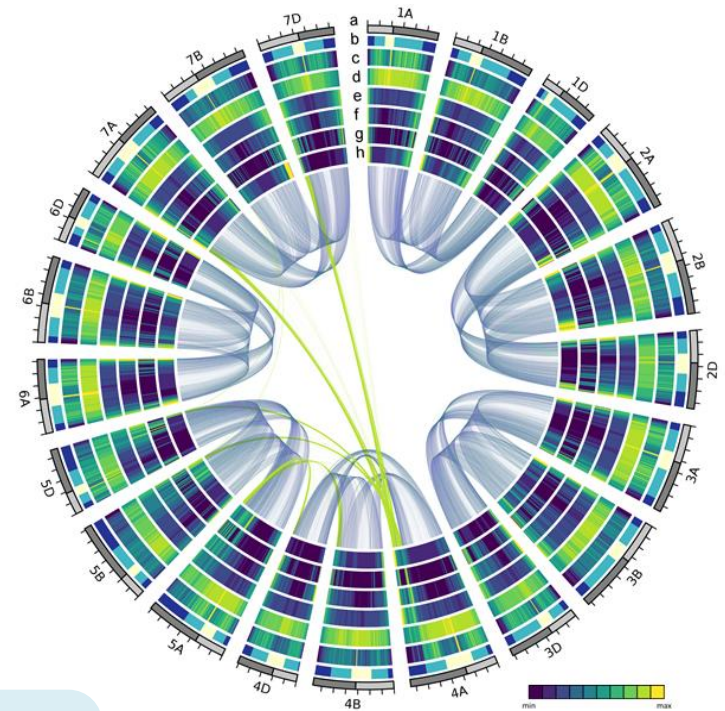
ATGAAGACCTTCATCATATTTGTCTCCTTGCTATGGTGATGAGCATCGCCAGTGCCACTAGGCAACTAA
GCCCTAGAAGCAAGGAGTTGCAAAACACCACAAGAACAATCCCCAACAGCAATTTCC TCAACCAACA
AATCTTCCAACAACAATCCCCAACACAACAATCCCCAGCAACCGCAACAATCCCCAACACA
ATCCCGCAACAGCAAAATCCCGCAACAACAATCCCCAACACAACA NNNNNNNNNNNNNNNNNNN
NNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNN
NNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNN
NNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNN // NNNNNNNNNNNNNNNNNNN
NNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNN
NNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNN
NNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNN
NNNNNNNNNACAACAATTTCCCAACAACAACAACAATTTCCCAACAACAATTTCCCATACCATACC
CACCCCAACAACAATCACAAGAAGCTTTCCCATACCAACAATACCAACAACAACAATCTGGGAGCGAC
GTTATAA
  
```

# ○ Predicting genes is still NOT routine

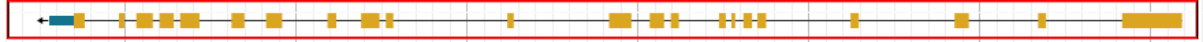
**GDEC** Clermont, France – *Rimbert Leroy Choulet et al.*  
**PGSB** Munich, Germany – *Spannagl Twardziok et al.*  
**EI** Norwich, UK – *Swarbreck Venturini et al.*

IWGSC RefSeq (Chinese Spring)

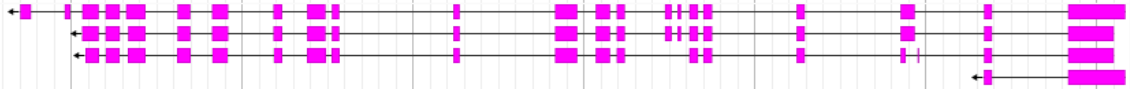
- 107,891 genes



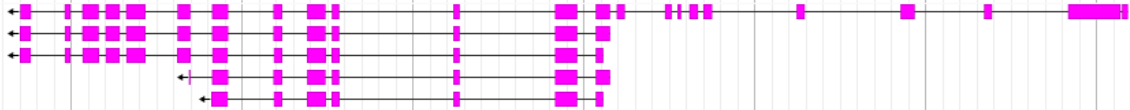
⊗ HighConfidenceGenesv1.1 (GOMAP annotation available)



⊗ Sorghum bicolor



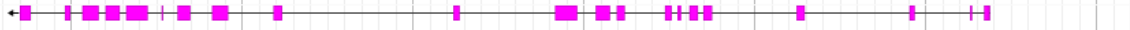
⊗ Hordeum vulgare var. distichum



⊗ Aegilops tauschii



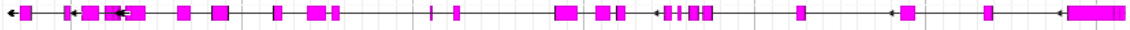
⊗ Triticum urartu



⊗ Brachypodium distachyon



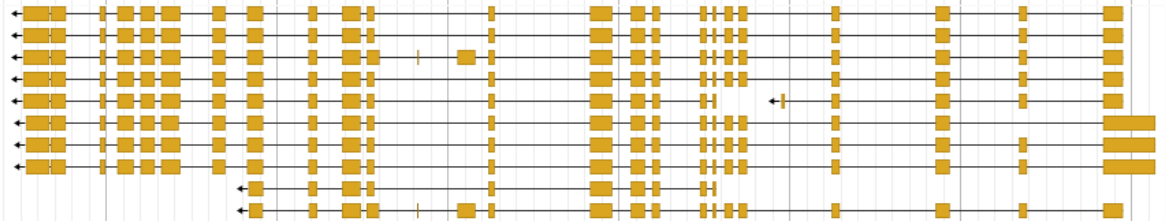
⊗ Oryza barthii

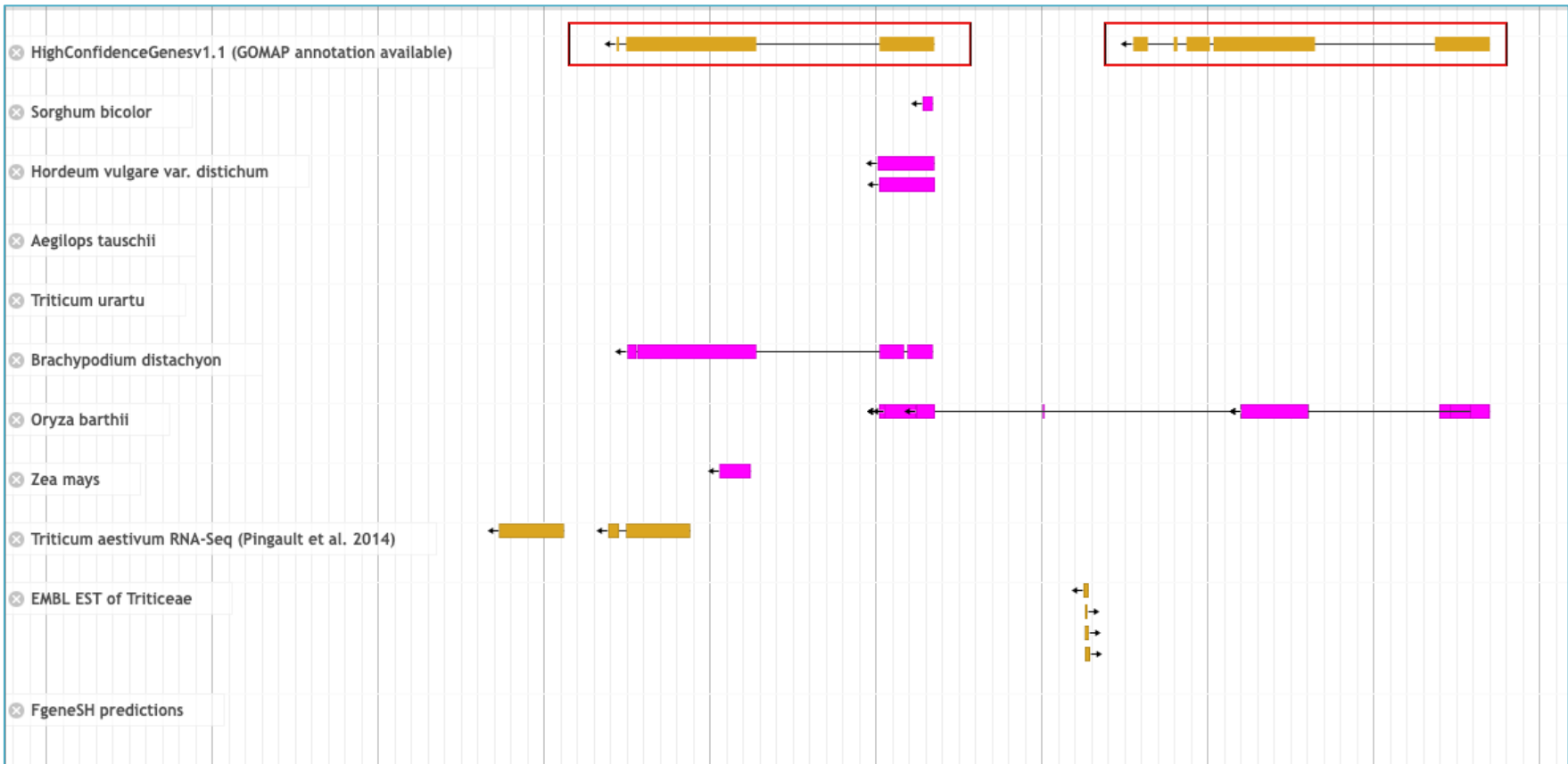


⊗ Zea mays

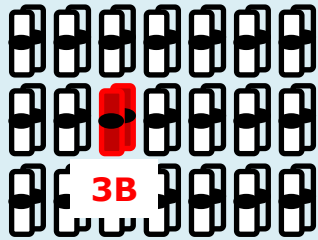


⊗ Triticum aestivum RNA-Seq (Pingault et al. 2014)



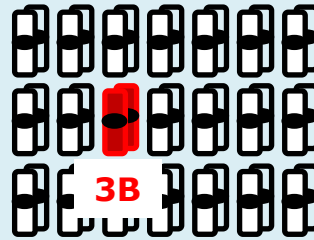


