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1 **Life history strategies of soil bacterial communities across global terrestrial biomes**

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16 **Abstract**

17 The life history strategies of soil microbes determine their metabolic potential and their response to  
18 environmental changes. Yet they remain poorly understood. Here we use shotgun metagenomes from  
19 terrestrial biomes to characterise overarching covariations of the genomic traits that captures dominant  
20 life history strategies in bacterial communities. The emerging patterns show a triangle of life history  
21 strategies shaped by two trait dimensions, supporting previous theoretical and isolate-based studies. The  
22 first dimension ranges from streamlined genomes with simple metabolisms to larger genomes and

23 expanded metabolic capacities. As metabolic capacities expand, bacteria communities increasingly  
24 differentiate along a second dimension that reflects a tradeoff between increasing capacities for  
25 environmental responsiveness or nutrient recycling. Random forest analyses shows that soil pH, C:N  
26 and precipitation patterns together drive the dominant life history strategy of soil bacteria communities  
27 and its biogeographic distribution. Our findings provide a trait-based framework to compare life history  
28 strategies of soil bacteria.

## 29 **Introduction**

30 Bacteria impact carbon (C) and nutrient cycling on a global scale<sup>1</sup>. Soil bacterial communities contain  
31 enormous, functionally uncharacterized genetic diversity<sup>2,3</sup> that hinders progress in predicting soil  
32 microbial responses to global change<sup>4,5</sup>. One approach to describe functional biodiversity is to collapse  
33 its complexity into one or more dimensions that capture the dominant associations and trade-offs  
34 between traits<sup>6-10</sup>. This multivariate trait space - or life history strategy scheme - provides a framework  
35 to compare broad organismal strategies<sup>6,8,10</sup>.

36 While the trait dimensions shaping plant life history strategies is now well established<sup>6</sup>, trait  
37 associations for soil microorganisms remain less clear. Initially, studies applied the ‘Competitor’, ‘Stress  
38 tolerant’, and ‘Ruderal’ (CSR) strategies proposed for plants<sup>7</sup> to soil bacteria<sup>1,11</sup>. This scheme  
39 emphasises trade-offs often observed between traits related to maximizing resource capture  
40 (Competitor, C), persisting under low resource and stressful condition (Stress tolerant, S), and  
41 responding rapidly to exploit growing window between disturbances (Ruderals, R)<sup>1,7</sup>. Building on the  
42 CSR scheme, Malik et al. (2020)<sup>12</sup> emphasised differences between microbial yield (Y), resource  
43 acquisition (A) and stress tolerance (S) traits as important for soil carbon cycling<sup>12</sup>. While these  
44 theoretical papers provide valuable hypotheses on which traits are probably central to soil microbial  
45 adaptation, no clear consensus has emerged on the trait dimensions that shape life history strategies of  
46 soil bacteria<sup>1,11,12</sup> (Extended Data table 1). Recently, Westoby and co-workers (2021) analysed bacterial  
47 cultures isolated from diverse habitats for genomic and phenotypic traits<sup>13</sup>. This analysis revealed a

48 primary dimension associated with metabolic versatility that was highly correlated with genome size. A  
49 secondary dimension separated differences in maximum growth rate and was correlated with variation  
50 in ribosomal gene copy number<sup>14</sup>. However, there is a lot of variation in how well bacterial cultures  
51 represent *in situ* community biodiversity<sup>15-17</sup>. Thus, it remains to be tested if the life history strategies  
52 of soil bacterial communities matches either the theoretical or culture-based predictions of key trait  
53 dimensions.

54         One advantage of studying the traits of microorganisms over those of larger organisms is the  
55 ease of which collections of their traits can be measured on the community level. Community aggregated  
56 traits (CATs)<sup>18</sup> represent the average functional profile of the community emerging from the  
57 combination of organisms' traits and community composition (similar to the idea of community-  
58 weighted means of traits proposed for plants)<sup>19,20</sup>. Hence, it is important to note, that while suggestive,  
59 such CAT patterns do not directly inform on the within-organism tradeoffs. Nevertheless, CATs  
60 described using metagenomic sequences offer a way to characterize shifts in the organismal strategies  
61 dominating bacterial communities *in situ* (eg. <sup>21,22</sup>) and thus offer an approach to test theoretical life  
62 history strategy schemes to *in situ* microbial communities. In addition, information on the dominant  
63 strategy in a bacterial community might be used to predict the response to environmental changes of this  
64 key group for global biogeochemical cycles<sup>4,18</sup>. Elucidating the trait dimensions that shape the dominant  
65 life history strategies of soil bacteria would thus provide a framework for comparing soil bacterial  
66 communities and developing generic predictions in soil microbial ecology<sup>14</sup>.

67

68 In this study, we used a global dataset of soil metagenomic sequences from major biomes to quantify  
69 key trait dimensions of soil bacterial communities. We then identified primary environmental factors  
70 partitioning the trait dimensions and projected the global biogeography. Finally, we compared the  
71 emergent life history strategies with theoretical and culture-based predictions.

## 72 **Results**

73 *The trait dimensions of soil bacterial communities*

74 Using a multi-table co-inertia analysis (MCOA), we found that two dimensions captured half of the  
75 overall variation in metagenomic community aggregated traits (CATs). MCOA1 and MCOA 2 captured  
76 29% and 21% of metagenomic trait variation (Figure 1 and Extended Data Figure 1), while MCOA 3  
77 and MCOA4 explained 16% and 10% of this variation, respectively (Extended Data Figure 1 and 2).  
78 The MCOA revealed the most important associations between traits (Figure 1-2) including traits  
79 previously associated with life history strategies (Figure 1-3).

80 Average genome size had the highest contribution to MCOA1 (Figure 1A) with a  $R^2$  of 0.64 for the  
81 positive correlation between average genome size and MCOA1 (Extended Data Figure 3A). Mapping  
82 coverage decreased along this dimension (Extended Data Figure 4). The lower end of this dimension  
83 was characterised by bacterial communities with higher relative abundance of genes for primary  
84 metabolism (ie. essential process for survival and growth) and C acquisition machinery (Figure 1). In  
85 these communities, carbon acquisition enzymes involved in depolymerization of oligosaccharides were  
86 favored over enzymes targeting polysaccharides. This oligosaccharide-degradation enzyme class was  
87 dominated by the beta-glucosidases GH1, GH2 and GH3 CAZy families. Finally, chaperones were  
88 overrepresented. Thus, the lower end of MCOA1 were defined by communities with a streamlined  
89 metabolism (Figure 2).

90 The upper end of MCOA1 defined bacterial communities with a large genome and more complex  
91 metabolism and resource acquisition strategies (Figure 1-2). The enriched genes allowed for degradation  
92 of complex polysaccharides from fungi, animals and plant lignin. There was also a gene  
93 overrepresentation for direct plant pathogenic interactions and negative interactions with other  
94 microorganisms. Finally, communities carried a higher proportion of genes encoding for EPS  
95 production, Dormancy and Sporulation, membrane, and DNA repair (Figure 1-2). These functions were  
96 generally present in lower relative abundance in communities with small genomes at the opposite end

97 of MCOA1. Thus, the first trait dimension captured functional variation associated with genome size  
98 and expanded *metabolic capacities* (Figure 2).

99 Bacterial communities differentiated along a second dimension (MCOA2) but only when they increased  
100 their *metabolic capacities* along the first trait dimension (MCOA1), shaping a triangle (Figure 2). This  
101 distribution indicated that bacteria communities with low *metabolic capacities* and small average  
102 genome size are constrained along the second dimension. The MCOA2 separated communities  
103 according to genomic traits for *environmental responsiveness* and *nutrient recycling* (Figure 2).  
104 Communities associated with the lower end of MCOA2 were enriched in mineral and organic N and P  
105 assimilation genes (Figure 1-2). Furthermore, there were also higher relative frequencies of genes  
106 encoding for bacterial necromass degradation including peptidoglycan. Communities at the upper end  
107 of MCOA2 were defined by an ability to respond to a complex set of environmental cues. This was  
108 manifested by an increased presence of genes encoding for activity regulation, resistance to  
109 environmental stress, foraging of beneficial conditions, fast growth (*rrn* copy), and building and  
110 repairing the cell membrane (Figure 1-2). The communities were also enriched in genes encoding for  
111 carbohydrates metabolism of simple substrates like starch, glycogen, and oligosaccharides. Thus, the  
112 second trait dimension captured a gradient in the average environmental responsiveness that was  
113 positively associated with a specialisation in simple carbon substrate metabolism and negatively with  
114 nutrient assimilation and recycling capacities (Figure 2).

#### 115 *Drivers of the trait dimensions*

116 Using random forest analyses, we next found that common soil environmental factors distributed the  
117 soil bacterial community along global trait dimensions. Random forest models based on soil pH,  
118 precipitation and C:N could predict most of the variation in MCOA1 and MCOA2 with a  $R^2$  of 0.80  
119 and 0.58, respectively (Extended Data Figure 5). Mean decrease in mean square error (%MSE) and  $R$   
120 squared calculated based on a ten-fold cross-validation of the random forests indicated that soil pH and  
121 annual precipitation are the most important predictors for both MCOA1 and MCOA2. However, the two  
122 dimensions showed different response patterns to these variables, with MCOA1 decreasing with soil pH

123 but increasing with annual precipitation whereas MCOA2 decreased with both soil pH and annual  
124 precipitation, leading to unique position along MCOA1 and MCOA2 depending on the combination of  
125 pH and annual precipitation (Figure 3-4). MCOA1 and MCOA2 were also driven by precipitation  
126 seasonality whereas soil C:N controls only MCOA1 (Figure 3-4, Extended Data Figure 5). Next, we  
127 projected the global variation in the trait dimensions using these random forests (Figure 4 B and D) and  
128 global soil and climate databases. It is worth noting that this broad spatial resolution map, using  
129 averaged conditions across large spatial units, showed high consistency with values observed locally in  
130 our samples (Extended Data Figure 6). Thus, the identified trait dimensions showed a clear global  
131 biogeography.

