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MSc BOOST, Mentor Project 2022 Plant -Oomycete Interactions

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Plant – Oomycete Interactions



❖ Oomycetes: Filamentous, fungus-like pathogens

- Responsible for important Crop losses
- Major constraint for conventional and soil-less agriculture

→ **Need for new methods of control that are adapted and environmentally friendly.**

❖ The IPO team objective :

Better understand the plant – oomycete interactions

- Study both the **plant** and the **pathogen**
- Characterize the molecular mechanisms and molecular dialog established throughout infection.
- Models :

Arabidopsis thaliana

Tomato

Marchantia polymorpha

Hyaloperonospora arabidopsidis

(specialist, downy mildew, leaves)

Phytophthora parasitica (polyphagous,
roots and leaves rot)

Phytophthora palmivora
(polyphagous, roots, leaves and fruits rot)



The IPO Team



Celso

Harald

Valérie

Carlotta

Marie-Line

Agnès

Enora

Cathy



IPO MENTOR DAYS FOR MSc BOOST FEBRUARY 2023

A

Group	Group Size	Firstname	Lastname	email
Group A	6			
		Shantanu	barve	shantanu.barve@etu.univ-cotedazur.fr
		Juan	duarte torres	juan.duarte-torres@etu.univ-cotedazur.fr
		Safae	mouatamid	safae.mouatamid@etu.univ-cotedazur.fr
		Sharmistha	aryal	sharmistha.aryal@etu.univ-cotedazur.fr
		Oumaima	chihani	oumaima.chihani@etu.univ-cotedazur.fr
		Youssra	sakkali	youssra.sakkali@etu.univ-cotedazur.fr
Group B	6			
		Michael	pinse	michael.pinse@etu.univ-cotedazur.fr
		Victoria	faivre	victoria.faivre@etu.univ-cotedazur.fr
		Tigran	satourov	tigran.satourov@etu.univ-cotedazur.fr
		Julien	dou	julien.dou@etu.univ-cotedazur.fr
		Mariam	el khatib	mariam.el-khatib@etu.univ-cotedazur.fr
		Xiaoxuan	Zhou	
Group C	6			
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		Mohamad Hasan	konyali	mohamad-hasan.konyali@etu.univ-cotedazur.fr
		Anna	vorobieva	anna.vorobieva@etu.univ-cotedazur.fr
		Emma	bustin	emma.bustin@etu.univ-cotedazur.fr
		Samuel Paulo	matsinhe	samuel-paulo.matsinhe@etu.univ-cotedazur.fr
		Mathieu	Brisson	

B

C

IPO MENTOR DAYS FOR MSc BOOST FEBRUARY 2022/2023

16 M1 students (+2 visiting) students divided in 3 groups A, B, C;

14 workpackages, 3 days (D1,...,D4,D5)

Monday 13th + Thursday 16th and Friday 17th

14 Workpackages

1-*Arabidopsis* seeds sterilization and sowing.

2-*Marchantia* observation, gemmae propagation.

3- Preparation for *Agrobacteria* infiltration in *Nicotiana*. 3bis- Infiltration.

4-*Phytophthora palmivora/parasitica* -Observing hyphae, sporangia, fixed zoospores... Propagation by transferring agar blocs in V8 media under laminar flow hood.

5-Sporulation/release zoospores/Encysting zoospores to count cells; make dilution.

6-*Arabidopsis* seedling inoculation with *P. parasitica*.

7-*Marchantia thalli* inoculation with *P. palmivora*.

8-Observe swimming zoospore and attraction.

9-Image aggregation on *Arabidopsis* roots, analyze/quantify area with ImageJ.

10-Embedding/sectioning of infected *Marchantia thalli*.

11-Observe fluorescent *P. palmivora* infecting *Marchantia* at the confocal microscope.

12-Phenotyping disease symptoms on *Marchantia* (area quantification with ImageJ).

13-Observation/phenotyping of infiltrated *Nicotiana* leaves.

14-Observation of *Arabidopsis* (WT vs *pskr1*) cotyledons infected w/ *Hyaloperonospora arabidopsidis*. Aniline blue staining of hyphae/haustoria + observation.

cancelled



IPO MENTOR DAYS FOR MSc BOOST FEBRUARY 2023

16 M1 students (+2 visiting) students divided in 3 groups A, B, C.
Over 3 days, Monday 13th + Thursday 16th and Friday 17th February

A 1, 2, 5, 6, 13

Monday 13 th	1-Arabidopsis seeds sterilization and sowing. 2-Marchantia observation, gemmae propagation.
	5-Sporulation/release zoospores/Encysting zoospores to count cells; make dilution.
	6-Arabidopsis seedling inoculation with <i>P. palmivora</i> .
Thursday 16 th	Article Discussion/Free time Thursday Morning
	13-Observation/Phenotyping of infiltrated <i>Nicotiana</i> leaves.
Friday 17 th	cancelled
	Article Restitution

