

# Incorporating biological information into genomic prediction models

Andrea Rau, Fanny Mollandin, Pascal Croiseau

#### ▶ To cite this version:

Andrea Rau, Fanny Mollandin, Pascal Croiseau. Incorporating biological information into genomic prediction models. VistaMilk Artificial Intelligence in Agriculture Masterclass, Feb 2023, Online, Ireland. hal-04173250

HAL Id: hal-04173250 https://hal.inrae.fr/hal-04173250

Submitted on 28 Feb 2024

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

## Incorporating biological information into genomic prediction models

Fanny Mollandin, Pascal Croiseau, Andrea Rau

VistaMilk Artificial Intelligence in Agriculture Masterclass @ Zoom February 8, 2023











#### INRAE Research Center @ Jouy en Josas:

- ✓ 1500+ staff
- ✓ Animal biology, microbiology, data science, systems biology

#### Animal Genetics & Integrative Biology (GABI) unit

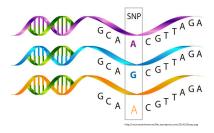
- ✓ Understanding & exploiting animal genetic variability
- Construction of phenotypes and their interaction with microbial ecosystems and environments
- ✓ Agroecological transition





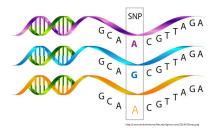
Objective: select the best animals for reproduction to obtain **genetic improvement** of the population on **traits of interest** 

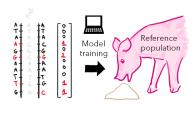
Objective: select the best animals for reproduction to obtain **genetic improvement** of the population on **traits of interest** 



- Low- to high-density genotyping chips (10k-100k SNPs)
  - $\rightarrow$  whole genome sequencing (10MM SNPs)

Objective: select the best animals for reproduction to obtain **genetic improvement** of the population on **traits of interest** 

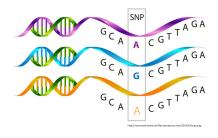




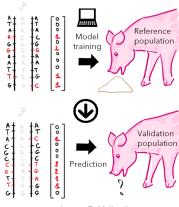
- Low- to high-density genotyping chips (10k-100k SNPs)
  - $\rightarrow$  whole genome sequencing (10MM SNPs)

Image: F. Mollandin

Objective: select the best animals for reproduction to obtain **genetic improvement** of the population on **traits of interest** 



 Low- to high-density genotyping chips (10k-100k SNPs)
 → whole genome sequencing (10MM SNPs)



#### Prediction models for genomic selection

Goal: given a **training set** of data  $(Y_i, X_i, Z_i)$  for i = 1, ..., n individuals

- $\bullet$   $Y_i = \text{trait}$
- $X_i$  = vector of (usually genome-wide) genotypes
- $Z_i$  = vector of covariates (age, location, sex, ...)

... predict the unobserved trait  $Y_{\star}$  of a future individual with corresponding  $X_{\star}$  and  $Z_{\star}$ 

#### Prediction models for genomic selection

Goal: given a training set of data  $(Y_i, X_i, Z_i)$  for i = 1, ..., n individuals

- $\bullet$   $Y_i = \text{trait}$
- $X_i$  = vector of (usually genome-wide) genotypes
- $Z_i$  = vector of covariates (age, location, sex, ...)

... predict the unobserved trait  $Y_{\star}$  of a future individual with corresponding  $X_{\star}$  and  $Z_{\star}$ 

- Introduced by Meuwissen et al. (2001)
- Successfully implemented in many plant/animal breeds for traits related to production, health, climate adaptation, ...
- Modest gains in predictions can have large economic impacts (reduced generation interval, reduced cost and labor for phenotyping)

#### Challenges of genomic prediction models

- Non-random association between alleles at neighboring loci (aka LD)
- Polygenic nature of complex traits
- Many more SNPs (variables) than individuals (observations) ⇒ curse of dimensionality
  - Including too many predictors in a model risks over-fitting, poor generalizability, and problems with model estimation
  - ... but including only a small pre-identified subset of SNPs (e.g., significant GWAS hits) usually leads to poor predictions