chr3B BAC-by-BAC



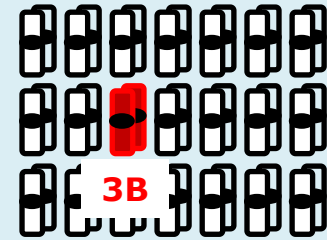
*Choulet et al. 2014*

TGAC\_v1



*Clavijo et al. 2017*

Chr. Survey Seq



*IWGSC 2014*

7264 genes

5728 genes

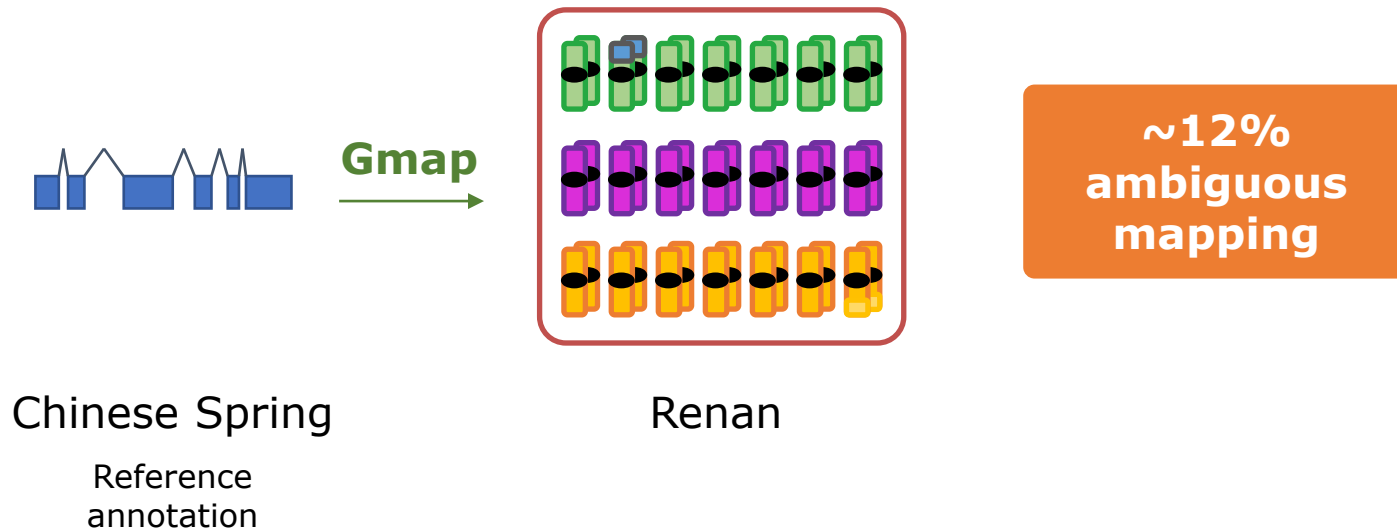
5862 genes

(**26%** identical)

(**12%** identical)

➤ *denovo* gene annotation not adapted to pangenomics

- Gene projections / Annotation transfer



- *denovo* gene annotation not adapted to pangenomics

- Gmap vs whole genome introduces mis-alignment errors

- Develop **MAGATT** pipeline

M

magatt

Project ID: 1089

☆ Star 1

120 Commits 1 Branch 5 Tags 5.5 MB Project Storage

Pipeline used to tranfert gene annotation (GFF3) between different versions of assemblies.

master

magatt

Find file

↓

Clone



Upload New environment file for Singularity build

Helene Rimbart authored 10 months ago

4df3e943



README

No license. All rights reserved

Name	Last commit	Last update
bin	NEW single-isoform mode for transfert b...	1 year ago
report	bug: if cds and mapping attributes existed...	1 year ago
rules	NEW single-isoform mode for transfert b...	1 year ago
.gitignore	ignore slurm log file	1 year ago
README.md	Update README.md	1 year ago
Singularity	Upload New Singularity definition file	10 months ago
Snakefile	bug: if cds and mapping attributes existed...	1 year ago
cluster-hpc2.json	NEW single-isoform mode for transfert b...	1 year ago
cluster-sibi.json	remove temporary fasta files to lower di...	1 year ago
config.yaml	NEW single-isoform mode for transfert b...	1 year ago
env.yaml	Upload New environment file for Singolari...	10 months ago
environment.yml	Update environment.yml with new depend...	1 year ago

README.md

## MAGATT pipeline

Marker Assisted Gene Annotation Transfert for Triticeae.

