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The structure of the food matrix at different length scales affects the mechanisms of digestion and the nutrient bioavailability

Didier Dupont

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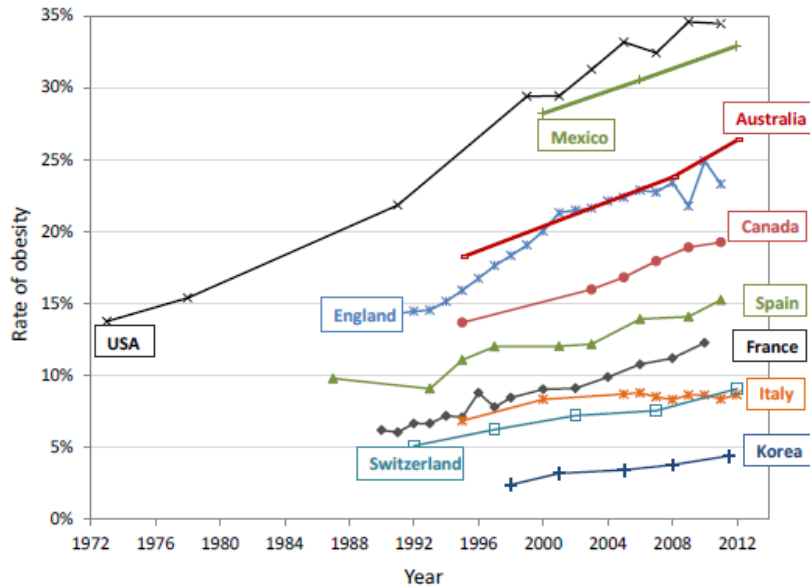
The structure of the food matrix at different length scales affects the mechanisms of digestion and the nutrient bioavailability



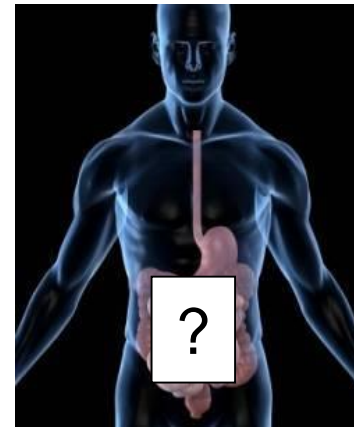
Dr Didier DUPONT, INRAE, STLO, Rennes, France



Food and human health: the key role of digestion



Diet-related diseases ↑
Prevent these pathologies rather than
cure them



Gut = interface between food and human body

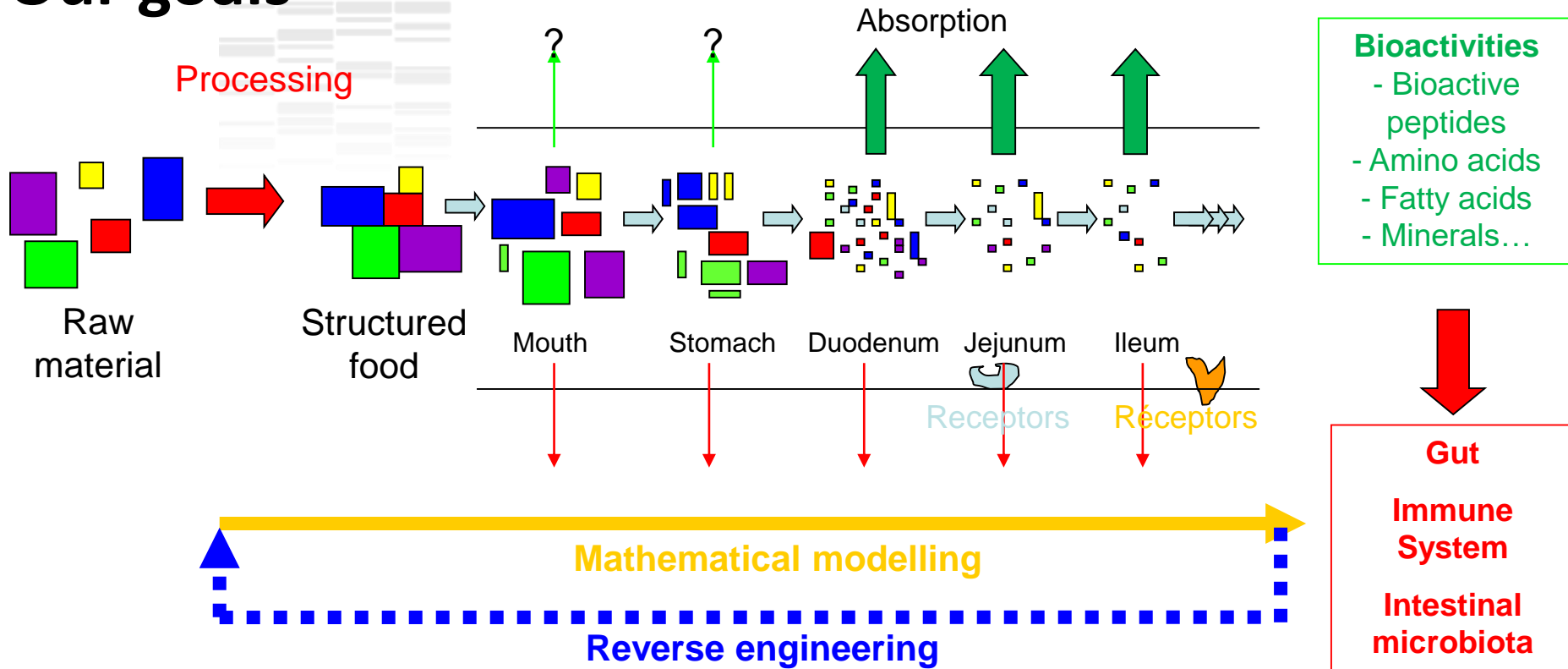
Digestion releases food components that can have a beneficial or a deleterious effect on human health

... but the mechanisms of food disintegration in the gastrointestinal tract remain unclear and the digestive process has been considered as a black box so far

By increasing our knowledge on food digestion, we will increase our knowledge on the effect of food on human health

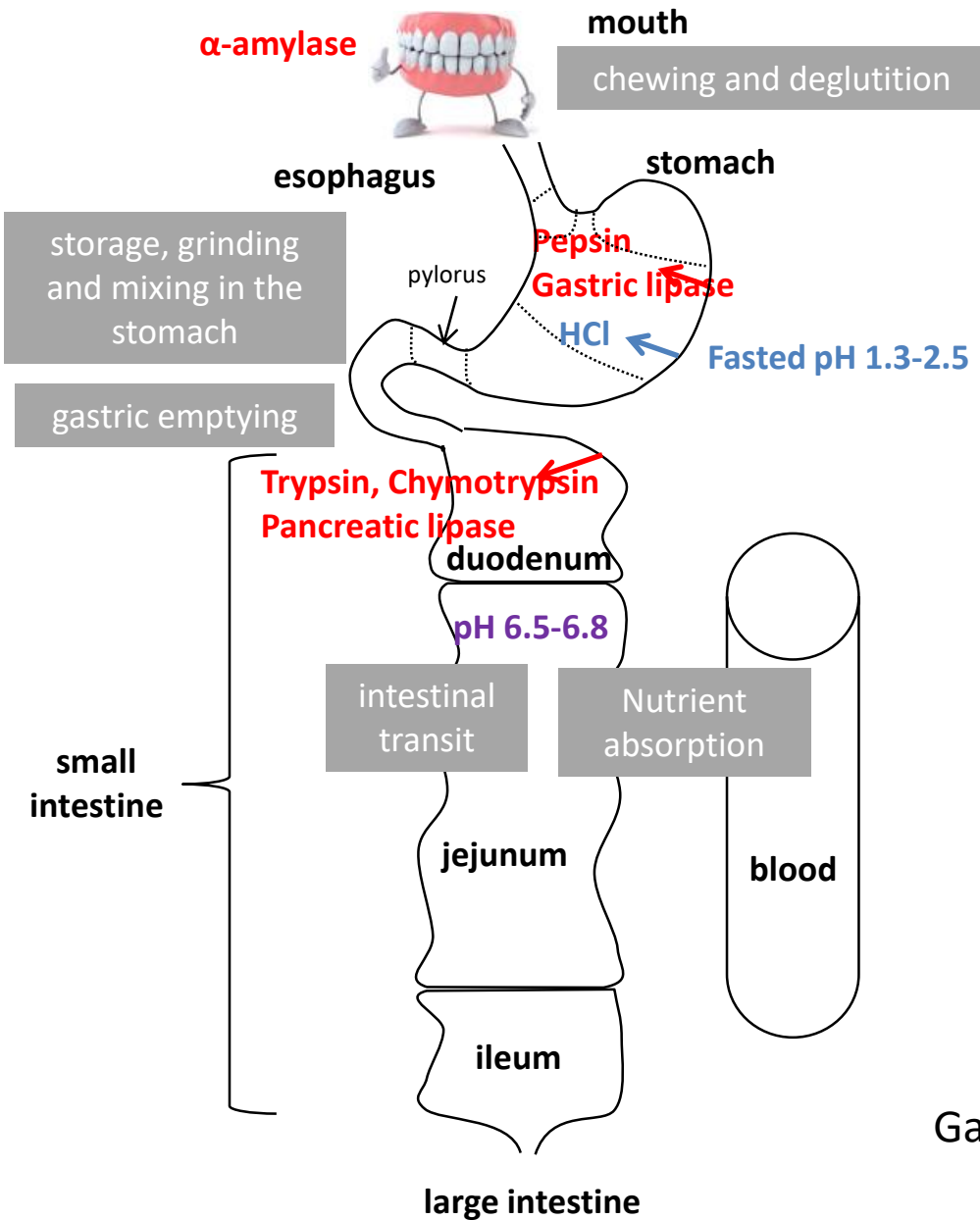
Our goals

Healthy Adult/ Infant / Elderly

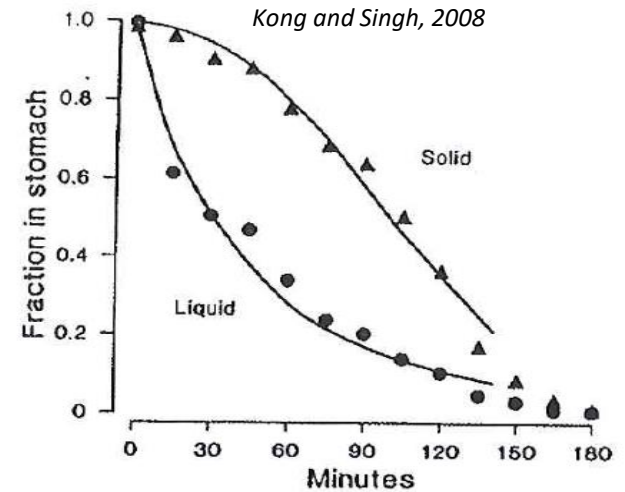


- ☞ To understand the mechanisms of breakdown of food matrices and their constituents in the gut and identify the beneficial/deleterious food components released during digestion
- ☞ To determine the impact of the structure of food matrices on nutrient bioavailability
- ☞ To model these phenomena in order to develop a reverse engineering approach

The digestive process



From Roger Lentle, Massey Univ. NZ



Gastric phase = a very complex but crucial step for the whole digestion process

Models available at INRAE for simulating digestion

Peng et al. 2021
 Halabi et al. 2021
 Giribaldi et al. 2021
 Chauvet et al. 2023
 Nebbia et al. 2022, 2023

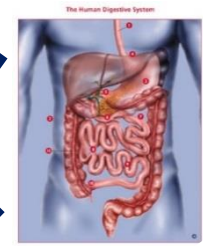
Menard et al. 2018, 2023
 Wang et al. 2022



*In vitro static models
 (infant, adult, elderly)*

Le Feunteun et al.
 2014, 2020

*In silico
 models*



*In vitro dynamic models
 (infant, adult, elderly)*

$$\Phi_{12} = k_{12whey} \times (V_1 - m_{caswpd1} \times \alpha) + k_{12aggr} \times m_{caswpd1} \times \alpha$$