 $\rightarrow$  Balance computational/statistical feasibility and biologically realistic assumptions

#### Challenges of genomic prediction models

- Non-random association between alleles at neighboring loci (aka LD)
- Polygenic nature of complex traits
- ullet Many more SNPs (variables) than individuals (observations)  $\Rightarrow$  curse of dimensionality
  - Including too many predictors in a model risks over-fitting, poor generalizability, and problems with model estimation
  - ... but including only a small pre-identified subset of SNPs (e.g., significant GWAS hits) usually leads to poor predictions
- $\rightarrow$  Balance computational/statistical feasibility and biologically realistic assumptions

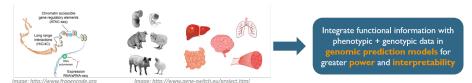
Can genomic prediction models be improved by better accounting for our **knowledge** about the **function** of certain regions of the genome?

#### Context: H2020 GENE-SWitCH project

The regulatory GENomE of Swine & Chicken: functional annotation during development

High-quality richly annotated maps of pig and chicken genomes:

- Development: early/late organogenesis, new born/hatched, adult
- Sexes: {M,F} × 3 biological replicates
- Tissues: liver, skeletal muscle, small intestine, cerebellum, dorsal epidermis, lung, kidney
- Assays: RNA-seq, ATAC-seq, ChIP-seq, smRNA-seq, methylation, Hi-C



#### Context: H2020 GENE-SWitCH project

The regulatory GENomE of Swine & Chicken: functional annotation during development

High-quality richly annotated maps of pig and chicken genomes:

- Development: early/late organogenesis, new born/hatched, adult
- Sexes: {M,F} × 3 biological replicates
- Tissues: liver, skeletal muscle, small intestine, cerebellum, dorsal epidermis, lung, kidney
- Assays: RNA-seq, ATAC-seq, ChIP-seq, smRNA-seq, methylation, Hi-C



But how?

#### First, back to basics: the linear model

The workhorse of genomic prediction is the multiple linear regression model:

$$Y = \mathbf{Z}\theta + \mathbf{X}\beta + \varepsilon$$

- Y = n-vector of traits
- $Z = n \times m$  matrix of covariates
- $\theta = m$ -vector of covariate effect parameters
- $\mathbf{X} = n \times p$  matrix of (suitably coded) genotypes
- $\beta = p$ -vector of genetic effect parameters
- $\varepsilon = n$ -vector of errors representing noise, assumed to be iid and (usually) normally distributed

#### Bayesian methods for genomic prediction

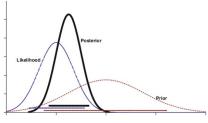
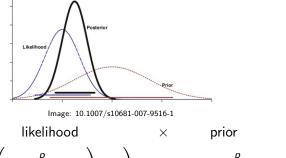


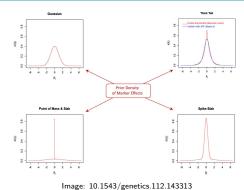
Image: 10.1007/s10681-007-9516-1

#### Bayesian methods for genomic prediction



$$\prod_{i=1}^{n} N\left(Y_{i} \middle| \left(\mu + \sum_{j=1}^{p} X_{ij} \beta_{j}\right), \sigma^{2}\right) \times p(\sigma^{2}) \prod_{j=1}^{p} p(\beta_{j} | \Psi)$$

- $\sigma^2$  often assigned a  $\chi^{-2}$  prior distribution
- Choice of prior for  $\beta_j$  should ideally reflect a trait's genetic architecture (and be computationally feasible...)



