

15 Juin  
2023

# Contribution des cellules souches à la croissance musculaire chez le porc

DESSAUGE Frédéric



UMR-1348

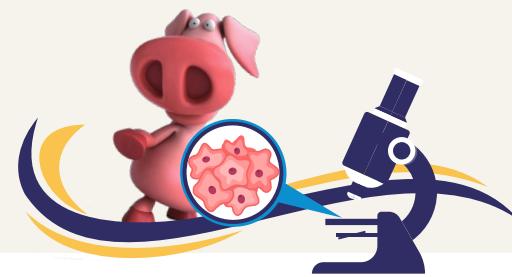
PHYSIOLOGIE, ENVIRONNEMENT  
ET GÉNÉTIQUE POUR L'ANIMAL  
ET LES SYSTÈMES D'ÉLEVAGE (PEGASE)

L'INSTITUT  
**agro** Rennes  
Angers

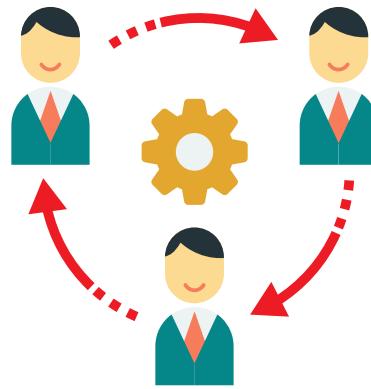
Journées Muscle et Qualités / 15-16 juin 2023



# Présentation



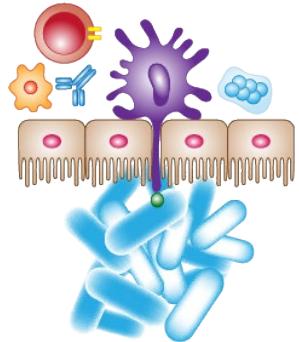
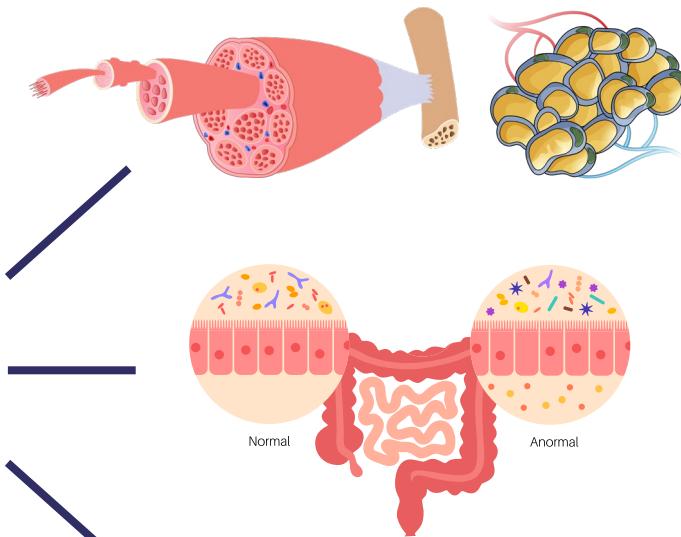
## L'UMR PEGASE en chiffres



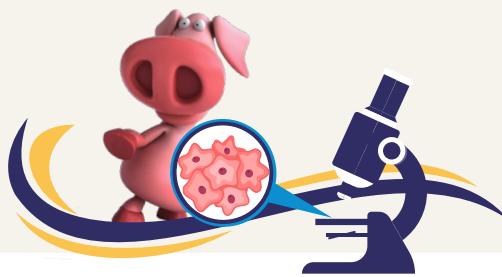
**120**  
collaborateurs

**4**  
Équipes de recherche

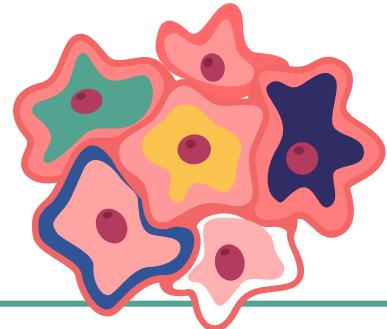
**1**  
Equipe Biologie des  
fonctions



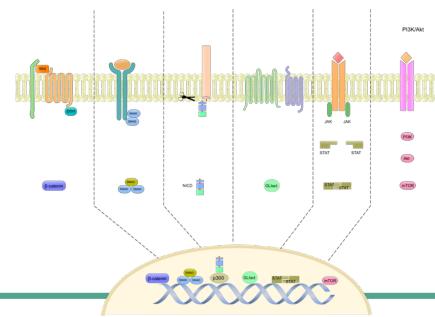
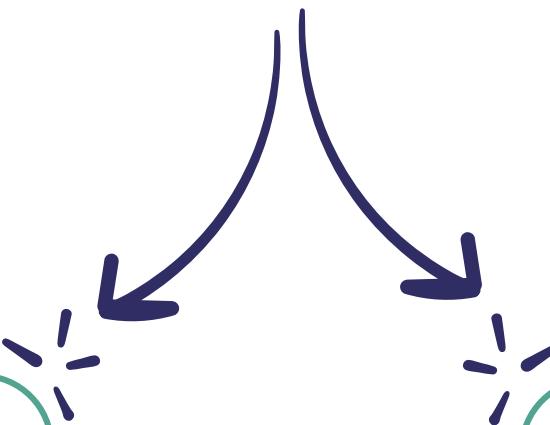
# Objectifs de recherche



Décrire comment des mécanismes biologiques spécifiques contrôlent la croissance et le développement des tissus musculaire et adipeux et contribuent à terme à la construction des qualités de la viande.

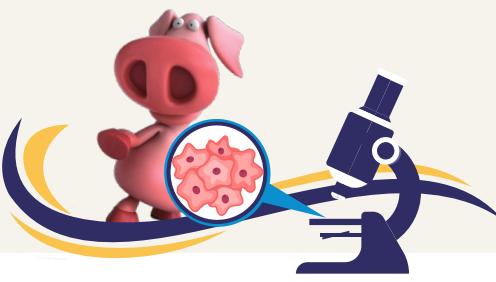
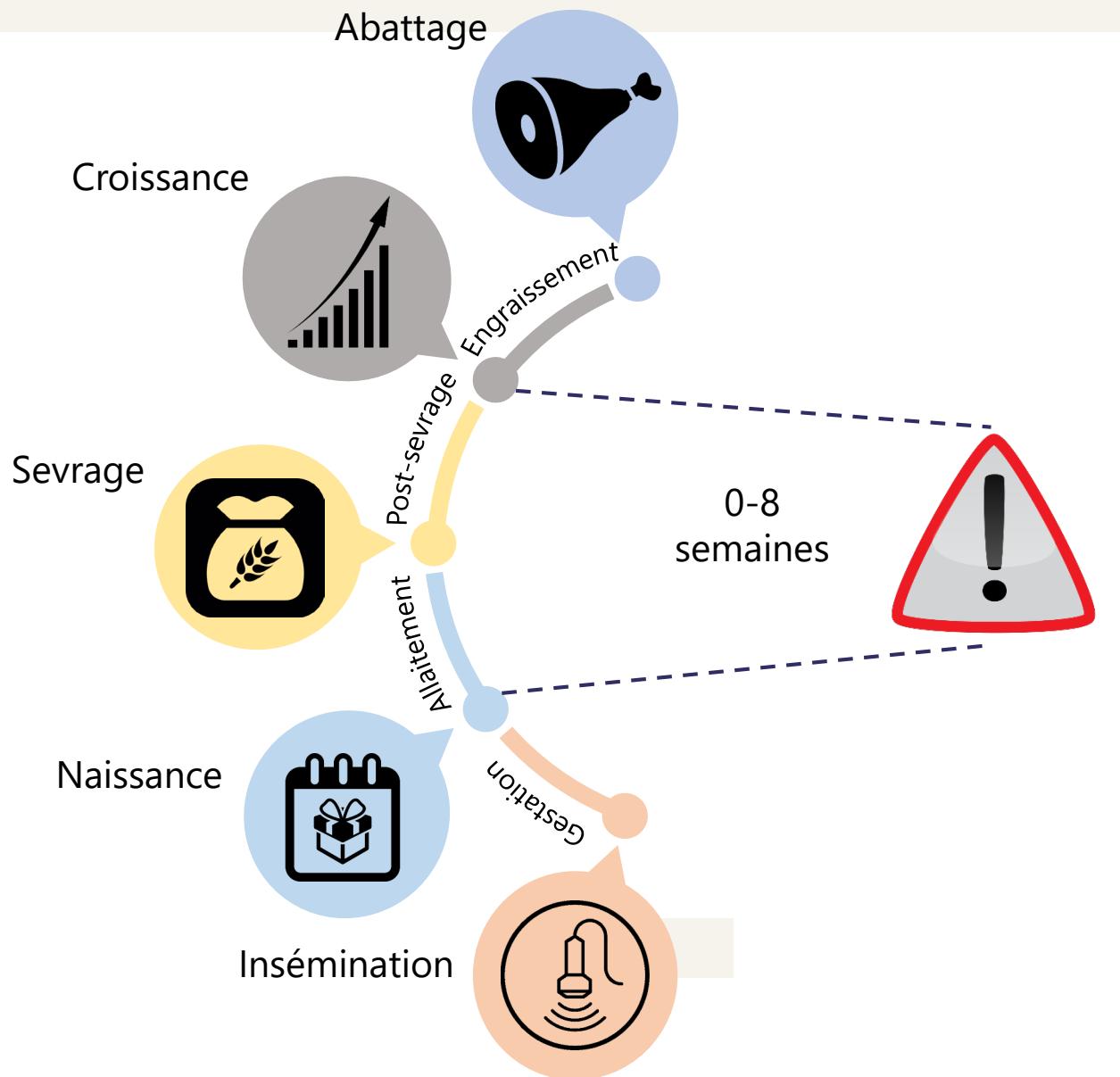


# Exploration de la diversité cellulaire au cours des premières semaines de vie



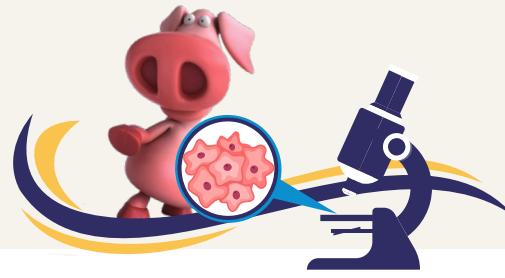
# Dynamique des voies moléculaires et cellulaires contrôlant les dépôts de masse maigre et grasse

# Contexte



**Période critique pour  
l'élevage du porc**

# Le porcelet, un animal fragile



## ALIMENTATION

- phase lactée
- sevrage

## ENVIRONNEMENT

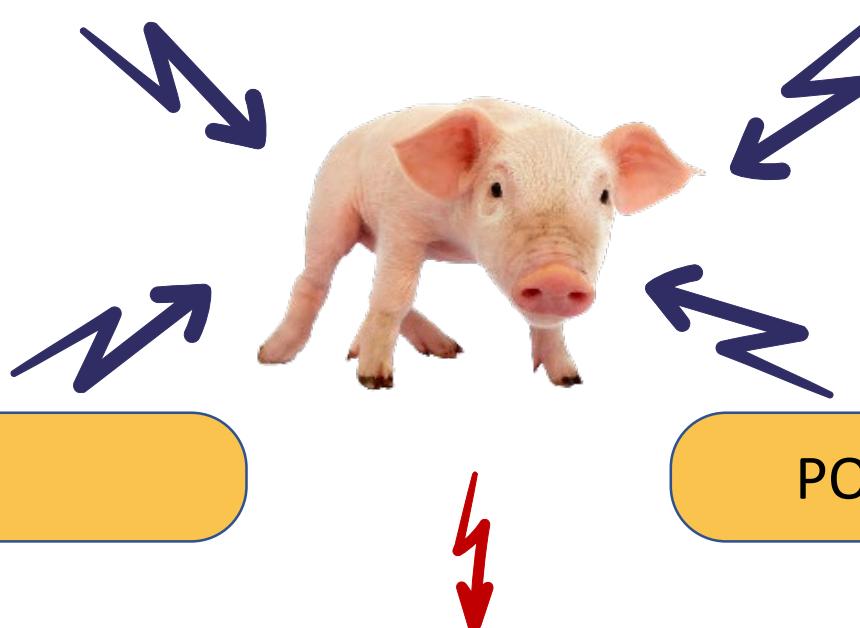
- température
- pathogènes
- exercice physique