Human models



Animal models

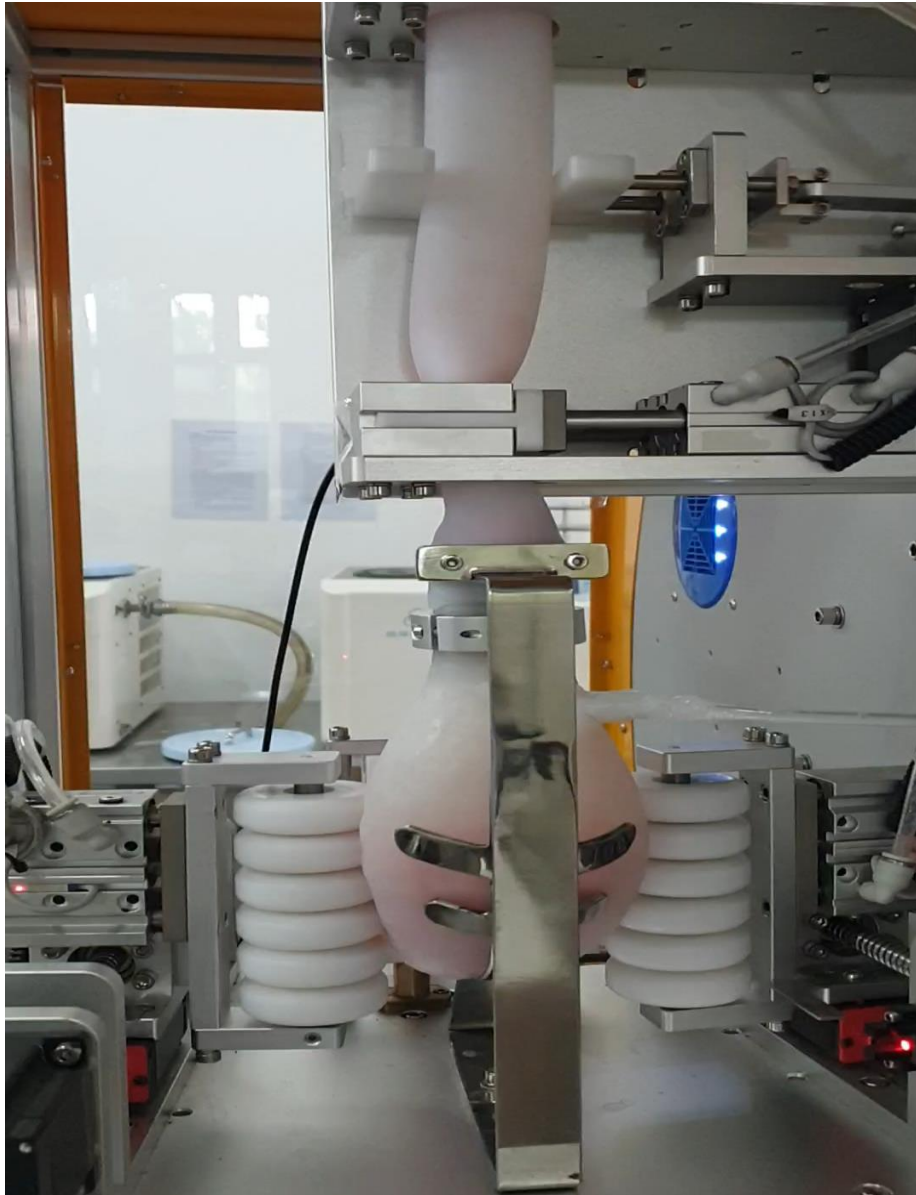


De Oliveira et al. 2016
 De Oliveira et al. 2017
 Buffière et al. 2020
 Boulier et al. 2023



Lemaire et al. 2021
 Nau et al. 2022
 Jimenez-Barrios et al. 2023
 Charton et al. 2022, 2023

NERDT™ : the NEar Real Digestive Tract



Xiao Dong Pro-Health
Smart Digestion
Suzhou University



INRAE

**L'INSTITUT
agro Rennes
Angers**





The molecular structure of food protein affects the kinetics of digestion



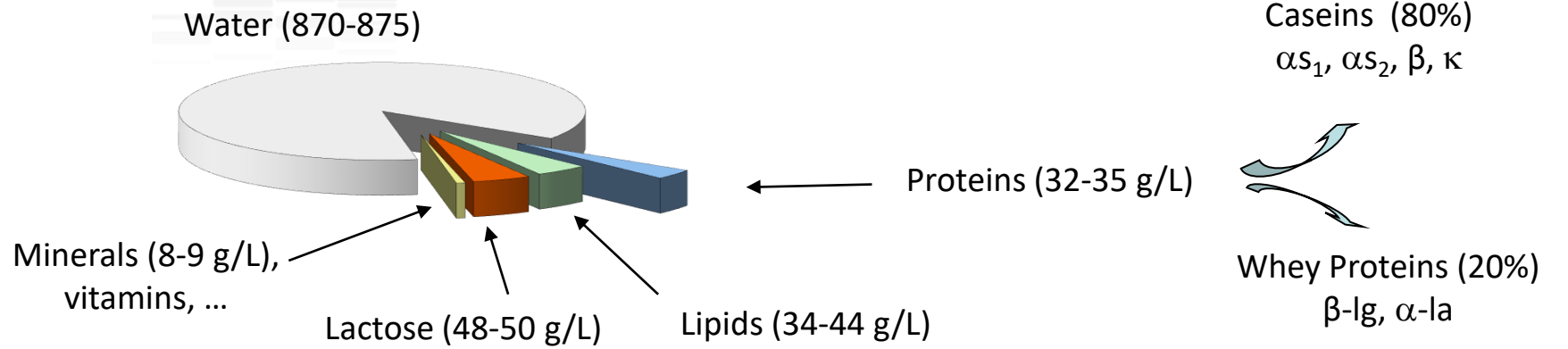
Dupont D.

INRAE, Rennes, France

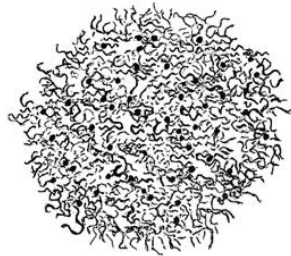


Milk

Molecular scale



$\varnothing \sim 200$ nm
(Holt, 1994)



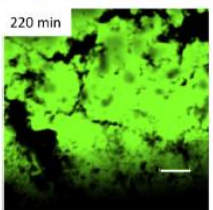
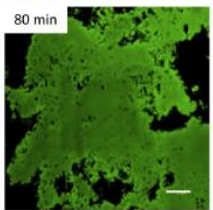
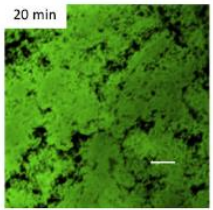
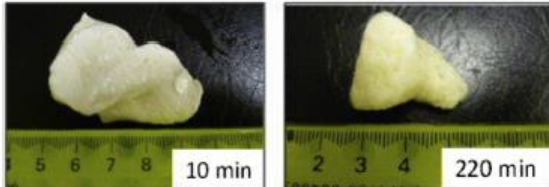
Casein organized into a supramolecular structure: the **casein micelle (CM)**



$\varnothing \sim 11$ nm
~15 casein molecules;
(Thomar *et al.* 2013)

Casein can also be extracted after acidification followed by neutralization: the **caseinate (CS)**

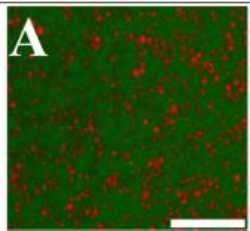
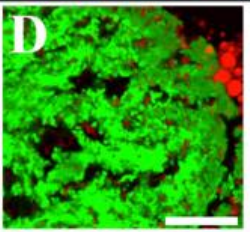

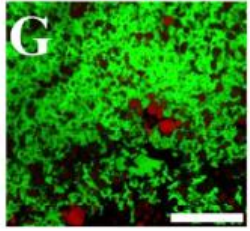

Milk coagulates in the stomach



In vitro demonstration using the HGS
Ye *et al.* 2016



Ye *et al.* 2019
In vivo evidence using a rat model

	Raw	
Initial		
36min		
182 min		

Mulet-Cabero *et al.* 2019
In vitro semi-dynamic model

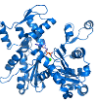


Skim Milk Powder



Sodium Caseinate

Casein micelles and sodium caseinate form different coagulums
Wang *et al.* 2018



The objectives of the study were to:

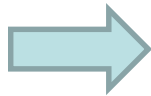
- 1 Determine whether gastric emptying of an isoproteic solution of CM and CS are different or not (exp. 1)
- 2 Characterize the structure of the resulting chyme and determine if CM and CS are differently metabolized (exp. 2)

Experiment 1 – Determination of Gastric Emptying

96 g of CM or CS
rehydrated in
800 ml of water

+12 g of glucose

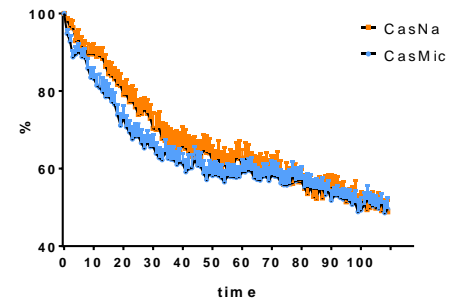
+ ^{99m}Tc -colloidal
(25Mbq)



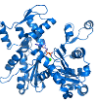
9 pigs (20-25 kg)



γ -scintigraphy over 120 min



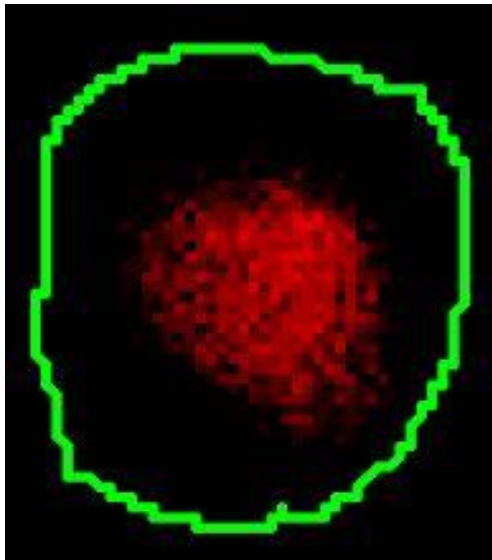
Gastric emptying half-time ($T_{1/2}$) and shape of the curve (β)



But a differential behaviour of CS and CM in the stomach

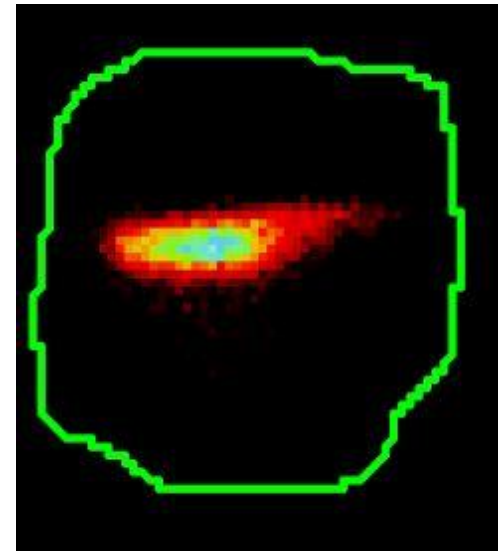
Exemple scintigraphic images at the beginning of gastric emptying (5-10 min after ingestion)

CM



Radioactivity fully fills the stomach

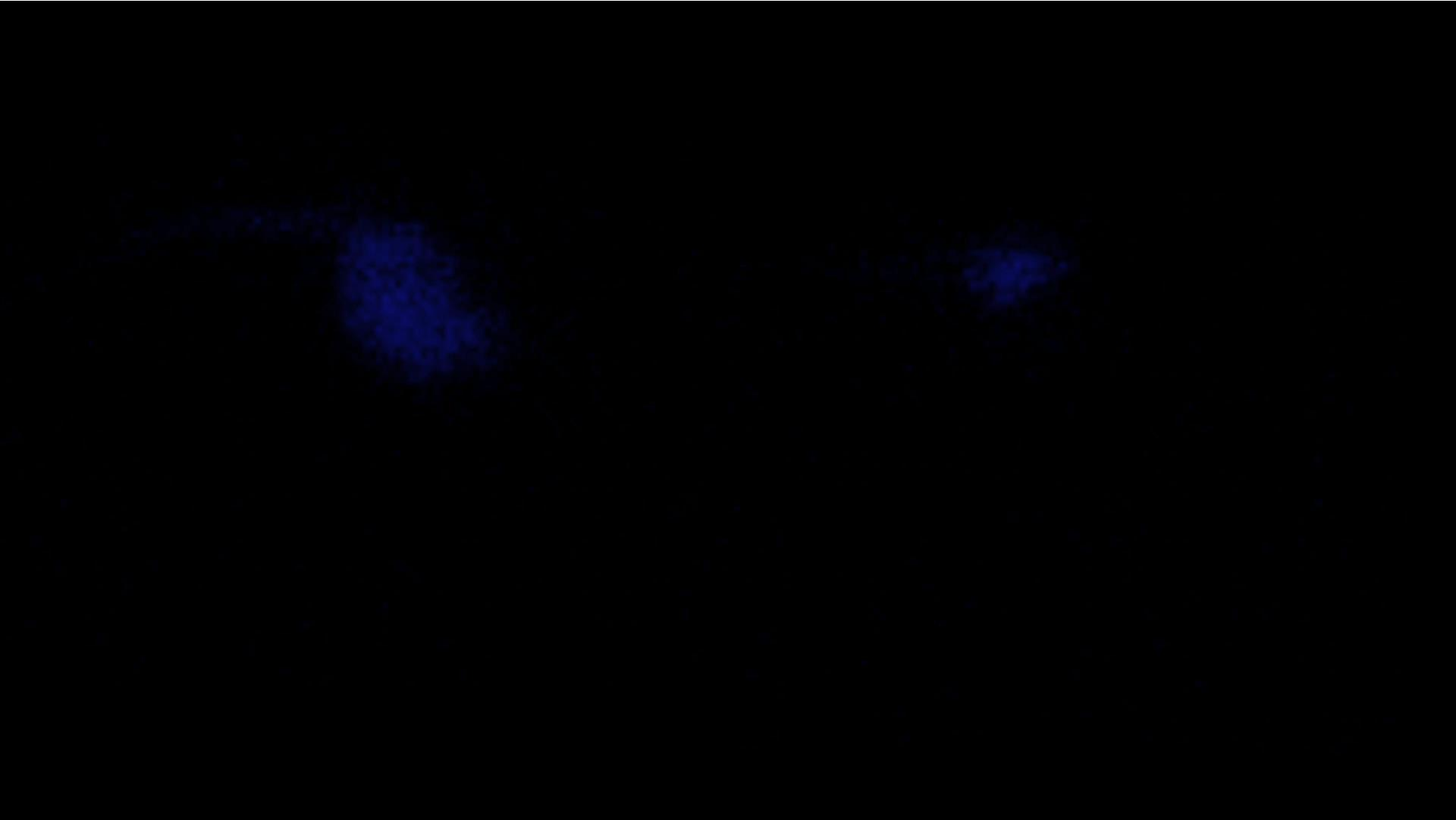
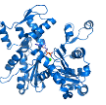
CS