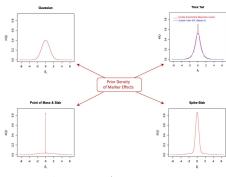


Image: 10.1543/genetics.112.143313

**GBLUP**:  $\beta_i \sim N(0, \sigma_\beta^2)$ 

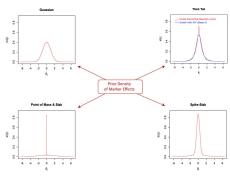


Image: 10.1543/genetics.112.143313

**GBLUP**:  $\beta_i \sim N(0, \sigma_\beta^2)$ 

**BayesA**:  $\beta_i \sim N(0, \sigma_{\beta_i}^2), \sigma_{\beta_i}^2 \sim \text{Inv } \chi^2(\nu, S^2)$ 

**BayesB**:  $\beta_i \sim N(0, \sigma_{\beta_i}^2), \sigma_{\beta_i}^2 \sim \pi \delta(0) + (1 - \pi) \text{Inv } \chi^2(\nu, S^2), \pi \text{ fixed}$ 

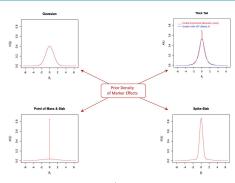


Image: 10.1543/genetics.112.143313

**GBLUP**:  $\beta_i \sim N(0, \sigma_\beta^2)$ 

**BayesA**:  $\beta_i \sim N(0, \sigma_{\beta_i}^2), \sigma_{\beta_i}^2 \sim \text{Inv } \chi^2(\nu, S^2)$ 

BayesB:  $\beta_i \sim N(0, \sigma_{\beta_i}^2), \sigma_{\beta_i}^2 \sim \pi \delta(0) + (1 - \pi) \text{Inv } \chi^2(\nu, S^2), \pi \text{ fixed}$ 

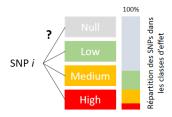
BayesC:  $\beta_i \sim \pi \delta(0) + (1-\pi)N(0,\sigma_\beta^2), \sigma_\beta^2 \sim \text{Inv } \chi^2(\nu,S^2)$ ,  $\pi$  fixed

**BayesC** $\pi$ : BayesC with  $\pi \sim \text{Unif}(0,1)$ 

## BayesR (Erbe et al., 2012)

$$\beta_i \sim \pi_1 \underbrace{\delta(0)}_{\text{null}} + \pi_2 \underbrace{N(0, 0.0001\sigma_g^2)}_{\text{small}} + \pi_3 \underbrace{N(0, 0.001\sigma_g^2)}_{\text{medium}} + \pi_4 \underbrace{N(0, 0.01\sigma_g^2)}_{\text{large}}$$

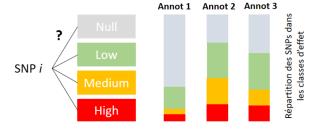
- $\pi \sim \mathsf{Dirichlet}(\alpha)$ , with  $\alpha = (1, 1, 1, 1)$
- Gibbs sampler for estimation



## Back to annotations: BayesRC (MacLeod et al., 2016)

$$f(\beta_i|\mathbf{C}_i = \mathbf{c}) = \sum_{k=1}^4 \pi_{\mathbf{c},k} f_k(\cdot|\theta_k)$$

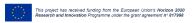
- SNPs assigned to disjoint "annotations", model is a factorized BayesR
- $\pi_c \sim \text{Dirichlet}(\alpha)$ , with  $\alpha = (1, 1, 1, 1)$
- Gibbs sampler for estimation