B 3, 5, 7, 3bis, 13, 12

Monday 13 th	3- Preparation for <i>Agrobacteria</i> infiltration in <i>Nicotiana</i> .
	5-Sporulation/release zoospores/Encysting zoospores to count cells; make dilution.
	7-Marchantia thalli inoculation with <i>P. palmivora</i> .
	3bis- <i>Agrobacteria</i> infiltration.
Thursday 16 th	13-Observation/Phenotyping of infiltrated <i>Nicotiana</i> leaves.
	Article Discussion/Free time Thursday Afternoon
Friday 17 th	12-Observation/Phenotyping disease symptoms on <i>Marchantia</i> (area quantification with ImageJ).
	Article Restitution

C 4, 5, 8, 6, 9, 14

Monday 13 th	4- <i>Phytophthora palmivora</i> , observing hyphae, propagation by transferring agar blocs.
	Article Discussion/ Free time Monday Morning
Thursday 16 th	5-Sporulation/release zoospores/Encysting zoospores to count cells; make dilution.
	8-Observe swimming zoospore and attraction.
	6-Arabidopsis seedling inoculation with <i>P. palmivora</i> .
	9-Image aggregation on <i>Arabidopsis</i> roots, analyze/quantify area with ImageJ.
Friday 17 th	14-Observation of <i>Arabidopsis</i> (WT vs <i>pskr1</i>) infected w/ <i>H. arabidopsidis</i> . Aniline blue staining and observation.
	Article Restitution

Aurélien
Marie-Line
Carlotta
Agnès
Enora/Valérie



Aurélien

Marie-Line

Carlotta

Agnès

Enora/Valérie

DAY 1

lecture | (Aurélien) Plant model organisms, evolutionary developmental biology... D206

9h30-12h | Transgénèse

1-Arabidopsis seeds sterilization and sowing. A

2-Marchantia observation, gemmae propagation. A

D216?

3- Preparation for Agrobacteria infiltration in Nicotiana. B

D218

4-Phytophthora palmivora, observing hyphae, propagation by transferring agar blocs. C

Lunch break 12h-13h30

lecture | (Agnès) Introduction to Oomycete life cycle, classification, host plant variety, symptoms... D206

D218 A

5-Sporulation/release zoospores/Encysting zoospores to count cells; make dilution. B

and Transgénèse

6-Arabidopsis seedling inoculation with P. palmivora. A

or Transgénèse

7-Marchantia thalli inoculation with P. palmivora. B

3bis- Agrobacteria infiltration in Nicotiana. B

DAY 4

lecture | (Agnès) Signaling during the first events of infection : from zoospore attraction to penetration. D206

9h30-12h | D218 C

5-Sporulation/release zoospores/Encysting zoospores to count cells; make dilution. C

8-Observe swimming zoospore and attraction. C

6-Arabidopsis seedling inoculation with P. palmivora. C

13-Observation/phenotyping of infiltrated Nicotiana leaves. B

Zeiss Confocal

Lunch break 12h-13h30

lecture | (Carlotta) Zoospores: biology of a swimming cell. D206

D218 C

9-Image aggregation on Arabidopsis roots, analyze/quantify area with ImageJ. C

Zeiss confocal

13-Observation/Phenotyping of infiltrated Nicotiana leaves. A

DAY 5

lecture | D206

cancelled

9h30-12h | Transgénèse B

12-Observation/Phenotyping disease symptoms on Marchantia (area quantification with ImageJ). B

cancelled

Zeiss Confocal A

14-Observation of Arabidopsis (WT vs pskr1) cotyledons infected w/ H. arabidopsidis. Aniline blue staining and observation. C

D216 + Axioplan

Lunch break 12h-14h

D206 A B C

14h-17h | Article Restitution

BOOST
Biocontrol Solutions for Plant Health

Each group chooses 1 article from the following:

1- A Phytophthora Effector Suppresses Trans-Kingdom RNAi to Promote Disease Susceptibility

<https://doi.org/10.1016/j.chom.2018.11.007>

2C-The Phytophthora capsici RxLR effector CRISIS2 triggers cell death via suppressing plasma membrane H⁺-ATPase in the host plant

<https://doi.org/10.1093/jxb/erac500>

3A- Elicitin recognition confers enhanced resistance to Phytophthora infestans in potato

<https://www.nature.com/articles/nplants201534>

4B- Conserved Biochemical Defenses Underpin Host Responses to Oomycete Infection in an Early-Divergent Land Plant Lineage

<https://doi.org/10.1016/j.cub.2019.05.078>

5-Developmental Modulation of Root Cell Wall Architecture Confers Resistance to an Oomycete Pathogen

