

Assessing the environmental impacts of conventional and organic scenarios of rainbow trout farming in France

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14 Abstract

15 In France, rainbow trout (Oncorhynchus mykiss) farming traditionally used flow-through 16 systems, which raised concerns about environmental impacts, including limited freshwater 17 availability, and the use of ingredients from intensive agriculture and fishing. To address the 18 growing demand for sustainable food products, there is an increasing interest in organic 19 aquaculture. In this study, we employed an attributional life cycle assessment (LCA) to analyse 20 the environmental impacts of rainbow trout production. We simulated conventional and organic 21 production practices in a hypothetical fish farm to evaluate the differences in environmental 22 impacts at the farm level. The potential impacts were calculated using a product-based 23 functional unit (one tonne of trout) under the two production scenarios and were also expressed 24 using a surface-based functional unit (m²y). Our life cycle impact assessment revealed that 25 organic farming significantly reduced environmental impacts per tonne of trout in seven out of 26 the nine selected impact categories. Notably, freshwater ecotoxicity exhibited the greatest 27 difference, with organic systems showing a 55% decrease. The only exceptions were freshwater 28 eutrophication and water dependence, where organic production led to higher impacts per tonne 29 of trout. In conventional farming, emissions amounted to 14 kg of P eq./tonne, whereas in 30 organic farming, the emissions were slightly higher (15 kg of P eq./tonne). For water 31 dependence, one tonne of trout production in the conventional system mobilized $128 \ 10^3 \ m^3 \ vs.$ 185 10³ m³ in the organic system. The environmental benefits of organic production were even 32 more marked when using a surface-based functional unit $(m^2 y)$. We demonstrated the benefits 33 34 of organic trout production from an environmental perspective. However, our findings highlight 35 the caution needed when interpreting LCA comparisons of such production systems that can be 36 highly influenced by methodological choices such as the functional unit used.

37 <u>Keywords:</u> Aquaculture systems; Conventional production; Fish; Life cycle assessment;
 38 Organic production

39 **1. Introduction**

40 Rainbow trout (Oncorhynchus mykiss) is the primary farmed fish species reared in France and 41 a significant salmonid species in the global aquaculture production (953,000 tonnes in 2021; 42 FAO, 2022). Only ~20% of this production is performed in seawater as done in Norway and 43 Chili while the vast majority is coming from freshwater production as practiced in Iran and 44 Turkey, the two main producing countries (FAO, 2023). Traditionally, freshwater trout farming 45 relied on flow-through systems with high water exchange. The lack of space for expansion and 46 new sites (due to competition with other uses and interests), limited freshwater availability, and 47 concerns over the sustainability of the aquafeeds are considered as key obstacles for further 48 expansion of conventional flow-through aquaculture systems (Albrektsen et al., 2022; Chen et 49 al., 2015; Maiolo et al., 2021). As consumer demand for sustainable and environmentally-50 friendly products grows, there is a rising interest in organic aquaculture, which aims to integrate 51 best environmental practices, natural resource preservation, and high animal welfare standards 52 (Ahmed et al., 2020).

53 Organic agriculture is often perceived as more sustainable than conventional farming 54 (Meemken and Qaim, 2018). Despite occupying only 1.6% of global agricultural land and 55 accounting for less than 10% of retail sales in most of the countries (Willer et al., 2023), organic 56 farming is one of the fastest-growing sectors in the food industry. Organizations such as the 57 International Federation of Organic Agriculture Movement (IFOAM), the Food and Agriculture 58 Organization (FAO) and the World Health Organization (WHO), through the Codex 59 Alimentarius, are working towards establishing an internationally agreed definition of organic 60 practices. In essence, organic farming is an agricultural system that places a high priority on the 61 well-being of ecosystems, encompassing soil, plants, animals, and humans. It relies on 62 ecological processes, biodiversity and cycles adapted to local conditions, rather than the use of 63 inputs with adverse effects. Moreover, organic farming promotes fair relationships and a good 64 quality of life for all involved (IFOAM, 2008). The magnitude of the benefits of organic farming can vary significantly depending on several factors, such as the farm-specific agricultural practices and management approaches, and local environmental conditions (Pépin et al., 2022; Smith et al., 2019). Thus, while organic farming generally fosters environmentally friendly practices, the actual environmental benefits can vary on a case-by-case basis (Meier et al., 2015). Therefore, a comprehensive assessment is necessary to accurately evaluate the overall environmental advantages of organic farming.

71 Different approaches have been employed to compare the environmental impacts of organic 72 and conventional farming systems, focusing on specific aspects such as biodiversity (e.g., Gabriel et al., 2013; Letourneau and Bothwell, 2008), land use (e.g., Badgley et al., 2007; 73 74 Connor, 2022; Gibson et al., 2007), or nutrient emissions (e.g., Nowak et al., 2013). However, 75 these assessments offer a limited perspective on the overall environmental impacts of 76 agricultural production. To provide a more comprehensive evaluation, efforts have been made 77 to develop multi-impact methods that can integrate various environmental impact categories, 78 enabling a holistic assessment. The reference method is the Life Cycle Assessment (LCA), 79 which examines the material and energy flows throughout a product's entire life cycle, 80 encompassing activities like raw material extraction, processing, manufacturing, transportation, 81 distribution, product use, maintenance, recycling, and waste management. LCA is recognized 82 as a comprehensive approach by researchers and international standards (ISO, 2006; Joint 83 Research Centre, 2010) and enables a thorough examination of the different stages and impacts 84 associated with a product's life cycle.

Tuomisto et al. (2012) and Meier et al. (2015) performed meta-analysis of Life Cycle Assessment (LCA) studies comparing the environmental impacts of organic and conventional terrestrial farming. Their findings indicate that organic farming practices generally yield positive environmental impacts per unit of area, although not necessarily per product unit. Organic production tends to exhibit higher levels of soil organic matter and reduced nutrient losses (such as nitrogen leaching, nitrous oxide emissions, and ammonia emissions). However, 91 when measured per product unit, organic systems were found to have higher levels of nutrient 92 emissions. Additionally, organic systems demonstrated lower energy requirements but higher 93 land use, eutrophication potential, and acidification potential per product unit. Nevertheless, 94 this meta-analysis only concerns land-based production In aquaculture, to the best of our 95 knowledge, only three case studies have been published in peer-reviewed literature: 96 comparisons of conventional and organic production of shrimps (Jonell and Henriksson, 2015) 97 and carp (Biermann and Geist, 2019), and comparison of ingredient types in salmon feeds 98 (Pelletier and Tyedmers, 2007).

99 This study aims at comparing the environmental impacts of conventional vs. organic rainbow 100 trout farming. To do that, we modelled a trout farm practicing either conventional or organic 101 rearing rainbow trout production. The model we have built aims to simulate a production farm 102 located in France, in Brittany, one of the main rainbow trout producing regions in the country.

103 **2. Materials and Methods**

104 2.1. Farm model

105 The farm model, developed using the R freeware (R Development Core Team, 2022), has been 106 partially adapted from previous investigations (Besson et al., 2017, 2016, 2014) to facilitate the 107 acquisition of input values required for conducting a LCA at the farm level. In the present study, the model was customized to simulate, on a daily basis, the production of rainbow trout (O. 108 109 mykiss) in a hypothetical flow-through farm, using actual farm data obtained from surveys 110 conducted in Britany in 2022. The hypothetical farm consisted of 12 concrete raceways, of 100 111 m³ each, for the pre-growing phase, and 24 concrete raceways of 250 m³ each for grow-out. 112 Among the 250 m³ raceways, 50% received first water, meaning that the water entered the tanks 113 directly from the river, while the remaining 50% received second water, supplied solely by the 114 outlet water from the upstream raceways (Figure 1). In addition to the raceways, the farm was 115 equipped with five feed storage silos and two warehouses measuring 60 and 80 m² (Figure 1).

Fish were initially stocked at 10 g and harvested at a fixed weight of 3,000 g that was assumed has the unique market size. The maximal annual production was fixed at 300 tonnes. Throughout the year, three batches of fry were stocked to stagger the sales period (Table 1). We simulated a production over 3 years and used the third year as the reference year for LCA (i.e. year where the first batches stocked in the first year reached market size).