132 The first trait dimension (MCOA1) mainly separated arid, alkaline regions from more acidic and wet  
133 ones. More precisely, bacterial communities characterised by a small genome size (i.e., low MCOA1  
134 value) were enriched under neutral to alkaline pH, low C:N, low annual precipitation but high  
135 precipitation seasonality (Figure 4A). Conversely, communities with larger genome sizes (high MCOA1  
136 value), were found in more acidic soils as well as soil with higher C:N and climate with elevated stable  
137 precipitation (Figure 4A). Globally, these environmental controls predicted low MCOA1 coordinates ( $<$   
138  $-1$ ) under arid and semi-arid climates at tropical and subtropical latitude as well as in the steppe zones  
139 of central Asia and North America (Figure 4B). Conversely, high MCOA1 coordinates ( $>1$ ) were seen  
140 in equatorial forests as well as some temperate zones in northern Europe, Western Canada, New Zealand  
141 and south Chile. Steep MCOA1 gradients were estimated to occur in regions separating arid and wet  
142 zones and medium coordinates ( $-1 < \text{MCOA dimension 1} < 1$ ) also covered most of temperate and high  
143 latitudinal regions (Figure 4B).

144 The second trait dimension (MCOA2) separated regions with high but stable precipitation from places  
145 with more seasonal climate and extremely acidic soils. The lower end of MCOA2 covered most high  
146 precipitation regions ( $>2500\text{mm}$ ) including equatorial zones of South-America and Asia and wet Europe  
147 and North America temperate zones. Medium-high coordinates ( $0 < \text{MCOA2} < 1$ ) covered most of the  
148 globe, characterising all tropical-dry, semi-arid and subarctic regions. The projection of this dimension

149 (Figure 4D) predicts very high coordinates (MCOA2 >1) under limited regions of subtropical and high  
150 latitudes combining low annual precipitation (<1000mm) and very acidic pH (<4).

151 Finally, we found that trait differences (defined based on euclidian distances along the two first  
152 dimensions of the MCOA) were significantly correlated with Unifrac phylogenetic distances ( $R^2=0.32$ ,  
153 Extended Data Figure 7). Communities with average genome size below its median values depicted a  
154 correlation between trait and phylogenetic distances significantly steeper (slope difference:  $p=0.00116$ )  
155 and tighter ( $R^2=0.46$ ) compared to communities with larger genomes ( $R^2=0.15$ , Extended Data Figure  
156 7).

## 157 **Discussion**

158 Our study describes two dominant dimensions of community aggregated traits variation across soil  
159 bacteria communities (Figure 2-3). In this trait space, communities are constrained in a triangle of three  
160 opposing life history strategies: low metabolic capacities; metabolic capacities expanded for  
161 environmental responsiveness; metabolic capacities expanded for nutrient recycling. These life history  
162 strategies incorporates traits previously identified as CSR strategies<sup>1,11,12</sup> (Extended Data Table 1).  
163 Moreover, it fits into a triangle like the original CSR model<sup>7,23</sup> (Figure 2-3) which suggests that the  
164 constraints on bacterial strategies might scale up to community level. Also consistent with CSR theory,  
165 both trait dimensions of our study capture competitor traits that tradeoff with traits of the other strategies.  
166 However, while one strategy generally dominates the traits of each end of the trait dimensions, our  
167 aggregated profiles often combine traits that had been associated with different strategies. In particular,  
168 one or more stress tolerance traits are part of all profiles (Figure 2-3). We hypothesise that these  
169 combinations indicate either that the communities are composed of taxa with different strategies or that  
170 the majority of bacteria living in soil need stress tolerant traits to survive in this challenging  
171 environment.

172 Bacteria with streamlined metabolism dominate the low end of the *metabolic capacity* dimension. The  
173 genomic traits of these bacterial communities with small average genome size have only few matches



174 with previous description of stress tolerance strategy (Extended Data Table 1)<sup>1,11,12</sup>. However, the clear  
175 association to arid biomes that we observed suggests that the streamlined bacteria are associated with  
176 stress tolerance strategy. This is consistent with recent studies showing that genome streamlining can  
177 play a role in adaptation to environmental stressful conditions (eg.<sup>24,25</sup>). In particular, Liu et al. (2023)  
178 used a joint species distribution model to show that soil bacteria with small genomes are selected under  
179 arid environments, as seen here. Moreover, these streamlined communities were associated with some  
180 low environmental constraints on resource acquisition (low soil C:N and pH near neutrality as observed  
181 in <sup>26</sup>) that might also reduce fitness benefits for gaining new capabilities<sup>27</sup>. Thus, genome streamlining  
182 and associated change in gene frequency might be central in the soil bacteria stress tolerance, especially  
183 in arid biomes.

184 Cells with larger genomes and a more complex metabolism dominate the other end of the *metabolic*  
185 *capacity* dimension. The associated variation in the functional gene frequency that we observed is also  
186 consistent with previous studies reporting that genome expansion in free-living bacteria is driven by  
187 gene additions encoding for new metabolic capabilities or regulation<sup>14,28</sup>. Large genomes, high catabolic  
188 diversity, and antibiotic resistance genes observed for this life history strategy were previously attributed  
189 to a competitor strategy (Extended Data Table 1)<sup>1,11</sup>. This supports the idea that complex substrates  
190 acquisition is a key trait of competitors as suggested by Malik et al. (2020). Consistent with competitor  
191 traits, these attributes are favoured under stable and wet climates, that reduce the benefits of desiccation  
192 stress traits and possibly leading to intense resource competition<sup>7</sup>. We also detected an enrichment in  
193 traits associated with sporulation and exopolysaccharides production, two traits often associated with  
194 stress tolerance or ruderality (Extended Data Table 1) that might also improve tolerance to antimicrobial  
195 compounds or nutritional constraints for such competitor profile<sup>29,30</sup>. Together, the first trait dimension  
196 appears to represent a gradient from stress tolerant communities with small genomes to communities  
197 dominated by bacteria with increased *metabolic capacities* associated with other strategies, especially  
198 competitors.

199 When average genome size increases, bacteria communities differentiate along the second dimension  
200 with opposing profiles of either increased capacities for *environmental responsiveness* or for *nutrient*  
201 *recycling*. At the high end of this dimension, communities with high *environmental responsiveness*  
202 shared numerous genomic features tied to both the ruderal and stress tolerant strategies (Extended Data  
203 Table 1). This includes traits to resist stress, sensing favourable environmental conditions, activate fast  
204 growth, and C acquisition. The reduced and fluctuating precipitation patterns associated with this profile  
205 are also consistent with original descriptions of these strategies<sup>1,7</sup>. At the opposite end of this second  
206 dimension, bacteria specialised in *nutrient recycling* show a resource acquisition strategy with a high  
207 number of transporters and bacterial biomass (Peptidoglycan) recycling and a higher investment towards  
208 nitrogen and phosphorus metabolism compared to carbon metabolism. Microbial mineralisation activity  
209 and biomass turnover release nutrients and necromass into soil that this profile seems optimised to  
210 recycle. Such traits might reflect a strategy that emphasises resource use efficiency and increased  
211 competitiveness for nutrients<sup>11,12</sup>. Further, the environmental parameters associated with this life history  
212 strategy (medium-low pH, high precipitation and low seasonality) are the most favourable for resource  
213 acquisition<sup>31</sup>, biomass turnover and yield<sup>32,33</sup>, reinforcing potential selection for competitor traits<sup>7</sup>. In  
214 summary, the second trait dimension reflects communities with increased metabolic capacities  
215 associated with either a combination of stress tolerance and ruderal traits that maximise their  
216 responsiveness or a reinforcement of competitor traits that favour nutrient recycling.

217 Overall, our dimension of *metabolic capacities* matches the versatility dimension described by Westoby  
218 et al. (2021) across cultured bacterial taxa, with both studies supporting that genome size plays a central  
219 role in differentiating bacteria strategies. Our dimension opposing *environmental responsiveness* and  
220 *nutrient recycling* also shows some consistencies with the second trait-dimension described by Westoby  
221 et al. (2021) capturing a rate-yield tradeoff, with rrn copy number as principal trait. Indeed, as discussed  
222 above, the traits of the *nutrient recycling* profile might favour growth yield, and high *environmental*  
223 *responsiveness* is associated with higher rrn copy number. However, these variations of rrn copy  
224 numbers have only a limited importance in the second trait dimension of our study, contrasting with the

225 observations of Westoby et al. (2021) for cultured bacteria from diverse habitats. This could be  
226 explained by the constraint range of this trait in soil. Indeed, variation in average rrn copy number  
227 observed across communities in our study is highly constrained (1 to 1.5 copies, Extended Data Figure  
228 3). These observations are consistent with Gao and Wu (2018) reporting that most soil bacteria have  
229 less than 2 rrn copies, whereas bacteria from other environments can have up to 15 copies<sup>34</sup>. Further,  
230 variation in the average rrn copy number of whole communities will be more constrained than variation  
231 across individual isolates within the community; indeed, some bacteria with more copies might be  
232 present in the soil community, with their populations increasing during resource flushes (eg.<sup>35</sup>). In the  
233 oligotrophic environment of soil, our results suggest that increased capacity to recycle resources  
234 efficiently, to sense favourable conditions and to survive or escape stressful ones represent more  
235 common adaptations for bacteria than growing more rapidly. Investigating the variation of these traits  
236 across taxa in soil and their distribution within communities represents a challenging, but fascinating  
237 perspective to disentangle how the trait dimensions across taxa scales up to the community level.  
238 Overall, life history strategies of soil bacteria that we described using aggregated traits at the community  
239 level show some important consistencies with life history strategies described across bacterial taxa from  
240 various habitats, but also highlights some specificities and challenges associated with soil environment.

241 Soil bacteria remain poorly characterised with a limited number of reference genomes and gene  
242 functional characterization<sup>36,37</sup>. This reduces annotation coverage of metagenomic data and can limit  
243 analysis conclusions. In our study, the proportion of reads annotated (between 5 and 15% depending on  
244 the database) were in the range of what is commonly obtained from soil metagenomes<sup>38</sup>. Our usage of  
245 stringent quality filtering criteria in the annotation<sup>2</sup> also reduced the annotation coverage but increased  
246 annotation confidence. Finally, the proportion of unannotated reads is increased by the sequencing error  
247 and our usage of short read sequencing technology and read-based profiling (as opposed to assembly  
248 based profiling with better annotation but very limited representativity of the community). Our  
249 annotation coverage also showed a decrease with genome sizes, as reported across taxa<sup>36,37</sup>. However,  
250 unannotated genes likely belong to accessory genes and not to core metabolism that are well represented

251 in current databases<sup>37</sup>. Thus, we can expect that increased annotation of large genomes would have  
252 accentuated evidence for our conclusion that our first trait dimension captured an increase in metabolic  
253 capacities. Overall, our trait dimensions are expected to capture at least the functional variations  
254 associated with core metabolism and provide some first elements about functional genes associated with  
255 expansion of metabolic capacities.