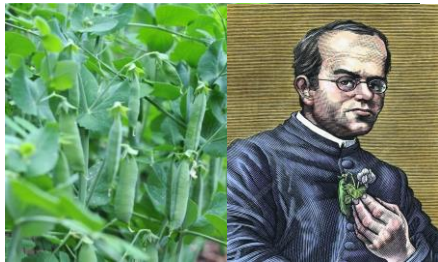
<https://doi.org/10.1016/j.cub.2020.08.011>

EVALUATION		Points (/20)	
Individual	<u>Participation to the Scientific Activities</u> (including punctuality, respect, curiosity, understanding, communication...)	4	
	<u>Overall quality of the oral presentation</u> -Quality and clarity of the oral presentation. -Attitude towards the audience (gestures, tone, expression, ...). -Quality of illustrations. -Respect of the speaking time.	4	
Collective (by group)	<u>Introduction</u> -The background information to understand the paper has been well delivered. -The main QUESTION the authors set out to answer in the published work was well introduced.	4	
	<u>Results</u> -The main findings are clearly presented at the level of scientific goals and experimental strategy. -Routine techniques are briefly exposed. -New techniques are sufficiently exposed without getting lost with details. -Clear & Correct conclusions are drawn from the presented data.	4	
	<u>Global Comprehension</u> Understanding the scientific context and interest of the study, the limitations as well.	4	

Model Organisms, what are they for?

• **Model organisms** are non-human species used in the lab to **help us understand a range of biological phenomena, with the hope that data, models and theories generated will be applicable to other organisms**, particularly those that are in some way more complex than the original.

• They should display particular **experimental advantages**. For example, they have **small physical and genomic sizes**; **low costs** to breed, maintain and transport; **short generation times and life cycles**; **high fertility rates**; and high mutation rates or high susceptibility to techniques for **genetic modification**. They may also occupy a **pivotal position in the evolutionary tree**...



Pea
- laws of inheritance

Amphibians
- developmental organizers

Drosophila
- chromosome theory
- developmental genetics

Mouse
- prime mammalian model for development and disease
- targeted genetics

Maize
- transposons
- epigenetics

Phages
- simplest life forms

Yeast
- basic eukaryotic cellular processes

Bacteria
- basic cellular processes

C. elegans
- cell death
- RNA interference

Zebrafish
- vertebrate developmental genetics

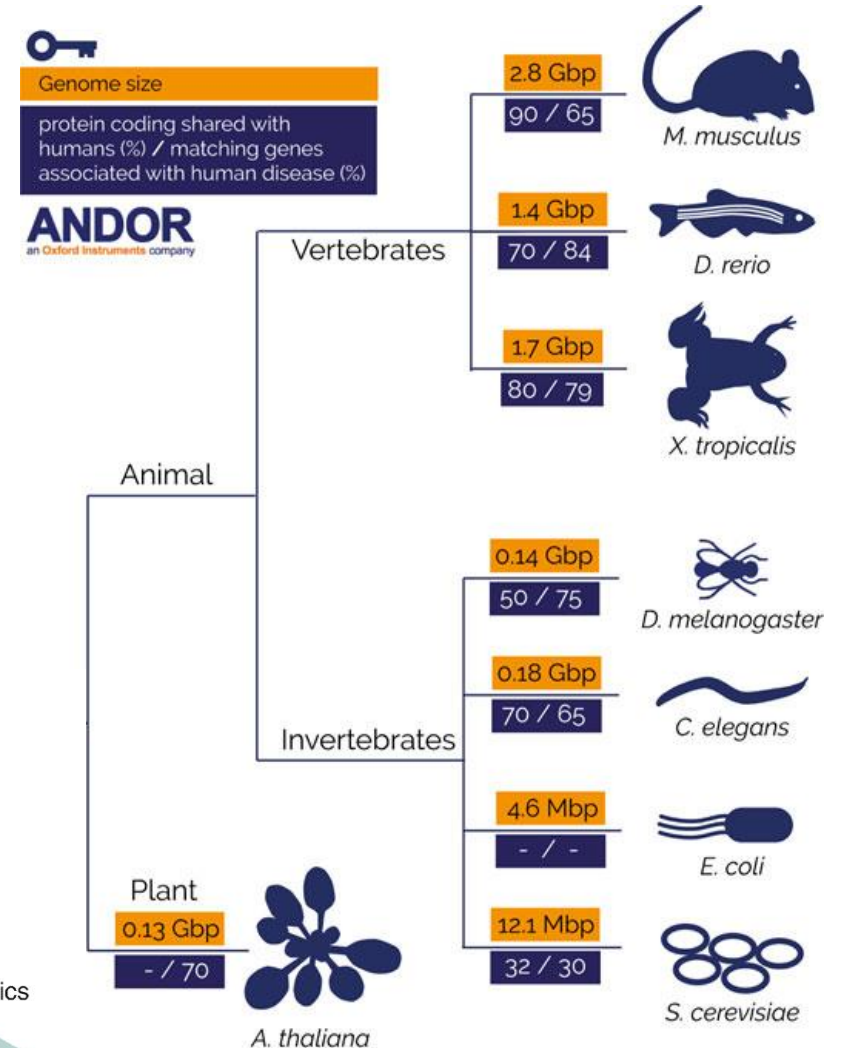
Arabidopsis
- plant developmental genetics