## GENETIQUE

- races
- lignées

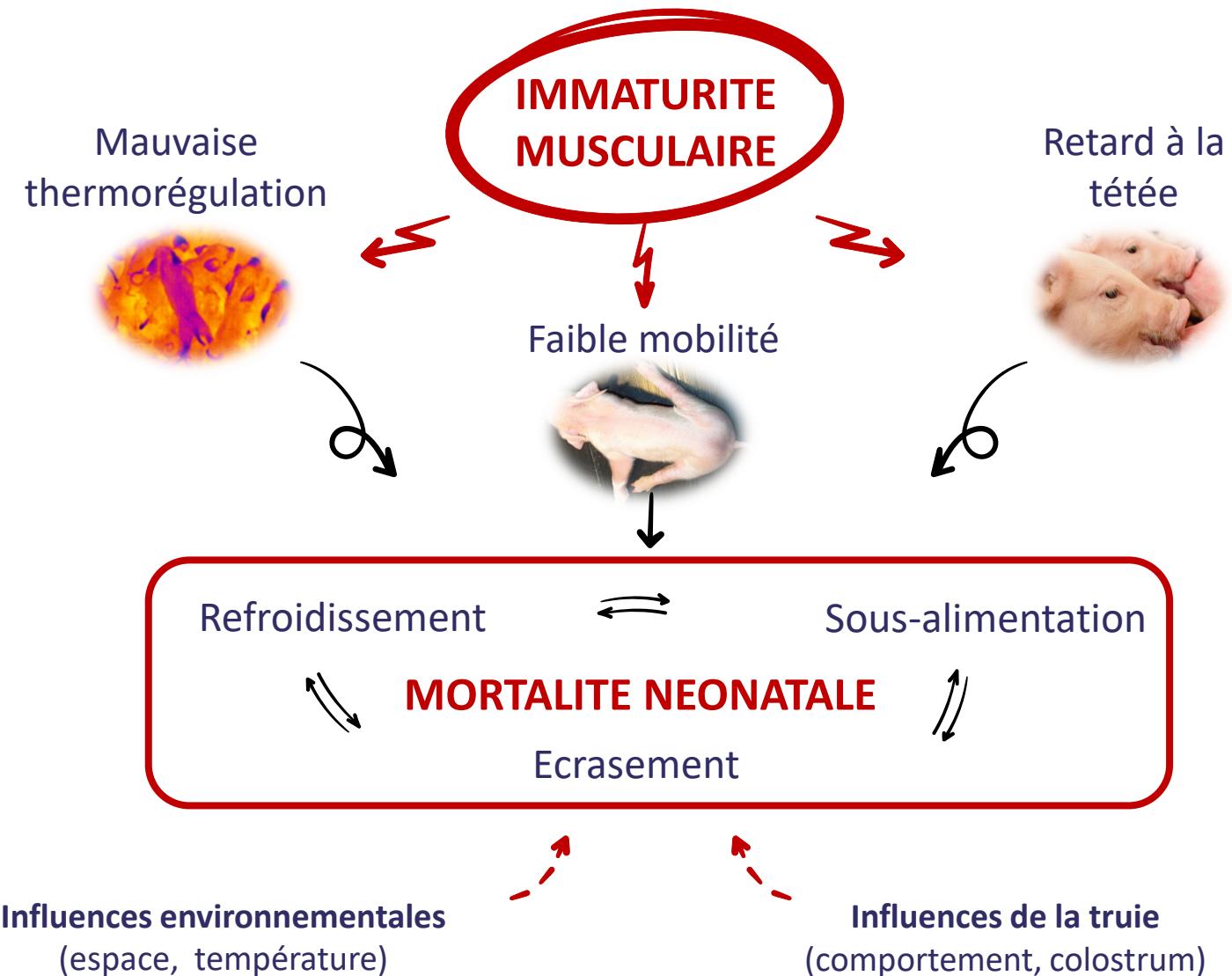
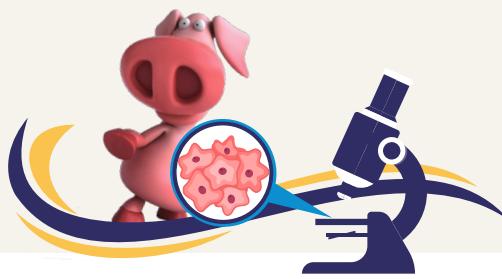
## POIDS VIF A LA NAISSANCE

- variabilité
- (im)maturité
- survie

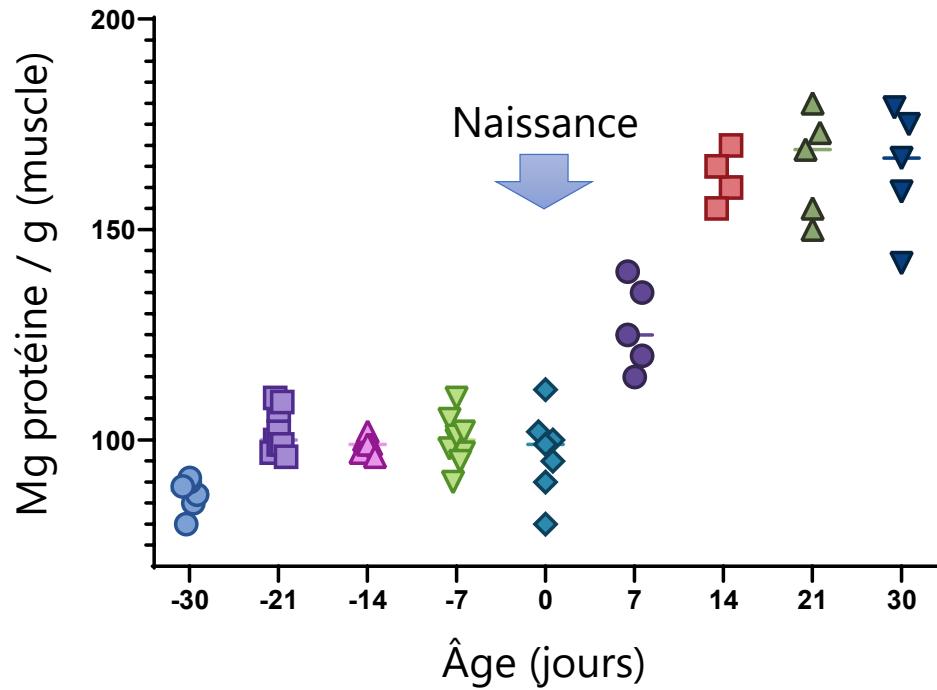
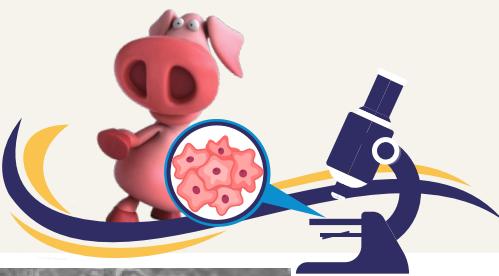


Vitalité néo-natale

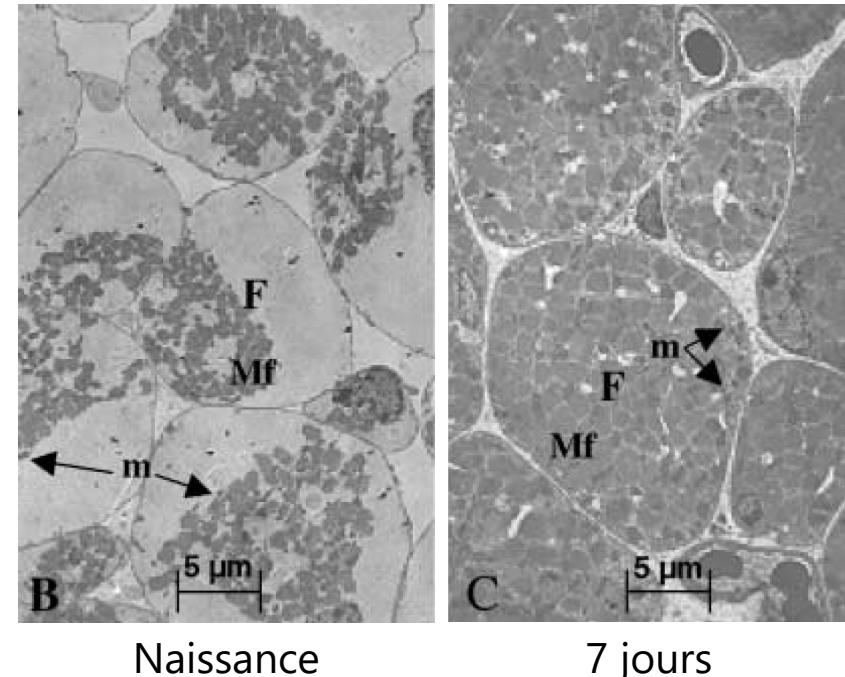
# Fonction musculaire et survie néo-natale



# Croissance et maturation musculaire chez le porc



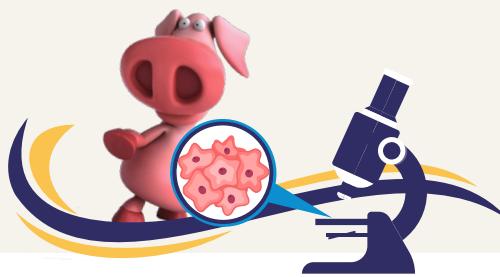
Forte synthèse protéique après la naissance



Forte maturation contractile et Métabolique après la naissance

Maturité musculaire déterminante pour la survie

# Développement précoce du tissu musculaire



1<sup>er</sup> tiers gestation

- 1<sup>ère</sup> génération de fibres
- Hyperplasie
- Différenciation

2<sup>ème</sup> tiers gestation

- 2<sup>ème</sup> génération de fibres
- Hyperplasie
- Différenciation

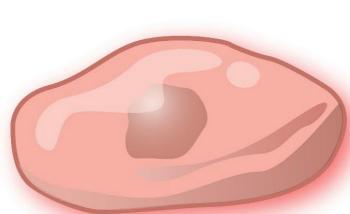
3<sup>ème</sup> tiers gestation

- Nombre total de fibres fixé
- Hypertrophie

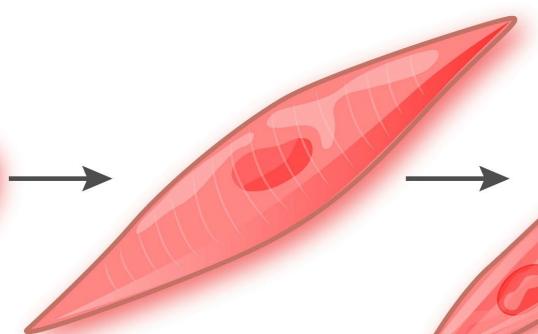


Naissance

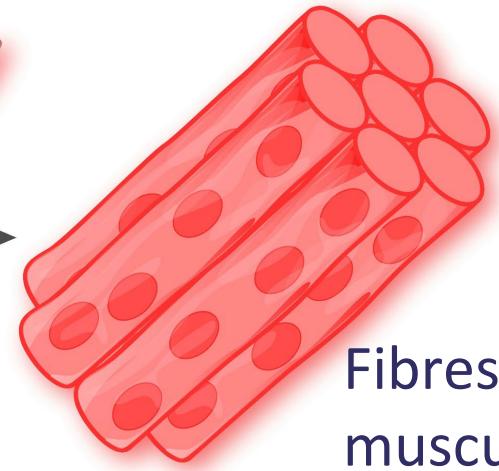
- 3<sup>ème</sup> génération de fibres
- Hypertrophie