256 We showed that communities with similar life history strategies tend to be phylogenetically closer,  
257 supporting a certain phylogenetic conservatism of the genomic traits shaping life history strategies<sup>39</sup>.  
258 However, this relationship weakens as genome size and metabolic capacities expand (Expanded Data  
259 Fig 7). This suggests that metabolic expansion during different evolutionary histories can converge to  
260 similar life history strategy<sup>40</sup>. Hence, phylogenetic distance become a poorer predictor of difference in  
261 life history strategies for soil bacterial communities with large genomes.

262 The biogeography of dominant life history strategies in soil bacterial communities is mainly driven by  
263 the combinations of soil pH and precipitation patterns across the globe. These environmental factors  
264 impact stress and competition intensity for soil bacteria, either through direct effect on their physiology  
265 and interaction<sup>41-43</sup> or indirectly through their modification of abiotic (eg. solubilization of toxic ions  
266 Al<sup>3+</sup>) and biotic (eg. plant and fungal communities) characteristics of the ecosystem<sup>44-46</sup>. The  
267 environmental distribution of the life history strategies suggests that bacteria expand their metabolic  
268 capacities to deal with conditions associated with increasing soil acidity and annual precipitation until a  
269 certain level (Figure 3). Then, expansion of *metabolic capacities* increases either *environmental*  
270 *responsiveness* to survive under more extreme pH and fluctuating precipitation or *nutrient recycling* to  
271 be competitive under higher precipitation levels. These global effects of pH and precipitation are  
272 consistent with previous studies of soil bacteria biogeography<sup>3,26,47</sup> and provide some new information  
273 on the traits associated with these environmental factors.

274 Our global projection (Figure 3B and D) aims at giving a picture of the general biogeographic patterns  
275 in the functional profiles of soil bacterial communities. However, it is important to note that

276 transposition of our trait dimensions at local scale will need further investigation. Values predicted for  
277 these broad resolution maps can be dissociated from the local situation if its conditions highly differ  
278 from the regional mean (Sup Figure 8) and should be used with caution. Despite outstanding issues that  
279 remain open, our study demonstrates how metagenomic approaches can provide substantial advance in  
280 our understanding of microbial community functioning. Altogether, our results suggest that land use and  
281 climate changes impacting soil pH and precipitation gradients at biogeographic scale might be central  
282 in shaping future functional potential of soil bacterial communities and thus global biogeochemical  
283 cycles.

## 284 **Methods**

### 285 *Soil sampling and characteristics*

286 We analysed a global dataset of 128 metagenomes each from unique soil samples distributed across  
287 continents and latitude (Extended Data Figure 8)<sup>2</sup>. We selected this dataset for our analysis because of  
288 its coverage and its use of a highly standardised protocol that: 1) sampled top-soils in spatially  
289 independent sites across the globe selected to represent all the most important vegetation types; 2)  
290 analysed soil chemistry and metagenomes<sup>2</sup>. All samples were processed using similar standardised  
291 protocols for their chemistry (carbon, nitrogen, phosphorus content and pH<sub>H2O</sub>) and metagenome (See<sup>2</sup>)  
292 for protocol details). We checked the global environmental coverage by comparing variation of the main  
293 environmental variables (mean annual temperature (MAT), mean annual precipitation (MAP), soil pH  
294 and net primary productivity (NPP)) in our dataset with global variation from the Atlas of the Biosphere  
295 (<https://nelson.wisc.edu/sage/data-and-models/atlas/maps.php>). This showed an almost complete global  
296 coverage, with only extreme MAT of very high latitude (below -11.33°C) and Sahelian Africa (above  
297 MAT 27.97°C) as well as very high pH (higher than 7.76) characterising some parts of North Africa,  
298 West Asia and Himalaya missing in our dataset (Extended Data Figure 8). As far as we know, when we  
299 conducted this analysis, this dataset was the only available with such precise characterization of soil  
300 environment done on the same sample as shotgun metagenomic analysis, making this dataset the most

301 robust for our objective to assess environmental drivers of metagenomic profiles. Nevertheless, potential  
302 to extend environmental range by adding all (excluding agricultural and contaminated) soil  
303 metagenomes available (accession date January 28 2021) from the main sequence repositories MG-  
304 RAST<sup>48</sup> and IMG:M<sup>49</sup> was also tested. This indicated that adding these data would not have extended  
305 environmental range (excepted a few samples from very cold sites with mean annual temperature lower  
306 than -11.5°C available on MG-RAST) and this would have greatly decreased precision of soil properties  
307 characterization (Extended Data Figure 9).

#### 308 *Metagenomic and amplicon sequencing data*

309 DNA extraction, sequencing (Illumina with RTA Version 1.18.54 and bcl2fastq v1.8.4), trimming and  
310 mapping approaches are detailed in Bahram et al. (2018). In this study, four community aggregated trait  
311 databases were built, corresponding to metagenomic reads mapping on different functional annotation  
312 systems by Bahram et al. (2018). An additional database was made for this study with genomic traits  
313 previously associated with bacterial life history strategies (See details below). Data from 16S rRNA  
314 gene amplicon sequencing were also used to characterise phylogenetic distances between bacterial  
315 communities using the Unifrac metric<sup>50</sup>

#### 316 *Bacterial community aggregated trait calculation*

317 Bahram et al. (2018) mapped reads to the functional databases (KEGG, eggNOG and CAZy). Data were  
318 aggregated at the (1) pathway (KEGG), (2) functional categories (eggNOG) levels, (3) SEED functional  
319 modules and (4). Glycolysis Hydrolases (GH) and Auxiliary Activities (AA) gene families from  
320 CAZy<sup>51</sup>. All read mapping was done competitively against both prokaryotic and eukaryotic functional  
321 databases and best bit score in the alignment and the taxonomic annotation was used to retrieve only  
322 reads annotated as bacteria.

323 We used output data from these four annotation processes to provide complementary classification of  
324 functional genes (e.g. eggNOG categories include Motility, Cell envelopes and Defense which are not  
325 included in SEED whereas SEED classes include Dormancy and Sporulation, Stress response,

326 Virulence, Carbon, Nitrogen and Phosphorus metabolism which are not included in eggNOG). The  
327 eggNOG annotation also differed from KEGG and SEED in the construction of orthologous groups with  
328 eggNOG using non-supervised construction increasing coverage whereas KEGG used supervised  
329 construction increasing annotation robustness. To obtain a more precise picture of C acquisition strategy,  
330 the CAZy annotated reads abundance were aggregated on the basis of their targeted substrates  
331 (Cellulose, Chitin, Glucan, Lignin, Peptidoglycan, Starch/Glycogen, Xylan, Other Animal  
332 Polysaccharides, Other Plant Polysaccharides, Oligosaccharides) using a curated database  
333 (Supplementary Table 2) based on previous works<sup>52-54</sup>. After mapping, the relative abundance of each  
334 gene (or aggregated group of genes) was normalised by the total number of bacteria-reads annotated for  
335 this sample on the same database. Such normalisation corrects for variation between samples in the  
336 quantity of annotated reads and avoids biases induced by contamination and sequencing error<sup>55</sup>. The  
337 obtained relative abundances inform on the relative importance of a gene (or gene group) compared to  
338 all the other annotated functions.

### 339 *Life history trait calculation*

340 An additional database was built with genomic traits previously associated with bacteria life history  
341 strategies (Extended Data Table 1). For this database, nine life history traits were calculated. Seven traits  
342 were calculated by summing the relative abundances of genes associated with Sigma factor<sup>56</sup>,  
343 Exopolysaccharides (EPS)<sup>57</sup>, Chaperons<sup>12,58</sup>, Chemotaxis, and Osmolytes<sup>59-62</sup>, antibiotic resistance and  
344 carbohydrates degradation enzymes (CAZyme). In addition, average genome size was calculated using  
345 MicrobeCensus<sup>63</sup> and rrn copy number using the method described in <sup>64</sup>. All sequences were used as  
346 input for average genome size and rrn copy number, after a verification that eukaryotic sequences were  
347 negligible (less than 2% of annotated reads for all databases verified for all samples) and therefore, that  
348 the samples mostly captured bacteria.

### 349 *Statistical analysis*

350 To identify the multivariate axes that best explain the global scale variation in metagenomic community  
351 aggregated traits of soil bacteria, we used a multi-table co-inertia analysis (MCOA), an exploratory  
352 analysis that leverages together the information from the 5 databases (genomic traits, eggNOG  
353 categories, SEED modules, KEGG pathway, CAZy types). This method identifies co-relationships  
354 between the different databases and uses a covariance optimization criterion to summarise in a common  
355 structure the information shared by multiple multivariate (eg. omic) tables<sup>65-67</sup>. All variables (CATs)  
356 were log transformed ( $\log X + 1$ ) before the analysis to improve normality<sup>67</sup> and standardised to a mean  
357 of zero and a variance of 1. The R package ade4 was used for the MCOA analysis<sup>68</sup>.