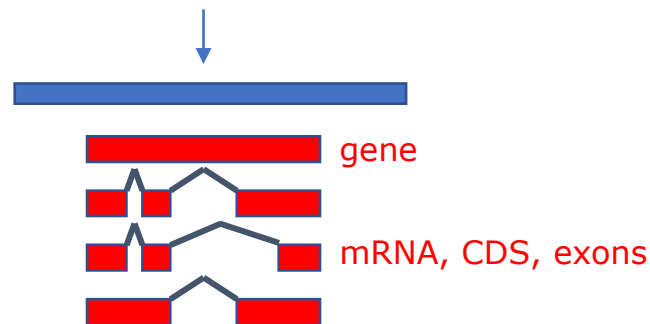
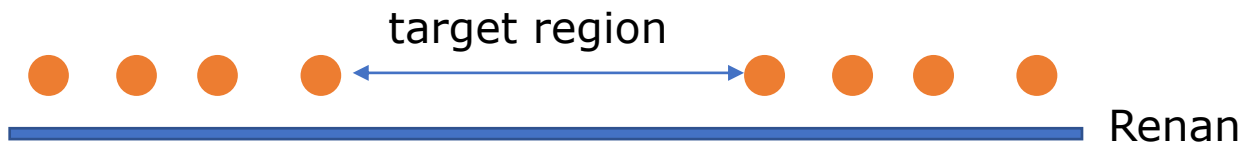
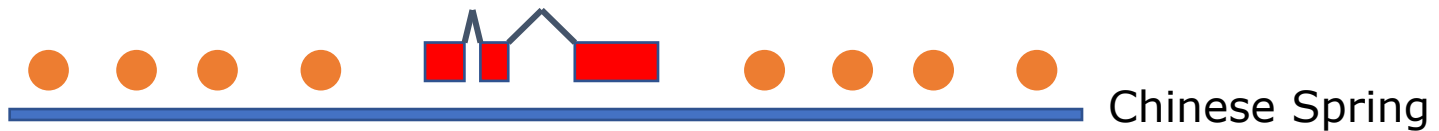
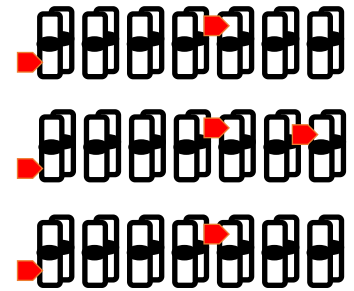
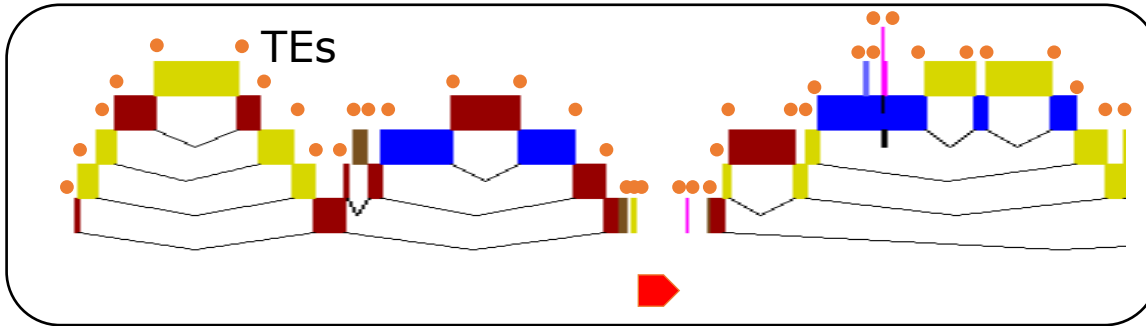
Snakemake pipeline used to transfert GFF annotation on a new assembly with a fine target mapping approach.

### Install the pipeline

```
$ git clone https://forgemia.inra.fr/umr-gdec/magatt.git
```

# □ **MAGATT** (Marker-Assisted Gene Annotation Transfer for *Triticeae*)

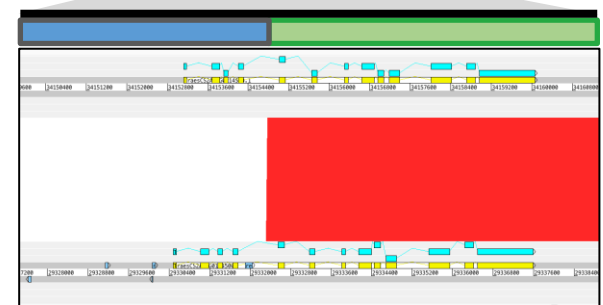
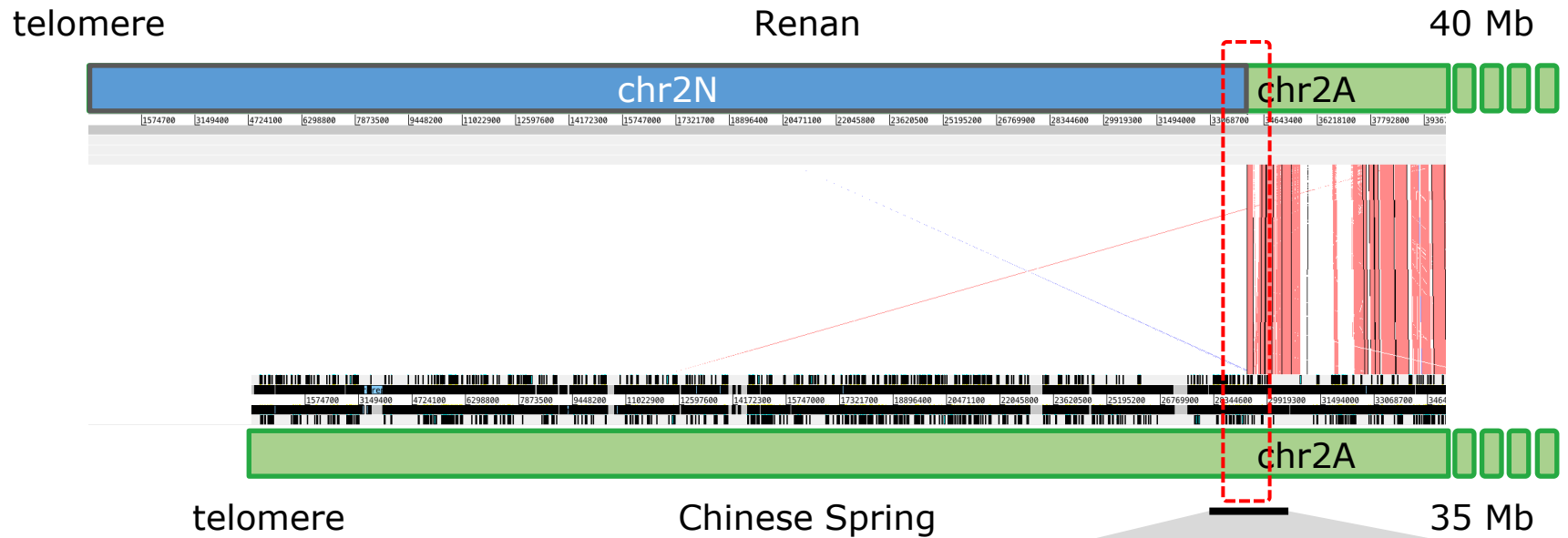
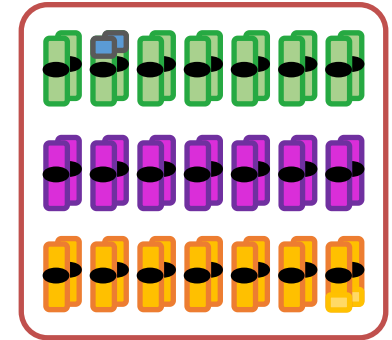
ISBPs (uniquely mappable)

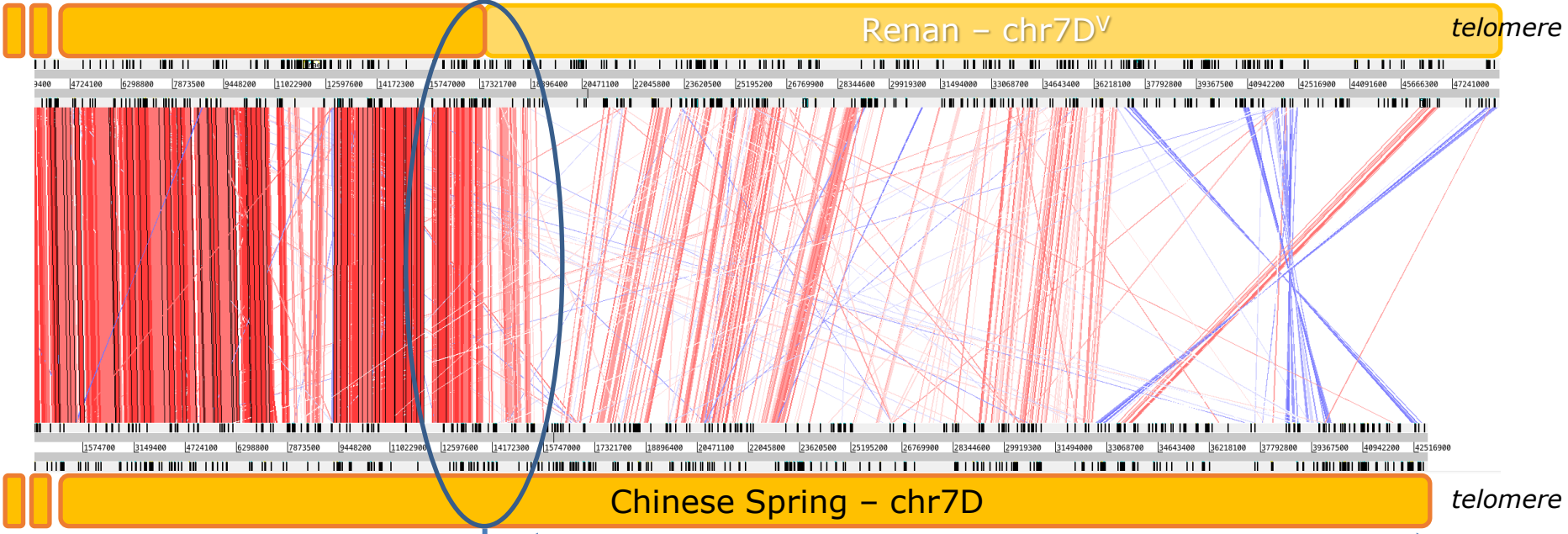


- 98% of genes mapped accurately
- + 4400 (4%) Renan specific genes

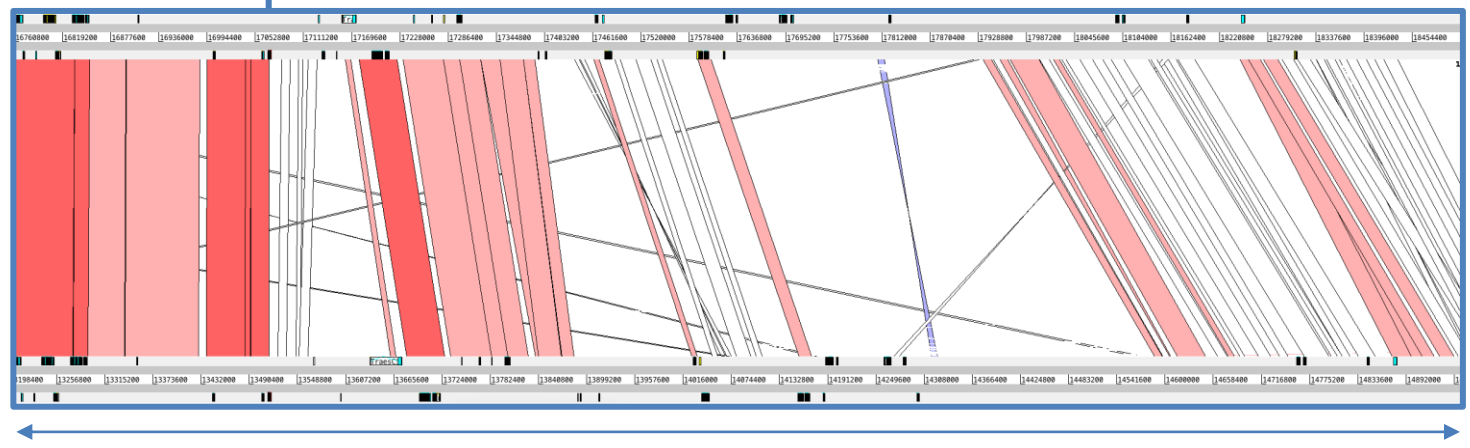


# □ *Introgressions*



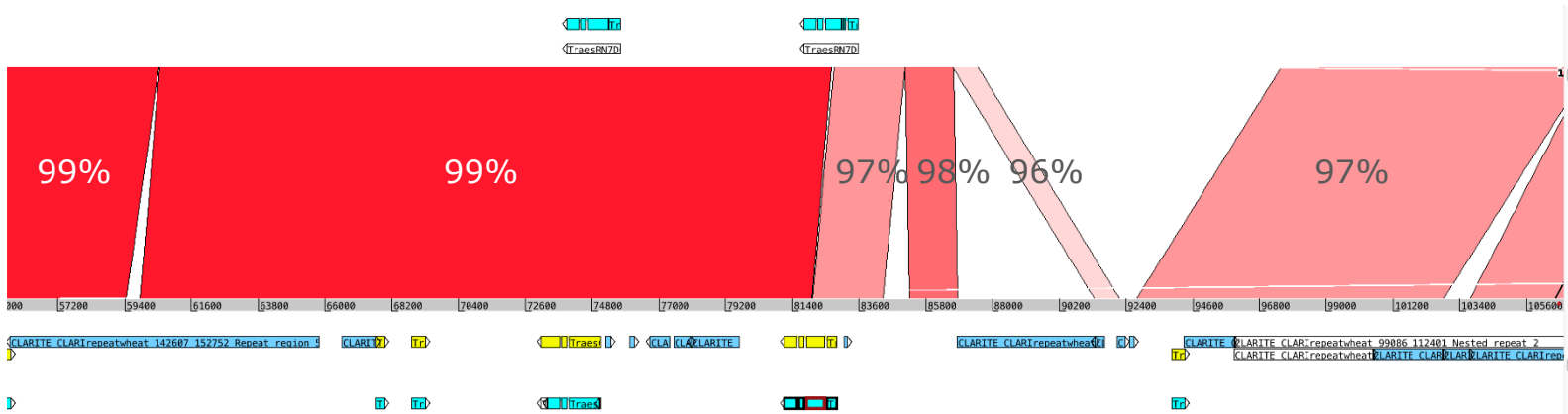


30 Mb, ~500 genes



## Renan – chr7D

## Renan – chr7D<sup>V</sup>




## Chinese Spring – chr7D

# □ TE dynamics

- PhD Nathan Papon (2019-2022)