Cellule souche

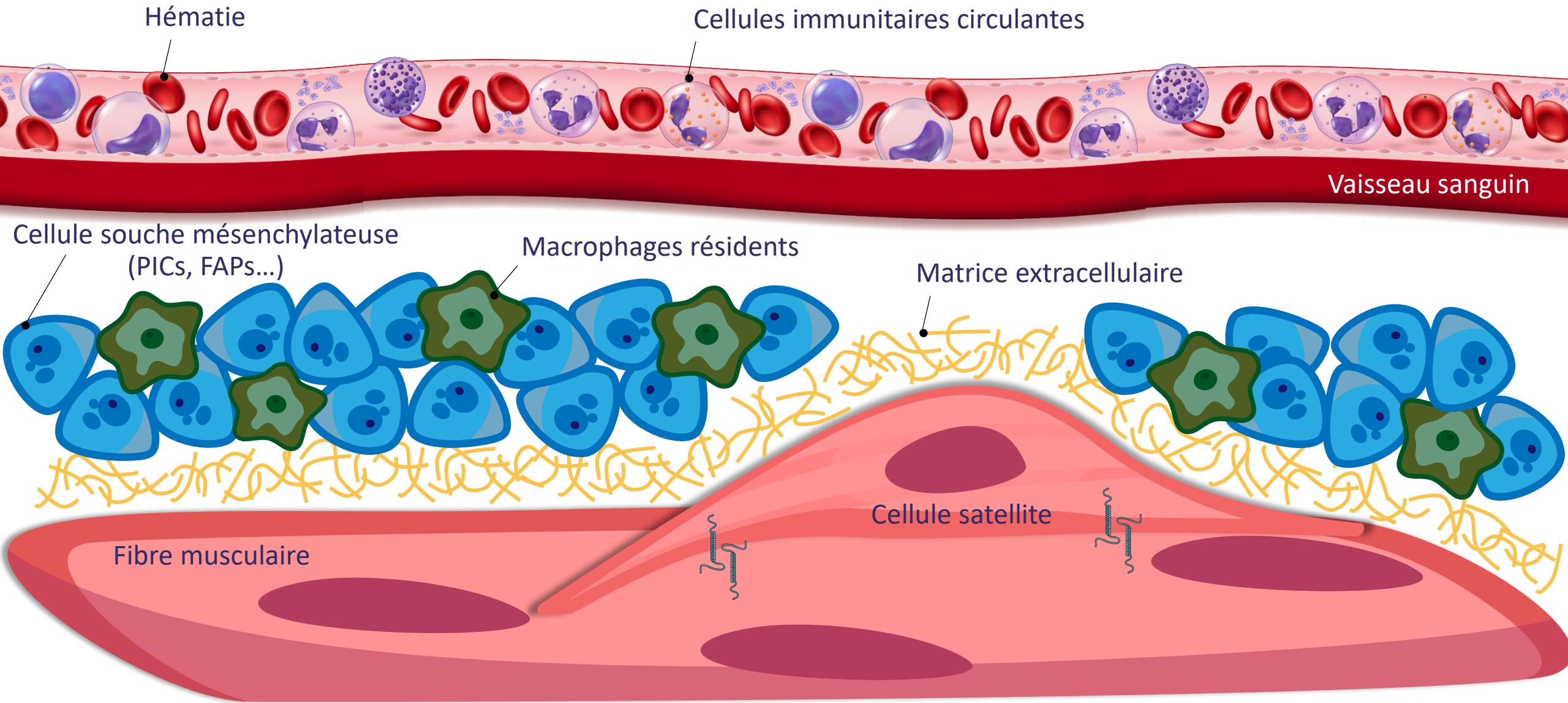
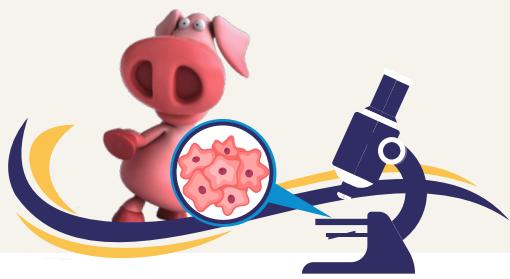


Myotube

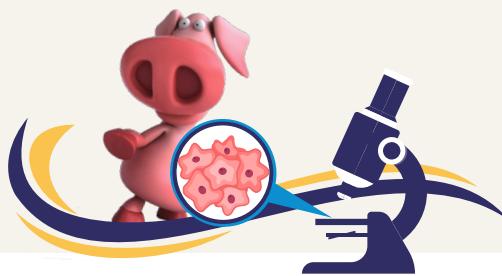


Fibres musculaires

# Une diversité de cellules souches dans le muscle



# Des cellules souches, des marqueurs de surface

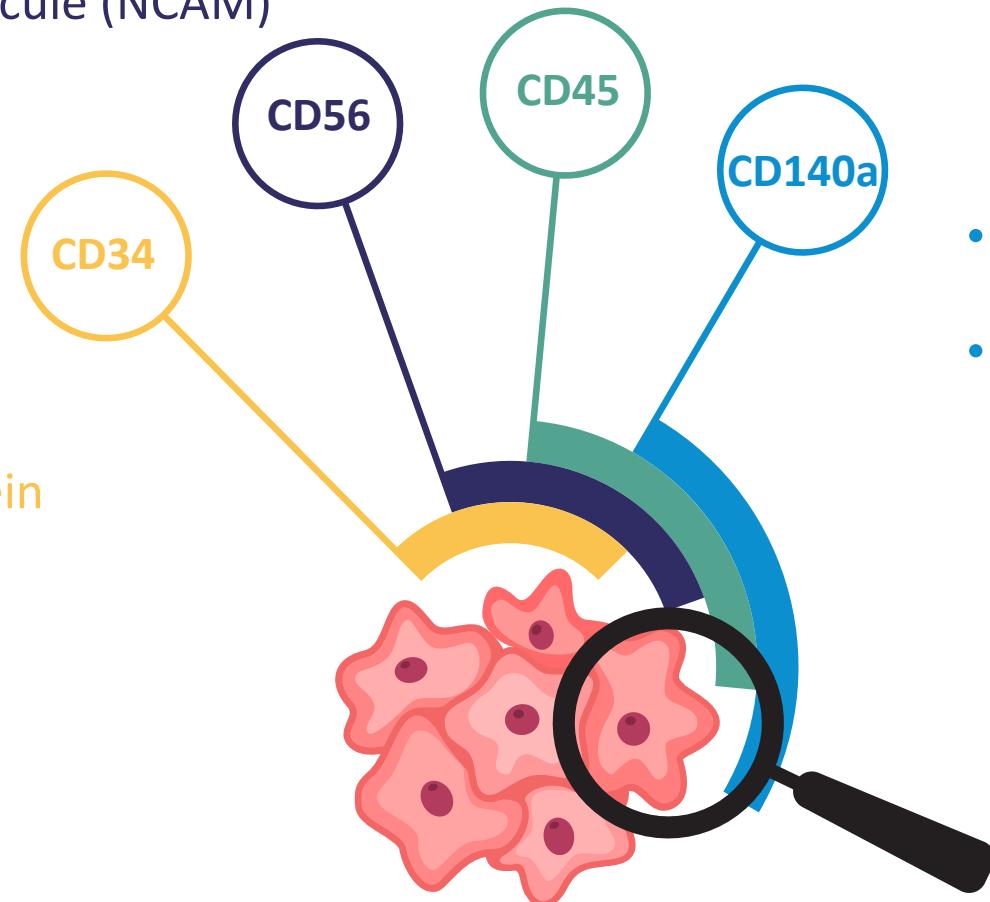


- Marqueur myogénique
- Neural Cell Adhesion Molecule (NCAM)

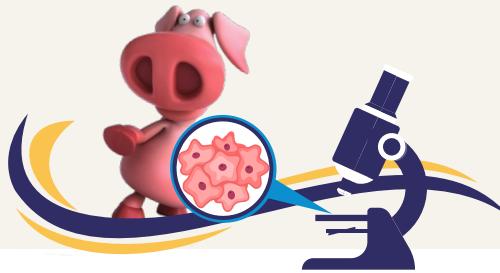
- Marqueur cellules souches hématopoïétiques
- Phospho-Glycoprotein Protein

- Marqueur cellules hématopoïétiques
- Protein Tyrosine Phosphatase

- Marqueur cellules fibro-adipogéniques
- Platelet-Derived Growth Factor Recept A



# Des marqueurs de surface qui évoluent chez l'adulte



Régime différencié (fibres, gras)

→ Perruchot *et al.*, 2021



Supplémentation acides aminés

→ Castellano *et al.*, 2017



Statut sanitaire

→ Quemener *et al.*, 2022



Statut Redox

→ Perruchot *et al.*, 2019



Diminution du réservoir de cellules souches avec l'âge

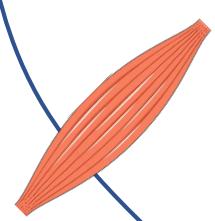


Nécessité de travailler dans les premiers jours de vie

# Stratégie expérimentale



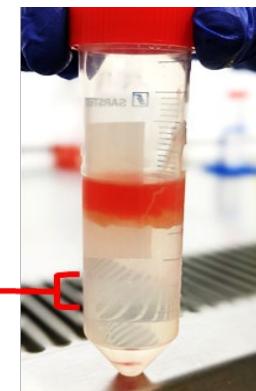
Porcelet J7 (3-3,2kg)



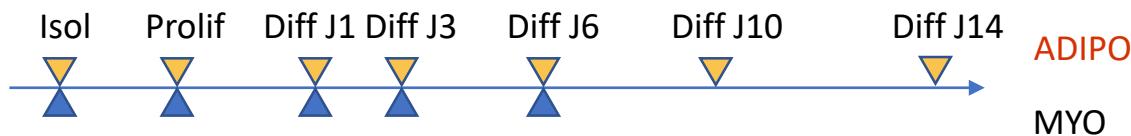
Isolement des cellules musculaires à partir du *longissimus dorsi*