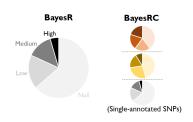


Genotype ...000001001201002100200010100001011001011110... Predict





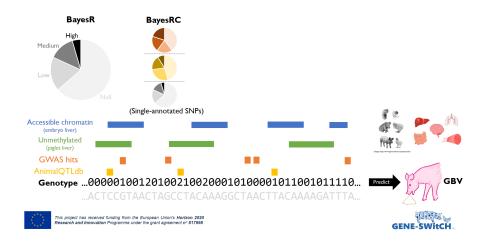


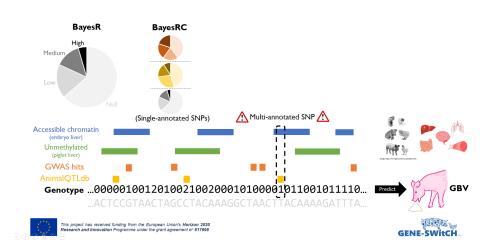


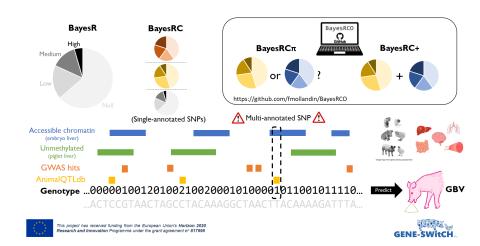












#### BayesRCO: BayesRC for Overlapping annotations

Two hypotheses = two models!

- lacktriangle Multi-annotations represent added confidenceightarrow BayesRC+
- **Q** Multi-annotations represent uncertainty  $\rightarrow$  BayesRC $\pi$

$$A \in \{0, 1\} = \begin{pmatrix} 1 & 1 & 1 & 1 \\ 0 & 0 & 1 & 1$$

	Method	SNP effect prior distribution	Annotations
	BayesR	$\beta_i \sim \sum_{K=1}^4 \pi_K \mathcal{N}(0, k \sigma_g^2)$	No
Cumulative	BayesRC	$\beta_i   a = A(i) \sim \sum_{K=1}^4 \pi_{K,a} \mathcal{N}(0, k\sigma_g^2)$	Yes, disjointed
	▶ BayesRC+	$\beta_i   a \in A(i) \sim \sum_{a \in A(i)} \sum_{K=1}^4 \pi_{K,a} N(0, k\sigma_g^2)$	Yes, overlapping
Preferential	$ ightharpoonup$ BayesRC $\pi$	$\beta_i a \in A(i) \sim \sum_{a \in A(i)} p_{i,a} \sum_{K=1}^4 \pi_{K,a} N(0, k\sigma_g^2)$	Yes, overlapping
assignment			

#### Simulation strategy

Phenotypes simulated from real cattle genotypes, 2500 animals:

- $-h^2 = \{0.2, 0.5\}$
- 5 large QTLs representing  $k = \{1\%, 2.5\%, 5\%\}$  of total additive variance  $\sigma_a^2$
- 300 medium QTLs representing 0.1% of  $\sigma_a^2$
- 4500 to 6500 low effect SNPs representing 0.01% of  $\sigma_a^2$
- 50 datasets generated for each setting

#### Scenarios

4 types of annotations possible:





strongly enriched: 5 large QTLs + 300 medium QTLs + 150 low or null SNPs



moderately enriched: 2 large QTLs + 100 medium QTLs + 300 low or null **SNPs** 

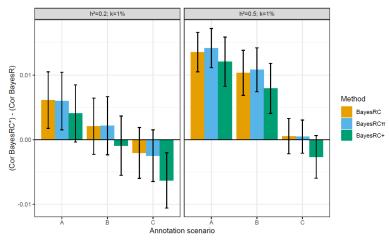


weakly enriched: 20 medium QTLs + 400 low or null SNPs



unenriched: 450 low or null SNPs

### Evaluating impact of using annotations on validation data



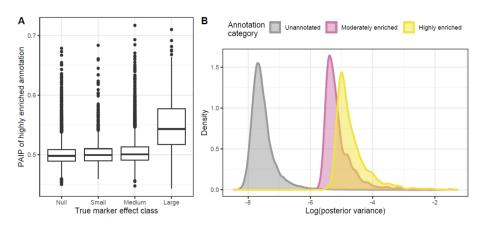
k= per-large QTL % of additive variance

A= 1 strongly enriched + 1 moderately enriched + unannotated;