121 The various parameters used and the constraints imposed, according to both conventional and 122 organic production scenarios, are elaborated in details below. We incorporated data from 123 surveys, scientific literature, and industry specifications to inform our analysis. Specifically, we 124 used the French production specifications for large trout provided by the Interprofessional 125 Committee for Aquaculture Products (CIPA, 2023) and the regulations for the organic 126 production of aquaculture species established by the French Ministry of Agriculture and 127 Fisheries (MAAP, 2010). A schematic representation of the modelling approach we employed 128 is depicted in Figure 2.

129 2.1.1. Environmental parameters

130 The daily temperature (T) was modelled using a sinusoidal function with a period of 365 days.

131 As suggested by Seginer and Halachmi (2008), T_n is given by:

132
$$T(n) = T_m - T_a \times \sin\left(2\pi \times \frac{n+\phi_T}{365}\right)$$
(1)

where n is a day from 1 to 365, T_m is the mean water temperature (13 °C), T_a is the amplitude of the variation (8°C corresponding to a difference of $2 \times 8 = 16$ °C between the minimum and maximum daily temperature across the whole year) and ϕ_T is the phase shift (time-delay of 27.36 d) (Figure 3A).

137 Dissolved oxygen concentration ($[O_2]$ in mg L⁻¹; Figure 3B) at day n in surface water was 138 calculated from Mortimer (1956) considering a standard pressure of 1 atm:

139
$$[0_2](n) = \exp^{7.7117 - \ln(T(n) + 45.93)}$$
 (2)

140 where T_n is the daily temperature (in °C).

141 The water flow within the fish farm, which experiences fluctuations throughout the year, was 142 simulated based on actual water flow data obtained from a river in Brittany. Data from the years 143 2018 to 2022, specifically from the Aulne River in Brittany, were collected from the reference 144 HydroPortail database version 3.1.4.3 (HydroPortail, 2023). Two constraints were considered when calculating the water flows: the inflow into the fish farm could not exceed 1.5 m³ s⁻¹, and 145 146 a maximum of 90% of the total river flow could be derived to the fish farm. To predict the daily 147 water inflows into the fish farm, a Generalized Additive Model (GAM) was then employed 148 considering the different constraints (Figure 3C).

149 2.1.2. Growth

The fish model described the daily weight and the daily weight gain of fish based on thermal growth coefficient (TGC). Considering that the relationship between growth rate and water temperature is non-linear, the TGC formula was corrected for the concave relationship between growth rate and temperature, using a corrected temperature K (Mallet et al., 1999) as suggested by Besson et al. (2016):

155
$$\operatorname{TGC} = \frac{W_{\mathrm{f}}^{\mathrm{b}} - W_{\mathrm{i}}^{\mathrm{b}}}{\Sigma_{\mathrm{i}=1}^{\mathrm{d}} \kappa_{\mathrm{i}}} \times 1000$$
 (3)

where W_f represents the final weight at harvest (3000 g), W_i denotes the initial weight at stocking (10 g), d is the rearing time in days and b is a weight coefficient set at 1/3 for the overall growing period even if this parameter can vary according to growth (Dumas et al., 2007).

The TGC values were adjusted to 1.80 and 1.45 ($g^{1/3} \circ C^{-1} d^{-1}$) in the conventional and organic production scenarios, respectively. We simulated a 24-month production cycle in conventional production and a 30-month production cycle in organic production (Figure 4). This rearing time difference corresponds to the expected growth differentials between triploid monosex trout, primarily used in conventional production, and male and female diploid trout (Aqualande Origins, 2019) used in organic production according to regulatory requirements (MAAP, 2010). In the conventional production scenario, the storage dates were kept constant throughout the three years and set at d 30 for the first batch, followed by intervals of 100 days (i.e., d 130 for batch 2 and d 230 for batch 3) over the course of a year. In the organic production scenario, the frequency of batch entries was set at 50 days (i.e., d 80 for batch 2 and d 130 for batch 3) to maintain the same rotation of harvests and stocking (3 entries and 3 harvests per year). This adjustment was necessary to accommodate the longer rearing duration (i.e. 30 vs 24 months) while ensuring consistent batch rotation in the organic production system.

173 The corrected temperature (K) at a given day n was calculated as follows:

174
$$K_{n} = \frac{T_{opt}(T_{n} - T_{min})(T_{n} - T_{max})}{(T_{n} - T_{min})(T_{n} - T_{max}) - (T_{n} - T_{opt})^{2}}$$
(4)

175 where $T_{min} \le K \le T_{max}$ and K = 0 for other values. Here, T_{min} and T_{max} represent the minimum 176 and maximum temperatures, respectively, below and above which growth does not occur. T_{opt} 177 refers to the optimal temperature for growth. Based on extrapolations from Bear et al. (2007), 178 the values for rainbow trout were set at 3 °C for T_{min} (K = 0), 13 °C for T_{opt} (K = 13), and 24 179 °C for T_{max} (K = 0). Consequently, for a positive growth rate, T_n must fall between 3 °C and 24 180 °C. The daily weight (W) and daily weight gain (DWG; g d⁻¹) can be calculated as follows at 181 day n:

182
$$W(n) = \left[W_i^{b} + \left(\frac{TGC}{1000} \times \sum_{i=1}^{n} K_i \right) \right]^{\frac{1}{b}}$$
(5)

183
$$DWG(n) = W(n) - W(n-1)$$
 (6)

184 Growth curves under the two production scenarios are presented in Figure 4A.

185 2.1.3. Mortality

In this study, a mortality rate of 15% was applied throughout the entire production cycle, spanning from 10 to 3000 g. It was assumed that the probability of daily mortality was not linear across the rearing period and is higher for younger individuals (Gåsnes et al., 2021). To model this, a Weibull function was considered for the lifetime distribution, as it is commonly used for

- 190 survival analysis (Carroll, 2003). So, the hazard function h which defines the death rate at a
- 191 given day (n) conditional on survival until time n or later can be calculated as follows:

192
$$h(n) = \frac{f(n)}{1 - F(n)}$$
 (7)

- 193 considering the Weibull density function $f(n) = 1 \exp^{-(\frac{n}{\lambda})^s}$ (8)
- 194 and the Weibull distribution function $F(n) = \frac{k}{\lambda} \left(\frac{n}{\lambda}\right)^{s-1} \exp^{-\left(\frac{n}{\lambda}\right)^s}$ (9)
- 195 While the shape parameter (s) was kept fixed at 0.5, the scale parameter (λ) was optimized for
- 196 each fish batch, ensuring a final mortality rate of 15% across the entire rearing duration.
- 197 2.1.4. Biomass
- 198 The biomass (BM) at a given day for each batch was determined as follows:

$$199 \quad BM(n) = W(n) \times SN(n) \tag{10}$$

- where W is the individual body weight at a given day n and SN the number of surviving fish at this day. In the same way, the dead biomass at day n was calculating by replacing SN by the number of dead fish at this day in the equation (10).
- The total production (in tonnes) was then calculated as the difference between the biomass at the harvest and the initial biomass at stocking. In the two different production scenarios, harvest took place at a constant weight of 3000 g.
- 206 2.1.5. Raceways occupation

207 In our model, the occupancy of the raceways was determined by the densities achieved, which 208 necessitates regular sorting of the fish during rearing. Initially, we assumed that each batch was stocked in a 100-m³ raceway. As the fish grow, they were periodically redistributed into 2 and 209 then 4 raceways of 100 m³ before ultimately occupying 4 then 8 raceways of 250 m³. The 210 211 maximum density constraints varied depending on the production scenario. In the conventional production scenario, the density limits applied were 50 kg m⁻³ when $W \le 50$ g, 70 kg m⁻³ when 212 213 50 g < W < 1000 g and then 90 kg m⁻³ when W > 1000 g (CIPA, 2023). For the organic production scenario, the density limits were as follows according to CIPA (2023): 25 kg m⁻³ 214

when $W \le 15$ g, 30 kg m⁻³ when 15 g < W \le 30 g and then 35 kg m⁻³ when W > 30 g. The percentage of occupancy of each rearing structure was calculated as the sum of the surface used per day divided by the total surface available over a year (expressed as m²y).