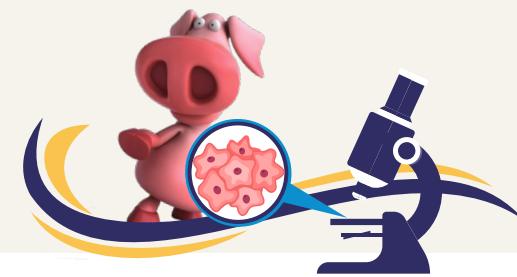
Anneau de cellules satellites



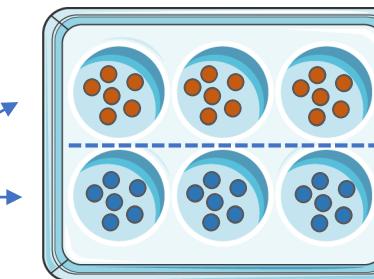
$0,5 \times 10^5$  cell / g



Phénotypage par cytométrie en flux



Analyse de gènes par qPCR

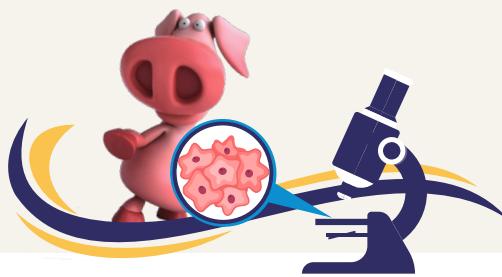


Prolifération puis différenciation en condition myogénique ou adipogénique

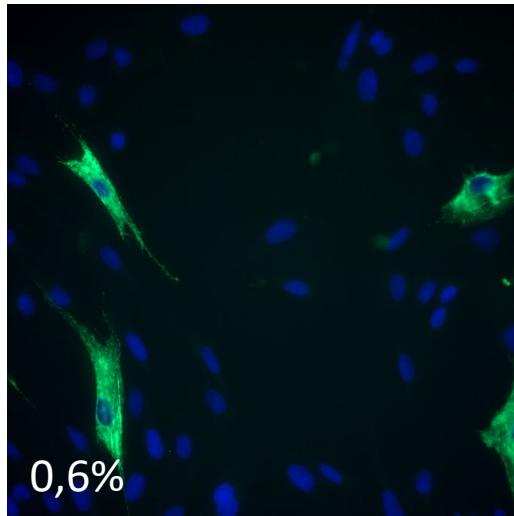


Mesure fusion par microscopie

# Cellules satellites en condition myogénique

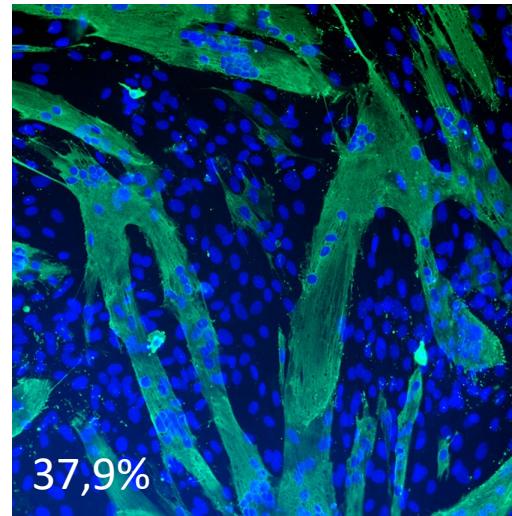


Prolif

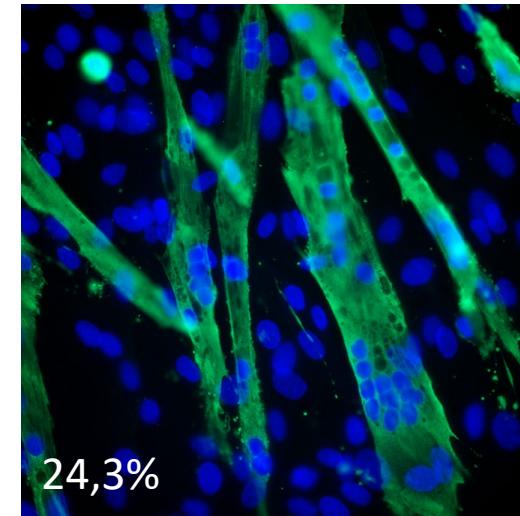


Marquage  
MF20

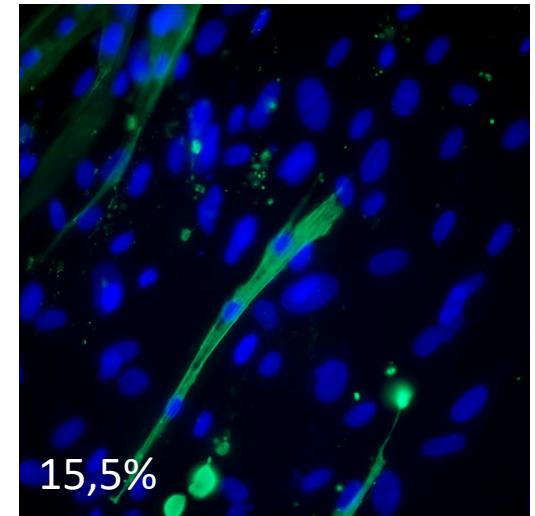
Diff J1



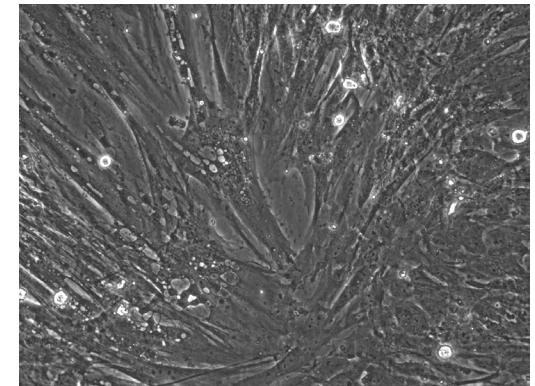
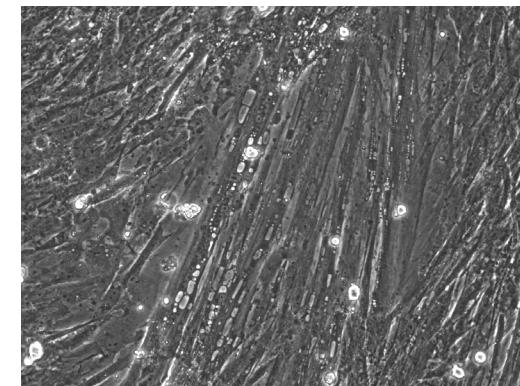
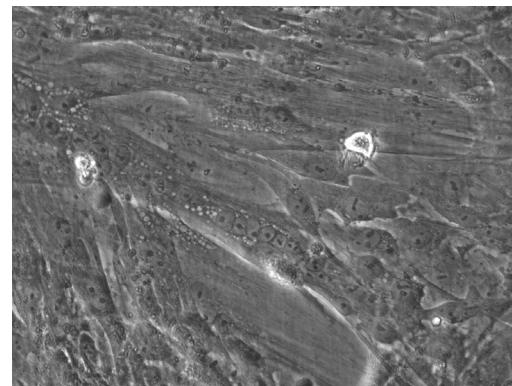
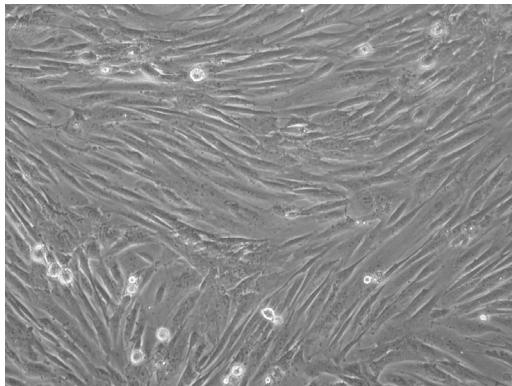
Diff J3



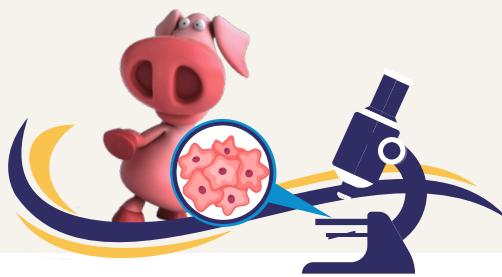
Diff J6



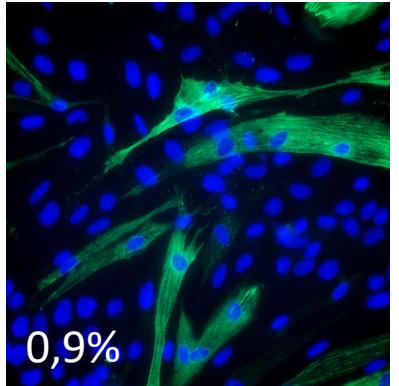
Contraste  
de phase



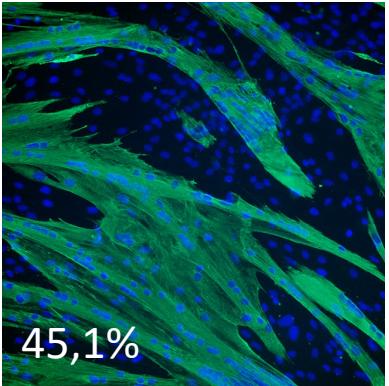
# Cellules satellites en condition adipogénique



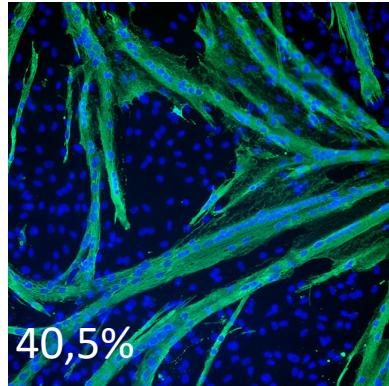
Prolif



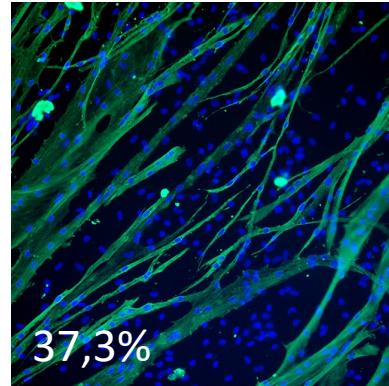
Diff J1



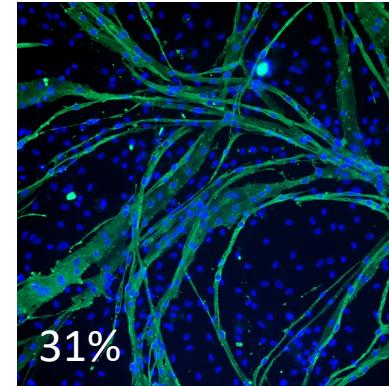
Diff J3



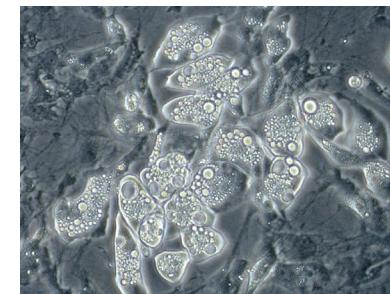
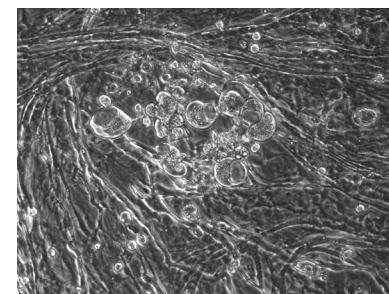
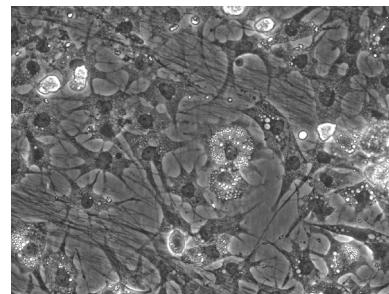
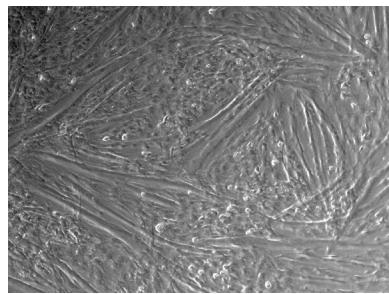
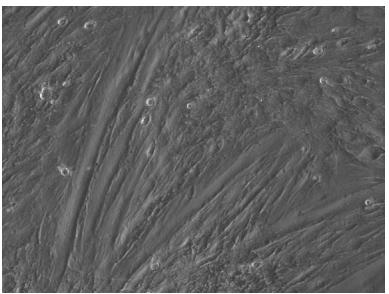
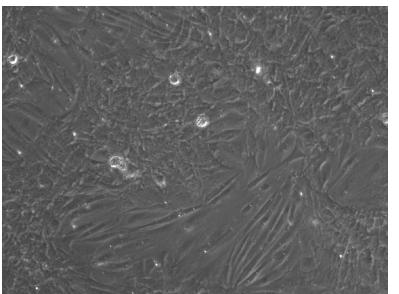
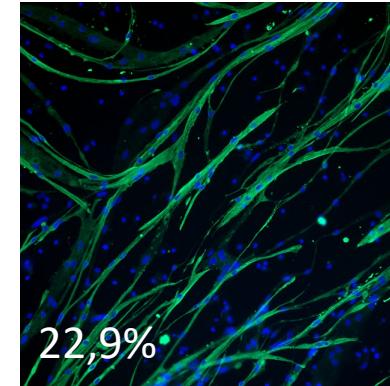
Diff J6