*Daron et al. 2015*

Daron et al. *Genome Biology* 2014, 15:546  
<http://genomebiology.com/2014/15/12/546>



**RESEARCH** **Open Access**

## Organization and evolution of transposable elements along the bread wheat chromosome 3B

Josquin Daron<sup>1,2</sup>, Natasha Glover<sup>1,2</sup>, Lise Pingault<sup>1,2</sup>, Sébastien Theil<sup>1,2</sup>, Véronique Jamilloux<sup>3</sup>, Etienne Paux<sup>1,2</sup>, Valérie Barbe<sup>4</sup>, Sophie Mangenot<sup>4</sup>, Adriana Alberti<sup>4</sup>, Patrick Wincker<sup>4,5,6</sup>, Hadi Quesneville<sup>3</sup>, Catherine Feuillet<sup>1,2</sup> and Frédéric Choulet<sup>1,2\*</sup>

*Wicker et al. 2018*

Wicker et al. *Genome Biology* (2018) 19:103  
<https://doi.org/10.1186/s13059-018-1479-0>



**RESEARCH** **Open Access**



## Impact of transposable elements on genome structure and evolution in bread wheat

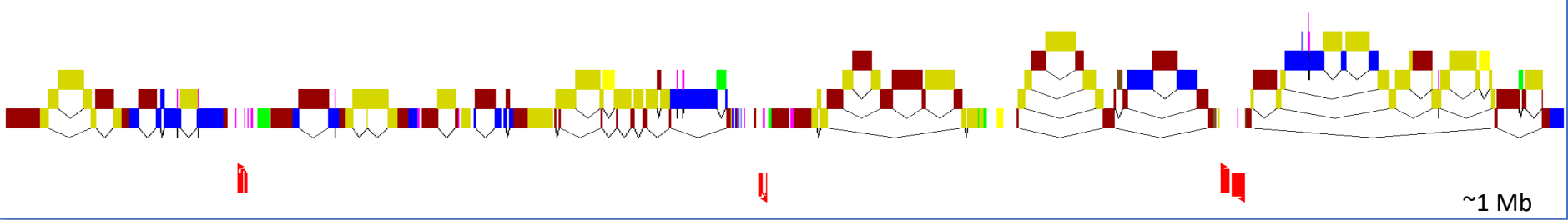