2.1.6. Feeds

218

Feed conversion ratio at a given day (FCR) was modelled by a third-order polynomial model based on fish body weight (W) using equation extrapolated from Bureau and Hua (2008) :

FCR(n) = $\alpha \times [(0.051 \times W(n)^3) - (0.261 \times W(n)^2) + (0.688 \times W(n)) + 0.65]$ (11) where α is a scaling factor to obtain a realized FCR of 1.30 kg kg⁻¹ over the production cycle for each batch in the two production scenarios assuming that the conventional and organic fish lines have the same feed efficiency (Figure 5). Daily feed intake (DFI, kg d⁻¹) is calculated back from FCR and DWG by:

226
$$DFI(n) = DWG(n) \times FCR(n)$$
 (12)

In our model, we considered the dynamic nature of fish feed composition, particularly in terms of protein and lipid content, throughout the rearing period. As a result, four different types of feed were incorporated based on the weight. Feed 1 was used for fish up to 50 g, feed 2 for fish up to 500 g, feed 3 for fish up 1500 g, and finally, feed 4 was used until reaching the harvest weight (W_f). This approach ensures that the nutritional needs of the fish are adequately met at each stage of their growth and development. Conventional or certified organic feeds were used depending on the production scenario.

234 2.1.7. Nutrient release

The concentration of nutrients (N and P) and chemical oxygen demand (COD) in effluent water was determined using a mass-balance approach (Aubin et al., 2011). To model excretion, the first step involved calculating the total nutrient amount provided by the feeds (N_{feed}), taking into account two fractions: the portion consumed (N_{eaten}) and the portion wasted (N_{waste}) on day n, along with the nutrient fixation by the fish (N_{fish}). It was assumed that 1% of the distributed

- 240 feeds remained uneaten (Boujard et al., 1995). The proximate composition of the different feeds
- can be found in Table 2.
- 242 $N_{\text{feeds}}(n) = N_{\text{content}} \times \text{DFI}(n)$ (13)
- 243 $N_{waste}(n) = N_{feeds}(n) \times 0.01$ (14)
- 244 $N_{eaten}(n) = N_{feeds}(n) N_{waste}(n)$ (15)

245
$$N_{fish}(n) = N_{fish_{body}} \times DWG(n) \times SN(n)$$
 (16)

- 246 where $N_{fish_{body}}$ is the nutrient composition of the fish (in kg kg⁻¹) set at 0.03 for N (Oz and
- 247 Dikel, 2015) and 0.004 for P (Kause et al., 2022).
- 248 The total nutrient excretion (N_{excretion}) was given by:

249
$$N_{\text{excretion}}(n) = N_{\text{eaten}}(n) - N_{\text{fish}}(n)$$
 (17)

250 Calculation of the suspended (N_{suspended}) and dissolved (N_{dissolved}) was given by:

251
$$N_{suspended}(n) = N_{eaten}(n) \times (1 - Dig_N)$$
 (18)

252
$$N_{dissolved}(n) = N_{excretion}(n) - N_{suspended}(n)$$
 (19)

where Dig_N is the digestibility coefficient set at 94% for proteins and 61% for P (Dalsgaard and
Pedersen, 2011).

The final amount of N release was then calculated considering that the sedimentation area used as water treatment is able to remove 20% of suspended N (Stewart et al., 2006):

257
$$N_{release}(n) = 0.8 \times N_{suspended}(n) + N_{dissolved}(n)$$
 (20)

COD at a given day n was calculated using feed quantity eaten (DFI) at day n, the proximate
protein, lipids and carbohydrates contents of the feeds (P_{feeds}, L_{feeds} and C_{feeds}) and their
respective digestibility (Dig) (i.e., 94% for proteins, 91% for lipids and 67% of carbohydrates;
Dalsgaard and Pedersen, 2011):

262
$$\operatorname{COD}(n) = \left[\left(P_{feeds}(n) \times (1 - \operatorname{Dig}_{P}) \times 1.66 \right) + \left(L_{feeds}(n) \times (1 - \operatorname{Dig}_{L}) \times 2.78 \right) + \right]$$

263
$$\left(C_{feeds}(n) \times (1 - \text{Dig}_{C}) \times 1.19\right) \times \text{DFI}(n)$$
 (21)

264 where the coefficients applied for protein, lipids and carbohydrates were coming from Meriac

265 et al. (2014).

266 2.1.8. Oxygen

267 In both production scenarios, the primary constraint for oxygen management was to maintain a 268 saturation level of 80% at the outlet of the raceways. However, the approach to O_2 269 supplementation differed between the two production scenarios. In conventional production, 270 liquid oxygen was used for O₂ supplementation, whereas in organic production, the use of 271 aerators was the only permissible method (MAAP, 2010). In our model, the amount of oxygen 272 added was determined based on the difference between the supply of oxygen through the water 273 inlet (O_{2inlet}), which could come directly from the river or from the upstream raceways (Figure 274 1), and the O_2 consumption by the fish (O_{2cons}). These two parameters were calculated using the 275 following equations:

276
$$O_{2_{inlet}}(n) = O_{2_{conc}}(n) \times Water_{flow}(n)$$
(22)

where O_{2conc} is the O_2 concentration from water inlet either coming from the river - in this case $O_{2conc} = [O_2](n)$ (see Section 2.1.1) or from the upstream raceway – in this case $O_{2conc} =$ $[O_2](n) - O_{2cons}(n)$ of the upstream raceway. Water_{flow} in a given raceway the water flow through the raceway calculated as follows:

281 Water_{flow}(n) =
$$\alpha \times \frac{Water_{total}(n)}{RN}$$

where Water_{total} is the water flow for the whole fish farm, RN is the number of raceways and a is a size coefficient (i.e. 0.29 for 100-m³ raceway and 0.71 for 250-m³ raceway). Then, O₂ consumption is given by:

285
$$0_{2_{\text{cons}}}(n) = \left[(\text{DFI}(n) \times P_{\text{feeds}}(n) \times \text{Dig}_{P}) - (\text{DFI}(n) \times P_{\text{fish}}(n)) \right] \times \frac{E_{P}}{Q_{\text{oxP}}} + \left[(\text{DFI}(n) \times P_{\text{feeds}}(n) \times \text{Dig}_{P}) - (\text{DFI}(n) \times P_{\text{fish}}(n)) \right] \times \frac{E_{P}}{Q_{\text{oxP}}} + \left[(\text{DFI}(n) \times P_{\text{feeds}}(n) \times \text{Dig}_{P}) - (\text{DFI}(n) \times P_{\text{fish}}(n)) \right] \times \frac{E_{P}}{Q_{\text{oxP}}} + \left[(\text{DFI}(n) \times P_{\text{fish}}(n) \right] \times \frac{$$

287
$$\left(\text{DFI}(n) \times C_{\text{fish}}(n)\right) \times \frac{E_{\text{C}}}{Q_{\text{oxC}}}$$
 (23)

where Q_{oxP} , Q_{oxL} and Q_{oxC} are the oxy-caloric coefficients of proteins (13.4 MJ kg O₂⁻¹), lipids

289 (13.7 MJ kg O_2^{-1}) and carbohydrates (14.8 MJ kg O_2^{-1}) (Brafield and Solomon, 1972; Elliott

and Davison, 1975) and E_P , E_L and E_C are the energy contents of proteins (23.6 MJ kg⁻¹), lipids

291 (39.5 MJ kg⁻¹) and carbohydrates (17.2 MJ kg⁻¹) (Brafield and Llewellyn, 1982).