358 Sample coordinates on the first and second dimension of the MCOA were extracted and used as latent  
359 variables representing bacterial community positions in the global trait space. Random forest models  
360 were then used to identify predictors of these coordinates among potential environmental drivers, which  
361 were the soil properties measured on the same sample as metagenome (see Soil sampling and  
362 characteristics) and climatic variables extracted from Worldclim2 : BIO1 = Annual Mean Temperature,  
363 BIO4 = Temperature Seasonality (standard deviation), BIO12 = Annual Precipitation and BIO15 =  
364 Precipitation Seasonality (standard deviation). First, we verified that all selected environmental drivers  
365 had spearman correlation coefficients lower than 0.7 to mitigate collinearity problems as recommended  
366 in <sup>69</sup>. Second, a variable selection process was carried out using the method implemented in the VSURF  
367 R package<sup>70</sup>. The number of predictors randomly tested at each node of the random forest tree (mtry)  
368 was optimised based on randomForest's tuneRF algorithm and the number of trees set to 1000. Third,  
369 the random forest models selected following the VSURF selection process were trained using ten-fold  
370 cross-validation (100 repetitions) implemented in the caret package<sup>71</sup> and model performance was  
371 assessed based on Root Mean Square Error (RMSE) and R squared. Finally, random forest predictive  
372 models were used to project a broad resolution map of trait dimension global biogeography using  
373 environmental maps (1600x1200 pixel) as predictors. For this projection, we used the the latest map  
374 (June 2022) released by ISRIC's World Soil Information Service  
375 ([https://files.isric.org/soilgrids/latest/data\\_aggregated/](https://files.isric.org/soilgrids/latest/data_aggregated/)) based on SoilGrids version 2.0<sup>72</sup>. Worldclim2



376 (<https://www.worldclim.org/>) was used for climatic variables. The raster R package was used for the  
377 spatial predication and projection. To validate the relevance of this broad resolution map to represent  
378 average local values, we tested the correlation between local observations and the predicted value of the  
379 cell in which the local observation was done.

380 Finally, we tested the relationship between phylogenetic composition of the bacterial communities and  
381 their positions in the MCOA trait space using linear correlation between Euclidean distances along the  
382 two first dimensions of the MCOA and Unifrac phylogenetic distance. The influence of average genome  
383 size on this relationship was then assessed by comparing the correlation coefficients for communities  
384 below and above the median average genome size in the dataset.

### 385 **Data availability**

386 The five CAT databases used to build the trait dimensions and the associated environmental variables  
387 are available on figshare repository : <https://doi.org/10.6084/m9.figshare.22620025> All the original  
388 sequences are available in the European Bioinformatics Institute Sequence Read Archive database:  
389 soil metagenomes, accession numbers PRJEB18701 (ERP020652), 16S metabarcoding sequences,  
390 accession numbers PRJEB19856 (ERP021922).

### 391 **Code availability**

392 Access to the code used in the analyses done for this research is available by request to the  
393 corresponding author.

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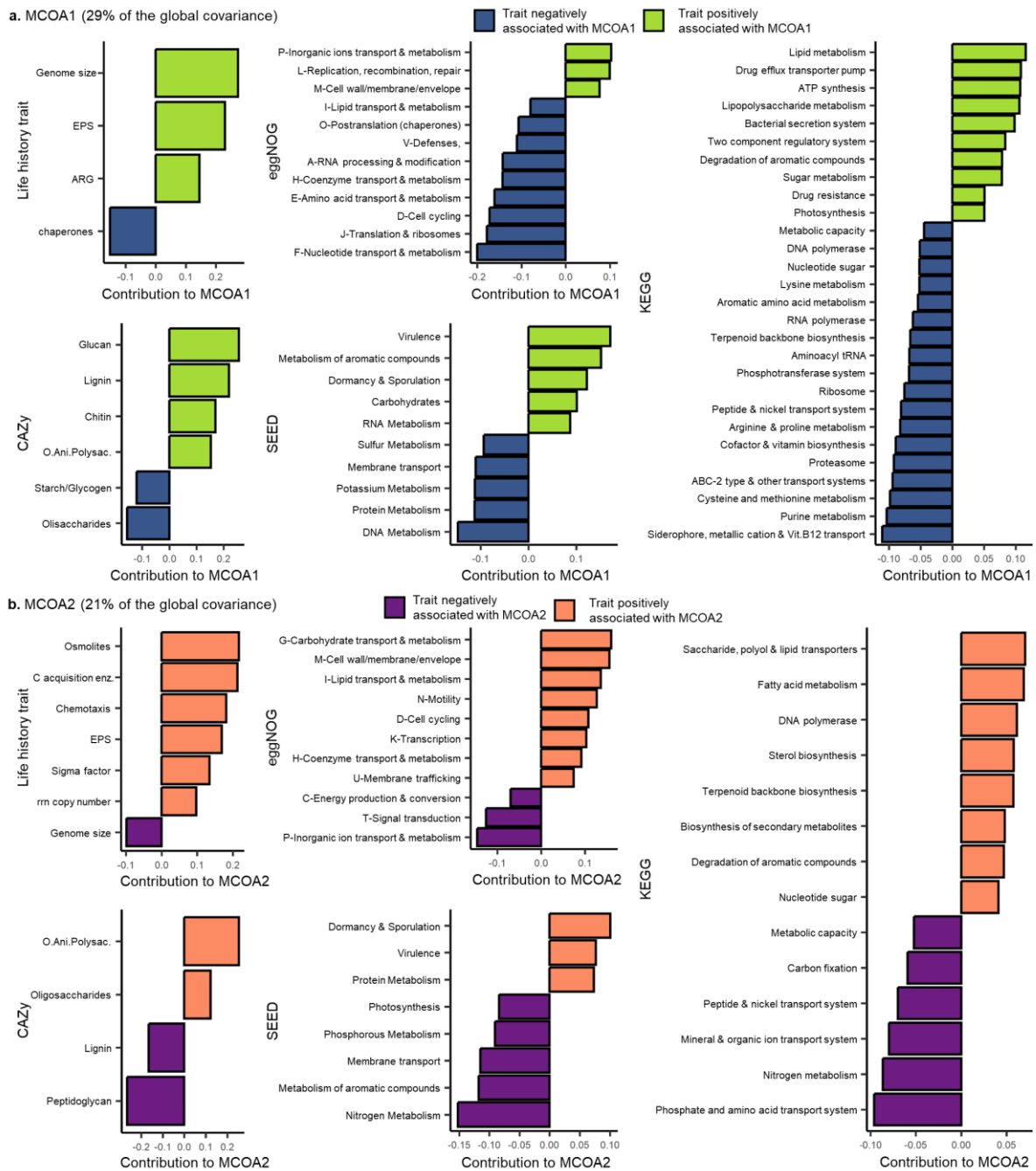
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404 project BBS/e/F/000Pr10355.

#### 405 **Authors contributions**

406 Data collection was designed and supervised by M.B. Initial bioinformatics analysis to obtain  
407 functional genes abundance tables (eggNOG, KEGG, SEED, CAZy) was designed and performed by  
408 F.H. Idea of this new analysis was conceived by G.P. with inputs from A.M., S.A, J.M. and K.T. New  
409 quantification of genomic traits, Unifrac and data analyses were performed by G.P. First draft and  
410 following editing was conducted by G.P. with inputs from all co-authors.

#### 411 **Competing interests**

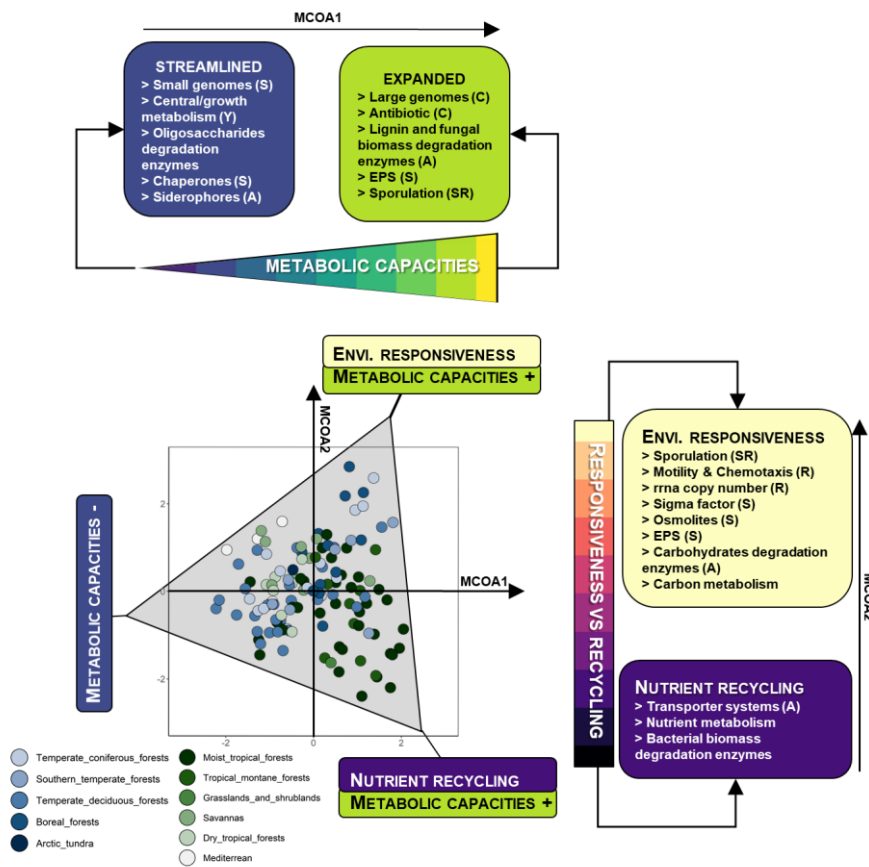
412 The authors declare no competing interests.



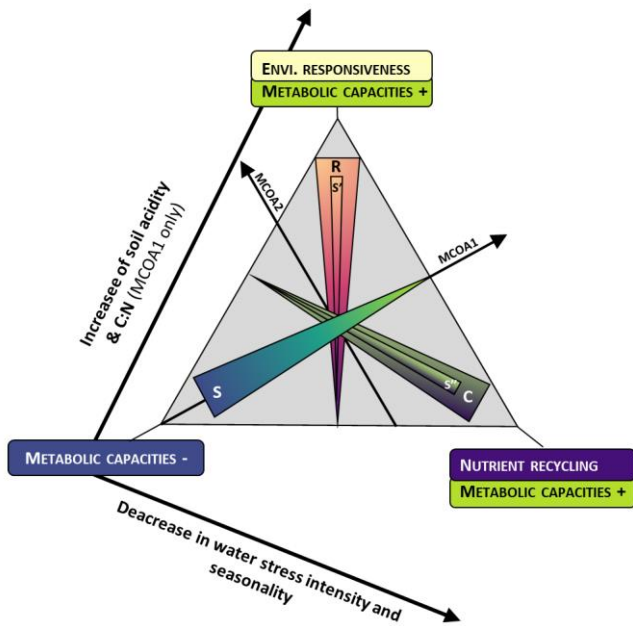
413

414 *Figure 1. Global trait dimensions of soil bacteria metagenomes. Variable contributions to the multiple*  
 415 *co-inertia analysis (MCOA) summarising in a common structure (MCOA dimensions 1 and 2) the*  
 416 *information shared by 5 community aggregated trait (CAT) databases (Life history trait, CAZY,*  
 417 *eggNOG, SEED and KEGG). Only the most important variables with significant correlation ( $p < 0.001$ )*  
 418 *with each dimension are reported in this figure. a and b panels present variable contributions to MCOA*

419 Dimension 1 and 2 respectively. Bar colours indicate the direction of the associations between the  
 420 variable and the MCOA dimensions.