Radioactivity is highly concentrated in the proximal part of the stomach

Gastric emptying

Molecular scale



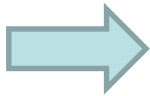
The objectives of the study were to:

- 1 Determine whether gastric emptying of an isoproteic solution of CM and CS are different or not (exp. 1)
- 2 Characterize the structure of the resulting chyme and determine if CM and CS are differently metabolized (exp. 2)

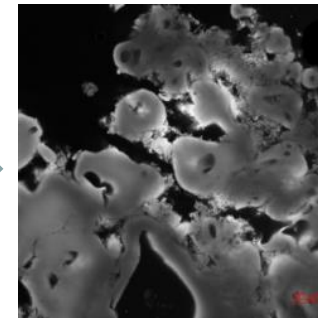
Experiment 2 – Chyme structure and protein metabolism

96 g of CM or CS rehydrated in 800 ml of water

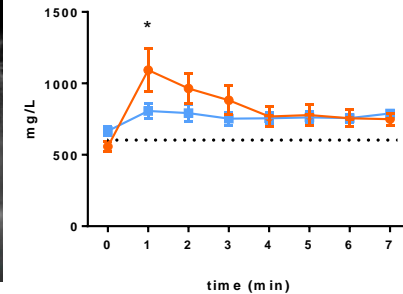
+12 g of glucose



10 catheterized pigs (20-25 kg)



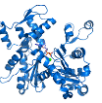
Characterization of the chyme structure (slaughtering after 10 min, n=4)



Free plasma amino acids over 7h n=6

Collection of the stomach contents

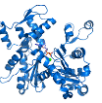
Molecular scale



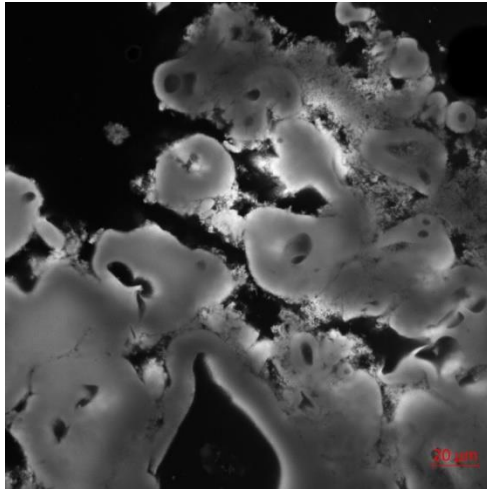
CM forms a large coagulum in the stomach whereas CS mainly remains in the liquid form

Microstructure of gastric chymes

Molecular scale



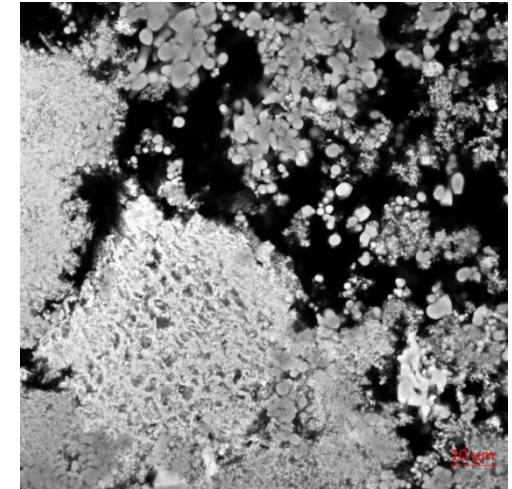
CM



CM Gels (left) are compact and dense =

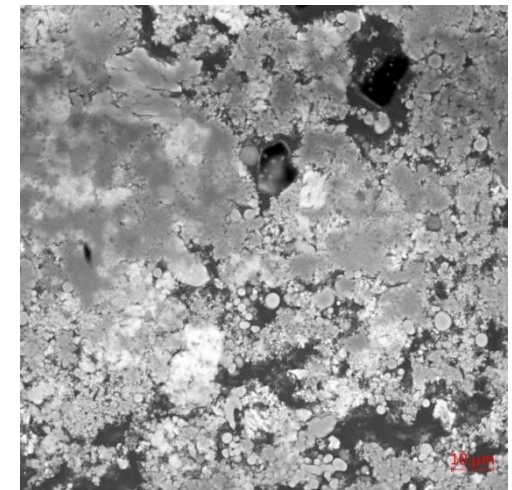
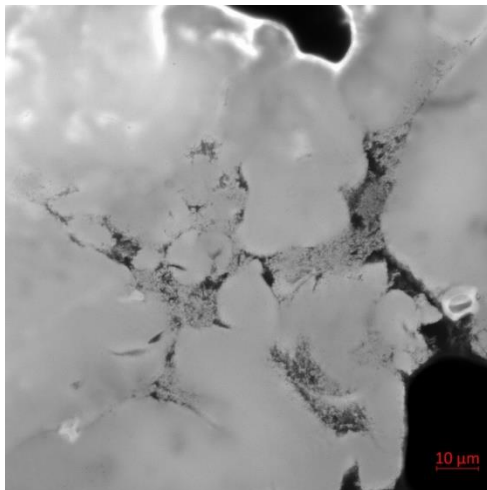
Strong coagulum

CS



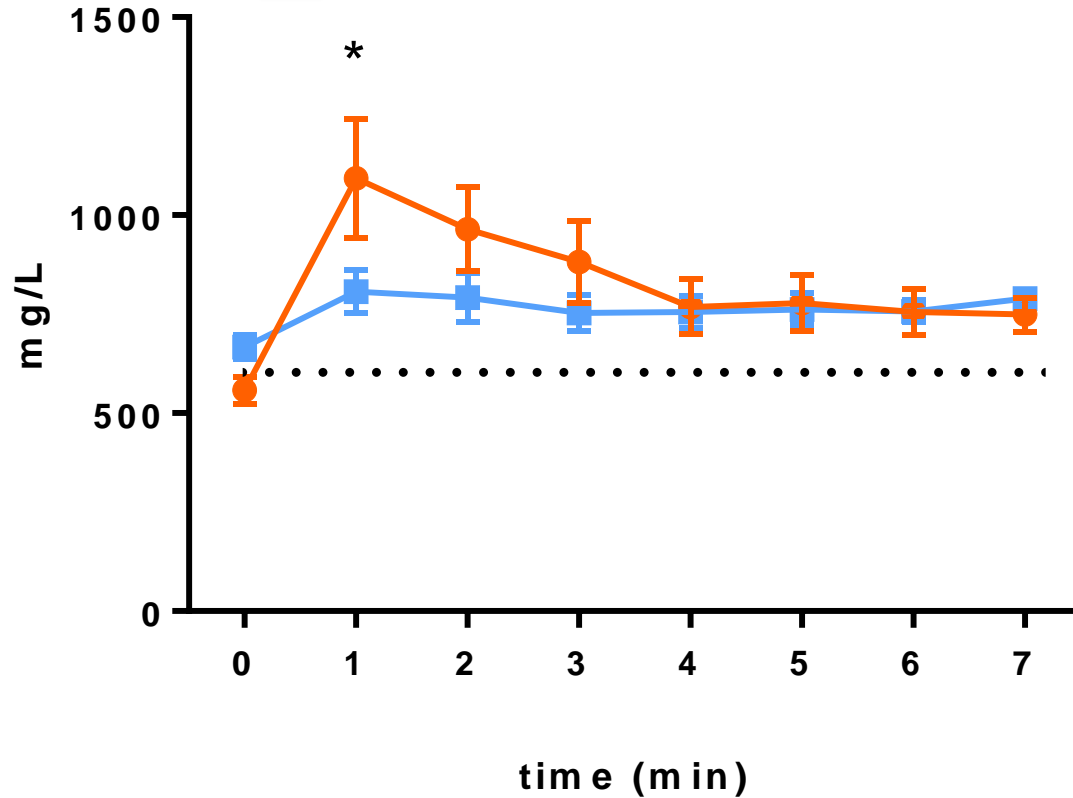
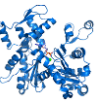
CS Gels (right) are an agglomerate of spherical particles that can easily dissociate. The gel have a very « loose » structure =

Protein precipitate



Plasma amino acids

Molecular scale



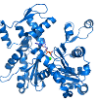
■ CasMic
● CasNa

casein x time
 $P = 0.0007$

**Boulier *et al.*
Food Chem. 2023**

AA peak after 1h for CM whereas the concentration remains stable for CS

Conclusion




- * CM form a strong coagulum in the stomach leading to a slow and constant release of plasma amino acids up to 7 h
- * CS form a loose precipitate in the proximal part of the stomach but most of the caseins remain solubilized in the liquid fraction
- * CS are rapidly metabolized in the small intestine leading to the appearance of a peak of plasma amino acids one hour after protein ingestion

CM = slow caseins, CS = fast caseins

Perspectives

Do some *in vivo* kinetics experiments

Use ^{15}N labelled-caseins to differentiate endo/exogenous proteins



From the protein molecular structure to the the food microstructure: The case of egg white gels



Dupont D.

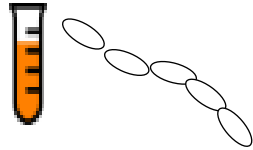
INRAE, Rennes, France



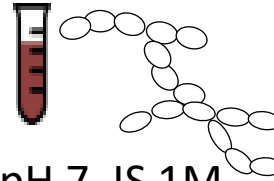
The microstructure of egg-white gels made from different types of aggregates affects the kinetics of proteolysis

Microscopic scale

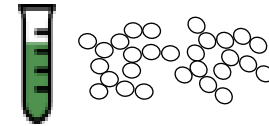
80°C/6h



pH 9, IS 1M





pH 7, IS 1M



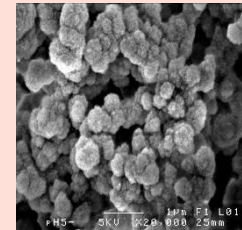
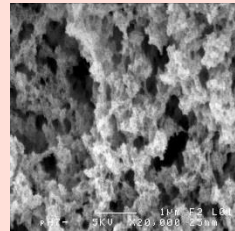
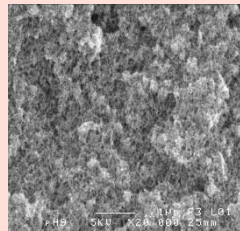
pH 5, IS 1M



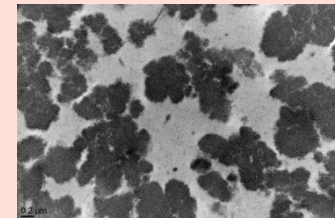
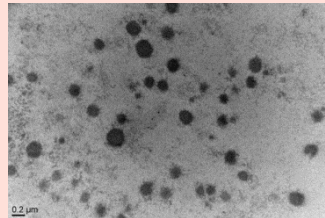
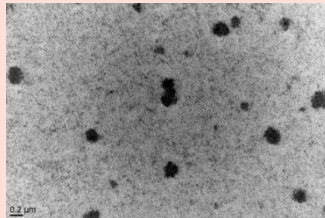
Aggregates	linear	branched	spherical
Rate of <i>in vitro</i> digestion	+++	++	+

Gels 90°C/2.5 h			
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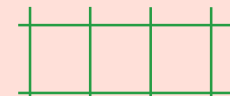
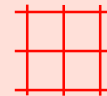
Nyemb *et al.*
Food Hydro. 2016
Nyemb *et al.*
Food Res Int 2016