Thomas Wicker<sup>1†</sup>, Heidrun Gundlach<sup>2†</sup>, Manuel Spannagl<sup>2</sup>, Cristóbal Uauy<sup>3</sup>, Philipp Borill<sup>3</sup>, Ricardo H. Ramírez-González<sup>2</sup>, Romain De Oliveira<sup>4</sup>, International Wheat Genome Sequencing Consortium<sup>5</sup>, Klaus F. X. Mayer<sup>2,6</sup>, Etienne Paux<sup>4</sup> and Frédéric Choulet<sup>1†\*</sup>

*De Oliveira et al. 2020*

Frontiers in Genetics | [www.frontiersin.org](http://www.frontiersin.org) August 2020 | Volume 11 | Article 891

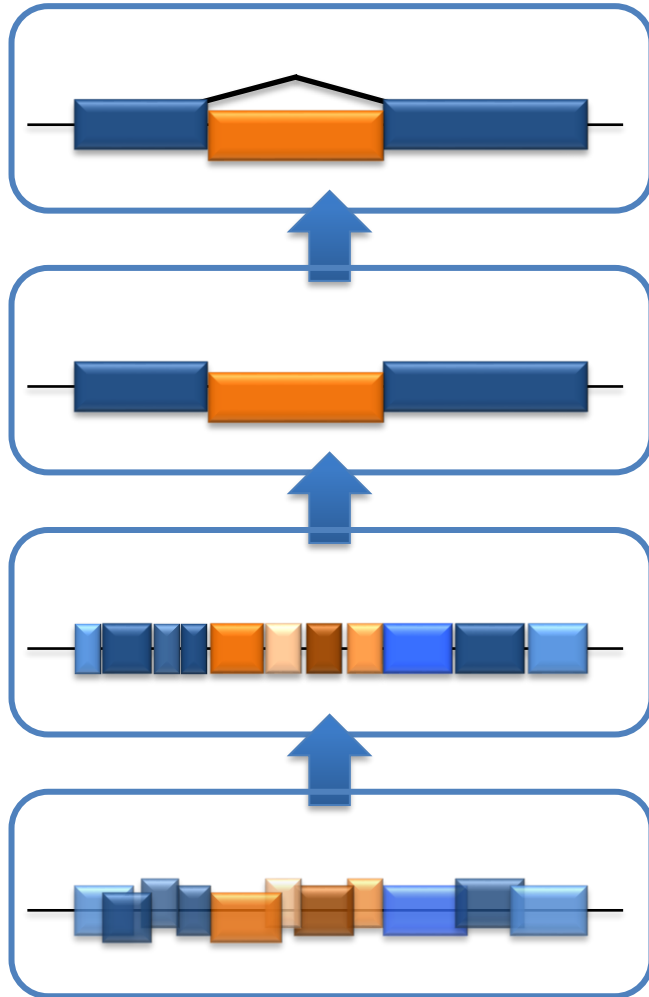
## Structural Variations Affecting Genes and Transposable Elements of Chromosome 3B in Wheats

Romain De Oliveira<sup>1</sup>, Hélène Rimbart<sup>1</sup>, François Balfourier<sup>1</sup>, Jonathan Kitt<sup>1</sup>, Emeric Dynomant<sup>1</sup>, Jan Vrána<sup>2</sup>, Jaroslav Doležel<sup>2</sup>, Federica Cattonaro<sup>3</sup>, Etienne Paux<sup>1</sup> and Frédéric Choulet<sup>1\*</sup>

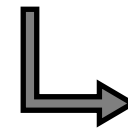