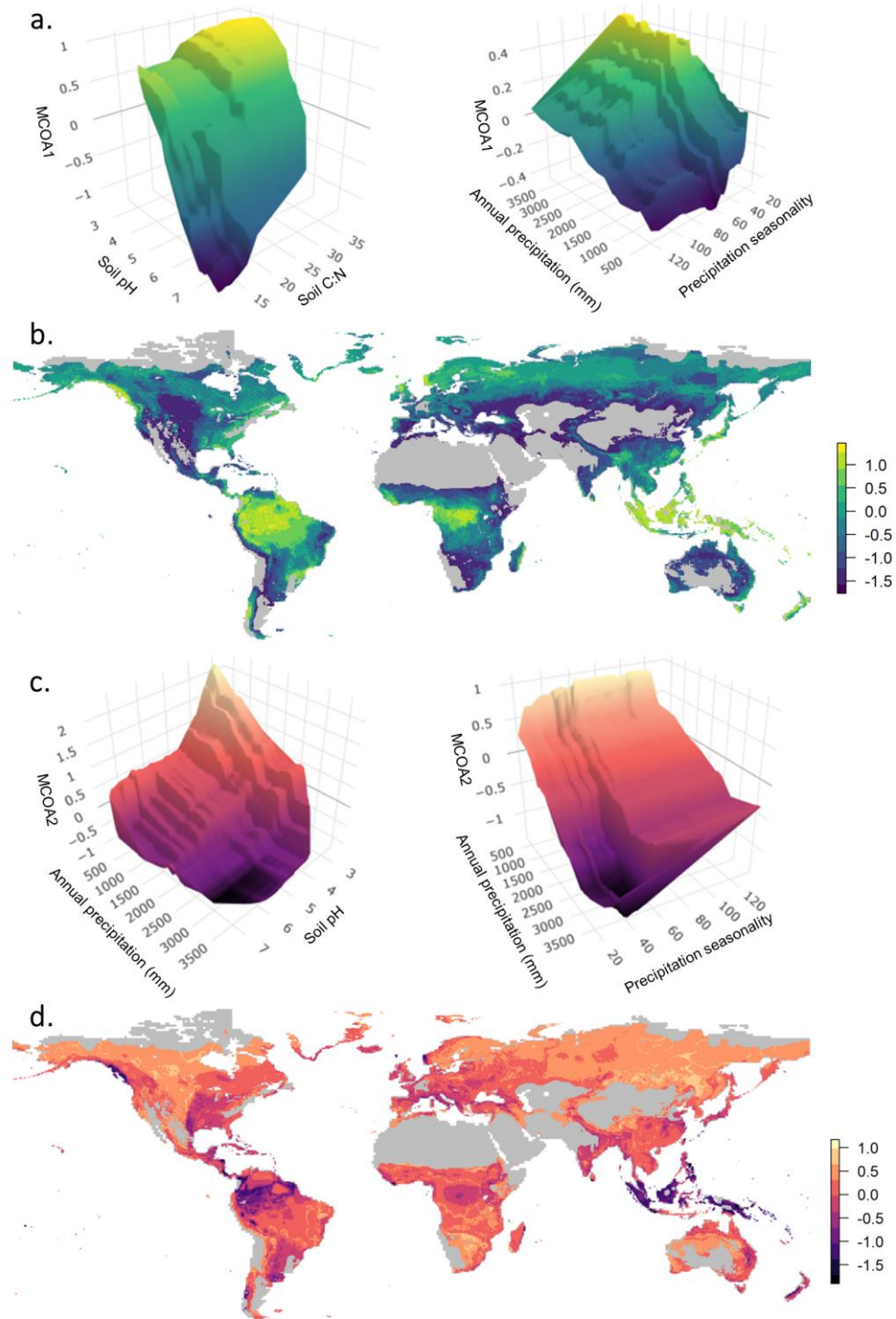


421  
 422 Figure 2. The global life history strategies of soil bacteria communities. Two-dimensional trait space  
 423 from a MCOA depicting trait associations across soil bacteria communities, with traits inferred from  
 424 enriched genes in bacteria metagenomes. Dots represent the positions of the 128 bacterial communities  
 425 used in this study along these two dimensions. In the trait lists, letters in brackets represent how CSR  
 426 (Competitors, Stress tolerant, Ruderal) and YAS (High Yield, resource Acquisition, Stress tolerance)  
 427 strategies have been associated with these traits in previous theoretical works (Extended Data Table1).



428

429 *Figure 3. Hypothesised role of competitor (C), Ruderal (R) and Stress tolerant (S) traits in shaping the*  
 430 *life history strategy observed at the community level and associated environmental gradients. Details*  
 431 *of CSR traits association are provided in Figure 1 and 2. S, S' and S'', represent the different S traits*  
 432 *associated with each dimension and detailed in Figure 1 and 2.*



433

434 *Figure 4. Environmental control and global scale projection of bacterial communities' coordinates*  
 435 *along MCOA dimension 1 and 2. a and c, random forest partial dependence plots describing*  
 436 *relationships between bacterial communities' coordinates along MCOA dimension 1 (a) and 2 (c) and*  
 437 *their most significant environmental predictors (Extended Data Figure 5). b and d, random forest*

438 predictions for MCOA dimension 1 (**b**) and 2 (**d**) projected across the globe using broad resolution map  
439 of mean soil and climate conditions (1600x1200 pixel), with land out of the dataset range in grey. Colour  
440 bars represent the predicted coordinates along MCOA dimension 1 (**b**) and MCOA dimension 2 (**d**).  
441 SoilGrids version 2.0 was used for soil properties and Worldclim2 for climate variables. Accuracy of  
442 the prediction was verified by ten-fold cross-validation of the random forest (Extended Data Figure 5)  
443 and by comparing the predicted values of the broad resolution projection with local observations  
444 (Extended Data Figure 6).

445

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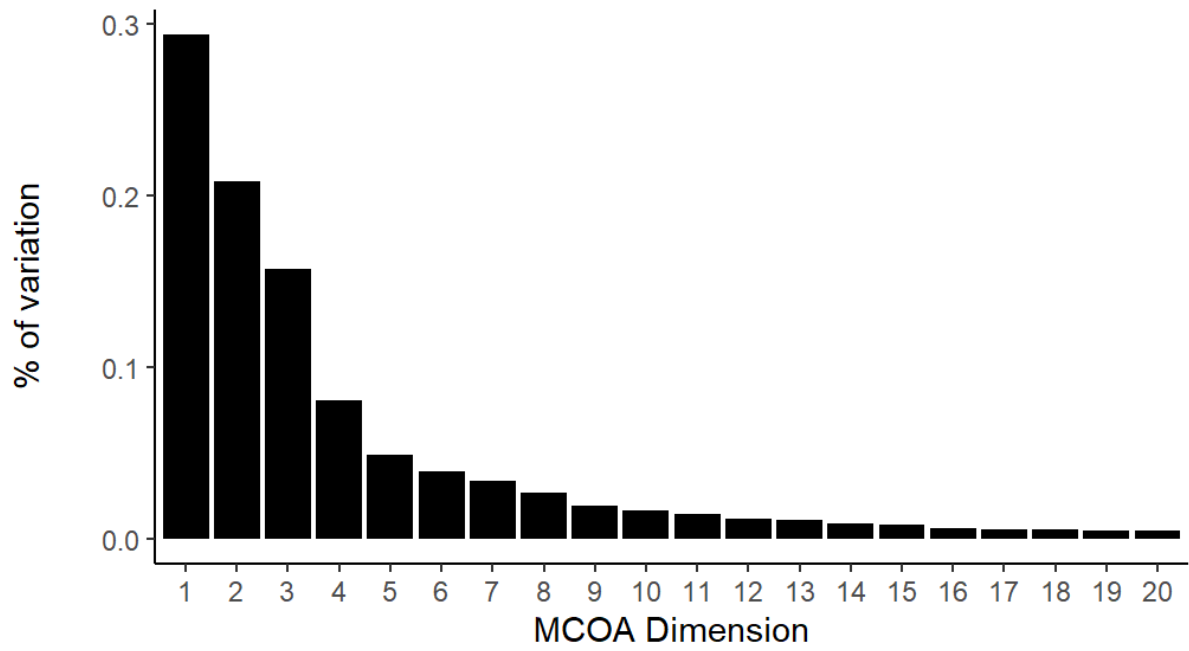
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607

608 **Extended Data**

609 *Extended Data Table 1. Life history traits used in this study. Traits were selected based on their previous*  
610 *association with CSR ('Competitor', 'Stress tolerant', and 'Ruderal') strategies by Fierer (2017) [1] or*  
611 *Krause et al. (2014) [2] or YAS strategies ("Yield", "Resource acquisition", and "Stress tolerant") by*  
612 *Malik et al. (2020) [3]. Cells associated with CSR and YAS have been greyed based on the strategy to*  
613 *facilitate comparisons between references. Same gray has been used for C and A, and for R and Y*  
614 *strategies as they have some important theoretical linkages (Malik et al. 2020).*