SEM



CRYO-TEM



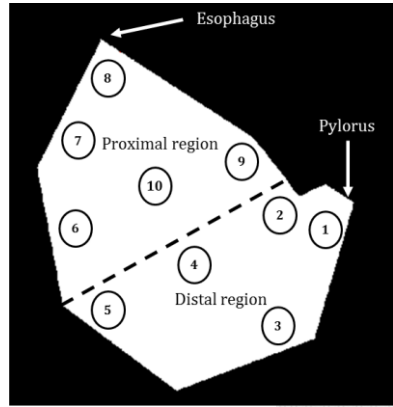
Rate of *in vitro* digestion +

++

+++

Spatial-temporal evolution of pH during an *in vivo* digestion

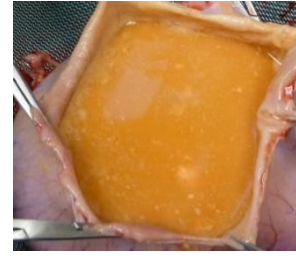
Microscopic scale



20 min



60 min

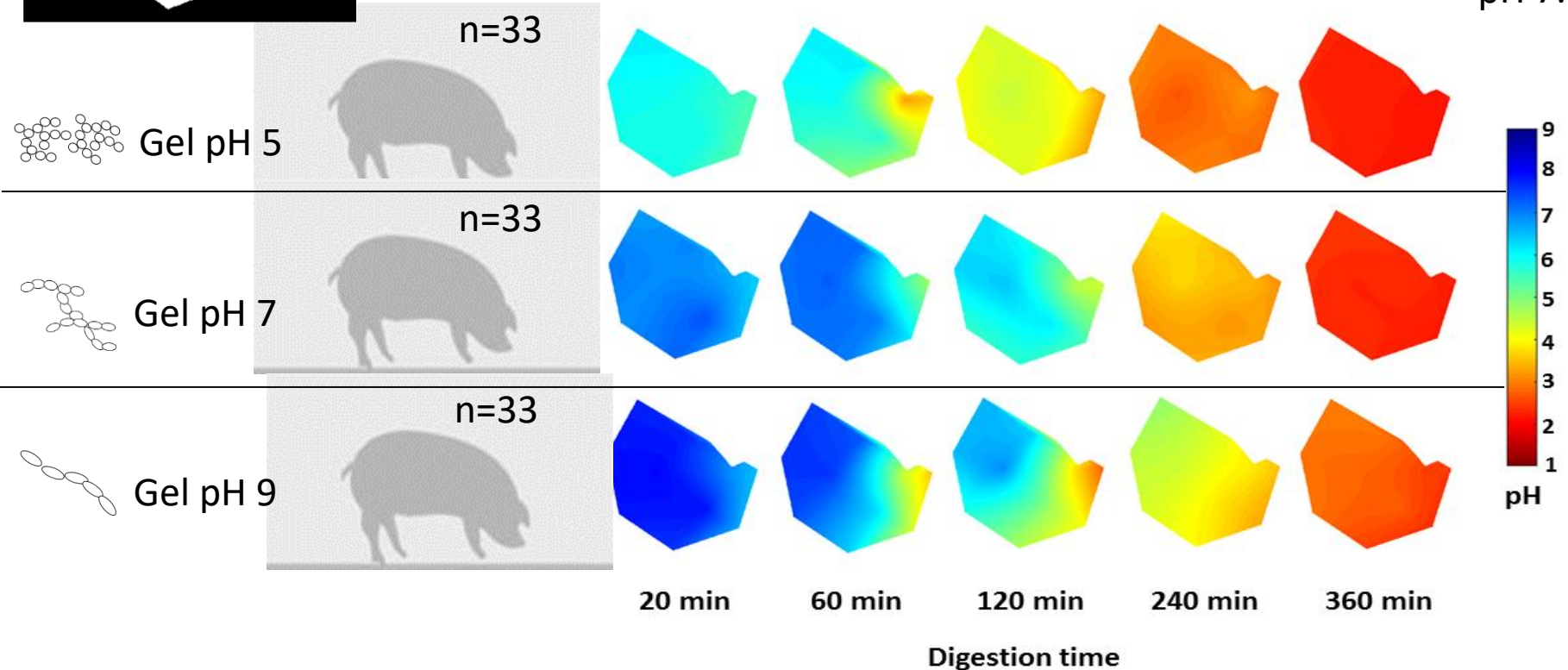


120 min

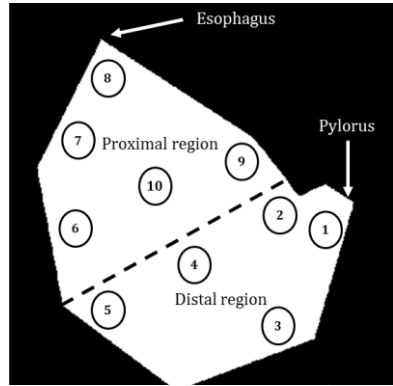


240 min

pH 7.0



Spatial-temporal evolution of pH during an *in vivo* digestion



20 min



60 min

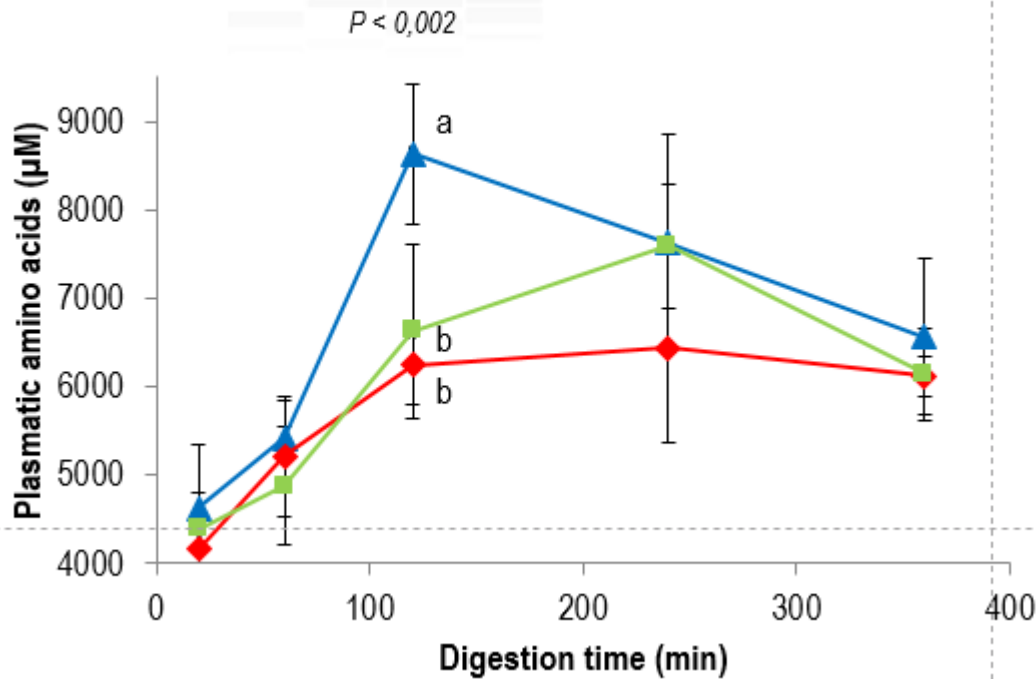


120 min



240 min

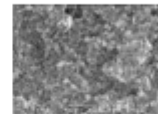
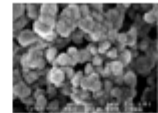
pH 7.0



▲ Granular-spongy EWG

■ Smooth-rigid EWG

◆ Intermediate EWG



Microscopic scale





Food structure as modified by processing affects the kinetics of food digestion



Dupont D.

INRAE, Rennes, France



Comparison of 6 dairy products of identical composition but different structure



Macroscopic scale

Ultra Low Heat powder



rehydration in water 14.5%

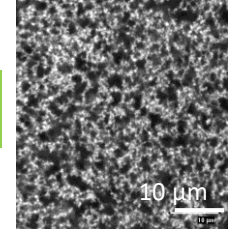


unheated milk ("raw" milk)

macrostructure

24h-20°C,
rennet 0.003 % v/w

rennet gel



pH 6.6



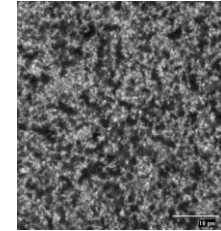
heat treatment
90°C-10 min

heated milk

microstructure

24h-20°C,
rennet 0.3 % v/w

rennet gel

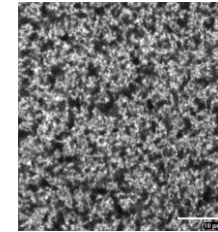


pH 6.6



24h-20°C,
GDL 3 % w/w

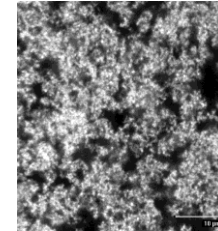
acid gel



pH 4

24h-20°C,
GDL 3 % w/w +
mixer 2 min

stirred acid gel



pH 4

Fat-free matrices:

40 g/L caseins, 10 g/L whey proteins,
95 g/L lactose and minerals

+ marker of the meal transit (Cr^{2+} -EDTA) → Gastric emptying half-time

The multi-cannulated mini-pigs

SOLID

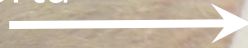
LIQUID



Macroscopic scale

6 minipigs (20 ± 1kg)

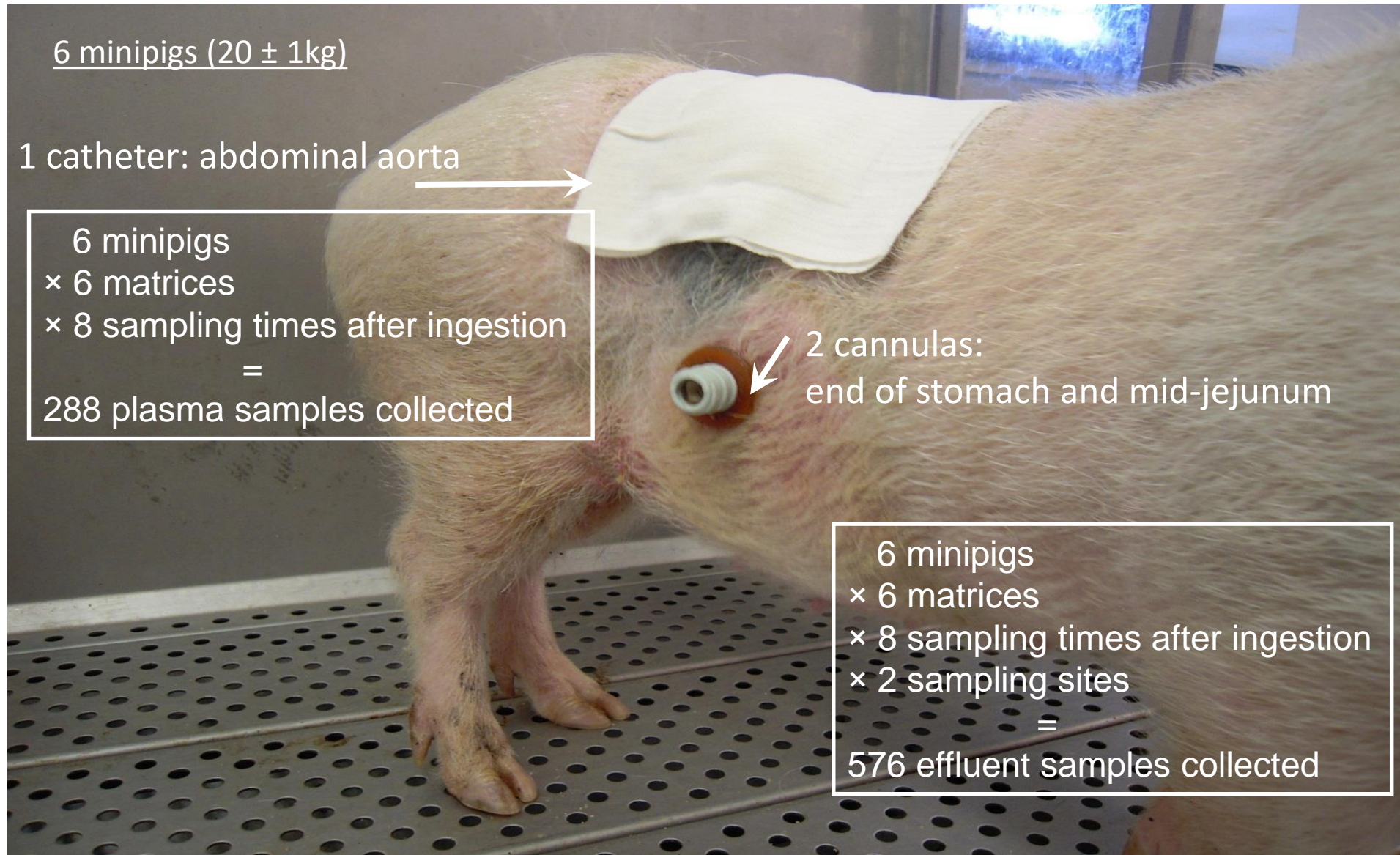
1 catheter: abdominal aorta



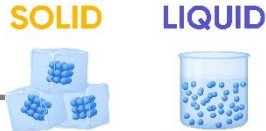
6 minipigs
× 6 matrices
× 8 sampling times after ingestion
=
288 plasma samples collected