If the difference between O_{2inlet} and O_{2cons} was higher than the 80% saturation O_2 concentration ($O_{2_{-80\%}} = 0.8 [O_2]$), it indicated that no oxygenation or aeration is required. Conversely, a result lower than $O_{2_{-80\%}}$ indicated the need for O_2 supplementation (O_{2sup}) either through the addition of liquid O_2 or by aeration:

296
$$O_{2_{sup}}(n) = 0$$
 when $O_{2_{inlet}}(n) - O_{2_{cons}}(n) > O_{2_{_{80\%}}}(n)$ (24)

297
$$O_{2_{sup}}(n) = |O_{2_{inlet}}(n) - O_{2_{cons}}(n)|$$
 when $O_{2_{inlet}}(n) - O_{2_{cons}}(n) < O_{2_{\underline{80\%}}}(n)$ (25)

The electricity consumption of the farm was modelled taking into account water filtration, oxygenation and recirculation processes. A drum filter (1 kWh) and a recirculation pump (20kWh) operated during periods when the water flow was at its lowest, typically between May and September. Their purpose was to ensure effective water recirculation during this period under both conventional and organic production scenarios. Electricity consumption by the filter (Efilter) and the recirculation pump (E_{pump}) at a given day n has been calculated as follows:

305
$$E_{filter}(n) = 1 \times 24$$
 for May < n < September and $E_{filter}(n) = 0$ for other dates (26)

306
$$E_{pump}(n) = 20 \times 24$$
 for May < n < September and $E_{pump}(n) = 0$ for other dates (27)

307 One key distinction between estimating electricity consumption for conventional and organic 308 production lies in the method employed for water oxygenation. In conventional production, 309 liquid oxygen was added using an oxygen cone and two pumps with a power consumption of 310 20 kWh each. In this case, the electrical consumption at a given day n was calculated as follows:

311
$$E_{oxygen}(n) = 2 \times 20 \times 24 \text{ for } O_{2_{sup}}(n) > 0$$
 (28)

312
$$E_{oxygen}(n) = 0 \text{ for } O_{2_{sup}}(n) = 0$$
 (29)

13

In the organic production scenario, the use of aerators replaced liquid oxygen. These aerators enable the addition of 1.5 kg of oxygen per kilowatt-hour (kWh) of electricity consumed (Ahmad and Boyd, 1988; Brown et al., 2014). Consequently, the calculation for electrical consumption associated with the aerators has been calculated as follows:

317
$$E_{oxygen}(n) = \frac{O_{2sup}(n)}{1.5}$$
 (30)

318 The total electricity consumption (E_{total}) was determined by summing the electricity usage for 319 water filtration, oxygenation, and recirculation:

320
$$E_{total}(n) = E_{filter}(n) + E_{pump}(n) + [E_{oxygen}(n) \text{ or } E_{aeration}(n)]$$
(31)

321 2.2. Life Cycle Assessment (LCA)

322 2.2.1. Goal and scope

323 An attributional LCA was conducted according to the general requirements of the methodology 324 proposed by ILCD standards (Joint Research Centre, 2010). The methodology was adapted to 325 the characteristics of fish farming. The goal and scope of this study was the environmental 326 assessment of trout farming in a hypothetical farm producing large rainbow trout following 327 either (1) conventional or (2) organic practices in the same infrastructures. The system was 328 defined from cradle-to-farm-gate and included five distinct sub-systems (Figure 6): (1) 329 production of purchased feed, including cultivation of ingredients, processing, and 330 transportation; (2) production of energy expended at farm level (electricity); (3) production of 331 farming facilities and equipment used: (4) chemicals, including liquid oxygen, veterinary and 332 disinfection products, and their transportation (5) farm operations, including nutrient emissions 333 from the biological transformation of feed after onsite treatment of wastewater (see details in 334 Section 2.2.2). The functional unit in which environmental impacts were expressed was one 335 tonne of trout produced at farm level on a basis of one year of routine production. We also 336 expressed the environmental impacts using an surface-based functional unit (m^2y) as 337 recommended by Van der Werf et al. (2020). Here we considered only the surface directly 338 involved in the fish production.

339 2.2.2. Life cycle inventory

The life cycle inventory (LCI), presented in Table 3, was conducted by running our farm model with the specifications for both conventional and organic production scenarios. All the inputs and outputs were calculated using all the results from each batch of fish over one year of routine production generated as described in the farm model. The Agribalyse version 3.0 (Koch and Salou, 2022) and Ecoinvent version 3.8 (Wernet et al., 2016) databases were used to obtain the necessary data for conducting the assessment. Both databases are grounded on the recommendations in international standards (Wolf et al., 2012).

(1) Production of purchased feed - Crop-derived ingredients used in fish feed mainly originated 347 348 from Brazil and France (e.g. soybean meal from Brazil and wheat bran from France). Fish-349 derived ingredients originated from the Peruvian and the Norwegian fish milling industry (e.g. 350 fish meal from Peru and fish meal from fish trimming from Norway). The exact composition 351 of the different feeds used and their nutritional values were given by the feed manufacturer (Le 352 Gouessant, personal communication). The transport of feed ingredients to feed manufacturers 353 in France was by trans-oceanic ship and by lorry (>32 t), whereas the transport of feed from 354 France to the fish farm in Brittany was by lorry (>32 t). Road distances were calculated from 355 Google Maps and ocean distances were assessed from shiptraffic.net. Other data required to 356 compute the environmental impact of feed ingredients were based on the literature (Boissy et 357 al., 2011; Pelletier et al., 2009).

(2) *Production of energy expended on the farm* - The electricity used by the farm was coming
from the French energy mix in the Ecoinvent database. Annual on-site consumption from other
energy sources (diesel and gas) where considered negligible.

361 (3) *Production of farming facilities and equipment used* - We considered the construction of
362 two different buildings with a life span of 30 years. Nevertheless, the life span of each rearing
363 structures has been adjusted in LCA inventory according to the rearing structures' occupancy
364 (Table 4) calculated as described in Section 2.1.5 assuming that the actual life span of the

365 rearing span is related to their level of occupancy. The production of equipment used (i.e. pump,

tanks) was calculated using data from INRAE.

(4) *Chemicals* - This sub-system included the veterinary and disinfection products. While the
use of these products varies little between conventional and organic production, the main
difference is the inclusion in this sub-system of the liquid oxygen used only in conventional
production. Here we considered production of liquid oxygen from cryogenic air separation
process.

(5) *Farm operations* - The farm operation sub-system included the use of facilities and equipment and the emissions of pollutants from the biological transformation of the feed distributed to the fish. The amount of nitrogen (N), phosphorus (P) and chemical oxygen demand (COD) of the dissolved organic matter excreted by the fish in effluent water were calculated through mass balance (Papatryphon et al., 2005) considering the onsite treatment capacity of the sludge settling pond. Sludge produced by the farm was used for neighbourhood agricultural purposes and was not included in the analysis.

379 Gaps in the inventory were filled on the basis of the assumptions reported in Table 4.

380 2.2.3. Life cycle impact assessment

The assessment of the impact was carried out using ReCiPe 2016 Midpoint (H) version 1.07 (Huijbregts et al., 2017), which is a methodology based on the Eco-indicator and CML approaches. According to the European Commission/JRC (2010), ReCiPe represents the most up-to-date and standardized indicator approach available for life cycle impact assessment.

Table 5 provides a breakdown of the nine selected impact categories from ReCiPe, namely climate change (GWP), terrestrial acidification (TAP), freshwater eutrophication (FEP), marine eutrophication (MEP), terrestrial ecotoxicity (TETP), freshwater ecotoxicity (FETP), land use (LU), water dependence (WD) and the Cumulative Energy Demand method (CED; Frischknecht et al., 2007). These impact categories have been identified among the most suitable indicators of aquaculture impacts (Bohnes et al., 2019). To enable comparison with 391 previous studies on trout production systems, the CML baseline (Guinée, 2002) was used as an

alternative to the ReCiPe approach. The environmental impacts were calculated using Simapro
version 8.0 software (PRé Consultants, 2014).

394 2.2.4. Sensitivity analysis

395 Considering that the feed use is the major contributors to environmental impacts, a sensitivity 396 analysis was conducted on FCR. In this study, we ran the model, both for the conventional and 397 organic productions, to gauge the changes in the different LCA impact categories when FCR 398 varied from 1.0 to 1.6 in steps of 0.1.