1850

1900

1950

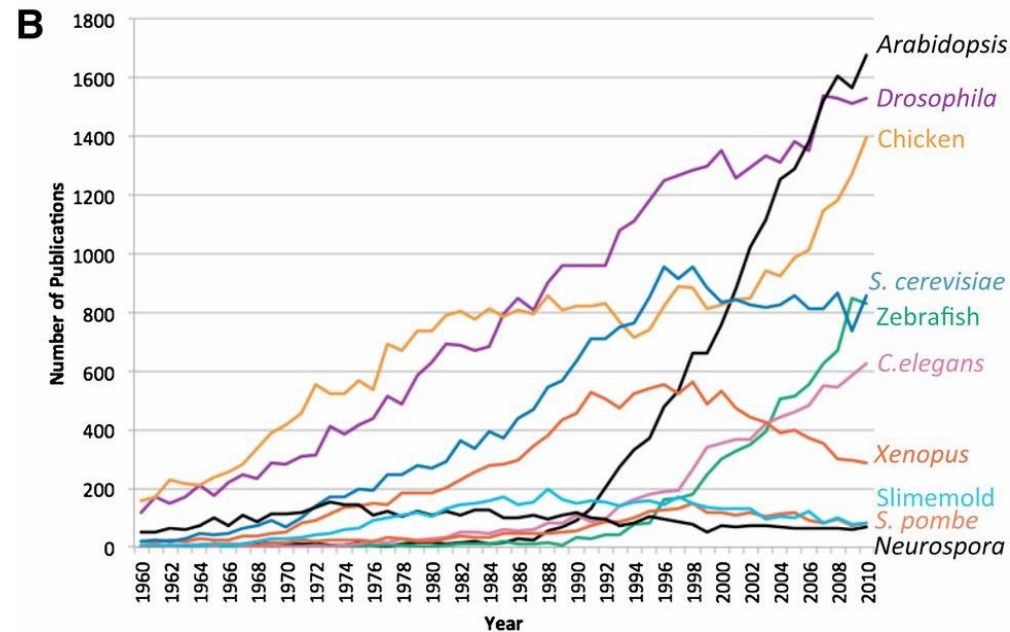
2000



Arabidopsis thaliana (Thale Cress): a weed, yes, but the Reference Plant Model of the last 4 decades!



- #The popularity of *A. thaliana* took off in the late 1980s, when researchers began to combine genetics with powerful molecular biology methods.
- #The *Arabidopsis* reference genome sequence was published as the first nuclear genome of a flowering plant in 2000.
- # Fostered enormous fundamental progress in our knowledge of the molecular principles of plant development, cell biology, metabolism, physiology, biotic and abiotic stress responses, genetics and epigenetics.



#Its uses continue to expand in the fields of systems biology and adaptation of molecular functions in natural environments.

#But it is only one plant species among 400,000! Can not apprehend alone the biodiversity of land plants in terms of ecosystems, architecture, biochemistry, reproductive systems...