2 cannulas:
end of stomach and mid-jejunum

6 minipigs
× 6 matrices
× 8 sampling times after ingestion
× 2 sampling sites
=
576 effluent samples collected

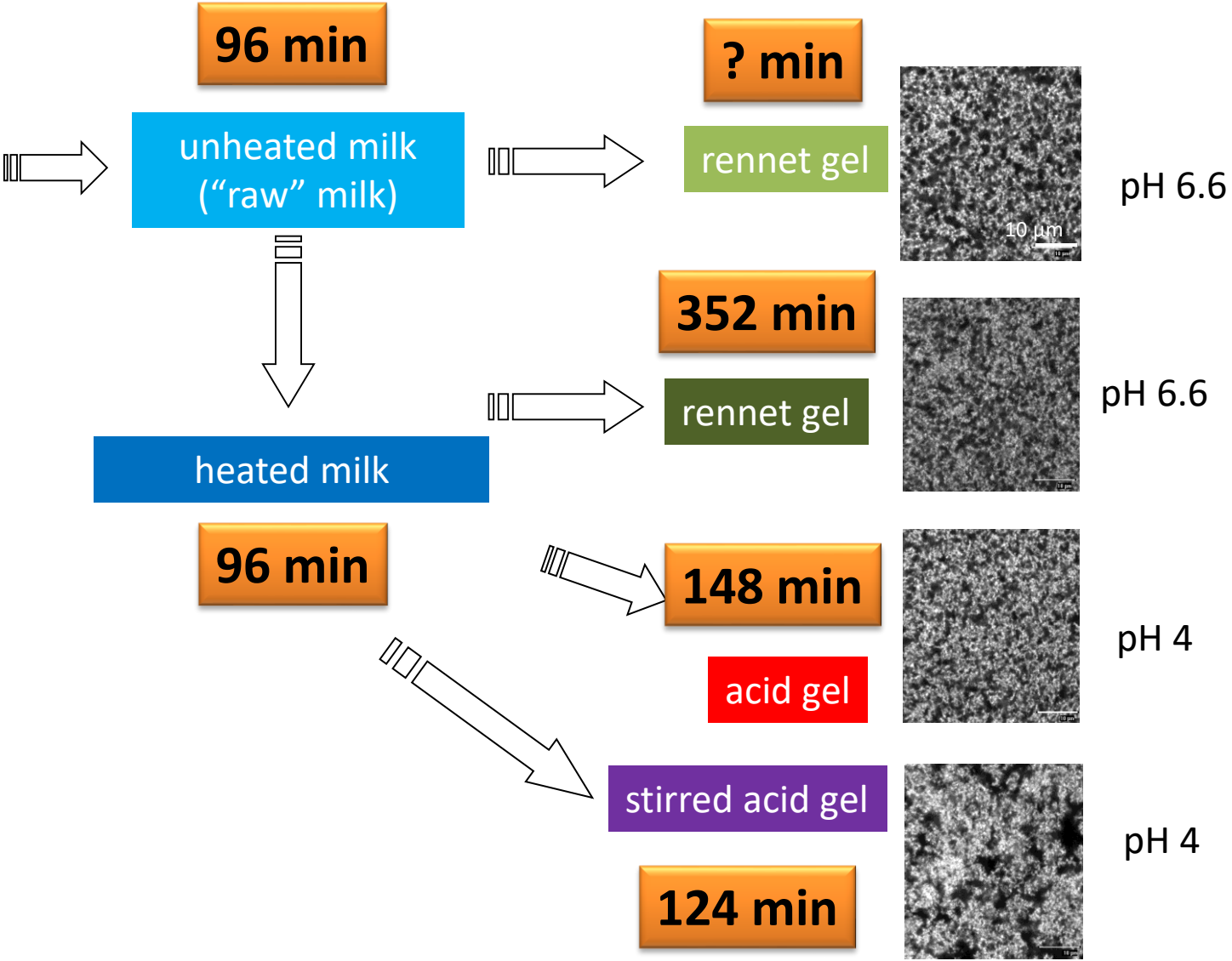


Gastric emptying half time

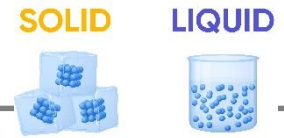


Macroscopic scale

Ultra Low Heat powder

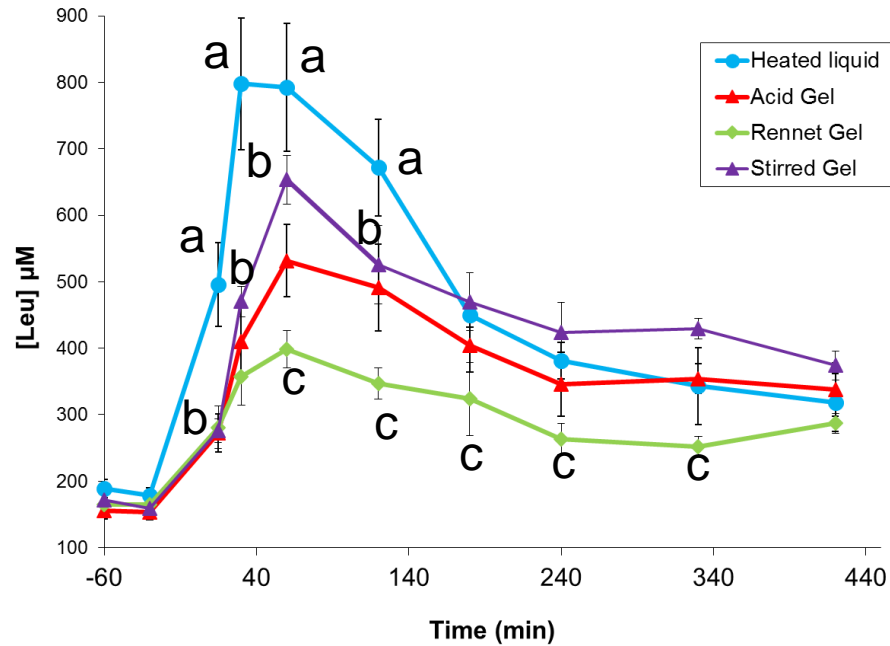


The liquid-gel transition



Macroscopic scale

Effect on absorption

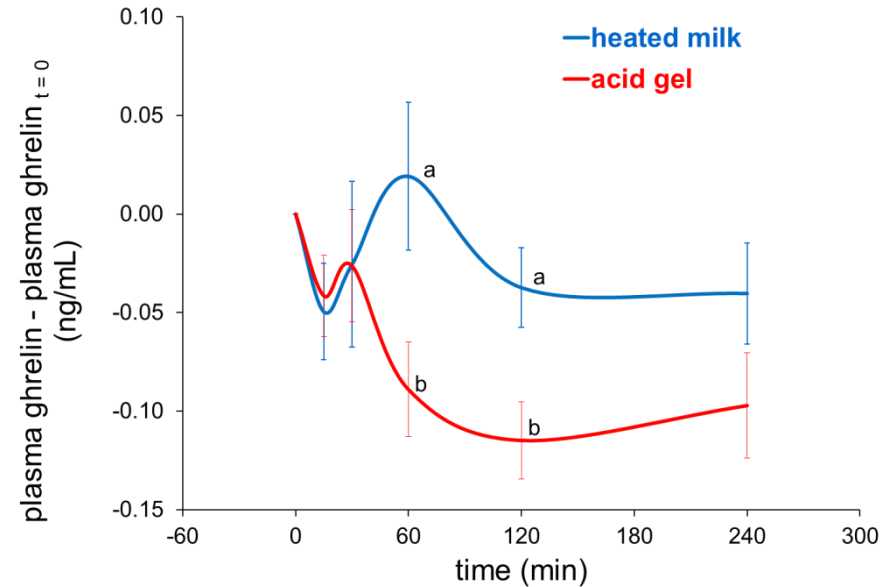


milk gelation:

→ delayed proteins transit → delayed AA absorption
maximal AA concentration in the plasma

Potential effect on satiety

ghrelin (gastrointestinal hormone → appetite stimulation)



milk gelation:

↘ postprandial ghrelin concentration =
↑ satiety ?

Bioactive peptides released during digestion differ from one matrix to another



Macroscopic scale

More than 16000 peptides identified by LC-MS-MS in the jejunum

Protein	Sequence	Activity	Reference	4	20	50	105	165	225	315
α s1	1-23	EMUL	Shimizu et al. (1984)	█						
α s1	23-34	HYP	Maruyama & Suzuki (1982)	█	█					
α s1	30-45	MB	Meisel et al. (1991)		█					
α s1	40-52	MB	Adamson & Reynolds (1996)				█	█	█	█
α s1	43-58	MB	Meisel et al. (1991)	█	█					
α s1	91-100	STRE	Miclo et al. (2001)							
α s1	99-109	MIC	McCann et al. (2006)	█	█					
α s1	167-180	MIC	Hayes et al. (2006)			█	█	█	█	
α s1	180-193	MIC	Hayes et al. (2006)	█	█	█	█	█	█	█
α s2	1-24	MB	Miquel et al. (2005)	█	█	█	█	█	█	█
α s2	124-146	MB	Miquel et al. (2005)							
α s2	183-206	TRAN	Kizawa et al. (1996)							
α s2	183-207	MIC	Recio & Visser (1999)							
α s2	189-197	HYP	Maeno et al. (1996)	█	█					
α s2	190-197	HYP	Maeno et al. (1996)	█	█					
β	1-24	MB	Bouhallab et al. (1999)	█	█	█	█	█	█	█
β	33-52	MB	Miquel et al. (2005)	█	█	█	█	█	█	█
β	60-80	OPI	Jinsmaa & Yoshikawa (1999)	█	█	█	█	█	█	█
β	98-105	OXI	Rival et al. (2001)	█	█	█	█	█	█	█
β	114-119	OPI	Jinsmaa & Yoshikawa (1999)	█	█	█	█	█	█	█
β	132-140	HYP	Robert et al. (2004)	█	█	█	█	█	█	█
β	192-209	IMM	Coste et al. (1992)	█	█	█	█	█	█	█
β	193-202	IMM	Kayser & Meisel (1996)	█	█	█	█	█	█	█
β	193-209	IMM	Coste et al. (1992)	█	█	█	█	█	█	█
κ	18-24	HYP	Lopez-Exposito et al. (2007)	█	█	█	█	█	█	█
κ	106-116	THR	Jolles et al. (1986)	█	█	█	█	█	█	█
β -lg	32-40	HYP	Pihlanto-Leppala et al. (2000)							
β -lg	92-100	MIC	Pellegrini et al. (2001)							
β -lg	142-148	HYP	Mullally et al. (1997)							

Acid Gel

Protein	Sequence	Activity	Reference	4	20	50	105	165	225	315
α s1	40-52	MB	Adamson & Reynolds (1996)							
α s1	43-58	MB	Meisel et al. (1991)							
α s1	99-109	MIC	McCann et al. (2006)							
α s1	167-180	MIC	Hayes et al. (2006)							
α s1	180-193	MIC	Hayes et al. (2006)							
α s2	1-24	MB	Miquel et al. (2005)							
α s2	189-197	HYP	Maeno et al. (1996)							
β	33-52	MB	Miquel et al. (2005)							
β	166-175	HYP	Hayes et al. (2007)							
β	193-202	IMM	Kayser & Meisel (1996)							
β -lg	92-100	MIC	(8)							
β -lg	142-148	HYP	(9)							

Rennet Gel

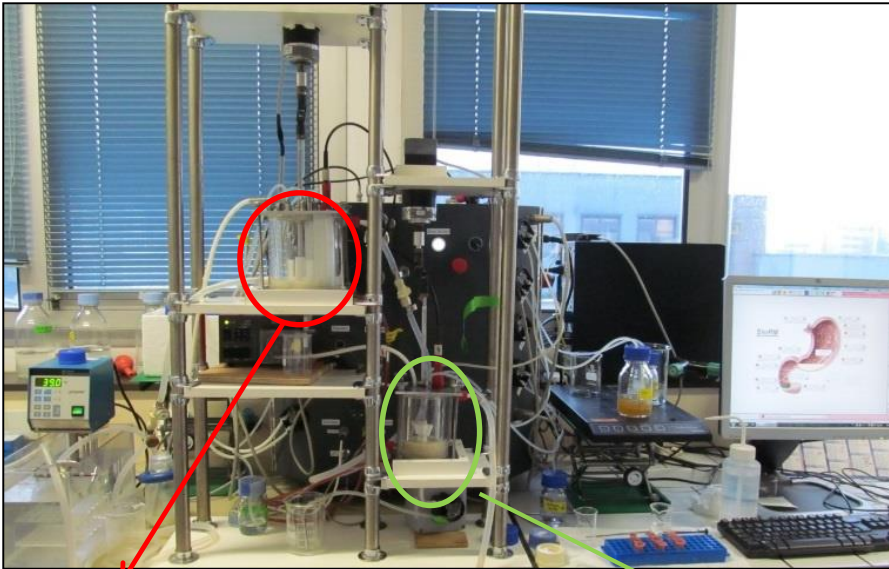
- More bioactive peptides identified during digestion of acid gel than rennet gel
- Nature of peptides is identical (clearly defined by the digestive enzyme specificity)
- Kinetics of release are different