399 **3. Results**

400 Figure 7 presents the level of the environmental impacts and the contribution of the system 401 components to the impacts for conventional and organic productions of rainbow trout. The 402 impacts are calculated according to ReCiPe method using two different functional units: per 403 tonne of trout (product-based) and per m^2y (surface-based). Among the nine impact categories 404 analyzed, the conventional production system exhibits higher impacts for all categories, except 405 for FEP and WD when the results are expressed per tonne of trout. For instance, in the 406 conventional production system, one tonne of trout emits 14 kg P eq. and depends on 128,000 407 m³ of water, while an equivalent quantity of organic trout emits 19 kg P eq. and depends on 408 185,000 m^3 of water. Obviously, when the results are expressed per m^2 y, organic production 409 shows lower environmental impacts for all considered categories, including FEP and WD 410 (Figure 7). Regardless of the functional units chosen (product-based or surface-based), other 411 impact categories also follow similar trends, with the surface-based functional unit leading to a 412 larger gap between the two production scenarios. The results presented in Figure 7 cannot be 413 compared with those presented in previous LCA publications, which were mostly obtained with 414 the CML baseline method. Thus, the environmental impacts of the production of one trout were 415 also assessed using the CML baseline methodology and compared with literature for 416 conventional production (Table 6). Overall, our results are consistent with those found in 417 previous studies. For the sake of clarity, the values presented in the subsequent paragraphs are

418 given for the ReCiPe method and per tonne of trout at market size, while the results expressed 419 per m^2y can be found in Figure 7.

420 The highest environmental gains observed in organic system compared to the conventional 421 production were for FETP (55% less in organic system): the production of one tonne of trout 422 induced 50 kg 1,4-DCB/tonne in conventional system, but the FETP value of the organic system 423 was noticeably lower with 33 kg 1,4-DCB/tonne. The energy requirements (CED) for producing 424 one tonne of trout were also noticeably different between the two production systems (30% less 425 in organic system) with values of 68 and 53 GJ/tonne in conventional and organic production, 426 respectively. Terrestrial acidification potential (TAP) was 28% less in organic system with 427 values of 15 and 12 kg SO_2 eq./tonne in conventional and organic production, respectively. 428 Differences between conventional and organic productions were less pronounced for the other 429 impact categories. GWP showed a reduction of 12% in the organic system with, per kg of fish 430 produced: 2602 kg CO_2 eq./tonne were estimated in conventional system vs. 2319 kg CO_2 431 eq./tonne for the organic system (Figure 7). The environmental gains through organic 432 production were equal for LU and TETP, both of these impact categories showing a reduction 433 of 11% in organic system while MEP is only diminished by 7% (Figure 7).

434 Contributions of the rearing system components (i.e., chemicals, dead biomass, energy, 435 equipment, feeds, fry, and farm functioning) varied according to the impact category and to the 436 production system (Figure 7). Overall, the ranking of the different contributors among the seven 437 impact categories remained relatively constant between conventional and organic productions 438 with the exception of chemicals, mostly driven by liquid oxygen, accounting for a non-439 negligible part of the environmental impacts in conventional production but not in organic 440 production (Figure 7). Results presented in Figure 7 show that, for MEP, FEP, and WD, farm 441 operations contributed the most to the impacts (81-84%, 90-93% and 93-98%, respectively), 442 and the second largest contributors are either feeds for MEP and FEP (15-18% and 5-6%, 443 respectively) or energy for WD (1-5%). For five out of nine impact categories (i.e., LU, TAP, 444 GWP, CED and TETP), exogenous feeds were the main contributors (96-97%, 79-90%, 66-445 85%, 50-79% and 53%, respectively), whatever the production systems. Equipment and 446 infrastructures are playing a significant role in the FETP and TETP impacts in the two 447 production systems (20-53%) while their role is relatively negligible in the other impact 448 categories. As mentioned earlier, the most remarkable difference in the contributions to 449 environmental impacts of the two production systems concerns the role of chemicals. Indeed, 450 chemicals include the use of antibiotics, other veterinary products and disinfectants, the use of 451 which remains relatively constant between conventional and organic production (Table 3). On 452 the other hand, the major difference is related to the use of liquid oxygen, included in chemicals, 453 only in conventional production (Table 3). Thus, while chemicals represent only < 2% of CED, 454 FETP, GWP, TAP and TETP and CED in organic production, they represent between 13% and 455 44% of the corresponding impacts in conventional production (Figure 7).

The sensitivity analysis results indicate a linear relationship between FCR and the environmental impacts of rainbow trout farming for the nine impact categories considered in this study (Figure 8). Across most of impact categories considered, a reduction of 0.1 kg kg⁻¹ in FCR leaded to a decrease of the environmental impacts decreased by 3 to 12%. Notably, the most substantial differences were observed for FEP. However, it is worth mentioning that improve feed efficiency had a negligible effect on WD, mostly linked to the water volume derived from the river and passing through the rearing structures, reducing it by less than 1%.

463 **4. Discussion**

Despite the rapid growth of organic agriculture production, organic finfish aquaculture remains relatively new and is still in its early stages (Mente et al., 2011). In Europe, the development of this sector has been hindered by technical challenges, such as the limited availability of organic feed and fry. Additionally, establishing effective communication strategies with clients proves difficult due to competition from other certification schemes, such as the Aquaculture Stewardship Council (ASC) or the Marine Stewardship Council (MSC) (European Comission, 2022). Furthermore, some organic farming systems experience lower yields, and previous research has suggested that the use of organic feed ingredients may lead to reduced farm ecoefficiency and increased environmental concerns (Pelletier and Tyedmers, 2007). However, there is a scarcity of peer-reviewed studies comparing the environmental impacts of conventional and organic aquaculture production systems (Biermann and Geist, 2019; Jonell and Henriksson, 2015).

476 The current studies in this field have predominantly followed a field-based approach, wherein 477 data was directly collected from both conventional and organic farms to establish the LCI. 478 However, employing such an approach may introduce certain bias, particularly regarding the 479 distinction between differences arising from the specific production systems (conventional or 480 organic) themselves and variations inherent to individual farming practices, which can 481 significantly impact the interpretation of the LCA results (Chen and Corson, 2014). It is 482 especially true in a context where the representativeness of farming practices is sometimes 483 called into question in the LCA studies carried out in animal production (Meier et al., 2015). In 484 this study, we employed a modelling approach, associated with LCA to compare environmental 485 impacts of conventional and organic rainbow trout production within a hypothetical farm. The 486 farm's infrastructures and available surface area for production were kept constant in the two 487 scenarios to determine the differences in environmental impacts between conventional and 488 organic production in the same infrastructures.

Before delving into the analysis of environmental impacts between the two studied production systems (conventional and organic), it is crucial to establish a reference point by comparing the results obtained in the conventional production scenario with those from existing literature. This step allows us to compare the results from modelling with those obtained from actual fish farm data. To achieve this, we have used not only the ReCiPe method but also CML baseline method (Guinée, 2002). The latter was commonly used in previous LCA studies focusing on 495 rainbow trout aquaculture while ReCiPe was only recently used in a rainbow trout aquaculture 496 context notably in Italy (Maiolo et al., 2021) and Spain (Sanchez-Matos et al., 2023). Overall, 497 the literature comparison corroborated our findings when expressed environment impact per 498 tonne of trout (Table 6). Indeed, our results are consistent with the literature (Aubin et al., 2009; 499 Boissy et al., 2011; Chen et al., 2015; d'Orbcastel et al., 2009; Dekamin et al., 2015; Maiolo et 500 al., 2021; Papatryphon et al., 2004; Samuel-Fitwi et al., 2013) even if the ranges of reported 501 values can be wide. Despite uncertainties related to varying inventory databases and CML 502 assessment method versions, another underlying cause of the differences in environmental 503 impacts among studies is the use of diverse production systems and varying FCRs to achieve 504 producing the same quantity of trout (Philis et al., 2019; Sanchez-Matos et al., 2023).

505 The choice of functional units in LCA is a crucial point considered to conduct comparisons of 506 production systems because its influences allocation decisions at the farm gate (Henriksson et 507 al., 2012). Van der Werf et al. (2020) highlighted the interest in combining product-based and 508 area-based LCA when comparing conventional and organic production systems. For instance, 509 although organic animal production generally emits fewer pollutants per unit of land occupied 510 than conventional agriculture (an surface-based approach), it may have higher impacts per unit 511 of product (e.g., land occupation, eutrophication and acidification) (Meier et al., 2015). Thus, 512 while we used one tonne of trout as a first functional unit we also expressed the environmental 513 impacts using a surface-based functional unit (m²y), an original approach in LCA aquaculture 514 studies (Bohnes et al., 2019; Pouil et al., 2023).