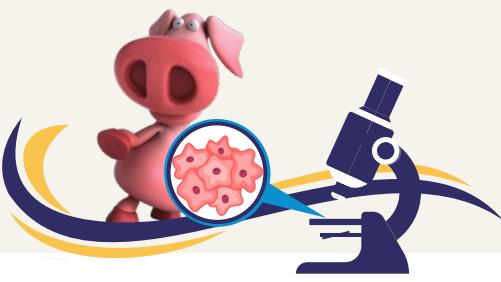
Diff J10



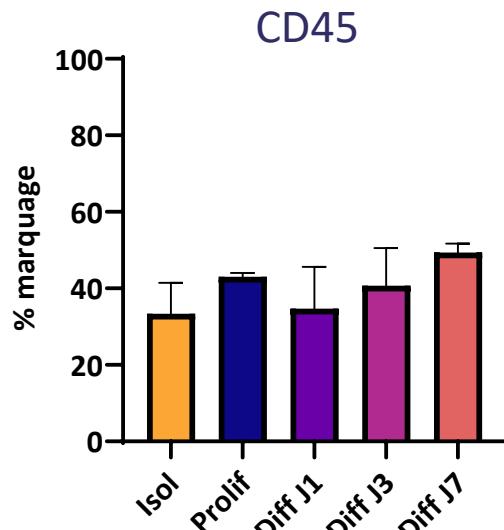
Diff J14



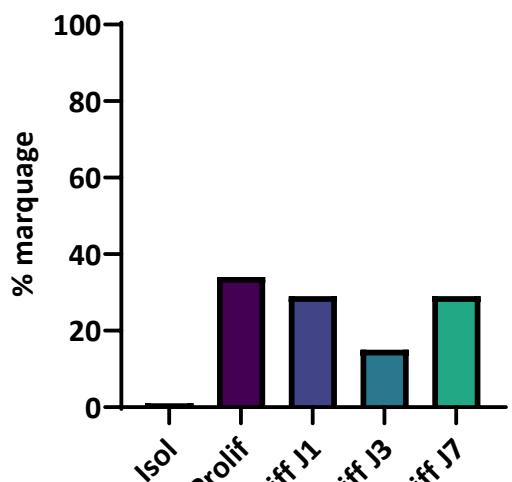
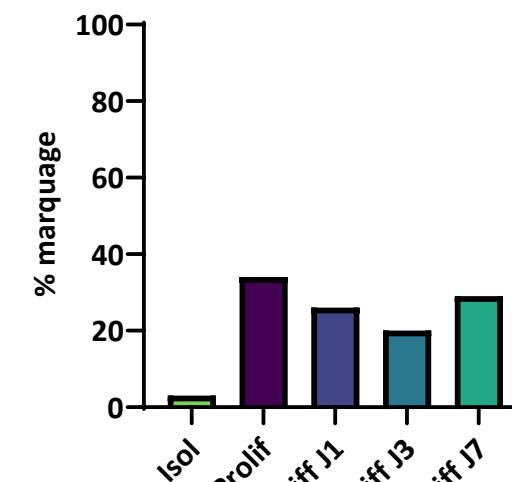
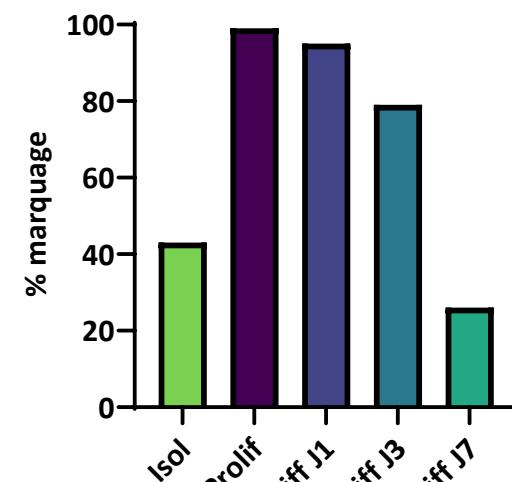
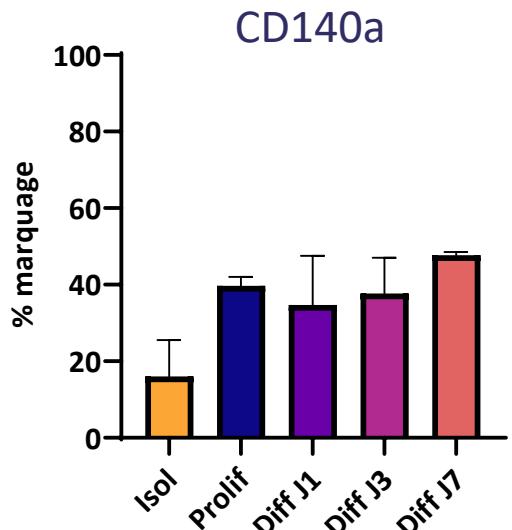
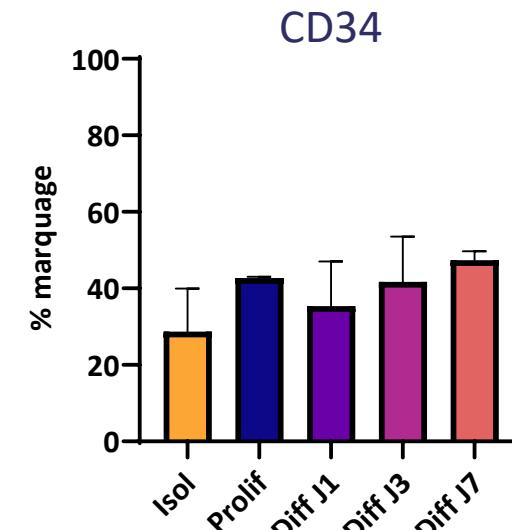
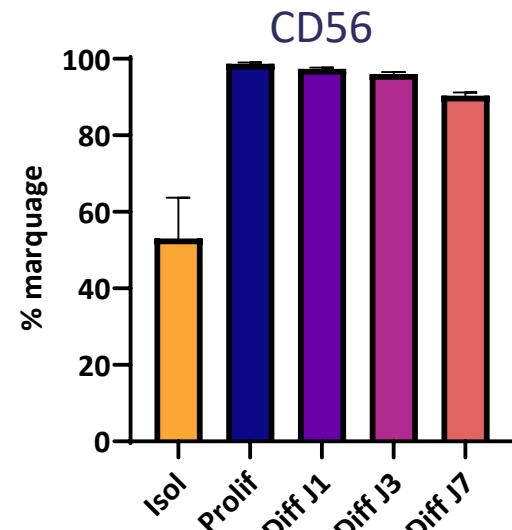
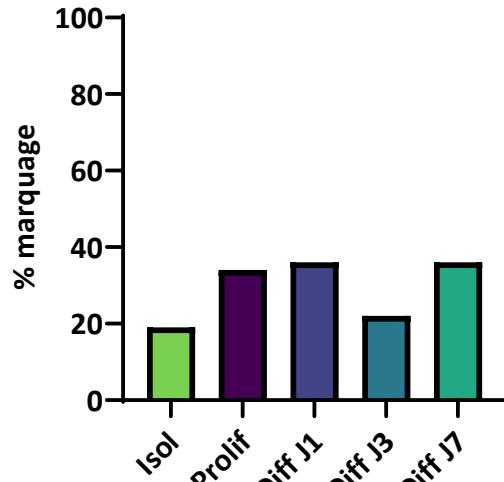
# Evolution des marqueurs de surface (cytométrie)



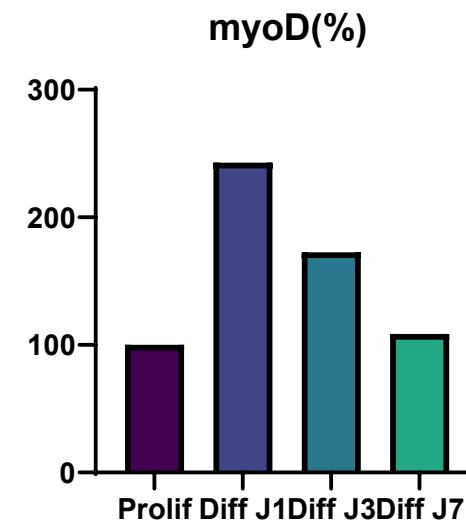
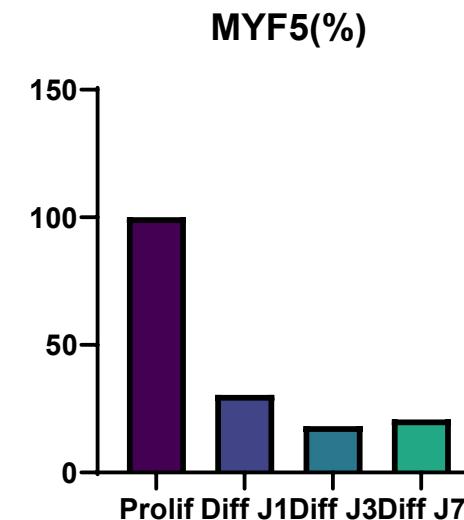
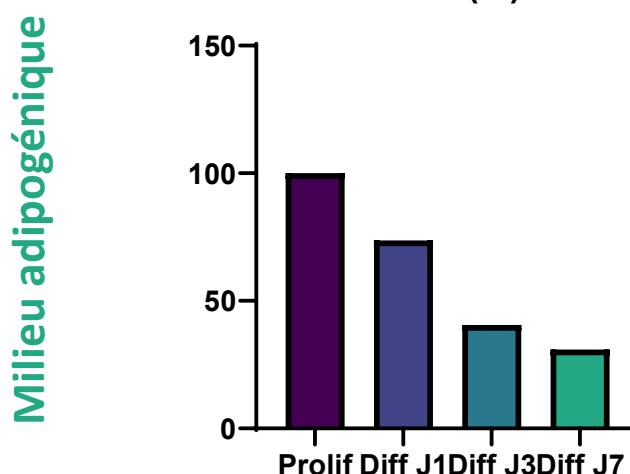
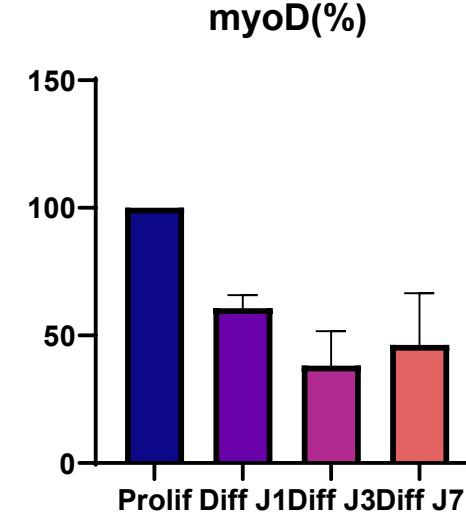
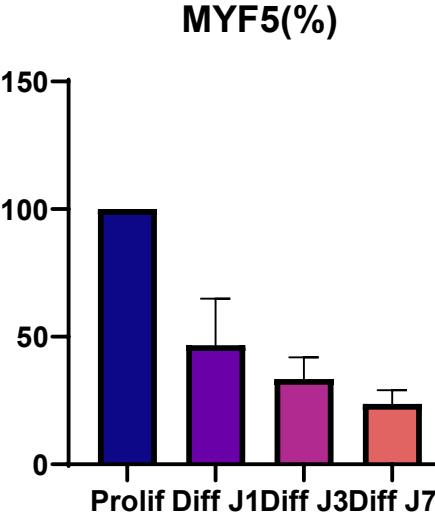
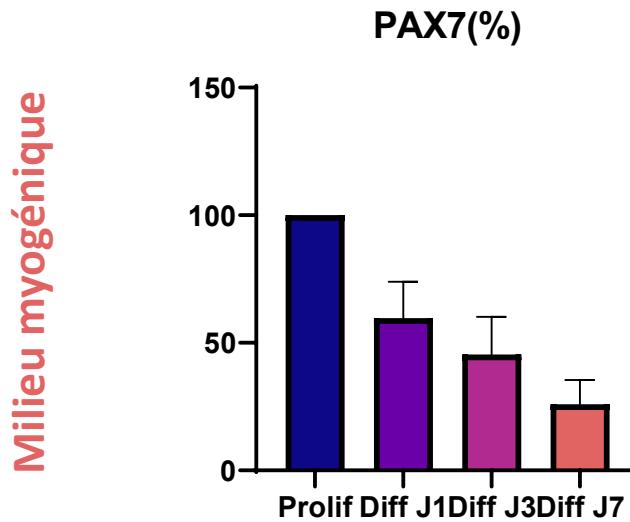
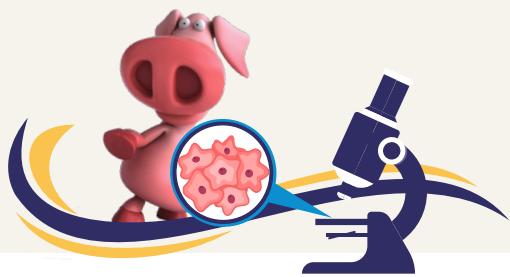
Milieu myogénique