# ○ TE modeling with **CLARITE** and **ClariTeRep**

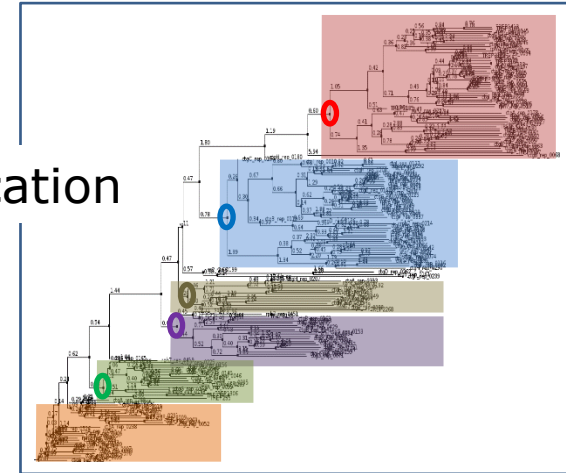
<https://github.com/jdaron/CLARI-TE>



TREP++  
5000 known wheat TEs

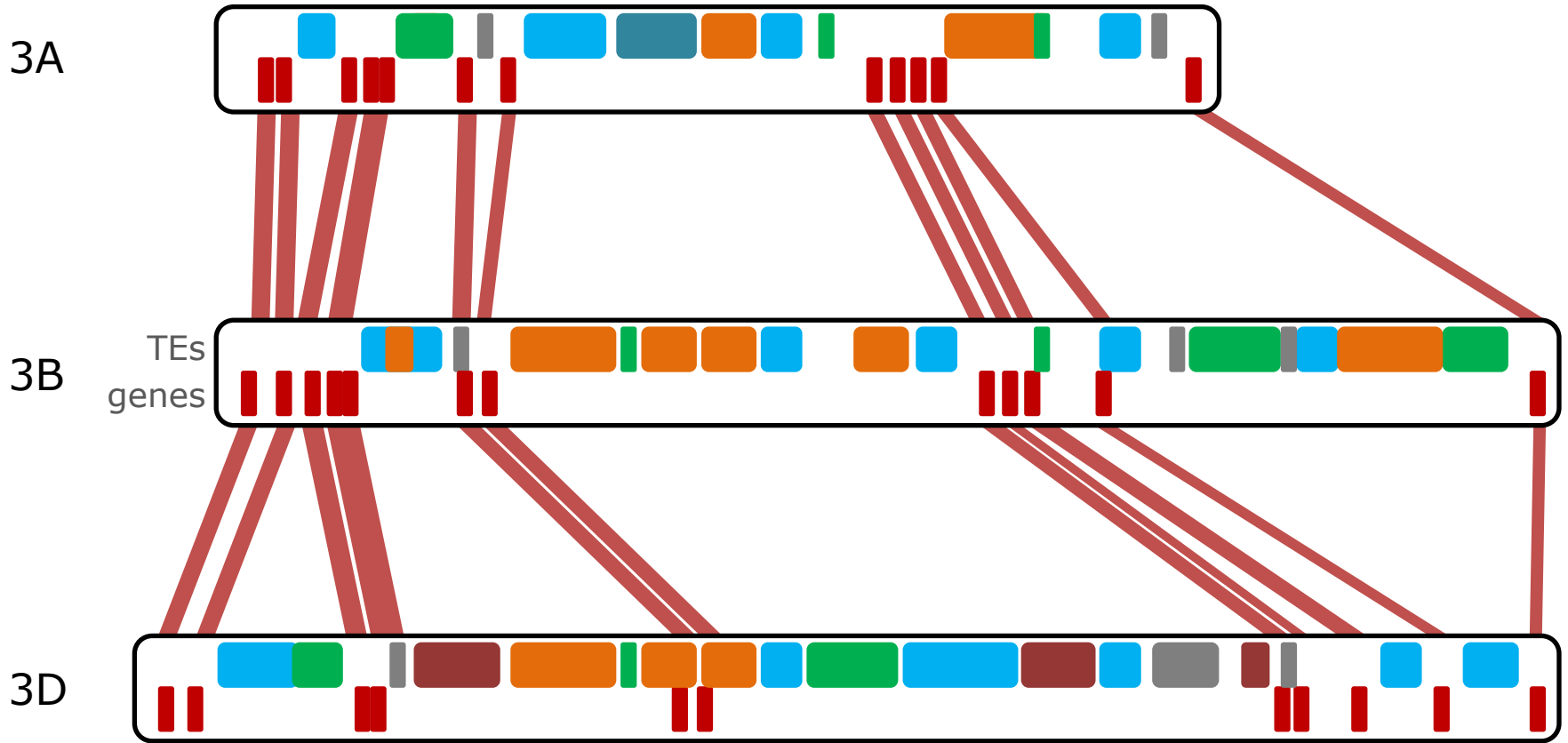


Classification

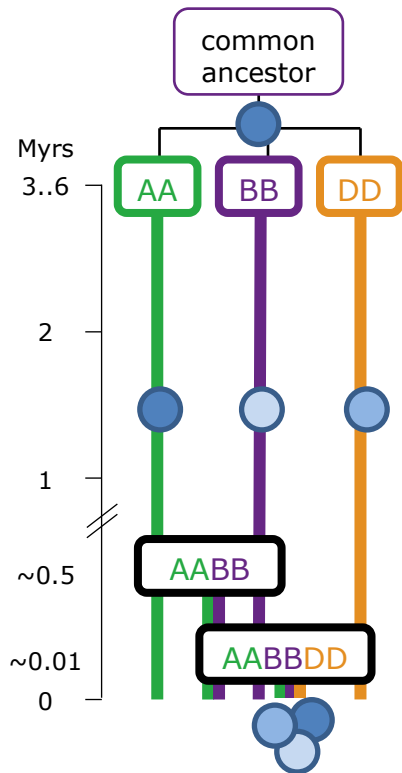


**ClariTeRep**  
~500 families

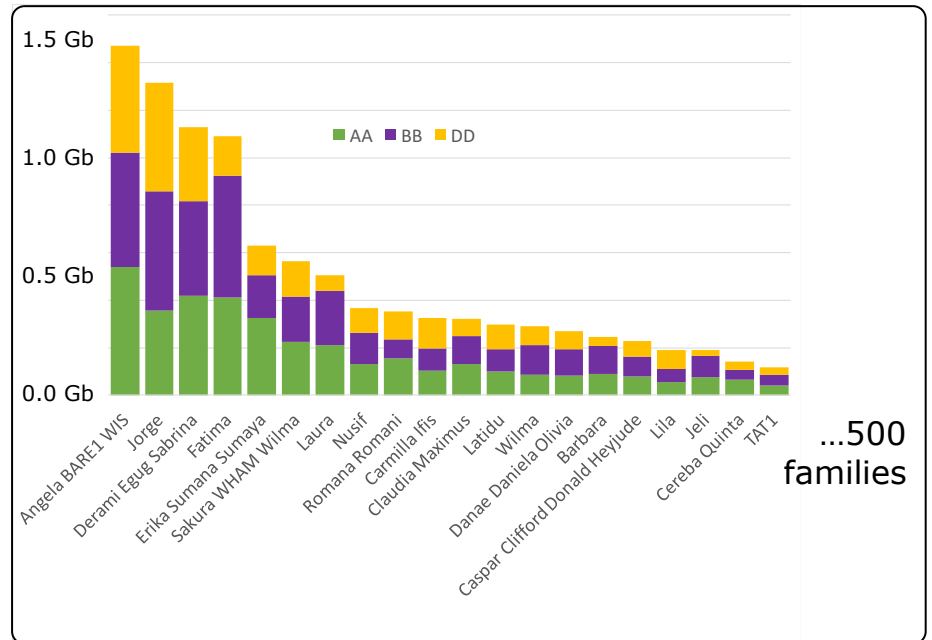




➤ Near-complete **TE turnover** since A-B-D divergence



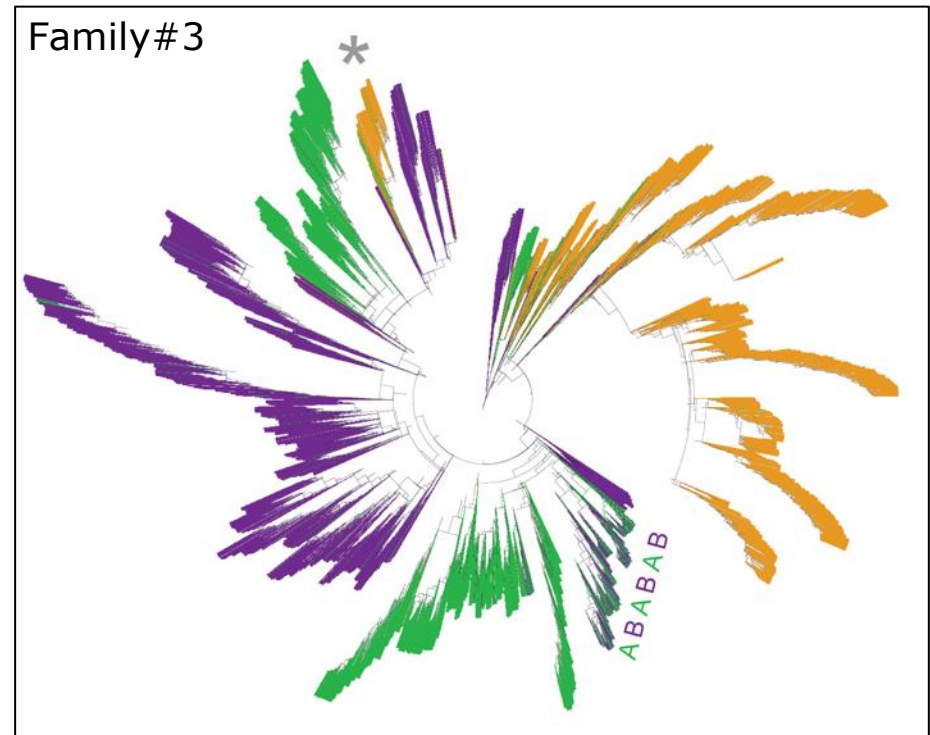
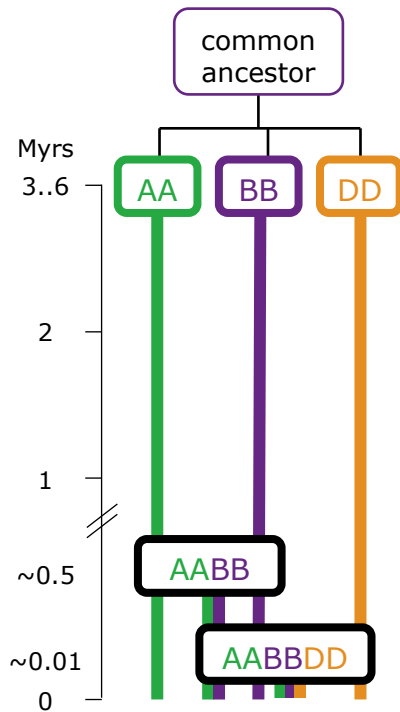
## Families



➤ A-B-D balanced proportions

➤ **0** family specific to 1 subgenome



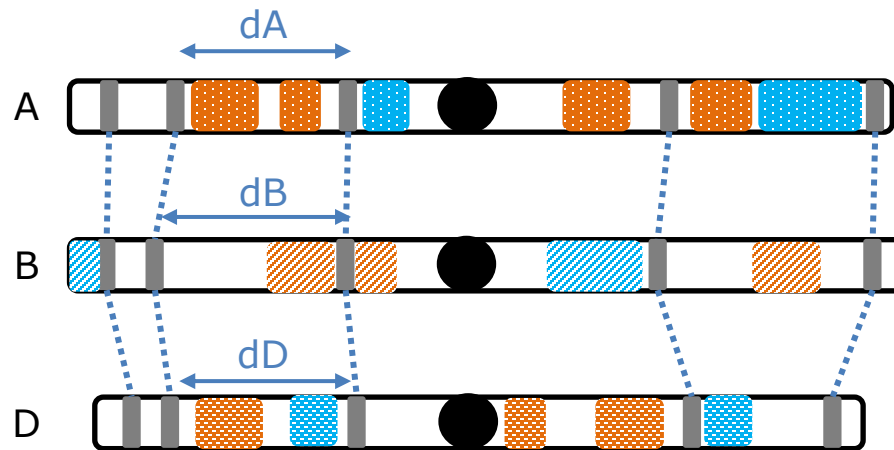


- Independent TE evolution in the diploids AA, BB, DD
- Some rare cases of TE amplif in the tetraploid AABB

A-B-D last  
common ancestor



~ 3 Myrs  
TE turnover



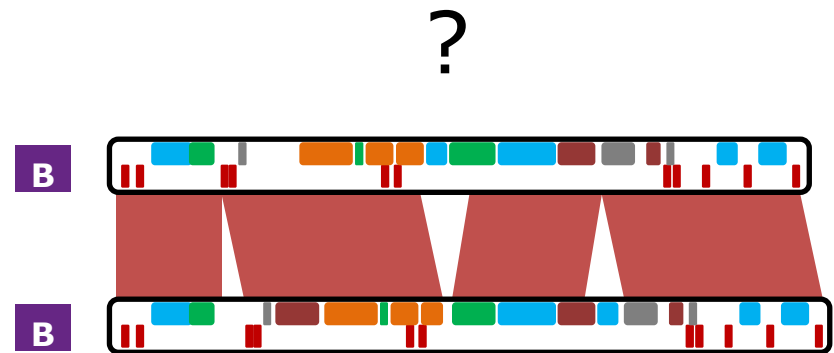
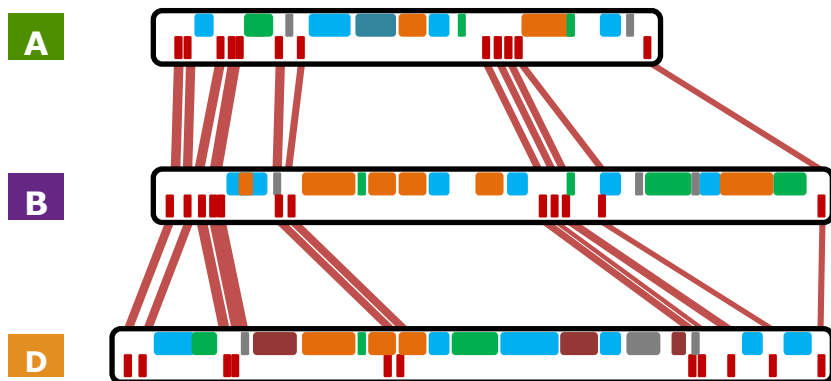
Most families were active since A-B-D divergence

➤ **Equilibrium**

Hypothesis of a structural role of TEs likely under selection pressure?