Dietrich et al., 2014 Genetics. <https://doi.org/10.1534/genetics.114.169714>

Krämer, 2015. Elife. <https://doi.org/10.7554/eLife.06100>

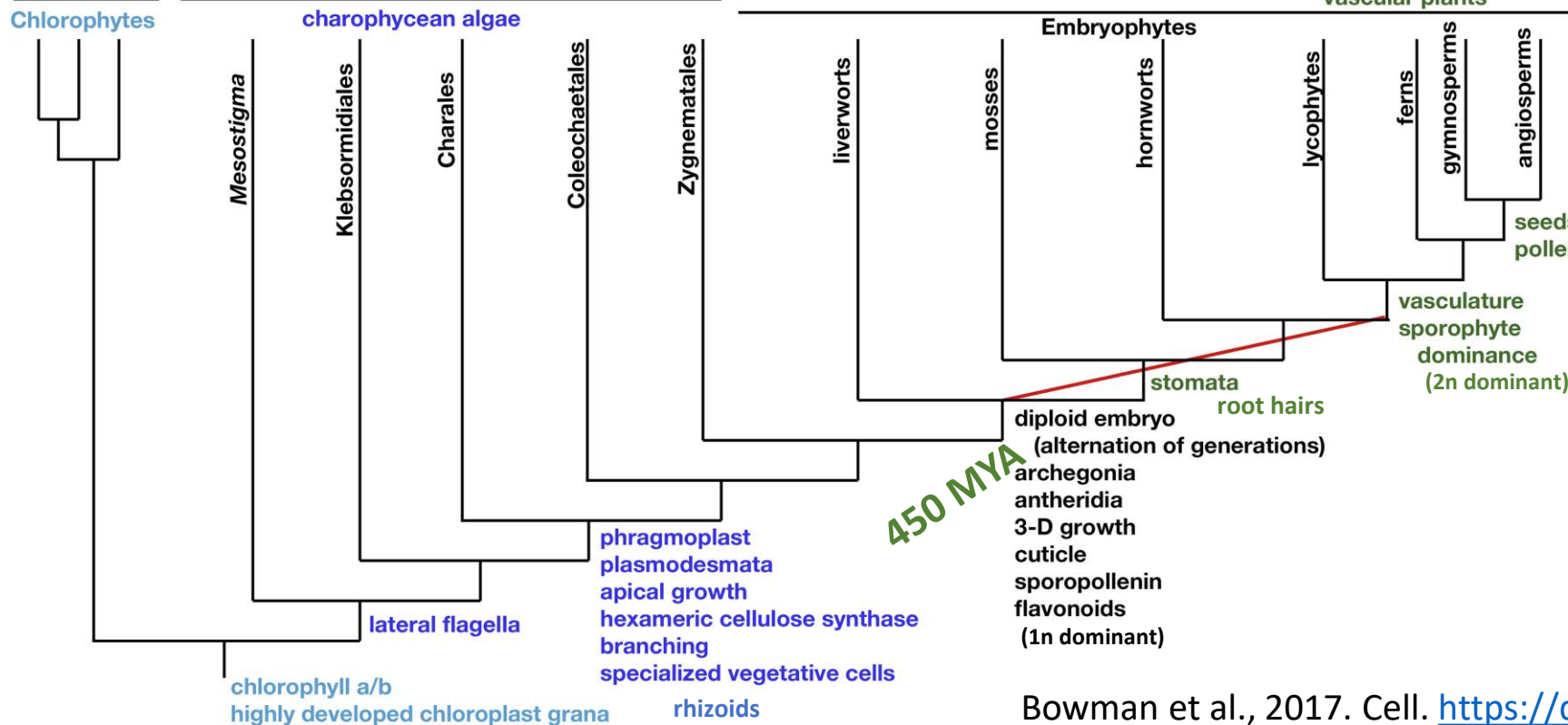
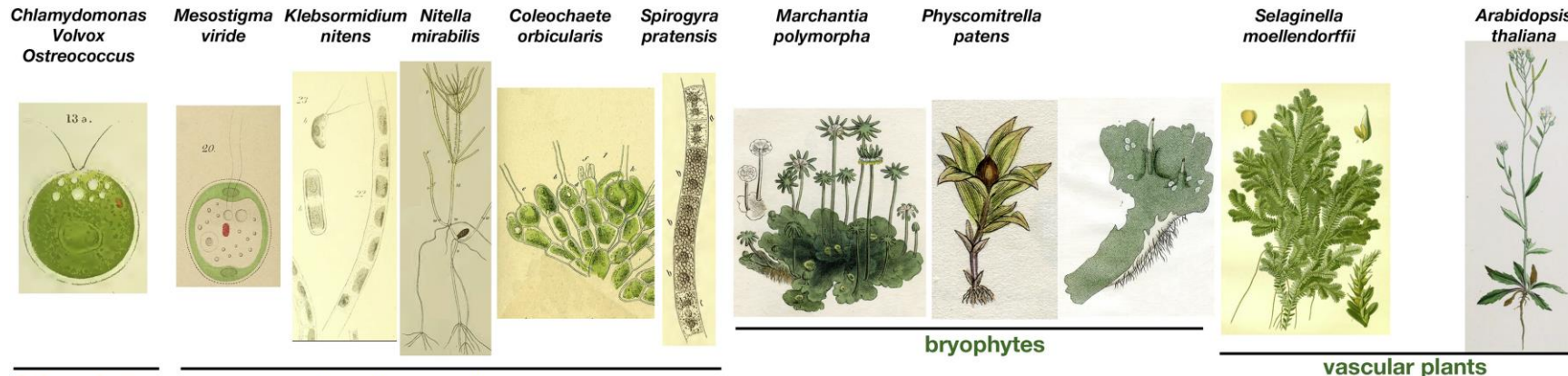
Chang et al., 2016. Cell. <https://doi.org/10.1016/j.cell.2016.08.031>