Barbé et al. 2014
Food Res Int

Differential behaviour of acid/rennet gels in gastric conditions

- ☞ Acid/Rennet gel: identical composition, similar rheological properties and pore size
- ☞ ≠ Time of residence in the stomach (Acid 148 min /Rennet 352 min)
 - ☞ How can we explain this difference? Dynamic *in vitro* digestion of the 2 gels

**Ménard *et al.*
Food Chem 2014**



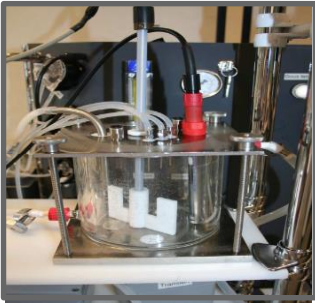
DIDGI®

StoRM® software

Stomach

Small intestine

- Pepsine
- Gastric lipase
- Simulated gastric fluid
- HCl



**Emptying :
Elashoff's model**



- Pancreatin
- Bile
- Simulated intestinal fluid
- NaHCO₃

**Emptying :
Elashoff's model**

Behaviour of acid and rennet gels in the stomach during *in vitro* dynamic digestion

Barbé et al.
Food Chem. 2014

Acid Gel



Rennet Gel



Formation of a strong coagulum with rennet gel → slow down the gastric emptying of caseins

The structure that a food adopts in the stomach is essential to understand its digestion

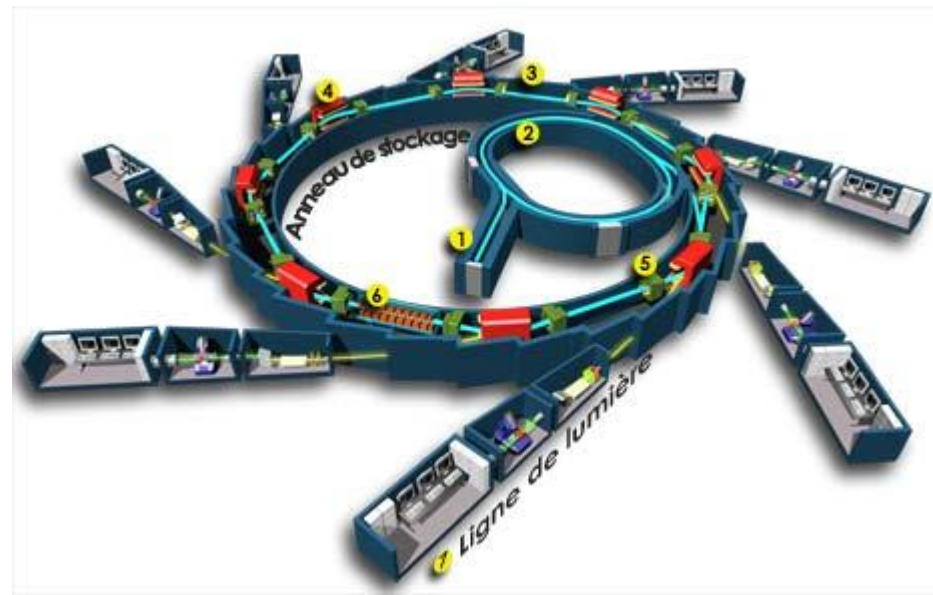


Soleil is a particle (electron) accelerator that produces the synchrotron radiation, an extremely powerful source of light that permits exploration of inert or living matter

DISCO

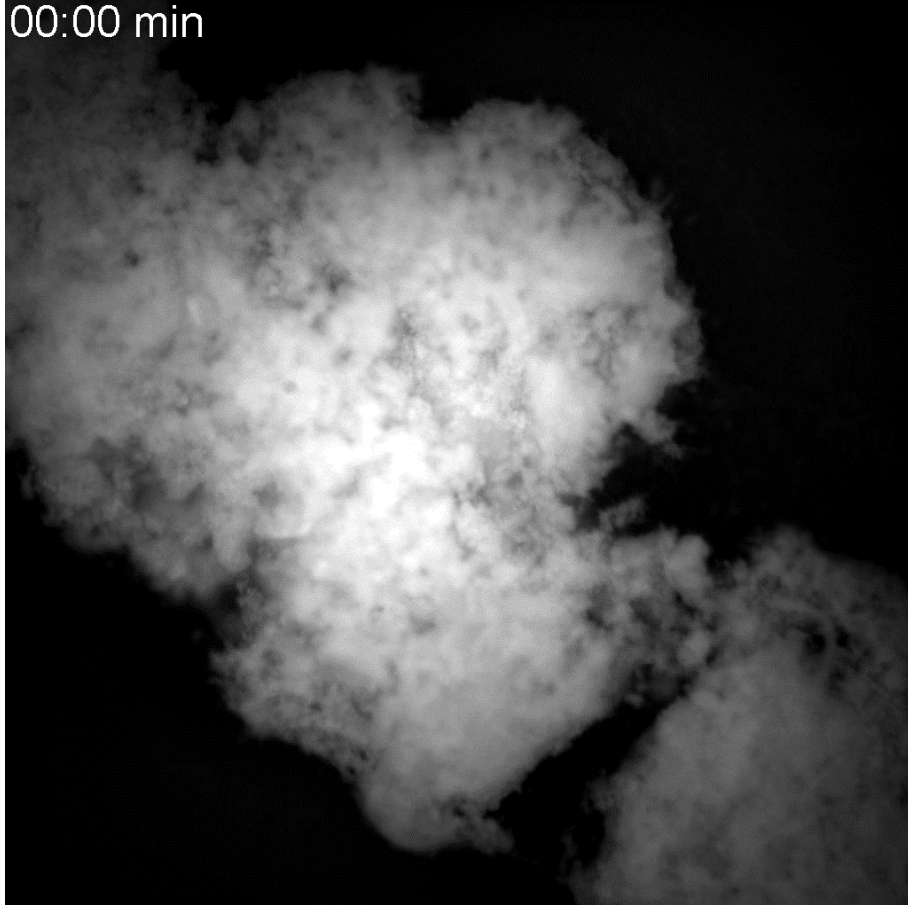
DISCO is a VUV to visible beamline dedicated to biochemistry, chemistry and cell biology. The spectral region is optimized between 60 and 700 nm with conservation of the natural polarization of the light

☞ Allow the imaging of protein intrinsic fluorescence with a UV microscope

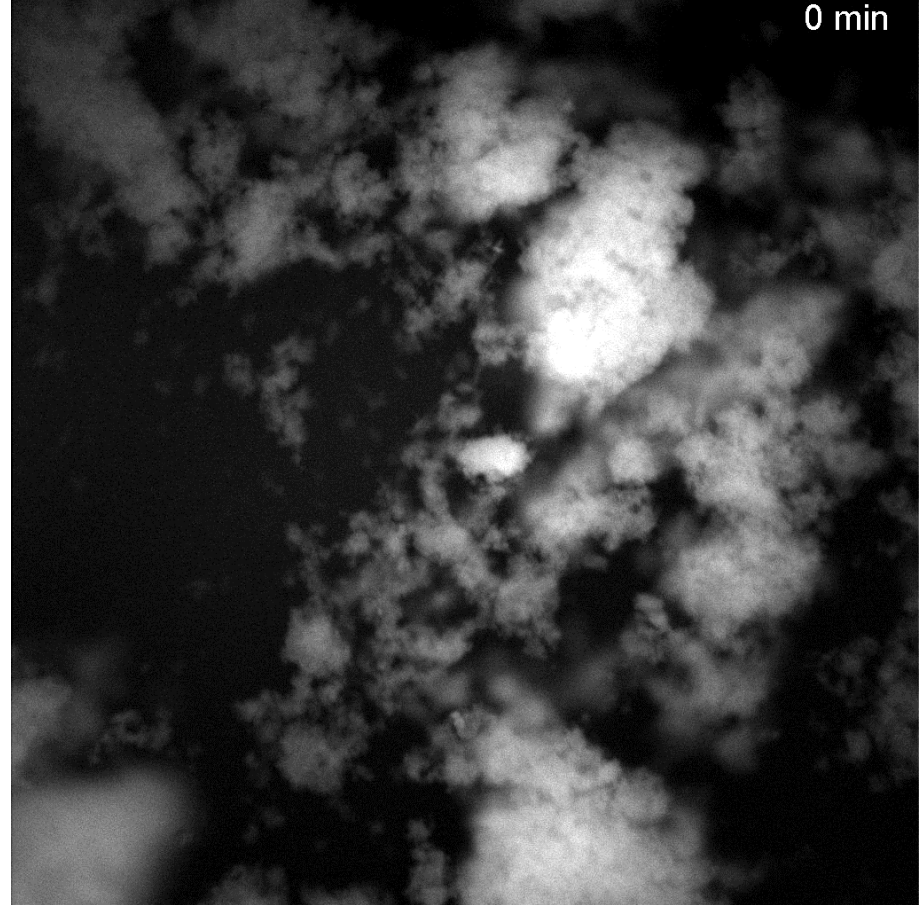


Kinetics of gel particles disintegration

00:00 min




0 min



Rennet Gel

Acid Gel



Understanding human milk digestion to design new infant formulas that will have the same behaviour in the GI tract



Deglaire A., Menard O., De Oliveira S., Bourlieu C.
& Dupont D.

INRAE, Rennes, France



Human/ bovine milk / Infant Formula

Lipid globule structure

Human milk

Bovine milk

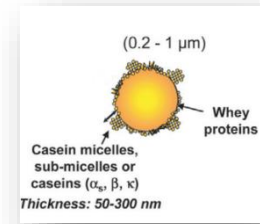
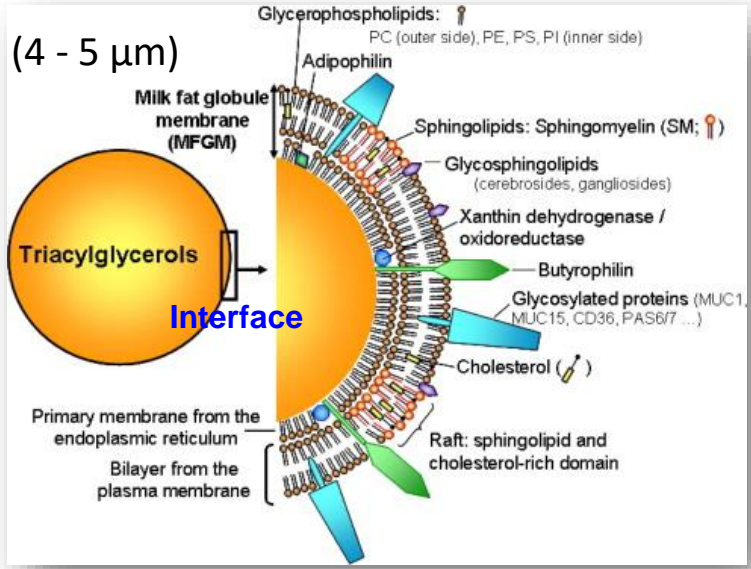
Infant Formula



Native milk fat globule

Lipid droplets

Triacylglycerols

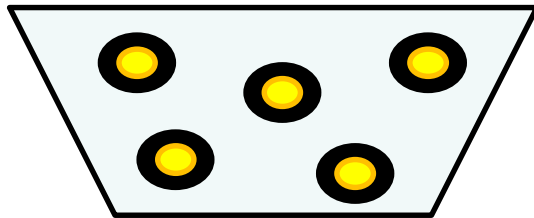


(Lopez, 2010)

(0,2 - 1 μm)
(Lopez and Briard-Bion, 2007)

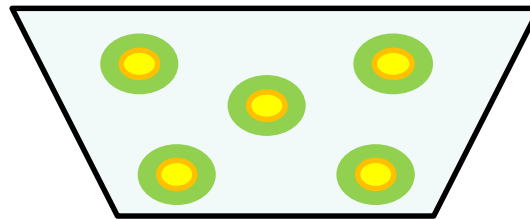
Infant formulas: can we create lipid structures biomimetic on the native fat globule?