515 Overall, our study highlights a significant lower level of environmental impacts of organic 516 production compared to conventional production. However, when impacts are expressed per 517 tonne of trout, the WD and the FED are higher in the organic system than in conventional 518 system. Nonetheless, it is important to be cautious when comparing the environmental 519 performance of the two production systems using a product-based functional unit because the 520 production capacity in the organic system is one-third lower. Specifically, the production of 521 organic trout is limited by the lower rearing densities and reduced inputs, such as the absence 522 of liquid oxygen (MAAP, 2010), while conventional intensive systems are managed with high 523 stocking rates and inputs to achieve high productivity (CIPA, 2023). As a result, in the 524 conventional production system, the environmental impacts are somewhat diluted by the larger 525 production volume. This limitation should be considered when comparing organic and 526 conventional systems using LCA and highlights the need to explore alternative surface-based 527 functional units to gain a more comprehensive understanding of the comparison (van der Werf 528 et al., 2020). By using a surface-based functional unit (m^2y) , we find that the FEP and the WD 529 become similar between the two production systems and even slightly lower in organic system 530 due to the absence of liquid oxygen usage. Our study demonstrates the benefits of organic trout 531 production in terms of overall environmental impacts. Considering the nuances related to 532 production capacity and LCA functional units is, however, crucial to gain a well-rounded 533 perspective on the environmental performance of both systems.

534 The significant importance of liquid oxygen usage in conventional production becomes 535 apparent when conducting a more detailed analysis of the contributions to environmental 536 impacts between the two production systems. This factor often serves as the key explanation 537 for the differences observed in impacts. Previous studies have also underscored the significance 538 of liquid oxygen in the environmental impacts associated with aquaculture production. For 539 instance, Song et al. (2019) highlighted that liquid oxygen contributed between 5% and 22% to 540 all LCA impact categories. Consequently, it is evident that such production inputs should not 541 be overlooked in LCA conducted for aquaculture production systems. Likewise, the role of 542 aquafeeds in influencing environmental impacts is fundamental, regardless of whether it is for 543 organic or conventional production. The importance of FCR and aquafeeds, in general, has been 544 emphasized by numerous LCA practitioners. Several studies have already concluded that feed 545 production constitutes a major environmental impact source (e.g., Aubin, 2013; Bohnes et al., 546 2019; Wilfart et al., 2023). Although organic feed helps reduce environmental impacts in many 547 categories, its higher proportion of fishmeal and fish oil, which are rich in P (Oliva-Teles et al., 548 2015), leads to a greater release of phosphate into the environment in the organic production 549 scenario, resulting in an increased risk of FEP as shown in Figure S1. It is worth noting that 550 while feed formulations cannot be entirely disclosed due to industrial secrecy, efforts have been 551 made to evolve these formulations. Nonetheless, these results align with the findings of Pelletier 552 and Tyedmers (2007) who reported considerably lower environmental impacts when feeds 553 contained reduced proportions of fish ingredients.

554 Given the paramount importance of feeds in determining the environmental impacts of our 555 production systems, we investigated the effects of a change in FCR on impact categories 556 encompassed in LCA. Our aim was to shed light on the relationship between FCR and 557 environmental impacts per tonne of trout in our production systems. Here, we established a 558 positive linear correlation between FCR and the environmental impacts observed. Such findings 559 agree with previous LCA studies reporting that all environmental impacts decrease in similar 560 proportions together with the improvement of FCR (d'Orbcastel et al., 2009; Jouannais et al., 561 2023; Elias Papatryphon et al., 2004). Our findings align with the conclusions drawn in a meta-562 analysis conducted by Philis et al. (2019), revealing a similar positive relationship between FCR 563 and environmental impacts when comparing the environmental impacts associated with 564 different salmonid production systems. This observation holds for changes of the FCR within 565 a same production system and does not hold anymore across systems (Jouannais et al., 2023). 566 Indeed, while the trend is quite clear in Recirculating Aquaculture Systems (RAS), it is notably 567 less when considering open production systems like land-based flow-through system or open 568 sea cages (Philis et al., 2019). Such dissimilarity can be attributed to inherent variations in the 569 studies themselves, which become more pronounced when analysing flow-through production 570 systems. The RAS, being more controlled, lend themselves to easier comparability across 571 studies. In contrast, the complexities and diverse factors associated with production in flow-572 through systems make it challenging to draw generalizable conclusions. Nevertheless, the 573 emergence of RAS as an alternative conventional rearing system to flow-through in trout 574 farming has brought about new challenges, including increased energy consumption, 575 dependence on equipment like pumps and filters, and potential greenhouse gas emissions and 576 environmental footprint associated with energy production and waste management (Ahmed and 577 Turchini, 2021; d'Orbcastel et al., 2009). Given the of absence of recent comparative LCA 578 available in the literature (d'Orbcastel et al., 2009; Dekamin et al., 2015; Samuel-Fitwi et al., 579 2013), it could be interesting to adapt our model for comparison between flow-through systems 580 and RAS in trout farming.

581 **5. Conclusion**

582 In our study, our modelling approach based on a hypothetical farm performing rainbow trout 583 production under conventional and organic production constraints combined with LCA 584 succeeded in drawing a fair description of conventional and organic scenarios of trout 585 production enable to compare the environmental impacts at the level of the same farm. Our 586 study demonstrates the benefits of organic trout production in terms of overall environmental 587 impacts, which is not common regarding livestock systems. Nonetheless, our findings 588 underscore the need for caution when interpreting LCA comparisons of such production 589 systems, as they can be significantly impacted by methodological choices such as the chosen 590 functional unit. Our analysis reveals that aquafeeds and liquid oxygen usage are key factors 591 contributing to the environmental impacts of conventional and/or organic trout production 592 systems. By recognizing and addressing the significance of these inputs, we can take further 593 steps towards sustainable finfish aquaculture practices.

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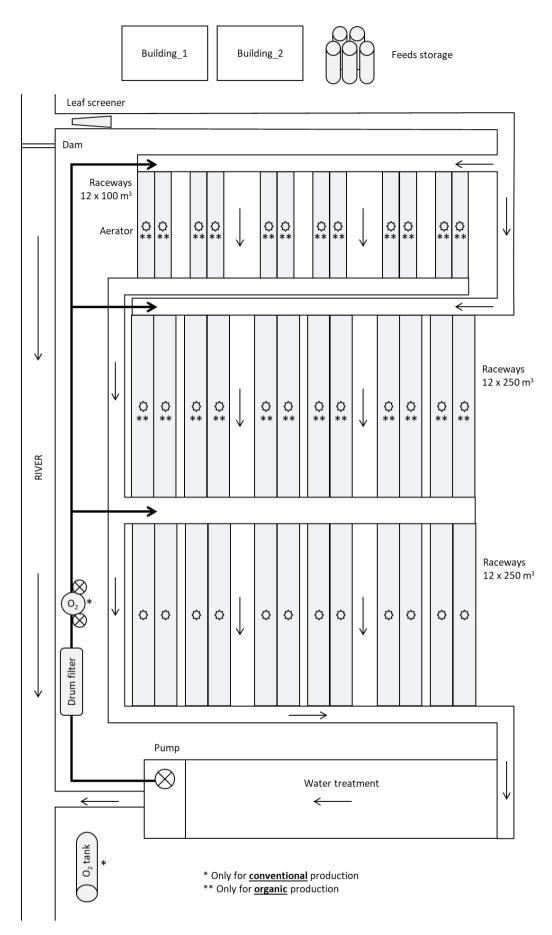
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- 835

836 Captions to figures

837

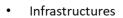
Figure 1. Schematic view of the hypothetical rainbow trout farm used to model conventionaland organic production. The equipment specific to the conventional and organic production

- system are annotated with the following symbols: * and **, respectively.
- Figure 2. Schematic view of the modelling approach we used. The values for the different model
- 842 models and details of the constraints applied for the conventional and organic systems are
- 843 detailed in the text.
- 844 Figure 3. Graphical representations of (A) the simulated annual temperature conditions with, in
- 845 insert, corrected temperature K as a function of temperature T and (B) the resulting oxygen
- 846 concentration in water. (C) the simulated annual water flow entering the fish farm.
- Figure 4. Graphical representations of (A) growth performances, from 0.01 to 3 kg and (B)
- 848 survival of the three fish batches in conventional and organic production systems.
- Figure 5. Estimated FCR of rainbow trout at increasing live weight extrapolated from Bureauand Hua (2008).
- Figure 6. System boundaries and flows of rainbow trout *Oncorhynchus mykiss* grow-outproduction.
- Figure 7. Contribution of each input or production step in environmental impacts in conventional and organic fish production system. Results are either expressed per tonne of trout
- at market size (product-based) or per m^2y (surface-based).
- 856 Figure 8. Influence of FCR variations in the environmental impacts per tonne of rainbow trout
- at market size in conventional and organic production system.