B= 1 strongly enriched + 1 moderately enriched + 1 weakly enriched + 1 unenriched + unannotated

C= 2 strongly enriched + 2 moderately enriched + 3 weakly enriched + 2 unenriched + unannotated

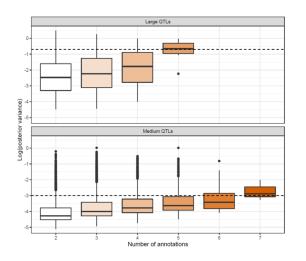
#### BayesRC $\pi$ assigns informative annotations to QTLs



 $h^2=0.5$ , k = 1%, scenario A PAIP = posterior annotation inclusion probability (BayesRC $\pi$  output)

## BayesRC+ assigns more weight to multi-annotated variants

Results



$$h^2 = 0.5$$
, k = 1%, scenario C

#### Application in backcross population of growing pigs

- n = 1297 backcross pigs (3/4 Large-White, 1/4 Creole), genetically related sows sired with 10 boars
  - Genotyped with Illumina Porcine 60k BeadChip array
  - Sibling-structured 10-fold cross validation procedure
- Traits pre-corrected for age, sex, farm
- Focus on average daily weight gain (ADG) and backfat thickness (BFT) at 23 weeks



#### Correlation of predicted traits in pig validation data

Annotations constructed using pigQTLdb for 11 trait sub-hierarchies

- Anatomy, behavioral, blood parameters, conformation, fatness, fatty acid content, feed conversion, growth, immune capacity, litter, reproductive organs
- Nearest up- and downstream neighboring markers also annotated

#### Correlation of predicted traits in pig validation data

Annotations constructed using pigQTLdb for 11 trait sub-hierarchies

- Anatomy, behavioral, blood parameters, conformation, fatness, fatty acid content, feed conversion, growth, immune capacity, litter, reproductive organs
- Nearest up- and downstream neighboring markers also annotated

	BayesR	BayesRC	BayesRCπ	BayesRC+
ADG	0.21 (±0.08)	+1.2 pts	+1.7 pts	+1.4 pts
BFT	0.26 (±0.16)	-0.6 pts	-1 pts	+0.6 pts

## Interpreting pigQTLdb annotations with BayesRC $\pi$



#### Conclusions: incorporating annotations with BayesRCO

#### BayesRCO:

- $\rightarrow$  **BayesRC** $\pi$  can assign informative annotations to multi-annotated SNPs to account for uncertainty in prior knowledge
- → BayesRC+ upweights multi-annotated SNPs and is robust to various annotation scenarios
  - ullet Fairly modest improvements in prediction ( $\sim$ 1-2 points) observed when incorporating biological annotations
    - Improved predictions and rankings of large QTLs in simulations, especially for highly informative annotations
    - Slight improvement in predictions for some traits in real data
    - Strategies for constructing annotation categories impact results

#### Take home messages

Can genomic prediction models be improved by better accounting for our knowledge about the function of certain regions of the genome?

#### Take home messages

Can genomic prediction models be improved by better accounting for our **knowledge** about the **function** of certain regions of the genome?

Yes, sometimes.

#### Take home messages

Can genomic prediction models be improved by better accounting for our **knowledge** about the **function** of certain regions of the genome?

#### Yes, sometimes.

- Models → BayesRCO for overlapping annotation categories, extensions in progress to handle quantitative annotations
- ullet Genotyping data o Capitalizing on annotation maps likely requires WGS resolution
- Validation data → Greater potential gains when prediction is performed on genetically distant populations
- ullet Traits o Heritability, genetic architecture, link with annotations, ...
- Annotations → Which molecular assays, in which tissues?

## Thank you!





Mollandin *et al.* (2022) Accounting for overlapping annotations in genomic prediction models of complex traits, *BMC Bioinformatics*, 23:65.



https://github.com/FAANG/BayesRCO