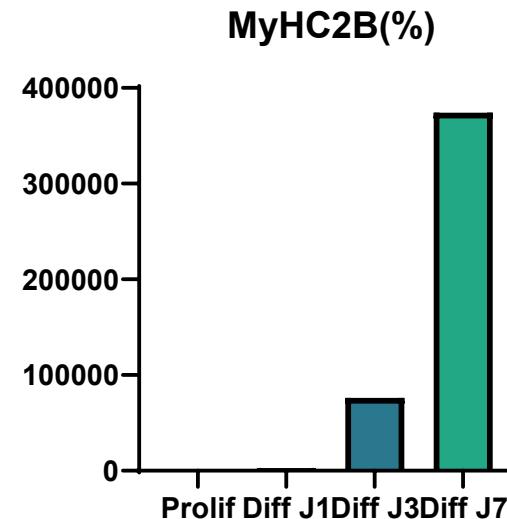
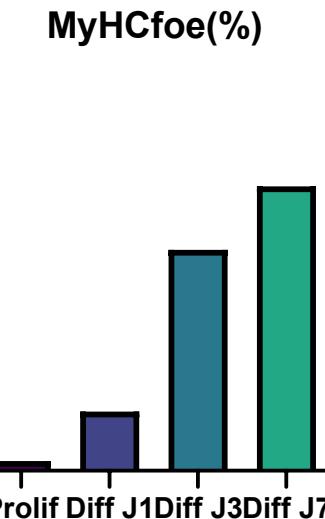
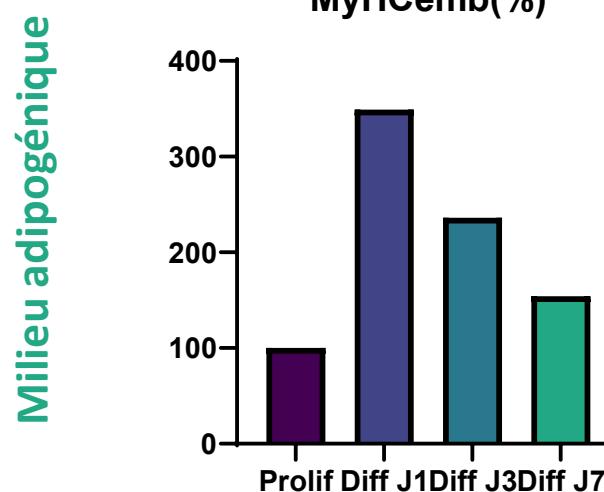
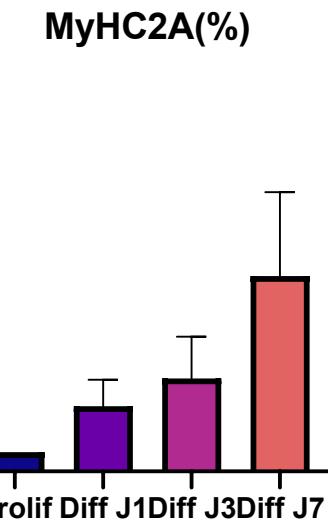
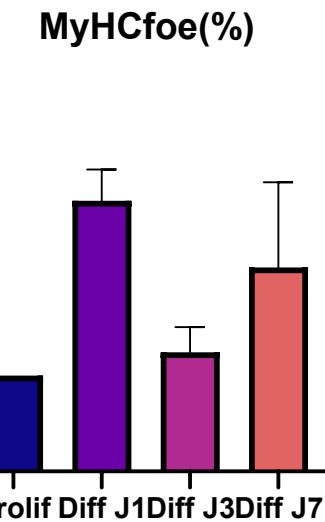
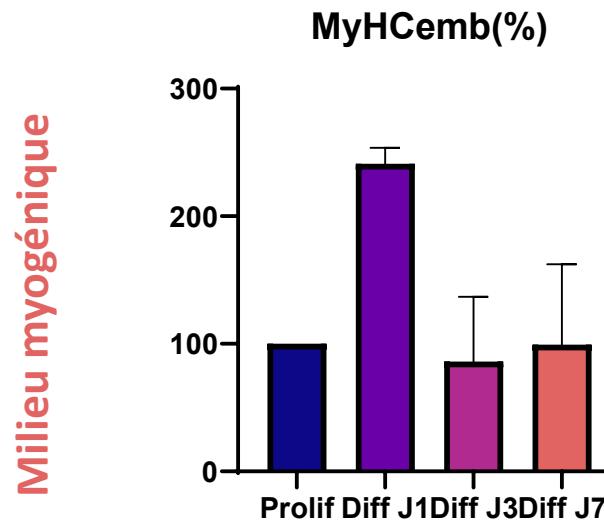
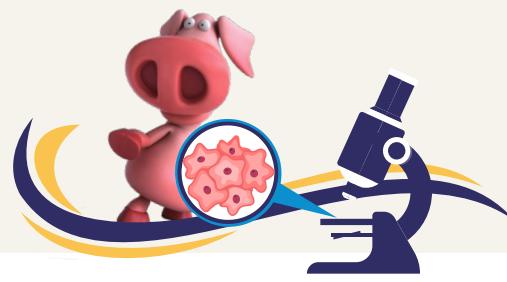
Milieu adipogénique



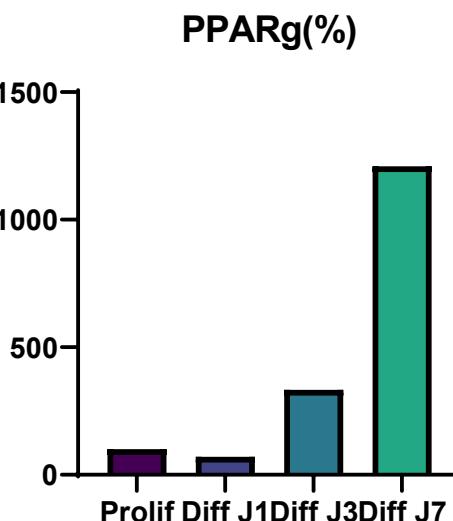
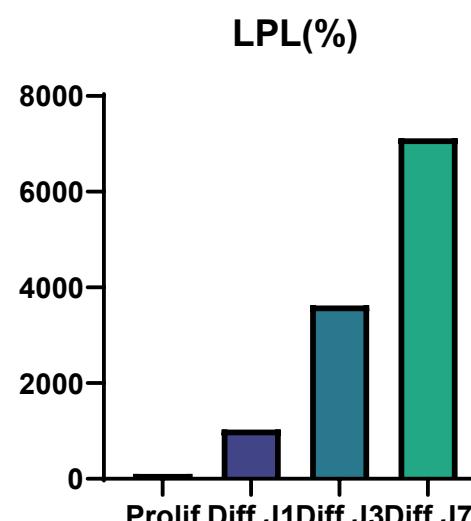
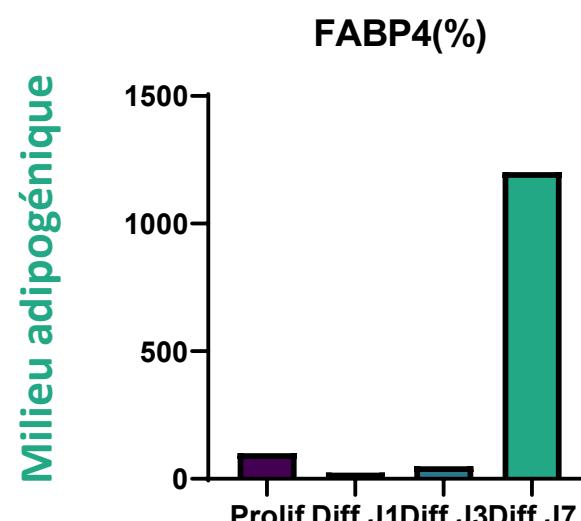
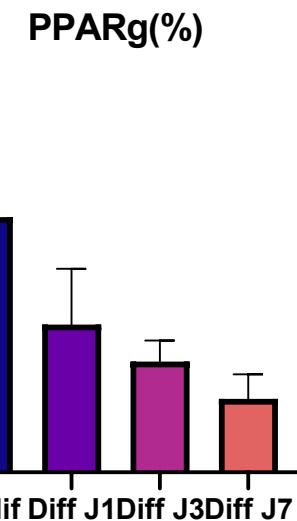
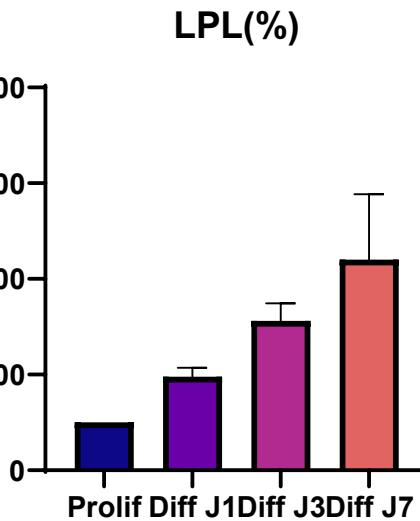
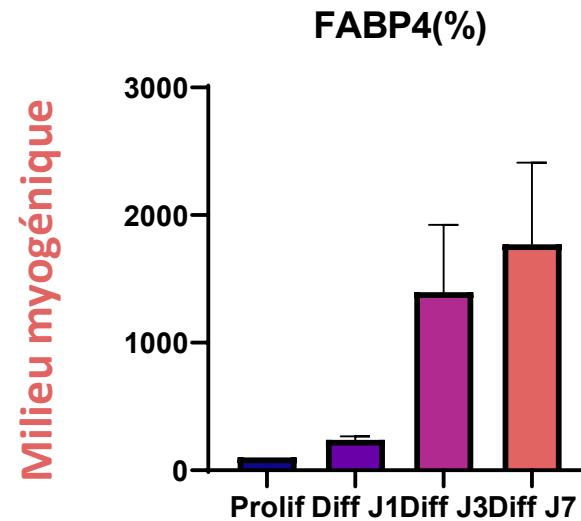
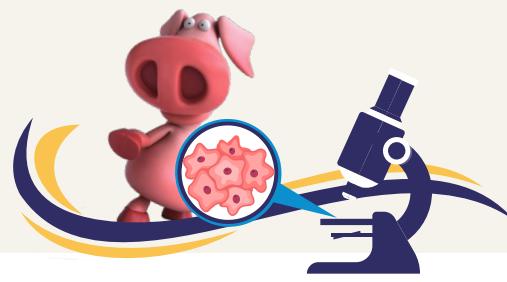
# Expression des marqueurs myogéniques (qPCR)