# □ Genomic variability affecting **TEs** in wheat **using assembled genomes**

*PhD Nathan Papon (2019-2022)*



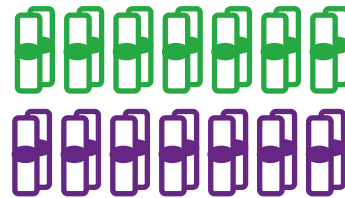
- Material = multiple RefSeq now available

*T. aestivum*



x13  
accessions

*T. dicoccoides*  
*T. durum*



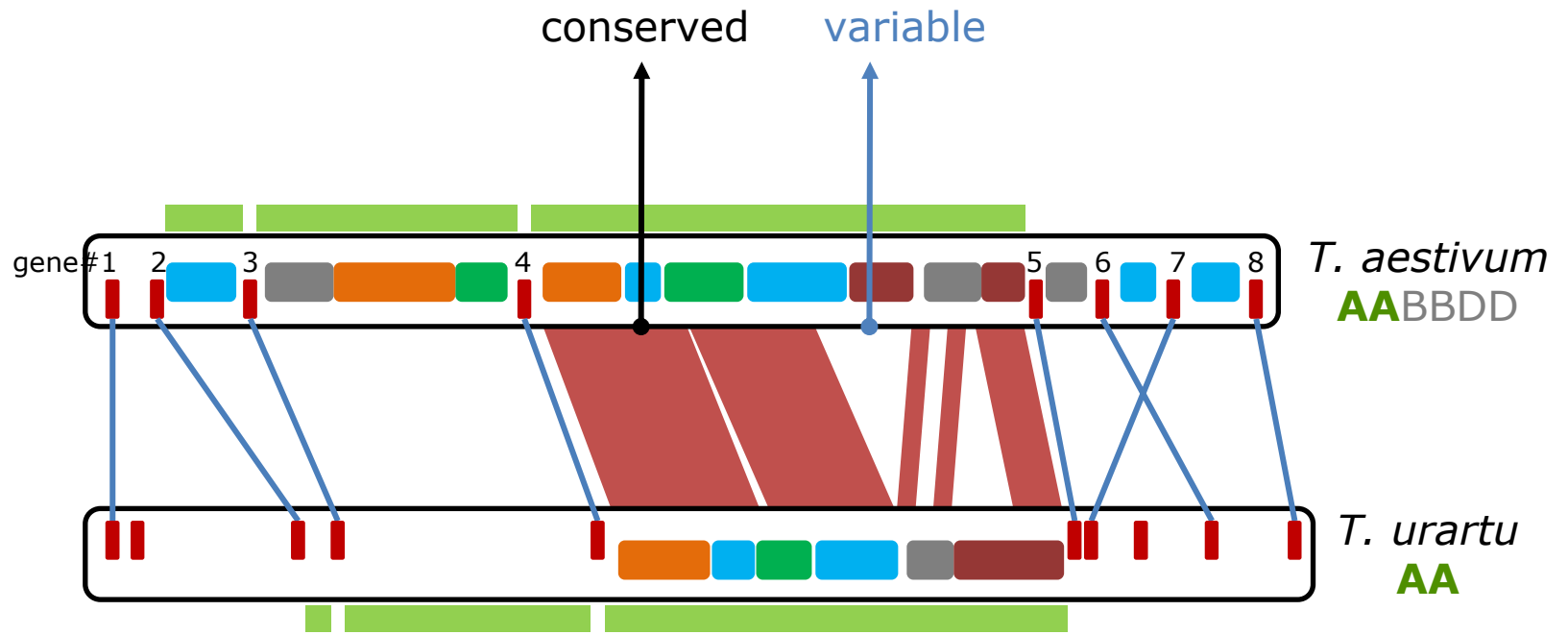
*T. urartu*



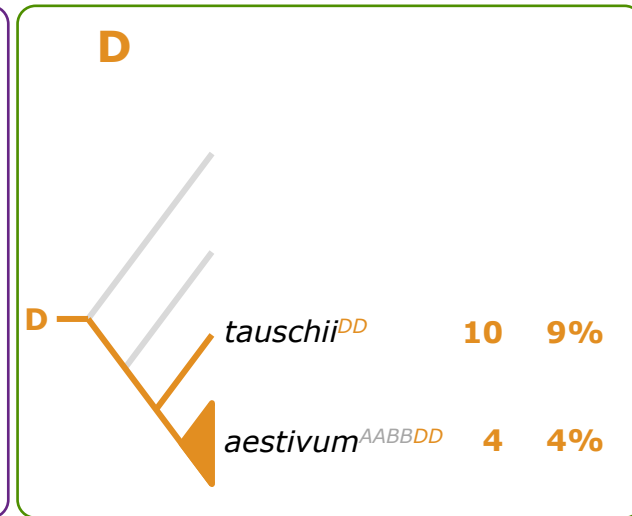
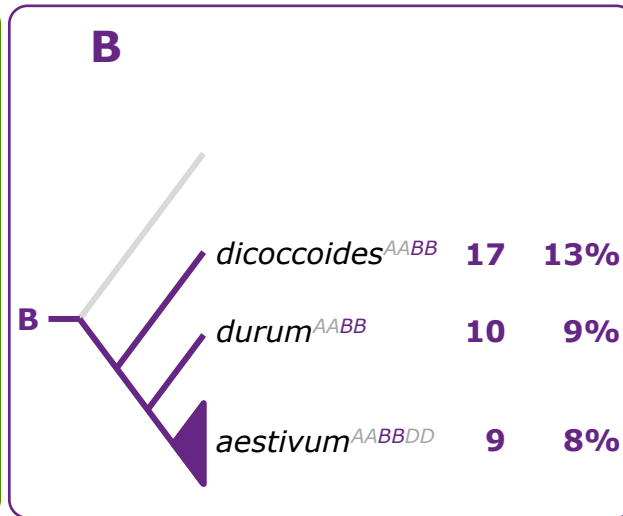
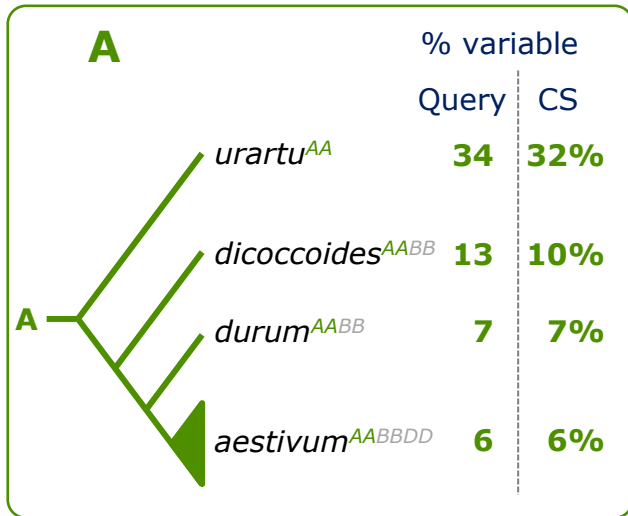
*Ae. tauschii*



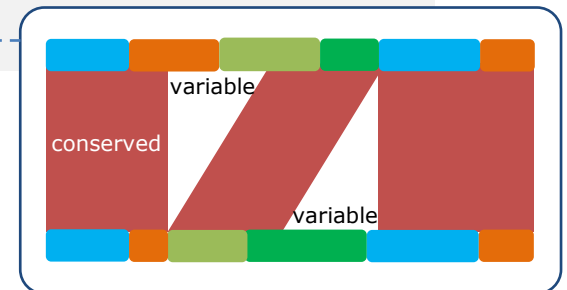
○ Method

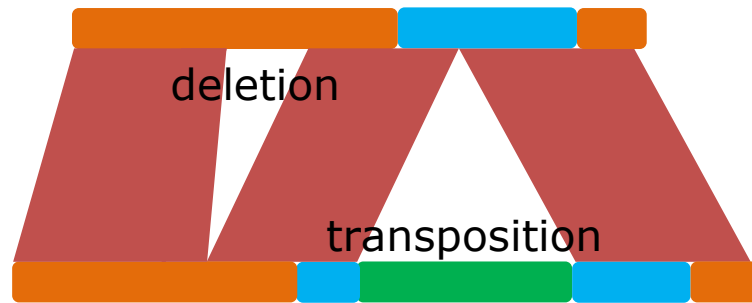


# Extent of variability affecting TEs



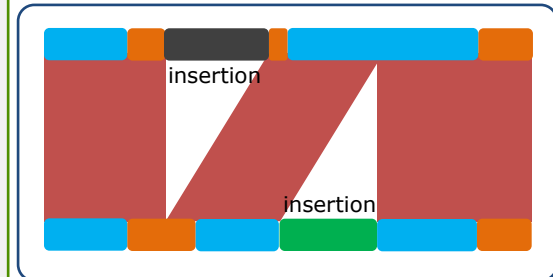
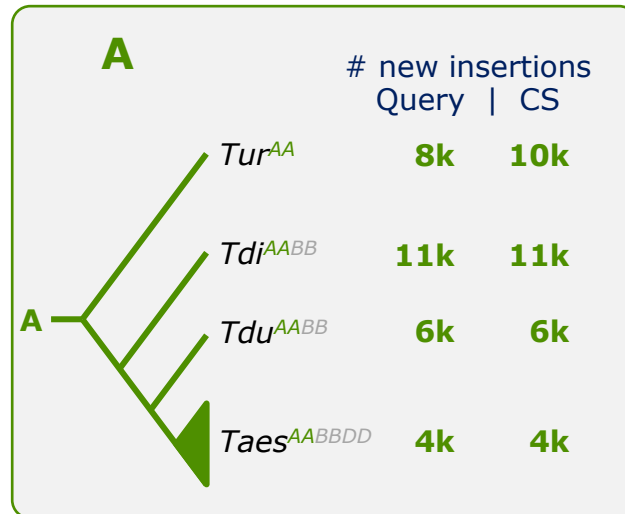
- 5% .. 15%
- B > A > D
- No genome expansion/contraction





## Recent TE insertions

- ~4,000–13,000 transposition events / subgenome (<1% of the genome)