Diversity of Plant Model Organisms

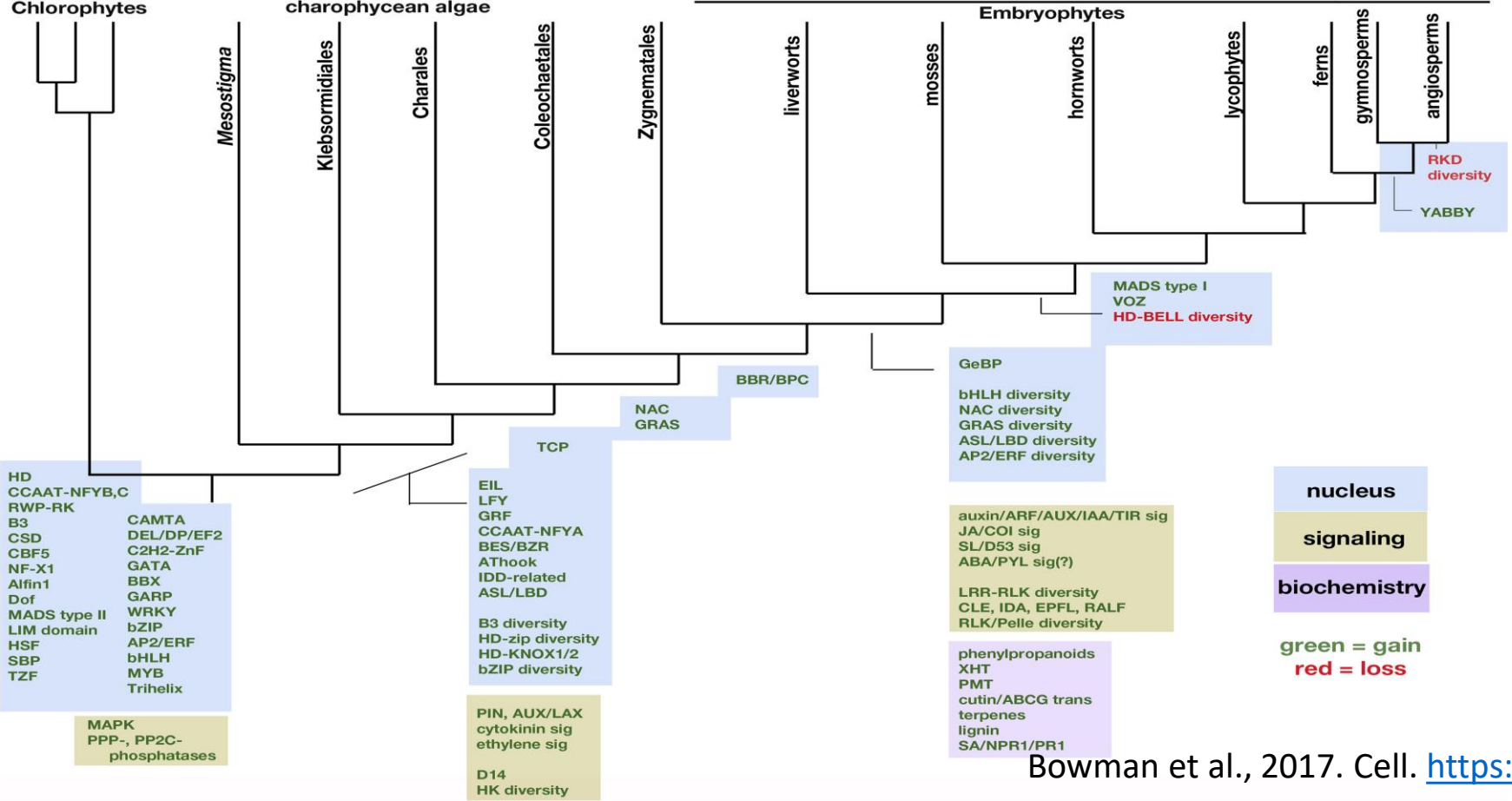
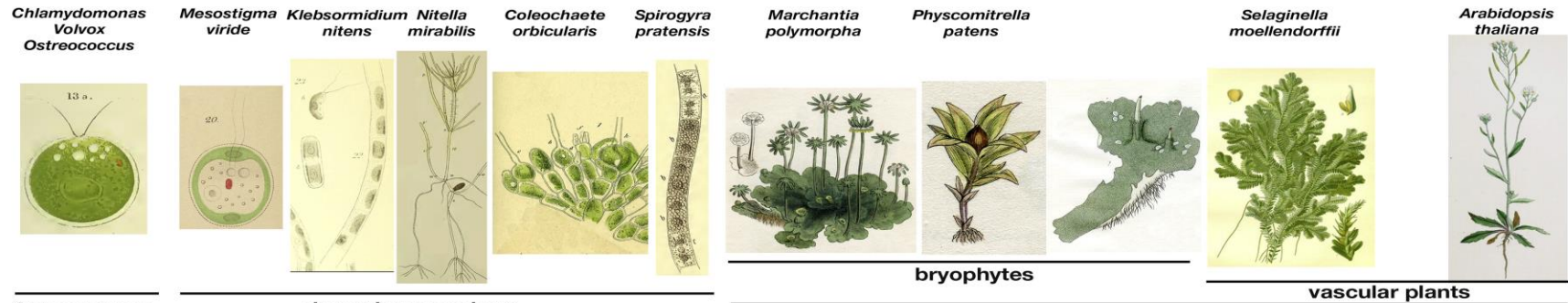
Classification	Family	Species Name	Properties
Angiosperm eudicot	Brassicaceae	Arabidopsis thaliana (mouse-ear cress)	predominant plant model system
		Arabidopsis halleri	heavy metal tolerance/accumulation
		Arabidopsis lyrata (lyrate rock cress)	self-incompatible A. thaliana relative
		Cardamine hirsute (bitter cress)	compound leaves
	Fabaceae	Lotus japonicas (birdsfoot trefoil)	nitrogen fixation
		Medicago truncatula (barrel medic)	nitrogen fixation
	Myrtaceae	Eucalyptus globulus (blue gum)	wood, lignification, biofuel
	Phrymaceae	Mimulus guttatus (seep monkeyflower)	ecological studies, flower evolution
	Rosaceae	Fragaria vesca (wild strawberry)	fruit development
		Prunus persica (peach)	fruit tree development
Salicaceae	Populus trichocarpa (black cottonwood)	wood	
Solanaceae	Solanum lycopersicum (tomato)	fruit ripening	
Vitaceae	Vitis vinifera (grape)	wine	
	Poaceae	Brachypodium distachyon	non-crop grass model
		Oryza sativa (rice)	major crop, grass model
		Setaria viridis (green foxtail)	non-crop grass
		Zea mays (maize)	genetics and development, major crop
Basal angiosperm	Amborellaceae	Amborella trichopoda	flowering plant evolution
Gymnosperm	Pinaceae	Picea abies/ Picea glauca (spruce)	wood
		Pinus taeda (loblolly pine)	wood
Pteridophyta	Azollaceae	Azolla filiculoides (water fern)	model fern, nitrogen fixation
Lycopodiophyta	Selaginellaceae	Selaginella moellendorffii (spikemoss)	clubmoss evolution
Anthocerotophyta	Anthocerotaceae	Antheroceros agresis (field hornwort)	hornwort evolution
	Funariaceae	Physcomitrella patens	moss evolution, moss development
Marchantiophyta	Marchantiaceae	Marchantia polymorpha	liverwort evolution
Charophyta	Klebsormidiaceae	Klebsormidium flaccidum	filamentous freshwater alga
Chlorophyta	Chlamydomonadaceae	Chlamydomonas reinhardtii	motile unicellular freshwater alga
	Volvocaceae	Volvox carteri	multicellular freshwater alga