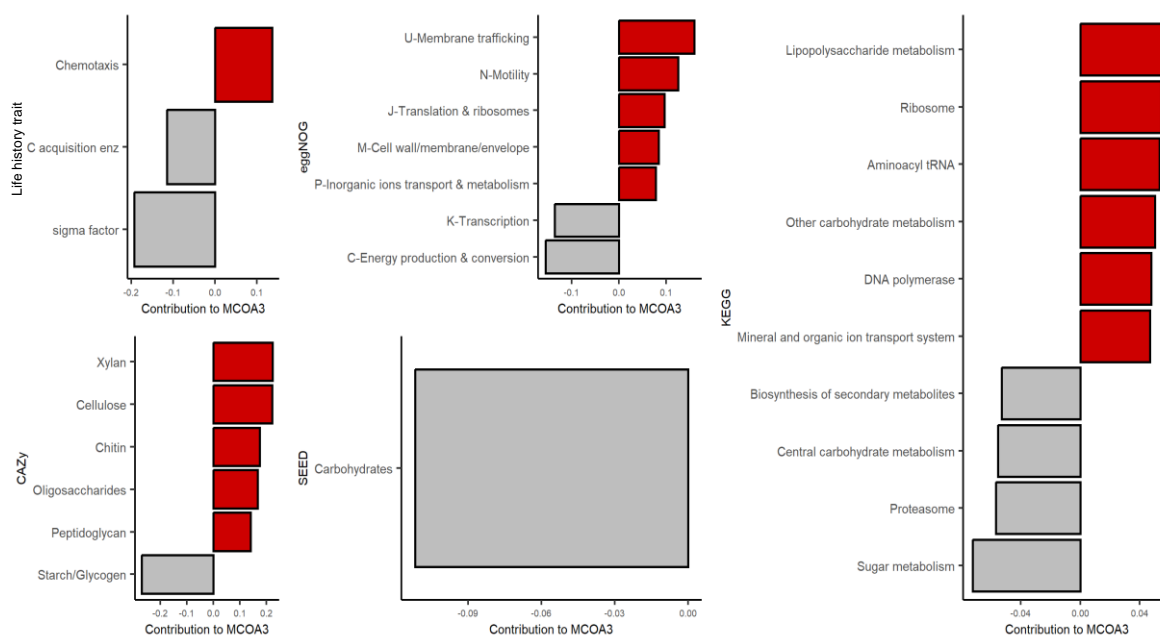
Life history traits	Associated metagenomic community aggregated traits used in this study	CSR [1]	CSR [2]	YAS [3]
Amino acid, fatty acid and nucleotide synthesis [3]	> eggNOG category, KEGG pathway and SEED modules associated with amino acid, lipid and nucleotide metabolism			Y
Chaperons [3]	> Chaperons genes : <i>GroEL</i> (COG0459), <i>dnaK</i> (COG0443) and <i>dnaJ</i> (COG0484) (Malik et al. 2020, Finn et al. 2020)			S
Siderophores [1,3]	> KEGG pathway "Metallic cation iron siderophore and vitamin B12 transport system "	C		A
Oligosaccharides degradation enzymes	> Genes associated with Oligosaccharides degradation among other GH and AA genes			
Carbohydrate central metabolism [3]	> KEGG pathway "Central_carbohydrate_metabolism"			Y
Primary metabolism	> eggNOG categories : F-Nucleotide transport and metabolism, J-Translation and ribosomes, D-Cell cycling, E-Amino acid transport and metabolism, H-Coenzyme transport and metabolism and A-RNA processing and modification, SEED modules : DNA and protein metabolisms. KEGG pathways: Purine, Cysteine, Methionine, Arginine, Proline and Lysine metabolism, Proteasome, cofactors and vitamins metabolisms and Ribosome, Aminoacyl tRNA, RNA and DNA polymerase and Nucleotide sugars			
Genome size [1,2]	> Average genome size (Nayfach and Pollard 2015)	C	R	
Complex polymers degradation enzymes [3]	> Genes associated with Lignin degradation among other GH and AA genes			A
Fungal biomass degradation enzymes	> Genes associated with Chitin and Glucan degradation among other GH and AA genes			
Antibiotic [1,2]	> Antibiotic Resistance Genes	C	C	
Pathogenic interactions with plants	> SEED module : Virulence			
Sporulation [1,2]	> SEED module "Dormancy_and_Sporulation"	R	S	
EPS [1,2,3]	> EPS genes : <i>WcaB</i> (COG1596), <i>WcaF</i> (COG0110), <i>Wza</i> (COG1596), <i>KpsE</i> and <i>RkpR</i> (COG3524) and <i>wcaK</i> (COG2327) (Cania et al. 2020)	S	S	S
Membrane synthesis and repair [3]	> eggNOG categories : L-Replication, recombination & repairs, M-Cell wall, membrane and envelope, KEGG pathways : Lipid and lipopolysaccharide metabolism			S
rRNA gene copies [1,2]	> Average rRNA copy number (Pereira-Flores et al. 2019)	R	C	
Motility [2,3]	> eggNOG category : "N-Motility"		R	A
Chemotaxis [2,3]	> Genes associated with chemotaxis : <i>CheA</i> ( COG0643), <i>CheY</i> (COG0784), <i>CheW</i> (COG0835), <i>CheB</i> (COG2201), <i>CheX</i> (COG1406), <i>CheD</i> (COG1871), Methyl-accepting chemotaxis proteins (COG0840, COG1352)		R	A
Sigma factor [3]	> $\sigma$ factor genes : $\sigma D$ , $\sigma S$ and $\sigma H$ (COG0568), $\sigma F$ and $\sigma B$ (COG1191), $\sigma N$ (COG1508) and extracytoplasmic function $\sigma$ factors (COG1595) (Chávez et al. 2020)			S
Osmolytes [3]	> Genes associated with Trehalose and glycine betaine (Malik et al. 2020, Sharma et al. 2020, Suriaty Yaakop et al. 2016, Bochet al. 1996, Wargo et al. 2013)			S
Exoenzymes (All) [2,3]	> GH and AA genes in global metabolism		S	A
Bacterial biomass degradation enzyme	> Genes associated with Peptidoglycan degradation			
Uptake system [2,3]	> KEGG pathway and SEED modules associated with transport systems		S	A

616



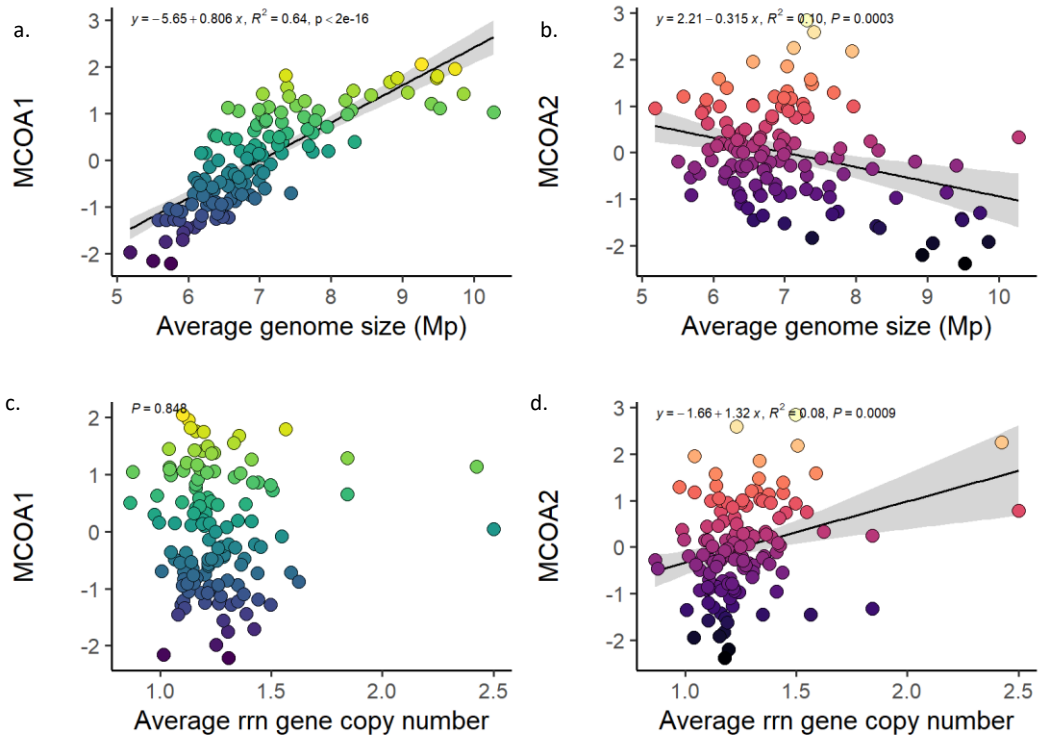
617

618 *Extended Data Figure 1. Stress plot representing the % of variation of the global dataset captured by*  
619 *each dimension of the MCOA*