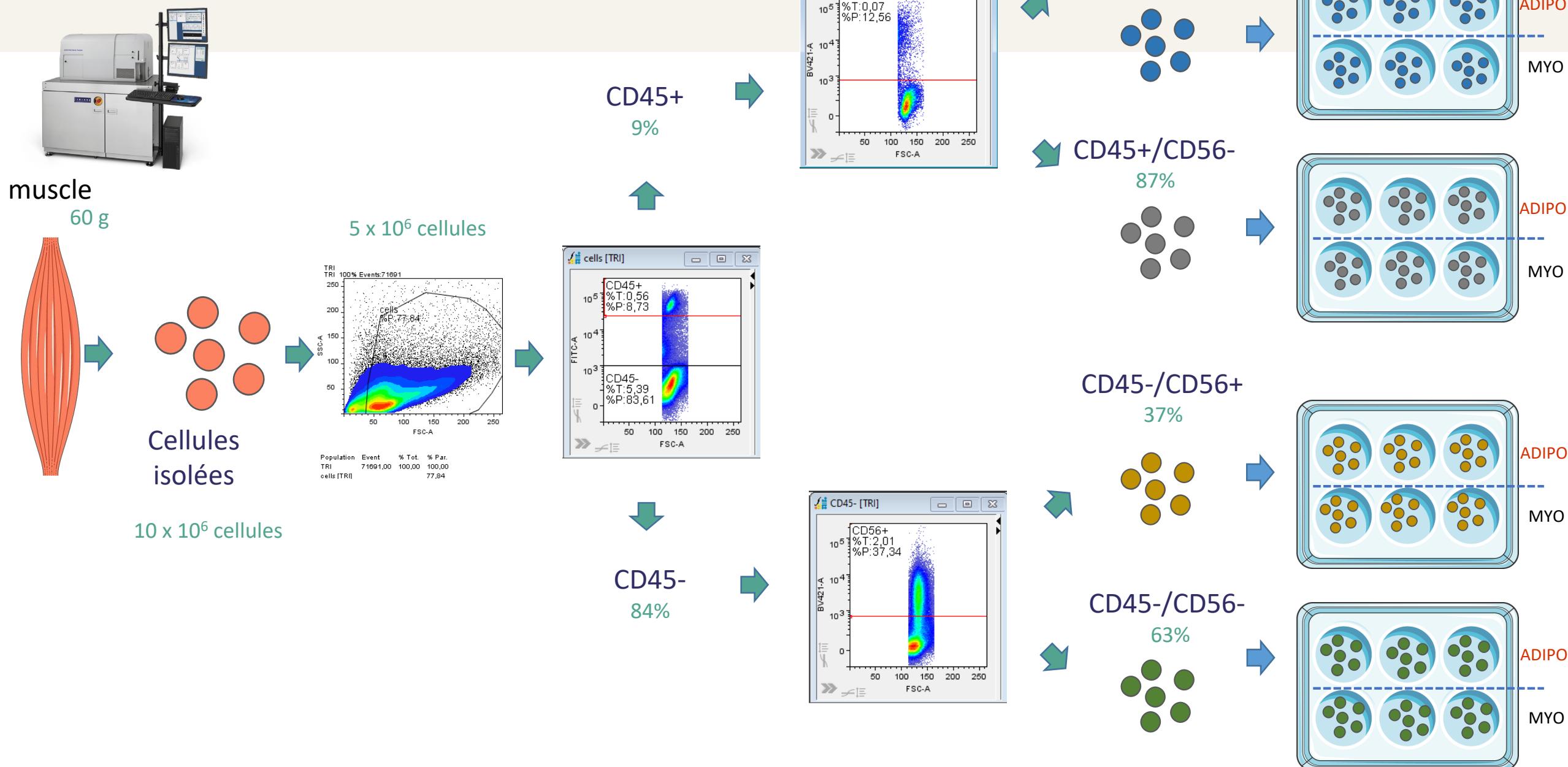
# Expression des marqueurs myogéniques (qPCR)



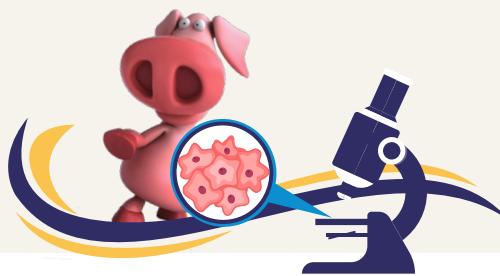
# Expression des marqueurs adipogéniques (qPCR)



# Stratégie de tri

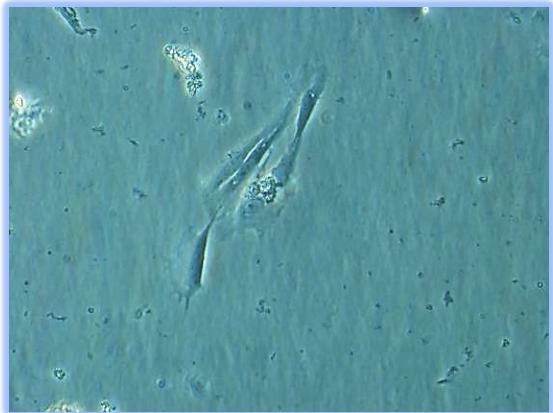


# Culture des populations triées



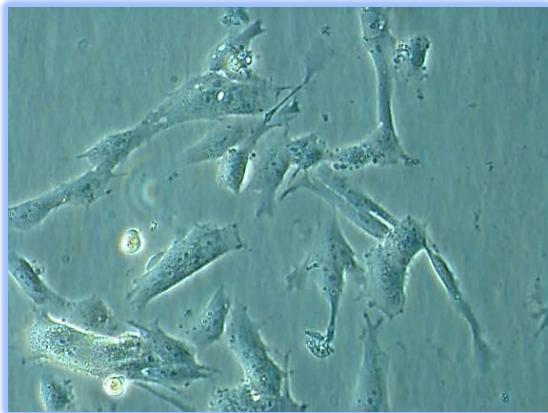
Milieu myogénique

CD45-/CD56-



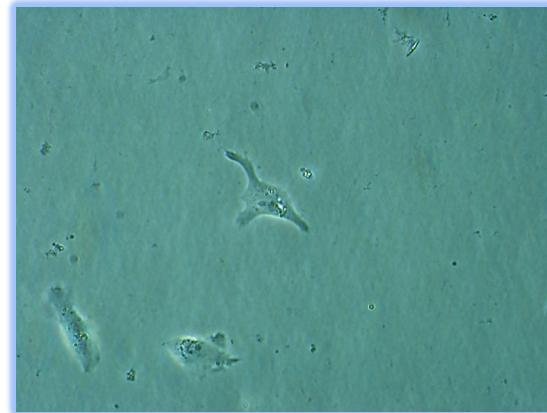
Prolif + dans les 2 conditions

CD45-/CD56+



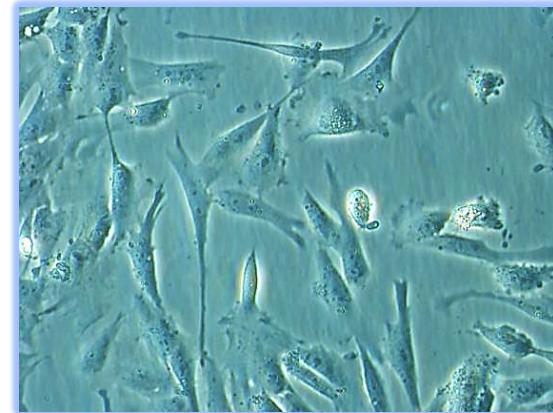
Prolif ++ dans les 2 conditions

CD45+/CD56-



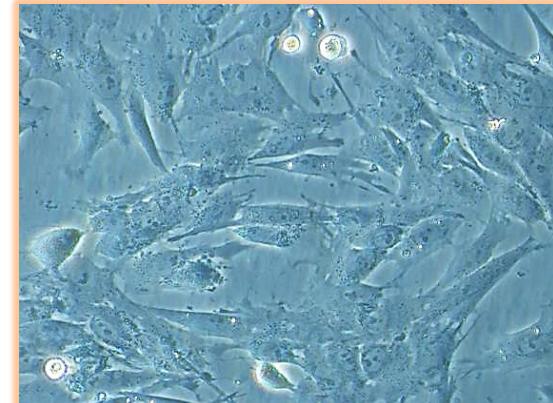
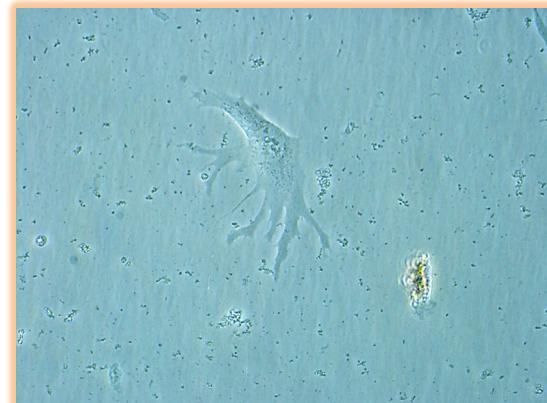
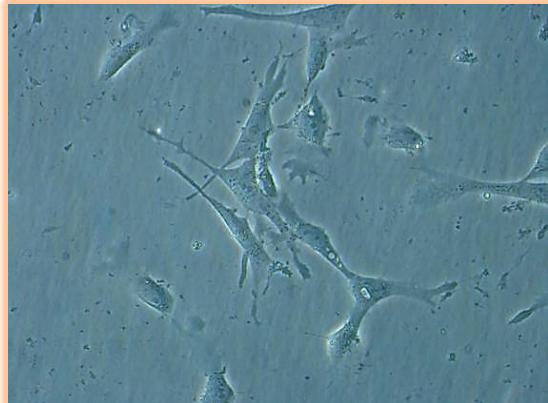
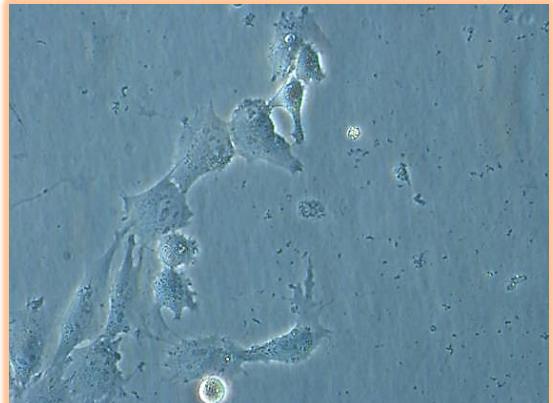
Peu de prolif et cellules dendritiques