Formula
T1



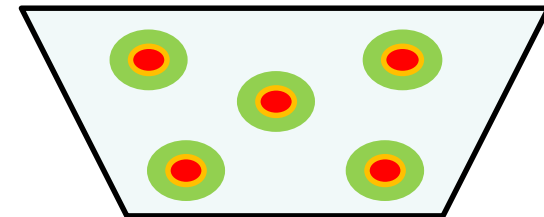
Interface 100 % Proteins
100% vegetable oil

Formula
T2

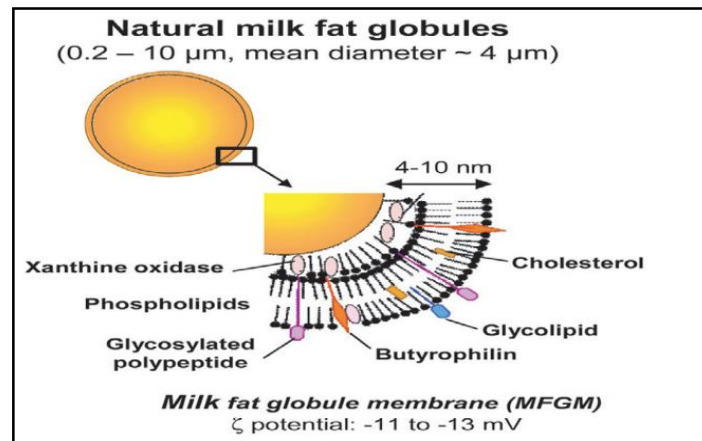


Interface 100 % phospholipids
100% vegetable oil

Formula
T3

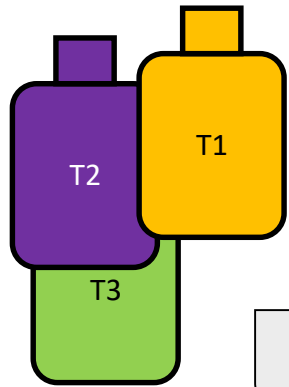


Interface 100 % phospholipids
40% vegetable oil + 60% milk fat



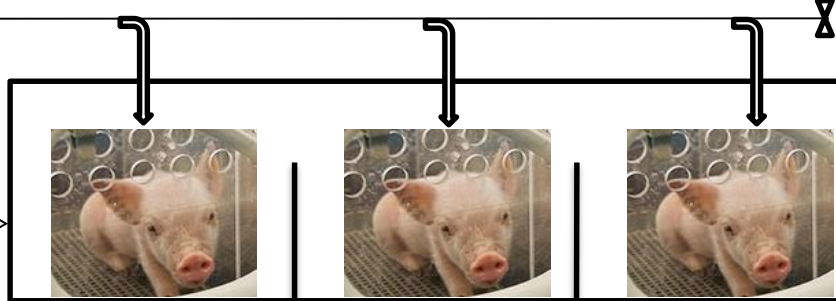
Lopez, (2007)

Can the composition of infant formula modulate the physiological response of the neonate?



- Veg
- Veg + PL
- Dairy Fat + PL

Automatic meal delivery (10 meals/ day)



+
Mother-fed piglets
(MF = + control)



Slaughtering after

7 days

28 days

(90 min postprandial)

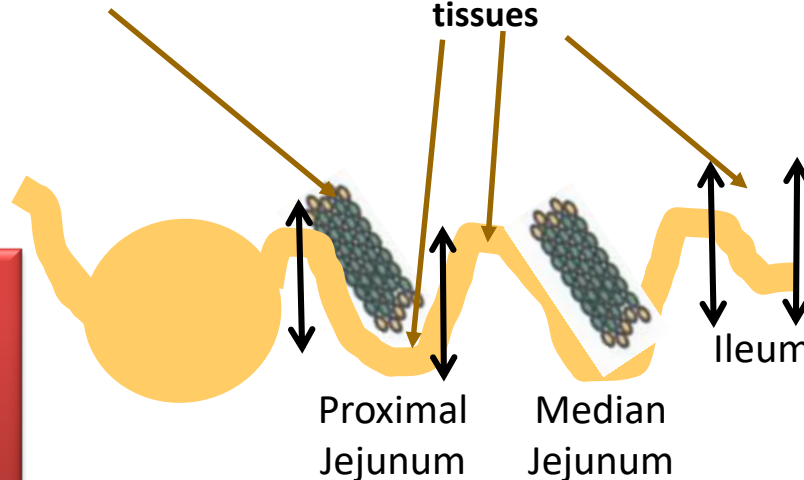
Mesenteric Lymph Nodes (MLN)

Collect of effluents and tissues

Effluents:
-SDS-PAGE
-Elisa

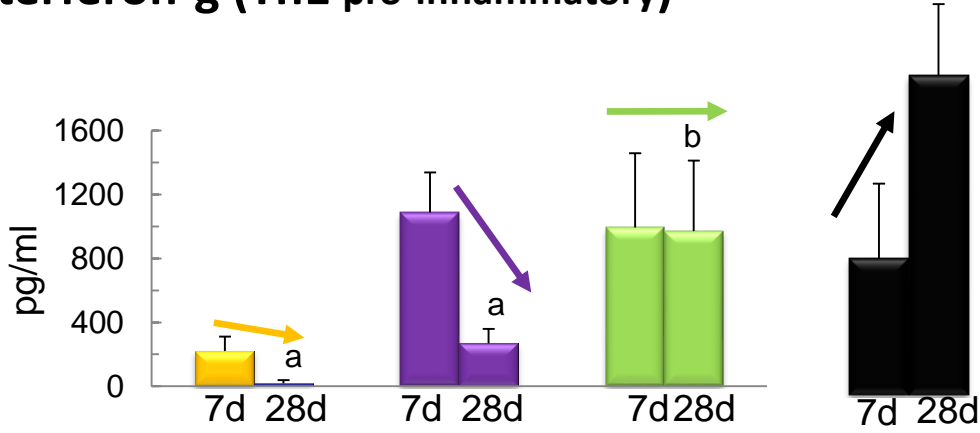
Tissues:

- Morphometry
- Enzyme Activities
- Intestinal Permeability
- Local immune response
- Microbiota

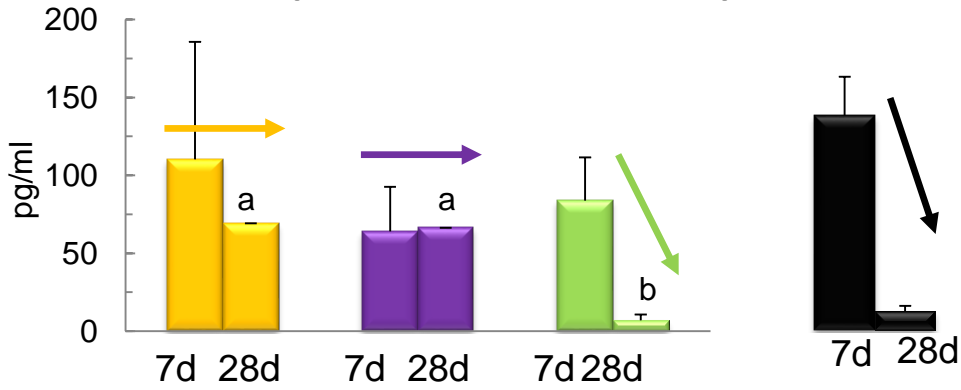


Secretory activity of MLN

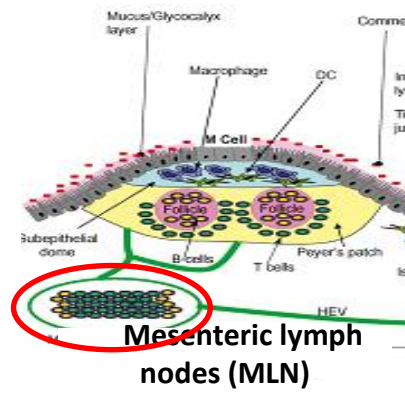
Interferon-g (Th1 pro-inflammatory)



Interleukine-10 (Th2 anti-inflammatory)



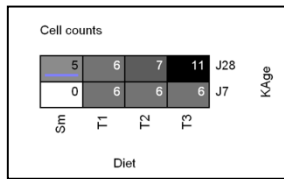
- Veg
- Veg + PL
- Dairy Fat + PL
- Porcelets SM



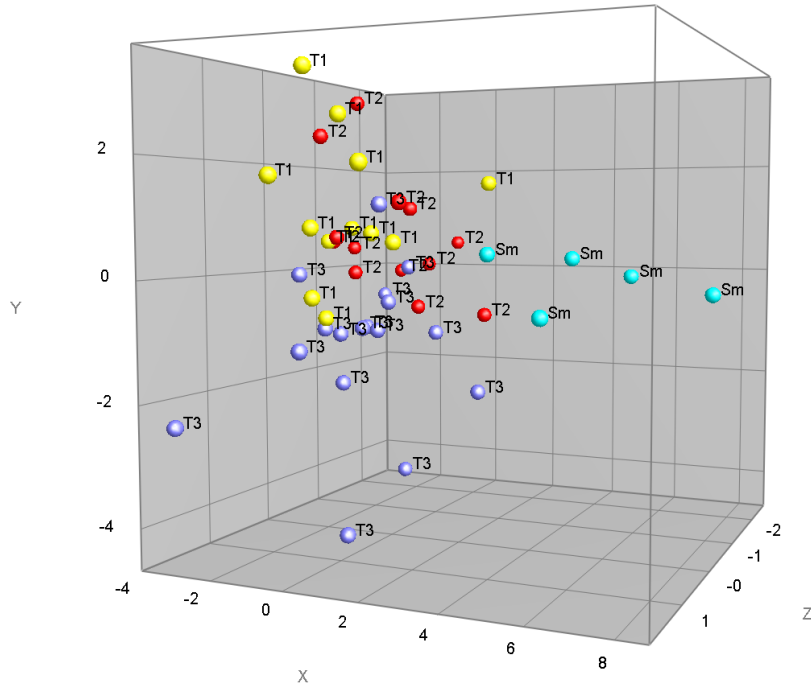
Milk lipids → maturation of the piglet's immune system more similar than with sow's milk