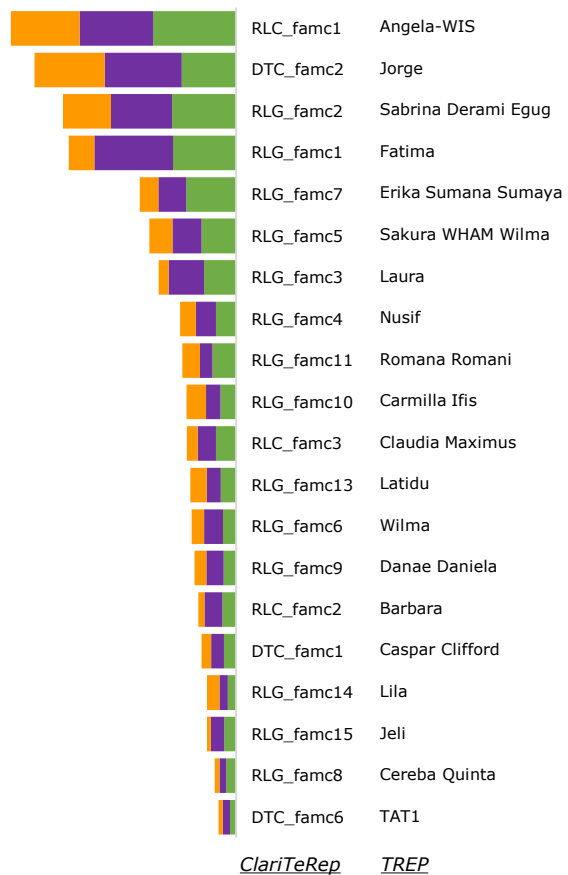
## Impact of polyploidy?

- none

## How many families were active recently?

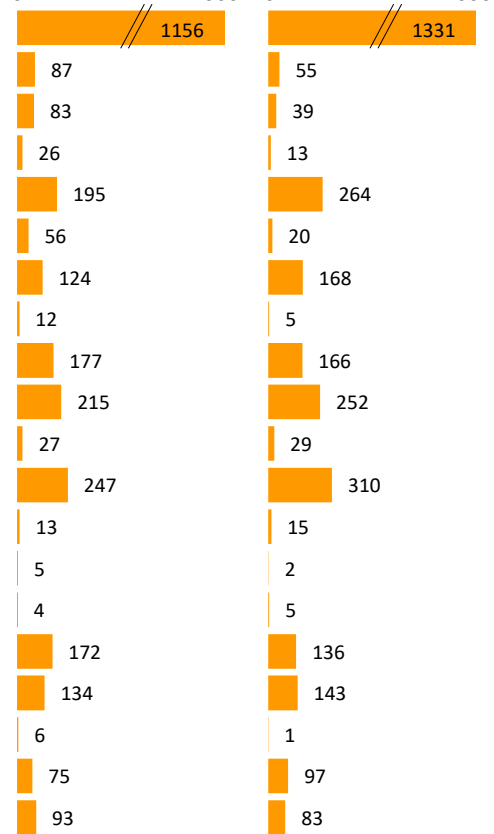
- 346 active families (=99.7% of all TEs)

## 20 most abundant families



## #specific TE insertions

0 1000 0 1000



CS<sup>AABBDD</sup>

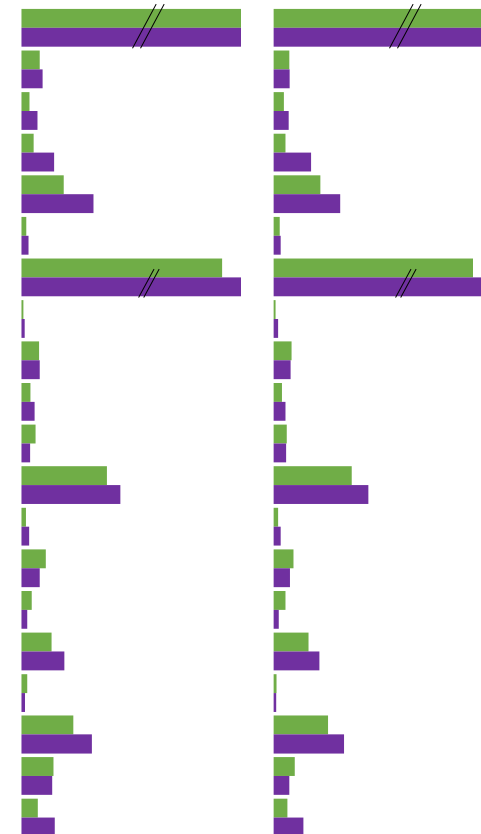
VS

*Ae. tauschii*<sup>DD</sup>

CS<sup>AABBDD</sup>

VS

*T. durum*<sup>AABB</sup>





# Conclusions

- No genome expansion/contraction due to TE transposition
  - No impact of polyploidy
  - No TE burst
  - All families were active recently
  - Transposition rate is stable
- 
- Equilibrium
  - Role of TEs in genome architecture under evolutionary constraints

# □ Acknowledgments

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SRESRI/Région ARA



La Région  
Auvergne-Rhône-Alpes