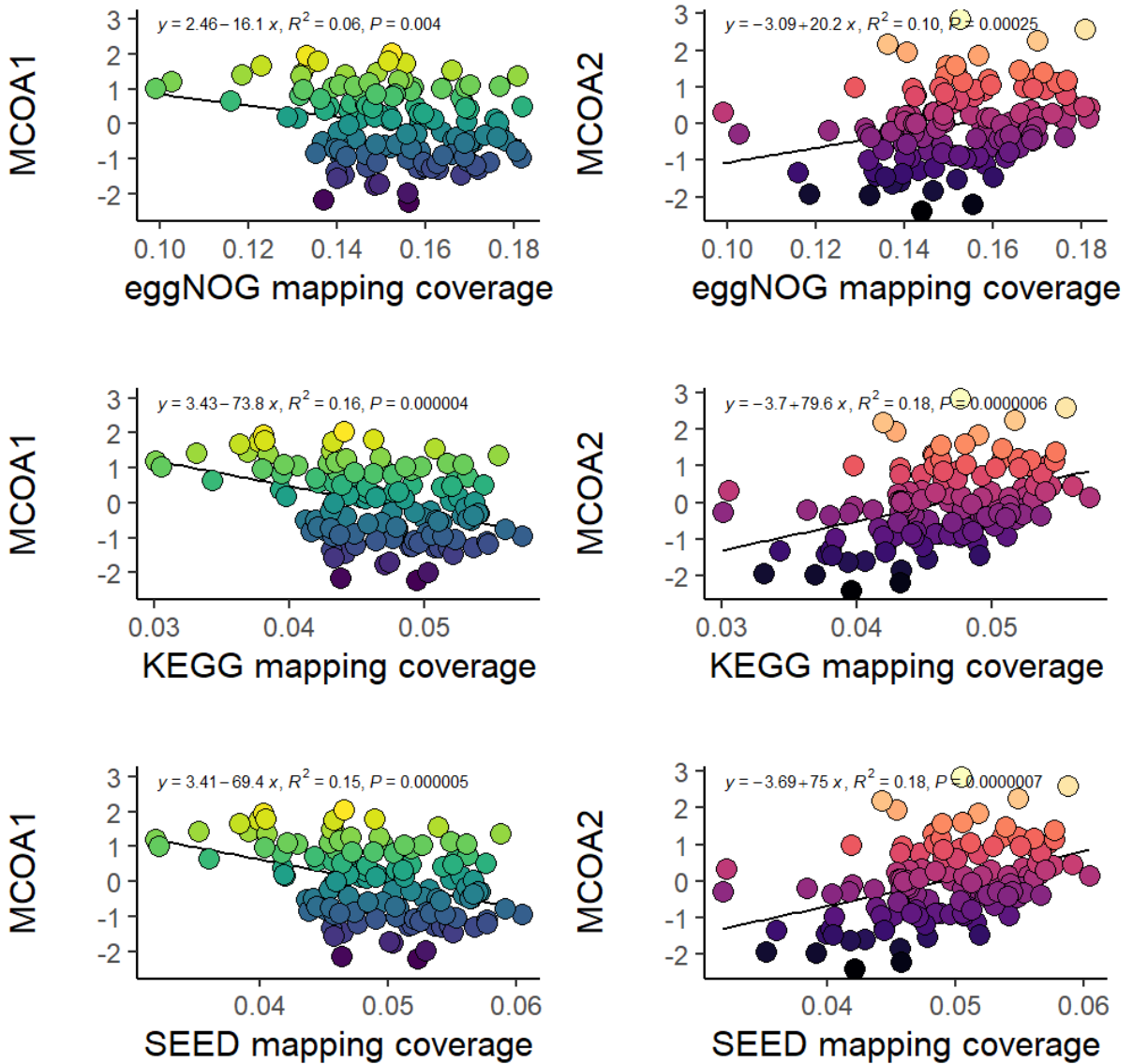
620

621 *Extended Data Figure 2. Variable contributions to the third trait dimension of the multiple co-inertia*  
 622 *analysis (MCOA). The MCOA summarizes in a common structure the information shared by 5*  
 623 *community aggregated trait (CAT) databases (Genomic trait, CAZy, eggNOG, SEED and KEGG). Only*  
 624 *the most important variables with significant correlation ( $p < 0.001$ ) with each dimension are reported*  
 625 *in this figure.*



626

627 *Extended Data Figure 3. Correlations between genomic traits and coordinates along dimensions 1 and*  
 628 *2 of the MCOA. The P value indicates the significance of the regression slope obtained using a t-test.*  
 629 *Shade represents the estimated 95% confidence interval. Color gradients follow MCOA dimensions and*  
 630 *match with figure 1 and 3 in the main text.*



631

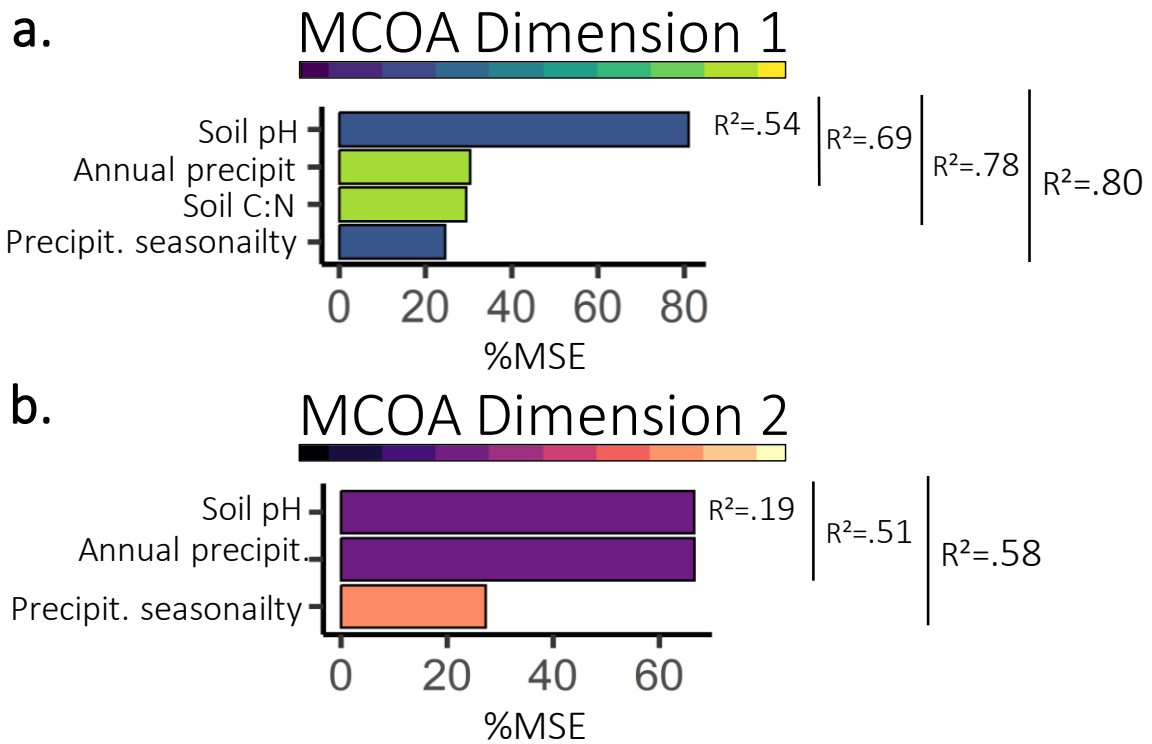
632 *Extended Data Figure 4. Correlations between MCOA dimensions (MCOA1 and MCOA2) and mapping*

633 *coverages on the 3 general databases (eggNOG, KEGG, SEED) used in this study. The P value indicates*

634 *the significance of the regression slope obtained using a t-test.*

635





636

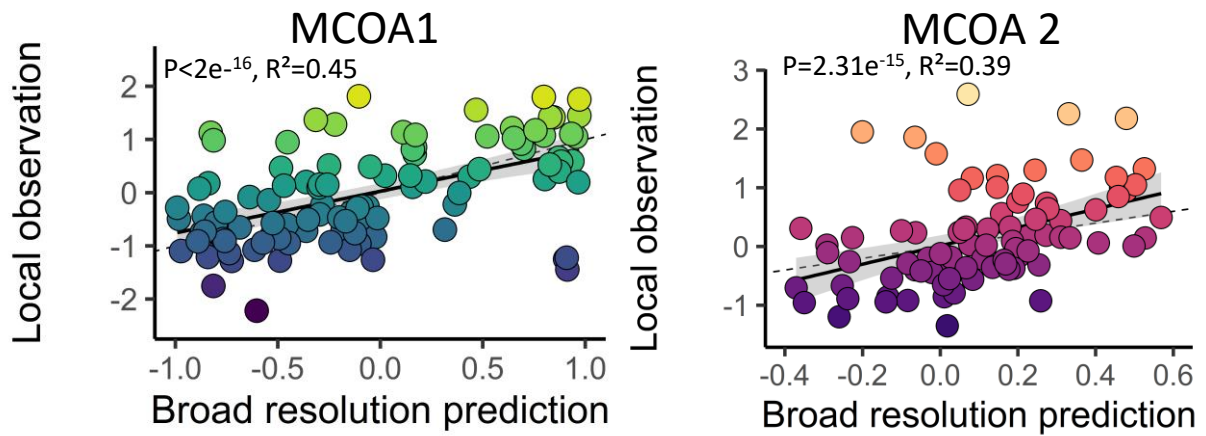
637 *Extended Data Figure 5. Environmental drivers of the bacterial community trait dimensions.*

638 *Environmental variable importances are represented as the mean decrease in mean square error*

639 *(%MSE) and R squared in random forest models predicting MCOA Dimension 1 (a) and 2 (b). Bar*

640 *colours indicate which end of the dimension (Figure 1 and 3) is positively correlated with the variable.*

641



642

643 *Extended Data Figure 6. Correlations between local trait dimension observations and global spatial*

644 *prediction. Correlations between local observations of bacterial community positions along the first*