Le Huerou et al.
Eur J Nutr 2017

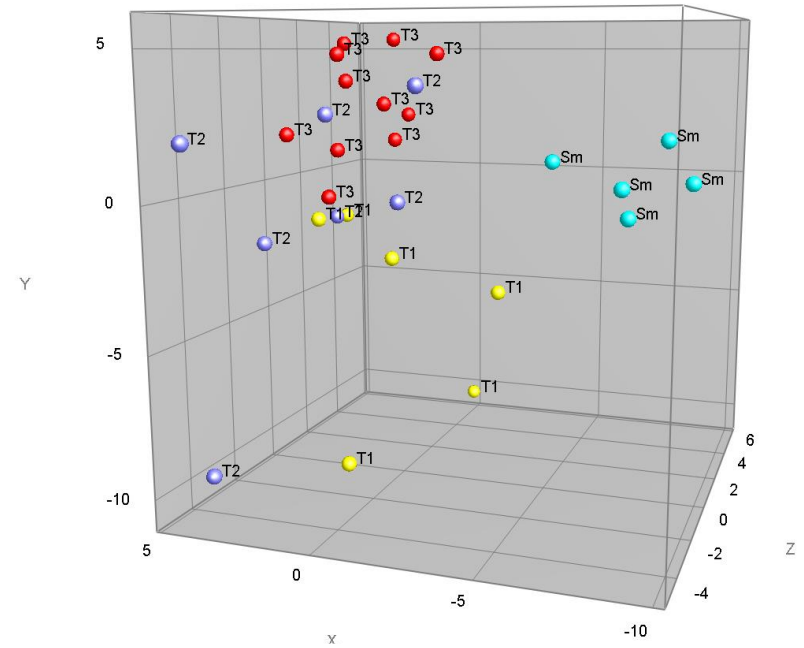
Microbiota by DHPLC



D7 & D28

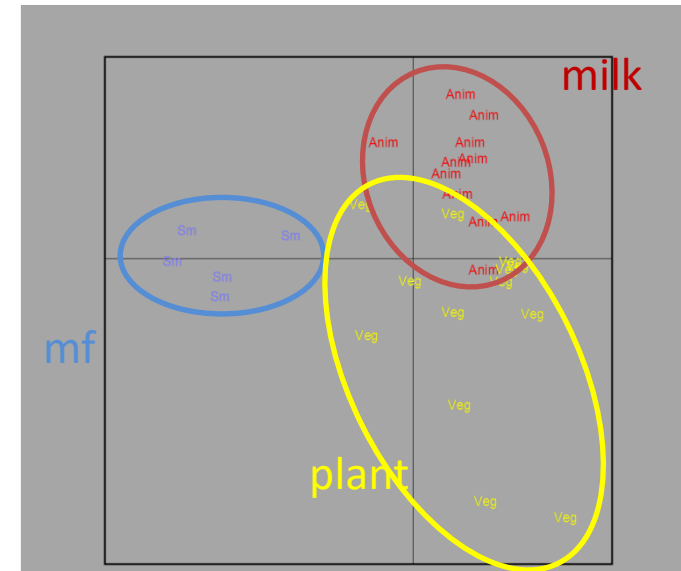


D28

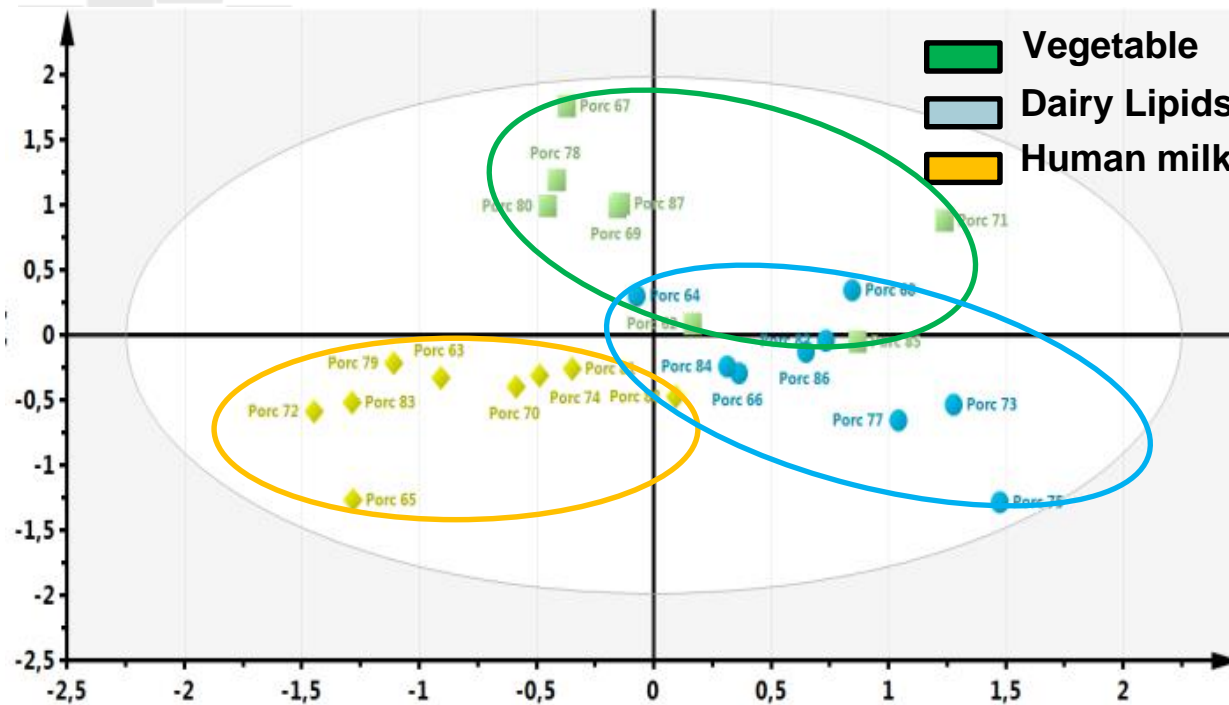


The composition/structure of the infant formula « orientates » the microbiota

More Proteobacteria with milk fat /
More Firmicutes with plant oil



What happens when they become older (140 d)?



If animals are submitted to a nutritional stress (high fat/sugar diet), some differences remain in:

- * the microbiota composition
- * the fecal metabolome with different metabolites (including propionate)
- * the immune system with a reduced susceptibility to inflammation with milk lipids

Conclusion

The structure/composition of food regulate the kinetics of protein digestion and the release of amino acids in the bloodstream

Gastric emptying rate will highly depend on the structure that the product will adopt in the stomach cavity.

Understanding the mechanisms of food particle breakdown in the stomach is critical to control the structure a food will adopt in gastric conditions

Being able to design food structures for controlling the kinetics of hydrolysis of macronutrients will allow to obtain food particularly adapted to specific population



The Bioactivity & Nutrition team at INRA Rennes



Head

Didier DUPONT - Senior Scientist

Scientists

Amélie DEGLAIRE – Lecturer

Juliane FLOURY – Lecturer

Catherine GUERIN - Lecturer

Steven LE FEUNTEUN – Senior Scientist

Joëlle LEONIL – Senior Scientist

Martine MORZEL – Senior Scientist

Françoise NAU – Professor

Frédérique PEDRONO – Lecturer

Xiaoxi YU – Post-doc

Guilherme FURTADO – Post-doc

PhD students

Yohan REYNAUD (2016-2019)

Amira HALABI (2017-2020)

Jun WANG (2018-2021)

Lea SALLELES (2018-2021)

Elise CHARTON (2019-2022)

Lucile CHAUVET (2019-2022)

Ousmane SUWAREH (2019-2022)

Technicians

Gwenaële HENRY

Yann LE GOUAR

Nathalie MONTHEAN

Engineers

Julien JARDIN

Olivia MENARD

Jordane OSSEMOND

Masters students

Improving health properties of food by sharing our knowledge on the digestive process

International Network

Dr. Didier DUPONT, Senior Scientist, INRAE, France

●
INFOGEST
●



Main objective: understanding the mechanisms of food digestion

- Develop new *in vitro*, *in vivo* and *in silico* digestion models including some for specific populations (infant, elderly)
- Harmonize the methodologies and propose guidelines for performing experiments
- Validate *in vitro* models towards *in vivo* data (animal and/or human)
- Identify the beneficial/deleterious components that are released in the gut during food digestion
- Determine the effect of the matrix structure on the bioavailability of food nutrients and bioactive molecules



Tech Univ Denmark Univ Aarhus Univ Copenhagen MTT Univ Oulu Univ Eastern Finland
 Norwegian Univ Life Sci Chalmers Univ Tech VTT Nofima Riga Stradin Univ Univ Ljubljana
 Univ Zagreb
 NIZO TNO Lund Univ

Wageningen UR Anabio
 Teagasc Univ Reading James Hutton Inst
 Univ College Cork Univ Birmingham
 Cent Rech Lippmann Univ Greenwich
 Univ Ghent Agroscope Posieux
 Inst Food Res Agrocampus Ouest
 Leatherhead Food Res ACW
 FIBL Univ Greifswald Univ Sevilla
 IRD Max Rubner-Institut
 INRA CNRS Univ Basque Country
 AgroParisTech Univ Murcia CTCPA
 CSIC Univ Granada

NIH Ricardo Jorge
 Univ Alto Douro
 ITQB Milan State Univ
 FEM CNR
 Univ Milan Univ Naples

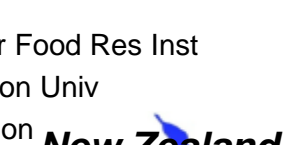
CONICET Univ Buenos Aires
 Soochow Univ
 Univ Medellin



Canada
 Laval Univ
 Univ Guelph
 Czech Univ Prague
 Inst Chem Technol



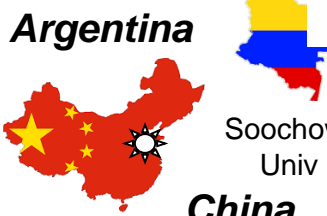
Kenya
 KTU Food Inst
 Lithuanian Univ HS
 Gdansk Univ Tech
 Maize Res Inst



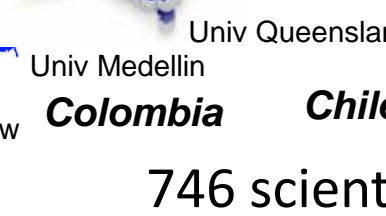
India
 NGO
 Univ Belgrade
 Polish Academy of Sci
 Univ Novi Sau



New Zealand
 Aristote Univ Thessaloniki
 Centr Food Res Inst
 Ben Gurion Univ Technion



Argentina



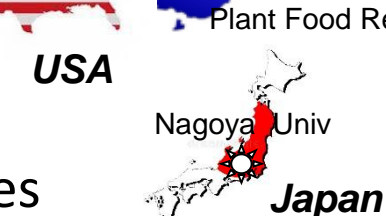
Colombia



Chile



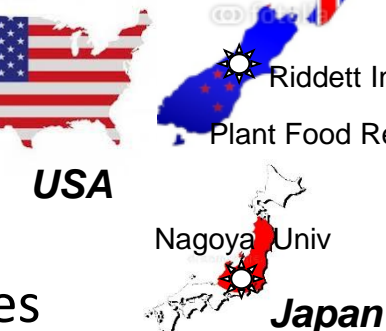
Albania



Tunisia



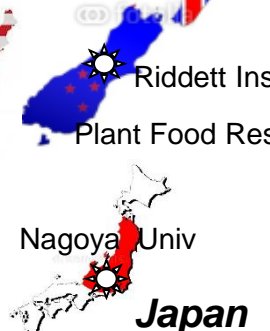
Montenegro



Brazil



USA



Japan

746 scientists - 250 institutes – 57 countries

Industry involvement

~ 60 private companies are following INFOGEST





INFOGEST



Chair

Didier Dupont - France
didier.dupont@inrae.fr



Vice-chair
Alan Mackie - UK

www.cost-infogest.eu

**In vitro
models of
digestion
WG1**

**Food
interaction –
meal digestion
WG2**

**Absorption
models
WG3**

**Digestive
lipases and
lipid digestion
WG4**

**Digestive
amylases and
starch
digestion
WG5**

**In silico
models of
digestion
WG6**



Isidra Recio



Pasquale Ferranti



Linda Giblin



Myriam Grundy



Daniela Freitas



Choi-Hong Lai



Andre
Brodkorb



Lotti Egger



Uri Lesmes



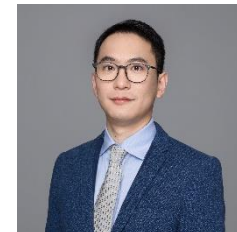
Brigitte Graf



Frederic
Carriere



Anabel
Mulet-Cabero



Bin Zhang



Steven Le Feunteun

Some outputs

In vitro gastrointestinal digestion Consensus INFOGEST protocol

Minekus et al. 2014
Food & Function, 5, 1113-1124
3125 citations

Oral phase

Mix 1:1 with Simulated Salivary Fluid (SSF)
salivary amylase (75 U/mL)
2 min, pH 7

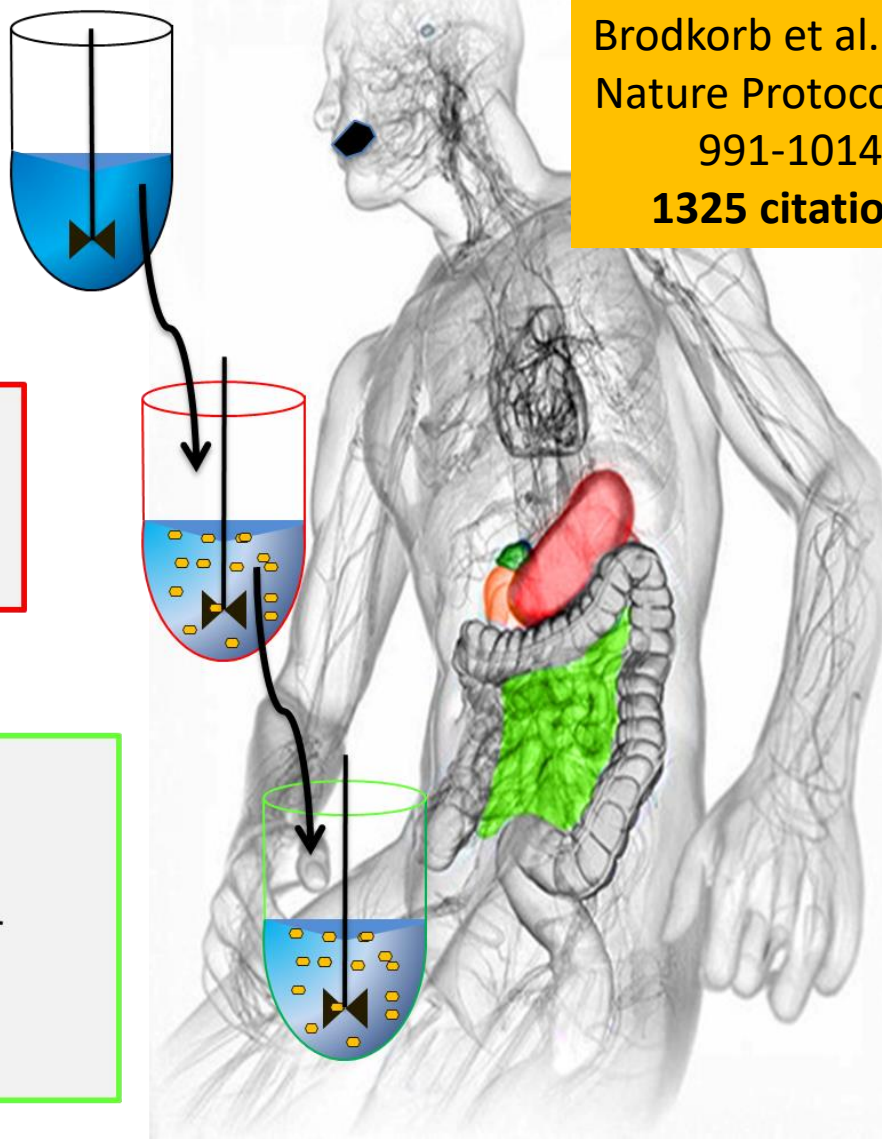
Gastric Phase

Mix 1:1 with Simulated Gastric Fluid (SGF)
Pepsin (2000 U/mL)
2h, pH 3

Intestinal Phase

Mix 1:1 with Simulated Intestinal Fluid (SIF)
Enzymes
Pancreatin (based on trypsin 100 U/mL) or
Pure enzymes
Bile (10mM)
2h, pH 7

Brodkorb et al. 2019
Nature Protocol, 14,
991-1014
1325 citations





We are pleased to announce the next
8th International Conference on Food Digestion



in Porto, Portugal, 9-11 April 2024