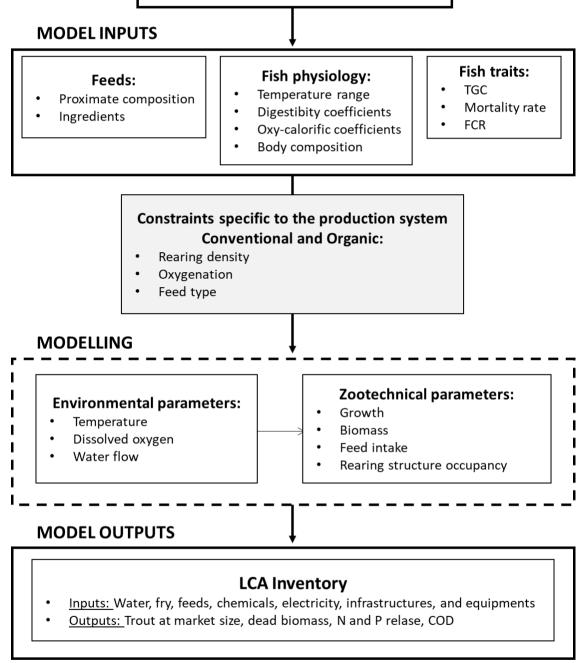




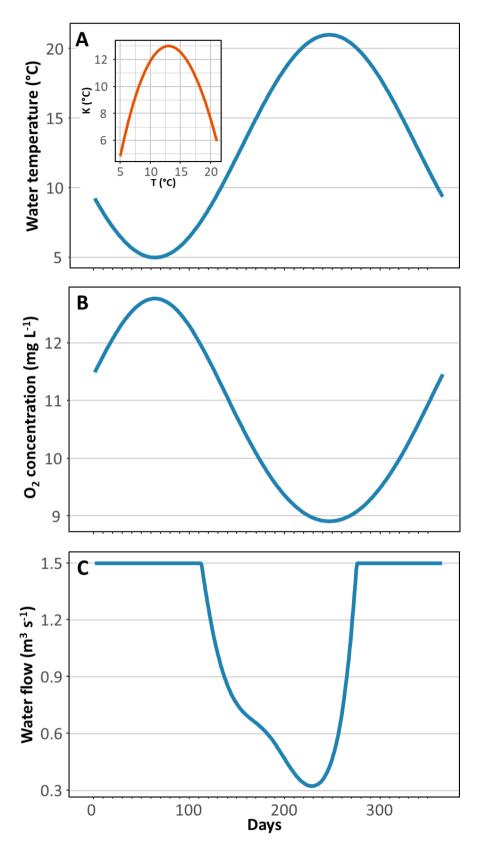
HYPOTHETICAL FARM



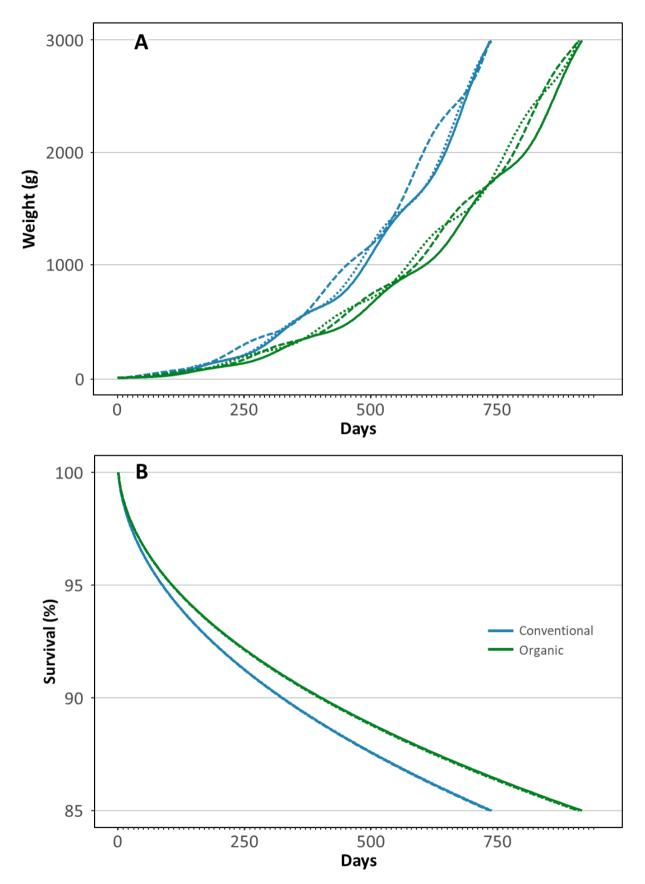
- Equipments
- Production (quota, market size...)
- Batches of fry (number, stocking dates)