CD45+/CD56+

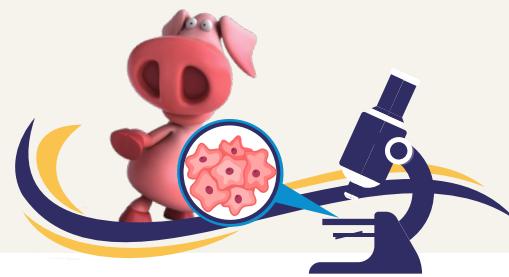


Prolif +++ dans les 2 conditions

Milieu adipogénique

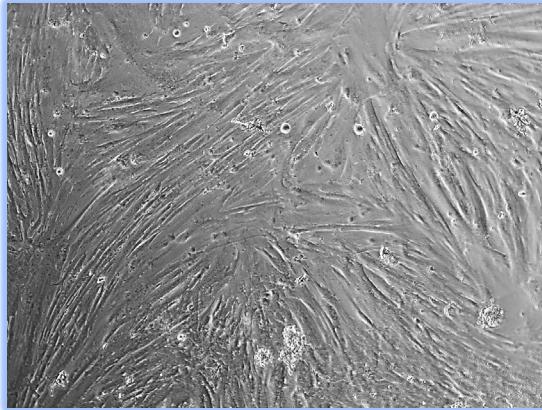


# Culture des populations triées (CD45<sup>-</sup>)

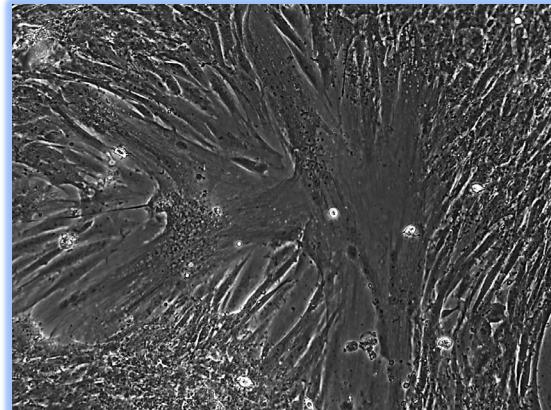


Milieu myogénique

CD45-/CD56-

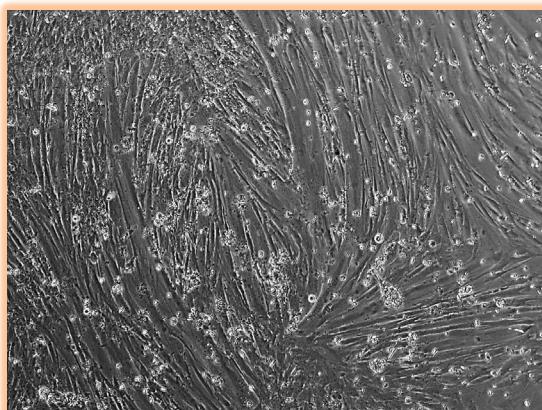


CD45-/CD56+

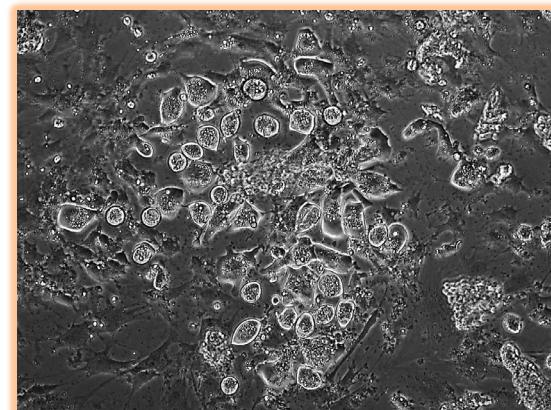


Milieu adipogénique

DIFF + dans les 2  
conditions



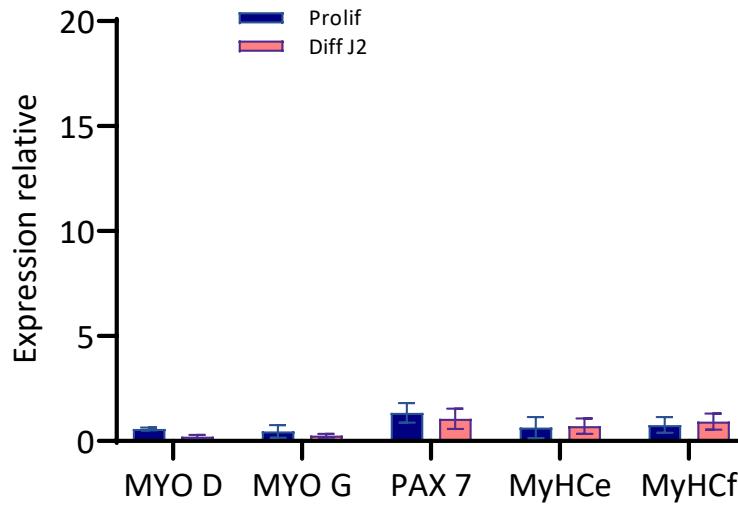
DIFF ++ dans les 2  
conditions



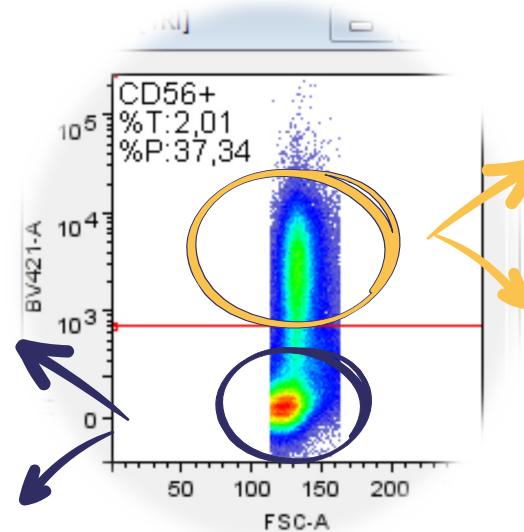
# Expression des marqueurs myogéniques après tri



Milieu myogénique

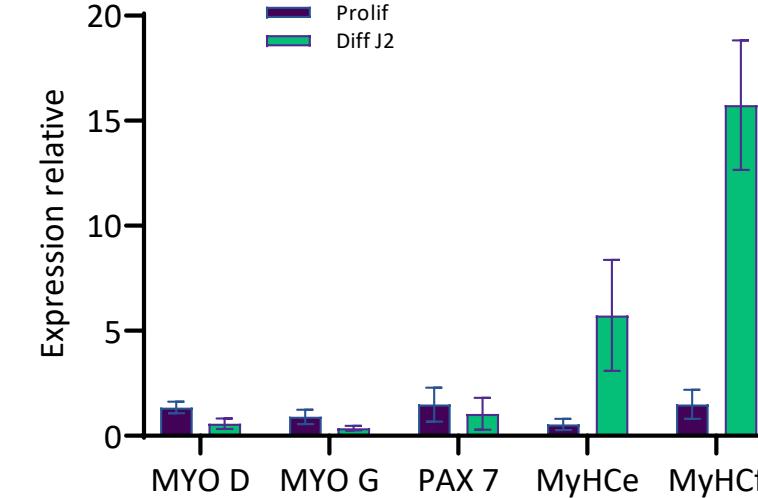
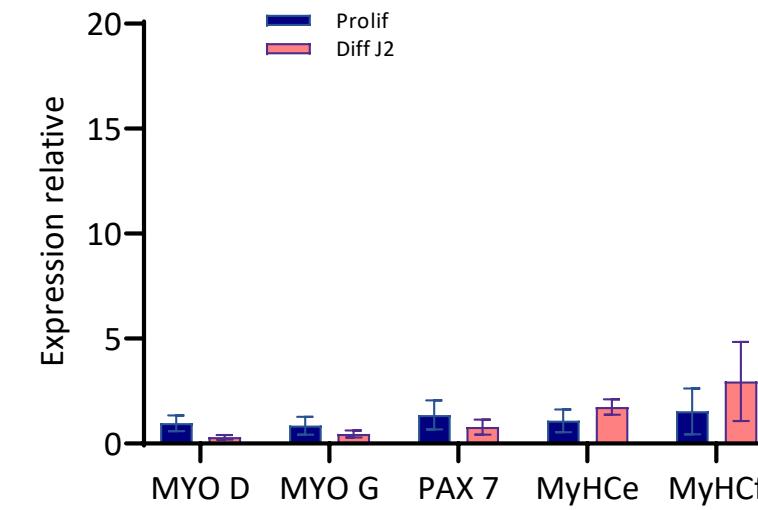
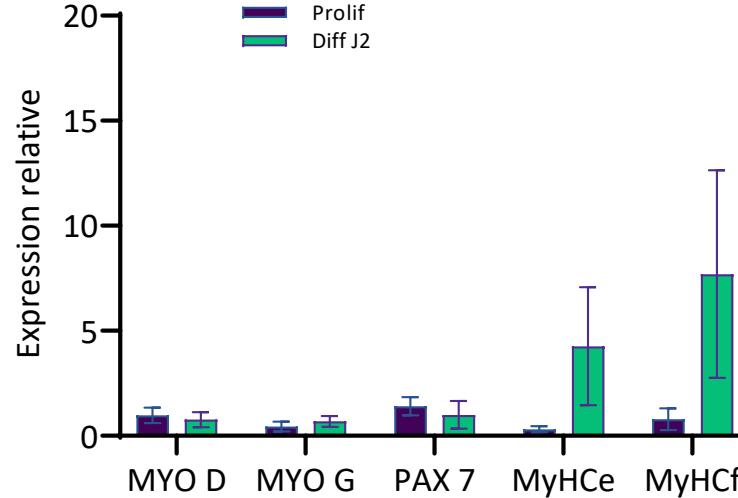


CD45-/CD56+

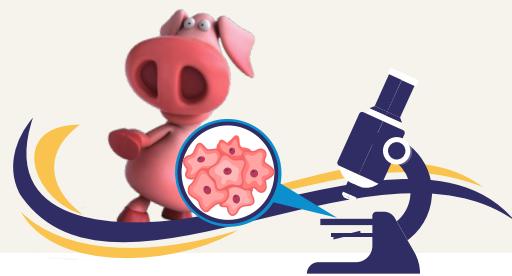


CD45-/CD56-

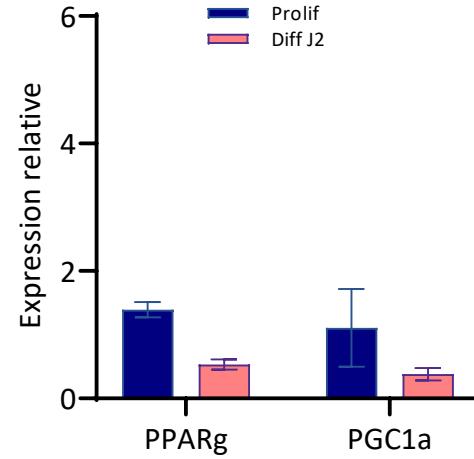
Milieu adipogénique



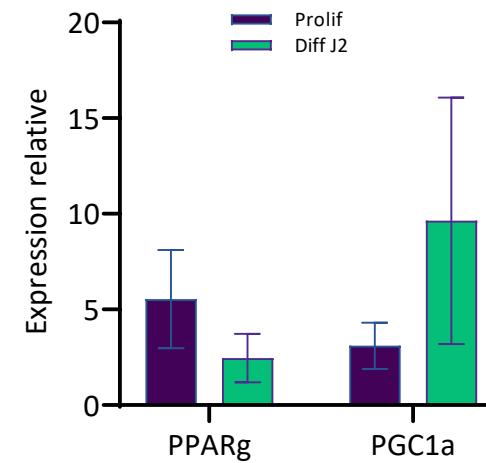
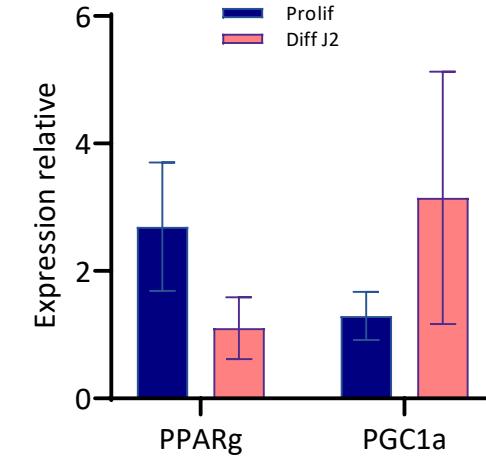
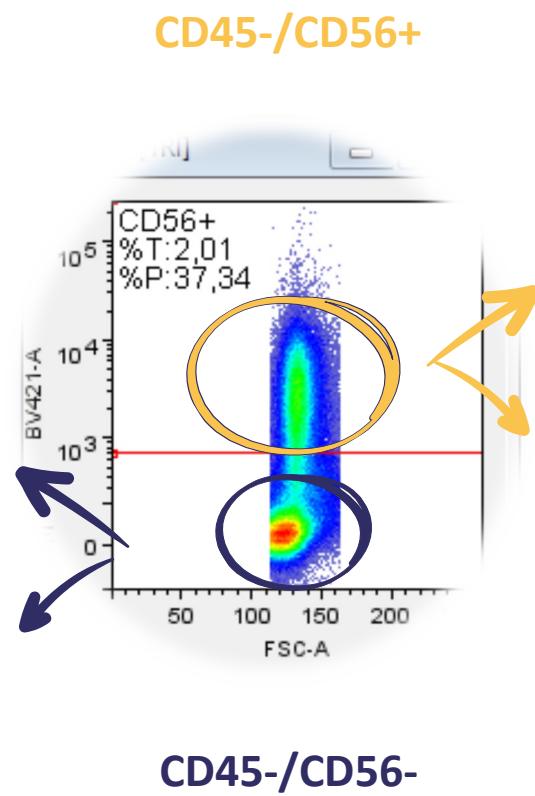
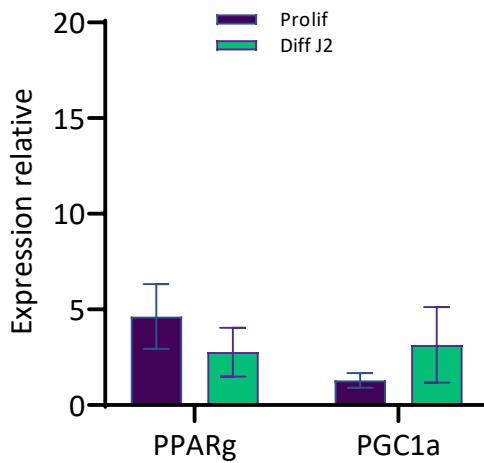
# Expression des marqueurs adipogéniques après tri



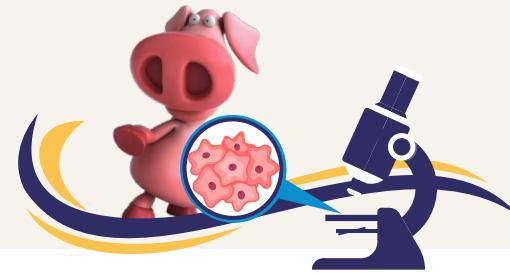
Milieu myogénique



Milieu adipogénique



# Conclusions



- ! Capacité à faire différencier les cellules souches en fibres musculaires et en adipocytes fonctionnels
- ! Expression des marqueurs de surface altérée par la culture primaire
- ! Tri cellulaire = méthodologie complexe pour caractériser les différentes populations cellulaires du muscle
- ! Relevance du marqueur myogénique CD56
- ! Le milieu adipogénique: un milieu prometteur (expression de myosines adultes !)