Major Evolutionary Innovations of Charophycean Algae and Land Plants

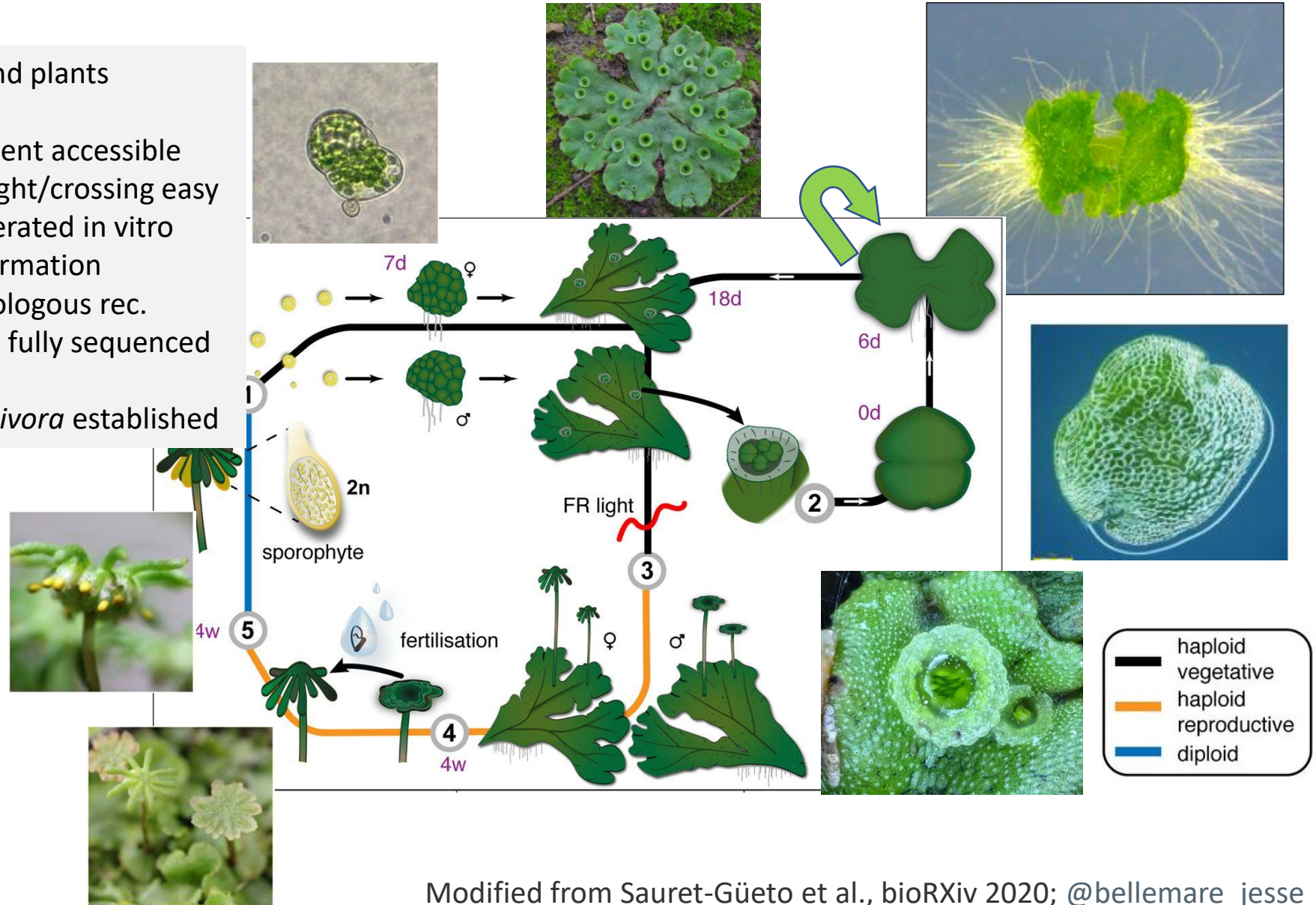


Major Evolutionary Innovations of Charophycean Algae and Land Plants



Marchantia polymorpha, the bryophyte model

- #Descendant of earliest land plants
- #Haploid genetics
- #Entire phase of development accessible
- #Gametes induced by FR light/crossing easy
- #Easily propagated /regenerated in vitro
- #High frequency of transformation
- #CRISPR technology /homologous rec.
- #Small and simple genome fully sequenced
- #Less genetic redundancy
- #Pathosystem with *P. palmivora* established





Aurélien

Marie-Line

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Agnès

Enora/Valérie

DAY 1

lecture | (Aurélien) Plant model organisms, evolutionary developmental biology... D206

9h30-12h | Transgénèse

1-*Arabidopsis* seeds sterilization and sowing. A

2-*Marchantia* observation, gemmae propagation. A

3- Preparation for *Agrobacteria* infiltration in *Nicotiana*. D216? B

4-*Phytophthora palmivora*, observing hyphae, propagation by transferring agar blocs. D218 C

Lunch break 12h-13h30

lecture | (Agnès) Introduction to Oomycete life cycle, classification, host plant variety, symptoms... D206

14h-17h | D218 A B

5-*Sporulation*/release zoospores/*Encysting* zoospores to count cells; make dilution. A B

and Transgénèse

6-*Arabidopsis* seedling inoculation with *P. palmivora*. A

or Transgénèse

7-*Marchantia thalli* inoculation with *P. palmivora*. B

3bis- *Agrobacteria* infiltration in *Nicotiana*.

DAY 4

lecture | (Agnès) Signaling during the first events of infection : from zoospore attraction to penetration. D206

9h30-12h | D218 C

5-*Sporulation*/release zoospores/*Encysting* zoospores to count cells; make dilution. C

8-Observe swimming zoospore and attraction. C

6-*Arabidopsis* seedling inoculation with *P. palmivora*. C

13-Observation/phenotyping of infiltrated *Nicotiana* leaves. Zeiss Confocal B

Lunch break 12h-13h30

lecture | (Carlotta) Zoospore mobility and attraction... D206

14h-17h | D218 C

9-Image aggregation on *Arabidopsis* roots, analyze/quantify area with *ImageJ*. C

13-Observation/Phenotyping of infiltrated *Nicotiana* leaves. Zeiss confocal A

DAY 5

lecture | D206

cancelled

9h30-12h | Transgénèse B

12-Observation/Phenotyping disease symptoms on *Marchantia* (area quantification with *ImageJ*). B

cancelled

Zeiss Confocal A

14-Observation of *Arabidopsis* (WT vs *pskr1*) cotyledons infected w/ *H. arabidopsidis*. Aniline blue staining and observation. C

D216 + Axioplan

Lunch break 12h-14h

14h-17h | D206 A B C

Article Restitution