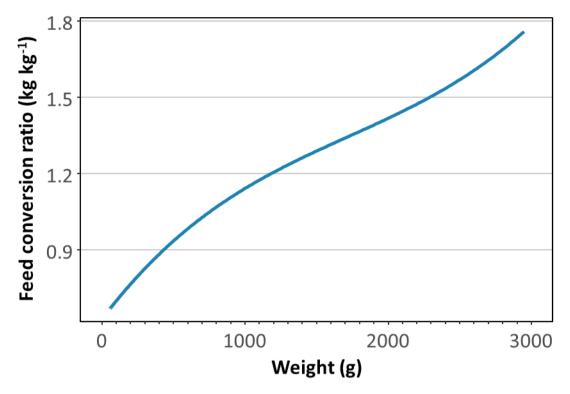
859 Figure 2



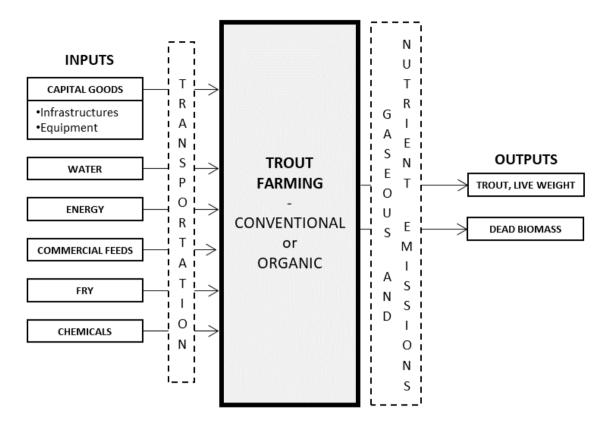
860 Figure 3



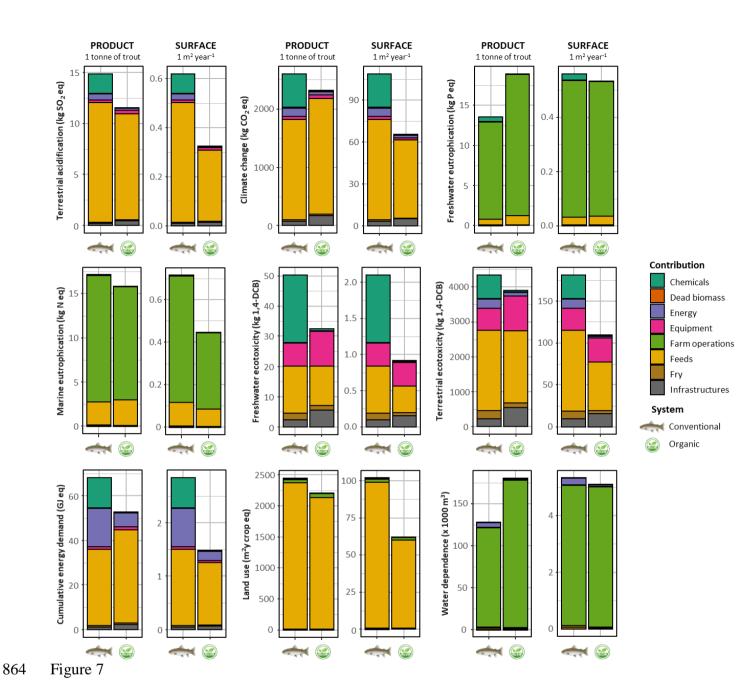
861 Figure 4

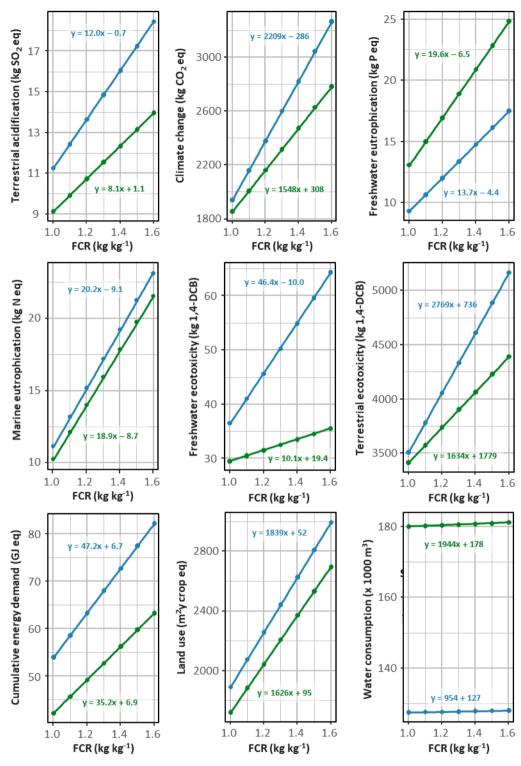


862 Figure 5



863 Figure 6





865 Figure 8

866 Table 1. Type of trout farms considered in the two different scenarios.

	Conventional	Organic
Production (t year ⁻¹)	300	203
Rearing duration (d)	737 ± 2	913 ± 4
FCR	1.3	1.3
Mortality rate (%)	15	15
Number of batches per year	3	3

867 FCR = Feed Conversion Ratio calculated as the ratio of feed intake to fish weight gain over one cycle

868 of production

	Conventional	Organic
Proteins (%)		
Feed 1	45	43
Feed 2	40	39
Feed 3	39	38
Feed 4	38	36
Lipids (%)		
Feed 1	21	21
Feed 2	23	24
Feed 3	27	26
Feed 4	30	28
Carbohydrates (%)		
Feed 1	12.0	13.0
Feed 2	13.9	14.0
Feed 3	12.8	13.6
Feed 4	12.8	11.4
Phosphorus (%)		
Feed 1	0.95	1.70
Feed 2	0.95	1.70
Feed 3	0.90	1.70
Feed 4	0.90	1.60

869 Table 2. Composition of the feeds used in the two different scenarios.

870

871 Table 3. Life Cycle Inventory for one year of production.

	Item	Unit	Conventional	Organic
INPUTS	Site surface	m ²	16000	16000
	Water	m ³	35785586	35785586
	Fry (10 g)			
	Triploid trout (\bigcirc_+)	u	120000	-
	Organic trout (\bigcirc/\bigcirc)	u	-	81000
	Feeds			
	feed_1/ feed_org_1	kg	3127	2120
	feed_2/ feed_org_2	kg	41192	29841
	feed_3/ feed_org_3	kg	120688	69479
	feed_4/ feed_org_4	kg	223868	151370
	Chemicals			
	Liquid oxygen	m ³	277036	-
	Antibiotics	kg	0.24	0.16
	Others	kg	4000	4000
	Electricity	kWh	427512	106440
	Infrastructures			
	60-m ² building	u	1	1
	80-m ² building	u	1	1
	100-m ³ raceways	u	12	12
	250-m ³ raceways	u	24	24
	Equipment			
	Feed storage silo	u	5	5
	Oxygen cone	u	2	-
	Oxygen tank	u	1	-
	Leaf screener	u	1	1
	Fish elevator	u	2	2
	Drum filter	u	1	1
	Electric generator	u	1	1
	Pumps	u	3	1
	Aerators	u	12	36
	PVC pipe	m	1500	1500
OUTPUTS	Trout at market size (3 kg)	kg	300478	202909
	Dead biomass (incinerated)	kg	9158	5990
	Water (back to river)	m ³	35785586	35785586
	Nitrogen (in river)	kg	14512	8793
	Phosphorus (in river)	kg	2254	2701
	COD (in river)	kg	62435	39631

872 u: unit; COD: Chemical Oxygen Demand; Transportation was included at each step when needed.

873 Table 4. Assumptions made to fill inventory gaps.

	Assumption(s)				
Wastewater treatment	We assumed that a sedimentation area is able to remove 20% of suspended N and P (Stewart et al., 2006)				
Lifespan of infrastructures and equipment	Adoption of the average lifespan (assuming only ordinary maintenance): equipment: 10-15 years; buildings and raceways: 30 years				
	The occupancy rates of the rearing struprocesses in the LCA:	The occupancy rates of the rearing structures were used as weights for these processes in the LCA:			
Rearing structures occupancy	Conventional production: 100-m ³ raceways: 62% 250-m ³ raceways: 45%	Organic production: 100-m ³ raceways: 70% 250-m ³ raceways: 87%			
Infrastructures weigh	Buildings: Walls: 0.15 m thick. Slab: 0.25 m thickFramework: 40 kg wood m-2Raceways: Walls: 0.15 m thick considering raceways of 1.5 m deep. Slab: 0.25 m thickConcrete density was considered equal to 2150 kg m-3Wood density was considered equal to 750 kg m-3				
Transport distances	Road distances were calculated from Google Maps; ocean distances (transport of aquafeed ingredients from South America to a French harbour) were assessed from shiptraffic.net				

874

875	Table 5.	Characteristics	of the	selected	impact	categories.
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Impact category	Abbreviation	Unit	Definition
Climate change potential	GWP	kg CO ₂ eq. to air	the contribution of greenhouse gases to global warming
Terrestrial acidification potential	ТАР	kg SO ₂ eq. to air kg	changes in acidity in the soil due to a change in acid deposition, which in turn is a consequence of changes in air emission of NO_x , NH_3 and SO_2
Freshwater eutrophication potential	FEP	kg P eq. to freshwater	a change in the levels of P in freshwater caused by emissions of nutrients into water and soil
Marine eutrophication potential	MEP	kg N eq. to freshwater	a change in the levels of N in marine water caused by emissions of nutrients into water and soil
Terrestrial ecotoxicity potential	TETP	kg 1,4-DCB eq. to soil	a change in the levels of toxic chemicals caused by emissions into the soil
Freshwater ecotoxicity potential	FETP	kg 1,4-DCB eq. to freshwater	a change in the levels of toxic chemicals caused by emissions into the water
Cumulative energy demand	CED	GJ eq.	the direct and indirect consumption of energy
Land use	LaU	m ² y crop eq.	the ground surface used directly (land occupied by ponds) and indirectly (land used to grow feed ingredients)
Water dependence	WD	m ³	the water flowing into the production system

876

	Global warming (kg CO ₂ eq.)	Acidification (kg SO ₂ eq.)	Eutrophication (kg PO ⁴⁻ eq.)	Terrestrial ecotoxicity (kg 1,4 DCB eq.)	Freshwater ecotoxicity (kg 1,4 DCB eq.)	Cumulative Energy Demand (GJ)
This study	2571	15	57	114	873	68
Literature						
Flow-through system	1157-3561	10-19	46-75	17-169	1290*	30-78
RAS	2043-13622	13-46	4-21	-	-	63**

877 Table 6. Comparison of the results assessed with the CML baseline method (Guinée, 2002) and Cumulative Energy Demand indicator (Frischknecht et

al., 2007) with literature data on conventional production systems. Impacts are scaled on 1 tonne of trout.

879 Values for flow-through systems were taken from eight studies (Aubin et al., 2009; Boissy et al., 2011; Chen et al., 2015; d'Orbcastel et al., 2009;

880 Dekamin et al., 2015; Maiolo et al., 2021; Papatryphon et al., 2004; Samuel-Fitwi et al., 2013) while values for RAS were taken from three studies

(d'Orbcastel et al., 2009; Dekamin et al., 2015; Samuel-Fitwi et al., 2013).

882 * Maiolo et al. (2021)

883 **d'Orbcastel et al. (2009)