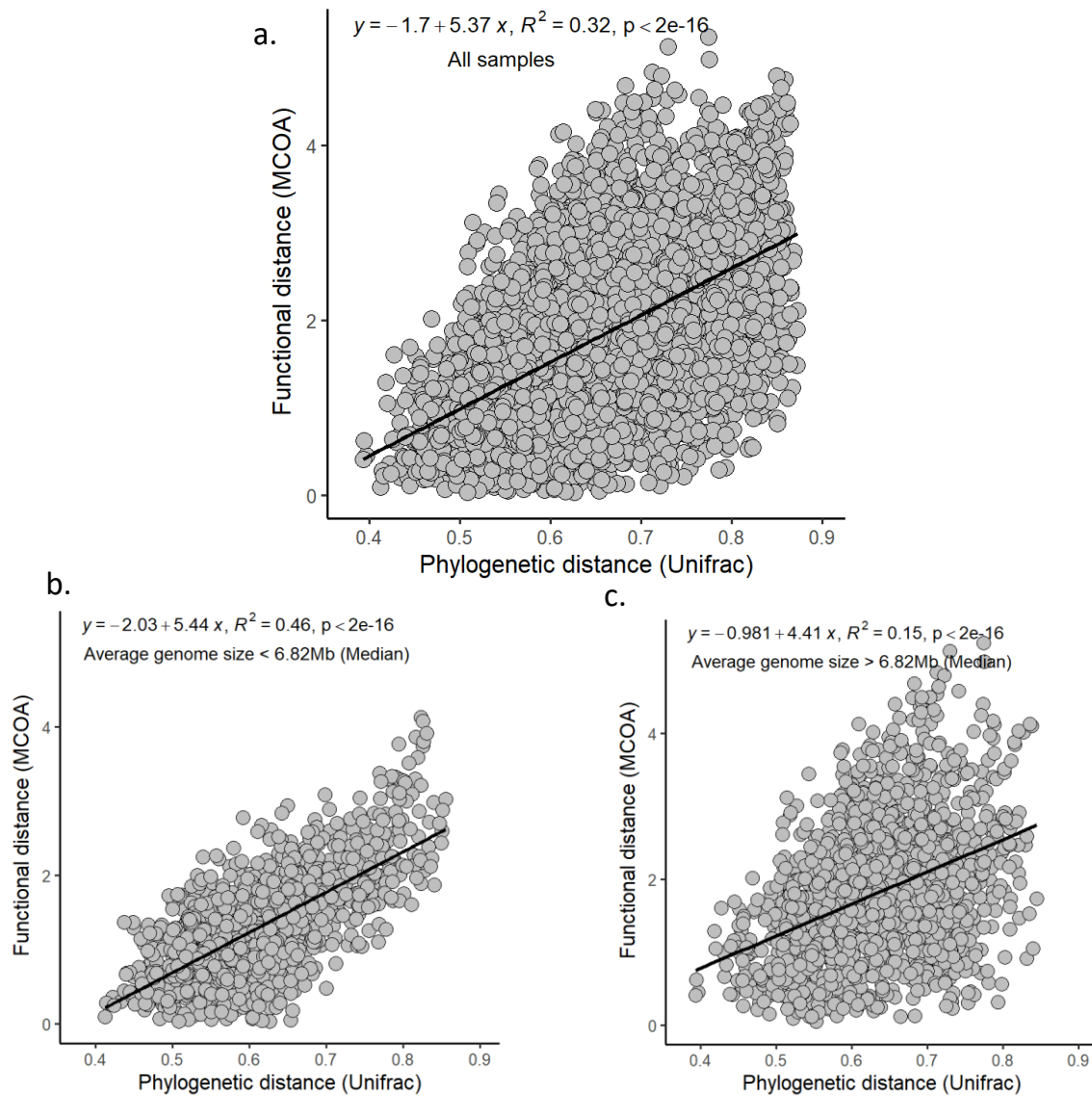
645 *and second trait dimensions from the MCOA (Figure 1-2) and the predicted value of the global map cell*

646 *(Figure 4) corresponding to where the local observations have been done. Dashed line represents a 1:1*

647 *correlation. The P value indicates the significance of the regression slope obtained using a t-test. Shade*

648 *represents the estimated 95% confidence interval. Color gradients follow MCOA dimension and match*

649 *with figure 1,2 and 4 in the main text.*

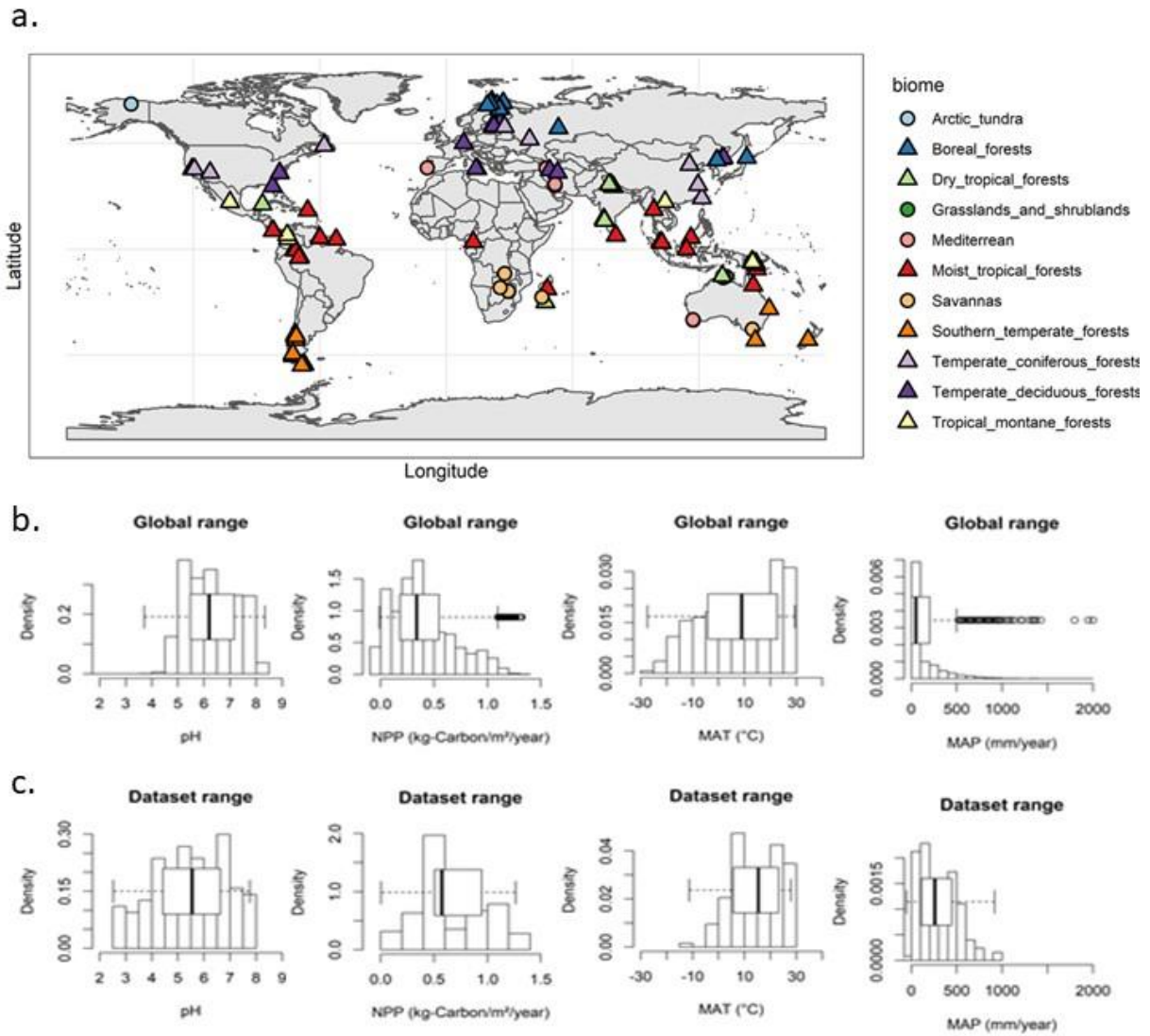


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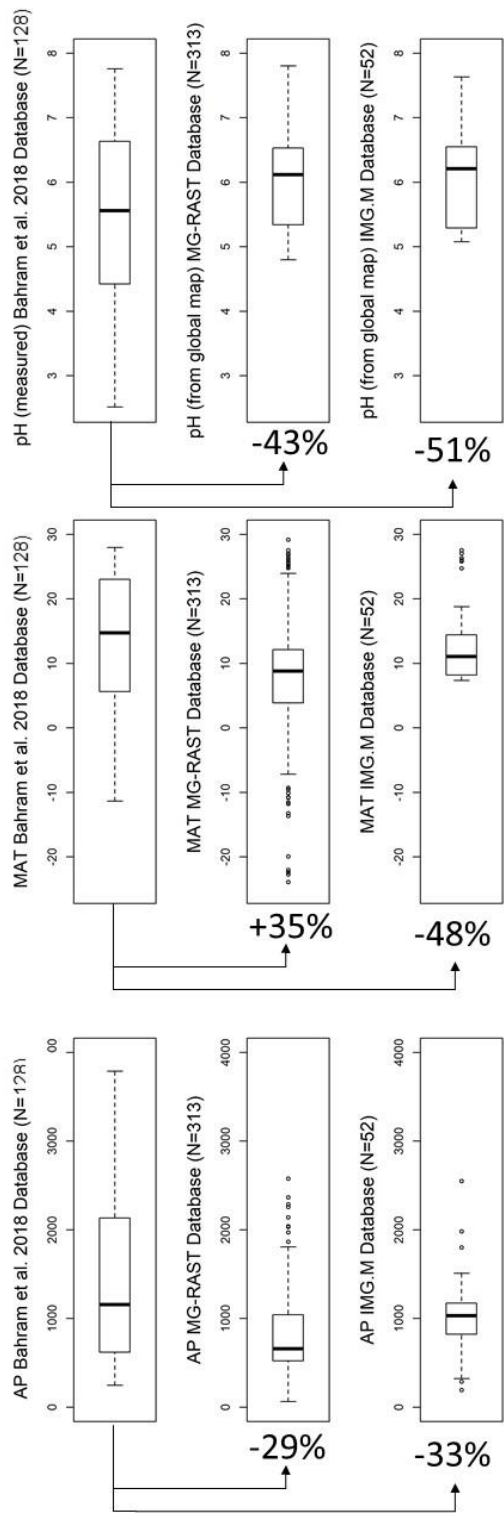
652 *Extended Data Figure 7. Correlation between phylogenetic distance (Unifrac metric) and functional*  
 653 *distance (Euclidian distance in MCOA space using coordinates of the two principal dimensions).*  
 654 *Correlation for all samples (a) and restricted to samples with average genome size below (b) and*  
 655 *above (c) its median value in the dataset. The P value indicates the significance of the regression slope*  
 656 *obtained using a t-test.*

657



658

659 *Extended Data Figure 8. Dataset distribution and environmental coverage. a. Sample localisations and*  
 660 *associated biomes b-c. Comparison between global range of environmental variables from the Atlas of*  
 661 *the Biosphere (b) and the environmental coverage of dataset (n=128) used in this study (c). Boxplot*  
 662 *elements: Center line=median; box limits=upper and lower quartiles; whiskers=1.5x interquartile*  
 663 *range; points=outliers. World map was done with rnaturlaearth R package*  
 664 *(<https://github.com/ropensci/rnaturlaearth>).*



665

666 *Extended Data Figure 9. Environmental coverage comparison between the database used in this study*

667 *from Bahram et al. (2018) and databases from the main metagenomes repositories (MG-RAST and*

668 *IMG:M). N corresponds to the number of metagenomes available in each database. MAT=Mean Annual*  
669 *Temperature, AP=Annual Precipitation. Boxplot elements: Center line=median; box limits=upper and*  
670 *lower quartiles; whiskers=1.5x interquartile range; points=outliers.*